## Aza-Wittig-Supported Synthesis of the A Ring of Nosiheptide\*\*

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In the constant quest for lead molecules to combat infectious diseases,<sup>[1]</sup> a promising class of natural products not used for human therapy are the thiopeptide antibiotics.<sup>[2]</sup> These heterocycle-rich molecules are biosynthesized from linear ribosomal peptides<sup>[3]</sup> and block the biosynthesis of the bacterial protein very efficiently.<sup>[2,4]</sup> The bismacrocyclic nosiheptide (1; Scheme 1) stands out among them as having the highest potency against multidrug-resistant *S. aureus* (MRSA) strains.<sup>[4d,5,6]</sup> Derivatives of 1 with improved biological properties have been identified,<sup>[7]</sup> but most synthetic studies have focused on the preparation of small frag-

ments.<sup>[8,9]</sup> Notably, a study on the thioester-containing Bring of **1** was recently reported.<sup>[10]</sup> Herein, we describe the synthesis of the fully functionalized A ring of **1** by using aza-Wittig transformations.<sup>[11,12]</sup>

Nosiheptide (1) is distinguished from other thiopeptide natural products<sup>[2]</sup> by an indolic acid macrothiolactone group which forms the smaller B ring ("southern hemisphere"),<sup>[10]</sup> and by a peculiar 3-hydroxypyridine group in the larger A ring ("northern hemisphere").<sup>[10]</sup> Retrosynthetic disconnection of the indole  $3^{[8d]}$  (I, II) and introduction of latent functionality and protecting groups leads to the A-ring



**Scheme 1.** Retrosynthetic analysis of nosiheptide (1) by thioesterifications (I/II), macrolactam formation (III/IV) and aza-Wittig ring closures (V, VIII, IX; rings in bold); Bn = benzyl, Boc = tert-butoxycarbonyl, Fmoc = 9-fluorenylmethyloxycarbonyl, TBS = tert-butyldimethylsilyl, TIPS = triisopropulsilyl, Tf = trifluormethylsulfonyl, Tr = triphenylmethyl, Ts = 4-toluenesulfonyl.

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scaffold 2 as a key synthetic target. We sought to disconnect the Aring into three fragments (III, IV, V): The thiazole segment 4, the 3-hydroxypyridine core 5, and the dipeptide side chain 6. Compound 4 should be available from two smaller building blocks (VI), and elimination of a side chain could deliver the enamide (VII). We have shown before that 1-azadiene cycloadditions efficiently furnish functionalized 3hydroxypyridines such as  $5^{[13]}$  The dipeptide 6 is easily available.<sup>[14]</sup> Overall, three aza-Wittig ring closures (V, VIII, IX) were planned for introducing the thiazole rings. In this synthesis design we planned to make ideal use of the mild aza-Wittig reaction, which is an acid- and base-free kinetic condensation reaction mediated by an intermediate iminophosphorane [Eq. (1)].<sup>[11,12]</sup> Liberal selection of the heteroatom  $(X = O, S, NR)^{[12]}$  as well as the degree of oxidation in the ring  $(4.5-H_2 \text{ or } 4.5-\Delta)^{[12]}$  would offer additional flexibility.



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The synthesis began with *trans*-4-hydroxyproline **7** (Scheme 2), which was protected at the nitrogen atom by a Boc group and then transformed under Mitsunobu conditions<sup>[15]</sup> into the crystalline bicyclic lactone **8** with an inverted



Scheme 2. Synthesis of thiazole amine 15. Reagents and conditions: a)  $Boc_2O$ ,  $10\% K_2CO_3$ , 1,4-dioxane,  $0 \rightarrow 20$  °C, 8 h; b) diisopropylazodicarboxylate (1.1 equiv), PPh<sub>3</sub> (1 equiv), THF, 0 °C, 4 h; c) TceOH (4 equiv), NaH, THF, -78 °C, 1 h; d) TBSCl, DMF, 20 °C, 6 h; e) RuCl<sub>3</sub> (1 mol%), NalO<sub>4</sub> (3 equiv), CCl<sub>4</sub>/CH<sub>3</sub>CN/H<sub>2</sub>O (1:9:15), 0 °C, 8 h; f) BnOH, NaH, THF, -78 °C; g) Zn<sup>0</sup>, THF, NaH<sub>2</sub>PO<sub>4</sub> (20 mM, pH 7.0), ultrasound, 16 h; h) EDC, HOBt, 13, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C, 2 h; j) PPh<sub>3</sub>, THF,  $-20 \rightarrow 20$  °C, 4 h; j) BrCCl<sub>3</sub>, DBU, CH<sub>2</sub>Cl<sub>2</sub>, 4 h; k) TFA/CH<sub>2</sub>Cl<sub>2</sub>, 1:2, 0 °C, 30 min. DBU = 1,8-diazabicyclo[5.4.0]undec-7-ene, DMF = N,N-dimethylformamide, EDC = N-ethyl-N'-(3-dimethylaminopropyl)carbodiimide, HOBt = 1-hydroxybenzotriazole, Tce = trichloroethyl, THF = tetrahydrofuran, TFA = trifluoroacetic acid.

