



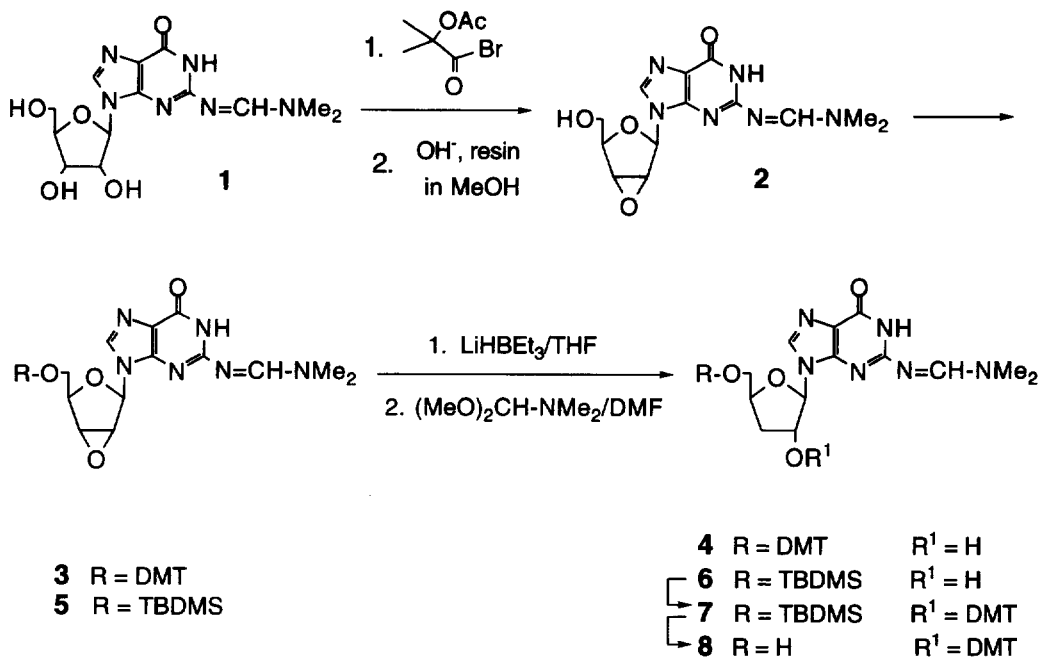
A Convenient Preparation of Protected 3'-Deoxyguanosine from Guanosine

Gong-Xin He* and Norbert Bischofberger

Gilead Sciences Inc., 353 Lakeside Drive, Foster City, CA 94404, USA

Abstract: Protected 3'-deoxyguanosine was synthesized from guanosine in five steps and 50% overall yield through reaction of the 2',3'-diol with α -acetoxyisobutyryl bromide followed by base treatment to form 2',3'-anhydroguanosine and subsequent selective reduction of the epoxide with LiHBEt₃.

3'-Deoxyribonucleosides have been studied for their potential antiviral, antiparasitic, and antitumor activities.¹ 3'-Deoxyadenosine, known as cordycepin, was the first reported nucleoside antibiotic.² Interest in this class of compounds also stems from their application in the sequencing of RNA³ and as probes for mechanism and structural studies.⁴ More recently, 3'-deoxynucleosides have been incorporated into oligonucleotides to study the effects of 2',5' linkages on hybridization with complementary oligonucleotides.⁵ It was found that 2',5' linked oligodeoxynucleotides associate through complementary base pairing, suggesting that 2',5' linked DNA comprises an alternative genetic material to 3',5' linked DNA. As part of a program to explore the structure-activity relationship of an oligodeoxynucleotide thrombin inhibitor,⁶ we required the preparation of 3'-deoxyguanosine on a preparative scale. A survey of the literature revealed that current methods are cumbersome and require difficult separations with low overall yields.⁷ Approaches which have been described for 3'-deoxyribonucleosides include attachment of the heterocyclic base to a suitably protected 3'-deoxyribose unit.⁸ This method requires a protected 3'-deoxyribose which has to be prepared in many steps. Another approach to 3'-deoxyribonucleosides involves the conversion of the corresponding ribonucleosides to the 3'-O-thiocarbonates⁹ or the 3'-deoxy-3'-halo derivatives¹⁰ followed by reduction. However, synthesis of 3'-deoxyguanosine by this method results in a mixture of 2' and 3'-deoxy isomers in low yield.^{9,10} Cordycepin can be synthesized from adenosine in high yield using α -acetoxyisobutyryl bromide or similar reagents.^{11,12} However, in our hands, guanosine itself did not react with α -acetoxyisobutyryl bromide in acetonitrile, presumably because of its low solubility, while N²-isobutyrylguanosine resulted in a mixture of products. In this paper, we wish to report that by using dimethylaminomethylene protecting group for the N²-



amino group of guanosine, a modified procedure can successfully be used to synthesize 3'-deoxyguanosine in high yield.

