# Synthesis of Cytotoxic Fluorinated Quassinoids 

Nobuhiro Ohno, ${ }^{a}$ Narihiko Fukamiya, ${ }^{a}$ Masayoshi Okano, ${ }^{\text {a,* }}{ }^{*}$ Kiyoshi Tagahara ${ }^{\text {b }}$ and Kuo-Hsiung Lee ${ }^{\text {c,* }}$<br>${ }^{a}$ Interdisciplinary Studies of Natural Environment, Faculty of Integrated Arts and Sciences, Hiroshima University, Higashi-Hiroshima 739, Japan<br>${ }^{b}$ Faculty of Pharmaceutical Sciences, Kobe Pharmaceutical University, Kobe 658, Japan<br>${ }^{\text {' Natural Products Laboratory, Divsion of Medicinal Chemistry and Natural Products, School of Pharmacy, }}$ University of North Carolina, Chapel Hill, NC 27599, U.S.A.


#### Abstract

The C-15 senecioyl side chain of brusatol was interchanged with fluorinated acyl groups, and the C-3 hydroxy group of bruceolide was esterified with fluorinated acyl chlorides. These fluorinated quassinoids 11, 12, 13, and $\mathbf{1 7}$ showed significant cytotoxic activity against eight human cancer cell lines including small and non-small cell lung, colon, CNS, ovarian and renal cancers, leukemia, and melanoma with 17 being about 100 times more potent than 11, 12, and 13 . The activity of $\mathbf{1 7}$ was similar to that of bruceantin (1) in this in vitro cell line panel. © 1997 Elsevier Science Ltd.


## Introduction

Bruceantin (1) has been tested in phase II clinical trials as an antitumor agent; however, it has not yet progressed to drug development. Oxidation of the side chain may cause deactivation of 1 and limit its efficacy. ${ }^{1}$ Accordingly, substituting the side chain with a moiety more stable to oxidation may prove beneficial. In this investigation, we have introduced a fluorinated moiety into the quassinoid side chain. This moiety was chosen based on several interesting properties: (1) the C-F bond energy ( $105.4 \mathrm{kcal} / \mathrm{mol}$ ) is higher than the $\mathrm{C}-\mathrm{H}$ bond energy ( $98.7 \mathrm{kcal} / \mathrm{mol}$ ), (2) generally, the activities and properties of fluorinated compounds mimic those of hydrogenated compounds, and (3) fluorinated compounds show increased hydrophobicity in comparison with analogous non-fluorinated compounds.

## Results

## Chemistry

The target fluorinated quassinoids were synthesized from brusatol (3), which differs from bruceantin (1) only at the C-15 side chain, or bruceolide (4). Acid hydrolysis of bruceoside-A (2), which was isolated from Brucea javanica, gave brusatol (3) as shown in Scheme 1. ${ }^{2}$ Bruceolide (4) was obtained by alkaline hydrolysis of 3.

Scheme 2 shows the preparation of the fluorinated acyl chlorides, 4,4,4-trifluoro-3-phenyl-2-butenoyl chloride (8) and 4,4,4-trifluoro-3-methylbutanoyl chloride (10). In the former, a Wittig reaction with trifluoroacetophenone (5) gave the unsaturated ester 6, which was saponified to give the acid 7 . Reaction of 7 with thionyl chloride gave the desired acid chloride 8. Compound 10 was prepared similarly from trifluoromethyl butanoic acid (9).

As shown in Scheme 3, acylation of bruceolide (4) with 8 afforded compounds 11 and 12, which were acylated at C-15 and at C-3, respectively. However, acylation of 4 with 10 occurred only at $\mathrm{C}-3$ to give compound 13 . This result suggests that the $\mathrm{C}-3$ hydroxyl is more reactive than the C-15 hydroxyl. ${ }^{3}$


(a)

(b)
4

Scheme 1. (a) $10 \% \mathrm{H}_{2} \mathrm{SO}_{4}$. (b) 0.5 N KOH .


Scheme 2.

Therefore, the $\mathrm{C}-3 \mathrm{OH}$ group of brusatol (3) was first protected with tert-butyldimethylsilyl chloride (TBDMS-C1) to afford compound 14 (Scheme 4). Then, the $\mathrm{C}-15$ side chain was removed by hydrolysis using MeOK to afford compound 15. Acylation of compound 15 with 10 gave compound 16 , which was treated with $\mathrm{Bu}_{4} \mathrm{NF}$ to remove the protecting group and afford the final compound $17 .{ }^{4}$

## Structural elucidation

Table 1 shows the ${ }^{1} \mathrm{H}$ NMR data for compounds 4 and 11-17. All compounds are colorless amorphous solids.

MS analysis shows that both compounds $\mathbf{1 1}$ and $\mathbf{1 2}$ have the same molecular formula, $\mathrm{C}_{31} \mathrm{H}_{31} \mathrm{~F}_{3} \mathrm{O}_{11}$, and contain one fluorinated moiety. The position of this moiety at C-15 in $\mathbf{1 1}$ and at $\mathrm{C}-3$ in $\mathbf{1 2}$ was confirmed by comparing the chemical shifts of $\mathrm{Me}-4$ in 11 and 12 ( $\delta 1.96$ and 1.72 , respectively) with that ( $\delta 1.96$ ) of the starting material, bruceolide (4), which is esterified at C-15.

Compound 13 has a molecular formula of $\mathrm{C}_{26} \mathrm{H}_{31} \mathrm{~F}_{3} \mathrm{O}_{11}$ and again, contains one fluorinated moiety. The upfield
shift of Me-4 ( $\delta 1.81$ in $13 ; \delta 1.96$ in 4) confirmed the esterification of C-3.

Compounds 14,15 , and 16 contain the appropriate $\mathrm{Me}-$ Si and $t \mathrm{Bu}-\mathrm{Si}$ signals in their NMR spectra (Table 1). The senecioyl moiety in $\mathbf{1 4}$ shows signals at $\delta 5.86$ for H $2^{\prime}$ and at $\delta 1.63$ and 2.15 for $\mathrm{Me}_{2}-3^{\prime}$. These signals were not observed in the spectrum of compound 15 . However, signals for the 4,4,4-trifluoro-3-methyl-butanoyl moiety were present in the spectrum of compound 16 $\left[\delta 2.91 \mathrm{~d}\left(\mathrm{H}-2^{\prime}\right), 2.53 \mathrm{~m}\left(\mathrm{H}-3^{\prime}\right)\right.$, and $\left.1.20 \mathrm{~d}\left(\mathrm{Me}-3^{\prime}\right)\right]$.

