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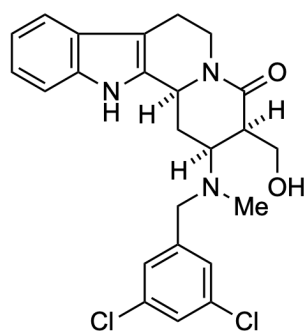
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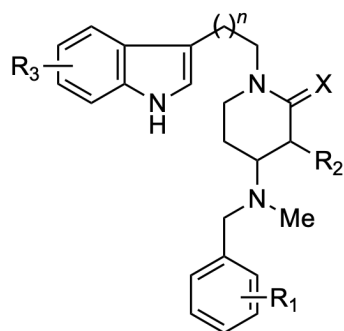
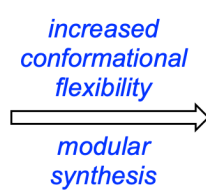
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T. brucei GI₅₀ = 25 μ M



T. brucei GI₅₀ 0.95 – 10 μ M

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Design, Synthesis, and Evaluation of Novel Anti-Trypanosomal Compounds

Lance T. Lepovitz[†], Alan R. Meis[‡], Sarah M. Thomas[§], Justin Wiedeman[§], Alexandra Pham[‡],
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This paper is dedicated to Professor Richard J. K. Taylor in recognition of his many successful years as an editor of *Tetrahedron*.

Supporting Information Placeholder

ABSTRACT: Human African trypanosomiasis (HAT) is a deadly neglected tropical disease caused by the protozoan parasite *Trypanosoma brucei*. During the course of screening a collection of diverse nitrogenous heterocycles, we discovered two novel compounds that contain the tetracyclic core of the *Yohimbine* and *Corynanthe* alkaloids, were potent inhibitors of *T. brucei* proliferation and *T. brucei* methionyl-tRNA synthetase (*TbMetRS*) activity. Inspired by these key findings, we prepared several novel series of hydroxyalkyl δ -lactam, δ -lactam, and piperidine analogs and tested their anti-trypanosomal activity. A number of inhibitors are more potent against *T. brucei* than these initial hits with one hydroxyalkyl δ -lactam derivative being 25-fold more effective in our assay. Surprisingly, most of these active compounds failed to inhibit *TbMetRS*. This work underscores the importance of verifying, irrespective of close structural similarities, that new compounds designed from a lead with a known biological target engage the putative binding site.

small molecular libraries derived from more than 40 different nitrogenous heterocycles were prepared using this approach, and compounds were submitted for screening as part of the NIH Molecular Libraries Initiative.²⁻⁷

One focused library contained the tetracyclic heterocyclic framework that is a structural subunit of the *Yohimbine* and *Corynanthe* alkaloids,⁷ and the representative compounds **1** and **2** (Figure 1) were found to inhibit the proliferation of the protozoan parasite *Trypanosoma brucei* (*T. brucei*), the infectious agent that causes human African trypanosomiasis (HAT). These compounds also inhibit *T. brucei* methionyl-tRNA synthetase (*TbMetRS*),⁸ which plays an essential role in protein synthesis. HAT is a deadly, yet sadly neglected, tropical disease with an at-risk population of more than 60 million people in sub-Saharan Africa.⁹ The progression of the disease is divided into two stages – haemolympathic and encephalitic. During the first stage, the parasite resides in the bloodstream and lymph of the host and causes a variety of flu-like symptoms and organ abnormalities. Progression to the second stage occurs when the parasite crosses the blood-brain barrier, leading to a variety of central nervous system disturbances such as dementia, sleep-wake cycle alteration, motor/visual system impairment, meningoencephalitis, and ultimately death. Unfortunately, treatment options for HAT are limited. Suramin and pentamidine are used to treat the first stage, whereas melarsoprol, eflornithine, and eflornithine/nifurtimox combination therapy are treatments for the second stage.^{9,10} However, each of these treatments suffer serious shortcomings, including toxicity and drug resistance. Recently fexinidazole, which is the first drug

Introduction

As part of a broad program directed toward the discovery of novel compounds that would serve as tools for chemical biology or as therapeutic leads, we recently developed a general platform for the rapid assembly of diverse molecular libraries of natural product-like compounds.¹ The approach features multi-component assembly processes (MCAPs) to create versatile intermediates that can then be quickly transformed by selective ring-forming reactions into various heterocyclic scaffolds bearing functional groups suitable for further diversification. A variety of

candidate for treating late-stage HAT in the past 30 years, was approved for use in non-European markets.¹¹

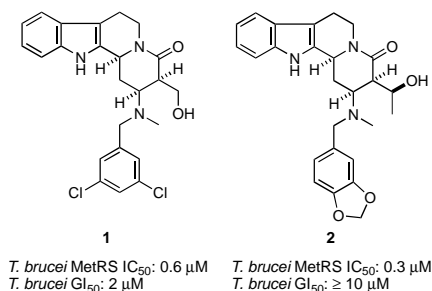


Figure 1. Initial screening hits identified for *T. brucei* and *TbMetRS*.⁸

In view of the deficiencies associated with drugs currently used to treat HAT, the unmet need for new chemotherapeutic agents with fewer off-target effects is apparent. Accordingly, the promising preliminary findings that **1** and **2** both decrease proliferation of *T. brucei* and inhibit *TbMetRS* inspired us to initiate a campaign to design and synthesize simplified analogs with improved physicochemical and biological properties. Owing to the substantial differences in active site residues of the parasite MetRS enzyme compared to its human ortholog,¹² we envisioned that such compounds for treating HAT would have minimal adverse side effects. Indeed, this approach was pioneered by Buckner and colleagues who developed and studied libraries of aminoquinoline and urea-based *TbMetRS* inhibitors.^{12,13} The lead compound **3** (Figure 2) that emerged from these studies significantly suppressed proliferation of *T. brucei rhodesiense* in mice, but the treatment was not curative because the mice succumbed to parasitemia.^{13a} This finding led to studies of other analogs.¹⁴

Comparing the structure of **3** with our initial hits **1** and **2** reveals some notable similarities and differences that provided a rationale for the design of analogs based on **1** and **2**. For example, **3** and related compounds possess heteroaromatic and *N*-benzyl substituents that are separated by a flexible linker. Compounds **1** and **2** likewise feature heteroaromatic and *N*-benzyl substituents, but these subunits are connected by a rigid quinolizidine core. In the crystal structure of **3** bound to *TbMetRS* (PDB: 4MVW),^{13b} the two aromatic substituents occupy separate pockets on the extended binding site of the enzyme. Specifically, the *N*-benzyl group occupies the methionine-binding pocket, and the heteroaromatic moiety resides in an auxiliary pocket; the flexible linker appears to allow these substituents to occupy their respective binding sites in favorable orientations. Based upon these observations and some preliminary modeling studies, we hypothesized that analogs of our initial hits **1** and **2** having the increased conformational flexibility of **3** would be superior inhibitors because the added flexibility allows the *N*-benzyl and indole moieties to interact more optimally with *TbMetRS* than is possible in the more rigid tetracyclic framework present in **1** and **2**.

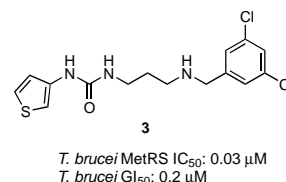


Figure 2. Flexible, urea-based inhibitor **3** of *TbMetRS*.¹³

To test this hypothesis, we envisioned scission of the C(2)–C(3) bond (Figure 3) of the core structure **4** leading to the set of compounds having the general structures **5–7**. In addition to increasing the conformational flexibility of the core framework using different linking subunits, novel inhibitors belonging to these new chemotypes may be accessed by a modular synthesis that enables facile late-stage diversification of the indole and the *N*-benzyl substituents. We also planned to examine the effects simplifying the core of **5** in an iterative fashion by removing the hydroxymethylene side-chain and reducing the lactam to give the three distinct series of hydroxyalkyl δ-lactam, lactam, and piperidine analogs as generally defined by **5–7**, respectively.

