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Synthesis of (3S, 4R)-bengamide E

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Abstract

(3S,4R)-Bengamide E (2) was synthesized starting from D-glucono- δ -lactone (3) and the key deoxygenation step from 13 to 15 was achieved by the application of NaBH₃CN and ZnI₂. Compared with natural bengamide E (1), the synthetic compound (3S,4R)-bengamide E (2) was inactive against the cell growth of HUVEC and cancer cells. These data represent the significance of the stereochemistry at C-3 and C-4 of bengamides for structural recognition and binding with the target(s).

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Bengamides were first isolated by Crews and co-workers from *Jaspis* sponges [1,2], which are classified as fused ketide-amino acid derivatives. It was demonstrated that bengamides are a family of potent cancer cell growth inhibitors [3,4] and anti-angiogenesis agents [5] as unique inhibitors of methionine aminopeptidase type I and type II (MetAP 1 and MetAP 2). Bengamide E (1) (Fig. 1), a representative member of bengamides, exhibits 3.3 μ mol/L against MDA-MB-435 (human breast carcinoma cell) [4]. To investigate the importance of the stereochemistry at C-3 and C-4 of bengamide ketide side chain, we herein report the synthesis and biological evaluation of (3*S*,4*R*)-bengamide E (2) (Fig. 1).

The (3S,4R)-bengamide E was synthesized starting from D-glucono- δ -lactone (3) (Scheme 1). Bis-acetonide 4 was prepared from 3 according to the literature [6]; *O*-Methylation of 4 was done by MeI and NaH to afford 5 in 72% yield at -20 °C in THF without C-2 epimerization. After the reduction of carboxyl group of 5 into terminal alcohol 6 by LiAlH₄, compound 7 was prepared from 6 by using benzyl bromide and NaH with the catalysis of Bu₄NI and diol 8 was generated by removal of terminal acetonide protection of 7. Then primary and secondary hydroxyl groups of 8 were protected by acetyl group and tetrahydropyranyl (THP) group, respectively. After a new chiral center was introduced by the THP protection, every succeeding product bearing THP has a diastereomer counterpart. Subsequently, treatment of the mixture of 9 with K₂CO₃ hydrolyzed acetyl group to give separable products of 10A (more polar, 51% yield) and 10B (less polar, 41% yield) and oxidation of 10A and 10B by Swern oxidation afforded aldehyde 11A and 11B, respectively. Julia olefination of aldehyde 11A or 11B gave low *E*-selectivity and yield (data not shown), whereas, Wittig–Horner reaction [7] of aldehyde 11A or 11B produced a single product of *E*-olefin 12A or

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Fig. 1. Structures of bengamide E (1) and (3S,4R)-bengamide E (2).

12B in a good yield as expected, which was the required geometrical isomer. Reaction of **12A** with Grignard reagent MeMgI in Et₂O at 0 °C generated a tertiary alcohol **13A**. Strategically, removal of allylic tertiary hydroxyl group in **13A** could achieve the same olefin moiety as bengamide E side chain. Many approaches including Pd/C in EtOH, Pd(OH)₂ in EtOH, Raney Ni in EtOH or in H₂O and Et₃SiH in CH₂Cl₂ failed to deoxygenate **13A**. At the end, deoxygenation of **13A** was accomplished by the reduction with NaBH₃CN and ZnI₂ in CH₂Cl₂, which was modified from the literature [8]. Reductive deoxygenation of **13A** by 3.0 equiv. NaBH₃CN and 1.0 equiv. ZnI₂ afforded desired key intermediate **15** as well as THP-protected **14A** that was easily transformed into **15** by PPTS in 72% yield. As a result, the two-step process generated **15** in 62% yield in total without obvious dimerized and olefin isomerized byproducts. Through the same reactions as **12A** to **15**, the key intermediate **15** was furnished with the similar yields starting from less polar **12B**. The diastereomer mixtures were used for the subsequent production of **15** on a larger scale.

With the key intermediate **15** in hand, the free secondary hydroxyl group was protected with acetyl group, and subsequent debenzylation of terminal protection was carried out by DDQ at room temperature to afford C-1 primary alcohol **17**. Sequential oxidations of **17** with Dess-Martin periodinane (DMP) and NaClO₂ buffered with KH₂PO₄ generated carboxylic acid **18** [9]. Activation of carboxylic acid **18** was carried out by *N*-hydroxysuccinimide (NHS) with *N*,*N*'-dicyclohexylcarbodiimide (DCC) and then coupling reaction was conducted by treatment of NHS-activated **18** with freshly prepared L-(-)- α -amino- δ -caprolactam [10] to furnish the coupled compound **19**. Finally, (3*S*,4*R*)-bengamide E (**2**) [11] was achieved by the treatment of **19** with dilute TFA at 0 °C (Scheme 2).