High-resolution MS of compound 17 shows a molecular formula of $\mathrm{C}_{26} \mathrm{H}_{31} \mathrm{~F}_{3} \mathrm{O}_{11}$. The Me-4 ${ }^{1} \mathrm{H}$ NMR signals were identical in $17(\delta 1.97)$ and $4(\delta 1.96)$, and established the position of the fluorinated moiety at C-15.

## Biological results

Recently, the NCI preclinical antitumor drug discovery screen has given results as $\mathrm{IC}_{50}$ mean graphs ${ }^{5}\left[\mathrm{IC}_{50}=\right.$ $50 \%$ Inhibition Concentration (M)] or $\mathrm{GI}_{50}$ mean graphs ${ }^{6}\left[\mathrm{GI}_{50}=50 \%\right.$ Growth Inhibition (M)]. These mean graphs provide a characteristic fingerprint,
(3:


Scheme 3.

Table 1. ${ }^{1} \mathrm{H}$ NMR data of compounds 4 and 11-17 (ppm in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}, J$ in Hz )

| Proton | $4^{\text {a }}$ | $11^{\text {b }}$ | $12^{\text {b }}$ | $13{ }^{\text {b }}$ | $14^{\text {a }}$ | $15^{\text {a }}$ | $16^{\text {c }}$ | $17^{\text {a }}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| H-1 $\alpha$ | $\begin{array}{r} \hline 2.57 \mathrm{~d} \\ (11) \end{array}$ | $\begin{gathered} 2.58 \mathrm{~d} \\ (10) \end{gathered}$ | $\begin{gathered} 2.58 \mathrm{~d} \\ (16) \end{gathered}$ | 2.66 | $\begin{gathered} 2.45 \mathrm{~d} \\ (15) \end{gathered}$ | $\begin{gathered} 2.45 \mathrm{~d} \\ (16) \end{gathered}$ | 2.47 | 2.50 |
| H-1 $\beta$ | $\begin{gathered} 3.31 \mathrm{~d} \\ (16) \end{gathered}$ | $\begin{gathered} 3.31 \mathrm{~d} \\ (16) \end{gathered}$ | $\begin{gathered} 3.30 \mathrm{~d} \\ (16) \end{gathered}$ | $\begin{gathered} 3.35 \mathrm{~d} \\ (16) \end{gathered}$ | $\begin{gathered} 3.22 \mathrm{~d} \\ (15) \end{gathered}$ | $\begin{gathered} 3.23 \mathrm{~d} \\ (16) \end{gathered}$ | $\begin{array}{r} 3.22 \mathrm{~d} \\ (16) \end{array}$ | $\begin{gathered} 3.28 \mathrm{~d} \\ (16) \end{gathered}$ |
| H-5 | $\begin{array}{r} 3.09 \mathrm{~d} \\ (13) \end{array}$ | 3.06 d <br> (13) | $\begin{gathered} 3.16 \mathrm{~d} \\ (13) \end{gathered}$ | 3.06 brs | 3.09 s | $\begin{gathered} 3.09 \mathrm{~d} \\ (13) \end{gathered}$ | NA ${ }^{\text {d }}$ | 3.00 brs |
| H-6 $\alpha$ | $\begin{array}{r} 2.29 \mathrm{~d} \\ (11) \end{array}$ | $\begin{gathered} 2.15 \mathrm{dd} \\ (13,13) \end{gathered}$ | $2.21 \mathrm{~d}$ <br> (7) | $\begin{array}{r} 2.33 d \\ (13) \end{array}$ | $\begin{gathered} 2.31 \mathrm{~d} \\ (15) \end{gathered}$ | $\begin{gathered} 2.28 \mathrm{~d} \\ (15) \end{gathered}$ | $\frac{2.32 \mathrm{dd}}{(14)}$ | $\begin{gathered} 2.32 \mathrm{~d} \\ (15) \end{gathered}$ |
| H-6 ${ }^{\text {a }}$ | $\begin{gathered} 1.71 \mathrm{dd} \\ (12,12) \end{gathered}$ | $\begin{gathered} 1.74 \\ (13,13) \end{gathered}$ | $\begin{gathered} 1.72 \mathrm{dd} \\ (12,12) \end{gathered}$ | $\begin{gathered} 1.66 \mathrm{dd} \\ (13,13) \end{gathered}$ | $\begin{gathered} 1.76 \mathrm{dd} \\ (12,12) \end{gathered}$ | $\begin{gathered} 1.72 \mathrm{dd} \\ (16,16) \end{gathered}$ | $\begin{array}{r} 1.78 \mathrm{dd} \\ (14,14) \end{array}$ | $\begin{gathered} 1.77 \mathrm{dd} \\ (13,13) \end{gathered}$ |
| H-7 | NA ${ }^{\text {d }}$ | $N A^{\text {d }}$ | $\mathrm{NA}^{\text {d }}$ | $\mathrm{NA}^{\text {d }}$ | 5.1 | $\mathrm{NA}^{\text {d }}$ | $N A^{\text {d }}$ | 5.10 s |
| H-9 | $\begin{gathered} 2.63 \mathrm{~d} \\ (5) \end{gathered}$ | 2.60 | 2.63 s | 2.69 | $\begin{gathered} 2.59 \mathrm{~d} \\ (5) \end{gathered}$ | $\begin{gathered} 2.60 \mathrm{~d} \\ (5) \end{gathered}$ | $N A^{\text {d }}$ | $\underset{(5)}{2.61 \mathrm{~d}}$ |
| H-11 | NA ${ }^{\text {d }}$ | 4.89 s | 4.79 s | 4.83 s | 4.78 s | $N A^{\text {d }}$ | 4.89 s | 4.78 s |
| H-12 | $N A^{\text {d }}$ | $\mathrm{NA}^{\text {d }}$ | $N A^{\text {d }}$ | $\mathrm{NA}^{\text {d }}$ | $\mathrm{NA}^{\text {d }}$ | $N A^{\text {d }}$ | $\mathrm{NA}^{\text {d }}$ | $N A^{\text {d }}$ |
| H-14 | $\begin{gathered} 3.63 \mathrm{~d} \\ (12) \end{gathered}$ | 3.96 | $\begin{gathered} 3.66 \mathrm{~d} \\ (13) \end{gathered}$ | $\begin{gathered} 3.68 \mathrm{~d} \\ (13) \end{gathered}$ | 4.03brs | $\begin{gathered} 3.63 \mathrm{~d} \\ (13) \end{gathered}$ | 3.96s | $\begin{gathered} 3.95 \mathrm{~d} \\ (13) \end{gathered}$ |
| H-15 | 6.06d <br> (13) | 6.94 s | $\begin{array}{r} 6.07 \mathrm{~d} \\ (13) \end{array}$ | $\begin{array}{r} 6.09 \mathrm{~d} \\ (13) \end{array}$ | 6.59 brs | $\begin{array}{r} 6.06 \mathrm{~d} \\ (13) \end{array}$ | 6.65 s | 6.58 brs |
| H-17 $\alpha$ | $\begin{gathered} 3.95 \mathrm{~d} \\ (8) \end{gathered}$ | $\mathrm{NA}^{\text {d }}$ | $3.95 \mathrm{~d}$ (7) | 3.99d <br> (7) | 3.94d <br> (7) | $\begin{gathered} 3.95 \mathrm{~d} \\ (7) \end{gathered}$ | 3.98 | 3.95 |
| H-173 | $N A^{\text {d }}$ | $N A^{\text {d }}$ | $N A^{\text {d }}$ | $\mathrm{NA}^{\text {d }}$ | 5.10 | $N A^{\text {d }}$ | 5.13 | 5.10 |
| Me-4 | 1.96 s | 1.96s | 1.72 s | 1.81 s | 1.86s | 1.86 s | 1.88 s | 1.97s |
| $\mathrm{Me}-10$ | 1.64 s | 1.63 s | 1.62 s | 1.81 s | 1.63 s | 1.63 s | 1.62 s | 1.63 s |
| $\mathrm{OMe}-20$ | 3.81 s | 3.84 s | 3.81 s | 3.83s | 3.76s | 3.81 s | 3.82s | 3.82 s |
| H-2' | - | $N A^{\text {d }}$ | $N A^{\text {d }}$ | $\begin{array}{r} 3.18 \mathrm{~d} \\ (19) \end{array}$ | 5.86 s | - | $\begin{array}{r} 2.91 \mathrm{~d} \\ (12) \end{array}$ | $\begin{array}{r} 2.95 \mathrm{~d} \\ (13) \end{array}$ |
| H-3' | - | - | - | 2.78m | - | - | 2.53 m | 2.50 m |
| $\mathrm{Me}-3^{\prime}$ | - | - | - | $1.27 \mathrm{~d}$ (7) | 1.63s, 2.15 s | - | $\begin{gathered} 1.20 \mathrm{~d} \\ (7) \end{gathered}$ | $\begin{gathered} 1.21 \mathrm{~d} \\ (6) \end{gathered}$ |
| Ph-3' | - | 7.45m | 7.45 m | - | - | - | - | - |
| $\mathrm{Me}-\mathrm{Si}$ | - | - | - | - | $0.36 \mathrm{~s}, 0.38 \mathrm{~s}$ | 0.35s, 0.36 s | 0.37s, 0.39s | - |
| $\mathrm{tBu}^{\text {-Si }}$ | - | - | - | - | 1.06 s | 1.05 s | 1.07 s | - |