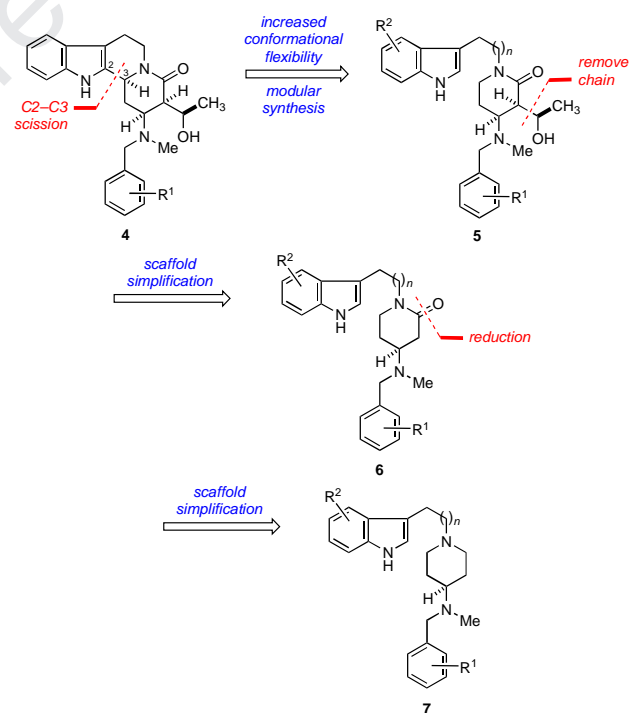
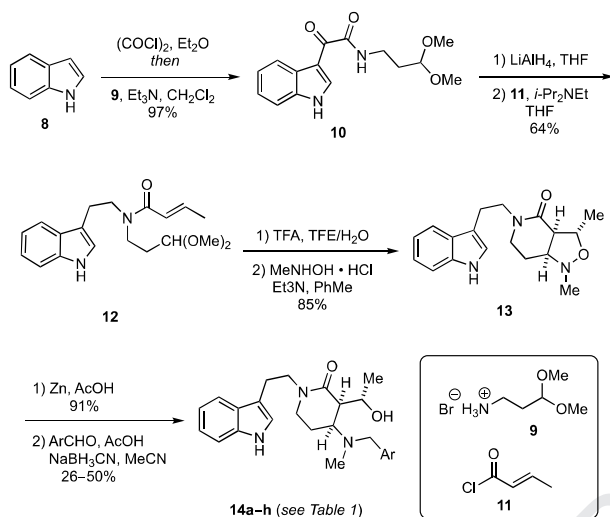


Figure 3. Design of more flexible scaffolds **5–7** from parent **4**.

Results and Discussion

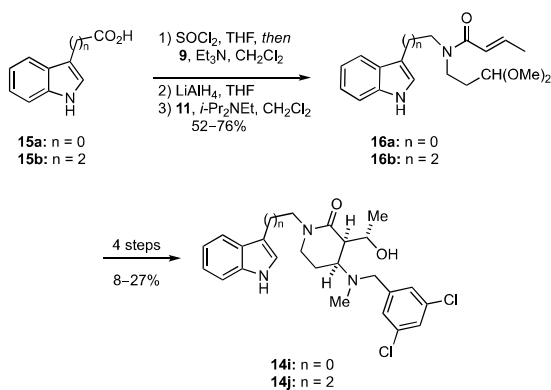
Synthesis. Synthesis of hydroxyalkyl δ-lactam analogs having the general form **5** was accomplished by first converting indole (**8**) into the amide **10** in a one-pot process involving sequential reaction of **8** with oxalyl chloride and the amine **9** (Scheme 1). Reduction of **10** furnished **12** in 62% overall yield from **8**. Acid-catalyzed removal of the acetal protecting group in **10** using CF₃CO₂H (TFA) in

aqueous trifluoroethanol (TFE) produced an unstable aldehyde that was treated directly with *N*-methylhydroxylamine, and the intermediate nitrene underwent an intramolecular dipolar cycloaddition upon heating to furnish **13** in 85% yield. Reductive opening of the isoxazolidine ring and subsequent reductive alkylation of the resultant secondary amine with a series of substituted benzaldehydes delivered the hydroxyalkyl δ -lactam analogs **14a-h** in overall yields ranging from 24–46%.



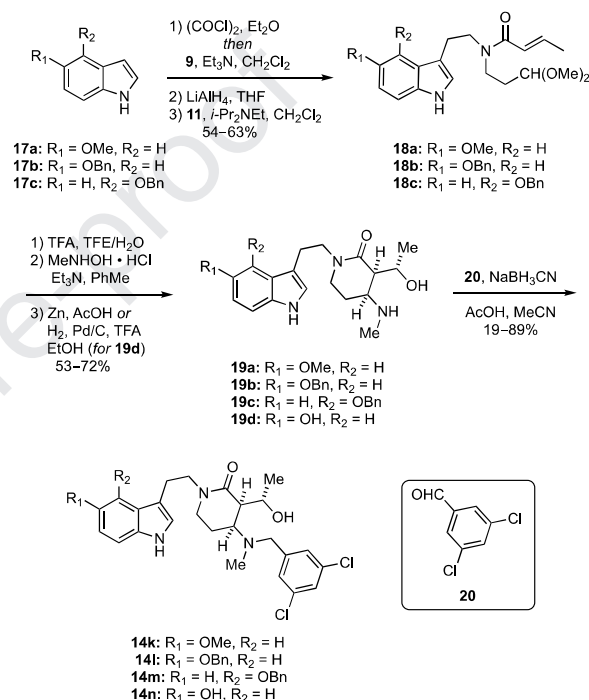
Scheme 1. Synthesis of hydroxyalkyl δ -lactam analogs **14a-h**.

In addition to probing the effects of varying the substitution on the *N*-benzyl group, we prepared one series of hydroxyalkyl δ -lactams in which the length of the chain linking the indole and lactam rings was varied (Scheme 2) and another series in which the nature of the substituents on the indole ring was changed (Scheme 3). Toward preparing analogs with variable linker lengths, the indole carboxylic acids **15a,b** were sequentially treated with thionyl chloride and then **9**, and the intermediate amides were reduced with LiAlH_4 to give amines that were acylated with **11** to furnish amides **16a,b** in 52–76% overall yields (Scheme 2). Elaboration of **16a,b** into **14i,j** was then achieved in four steps (8–27%, Scheme 2) following the same steps previously used to convert **12** into **14a-h** (Scheme 1).



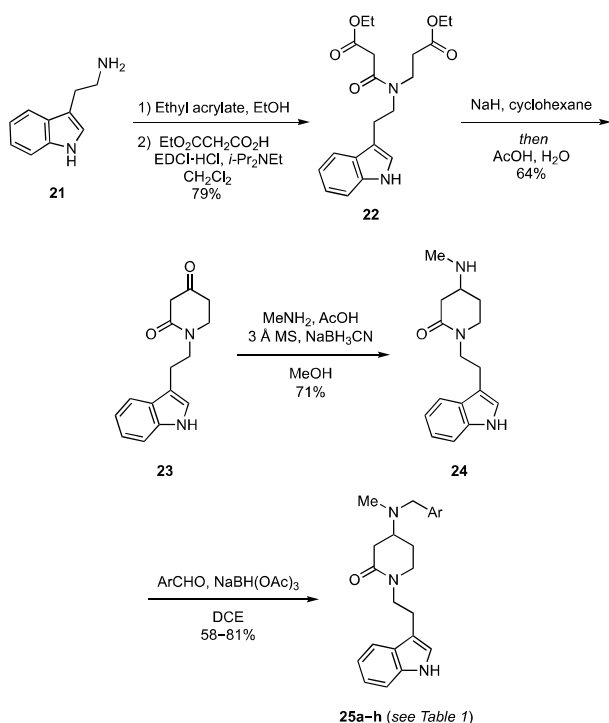
Scheme 2. Synthesis of hydroxyalkyl δ -lactam analogs **14i,j** with varying linker lengths.

Analogues bearing different oxygen substituents at the 4- and 5-positions of the indole ring were likewise prepared by the three-step transformation of indoles **17a-c** into the crotonamides **18a-c** in 54–63% overall yields following the procedures previously outlined in Scheme 1 to convert indole (**8**) into **12** (Scheme 3). These amides were then elaborated to their respective analogs **14k-m** in four steps (10–64% overall yield) as described for the syntheses of **14a-h** (Scheme 1). The 5-hydroxy analog **14n** was also prepared from **18b** in four steps (16% overall yield) involving nitrene formation and cycloaddition followed by hydrogenolysis to simultaneously remove the *O*-benzyl protecting group and cleave the isoxazolidine ring. The 5-hydroxy intermediate **19d** thus formed was then converted to **14n** via reductive *N*-alkylation.

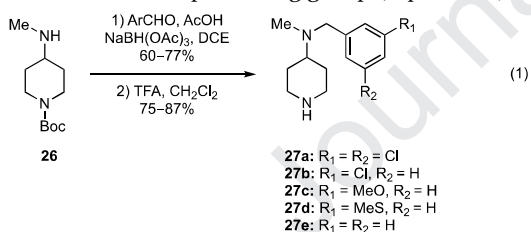


Scheme 3. Synthesis of hydroxyalkyl δ -lactam analogs **14k-n** with indolyl substituents.

The 4-amino- δ -lactams **25a-h** were then prepared from tryptamine (**21**) via the sequence of reactions summarized in Scheme 4. In the event, reaction of **21** with one equivalent of ethyl acrylate provided a secondary amine that was coupled with malonic acid monoethyl ester using *N*-ethyl-*N*-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDCI) to furnish **22** in 79% overall yield. Dieckmann cyclization of **22** followed by hydrolysis and decarboxylation provided the β -keto-lactam **23** in 64% yield. Reductive amination of **23** with methylamine then delivered the amine **24** (71% yield), which was reductively alkylated with a set of substituted benzaldehydes to deliver the **25a-h** in 58–81% yield.