The starting material N²-(dimethylaminomethylene)guanosine (**1**) was readily prepared from guanosine and dimethylformamide dimethyl acetal.⁷ To a solution of **1** (5.5 g, 16 mmol) and water (0.3 g, 17 mmol) in acetonitrile (200 mL), α -acetoxyisobutyryl bromide (13 g, 64 mmol) was added and the mixture was stirred at room temperature for 2 h resulting in a clear solution. The solution was poured into saturated NaHCO₃ (100 mL) and extracted with CH₂Cl₂ (4 x 200 mL), and the organic layers were dried (Na₂SO₄) and concentrated to give a white solid, which was a 1:1 mixture of N²-(dimethylaminomethylene)-2'-O-acetyl-3'-bromo-3'-deoxy-5'-O-(2,5,5-trimethyl-1,3-dioxolan-4-on-2-yl)guanosine and N²-(dimethylaminomethylene)-2'-O-acetyl-3'-bromo-3'-deoxyguanosine as indicated by TLC and ¹H-NMR. The solid was dissolved in methanol (200 mL) and stirred with 10 eq of basic ion-exchange resin [Amberlite, IRA-400(OH)]. After 2 h, TLC indicated complete consumption of starting materials. The resin was filtered off and washed with 10% HOAc in methanol (5 x 100 mL). The filtrate was neutralized with triethylamine and concentrated, yielding **2** (4 g) as a white solid.¹³ Reaction of **2** with DMT-Cl in dry pyridine gave **3** (7 g) as a pale yellow foam after column chromatography purification (SiO₂/CH₂Cl₂:CH₃OH:Et₃N =

100:5:1, 69% from 1). A solution of 3 in dry THF (400 mL) was treated with LiHBEt₃ (60 mmol, 1 M solution in THF) at 0°C and subsequently at room temperature for 10 h. After quenching with CH₃OH and 5% aqueous NaHCO₃, the mixture was concentrated. To the residue, 5% NaHCO₃ aqueous solution (200 mL) was added and the mixture was extracted with CH₂Cl₂ (3 x 200 mL). The combined organic layers were dried (Na₂SO₄) and concentrated resulting in a crude product. ¹H-NMR indicated selective epoxide opening with the 3'-deoxy isomer being formed. At the same time, however, the dimethylaminomethylene protecting group had been removed. Thus, the crude product was re-dissolved in dry DMF (100 mL) and reacted with dimethylformamide dimethyl acetal (6 g, 50 mmol) in the presence of CH₃OH (0.5 mL) at room temperature for 10 h. After work-up and column chromatography (SiO₂/CH₂Cl₂:CH₃OH:Et₃N = 100:10:1), pure 4¹⁴ was obtained as a white foam (5 g, 50% from 1).

A similar procedure was also used in the preparation of 2'-O-DMT protected 3'-deoxyguanosine 8. 5 was prepared from 2 in 70% yield using TBDMS-Cl/imidazole/DMF. Reduction of 5 with LiHBEt₃ followed by reprotecting the 2-amino group as described above gave 6 in 61% yield. Treatment of 6 with DMT-Cl/pyridine/DMAP followed by removal of the TBDMS-group (TBAF/THF) yielded 8¹⁵ (overall 40% yield from 2). The protected 3'-deoxyguanosine derivatives 4 and 8 obtained above can be easily converted to the corresponding H-phosphonates or amidite derivatives for incorporation into oligonucleotides by solid phase oligonucleotide synthesis using standard H-phosphonate¹⁶ or amidite coupling protocols.^{7,17}

Acknowledgment: This work was supported in part by SBIR Grant No. 1R43 HL484311-01.

References and Notes:

- Widell, A.; Hansson, B. G.; Oeberg, B.; Nordenfelt, E. *Antiviral Res.* **1986**, *6*, 103.
 - Nakatsugawa, S.; Dewey, W. C. *Int. J. Radiat. Oncol., Biol., Phys.* **1984**, *10*, 1425.
 - Kumar, A.; Khan, S. I.; Mangani, A.; Khan, Z. K.; Katti, S. B. *Nucleosides & Nucleotides* **1994**, *13*, 1049
- Cunningham, K. G.; Hutchinson, S. A.; Manson, W.; Spring, F. S. *J. Chem. Soc.* **1951**, 2299
- Axelrod, V.; Kramer, F. *Biochemistry* **1985**, *24*, 5716.
- Moran, S.; Kierzek, R.; Turner, D. H. *Biochemistry* **1993**, *32*, 5247.