[^0]

Scheme 4.

Table 2. $\log \mathrm{GI}_{50}$ values from the human disease-oriented cancer cell line screening panel for compounds 11, 12, 13, and $\mathbf{1 7}\left[\mathrm{GI}_{50}=50 \% \mathrm{growth}^{1}\right.$ inhibition ( $\mathrm{mol} / \mathrm{L}$ )]

| Panel/Cell Line | $11\left(\operatorname{Log~GI}{ }_{50}\right)$ | $12\left(\mathrm{Log} \mathrm{GI}_{50}\right)$ | $13\left(\operatorname{Log~GI}{ }_{50}\right)$ | $17\left(\log \mathrm{GI}_{50}\right)$ | $1\left(\mathrm{Log} \mathrm{GI} \mathrm{F}_{50}\right)$ |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Leukemia |  |  |  |  |  |
| CCRP-CEM | -5.41 | --5.06 | -5.39 | -7.92 | -8.68 |
| HL-60 (TB) | -6.44 | -5.94 | -6.57 | $<-8.43$ | -8.73 |
| K-562 | -5.31 | -5.03 | -5.22 | -7.80 | -8.42 |
| MOLT-4 | -6.00 | -5.67 | -6.11 | -8.33 | -8.42 |
| RPMI-8226 | -5.81 | -5.69 | -6.23 | -7.94 | -8.58 |
| SR | -6.18 | -5.48 | -5.56 | -8.37 | -8.74 |
| Non-Small cell lung cancer |  |  |  |  |  |
| A549/ATCC | -5.36 | -4.78 | $-4.76$ | -7.58 | -8.33 |
| EKVX | -4.92 | -4.41 | -4.58 | -6.89 | - |
| HOP-18 | -5.20 | -4.52 | -5.70 | -6.51 | -- |
| HOP-62 | $-5.22$ | -4.91 | -4.82 | -7.01 | -7.97 |
| HOP-92 | -5.67 | -5.23 | $-6.05$ | -8.02 | -7.74 |
| NCI-H226 | -5.69 | -5.19 | -5.69 | -7.97 | -8.69 |
| NCI-H23 | -5.83 | -5.34 | -5.86 | -7.90 | -8.41 |
| NCI-H322M | -4.94 | -4.52 | -4.65 | -6.40 | -7.95 |
| $\mathrm{NCI}-\mathrm{H} 460$ | -5.99 | -5.61 | -5.75 | -8.27 | -8.48 |
| $\mathrm{NCl}-\mathrm{H} 522$ | -5.62 | -5.20 | $-5.46$ | -8.05 | -8.60 |
| LXFL 529 | -5.58 | -5.13 | -5.42 | -8.14 | - |
| Small cell lung cancer $\quad-38$ |  |  |  |  |  |
| DMS 114 | -5.38 | -5.37 | $-5.84$ | -8.02 | - |
| DMS-273 | -6.01 | -5.46 | $-6.42$ | -8.16 | - |
| Colon cancer |  |  |  |  |  |
| COLO 205 | -5.18 | -4.95 | -5.17 | -7.00 | -8.61 |
| DLD-1 | -5.27 | -5.13 | $-5.56$ | -7.06 | - |
| HCC-2998 | -5.75 | -5.46 | $-5.70$ | -7.98 | - |
| HCT-116 | -5.61 | $-5.06$ | -5.61 | -8.00 | -8.49 |
| НСТ-15 | -5.10 | -4.62 | $>-4.20$ | $-6.78$ | - |
| HT29 | -5.66 | -5.17 | -5.16 | -7.88 | -8.49 |
| KM 12 | -5.24 | -4.84 | -4.95 | -7.49 | -- |
| KM20L2 | -5.41 | $-5.07$ | -5.42 | -7.71 | - -7 |
| SW-620 | - | - | -5.92 | -7.66 | -7.86 |
| CNS cancer |  |  |  |  |  |
| SP-268 | -5.06 | -4.95 | $-5.16$ | -7.06 | $-7.71$ |
| SF-295 | -5.38 | -5.10 | -5.29 | -7.35 | -7.76 |
| SF-539 | -6.00 | -5.42 | -5.82 | -7.98 | -8.07 |
| SNB-19 | -5.44 | -5.33 | -6.09 | -8.10 | -8.29 |
| SNB-75 | -5.11 | -5.40 | -5.68 | -7.35 | -7.84 |
| SNB-78 | -5.04 | -4.95 | -4.53 | -6.69 | - |
| U251 | -5.36 | -5.18 | -5.32 | -7.69 | - |
| XP498 | $-5.65$ | -5.01 | -6.89 | -8.28 | - |
| Melanoma |  |  |  |  |  |
| MALME-3M | -5.94 | -5.10 | -6.67 | -8.00 | -8.47 |
| M14 | -5.45 | $-5.06$ | -5.70 | -7.46 | -7.92 |
| M19-MEL | -5.92 | -5.29 | -6.36 | -7.79 | -75 |
| SK-MEL-2 | -5.08 | $-5.10$ | -5.53 | -7.11 | -7.58 |
| SK-MEL-28 | -5.33 | -5.27 | $-5.82$ | -7.92 | -8.14 |
| SK-MEL-5 | -5.48 | -5.32 | -5.79 | -7.90 | -8.42 |
| UACC-257 | -5.15 | -5.06 | $-5.53$ | -7.12 | - |
| UACC-62 | -5.69 | $-5.63$ | -5.95 | -8.13 | -7.89 |
| Ovarian cancer |  |  |  |  |  |
| 1GROV1 | -5.67 | -5.41 | -5.65 | $-7.77$ | -8.26 |
| OVCAR-4 | $-5.20$ | -4.75 | -4.83 | -7.17 | -8.38 |
| OVCAR-5 | -5.06 | -4.66 | -4.71 | -6.84 | -7.78 |
| OVCAR-8 | -4.98 | -4.92 | -5.33 | -7.24 | -7.90 |
| SK-OV-3 | -5.14 | -5.10 | $-5.28$ | -7.29 | -8.01 |
| Renal cancer |  |  |  |  |  |
| 786-0 | -5.10 | -5.04 | $-5.72$ | -7.19 | -7.57 |