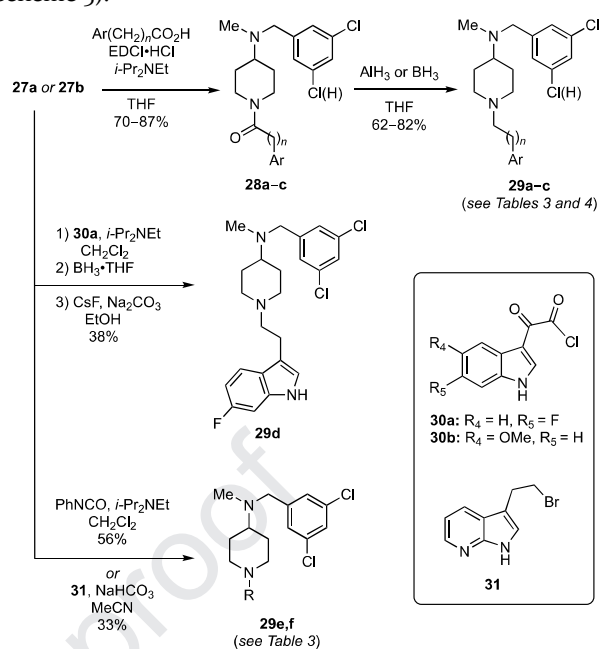
Scheme 4. Synthesis of δ -lactam analogs **25a-h**.

Toward preparing the 4-aminopiperidines **29a-l**, the key intermediates **27a-e** were first synthesized from the Boc-protected piperidine **26**, itself prepared from *N*-Boc-4-piperidinone (see SI) by sequential reductive alkylation with various substituted benzaldehydes and acid-catalyzed removal of the Boc-protecting group (Equation 1).

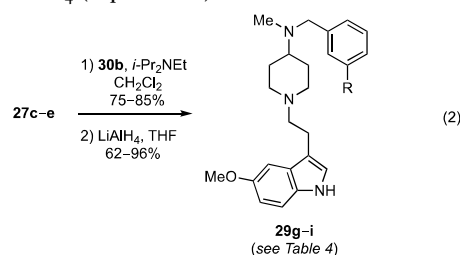


Compounds **27a-e** were then elaborated via divergent synthetic sequences to deliver the desired analogs. For example, coupling of **27a** or **27b** with various heteroaromatic alkyl carboxylic acids using EDCI gave the *N*-acyl piperidines **28a-c** (70–87% yields) that were selectively reduced with either alane or borane to give **29a-c** in 62–82% overall yields (Scheme 5). Use of a Lewis-acidic hydride reducing agent was crucial to avoid reduction of the aryl chloride substituents, as was observed when performing the reduction with lithium aluminum hydride (LiAlH_4). The related fluoro-indole analog **29d** was prepared from **27a** in a similar fashion except the acid chloride **30a** was used as an acylating agent, and $\text{CsF}/\text{Na}_2\text{CO}_3$ was used in an additional step after borane-mediated ketoamide reduction to decompose the amine-borane adducts. This step was required rather than the alternative aqueous acid workup because the addition of aqueous HCl following the ketoamide reduction led to reduction of the indole ring.¹⁵ The phenylurea and 7-azaindole derivatives **29e** and **29f** were prepared

by treating **27a** with phenyl isocyanate and **31**, respectively (Scheme 5).¹⁶

Scheme 5. Synthesis of piperidine analogs **29a-f**.

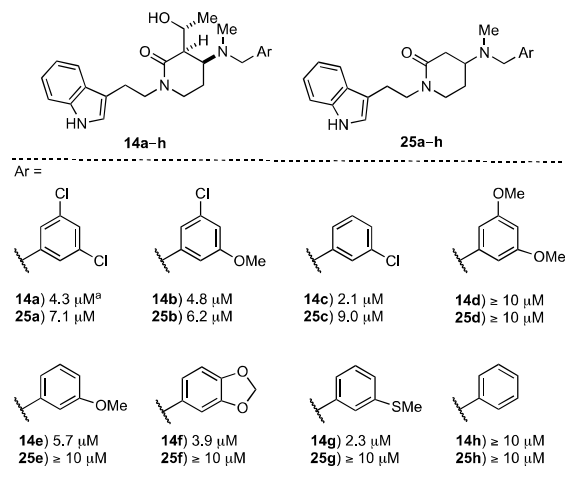
Finally, analogs **29g-i** were prepared in 62–96% overall yields from **27c-e** via acylation with **30b** followed by reduction with LiAlH_4 (Equation 2).



Biological Analysis. Having prepared a suite of different analogs of our initial leads **1** and **2**, we turned to the evaluation of their efficacy using a phenotypic screen to determine their effects on the growth of *T. brucei* *in vitro*. In order to ensure meaningful comparisons of structure activity relationships (SAR) relative to the more potent lead **1** (Figure 1), we assayed **1** using the same growth inhibition assay that we would employ to evaluate analogs of **1** and found that it displayed a GI_{50} value of 25 μM . Owing perhaps to variations in experimental conditions, this value is substantially different from the 2 μM value originally determined (see Figure 1).⁸ Analysis of the GI_{50} values of the hydroxyalkyl δ -lactams **14a-h** and the δ -lactams **25a-h** reveals how increasing the flexibility of the central core, modifying the substitution on the δ -lactam ring, and varying the nature of the *N*-benzyl group affects potencies (Table 1). Indeed, both **14a** and **25a** are roughly five-fold more active against *T. brucei* than **1**, thus supporting our central hypothesis that increased flexibility would improve activity. As evidenced by the observation that **25a-h** are less active than their counterparts **14a-h**, removing the hydroxyethyl side chain on the δ -lactam ring consistently reduces the ability of compounds to inhibit growth. The SAR asso-

ciated with the *N*-benzyl substituent on the various scaffolds roughly tracks trends observed by Buckner and co-workers. For example, the dimethoxy analogs **14d** and **25d** and the unsubstituted benzyl analogs **14h** and **25h** do not exhibit any activity,^{12,13} whereas compounds having 3-chlorobenzyl groups appear to outperform other substituted analogs.

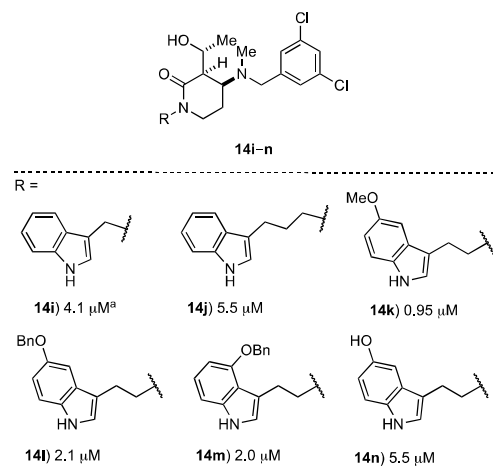
Table 1. *T. brucei* GI₅₀ values hydroxyalkyl δ -lactams **14a–h** and δ -lactams **25a–h**.



^a error values were consistently <10% with exceptions of **14f** (18%) and **14g** (26%)

The GI₅₀ values for the set of hydroxyalkyl δ -lactam analogs **14i–n** enable us to assess the effects of varying the length of the chain linking the indole ring and the δ -lactam core and of modifying the nature of substituents on the indole ring (Table 2). With regard to the linker length, compounds having one to three carbon atoms in the chain (e.g., **14a**, **14i**, and **14j**, respectively) are approximately equipotent. Based upon our original design rationale, this result is somewhat surprising. Specifically, we hypothesized that the length of the linking unit would modulate potency because the increased flexibility that accompanies longer chains could affect how the two aromatic moieties on the δ -lactam ring interact with their respective binding pockets. The presence of alkoxy groups on the indole ring as found in **14k–m** offers a 2–5-fold increase in growth inhibitory activity relative to the unsubstituted parent **14a**, whereas the 5'-hydroxy analog **14n** is equipotent with **14a**.

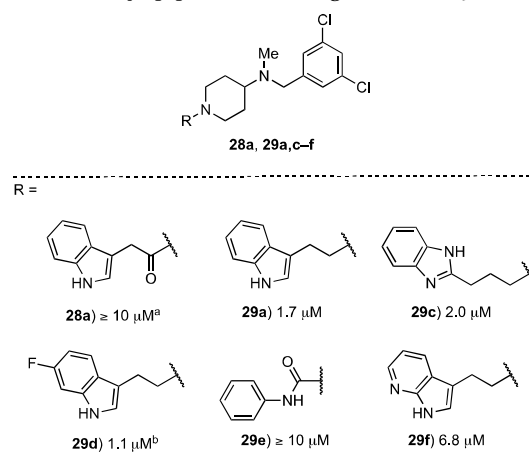
Table 2. *T. brucei* GI₅₀ values of hydroxyalkyl δ -lactams **14i–n**.