configuration (62%). Compound **8** was transesterified with TceOH, protected with a TBS group ( $\rightarrow$ 9, 69%), and regioselectively oxidized to lactam **10** (76%) using a catalytic amount of RuO<sub>4</sub>.<sup>[16]</sup> Ring opening of **10** was achieved with NaOBn at low temperature (87%). The resulting orthogonally protected 4-hydroxyglutamate **11** was converted into acid **12** by reduction with Zn<sup>0</sup>. Thioester formation with azidothiol **13**,<sup>[12]</sup> aza-Wittig ring closure with PPh<sub>3</sub>, and oxidation delivered building block **14** in excellent yield and purity (79%, d.r. > 98:2), which was swiftly converted into the labile amine **15** by removal of the protecting groups.

Furthermore, threonine **16** was converted into thiazole **17** by using an aza-Wittig reaction (89%; Scheme 3).<sup>[12]</sup> Removal of the *t*Bu and Boc groups and subsequent selective chain extension at the nitrogen atom was carried out on Fmocprotected Thr using EDC/HOBt ( $\rightarrow$ **18**, 99%). We found that the crucial enamide could be cleanly installed by using the method developed by Grieco et al. ( $\rightarrow$ **19**, 90%).<sup>[17]</sup> In contrast, activation of the OH group (Ms, Ts) and elimination (DBU, DMAP) gave **19** with inferior results. Palladium-



Scheme 3. Preparation of bisthiazole peptide 21. Reagents and conditions: a) TFA/CH<sub>2</sub>Cl<sub>2</sub> (1:1),  $0 \rightarrow 20$ °C, 1 h; b) Fmoc(tBu)ThrOH, EDC, HOBt, CH<sub>2</sub>Cl<sub>2</sub>/DMF (10:1),  $0 \rightarrow 20$ °C, 4 h; c) o-NO<sub>2</sub>C<sub>6</sub>H<sub>4</sub>SeCN (2 equiv), PBu<sub>3</sub> (2 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 20°C, 16 h; d) 1% H<sub>2</sub>O<sub>2</sub>, 20°C, 30 min; e) [Pd(PPh<sub>3</sub>)<sub>4</sub>], PhSiH<sub>3</sub> (2 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 20°C, 10 min; f) 15, HOBt, EDC,  $0 \rightarrow 20$ °C, 5 h; g) 1% DBU, 5% piperidine in CH<sub>2</sub>Cl<sub>2</sub>, 0°C, 30 min; h) 50% TFA,  $0 \rightarrow 20$ °C, 20 min.

mediated deallylation under neutral reaction conditions<sup>[18]</sup> then provided acid **20**, which was coupled to amine **15** and delivered segment **4** (58%). Removal of the Fmoc group provided amine **21**, which was ready for extension (96%).

The hydroxypyridine core was elaborated by a hetero-Diels-Alder cycloaddition.<sup>[13]</sup> The regiochemistry was unequivocally established by X-ray crystal structure analysis of ketone 22,<sup>[14]</sup> which was then converted into bisthiazolyl pyridine 5 by a racemization-free Hantzsch annelation (Scheme 4).<sup>[13a]</sup> The hydrolysis of diester 5 initially proved nonselective under a variety of reaction conditions. We found, however, that catalytic amounts of  $Sc(OTf)_3^{[19]}$  removed the methyl ester group on the pyridine ring selectively if a free hydroxy group at C3 was present  $(23 \rightarrow 24)$ .<sup>[20]</sup> The synthesis was initially carried on with the mandatory<sup>[13a]</sup> Boc-protected thioaminal group, but all attempts to unmask the cysteine residue at a later stage in the synthesis were unsuccessful. Therefore, the thioaminal 24 had to be cleaved at this stage. The free thiol was captured with TrCl, and an Alloc group was introduced on the nitrogen atom  $(\rightarrow 25, 82\%)$ .