| A498 | -5.74 | -6.28 | -6.81 | -7.67 | -7.78 |
| ACHN | -5.38 | -4.86 | -5.08 | -7.64 | -8.39 |
| CAKI-I | -5.12 | -4.70 | $-5.03$ | -7.25 | -7.65 |
| RXF-393 | -5.22 | -4.77 | -5.33 | -7.04 | - |
| SN12C | --5.06 | $-5.10$ | $-5.70$ | -6.84 | - |
| TK-10 | -5.11 | $-5.00$ | -5.26 | -7.18 | - |
| UO-31 | -5.17 | -4.45 | -4.97 | $-6.83$ | - |
| MG-MID | -5.44 | -5.13 | -5.54 | $-7.56$ | -8.10 |

displaying the individual cell lines that are proportionally more sensitive than average or proportionally less sensitive than average to the test compound.
$\log \mathrm{GI}_{50}$ values from the human disease-oriented cancer cell line screeening panel for compounds 11, 12, 13, and 17 are shown in Table 2, together with the data for bruceantin (1). The values of 'MG-MID' are mean values of all $\log \mathrm{GI}_{50}$ values, which for $\mathbf{1 1}, \mathbf{1 2}, \mathbf{1 3}$, and 17 are $-5.44,-5.13,-5.54$, and -7.56 , respectively, compared with -8.10 for 1 . On the other hand, mean $\log \mathrm{IC}_{50}$ values for 5 -fluorouracil, 5 -fluorodeoxyuridine, and bleomycin are $-3.5,-4.7$, and -5.2 , respectively. ${ }^{5}$ These data indicate that the fluorinated quassinoids are potent cytotoxic agents. Notably, the cytotoxic activity of 17 is about 100 -times higher than that of 11,12 , and 13. The position of the 4,4,4-trifluoro-3-phenyl-2-butenoyl group did not affect the potency (compare 11 to $\mathbf{1 2}$ ). However, with a 4,4,4-trifluoro-3-methylbutanoyl group, the C-15 esterified compound (17) was much more active than the $\mathrm{C}-3$ substituted analogue (13). In this in vitro cell line panel, compound 17 was approximately as active as bruceantin (1). Therefore, this fluorinated component is an ideal lead compound for further study.

## Experimental

## General experimental procedures and materials

Melting points were measured on an MRK air-bathtype melting point apparatus and are uncorrected. Specific rotations were obtained on a Jasco DIP-370 polarimeter $(L=0.5 \mathrm{dm})$. IR and UV spectra were recorded on a Jasco IR-810 spectrometer and a Hitachi $320-\mathrm{S}$ spectrometer, respectively. ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR spectra were determined on a Varian XL-200 (200.6 MHz for ${ }^{1} \mathrm{H}$ NMR ), a Varian VXR-500 ( 499.8 MHz for ${ }^{1} \mathrm{H}$ NMR and 125.7 MHz for ${ }^{13} \mathrm{C}$ NMR) and a Jasco GSX-500 ( 500.1 MHz for ${ }^{1} \mathrm{H}$ NMR and 125.7 MHz for ${ }^{13} \mathrm{C}$ NMR) instrument in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}$ or $\mathrm{CDCl}_{3}$, using TMS as an internal standard. Mass spectra were recorded on a Hitachi M80 or a Hitachi M-2500 instrument.

Analytical HPLC was performed on a Tosoh liquid chromatograph equipped with a UV detector at 254 nm and a reverse-phase column (TSK-gel ODS-80T ${ }_{\mathrm{M}}$, $4.6 \times 150 \mathrm{~mm}$ ), using a mixed solvent of $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}$ or $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}-\mathrm{HOAc}$. Preparative HPLC was carried out on a Waters or a Tosoh liquid chromatograph equipped with a reverse-phase column (Lichrosorb RP$18,10 \times 250 \mathrm{~mm}$ or Dynamax-60A, $21.4 \times 250 \mathrm{~mm}$ ) and a UV monitor set at 254 nm , using the same solvents as those used in the analytical HPLC. The final compounds 11, 12, 13, and 17 were homogeneous as determined by analytical HPLC.