^a error values were consistently <10% with exceptions of **14i** (15%), **14l** (19%), and **14n** (11%)

The GI₅₀ values for the *N*-(3,5-dichlorobenzyl) piperidines **28a** and **29a,c–f** (Table 3) provide additional SAR data for analogs of **1**. For example, **28a**, which has an amide linker between the piperidine ring and the indole groups, displays significantly diminished activity relative to its alkyl counterpart **29a**. We surmise that this difference likely arises from a conformational effect, because the potencies of the lactams **25a–h** (Table 1) show that a basic nitrogen atom in the piperidine ring is not essential for activity. Having a fluorine substituent on the indole ring as in **29d** or replacing the indole moiety with a benzimidazole ring as in **29c** has no notable effect on potency, whereas replacing the indole with the 7-azaindole ring in **29f** leads to a four-fold loss in growth inhibitory activity. Finally, the phenyl-urea analog **29e**, which we prepared for a direct comparison of our piperidine scaffold with Buckner's phenyl-urea diaminopropane inhibitors, is inactive, perhaps because of conformational effects.

Table 3. *T. brucei* GI₅₀ values of *N*-(3,5-dichlorobenzyl)piperidine analogs **28a** and **29a,c–f**.



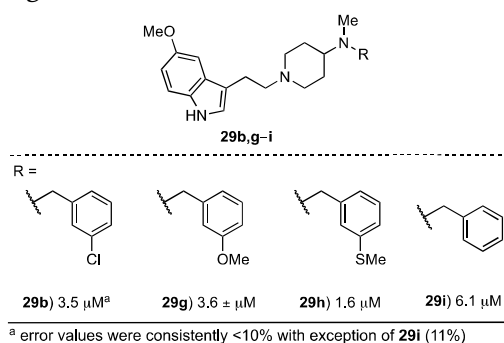
^a error values were consistently <10% with exception of **29c** (30%).

^b error not determined.

The set of compounds **29b,g–i** was designed to explore the SAR of a set of analogs of **1** having a piperidine core in the spacer linking the 5-methoxyindole, which appears to be an optimal indole substituent (see **14k**, Table 2), and the *N*-benzyl groups (Table 4). Of these compounds, the thi-

omethyl compound **29h** ($GI_{50} = 1.6 \mu\text{M}$) is the most potent, while the potencies of the other *N*-benzyl analogs **29b,g,i** are 2–4-fold less active.

Table 4. *T. brucei* GI_{50} values of piperidine analogs **29b,g-i**.



Examination of the growth inhibition data for our analogs of **1** (see Tables 1–4) reveals a relatively flat SAR in which a fairly large number of structural modifications seem to have little impact on the observed GI_{50} values. We initially focused upon GI_{50} values because we lacked the capability of performing the established radiolabeled methionine assay for *TbMetRS* activity that was employed in the initial screening.¹³ However, we also wanted to show that the new analogs of **1** inhibited *TbMetRS*, so we assessed the inhibitory activity of our compounds towards recombinant *TbMetRS* using a colorimetric pyrophosphatase-coupled enzyme inhibition assay.¹⁷ We first tested the original hit **1** and found that it exhibited an IC_{50} value of $4.9 \pm 1.5 \mu\text{M}$ in this assay. The discrepancy between this IC_{50} value and the $0.6 \mu\text{M}$ value (Figure 1) is expected, because the concentration of *TbMetRS* and other reagents in the colorimetric assay is much higher than those used in the radiolabeled methionine assay. Indeed, variations in IC_{50} values arising from concentration differences are common, so it is important to ensure concentrations are the same before making any correlations of IC_{50} values. Surprisingly, however, although a few compounds having the hydroxyalkyl δ -lactam scaffold found in **1** (i.e., **14a,b,h**, and **n**) display 15–30 % inhibition of *TbMetRS* at $200 \mu\text{M}$, most analogs are inactive against *TbMetRS* at concentrations up to $200 \mu\text{M}$. Even for those compounds that inhibit *TbMetRS*, growth inhibition values and enzyme inhibition data correlate poorly, suggesting that the mechanism of action for the growth inhibition of more flexible analogs of **1** likely differs from that of **1**.

Summary and Conclusion.

In summary, following the discovery of **1** and **2** as novel leads against HAT, we designed several sets of analogs as molecular hybrids of **1** and **3**, a more flexible inhibitor of *TbMetRS*, that we hypothesized would bind with higher affinity to *TbMetRS*. The rationale for these experiments was based upon preliminary modeling studies suggesting that the terminal aromatic rings, which comprise the pharmacophore of *TbMetRS* inhibitors, in more flexible analogs of **1** would interact better with the enzyme. Small collections of hydroxyalkyl δ -lactams, δ -lactams and piperidines were thus prepared and evaluated in growth inhibi-

tion assays of *T. brucei*. Although a number of compounds were discovered that are up to 25-fold more potent than **1** in our assay, there is little variation overall in the SAR. Moreover, each of the new compounds is less active against *TbMetRS* than **1**. Accordingly, the majority of analogs in this study appear to inhibit proliferation of *T. brucei* by an alternate, as yet unknown, mechanism that does not involve inhibition of *TbMetRS*. This work underscores the importance of ensuring that new compounds that are designed from an initial hit engage the same binding site; nothing should be assumed, irrespective of close structural similarities.

EXPERIMENTAL SECTION

General. Solvents and reagents were reagent grade and were used without purification unless otherwise noted. Methanol (MeOH) and *N,N*-dimethylformamide (DMF) were dried by filtration through two columns of activated molecular sieves prior to use. Tetrahydrofuran (THF) was passed through two columns of activated neutral alumina prior to use. Benzene was distilled from sodium and benzophenone. Methylene chloride, diisopropylamine, triethylamine, and diisopropylethylamine were distilled from calcium hydride immediately prior to use. Pyridine was distilled from potassium hydroxide (KOH) and calcium hydride and stored over KOH. Dioxane was distilled from sodium metal and benzophenone prior to use. All solvents were determined to have less than 50 ppm H_2O by Karl Fischer coulometric moisture analysis. All reactions were performed under an atmosphere of argon or nitrogen. Removal of solvent under reduced pressure was performed using a rotary evaporator. Flash chromatography was performed using SiliaFlash[®] F60 silica gel (Silicycle, 40–63 μM , 60 Å). Infrared (IR) spectra were obtained either neat or as solutions in the solvent indicated and reported as wavenumbers (cm^{-1}). ^1H and ^{13}C NMR spectra were obtained at the specified frequency as a solution in CDCl_3 unless otherwise indicated. Chemical shifts are reported in parts per million (ppm) downfield from tetramethylsilane (TMS) (δ 0.00) and referenced relative to deuterated solvent. Coupling constants (*J*) are reported in Hz, and the splitting abbreviations used are: s, singlet; d, doublet; t, triplet; q, quartet; dd, doublet of doublets; ddd, doublet of doublets of doublets; m, multiplet; comp, overlapping multiplets of magnetically non-equivalent protons; br, broad. For HRMS with compounds containing chlorine, the calculated and measured masses are for the isotope ^{35}Cl .

***N*-(3,3-Dimethoxypropyl)-2-(1*H*-indol-3-yl)-2-oxoacetamide (10).** A solution of oxalyl chloride (1.52 g, 12.0 mmol) in ether (1 mL) was added dropwise over 5 min to a solution of indole (1.41 g, 12.0 mmol) in ether (24 mL) at 0°C . The reaction was stirred at 0°C for 1 h, then warmed to room temperature, and stirred for an additional 1 h. The yellow precipitate was collected via vacuum filtration and dried *in vacuo* to give 2.28 g (87%) of indole-3-glyoxal chloride. A suspension of indole-3-glyoxal chloride (1.91 g, 8.7 mmol) in CH_2Cl_2 (20 mL) was added dropwise over 30 min to a solution of **9** (1.47 g, 7.3 mmol) and triethylamine (3.70 g, 36.5 mmol) in CH_2Cl_2 (75 mL) at 0°C .