To install the side chain, the hydroxy acid 25 was activated with phosgene and treated with peptide thiol 6, which was prepared in situ from the stable peptide 26 (5 % TFA, quant.). Immediate aza-Wittig ring closure gave the thiazoline, which was directly oxidized to the tris-thiazolyl pyridine 27 (46%, over 4 steps). Protection of the hydroxy group at C3 using a sulfonate group had to be carefully controlled ( $\rightarrow$ 28, 80% based on recovered starting material), then acid 29 was released using Me<sub>3</sub>SnOH.<sup>[21]</sup> Coupling of 29 to amine 21 proved challenging under many reaction conditions, but reliable transformation into 30 was achieved with DEPBT as the activating reagent (87%; 47% after preparative HPLC).<sup>[22]</sup> Parallel removal of the allyl-based protecting groups could then be cleanly achieved-despite the sulfurrich substrate 30-with Pd<sup>0</sup>/PhSiH<sub>3</sub> under neutral reaction conditions (99%).



Scheme 4. Synthesis of peptide 31. Reagents and conditions: a) (Bu<sub>4</sub>N)OH (2 equiv), 1,4-dioxane, 20°C, 5 min; b) Sc(OTf)<sub>3</sub> (5 mol%), 1,4-dioxane/H<sub>2</sub>O (3:1), pH 8.5, 60°C, 8 h; c) TFA/CH<sub>2</sub>Cl<sub>2</sub>/Et<sub>3</sub>SiH (13:13:1), 20°C, 30 min; d) TrCl, DMF, 20°C, 14 h; e) AllocCl, NaHCO<sub>3</sub>, THF/H<sub>2</sub>O (5:1); f) COCl<sub>2</sub> (20% in toluene, 1.1 equiv), NEt<sub>3</sub>, THF, -40°C, 2 h; then **6** (1.2 equiv), DMAP (0.1 equiv); g) PPh<sub>3</sub>, THF,  $-20 \rightarrow 40°C$ , 20 h; h) BrCCl<sub>3</sub>, DBU, CH<sub>2</sub>Cl<sub>2</sub>,  $-20 \rightarrow 20°C$ , 2 h; i) TSCl, NEt<sub>3</sub>, DMAP (0.1 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 0°C, 2 h; j) Me<sub>3</sub>SnOH (9 equiv), 1,2-dichloroethane, 80°C, 4 h; k) DEPBT (3 equiv), NaHCO<sub>3</sub> (10 equiv), THF, **21**, 20°C, 19 h; l) PhSiH<sub>3</sub>, [Pd(PPh<sub>3</sub>)<sub>4</sub>], CH<sub>2</sub>Cl<sub>2</sub>. Alloc = allyloxycarbonyl, DEPBT = 3-(diethoxyphos-phoryloxy)-1,2,3-benzotriazin-4(3H)-one, DMAP = 4-dimethylaminopyridine.

The stage was now set for formation of the macrolactam (Scheme 5). In the event, HATU proved to be a superb mediator of this ring-closing reaction,<sup>[23]</sup> but excess HOAt had to be minimized to avoid the formation of side products. The slow addition of amino acid **31** in nonpolar solvent gave the best results and consistently delivered the fully functionalized A ring **2** in excellent yield (82%; 56% after preparative HPLC). Notably, other cyclization strategies were less productive (data not shown), thus suggesting a very favorable conformational preorganization of **31**.

Preliminary deprotection studies of 2 showed that the thiol group could be cleanly unmasked (-33; Scheme 5). Alkylation delivered the stable thioether 34. The silyl and *t*Bu groups could be removed under standard reaction conditions (HF/pyridine, 30% TFA), with removal of the TIPS group being the most labile. Removal of the TFA-stable Ts group was achieved in parallel to base-mediated cleavage of the Bn ester group (32). These results indicate that the A-ring scaffold 2 is suited well for access to nosiheptide (1) and its derivatives.