Silica gel (Merck, type 60, 70-230 mesh) was used for column chromatography. Precoated Si gel plates (Merck, $60 \mathrm{~F}_{254}$ ) of 0.25 mm thickness were used for analytical TLC, and plates of 1 mm and 2 mm thickness
were used for preparative TLC. Components were detected by using a UV lamp ( 254 nm ). Bruceoside-A (2), was obtained from the fruits of Brucea javanica, as previously described in the literature. ${ }^{2}$ Compounds 5 and 9 used in this investigation were commercially available.

Brusatol (3). Bruceoside-A (2, $209 \mathrm{mg}, 0.306 \mathrm{mmol}$ ) was dissolved in $\mathrm{MeOH}\left(40 \mathrm{~mL}\right.$ ), and $10 \% \mathrm{H}_{2} \mathrm{SO}_{4}(20$ mL ) was added to the solution. The mixture was refluxed for 6 h . The reaction product was extracted with $\mathrm{CHCl}_{3}$ and purified by preparative HPLC to afford brusatol (3, $137 \mathrm{mg}, 0.263 \mathrm{mmol}, 86 \%$ yield) as a colorless amorphous solid. It was identified by comparing its TLC, HPLC, and IR spectra with those of an authentic sample.

Bruceolide (4). Brusatol (3, $39.2 \mathrm{mg}, 0.075 \mathrm{mmol}$ ) was dissolved in MeOH ( 4 mL ), and $0.5 \mathrm{~N} \mathrm{KOH}-\mathrm{MeOH}$ solution ( 1.2 mL ) was added to the solution. The mixture was stirred at room temperature for 8.5 h . The reaction mixture was neutralized with anion exchange resin (Dowex 50W-X2) and extracted with MeOH to afford crude bruceolide ( $4,37.6 \mathrm{mg}$ ). This compound was purified by preparative HPLC (Lichrosorb RP-18, $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}, 7: 3, \mathrm{v} / \mathrm{v}$ ) to afford pure bruceolide (4, 14.5 $\mathrm{mg}, 0.033 \mathrm{mmol}, 44 \%$ yield) as a colorless amorphous solid. It was identified by comparing its TLC, HPLC, and IR spectra with those of an authentic sample.

Ethyl-3-phenyl-4,4,4-trifluoro-2-butenoic acid (6). Triethyl phosphonoacetate ( $4.93 \mathrm{~g}, 22 \mathrm{mmol}$ ) was dissolved in dry $\mathrm{Et}_{2} \mathrm{O}(40 \mathrm{~mL})$ with stirring at $0{ }^{\circ} \mathrm{C}$ under $\mathrm{N}_{2}$. Butyl lithium ( $15 \%$ hexane solution, 15 mL , 24 mmol ) was added dropwise to the solution at $0{ }^{\circ} \mathrm{C}$ over 10 min , and the mixture was stirred at room temperature for 30 min . 2,2,2-Trifluoroacetophenone $(5,3.49 \mathrm{~g}, 20 \mathrm{mmol})$ was dissolved in dry $\mathrm{Et}_{2} \mathrm{O}(6 \mathrm{~mL})$ and added to the reaction mixture. Stirring was then continued at room temperature for 17 h . The reaction product was extracted with $\mathrm{Et}_{2} \mathrm{O}$; evaporation of the solvent afforded compound $6(4.70 \mathrm{~g}, 19 \mathrm{mmol}, 95 \%$ yield) as a yellow liquid; IR (film) $\mathrm{cm}^{-1} 1730(\alpha, \beta-$ unsaturated $\mathrm{C}=\mathrm{O}), 1650(\mathrm{C}=\mathrm{C})$; ${ }^{1} \mathrm{H}$ NMR ( $\mathrm{CDCl}_{3}$ ) $\delta 7.41$ (brs, 5 H ), 6.36 (brs, 1 H ), 4.32 (q, $J=7 \mathrm{~Hz}, 2 \mathrm{H}$ ), $1.33(\mathrm{t}, J=7 \mathrm{~Hz}, 3 \mathrm{H})$. HPLC showed the compound to be a mixture of the $E$ and $Z$ isomers.
(Z)-3-Phenyl-4,4,4-trifluoro-2-butenoic acid (7). Compound 6 ( $E, Z$ mixture, $1.35 \mathrm{~g}, 5.5 \mathrm{mmol}$ ) was dissolved in $\mathrm{MeOH}(15 \mathrm{~mL})$, and $10 \% \mathrm{KOH}$ aqueous solution ( 15 mL ) was added to the solution, and the mixture was stirred at $0{ }^{\circ} \mathrm{C}$ for 3 h . Trifluoroacetic acid ( 2 mL ) was added to the reaction mixture, and the product was extracted with $\mathrm{Et}_{2} \mathrm{O}$ to afford compound $7(E, Z$ mixture, $1.08 \mathrm{~g}, 5.00 \mathrm{mmol}, 90.4 \%$ yield). The $E$ and $Z$ isomers of 7 were separated by preparative TLC ( $\mathrm{EtOAc}: \mathrm{Et}_{2} \mathrm{O}, 1: 1, \mathrm{v} / \mathrm{v}$ ) to afford the pure $Z$ isomer (440 $\mathrm{mg}, 2.04 \mathrm{mmol}, 63.8 \%$ yield) as pale-yellow needles; IR ( KBr ) $\mathrm{cm}^{-1} 3000$ (carboxylic OH ), 1720 ( $\alpha, \beta$-unsaturated $\mathrm{C}=\mathrm{O}), 1650(\mathrm{C}=\mathrm{C}) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.42$ (brs, 5 H ), $6.59(\mathrm{~s}, 1 \mathrm{H})$.
(Z)-3-Phenyl-4,4,4-trifluoro-2-butenoyl chloride (8). A mixture of $7(Z$ isomer, $186 \mathrm{mg}, 0.860 \mathrm{mmol}), \mathrm{SOCl}_{2}$ ( $660 \mu \mathrm{~L}$ ), and dry benzene ( 2 mL ) was refluxed for 11 h under $\mathrm{N}_{2}$. Excess $\mathrm{SOCl}_{2}$ and benzene were removed by evaporation to afford compound 8 ( $194 \mathrm{mg}, 0.826$ mmol, $96 \%$ yield) as a brown liquid: IR (film) $\mathrm{cm}^{-1}$ $1800(\mathrm{C}=\mathrm{O}), 1645(\mathrm{C}=\mathrm{C}) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CHCl}_{3}\right) \delta 7.53$ (brs, 5H), 6.80 (brs, 1H).

