## **ARTICLE IN PRESS**

#### Tetrahedron Letters xxx (2015) xxx-xxx

Contents lists available at ScienceDirect



**Tetrahedron Letters** 

journal homepage: www.elsevier.com/locate/tetlet

# Synthesis of a monofluoro 3-alkyl-2-hydroxy-1,4-naphthoquinone: a potential anti-malarial drug

Eliana E. Kim, Evans O. Onyango, Liangfeng Fu, Gordon W. Gribble\*

Department of Chemistry, Dartmouth College, Hanover, NH 03755-3564, USA

#### ARTICLE INFO

Article history: Received 27 August 2015 Revised 10 October 2015 Accepted 15 October 2015 Available online xxxx

## ABSTRACT

Monofluorinated 3-alkyl-2-hydroxy-1,4-naphthoquinone **4** was prepared in eight steps from commercially available 8-bromooctanoic acid (**10**). The key step involved L-proline-catalyzed three-component reductive alkylation (TCRA) of 2-hydroxy-1,4-naphthoquinone (**5**) with the optically active aldehyde **7**. © 2015 Elsevier Ltd. All rights reserved.

Keywords: Three-component reductive alkylation Nucleophilic fluorination Naphthoquinone Malaria Atovaquone

Atovaquone (**1**, Fig. 1) is a 2-hydroxynaphthoquinone that is a potent anti-malarial compound in current clinical use, and which competitively inhibits the cytochrome  $bc_1$  complex of the malaria parasite *Plasmodium falciparum*.<sup>1</sup>

Due to the resistance developed to atovaquone,<sup>2</sup> other hydroxynaphthoquinones have been investigated for comparable antimalarial properties.<sup>3–5</sup> For example, we found that S-10576 ( $\mathbf{2}$ ) is a potent inhibitor of the yeast cytochrome  $bc_1$  complex that exhibits species selectivity higher than that of atovaquone.<sup>3</sup> Despite its efficacy, S-10576 (2) is readily metabolized in human cells via hepatic P450-mediated hydroxylation and subsequent oxidative carboxylation at the terminal position of the alkyl chain.<sup>6</sup> On the other hand, NQ1 (3), the trifluorinated analog of 2, is metabolically stable and strongly inhibits atovaquone-resistant P. falciparum sporozoites. However, its species selectivity is significantly lower than that of S-10576 (**2**).<sup>4,7</sup> Based on molecular modeling studies and biological assays, we and others believe that the bulkiness of the trifluoromethyl group may be responsible for this reduced selectivity.<sup>5,7</sup> Thus, we surmised that a monofluorinated 8-carbon side chain would enhance the poor species selectivity of 3, while retaining its metabolic stability, and would also recover the inhibition potency of **2**.<sup>5,7</sup> This led us to pursue the synthesis of monofluorinated 3-alkyl-2-hydroxy-1,4-naphthoquinone derivative 4.

The retrosynthetic analysis of **4** is outlined in Scheme 1. Our target molecule **4** can be partitioned into 2-hydroxy-1,4-naphthoquinone (**5**) and S-aldehyde **7**. We envisioned that **4** might be

http://dx.doi.org/10.1016/j.tetlet.2015.10.054 0040-4039/© 2015 Elsevier Ltd. All rights reserved. generated by L-proline-catalyzed three-component reductive alkylation (TCRA) of **5** with **7**. The optically active aldehyde **7** would be obtained by chiral auxiliary-mediated asymmetric  $\alpha$ -methylation of commercially available 8-bromocarboxylic acid **10** or 8-fluorocarboxylic acid **9**, itself obtained by fluorination of **10**.