The reaction was stirred at 0 °C for 0.5 h, then warmed to room temperature and stirred for an additional 1 h. Saturated aqueous NaHCO₃ (100 mL) was added to the reaction and stirred for 15 min. The mixture was extracted with CH₂Cl₂ (3 x 100 mL), and the combined organic extracts were washed with aqueous NaOH (1 M, 2 x 100 mL), water (100 mL), and brine (100 mL), dried (Na₂SO₄), filtered, and concentrated *in vacuo* to give 2.23 g (97%) of crude **10**. The crude material was >90% purity, and was further purified by dissolving in CH₂Cl₂ (300 mL) and washing successively with saturated aqueous NH₄Cl (200 mL), saturated aqueous NaHCO₃ (200 mL), water (200 mL), and brine (200 mL). The organic fraction was dried (Na₂SO₄), and concentrated *in vacuo* to give 1.91 g (90%) of **10** as a pale yellow solid (>95% purity, ¹H NMR). ¹H NMR (400 MHz) δ 9.99 (s, 1 H), 9.01 (d, *J* = 3.2 Hz, 1 H), 8.41 (d, *J* = 7.2 Hz, 1 H), 7.97 (t, *J* = 6.0 Hz, 1 H), 7.40 – 7.38 (m, 1 H), 7.31 (ddd, *J* = 8.4, 7.2, 1.2 Hz, 1 H), 7.25 (ddd, *J* = 8.4, 7.2, 1.2 Hz, 1 H), 4.48 (t, *J* = 6.4 Hz, 1 H), 3.47 (q, *J* = 6.4 Hz, 2 H), 3.36 (s, 6 H), 1.91 (q, *J* = 6.4 Hz, 2 H). ¹³C NMR (100 MHz, CD₃CN) δ 181.6, 163.0, 139.1, 136.5, 126.9, 124.0, 123.1, 121.9, 112.8, 112.5, 103.7, 53.1, 35.1, 32.3. HRMS (ESI) *m/z* calcd for C₁₅H₁₈N₂O₄ (M+Na)⁺, 313.1159; found, 313.1166.

(E)-N-(2-(1*H*-Indol-3-yl)ethyl)-N-(3,3-dimethoxypropyl)but-2-enamide (12). A solution of **10** (0.50 g, 1.70 mmol) in THF (5 mL) was added dropwise over 15 min to a stirred suspension of lithium aluminum hydride (0.65 g, 17.0 mmol) in THF (40 mL) at 0 °C. The reaction was heated at 65 °C for 14 h. The reaction was cooled to 0 °C, and the reaction was quenched via successive addition of water (0.6 mL), aqueous sodium hydroxide (15% w/v, 0.6 mL), and water (3 mL). The suspension was warmed to room temperature and MgSO₄ was added. The solution was filtered through a fritted funnel, the solids were washed with excess CH₂Cl₂ and the filtrate was concentrated *in vacuo*. The crude material was taken up in aqueous HCl (0.2 M, 150 mL) and washed with ether (2 x 100 mL). The aqueous fraction was basified with aqueous NaOH (40% w/v) to pH 12-14, as judged by pH paper. The basic solution was extracted with CH₂Cl₂ (3 x 200 mL). The combined organic extracts were dried (Na₂SO₄), filtered, and concentrated to give 0.41 g (90%) of *N*-(2-(1*H*-indol-3-yl)ethyl)-3,3-dimethoxypropan-1-amine as a viscous oil (>95% purity, ¹H NMR); ¹H NMR (400 MHz) δ 8.61 (s, 1 H), 7.57 (d, *J* = 7.2 Hz, 1 H), 7.42 (dt, *J* = 8.0, 1.2 Hz, 1 H), 7.27 – 7.20 (m, 1 H), 7.17 (dd, *J* = 8.0, 1.2 Hz, 1 H), 7.14 (ddd, *J* = 8.0, 7.2, 1.2 Hz, 1 H), 4.25 (t, *J* = 4.0 Hz, 1 H), 3.20 – 3.18 (comp, 4 H), 3.03 (t, *J* = 5.6 Hz, 2 H), 3.00 (s, 6 H), 1.88 (tt, *J* = 7.2, 4.0 Hz, 2 H). ¹³C NMR (100 MHz) δ 134.0, 123.8, 121.5, 120.0, 117.4, 115.6, 109.2, 105.8, 101.3, 52.3, 45.6, 41.1, 25.8, 19.6; IR (neat) 3411, 2337, 2173, 1640, 1458 cm⁻¹. HRMS (CI) *m/z* calcd for C₁₅H₂₃N₂O₂ (M+H)⁺, 263.1760; found, 263.1760.

A solution of crotonoyl chloride (51 mg, 0.48 mmol) in CH₂Cl₂ (1 mL) was added dropwise to a solution of triethylamine (82 mg, 0.81 mmol) and *N*-(2-(1*H*-indol-3-yl)ethyl)-3,3-dimethoxypropan-1-amine (106 mg, 0.40 mmol) in CH₂Cl₂ (1 mL) at -78 °C. The reaction was stirred for 2 h at -78 °C, whereupon saturated aqueous NaHCO₃ (5 mL) was added. The phases were separated, and the aqueous phase

was extracted with CH₂Cl₂ (3 x 10 mL). The combined organic fractions were dried (Na₂SO₄), filtered, and the solvent removed *in vacuo*. The resulting residue was purified via flash column chromatography eluting with CH₂Cl₂ : methanol (99 : 1 → 97 : 1 along a gradient) to give 94 mg (71%) of **12** as a clear oil. ¹H NMR (400 MHz, CD₃CN) (1 : 1 rotamer mixture) δ 9.24 (brs, 0.5 H), 9.20 (brs, 0.5 H), 7.65 (d, *J* = 8.0 Hz, 0.5 H), 7.58 (d, *J* = 8.0 Hz, 0.5 H), 7.42 (d, *J* = 3.6 Hz, 0.5 H), 7.40 (d, *J* = 3.6 Hz, 0.5 H), 7.17 – 7.14 (comp, 1 H), 7.10 – 7.06 (comp, 2 H), 6.87 (dq, *J* = 14.8, 7.2 Hz, 0.5 H), 6.64 (dq, *J* = 14.8, 7.2 Hz, 0.5 H), 6.40 (dd, *J* = 14.8, 1.6 Hz, 0.5 H), 6.16 (dd, *J* = 14.8, 1.6 Hz, 0.5 H), 4.38 – 4.34 (comp, 1 H), 3.63 (t, *J* = 6.4 Hz, 1 H), 3.61 (t, *J* = 6.4 Hz, 1 H), 3.39 – 3.35 (comp, 2 H), 3.27 (s, 6 H), 3.00 (t, *J* = 8.0 Hz, 1 H), 2.98 (t, *J* = 8.0 Hz, 1 H), 1.99 (dd, *J* = 6.8, 1.6 Hz, 1.5 H), 1.83 – 1.79 (comp, 2 H), 1.69 (dd, *J* = 6.8, 1.6 Hz, 1.5 H). ¹³C NMR (100 MHz, CD₃CN) δ 136.8, 127.8, 127.7, 123.4, 122.7, 122.6, 122.6, 121.8, 121.7, 119.1, 119.0, 118.9, 118.5, 112.9, 111.9, 111.7, 111.6, 103.1, 102.6, 52.8, 52.5, 48.8, 47.5, 44.0, 42.5, 32.7, 31.1, 25.3, 23.7, 17.5, 17.3. HRMS (ESI) *m/z* calcd for C₁₉H₂₆N₂O₃ (M+Na)⁺, 353.1836; found, 353.1836.

(3*S*,3*R*,7*aS*)-5-(2-(1*H*-Indol-3-yl)ethyl)-1,3-dimethylhexahydroisoxazolo[4,3-*c*]pyridin-4(1*H*)-one (13). A solution of trifluoroacetic acid (5 drops) and **12** (10 mg, 0.33 mmol) in TFE/water (3 : 1, 7 mL) was stirred for 1 h at room temperature. Saturated aqueous NaHCO₃ (20 mL) was added, and the aqueous layer extracted with CH₂Cl₂ (3 x 20 mL). The combined organic fractions were dried (Na₂SO₄), filtered, and concentrated *in vacuo*. The crude mixture was dissolved in toluene (5 mL), followed by addition of *N*-methylhydroxylamine hydrochloride (41 mg, 0.50 mmol) and triethylamine (83 mg, 0.83 mmol). The reaction was heated under reflux for 1 h, then cooled to room temperature, and partitioned between saturated aqueous NaHCO₃ (20 mL), ethyl acetate (20 mL), and methanol (2 mL). The layers were separated, and the aqueous layer was extracted with ethyl acetate (3 x 20 mL). The combined organic fractions were washed with water (30 mL), brine (30 mL), dried (Na₂SO₄), filtered, and concentrated *in vacuo* to give 88 mg (85%) of **13** as an oil (>90% purity, ¹H NMR). ¹H NMR (500 MHz) δ 8.26 (brs, 1 H), 6.62 (d, *J* = 8.1 Hz, 1 H), 7.32 (d, *J* = 8.1 Hz, 1 H), 7.16 (t, *J* = 7.1 Hz, 1 H), 7.09 (t, *J* = 7.1 Hz, 1 H), 6.99 (d, *J* = 1.7 Hz, 1 H), 3.89 (p, *J* = 6.1 Hz, 1 H), 3.73 (m, 1 H), 3.64 (m, 1 H), 3.53 (td, *J* = 12.0, 2.9 Hz, 1 H), 2.99 – 2.95 (comp, 3 H), 2.81 – 2.76 (comp, 2 H), 2.63 (s, 3 H), 1.75 (ddt, *J* = 14.5, 11.2, 4.1 Hz, 1 H), 1.59 (dq, *J* = 14.5, 3.2 Hz, 1 H), 1.44 (d, *J* = 5.9 Hz, 3 H). ¹³C NMR (125 MHz) δ 169.0, 136.3, 127.4, 122.0, 121.9, 119.3, 118.7, 112.8, 111.2, 77.4, 66.1, 55.3, 48.5, 44.0, 43.1, 25.2, 23.4, 19.2. HRMS (CI) *m/z* calcd for C₁₈H₂₃N₃O₂ (M+H)⁺, 314.1869; found, 314.1867.