3-Trifluoromethylbutanoyl chloride (10). Compound 10 ( $583 \mathrm{mg}, 3.3 \mathrm{~mol}, 50 \%$ yield) was obtained from $3-$ trifluoromethylbutanoic acid $(9,1.05 \mathrm{~g}, 6.7 \mathrm{mmol})$ in an identical manner to that of compound 8 from compound 7. Colorless liquid: IR $\left(\mathrm{CHCl}_{3}\right) \mathrm{cm}^{-1} 1800$ $(\mathrm{C}=\mathrm{O}) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 3.33(\mathrm{~d}, J=12 \mathrm{~Hz}, 2 \mathrm{H})$, $2.53(\mathrm{~m}, 1 \mathrm{H}), 1.24(\mathrm{~d}, J=7 \mathrm{~Hz}, 3 \mathrm{H})$.

15- and 3-(4', $4^{\prime}, 4^{\prime}$-Trifluoro- $3^{\prime}$-Phenyl- $\mathbf{2}^{\prime}$-butenoyl)-bruceolide (11 and 12). Bruceolide ( $4,26.5 \mathrm{mg}, 0.061$ mmol ) was dissolved in dry pyridine ( $400 \mu \mathrm{~L}$ ), and compound $\mathbf{8}$ ( $64.7 \mathrm{mg}, 0.276 \mathrm{mmol}$ ) was dissolved in dry $\mathrm{CHCl}_{3}(500 \mu \mathrm{~L})$. The solution of 8 was added to the solution of 4 at $0^{\circ} \mathrm{C}$, and the mixture was stirred at $57^{\circ} \mathrm{C}$ for 48 h under $\mathrm{N}_{2}$. Excess 8 was decomposed with EtOH $(500 \mu \mathrm{~L})$, and the acylation product was extracted with $\mathrm{CHCl}_{3}$ to afford a brown oil ( 56 mg ) that contained compounds 11 and 12. Pure compounds $11(2.1 \mathrm{mg}$, $0.0033 \mathrm{mmol}, 5.5 \%$ yield) and $12(3.7 \mathrm{mg}, 0.0058 \mathrm{mmol}$, $9.6 \%$ yield) were obtained by preparative HPLC (Lichrosorb RP-18, $\mathrm{H}_{2} \mathrm{O}: \mathrm{MeOH}: \mathrm{AcOH}, ~ 49.5: 49.5: 1$, $\mathrm{v} / \mathrm{v}$ ).

15-( $\mathbf{4}^{\prime}, 4^{\prime}, 4^{\prime}$-Trifluoro- $3^{\prime}$-phenyl- $\mathbf{2}^{\prime}$-butenoyl)-bruceolide (11). Colorless amorphous solid; mp $154-156^{\circ} \mathrm{C} ;[\alpha]_{\mathrm{D}}{ }^{27}$ $-13.3^{\circ}$ (c 0.030, EtOH); UV (EtOH) nm 270 ( $\varepsilon$ 1900); IR ( KBr ) $\mathrm{cm}^{-1} 3400(\mathrm{OH}), 1740(\delta$-lactone and ester $\mathrm{C}=\mathrm{O}), 1680(\alpha, \beta$-unsaturated $\mathrm{C}=\mathrm{O}), 1645(\mathrm{C}=\mathrm{C}) ;{ }^{1} \mathrm{H}$ NMR see Table 1; EIMS m/z $636\left(\mathrm{M}^{+}, 6.5 \%\right), 437$ ([M side chain $\left.\mathrm{C}_{10} \mathrm{H}_{6} \mathrm{~F}_{3} \mathrm{O}\right]^{+}, 47.9 \%$ ), 199 ([side chain $\left.\left.\mathrm{C}_{10} \mathrm{H}_{6} \mathrm{~F}_{3} \mathrm{O}\right]^{+}, 100 \%\right) ;$ CIMS $m / z 637\left([\mathrm{M}+\mathrm{H}]^{+}, 6.7 \%\right)$, 437 ([M - side chain $\left.\mathrm{C}_{10} \mathrm{H}_{6} \mathrm{~F}_{3} \mathrm{O}\right]^{+}$, $16.4 \%$, 199 ([side chain $\left.\mathrm{C}_{10} \mathrm{H}_{6} \mathrm{~F}_{3} \mathrm{O}\right]^{+}, 100 \%$ ).

3-( $\mathbf{4}^{\prime}, 4^{\prime}, 4^{\prime}$-Trifluoro- $\mathbf{3}^{\prime}$-phenyl- $\mathbf{2}^{\prime}$-butenoyl)-bruceolide (12). Colorless amorphous solid; mp $147-149^{\circ} \mathrm{C}$; $[\alpha]_{\mathrm{D}}{ }^{26}$ $+13.3^{\circ}(c 0.030, \mathrm{EtOH})$; UV (EtOH) nm $243(\varepsilon 3600)$; IR ( KBr ) cm ${ }^{-1} 3400(\mathrm{OH}), 1740$ ( $\delta$-lactone and ester $\mathrm{C}=\mathrm{O}$ ), 1680 ( $\alpha, \beta$-unsaturated $\mathrm{C}=\mathrm{O}$ ), $1645(\mathrm{C}=\mathrm{C}) ;{ }^{1} \mathrm{H}$ NMR see Table 1; EIMS m/z 636 ( $\mathrm{M}^{+}, 1.0 \%$ ), 437 ([M side chain $\left.\left.\mathrm{C}_{10} \mathrm{H}_{6} \mathrm{~F}_{3} \mathrm{O}\right]^{+}, 47.9 \%\right), 199([$ side chain $\left.\left.\mathrm{C}_{10} \mathrm{H}_{6} \mathrm{~F}_{3} \mathrm{O}\right]^{+}, 38.6 \%\right) ;$ CIMS m/z $637\left([\mathrm{M}+\mathrm{H}]^{+}\right.$, $3.2 \%$ ), 199 ([side chain $\left.\mathrm{C}_{10} \mathrm{H}_{6} \mathrm{~F}_{3} \mathrm{O}\right]^{+}, 100 \%$ ).