etrahedro

We initially attempted the synthesis of 8-fluorocarboxylic acid 9 by treating the commercially available 8-bromooctanoic acid (10) with TBAF at 70 °C in tert-butanol. However, this reaction gave an inseparable mixture consisting majorly of the corresponding nine-membered lactone (not shown) and only a trace of the desired product 9. A search of the literature revealed a precedent for TBAF-induced intramolecular S<sub>N</sub>2-type cyclization/lactonization of halo-carboxylic acids.<sup>8</sup> To circumvent this side reaction, the carboxylic acid moiety in 10 was masked as its methyl ester 11 (Scheme 2). In the event, treating 10 with K<sub>2</sub>CO<sub>3</sub> and iodomethane afforded a 2:1 mixture (85%) of the desired product 11 and its iodo-analog **12**, respectively.<sup>9</sup> No attempt was made to separate the mixture of 11 and 12 since both were anticipated to undergo fluorination in the next step.<sup>10</sup> The fluorination of the mixture of 11 and 12 gave the desired product 13 (75%) and a small amount of the Hoffmann elimination side product 14 (5%).<sup>1</sup> Attempts to separate the mixture by column chromatography were not successful. Fortunately, though, pure fluoro ester 13 was obtained by vacuum distillation using a Vigreux column.<sup>12</sup> Finally, saponification (LiOH, H<sub>2</sub>O/MeOH) of fluoro ester 13 furnished the corresponding fluoro acid 9 in 95% yield. Overall, even with two additional steps for the protection of the carboxylic acid moiety and subsequent deprotection, this three-step fluorination method is a more efficient alternative for the preparation of fluoro acid 9

<sup>\*</sup> Corresponding author. Tel.: +1 603 646 3118; fax: +1 603 646 3946. *E-mail address*: ggribble@dartmouth.edu (G.W. Gribble).

## **ARTICLE IN PRESS**

E. E. Kim et al./Tetrahedron Letters xxx (2015) xxx-xxx



**Figure 1.** 3-Alkyl-2-hydroxy-1,4-naphthoquinones that inhibit the cytochrome *bc*<sub>1</sub> complex and our synthetic target **4**.

than that described in the literature, in which  ${\bf 9}$  was synthesized in 5 steps (61%).  $^{13}$ 

The enantioselective  $\alpha$ -methylation was performed as shown in Scheme 3. The  $\omega$ -fluorocarboxylic acid **9** was first treated with oxalyl chloride to form 8-fluorocctanoyl chloride, followed by the addition of (*S*)-4-benzyl-2-oxazolidinone using triethylamine and DMAP to give **15** in 87% yield.<sup>14</sup>

The  $\alpha$ -methylation using NaHMDS and iodomethane furnished the oxazolidinone **16** in 83% yield.<sup>14</sup> Reductive cleavage of the chiral auxiliary from **16** using LiAlH<sub>4</sub> produced the alcohol **17** in a yield of 92%.<sup>15</sup> Dess–Martin oxidation provided aldehyde **7** (92%) that was used without further purification.<sup>16</sup>

We conducted the L-proline-catalyzed three-component reductive alkylation of naphthoquinone 5 with aldehyde 7 and the Hantzsch ester (18) under the conditions reported by Ramachary.<sup>17–19</sup> Even though the authors reported good yields during the synthesis of several 3-substituted 2-hydroxy-1,4-naphthoquinones at room temperature,<sup>17</sup> in our case, the reactions at room temperature furnished the desired product **4** in only 33% yield even with extended reaction times (more than 48 h). However, refluxing CH<sub>2</sub>Cl<sub>2</sub> not only accelerated the reaction but also improved the vield from 33% to 84% (Scheme 4).<sup>20,21</sup> Overall, the best yields were obtained by using 2 equiv of the aldehyde 7, consistent with the examples reported by Ramachary.<sup>17</sup> Purification of the final product **4** was complicated by the presence of the pyridine by-product (19) from the oxidation of the Hantzsch ester (18), which exhibited the same  $R_f$  value as **4**. Attempts to remove **19** by washing the mixture with 2 N HCl failed. Alternatively, stirring the mixture with LiOH in H<sub>2</sub>O/MeOH at room temperature for 4 h followed by acid work up afforded the desired product **4** as a pale yellow solid. It is important to note that the addition of LiOH to the H<sub>2</sub>O/MeOH solution of the crude reaction mixture could have certainly furnished the final product 4; however, we ran column



Scheme 2. Synthesis of 8-fluorooctanoic acid (9).



Scheme 3. Synthesis of aldehyde 7.

chromatography prior to hydrolysis in order to recover and recycle the excess aldehyde **7**.