(±)-(3*R*,4*S*)-1-(2-(1*H*-Indol-3-yl)ethyl)-3-((*R*)-1-hydroxyethyl)-4-(methylamino)piperidin-2-one. Zinc (powder, 530 mg, 8.14 mmol) was added in three portions over 1.5 h to a solution of **13** (85 mg, 0.27 mmol) in acetic acid (aq 80%, 20 mL) at 60 °C, and stirred at 60 °C for an additional 1 h. The reaction was cooled to room temperature, whereupon excess zinc was filtered and washed with ethyl acetate. Ethyl acetate (100 mL) was added to the filtrate and zinc acetate immediately precipitated out of solu-

tion as a fluffy white solid. The zinc acetate was filtered, and washed with excess ethyl acetate. The solvent was removed *in vacuo*, and the crude material was taken up in aqueous HCl (1 M, 30 mL) and washed with ether (2 x 30 mL). The aqueous fraction was basified with aqueous NaOH (40% w/v) to pH ~ 14. The basic solution was extracted with CH₂Cl₂ (4 x 50 mL). The combined organic fractions were washed with saturated aqueous NaHCO₃ (100 mL), water (100 mL), brine (1 x 100 mL), dried (Na₂SO₄), filtered, and concentrated *in vacuo* to give 78 mg (91%) of (±)-(3*R*,4*S*)-1-(2-(1*H*-indol-3-yl)ethyl)-3-((*R*)-1-hydroxyethyl)-4-(methylamino)piperidin-2-one as an oil (>95% purity, ¹H NMR). ¹H NMR (500 MHz, CD₃OD) δ 7.58 (dt, *J* = 7.8, 0.7 Hz, 1 H), 7.32 (dt, *J* = 7.8, 0.7 Hz, 1 H), 7.09 – 7.05 (comp, 2 H), 7.00 (td, *J* = 7.1, 1.0 Hz, 1 H), 4.37 (pent, *J* = 6.3 Hz, 1 H), 3.67 – 3.60 (m, 1 H), 3.56 – 3.49 (m, 1 H), 3.33 – 3.17 (comp, 1 H), 3.15 (pent, *J* = 3.9 Hz, 1 H), 3.11 – 3.04 (comp, 3 H), 2.40 (dd, *J* = 6.1, 4.2 Hz, 1 H), 2.34 (s, 3 H), 1.97 (app sex, *J* = 7.8 Hz, 1 H), 1.74 (dtd, *J* = 13.9, 7.1, 3.2 Hz, 1 H), 1.23 (d, *J* = 6.4 Hz, 3 H). ¹³C NMR (125 MHz, CD₃OD) δ 171.0, 138.1, 128.9, 123.6, 122.4, 119.7, 119.3, 113.1, 112.3, 67.5, 56.1, 51.4, 45.7, 33.5, 30.8, 24.8, 23.7, 22.5. HRMS (ESI) *m/z* calcd for C₁₈H₂₅N₃O₂ (M+H)⁺, 316.2020; found, 316.2024.

(3*R*,4*S*)-1-(2-(1*H*-Indol-3-yl)ethyl)-4-((3,5-dichlorobenzyl)(methylamino)-3-((*R*)-1-hydroxyethyl)piperidin-2-one (14a). A solution of 4.28 (50 mg, 0.16 mmol) and 3,5-dichlorobenzaldehyde (45 mg, 0.26 mmol) in acetonitrile (1 mL) was heated under reflux for 2 h. The reaction was cooled to room temperature, at which time NaBH₃CN (22 mg, 0.35 mmol) and acetic acid (53 μL, 0.89 mmol) were added. The reaction was stirred at room temperature for 20 h. The mixture was partitioned between saturated aqueous NaHCO₃ (6.5 mL), ethyl acetate (6.5 mL), and methanol (0.5 mL). The two phases were separated, and the aqueous phase was extracted with ethyl acetate (2 x 6.5 mL). The combined organic fractions were washed with brine (6.5 mL), dried (Na₂SO₄), filtered, and concentrated *in vacuo*. The crude material was purified via flash column chromatography eluting with a gradient of hexanes : ethyl acetate (1 : 1) → ethyl acetate : methanol (20 : 1), to give 36 mg (48%) of 14a as a white solid; ¹H NMR (600 MHz) δ 8.15 (d, *J* = 1.9 Hz, 1 H), 7.55 (d, *J* = 8.2 Hz, 1 H), 7.34 (dt, *J* = 8.1, 0.8 Hz, 1 H), 7.27 (t, *J* = 1.9 Hz, 1 H), 7.18 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1 H), 7.12 – 7.08 (comp, 3 H), 7.02 (d, *J* = 1.9 Hz, 1 H), 4.35 (dq, *J* = 6.5, 6.2 Hz, 1 H), 3.74 – 3.78 (m, 1 H), 3.64 – 3.69 (m, 1 H), 3.41 – 3.26 (comp, 3 H), 3.24 (dt, *J* = 13.4, 5.4 Hz, 1 H), 3.12 (dt, *J* = 16.9, 9.2 Hz, 1 H), 3.04 – 2.99 (m, 1 H), 2.97 – 2.92 (m, 1 H), 2.31 (t, *J* = 5.2 Hz, 1H), 2.03 (s, 3 H), 1.83 – 1.87 (comp, 3 H), 1.15 (d, *J* = 6.5 Hz, 3 H); ¹³C NMR (150 MHz) δ 171.7, 140.6, 136.3, 135.2, 128.1, 127.6, 127.3, 122.4, 122.1, 119.4, 118.4, 112.3, 111.5, 66.2, 58.9, 48.0, 47.4, 44.9, 37.3, 29.7, 23.2, 21.7, 20.1; HRMS (ESI) *m/z* calcd for C₂₅H₂₉Cl₂N₃O₂ (M+H)⁺, 474.1710 and 476.1686; found, 474.1712 and 476.1690.

Ethyl 3-((2-(1*H*-indol-3-yl)ethyl)(3-ethoxy-3-oxopropyl)amino)-3-oxopropanoate (22). Ethyl 3-((2-(1*H*-indol-3-yl)ethyl)amino)propanoate¹⁸ (3.00 g, 11.5 mmol) and 3-ethoxy-3-oxopropanoic acid (1.68 g, 12.6 mmol) were dissolved in CH₂Cl₂ (250 mL) and the solution was cooled

to 0 °C, whereupon EDCI•HCl (2.43 g, 12.6 mmol) and *i*-Pr₂NEt (3.30 g, 25.2 mmol, 4.38 mL) were added. The reaction was warmed to room temperature and stirred for 40 h, whereupon the reaction was diluted with CH₂Cl₂ (250 mL) and washed with 1 M HCl (500 mL). The aqueous layer was then extracted with an additional portion of CH₂Cl₂ (250 mL). The combined organic layers were washed with brine (500 mL), dried (Na₂SO₄) and concentrated under reduced pressure to yield 4.3g (99%) of crude 22 as an amber oil (4.3g 99%) which was judged to be >90% pure by ¹H NMR and carried forward without further purification. For characterization, a portion of the crude product was purified via column chromatography, eluting with hexanes/EtOAc (3:1 → 1:1) to provide 22 as an off-white solid. ¹H NMR (3:2 rotamer mixture) (400 MHz) δ 8.24 (s, 0.5 H), 8.15 (s, 0.3 H), 7.71 (d, *J* = 7.8 Hz, 0.3 H), 7.59 (d, *J* = 7.8 Hz, 0.6 H), 7.40 (t, *J* = 8.7 Hz, 0.9 H), 7.25 – 7.18 (m, 1 H), 7.18 – 7.11 (m, 0.9 H), 7.06 (d, *J* = 2.3 Hz, 0.3 H), 7.00 (d, *J* = 2.4 Hz, 0.6 H), 4.26 (q, *J* = 7.1 Hz, 0.7 H), 4.18 – 4.11 (comp, 3 H), 3.70 (t, *J* = 7.0 Hz, 1.2 H), 3.65 – 3.59 (comp, 1.9 H), 3.58 (s, 0.7 H), 3.55 (t, *J* = 7.1 Hz, 0.7 H), 3.16 (s, 1.2 H), 3.08 – 3.01 (comp, 1.9 H), 2.69 (t, *J* = 7.0 Hz, 1.2 H), 2.57 (t, *J* = 7.1 Hz, 0.7 H), 1.32 (t, *J* = 7.1 Hz, 1.2 H), 1.28 – 1.22 (comp, 4.7 H); ¹³C NMR (100 MHz) (3:2 rotamer mixture) δ 172.1, 170.1, 167.8, 167.6, 166.4, 166.1, 136.3, 127.3, 126.9, 122.4, 122.3, 122.1, 122.0, 119.7, 119.4, 118.8, 118.1, 112.9, 111.7, 111.4, 111.1, 61.4, 61.3, 61.0, 60.6, 49.9, 46.9, 44.5, 42.4, 41.2, 40.9, 36.6, 32.6, 29.7, 24.8, 23.2, 14.2, 14.1, 14.0; IR (NaCl, film) 3417, 2983, 2920, 2361, 2255, 2128, 1731, 1642, 1458, 1370, 1319, 1260, 1190, 1159, 1097, 1027, 1007, 824, 746, 618 cm⁻¹; HRMS (ESI) *m/z* calcd for C₂₀H₂₆N₂O₅ (M+Na)⁺ 397.1734; found 397.1733.