In summary, an efficient synthesis to the fully functionalized A Ring of nosiheptide (1) was presented. En route we have demonstrated that aza-Wittig ring closures allow challenging thiopeptide functionality to be mastered. A novel  $Sc^{III}$ -mediated regioselective ester hydrolysis, highly efficient formation of a macrolactam, and manipulation strategies for the A ring featuring the unique 3-hydroxypyridine nucleus have been developed. These results will prove highly valuable for the efficient synthesis of thiopeptides such as nosiheptide,<sup>[5,7]</sup> and thus facilitate their further exploration.



**Scheme 5.** Manipulation of the A ring **2**. Reagents and conditions: a) HATU,  $EtNiPr_2$ ,  $CH_2Cl_2/DMF$  (18:1), slow addition of **31** (0.8 mM final concentration); b)  $Et_3SiH$ ,  $TFA/CH_2Cl_2$  (1:19); c) NaOH (0.35 M) in  $CH_2Cl_2/MeOH$  (1:3); d)  $ICH_2CONH_2$ , DMF. HOAt = 1-hydroxy-7-azabenzotriazole, HATU = O-(7'-azabenzotriazol-1'-yl)-1,1,3,3-tetrame-thyluronium hexafluorophosphate.

## Communications

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- a) R. O'Shea, H. E. Moser, J. Med. Chem. 2008, 51, 2871-2878; Reviews: b) F. von Nussbaum, M. Brands, B. Hinzen, S. Weigand, D. Häbich, Angew. Chem. 2006, 118, 5194-5254; Angew. Chem. Int. Ed. 2006, 45, 5072-5129; c) K. C. Nicolaou, J. S. Chen, D. J. Edmonds, A. A. Estrada, Angew. Chem. 2009, 121, 670-732; Angew. Chem. Int. Ed. 2009, 48, 660-719.
- [2] Reviews: a) M. C. Bagley, J. W. Dale, E. A. Merritt, X. Xiong, *Chem. Rev.* 2005, 105, 685–714; b) R. A. Hughes, C. J. Moody, *Angew. Chem.* 2007, 119, 8076–8101; *Angew. Chem. Int. Ed.* 2007, 46, 7930–7954.
- [3] For a discussion, see: H.-D. Arndt, S. Schoof, J.-Y. Lu, Angew. Chem. 2009, 121, 6990–6994; Angew. Chem. Int. Ed. 2009, 48, 6770–6773.
- [4] Recent progress: a) H. R. A. Jonker, S. Ilin, S. K. Grimm, J. Wöhnert, H. Schwalbe, *Nucleic Acids Res.* 2007, 35, 441–454;
  b) J. M. Harms, D. N. Wilson, F. Schlünzen, S. R. Connell, T. Stachelhaus, Z. Zaborowska, C. M. T. Spahn, P. Fucini, *Mol. Cell* 2008, 30, 26–38; c) S. Baumann, S. Schoof, S. D. Harkal, H.-D. Arndt, *J. Am. Chem. Soc.* 2008, 130, 5664–5666; d) S. Schoof, S. Baumann, B. Ellinger, H.-D. Arndt, *ChemBioChem* 2009, 10, 242–245.
- [5] a) T. Prange, A. Ducruix, C. Pascard, J. Lunel, *Nature* 1977, 265, 189–190; b) C. Pascard, A. Ducruix, J. Lunel, T. Prange, *J. Am. Chem. Soc.* 1977, 99, 6418–6423.
- [6] F. Benazet, M. Cartier, J. Florent, C. Godard, G. Jung, J. Lunel, D. Mancy, C. Pascal, J. Renaut, P. Tarridec, J. Theilleux, R. Tissier, M. Dubost, L. Ninet, *Experientia* **1980**, *36*, 414–416.
- [7] K. L. Constantine, L. Müller, S. Huang, S. Abid, K. S. Lam, W. Y. Li, J. E. Leet, J. Am. Chem. Soc. 2002, 124, 7284–7285.
- [8] Nosiheptide fragments: a) M. Iwakawa, Y. Kobayashi, S. Ikuta, J. Yoshimura, *Chem. Lett.* **1982**, 1975–1978; b) K. Koerber-Plé, G. Massiot, *J. Heterocycl. Chem.* **1995**, *32*, 1309–1315; c) K. Umemura, T. Tate, M. Yamaura, J. Yoshimura, Y. Yonezawa, C.-g. Shin, *Synthesis* **1995**, 1423–1426; d) C.-g. Shin, Y. Yamada, K. Hayashi, Y. Yonezawa, K. Umemura, T. Tanji, J. Yoshimura, *Heterocycles* **1996**, *43*, 891–898; e) K. Umemura, H. Noda, J. Yoshimura, A. Konn, Y. Yonezawa, C.-g. Shin, *Tetrahedron Lett.*