The alternative strategy, which delayed the introduction of the terminal fluorine until the final step, achieved the synthesis of brominated 3-alkyl-2-hydroxy-1,4-naphthoquinone **6** in five steps from bromooctanoic acid **10** (Scheme 5). This approach essentially mirrored the sequence of steps presented in Scheme 3. Moreover, the TCRA reaction of naphthoquinone **5** with aldehyde **8** (Scheme 1), the bromo analog of the aldehyde **7**, furnished the bromide **6** in 78% yield. Crystallization gave needle crystals of **6**, enabling us to assign its structure and absolute configuration using X-ray crystallography. However, the conversion of **6** to **4** using TBAF and *tert*-butanol was plagued by purification issues, in which **4** could not be separated from its mixture with other side products. In an attempt to optimize nucleophilic fluorination, we screened



Please cite this article in press as: Kim, E. E.; et al. Tetrahedron Lett. (2015), http://dx.doi.org/10.1016/j.tetlet.2015.10.054

## **ARTICLE IN PRESS**

E. E. Kim et al. / Tetrahedron Letters xxx (2015) xxx-xxx



Scheme 5. Summary of two different approaches to the total synthesis of 4.

different reaction conditions: CsF in *tert*-amyl alcohol,<sup>22</sup> TBAF in *tert*-amyl alcohol,<sup>11</sup> and TBAF in acetonitrile in 'wet' conditions.<sup>23,24</sup> All these conditions invariably produced mixtures consisting of the product **4**, alkene elimination product, and other unidentified side products that were inseparable by chromato-graphic techniques. Also, vacuum distillation, which had completely removed the fluorination side products in the first synthetic approach (vide supra), could not be employed in this case due to the solid nature of **4**. Thus, even though the second approach with fluorination at the last step would have shortened the total synthesis by two steps, the first approach with early fluorination was ultimately used to obtain pure and reliable quantities of the target compound **4**.

In conclusion, we have synthesized monofluorinated 3-alkyl-2hydroxy-1,4-naphthoquinone **4** in 8 steps in 31% overall yield from 8-bromooctanoic acid (**10**).<sup>25–31</sup> Biological evaluation of this compound as an inhibitor of the cytochrome  $bc_1$  complex of the malaria parasite is in progress and will be reported in due course.

### Acknowledgments

E.E.K. acknowledges support from the Zabriskie Fellowship Fund at the Dartmouth College. This work was supported in part by the Donors of the Petroleum Research Fund administered by the American Chemical Society.

#### Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.tetlet.2015.10. 054.

#### **References and notes**

- Kessl, J. J.; Lange, B. B.; Merbitz-Zahradnik, T.; Zwicker, K.; Hill, P.; Meunier, B.; Pálsdóttir, H.; Hunte, C.; Meshnick, S.; Trumpower, B. L. J. Biol. Chem. 2003, 278, 31312–31318.
- 2. Kessl, J. J.; Meshnick, S. R.; Trumpower, B. L. Trends Parasitol. 2007, 23, 494–501.
- Kessl, J. J.; Moskalev, N. V.; Gribble, G. W.; Nasr, M.; Meshnick, S. R.; Trumpower, B. L. Biochim. Biophys. Acta 2007, 1767, 319–326.
- Hughes, L. M.; Lanteri, C. A.; O'Neil, M. T.; Johnson, J. D.; Gribble, G. W.; Trumpower, B. L. *Mol. Biochem. Parasitol.* 2011, 177, 12–19.
- Barton, V.; Fisher, N.; Biagini, G. A.; Ward, S. A.; O'Neill, P. M. Curr. Opin. Chem. Biol. 2010, 14, 440–446.
- 6. Fieser, L. F.; Chang, F. C. J. Pharmacol. Exp. Ther. 1948, 94, 85–96.
- Hughes, L. M.; Covian, R.; Gribble, G. W.; Trumpower, B. L. Biochim. Biophys. Acta 2010, 1797, 38–43.
- Wu, C.; Brik, A.; Wang, S.; Chen, Y.; Wong, C. *ChemBioChem* 2005, *6*, 2176–2180.
  Xia, Y.; Qu, F.; Maggiani, A.; Sengupta, K.; Liu, C.; Peng, L. Org. Lett. 2011, *13*,
- 4248-4251.
- **10.** Buckle, F. J.; Pattison, F. L. M.; Saunders, B. C. J. Chem. Soc. **1949**, 1471–1479.
- 11. Kim, D. W.; Jeong, H.; Lim, S. T.; Sohn, M. Tetrahedron Lett. 2010, 51, 432–434.
- 12. Ji, Q.; Miljanic, O. J. Org. Chem. **2013**, 78, 12710–12716.
- 13. Nagatsugi, F.; Sasaki, S.; Maeda, M. J. Fluorine Chem. **1992**, 56, 373–383.
- 14. Chen, H.; Feng, Y.; Xu, Z.; Ye, T. Tetrahedron 2005, 61, 11132–11140.