1-(2-(1*H*-Indol-3-yl)ethyl)piperidine-2,4-dione (23). A suspension of NaH (60% w/w in mineral oil, 135 mg, 3.80 mmol) in cyclohexane (20 mL) was heated under reflux, and a solution of 22 (700 mg, 1.90 mmol) in toluene (3 mL) was added dropwise. The reaction was stirred under reflux for 6 h during which time a tan precipitate formed. The reaction was then cooled to room temperature, and the solvent was removed under reduced pressure. The flask was then cooled to 0 °C, EtOH (20 mL) was added, and the reaction was stirred for an additional 2 h. The solution was then diluted with H₂O (200 mL) and 1 M HCl was added until pH = 1, and the aqueous solution was extracted with CH₂Cl₂ (3x 100 mL). The combined organic layers were washed with brine (100 mL), dried (MgSO₄) and concentrated under reduced pressure to afford a tan solid which was then resuspended in AcOH (2 mL) and H₂O (18 mL). The solution was heated under reflux until all starting material was consumed, as judged by LC-MS. The solution was cooled to room temperature, neutralized with sat. NaHCO₃ (100 mL), and extracted with CH₂Cl₂ (3x 100 mL). The combined organic extracts were then washed with brine (200 mL), dried (MgSO₄) and concentrated under reduced pressure to afford crude 23. The crude material was purified via flash column chromatography, eluting with hexanes:EtOAc (75:25 → 40:60) to furnish 313 mg (64%) of 23 as an off-white solid. ¹H NMR (400 MHz) δ 8.04 (bs, 0.5 H), 7.65 (d, *J* = 7.8 Hz, 0.99 H), 7.39 (d, *J* = 8.0 Hz, 1.05 H), 7.23 (t, *J* = 7.2 Hz, 1.18 H), 7.16 (t, *J* = 7.2, 1.08 H), 7.07 (s, 0.96 H), 3.85 (t, *J* =

6.9 Hz, 2.00 H), 3.34 (t,s overlapping, 3.90 H), 3.11 (t, $J = 7$ Hz, 2.05 H), 2.34 (t, $J = 6.1$ Hz, 1.91 H); ^{13}C NMR (100 MHz) δ 203.8, 168.1, 136.3, 127.3, 122.3, 122.1, 119.6, 118.5, 112.9, 111.4, 49.0, 44.4, 38.5, 23.7; IR (NaCl, film) 3283, 2924, 2854, 2362, 1726, 1644, 1489, 1644, 1489, 1457, 1350, 1227, 1095, 1011, 745 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{16}\text{N}_2\text{O}_2$ ($\text{M}+\text{Na}$)+ 279.1104; found 279.1100.

1-(2-(1H-Indol-3-yl)ethyl)-4-(methylamino)piperidin-2-one (24). To a solution of ketone **23** (65 mg, 0.25 mmol) in methanol (2.5 mL) was added methylamine (78 mg, 2.5 mmol, 33% wt. soln. in EtOH), AcOH (150 mg, 2.5 mmol), and NaBH_3CN (31 mg, 0.5 mmol), and the resulting mixture was stirred for 16 h at room temperature, whereupon an additional portion of NaBH_3CN (31 mg, 0.5 mmol) was added. After stirring for an additional 3 h, the reaction was diluted in 1 M HCl (50 mL) and washed with Et_2O (50 mL). The aqueous layer was treated with 1 M NaOH until the solution turned cloudy and $\text{pH} > 10$, and the resulting mixture was diluted with brine (20 mL) and extracted with CH_2Cl_2 (3 x 50 mL). The combined organic layers were dried (Na_2SO_4) and concentrated under reduced pressure. The crude material was purified via column chromatography, eluting with CH_2Cl_2 :MeOH: Et_3N (95 : 5 : 1) to yield 45 mg (66%) of **24** as a white foam. ^1H NMR (400 MHz) δ 8.02 (s, 1 H), 7.66 (d, $J = 7.9$ Hz, 1 H), 7.37 (dt, $J = 8.1, 1.0$ Hz, 1 H), 7.20 (ddd, $J = 8.1, 7.0, 1.3$ Hz, 1 H), 7.13 (ddd, $J = 7.9, 7.0, 1.1$ Hz, 1 H), 7.06 (d, $J = 2.3$ Hz, 1 H), 3.66 (td, $J = 7.1, 2.9$ Hz, 2 H), 3.25 – 3.18 (m, 1 H), 3.16 – 3.08 (m, 1 H), 3.07 – 3.02 (comp, 2 H), 2.86 – 2.78 (m, 1 H), 2.68 (ddd, $J = 17.1, 5.3, 1.8$ Hz, 1 H), 2.41 (s, 3 H), 2.22 (dd, $J = 17.1, 8.6$ Hz, 1 H), 1.90 (dt, $J = 8.1, 5.0$ Hz, 1 H), 1.58 – 1.46 (m, 1 H); ^{13}C NMR (100 MHz, CDCl_3) δ 168.3, 136.3, 127.5, 122.00, 121.97, 119.3, 118.7, 113.1, 111.2, 53.2, 48.12, 45.7, 43.42, 38.8, 33.3, 28.4, 22.9; IR (NaCl, film) 3252, 3053, 2926, 2856, 2799, 2361, 1624, 1498, 1456, 1344, 1302, 1264, 1233, 1158, 1102, 1011 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{16}\text{H}_{21}\text{N}_3\text{O}$ ($\text{M}+\text{Na}$)+ 294.1577; found 294.1574.

1-(2-(1H-Indol-3-yl)ethyl)-4-((3,5-dichlorobenzyl)(methylamino)piperidin-2-one (25a). To a stirred solution of **24** (16 mg, 0.11 mmol) and 3,5-dichlorobenzaldehyde (25 mg, 0.314 mmol) in 1,2-dichloroethane (1.4 mL) was added sodium triacetoxyborohydride (45 mg, 0.21 mmol). The reaction was stirred for 4 h, whereupon it was diluted with CH_2Cl_2 (50 mL) and washed with sat. NaHCO_3 (50 mL) and brine (50 mL). The organic layer was dried (Na_2SO_4) and concentrated under reduced pressure to afford crude **25a**. The crude material was purified via flash column chromatography, eluting with hexanes:EtOAc:TEA (40:60:1) to furnish 17 mg (68%) of **25a** as a yellow solid. ^1H NMR (400 MHz) δ 8.10 (s, 1 H), 7.66 (d, $J = 7.9$ Hz, 1 H), 7.37 (d, $J = 8.1$ Hz, 1 H), 7.24 (s, 1 H), 7.31 – 7.25 (comp, 3 H), 7.18 (td, $J = 8.1, 7.0$ Hz, 1 H), 7.10 (td, $J = 7.9, 7.0$ Hz, 1 H), 7.05 (d, $J = 2.3$ Hz, 1 H), 3.74 – 3.50 (comp, 4 H), 3.17 – 3.00 (comp, 4 H), 2.94 – 2.81 (m, 1 H), 2.65 (dd, $J = 17.2, 5.2$ Hz, 1 H), 2.51 – 2.41 (m, 1 H), 2.16 (s, 3 H), 2.05 – 1.91 (m, 1 H), 1.60 (qd, $J = 11.5, 6.0$ Hz, 1 H); ^{13}C NMR (100 MHz) δ 144.0, 137.0, 135.6, 128.2, 127.9, 127.4, 122.7, 122.5, 120.0, 119.4, 113.9, 111.8, 57.31, 57.29, 48.4, 46.9, 37.6, 35.0, 26.6, 23.7; IR (NaCl, film) 3624, 2509, 2929, 2855, 2793, 2242, 1623, 1569, 1499, 1456, 1433, 1345, 1303, 1231, 1209,

1158, 1125, 1098, 1074, 1036 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{25}\text{Cl}_2\text{N}_3\text{O}$ ($\text{M}+\text{H}$)+ 430.1447; found 430.1448.