**1997**, *38*, 3539–3542; f) D. J. Bentley, J. Fairhurst, P. T. Gallagher, A. K. Manteuffel, C. J. Moody, J. L. Pinder, *Org. Biomol. Chem.* **2004**, *2*, 701–708; g) Y. Yonezawa, A. Konn, C. G. Shin, *Heterocycles* **2004**, *63*, 2735–2746; h) T. Belhadj, A. Nowicki, C. J. Moody, *Synlett* **2006**, 3033–3036.

- [9] Recent synthesis of Micrococcin P1: D. Lefranc, M. A. Ciufolini, Angew. Chem. 2009, 121, 4262-4265; Angew. Chem. Int. Ed.
   2009, 48, 4198-4201; earlier syntheses of thiopeptides: Refs. [1c,2].
- [10] M. C. Kimber, C. J. Moody, Chem. Commun. 2008, 591-593.
- [11] Recent review: F. Palacios, C. Alonso, D. Aparicio, G. Rubiales, J. M. de Los Santos, *Tetrahedron* 2007, 63, 523-575.
- [12] a) M. Riedrich, S. D. Harkal, H.-D. Arndt, Angew. Chem. 2007, 119, 2755–2758; Angew. Chem. Int. Ed. 2007, 46, 2701–2703;
  b) P. Loos, M. Riedrich, H.-D. Arndt, Chem. Commun. 2009, 1900–1902.
- [13] a) J.-Y. Lu, H.-D. Arndt, J. Org. Chem. 2007, 72, 4205–4212;
  b) J.-Y. Lu, W.-Z. Shen, H. Preut, H.-D. Arndt, Acta Crystallogr. Sect. E 2008, 64, 0602;
  c) J.-Y. Lu, J. Keith, W.-Z. Shen, M. Schürmann, H. Preut, T. Jacob, H.-D. Arndt, J. Am. Chem. Soc. 2008, 130, 13219–13221.
- [14] See the Supporting Information for details.
- [15] M. M. Bowers-Nemia, M. M. Joullié, *Heterocycles* 1983, 20, 817– 828.
- [16] a) X. Zhang, A. C. Schmitt, W. Jiang, *Tetrahedron Lett.* 2001, 42, 5335–5338; b) H. Waldmann, Y.-P. He, H. Tan, L. Arve, H.-D. Arndt, *Chem. Commun.* 2008, 5562–5564; c) review: B. Plietker, *Synthesis* 2005, 2453–2472.
- [17] P. A. Grieco, S. Gilman, M. Nishizawa, J. Org. Chem. 1976, 41, 1485–1486.
- [18] M. Dessolin, M.-G. Guillerez, N. Thieriet, F. Guibe, A. Loffet, *Tetrahedron Lett.* 1995, 36, 5741–5744.
- [19] Sc(OTf)<sub>3</sub> in transesterification: N. Remme, K. Koschek, C. Schneider, *Synlett* 2007, 491–493.
- [20] Sc<sup>III</sup> may chelate the 3-hydroxypyridine ester. Further research is underway in our laboratory to clarify this point.
- [21] K. C. Nicolaou, A. A. Estrada, M. Zak, S. H. Lee, B. S. Safina, Angew. Chem. 2005, 117, 1402–1406; Angew. Chem. Int. Ed. 2005, 44, 1378–1382.
- [22] H. T. Li, X. H. Jiang, Y. H. Ye, C. X. Fan, T. Romoff, M. Goodman, Org. Lett. 1999, 1, 91–93.
- [23] a) L. A. Carpino, J. Am. Chem. Soc. 1993, 115, 4397-4398; b) A. Ehrlich, S. Rothemund, M. Brudel, M. Beyermann, L. A. Carpino, M. Bienert, Tetrahedron Lett. 1993, 34, 4781-4784.