- Evans, D. A.; Ennis, M. D.; Mathre, D. J. J. Am. Chem. Soc. 1982, 104, 1737–1739.
  Feutrill, J. T.; Lilly, M. J.; White, J. M.; Rizzacasa, M. A. Tetrahedron 2008, 64, 4880–4895.
- 17. Ramachary, D. B.; Kishor, M. Org. Biomol. Chem. 2010, 8, 2859–2867.
- 18. Ramachary, D. B.; Jain, S. Org. Biomol. Chem. 2011, 9, 1277–1300.
- 19. Ramachary, D. B.; Barbas, C. F. Chem. Eur. J. 2004, 10, 5323–5331.
- Mahrwald, R. In Modern Aldol Reactions. Vol. 1: Enolates, Organocatalysis, Biocatalysis and Natural Product Synthesis; WILEY-VCH Verlag GmbH & Co. KGaA, 2004; Vol. 1, p 344.
- 21. A mixture of the aldehyde 7 (100 mg, 0.624 mmol), 2-hydroxy-1,4naphthoquinone (54.0 mg, 0.312 mmol), the Hantzsch ester (79.0 mg, 0.312 mmol), and L-Proline (18.0 mg, 0.156 mmol) in  $CH_2Cl_2$  (30 mL) was refluxed for 12 h. After cooling to room temperature, SiO<sub>2</sub> (1 g) was added to the reaction mixture, which was concentrated in vacuo, then purified via drypack column chromatography (SiO2, 10:1 hexanes/ethyl acetate). <sup>1</sup>H NMR confirmed **4** as the major product contaminated with diethyl 2,6dimethylpyridine-3,5-dicarboxylate (19). LiOH·H<sub>2</sub>O (82 mg, 1.2 mmol) was added, and the mixture was allowed to stir in 3:1 MeOH/H<sub>2</sub>O (10 mL) for 4 h. After the removal of MeOH under reduced pressure, the mixture was washed with CH<sub>2</sub>Cl<sub>2</sub> and the organic extracts were discarded. The aqueous layer was covered with CH2Cl2 and acidified with 2 N HCl. After the two layers were separated, the aqueous layer was further extracted with CH<sub>2</sub>Cl<sub>2</sub> three times and the combined organic extracts were washed with brine, dried over MgSO4, and concentrated in vacuo to give **4** as a yellow solid (50 mg, 63%).  $R_f = 0.70$  (3:1 hexanes/ethyl acetate). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.11–8.13 (dd, J = 7.81, 0.79 Hz, 1H), 8.08–8.09 (dd, J = 7.61, 0.79 Hz, 1H), 7.74–7.77 (td, J = 7.75, 1.26 Hz, 1H), 7.67–7.70 (td, J = 7.55, 1.26 Hz, 1H), 7.29 (s, 1H), 4.38– 4.48 (dt, J = 47.40, 6.27 Hz, 2H), 2.58–2.62 (dd, J = 12.58, 6.15 Hz, 1H), 2.43– 2.46 (dd, J = 12.63, 8.33 Hz, 1H), 1.81-1.86 (m, 1H), 1.64-1.73 (m, 2H), 1.21-1.44 (m, 8H), 0.88 (d, J = 6.61 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  185.1, 181.6, 153.7, 135.1, 133.2, 133.1, 129.7, 127.1, 126.3, 124.2, 85.0, 83.9, 37.4, 33.0, 31.0, 30.7, 30.5, 29.7, 27.2, 25.4, 25.4, 19.9. FT-IR (CH<sub>2</sub>Cl<sub>2</sub>, cm<sup>-1</sup>) estim. 2922, 2851, 1640, 1589, 1363, 1272, 1223, 726. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) δ -217.95. HRMS (ESI) calcd for C<sub>19</sub>H<sub>24</sub>O<sub>3</sub>F [M+H]<sup>+</sup>: 319.1709, found 319.1702.  $[\alpha]_{D}^{23}$  +7.9 (CH<sub>2</sub>Cl<sub>2</sub>).
- Kim, D. W.; Ahn, D.; Oh, Y.; Lee, S.; Kil, H. S.; Oh, S. J.; Lee, S. J.; Kim, J. S.; Ryu, J. S.; Moon, D. H.; Chi, D. Y. J. Am. Chem. Soc. 2006, 128, 16394–16397.
- 23. Landini, D.; Maia, A.; Rampoldi, A. J. Org. Chem. 1989, 54, 328–332.
- 24. Albanese, D.; Landini, D.; Penso, M. J. Org. Chem. 1998, 63, 9587-9589.
- 25. Methyl 8-fluorooctanoate (**13**). Clear oil.  $R_f$  = 0.75 (3:1 hexanes/ethyl acetate). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  4.37–4.49 (dt, *J* = 47.47, 6.26 Hz, 2H), 3.67 (s, 3H), 2.29–2.32 (t, *J* = 7.52 Hz, 2H), 1.60–1.73 (m, 4H), 1.33–1.40 (m, 6H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  174.4, 84.9, 83.8, 51.7, 34.2, 30.6, 30.5, 29.2, 29.1, 27.8, 25.2, 25.0. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>)  $\delta$  –218.15.
- 26. 8-Fluorooctanoic acid (9). Clear oil.  $R_r$  = 0.45 (3:1 hexanes/ethyl acetate). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  4.38–4.49 (dt, *J* = 47.42, 6.13 Hz, 2H), 2.34–2.37 (t, *J* = 7.59 Hz, 2H), 1.62–1.74 (m, 4H), 1.35–1.43 (m, 6H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  179.0, 84.9, 83.8, 34.0, 30.6, 30.5, 29.1, 25.2, 24.8. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>)  $\delta$  –218.16.