Scheme 1. Condition and regent: (a) Ref. [6]; (b) NaH, MeI, THF, $-20 \degree C$, 72%; (c) LiAlH₄, THF, $0 \degree C$, 70%; (d) BnBr, NaH, Bu₄N⁺I⁻, THF, 86%; (e) 50% HOAc, 67%; (f) pyridine, AcCl, CH₂Cl₂, $-80 \degree C$, 63%; (g) DHP, PPTS, CH₂Cl₂; (h) K₂CO₃, MeOH/H₂O, 51% for **10A** (the more polar product) and 41% for **10B** (the less polar product); (i) Swern oxidation; (j) NaH, THF, (EtO)₂P(O)CH₂CO₂Et, $0 \degree C$ to rt, 82% for**12A** from **10A** and 72% for **12B** from **10B**; (k) MeMgI, Et₂O, $0 \degree C$, 77% for **13A**, 86% for **13B**; (l) NaBH₃CN, ZnI₂, CH₂Cl₂, rt, 22% for **14A** and 47% for **15** from **13B**; (m) PPTS, EtOH, 50 °C, 72%.



Scheme 2. Condition and regent: (a) Ac₂O, TEA, CH₂Cl₂, 89%; (b) DDQ, CH₂Cl₂, 61%; (c) DMP, CH₂Cl₂; (d) NaClO₂, KH₂PO₄, acetone/H₂O, 86%; (e) NHS, DCC, DMAP, CH₂Cl₂, rt; (f) dioxane, saturated K₂CO₃, L-(-)- α -amino- δ -caprolactam, 42%; (g) 3:1:3 THF-H₂O-TFA, 0 °C, 61%.

Compound **2** was inactive up to 50.0 μ mol/L against the cell growth of HUVEC (human umbilical vein endothelial cell), A549 (human non-small cell lung cancer cell), Bel-7402 (human hepatocarcinoma cell) and MCF-7 (human breast cancer cell) [12,13]. These structural modifications at C-3 and C-4 of bengamide side chain damaged biological activity severely, thus, suggesting that strict stereochemistry at C-3 and C-4 of bengamides is important for structural recognition and binding with the target(s).

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- [11] Compound 2: colorless semisolid; $[\alpha]_D^{20} + 33.8 (c \ 0.71, CH_3OH)$; ¹H NMR (400 MHz, CDCl₃): δ 7.65 (m, 1H), 6.40 (m, 1H), 5.76 (dd, 1H, J = 6.4, 15.6 Hz), 5.49 (m, 1H), 4.58 (m, 1H), 4.25 (m, 1H), 4.11 (m, 1H), 3.84 (m, 1H), 3.72 (m, 1H), 3.48 (s, 3H), 3.28 (m, 2H), 2.29 (m, 1H), 2.03 (m, 1H), 1.83 (m, 2H), 1.62 (m, 1H), 1.43 (m, 1H), 0.99 (d, 6H, J = 6.8 Hz); ¹H NMR (400 MHz, DMSO- d_6): δ 8.00 (m, 1H), 7.79 (d, 1H, J = 6.0 Hz), 5.58 (dd, 1H, J = 6.4, 15.6 Hz), 5.49 (dd, 1H, J = 6.0 Hz), 5.58 (dd, 1H, J = 6.4, 15.6 Hz), 5.49 (dd, 1H, J = 6.0, 16.0 Hz), 4.68 (d, 1H, J = 5.7 Hz), 4.49 (d, 1H, J = 6.4 Hz), 4.34 (m, 1H), 4.29 (d, 1H, J = 6.4 Hz), 3.87 (m, 1H), 3.68 (d, 1H, J = 5.2 Hz), ~3.35 (s, 3H, overlapped with HDO/H₂O), 3.19 (m, 1H), 3.08 (m, 1H), 2.26 (m, 1H), 1.92 (m, 2H), 1.75 (m, 1H), 1.67 (m, 1H), 1.35 (m, 1H), 1.16 (m, 1H), 0.96 (d, 6H, J = 6.4 Hz). ¹³C NMR (100 MHz, DMSO- d_6): δ 174.6, 169.9, 137.8, 129.0, 83.8, 73.8, 71.7, 70.9, 58.7, 51.8, 41.2, 31.2, 30.7, 29.1, 28.0, 22.8, 22.7; HRESIMS calcd. for C₁₇H₃₀N₂NaO₆ 381.1996, found 381.1991 [M+Na]⁺.
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