3-[3'-(Trifluoromethyl)butanoyl]-bruceolide (13). Bruceolide ( $4,30.0 \mathrm{mg}, 0.068 \mathrm{mmol}$ ) was reacted with compound 10 ( $70.4 \mathrm{mg}, 0.403 \mathrm{mg}$ ) in an identical manner to the reaction of 4 and 8 . Pure compound 13 $(9.9 \mathrm{mg}, 0.017 \mathrm{mmol}, 25 \%$ yield) was obtained by preparative HPLC (Lichrosorb RP-18, $\mathrm{H}_{2} \mathrm{O}: \mathrm{MeOH}$ : $\mathrm{AcOH}, 49.5: 49.5: 1, \mathrm{v} / \mathrm{v}$ ). Colorless amorphous solid; mp $168-170^{\circ} \mathrm{C} ;[\alpha]_{\mathrm{D}}{ }^{27}+16.3^{\circ}$ (c 0.049, EtOH); UV (EtOH) nm 243 ( $\varepsilon 9200$ ); IR (KBr) cm ${ }^{-1} 3450(\mathrm{OH}), 1740(\delta-$
lactone and ester $\mathrm{C}=\mathrm{O}$ ), $1680(\alpha, \beta$-unsaturated $\mathrm{C}=\mathrm{O})$; ${ }^{1} \mathrm{H}$ NMR see Table 1; CIMS m/z 577 ([M+H] $\left.{ }^{+}, 1.3 \%\right)$, 139 ([side chain $\left.\mathrm{C}_{5} \mathrm{H}_{6} \mathrm{~F}_{3} \mathrm{O}\right]^{+}, 55.2 \%$ ).

3-(tert-Butyldimethylsilyl)-brusatol (14). A mixture of 3 ( $105 \mathrm{mg}, 0.202 \mathrm{mmol}$ ), tert-butyldimethylsilyl chloride (TBDMS-Cl, $103 \mathrm{mg}, 0.685 \mathrm{mmol}$ ), and imidazole ( 104 $\mathrm{mg}, 1.52 \mathrm{mmol}$ ) was stirred at room temperature for 3.5 $h$. Water ( 1 mL ) was added to the reaction mixture, and colorless crystals ( 119 mg ) were obtained. The crystals were subjected to preparative HPLC (Dynamax-60A, $\left.\mathrm{H}_{2} \mathrm{O}: \mathrm{MeOH}, 1: 4\right)$ to afford pure compound 14. Colorless amorphous solid; mp $230-233{ }^{\circ} \mathrm{C} ;[\alpha]_{\mathrm{D}}{ }^{26}+14.6^{\circ}$ ( $c 0.123, \mathrm{EtOH}) ; \mathrm{UV}(\mathrm{EtOH}) \mathrm{nm} 220(\varepsilon 12100), 270(\varepsilon$ 7300 ); IR ( KBr ) $\mathrm{cm}^{-1} 3450(\mathrm{OH}), 1740$ ( $\delta$-lactone and ester $\mathrm{C}=\mathrm{O}$ ), 1680 ( $\alpha, \beta$-unsaturated $\mathrm{C}=\mathrm{O}$ ), 1645 ( $\mathrm{C}=\mathrm{C}$ ); ${ }^{1} \mathrm{H}$ NMR see Table 1; EIMS $m / z 577$ ([M $\left.\mathrm{C}_{4} \mathrm{H}_{9}\right]^{+}, 100 \%$ ), 83 ([side chain $\left.\mathrm{C}_{5} \mathrm{H}_{7} \mathrm{O}\right]^{+}, 19.7 \%$ ); CIMS $m / z 577\left(\left[\mathrm{M}-\mathrm{C}_{4} \mathrm{H}_{9}\right]^{+}, 100 \%\right), 83$ ([side chain $\left.\mathrm{C}_{5} \mathrm{H}_{7} \mathrm{O}\right]^{+}$, $100 \%$ ). ( $57.0 \mathrm{mg}, 0.090 \mathrm{mmol}, 45 \%$ yield). ${ }^{7}$

3-(tert-Butyldimethylsilyl)-bruceolide (15). Compound 14 ( $54.7 \mathrm{mg}, 0.086 \mathrm{mmol}$ ) was dissolved in dry MeOH ( 1 mL ). Then, $1 \mathrm{~N}-\mathrm{MeOK}$ solution ( $\mathrm{MeOH}, 1 \mathrm{~mL}$ ) was added dropwise at $0^{\circ} \mathrm{C}$, and the mixture was stirred at 0 ${ }^{\circ} \mathrm{C}$ for 6 h . The reaction mixture was treated with cation exchange resin (AMBERLITE IRC-50) and extracted with $\mathrm{CHCl}_{3}$ to afford crude crystals ( 40 mg ). Purification by preparative HPLC (Lichrosorb RP-18, $\mathrm{H}_{2} \mathrm{O}: \mathrm{MeOH}, 3: 7, \mathrm{v} / \mathrm{v}$ ) afforded compound 15 (19.0 $\mathrm{mg}, 0.035 \mathrm{mmol}, 40 \%$ yield). Colorless amorphous solid; mp 238-240 ${ }^{\circ} \mathrm{C} ;[\alpha]_{\mathrm{D}}{ }^{27}-11.1^{\circ}$ (c 0.072, EtOH); UV (EtOH) nm 270 ( $\varepsilon 6400$ ); IR (KBr) $\mathrm{cm}^{-1} 3450$ $(\mathrm{OH}), 1740$ ( $\delta$-lactone and ester $\mathrm{C}=\mathrm{O}$ ), 1670 ( $\alpha, \beta-$ unsaturated $\mathrm{C}=\mathrm{O}), 1610(\mathrm{C}=\mathrm{C}) ;{ }^{1} \mathrm{H}$ NMR see Table 1 ; EIMS $m / z 495\left(\left[\mathrm{M}-\mathrm{C}_{4} \mathrm{H}_{9}\right]^{+}, 100 \%\right.$; CIMS $m / z 495$ ([M$\left.\mathrm{C}_{4} \mathrm{H}_{9}\right]^{+}, 100 \%$ ).