- 27. (S)-4-Benzyl-3-(8-fluorooctanoyl)oxazolidin-2-one (**15**). Clear oil.  $R_f$  = 0.50 (3:1 hexanes/ethyl acetate). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.20–7.35 (m, 5H), 4.65–4.70 (m, 1H), 4.38–4.50 (dt, *J* = 47.49, 6.41 Hz, 2H), 4.15–4.22 (m, 2H), 3.28–3.31 (dd, *J* = 13.29, 3.08 Hz, 1H), 2.87–3.01 (m, 2H), 2.74–2.79 (dd, *J* = 13.90, 9.72 Hz, 1H), 1.65–1.75 (m, 4H), 1.37–1.45 (m, 6H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  173.5, 153.7, 135.5, 129.6, 129.2, 127.6, 84.9, 83.8, 66.4, 55.4, 38.2, 35.7, 29.2, 25.3, 25.2, 24.3. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>)  $\delta$  –218.10. FT-IR (CH<sub>2</sub>Cl<sub>2</sub>, cm<sup>-1</sup>) estim. 2932, 1781, 1698, 1387, 1212, 703. HRMS (ESI) calcd for C<sub>18</sub>H<sub>24</sub>NO<sub>3</sub>FNa [M+Na]<sup>+</sup>: 344.1638, found 344.1634.
- 28. (*S*)-4-Benzyl-3-((*S*)-8-fluoro-2-methyloctanoyl)oxazolidin-2-one (**16**). Clear oil.  $R_f$  = 0.60 (3:1 hexanes/ethyl acetate). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.21–7.35 (m, 5H), 4.65–4.70 (m, 1H), 4.37–4.49 (dt, *J* = 47.26, 6.19 Hz, 2H), 4.16–4.22 (m, 2H), 3.69–3.73 (m, *J* = 6.85 Hz, 1H), 3.25–3.28 (dd, *J* = 13.24, 3.44 Hz, 1H), 2.74–2.79 (dd, *J* = 13.54, 9.54 Hz, 1H), 1.63–1.79 (m, 3H), 1.30–1.44 (m, 7H), 1.21 (d, *J* = 7.05 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  177.5, 153.3, 135.5, 129.2, 127.6, 84.9, 83.8, 66.2, 55.6, 38.1, 37.9, 33.5, 30.6, 29.4, 27.3, 25.3. FT-IR (CH<sub>2</sub>Cl<sub>2</sub>, cm<sup>-1</sup>) estim. 2931, 1774, 1697, 1386, 1208. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>)  $\delta$  –218.10. HRMS (ESI) calcd for C<sub>19</sub>H<sub>26</sub>NO<sub>3</sub>FNa [M+Na]\*: 358.1794, found 358.1787. [ $\alpha$ ]<sup>23</sup> +76.3 (CH<sub>2</sub>Cl<sub>2</sub>).
- 29. (*S*)-8-Fluoro-2-methyloctan-1-ol (**17**). Clear oil.  $R_f = 0.50$  (3:1 hexanes/ethyl acetate). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  4.38–4.49 (dt, *J* = 47.40, 6.26 Hz, 2H), 3.49–3.52 (dd, *J* = 10.43, 5.87 Hz, 1H), 3.40–3.44 (dd, *J* = 10.41, 6.48 Hz, 1H), 1.64–1.74 (m, 2H), 1.58–1.63 (m, 1H), 1.24–1.41 (m, 8H), 0.91 (d, *J* = 6.76, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  129.1, 85.0, 83.9, 68.6, 36.0, 33.2, 29.7, 27.1, 25.4, 25.3, 16.8. FT-IR (CH<sub>2</sub>Cl<sub>2</sub>, cm<sup>-1</sup>) estim. 3355, 2930, 2858, 2360, 1458, 1038. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>)  $\delta$  –218.05. [ $\alpha$ ]<sub>D</sub><sup>23</sup> –11.7 (CH<sub>2</sub>Cl<sub>2</sub>).
- 30. (5)-8-Fluoro-2-methyloctanal (7). Clear oil.  $R_f = 0.80$  (3:1 hexanes/ethyl acetate). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.61–9.62 (d, J = 2.04 Hz, 1H), 4.38–4.49 (dt, J = 47.25, 6.11 Hz, 2H), 2.30–2.37 (m, 1H), 1.69–1.74 (m, 2H), 1.63–1.68 (m, 2H), 1.33–1.43 (m, 7H), 1.09 (d, J = 7.18 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  205.5, 85.0, 83.7, 46.5, 30.6, 30.4, 29.4, 27.0, 25.3, 13.6. FT-IR (CH<sub>2</sub>Cl<sub>2</sub>, cm<sup>-1</sup>) estim. 2936, 2860, 2359, 1811, 1706, 1457, 1004. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>)  $\delta$  –218.26. HRMS (ESI) calcd for C<sub>9</sub>H<sub>11</sub>OF [M+H]<sup>+</sup>: 159.1174, found 159.1184. [ $\alpha$ ]<sub>D</sub><sup>23</sup> +11.7 (CH<sub>2</sub>Cl<sub>2</sub>).
- (5)-2-(8-Bromo-2-methyloctyl)-3-hydroxynaphthalene-1,4-dione (6). Yellow solid. Mp 81–82 °C. R<sub>f</sub> = 0.70 (3:1 hexanes/ethyl acetate). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) & 8.10–8.12 (dd, J = 7.63, 0.83 Hz, 1H), 8.07–8.08 (dd, J = 7.57, 0.91 Hz,

4

## **ARTICLE IN PRESS**

E. E. Kim et al./Tetrahedron Letters xxx (2015) xxx-xxx

1H), 7.73–7.76 (td, *J* = 7.48, 1.33 Hz, 1H), 7.66–7.69 (td, *J* = 7.45, 1.21 Hz, 1H), 7.33 (s, 1H), 3.39 (t, *J* = 7.21 Hz, 2H), 2.57–2.61 (dd, *J* = 12.31, 5.96 Hz, 1H), 2.42–2.45 (dd, *J* = 12.51, 8.41 Hz, 1H), 1.81–1.85 (m, *J* = 7.56 Hz, 3H), 1.29–1.42 (m, 8H), 0.88 (d, *J* = 6.52 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  185.1, 181.6,

153.7, 135.1, 133.2, 133.1, 129.7, 127.0, 126.3, 124.2, 37.3, 34.2, 33.0, 33.0, 30.9, 29.1, 28.4, 27.1, 19.9. FT-IR (CH<sub>2</sub>Cl<sub>2</sub>, cm<sup>-1</sup>) estim. 3350, 2900, 1630, 1600, 1350, 1250, 700. HRMS (ESI) calcd for C<sub>19</sub>H<sub>24</sub>O<sub>3</sub>Br: 379.0909, found 379.0894.  $[\alpha]_D^{23}$ +0.9 (CH<sub>2</sub>Cl<sub>2</sub>).