5. a) Dougherty, J. P.; Rizzo, C. J.; Breslow, R. *J. Am. Chem. Soc.* **1992**, *114*, 6254.
b) Hashimoto, H.; Switzer, C. *J. Am. Chem. Soc.* **1992**, *114*, 6255.
c) Jung, K.-E.; Switzer, C. *J. Am. Chem. Soc.* **1994**, *116*, 6059.
6. Wang, K. Y.; Krawczyk, S. H.; Bischofberger, N.; Swaminathan, S.; Bolton, P. H. *Biochemistry* **1993**, *32*, 11285.
7. Sheppard, T. L.; Rosenblatt, A. T.; Breslow, R. *J. Org. Chem.* **1994**, *59*, 7243.
8. a) Jenkins, S. R.; Holly, F. W.; Walton, E. *J. Org. Chem.* **1965**, *30*, 2851
b) Novak, J. J. K.; Sorm, F. *Coll. Czech. Chem. Comm.* **1973**, *38*, 1173.
c) Rizzo, C. J.; Dougherty, J. P.; Breslow, R. *Tetrahedron Lett.* **1992**, *33*, 4129.
9. Ogilvie, K. K.; Hakimelahi, G. H.; Proba, Z. A.; Usman, N. *Tetrahedron Lett.* **1983**, *24*, 865.
10. a) Mengel, R.; Muhs, W. *Nucleic Acid Res., Spec. Publ.* **1975**, *1*, S41.
b) Mengel, R.; Muhs, W. *Chem. Ber.* **1979**, *112*, 625.
11. Norman, D. G.; Reese, C. B. *Synthesis*, **1983**, 304.
12. a) Hansske, F.; Robins, M. J. *Tetrahedron Lett.* **1985**, *26*, 4295.
b) Bazin, H.; Chattopadhyaya, J. *Synthesis*, **1985**, 1108.
13. Compound 2: $^1\text{H-NMR}$ (300 MHz, DMSO- d_6) δ 11.32(br., NH, 1H), 8.61(s, =CH-, 1H), 8.00(s, 8-H, 1H), 6.12(s, 1'-H, 1H), 5.02(br., OH, 1H), 4.38(d, $J = 2.5$ Hz, 2'-H, 1H), 4.21(d, $J = 2.5$ Hz, 3'-H, 1H), 4.15(t, $J = 5.5$ Hz, 4'-H, 1H), 3.53(br, 5', 5''-H, 2H), 3.16(s, CH₃, 3H), 3.03(s, CH₃, 3H).
14. Compound 4: $^1\text{H-NMR}$ (300 MHz, CDCl₃) δ 9.48(br., NH, 1H), 8.45(s, =CH-, 1H), 7.61(s, 8-H, 1H), 7.16-7.41(m, Ar-H, 11H), 6.78(d, $J = 9$ Hz, Ar-H, 4H), 5.87(d, 3 Hz, 1'-H, 1H), 5.22(br., OH, 1H), 4.86(br., 2'-H, 1H), 4.60(br., 4'-H, 1H), 3.76(s, OCH₃, 6H), 3.32(br., 5', 5''-H, 2H), 3.07(s, CH₃, 3H), 3.01(s, CH₃, 3H), 2.10-2.30(m, 3', 3''-H, 2H). The ribo configuration of the 2'-hydroxyl was confirmed by 2D NMR techniques (HMBC and HMQC).
15. Compound 8: $^1\text{H-NMR}$ (300 MHz, CDCl₃) δ 8.92(br., NH, 1H), 8.25(s, =CH-, 1H), 7.47(s, 8-H, 1H), 7.12-7.32(m, Ar-H, 11H), 6.70(d, $J = 9$ Hz, Ar-H, 2H), 6.65(d, 9 Hz, Ar-H, 2H), 5.65(d, 5.5 Hz, 1'-H, 1H), 4.85(dd, $J = 7, 12.5$ Hz, 2'-H, 1H), 4.45(br., 4'-H, 1H), 4.27(d, $J = 12$ Hz, OH, 1H), 3.80(d, $J = 12$ Hz, 5'-H, 1H), 3.76(s, OCH₃, 3H), 3.73(s, OCH₃, 3H), 3.32(t, $J = 12$ Hz, 5''-H, 1H), 3.16(s, CH₃, 3H), 3.06(s, CH₃, 3H), 1.93-2.06(m, 3', 3''-H, 2H).
16. a) Marugg, J. E.; Tromp, M.; Kuyl-Yeheskiely, E.; van der Marel, G. A.; van Boom, J. H. *Tetrahedron Lett.* **1986**, *27*, 2661.
b) Froehler, B.; Ng, P.; Matteucci, M. *Nucleic Acid Res.* **1986**, *14*, 5399.
17. a) Sinha, N. D.; Biernat, J.; Koster, H. *Tetrahedron Lett.* **1983**, *24*, 5843.
b) Sinha, N. D.; Biernat, J.; McManus, J.; Koster, H. *Nucleic Acid Res.* **1984**, *12*, 4539.

(Received in USA 12 June 1995; revised 20 July 1995; accepted 27 July 1995)