3-(tert-Butyldimethylsilyl)-15-[3'-(trifluoromethyl)-butan-oyl]-bruceolide (16). Compound 15 ( $16.0 \mathrm{mg}, 0.029$ $\mathrm{mmol})$ and compound $10(32.2 \mathrm{mg}, 0.184 \mathrm{mmol})$ were reacted as described for the reaction of 4 and 8 . Pure compound 16 ( $6.8 \mathrm{mg}, 0.0099 \mathrm{mmol}, 34 \%$ yield) was obtained by preparative HPLC. Colorless amorphous solid; mp 174-177 ${ }^{\circ} \mathrm{C} ;[\alpha]_{\mathrm{D}}{ }^{25}+23.5^{\circ}$ (c 0.068, EtOH); UV (EtOH) nm 270 ( $\varepsilon 6400$ ); IR ( KBr ) $\mathrm{cm}^{-1} 3450$ $(\mathrm{OH}), 1740$ ( $\delta$-lactone and ester $\mathrm{C}=\mathrm{O}$ ), $1680(\alpha, \beta-$ unsaturated $\mathrm{C}=\mathrm{O}), 1645(\mathrm{C}=\mathrm{C}) ;{ }^{1} \mathrm{H}-\mathrm{NMR}$ see Table 1; EIMS $m / z 633\left(\left[\mathrm{M}-\mathrm{C}_{4} \mathrm{H}_{9}\right]^{+}, 100 \%\right.$ ), 139 ([side chain $\left.\mathrm{C}_{5} \mathrm{H}_{6} \mathrm{~F}_{3} \mathrm{O}\right]^{+}, 55.2 \%$ ); CIMS m/z 633 ([M - $\left.\mathrm{C}_{4} \mathrm{H}_{9}\right]^{+}$, $100 \%$ ).

15-[(3'-Trifluoromethyl)-butanoyl]-bruceolide (17). Tetrabutyl ammonium fluoride $\left(1.0 \mathrm{M}, \mathrm{Bu}_{4} \mathrm{NF}-\mathrm{THF}\right.$ solution, $40 \mu \mathrm{~L}, 0.04 \mathrm{mmol}$ ) was added to a solution of compound $16(5.7 \mathrm{mg}, 0.008 \mathrm{mmol}$, dry THF $500 \mu \mathrm{~L})$, and the mixture was stirred at room temperature for 15 min . The reaction mixture was poured into ice-water and extracted with $\mathrm{CHCl}_{3}$ to afford crude crystals (8.0 mg ), which were purified by preparative HPLC (Lichrosorb RP-18, $\mathrm{H}_{2} \mathrm{O}: \mathrm{MeOH}, 1: 1, \mathrm{v} / \mathrm{v}$ ) to afford pure compound 17 ( $2.5 \mathrm{mg}, 0.0043 \mathrm{mmol}, 53 \%$ yield).

Colorless amorphous solid; mp $146-148{ }^{\circ} \mathrm{C} ;[\alpha]_{\mathrm{D}}{ }^{28}$ $+4.0^{\circ}(c 0.025, \mathrm{EtOH}) ; \mathrm{UV}(\mathrm{EtOH}) \mathrm{nm} 277$ ( $\varepsilon 6600$ ); IR ( KBr ) $\mathrm{cm}^{-1} 3450(\mathrm{OH}), 1740$ ( $\delta$-lactone and ester $\mathrm{C}=\mathrm{O}$ ), 1680 ( $\alpha, \beta$-unsaturated $\mathrm{C}=\mathrm{O}$ ) $;{ }^{1} \mathrm{H}$ NMR see Table 1; EIMS $m / z 576\left([M]^{+}, 5.7 \%\right), 139$ ([side chain $\left.\mathrm{C}_{5} \mathrm{H}_{6} \mathrm{~F}_{3} \mathrm{O}\right]^{+}, 49.3 \%$ ); High-resolution MS m/z 576.1842 $\left(\mathrm{M}^{+}\right)$(Calcd for $\mathrm{C}_{26} \mathrm{H}_{31} \mathrm{~F}_{3} \mathrm{O}_{11}: 576.1817$ ).

## Cytotoxicity assay

Compounds 11, 12, 13, and 17 were tested against 60 cell lines of eight human cancers including leukemia, non-small cell lung cancer, small cell lung cancer, colon cancer, CNS cancer, melanoma, ovarian cancer, and renal cancer according to literature methods ${ }^{8,9}$ at the National Cancer Institute.

## Acknowledgements

The authors thank Dr S. Ohta, Instrument Center for Chemical Analysis at Hiroshima University, for his measurement of NMR spectra; Drs M. Sugiura, K. Saiki, and T. Sai, Kobe Women's College of Pharmacy, for their measurements of NMR and MS (EI, CI, and High Resolution); Dr D. Lednicer, the National Cancer Institute (USA), for the cytotoxicity assay of compounds 11, 12, 13, and 17; Mr K. Ishikawa, under-
graduate student of Hiroshima University (present: Dai Nippon Printing Ltd), for his technical assistance. This investigation was supported in part by a grant from the National Cancer Institute (CA 17625) awarded to K.-H. Lee.

## References

1. Chien, M. M. and Rosazza, J. P. J. Chem. Soc. Perkin Trans. 1981, 1, 1352.
2. Lee, K. H.; Imakura, Y.; Wu, R. Y.; and Hall, I. H. J. Org. Chem. 1979, 44, 2180.
3. Okano, M.; Lee, K. H. J. Org. Chem. 1981, 46, 1138.
4. Lee, K. H.; Tani, S.; Imakura, Y. J. Nat. Prod. U.S.A. 1987, 50, 847.
5. Boyd, M. R. Principles and Practices of Oncology 1989, 3, 1.
6. Boyd, M. R. In Current Therapy in Oncology; Neiderhuber, J. E., Ed.; Decker: Philadephia, 1992.
7. We also obtained a silyl derivative of a tautomer (A-ring) of compound 3 in a $30 \%$ yield.
8. Boyd, M. R. In Cancer: Principles and Practice of Oncology Updates; DeVita, V. T.; Hellman, S.; Rosenberg, S. A., Eds; Lippincott: Philadelphia, 1989; pp 1-12.
9. Monks, A.; Scudiero, D.; Skehan, P.; Shoemaker, R.; Paull, K.; Vistica, D.; Hose, C.; Langley, J.; Cronise, P.; VaigroWolff, A.; Gray-Goodrich, M.; Campbell, H.; Mayo, J.; Boyd, M. J. Natl. Cancer Inst. 1991, 83, 757.
(Received in U.S.A. 15 October 1996; accepted 6 March 1997)

[^0]:    ${ }^{\text {a }}$ Measured at 500 MHz .
    ${ }^{\mathrm{b}}$ Measured at 200 MHz .
    ${ }^{c}$ Measured at 270 MHz .
    ${ }^{\mathrm{d}} \mathrm{NA}=$ Not assignable.