N-(3,5-Dichlorobenzyl)-N-methylpiperidin-4-amine (27a). A solution of *tert*-butyl 4-(methylamino)piperidine-1-carboxylate (44 mg, 0.2 mmol), 3,5-dichlorobenzaldehyde (53 mg, 0.3 mmol), acetic acid (12 mg, 0.2 mmol), and sodium triacetoxyborohydride (85 mg, 0.4 mmol) in 1,2-dichloroethane (2 mL) was stirred at room temperature for 18 h, whereupon the solution was diluted with 1 M NaOH (15 mL) and extracted with Et_2O (3 x 15 mL). The combined organic extracts were dried (Na_2SO_4) and concentrated via rotary evaporation. The crude material was purified by flash chromatography eluting with hexane:EtOAc: Et_3N (95:5:1) to give 48 mg (64%) of *tert*-butyl 4-((3,5-dichlorobenzyl)(methylamino)piperidine-1-carboxylate as an opaque white oil. ^1H NMR (400 MHz) δ 7.23 – 7.19 (comp, 3 H), 4.16 (s, 2 H), 3.50 (s, 2 H), 2.75 – 2.61 (comp, 2 H), 2.55 (tt, $J = 11.5, 3.6$ Hz, 1 H), 2.17 (s, 3 H), 1.83 – 1.70 (comp, 2 H), 1.45 (comp, 11 H). ^{13}C NMR (100 MHz) δ 154.7, 143.9, 134.7, 126.9, 126.7, 79.4, 61.1, 57.0, 43.1, 37.7, 28.4, 27.9; IR (NaCl, film) 2975, 2940, 2854, 2788, 1694, 1590, 1569, 1451, 1425, 1365, 1330, 1275, 1244, 1159, 1111, 1046, 1004 cm^{-1} . HRMS (ESI) m/z calcd for $\text{C}_{18}\text{H}_{26}\text{Cl}_2\text{N}_2\text{O}_2$ ($\text{M}+\text{H}$)+ 373.1444; found 373.1454.

A solution of *tert*-butyl 4-((3,5-dichlorobenzyl)(methylamino)piperidine-1-carboxylate (52 mg, 0.14 mmol) and trifluoroacetic acid (160 mg, 1.4 mmol) in CH_2Cl_2 (2.8 mL) was stirred at room temperature for 24 h, whereupon the solvent was removed by rotary evaporation. The resulting solid was resuspended in 1 M HCl (20 mL) and washed with Et_2O (20 mL). 3 M NaOH was added to the aqueous layer until the solution became cloudy, whereupon it was extracted with CH_2Cl_2 (3 x 20 mL), dried (Na_2SO_4) and concentrated via rotary evaporation to give 29 mg (76%) of **27a** as a yellow paste. ^1H NMR (400 MHz) δ 7.22 (t, $J = 0.6$ Hz, 3 H), 3.52 (s, 2 H), 3.18 (d, $J = 12.2$ Hz, 2 H), 2.66 – 2.46 (m, 4 H), 2.19 (s, 3 H), 1.51 (qd, $J = 12.1, 4.1$ Hz, 3 H); ^{13}C NMR (100 MHz) δ 144.1, 134.7, 126.9, 126.8, 61.1, 56.9, 46.1, 37.7, 29.0; IR (NaCl, film) 2941, 2851, 2791, 1589, 1568, 1449, 1432, 1384, 1353, 1273, 1208, 1045 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{13}\text{H}_{18}\text{Cl}_2\text{N}_2$ ($\text{M}+\text{H}$)+ 273.0920; found 273.0923.

1-(4-((3,5-Dichlorobenzyl)(methylamino)piperidin-1-yl)-2-(1H-indol-3-yl)ethan-1-one (28a). A solution **27a** (33 mg, 0.12 mmol), indole-3-acetic acid (32 mg, 0.18 mmol), EDCI-HCl (35 mg, 0.18 mmol), and $i\text{Pr}_2\text{NEt}$ (0.8 mL, 0.48 mmol) in THF (1.2 mL) was stirred at rt for 2 h. The reaction was then diluted with 1 M aq. NaOH (20 mL) and extracted with CH_2Cl_2 (3 x 15 mL). The combined organic extracts were dried (Na_2SO_4) and concentrated via rotary evaporation. The crude material was purified by flash chromatography, eluting with hexanes:EtOAc: Et_3N (40:60:1) to give **28a** (40 mg, 78%) as a white solid. ^1H NMR (400 MHz) δ 8.17 (s, 1 H), 7.65 (dq, $J = 7.9, 0.9$ Hz, 1 H), 7.35 (dt, $J = 8.1, 1.0$ Hz, 1 H), 7.22 (t, $J = 2.0$ Hz, 1 H), 7.19 (ddd, $J = 8.2, 7.0, 1.2$ Hz, 1 H), 7.16 – 7.14 (m, 2 H), 7.13 (ddd, $J = 8.0, 7.1, 1.1$ Hz, 1 H), 7.09 (dd, $J = 2.3, 1.1$ Hz, 1 H), 4.74 (d, $J = 13.7$ Hz, 1 H), 4.05 – 3.93 (m, 1 H), 3.86 (dd, $J = 4.4, 1.0$ Hz, 2 H), 3.37 (s, 2 H), 2.93 (td, $J = 12.9, 2.7$ Hz, 1 H), 2.55 (td, $J = 11.2,$

10.6, 3.0 Hz, 2 H), 2.07 (s, 3 H), 1.80 (d, $J = 13.0$ Hz, 1 H), 1.58 (d, $J = 13.0$ Hz, 1 H), 1.40 (qd, $J = 12.3, 4.4$ Hz, 1 H), 1.12 (qd, $J = 12.3, 4.3$ Hz, 1 H); ^{13}C NMR (126 MHz) δ 170.1, 144.0, 136.4, 135.0, 127.3, 127.3, 126.9, 122.6, 122.5, 119.9, 119.1, 111.4, 109.8, 61.1, 57.1, 46.1, 41.8, 37.9, 31.9, 28.7, 27.7; IR (NaCl, film) 3058, 2926, 2857, 2790, 1624, 1568, 1454, 1434, 1353, 1268, 1227, 1149, 1125, 1098, 1047 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{25}\text{Cl}_2\text{N}_3\text{O}$ (M+Na) $^{+}$ 452.1267; found, 452.1268.

1-(2-(1H-Indol-3-yl)ethyl)-N-(3,5-dichlorobenzyl)-N-methylpiperidin-4-amine (29a). To a solution of **28a** (21 mg, 0.05 mmol) in THF (1 mL) was added a 0.5 M solution of alane in toluene (0.15 mmol, 0.3 mL) and stirred for 3 h at room temperature. The reaction was then diluted with 1 M NaOH (20 mL) and extracted with CH_2Cl_2 (3 x 15 mL). The combined organic extracts were dried (Na_2SO_4) and concentrated via rotary evaporation. The crude material purified by flash chromatography eluting with hexanes/EtOAc/ Et_3N (40:60:1) to give 15 mg (70%) of **29a** as a yellow paste. ^1H NMR (400 MHz) δ 8.13 – 8.05 (s, 1 H), 7.61 (d, $J = 7.8$ Hz, 1 H), 7.34 (dt, $J = 8.0, 0.9$ Hz, 1 H), 7.23 (s, 3 H), 7.21 – 7.16 (m, 1 H), 7.15 – 7.09 (m, 1 H), 7.01 (d, $J = 2.2$ Hz, 1 H), 3.80 – 3.71 (m, 1 H), 3.53 (s, 2 H), 3.21 – 3.11 (comp, 2 H), 3.02 – 2.93 (comp, 2 H), 2.75 – 2.65 (comp, 2 H), 2.46 (tt, $J = 11.5, 3.8$ Hz, 1 H), 2.21 (s, 3 H), 2.05 (td, $J = 11.8, 2.4$ Hz, 2 H), 1.85 (comp, 3 H), 1.70 (qd, $J = 12.0, 3.8$ Hz, 2 H). ^{13}C NMR (100 MHz) δ 144.2, 136.2, 134.7, 127.4, 126.9, 126.8, 122.0, 121.4, 119.2, 118.8, 114.5, 111.1, 61.3, 59.3, 57.0, 53.4, 37.9, 27.9, 23.2. IR (NaCl, film) 3412, 3179, 3057, 2943, 2853, 2808, 2360, 1620, 1589, 1568, 1455, 1433, 1376, 1353, 1303, 1228, 1145, 1097, 1049, 1014 cm^{-1} . HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{27}\text{Cl}_2\text{N}_3$ (M+H) $^{+}$ 416.1655; found 416.1656.

ASSOCIATED CONTENT

Supporting Information

Experimental procedures and analytical data for all new compounds (PDF), purification of recombinant *TbMetRS* and *TbMetRS* aminoacylation assay procedures.

AUTHOR INFORMATION

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Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: