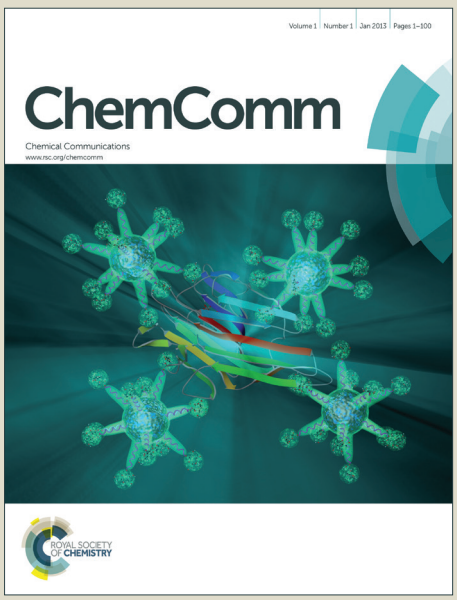
View Article Online
View Journal

ChemComm

Accepted Manuscript

This article can be cited before page numbers have been issued, to do this please use: D. Hymel, S. Cai, Q. Sun, R. S. Henkhaus, C. Perera and B. R. Peterson, *Chem. Commun.*, 2015, DOI: 10.1039/C5CC06325F.



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the Information for Authors.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard Terms & Conditions and the Ethical guidelines still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

ROYAL SOCIETY OF CHEMISTRY

www.rsc.org/chemcomm

Please do not adjust margins

View Article Online
DOI: 10.1039/C5CC06325F

Journal Name

COMMUNICATION

Published on 13 August 2015. Downloaded by Old Dominion University on 13/08/2015 16:13:55.

Fluorescent Mimics of Cholesterol that Rapidly Bind Surfaces of Living Mammalian Cells

Received 00th January 20xx,
Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

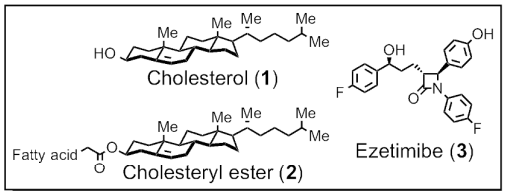
www.rsc.org/

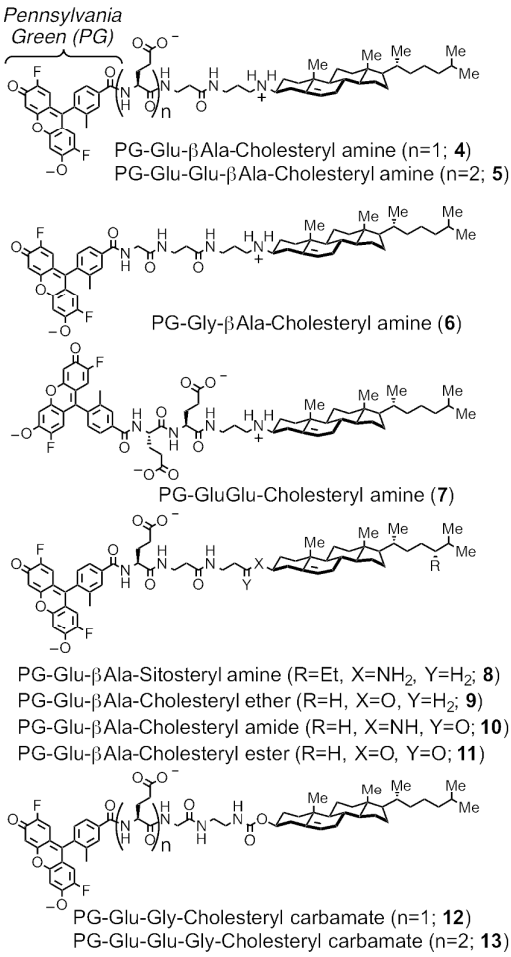
David Hymel, Sutang Cai, Qi Sun, Rebecca S. Henkhaus, Chamani Perera, and Blake R. Peterson*

Mammalian cells acquire cholesterol, a critical membrane constituent, through multiple mechanisms. We synthesized mimics of cholesterol, fluorescent *N*-alkyl-3β-cholesterylamine-glutamic acids, that are rapidly incorporated into cellular plasma membranes compared with analogous cholesteryl amides, ethers, esters, carbamates, and a sitosterol analogue. This process was inhibited by ezetimibe, indicating a receptor-mediated uptake pathway.

Cholesterol (**1**) is a critical constituent of membranes of mammalian cells. Cells acquire exogenous forms of this sterol through multiple mechanisms involving structurally distinct cell surface receptors. Lipoprotein particles such as low-density lipoprotein (LDL) and high-density lipoprotein (HDL) carry cholesteryl esters (**2**) and associated protein and lipid components throughout the bloodstream.[1] Cells expressing LDL and HDL receptors actively internalize these natural nanoparticles via receptor-mediated endocytosis. In contrast, Niemann-Pick C1 Like 1 protein (NPC1L1) plays key roles in the cellular uptake of dietary (unesterified) cholesterol (**1**), as found in mixed micelles.[2] This receptor was identified in 2004[3] as a target of ezetimibe (**3**), a drug used to treat hypercholesterolemia. More recent studies suggest that although NPC1L1 is a primary target of ezetimibe and its active glucuronide metabolite,[4-6] other proteins, such as the HDL receptor SR-BI, can also be inhibited by this drug.[7] Recent proteomics experiments have identified over 250 cholesterol-binding proteins, including receptors, channels, and enzymes.[8]

Derivatives of cholesterol have numerous biological applications. These compounds have been used to facilitate the delivery of small inhibitory RNA (siRNA),[9] enhance DNA transfection,[10] probe cellular membrane subdomains,[11] and assay cholesterol transport processes.[12-17] Cholesteryl carbamates have been extensively investigated, and cellular uptake of cholesteryl carbamate-conjugated siRNA has been reported to be similar to uptake of cholesteryl esters, requiring





The University of Kansas, Department of Medicinal Chemistry, Lawrence, KS 66045, E-mail: brpeters@ku.edu

† Electronic Supplementary Information (ESI) available: Experimental details, characterization data, and additional figures. See DOI: 10.1039/x0xx00000x

ChemComm Accepted Manuscript

Please do not adjust margins

View Article Online
DOI: 10.1039/C5CC06325F

Published on 13 August 2015. Downloaded by Old Dominion University on 13/08/2015 16:13:55.

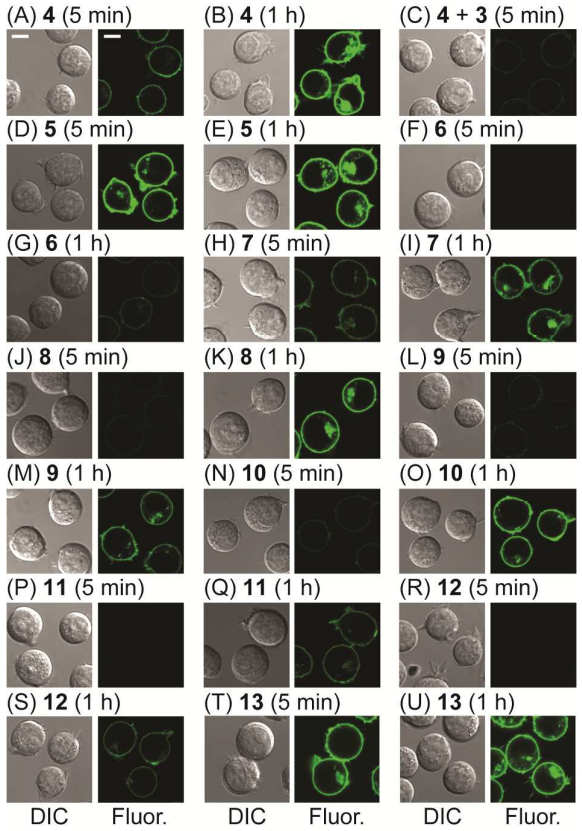


Fig. 1. Differential interference contrast (DIC) and confocal laser scanning microscopy of living Jurkat lymphocytes in media containing 10% serum. Cells were treated with fluorescent compounds **4–13** (2 μM) at 37 °C for 5 min or 1 h. In panel C, ezetimibe (**3**, 200 μM in 0.2% DMSO) was included to illustrate competitive inhibition of uptake of **4**. Scale bar = 7.5 microns.

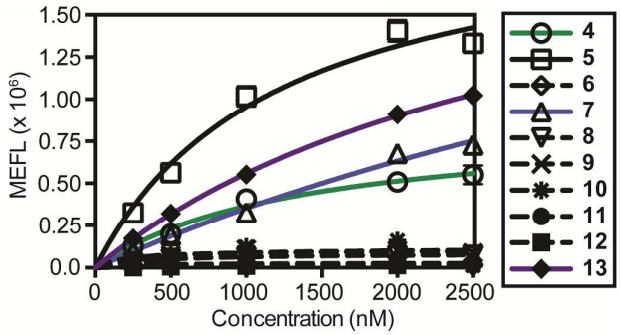


Fig. 2. Specific binding of compounds **4–13** to plasma membranes of living Jurkat lymphocytes in media containing 10% serum. Cells were treated with **4–13** for 5 min at 37 °C and analyzed by flow cytometry with and without excess ezetimibe (**3**, 200 μM) in saturation binding experiments. The linear non-specific binding component was subtracted from the total binding data followed by curve fitting with a one-site binding model (GraphPad Prism 6).

binding to HDL or LDL,[9] followed by internalization via HDL or LDL receptors. This initial lipoprotein-binding step can slow cellular uptake, and the presence of high concentrations of serum (e.g. 10%) in media typically reduces the activity of these compounds, likely because of competition between serum lipoproteins and cognate cell surface receptors.

In an effort to mimic the molecular recognition properties of free cholesterol (**1**), we hypothesized that the protonated secondary amino group of *N*-alkyl-3β-cholesterylamines,[18, 19] as found in compounds **4**-**7**, might function as a bioisostere for the 3β-hydroxyl group of cholesterol. Thus, in contrast to cholesteryl esters, cholesteryl carbamates, and structurally related compounds, which may require lipoprotein-mediated cellular uptake, these compounds might bind to receptors on cell surfaces that recognize free cholesterol or structurally similar metabolites through alternative mechanisms. We further hypothesized that the addition of anionic amino acids might affect binding to serum proteins and increase the affinity of these compounds for cells.

To investigate how structural features affect recognition of cholesterol derivatives and related compounds by proteins on the surface of cells, we synthesized the fluorescent molecular probes **4**-**13**. For probes derived from 3β-cholesterylamine, this steroid building block,[20] and some cholesterylamine-derived intermediates,[21] were prepared as previously reported. The novel building block 3β-sitosterylamine was prepared from sitosterol using methodology described for the synthesis of 3β-cholesterylamine.[20] The 4-carboxy Pennsylvania Green fluorophore was prepared as previously described.[22] Full synthetic details are provided in Scheme S1 and Scheme S2 of the supporting information. These probes were designed to systematically compare membrane anchors derived from *N*-alkyl-3β-cholesterylamines (**4**-**7**), a sitosteryl analogue (**8**), a *N*-acyl-3β-cholesterylamine (**10**), or cholesterol (**9**, **11**–**13**) linked to the hydrophobic Pennsylvania Green[23, 24] (PG) fluorophore through amino acid subunits. We hypothesized that the carbonyl linked to the steroid in amide **10**, ester **11** or carbamates **12** and **13** would be similar to natural cholesteryl esters, and this structural modification might correspondingly affect their ability to bind cell surfaces.

As shown in Figure 1, confocal laser scanning microscopy was employed to compare living human Jurkat lymphocyte cells after treatment with **4–13**. These experiments demonstrated that a brief (5 minute) treatment of cells with **4** or **5** (2 μM) at 37 °C results in robust fluorescent staining of cellular plasma membranes. Examination of these cells after 1 hour showed enhanced cellular binding, uptake of the probe, and localization in transferrin-positive early/recycling endosomes (Figure 1 and Figure S1 of the supporting information). Binding of **4** (and **5**) to cell surfaces was predominantly receptor-mediated as evidenced by up to 80% inhibition upon coaddition with excess (200 μM) ezetimibe (**3**, Compare Figure 1A and 1C, and see Figure S3 of the supporting information). This inhibition indicates that proteins on cell surfaces may recognize these compounds as mimics of free cholesterol or related cholesterol metabolites. These metabolites might include structurally related cholesterol sulfate[25] and cholesterol glucuronide,[26] which are present in micromolar concentrations in the bloodstream of animals. Comparison of **4** with **6** lacking the glutamic acid residue in the linker region revealed that the anionic moiety of **4** is critical for rapid high affinity/efficacy binding to cells (Figure 1, compare panels B and G). Moreover, despite the presence of a

ChemComm Accepted Manuscript

This journal is © The Royal Society of Chemistry 20xx

Please do not adjust margins

View Article Online
DOI: 10.1039/C5CC06325F

Published on 13 August 2015. Downloaded by Old Dominion University on 13/08/2015 16:13:55.

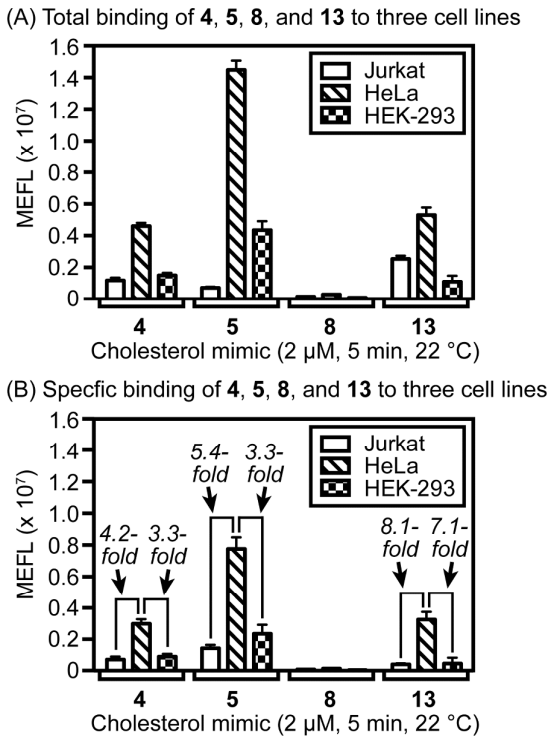


Fig. 3. Total binding (A) and specific binding (B) of compounds **4**, **5**, **8**, and **13** to three mammalian cell lines. Cells in media containing 10% serum were treated with compounds, without (A) and with (B) excess ezetimibe (**3**, 200 μM), in triplicate at 22 °C for 5 min. Jurkat, HeLa, and HEK-293 cells were suspended in media prior to treatment and analysis of fluorescence by flow cytometry.

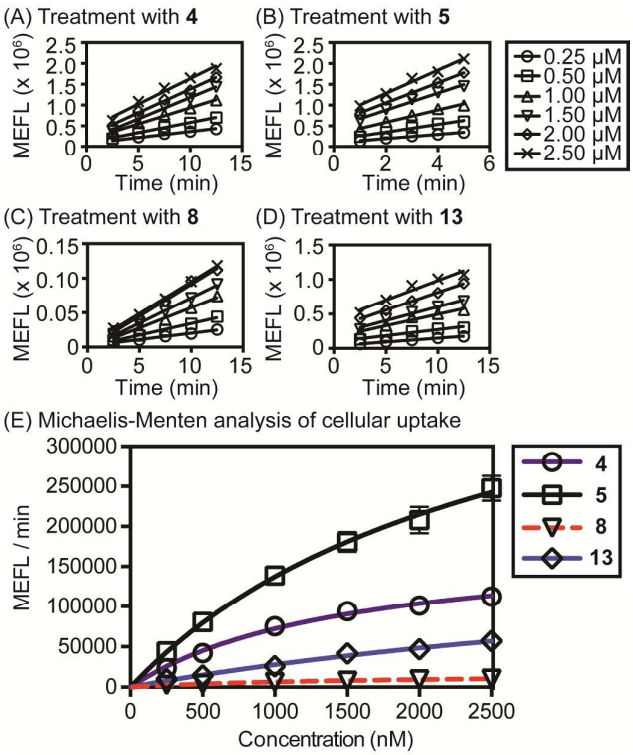


Fig. 4. Analysis of the kinetics of cellular uptake of compounds **4**, **5**, **8**, and **13**. Panels A-D: Living Jurkat lymphocytes in media containing 10% serum were treated with compounds in triplicate at 22 °C, aliquots were sampled at the times shown and fluorescence analyzed by flow cytometry. Panel E: Values of MEFL/min, obtained by linear regression of the data shown in A-D, was analyzed with a Michaelis-Menten model (GraphPad Prism 6).

structurally analogous glutamic acid, the cholesteryl carbamate **12** showed low cellular binding and cellular uptake compared with **4**, supporting the hypothesis that carbamates engage a mechanistically distinct cellular uptake pathway. However, this loss of activity of **12** could be at least partially rescued by addition of a second glutamic acid, as found in **13**.

Using ezetimibe (**3**, 200 μM, 0.2% DMSO) as a specific competitor, we quantified the relative affinities (K_d, app) and efficacies (B_{max}) of rapid binding of **4**-**13** to Jurkat lymphocytes

Compd.	$K_{d,\ app}$ (μM)	B_{max} (MEFL × 10^6)	K_M (μM)	V_{max} (MEFL / min × 10^5)
4	1.4 ± 0.5	0.9 ± 0.1	1.5 ± 0.3	1.8 ± 0.2
5	1.2 ± 0.2	2.1 ± 0.2	2.6 ± 0.7	5.0 ± 0.8
6	NC	NC	ND	ND
7	7.7 ± 4.1	3.1 ± 1.3	ND	ND
8	NC	NC	1.8 ± 0.5	0.2 ± 0.1
9	NC	NC	ND	ND
10	NC	NC	ND	ND
11	NC	NC	ND	ND
12	NC	NC	ND	ND
13	3.2 ± 0.6	2.34 ± 0.28	5.8 ± 2.2	1.9 ± 0.5

Table 1. Left columns: Apparent affinity (K_d, app) and efficacy (B_{max}) of binding of **4–13** to plasma membranes of living Jurkat cells. Cells were treated with compounds at 37 °C for 5 min in media containing 10% FBS (± SEM). Non-specific binding was quantified with ezetimibe (200 μM) as a competitor; vehicle = 0.2% DMSO; NC: Not calculated due to low efficacy. Right columns: Values of K_M and V_{max} calculated from Michaelis-Menten analysis of rapid time-dependent cellular uptake at 22 °C (Figure 3). ND: Not determined. MEFL: molecules of equivalent fluorescein. Data, based on measurements in triplicate, is reported as mean ± SEM.

in media containing 10% serum. Data from saturation binding experiments after treatment for five minutes, designed to limit cellular uptake by endocytosis, are shown in Figure 2, Table 1, and the supporting information. The high affinity and efficacy of binding of **4** and **5** to cell surfaces compared to **6**-**12** revealed that *N*-alkyl cholesterylamines bearing a anionic functional group and a spacer residue such as β-alanine most efficiently bind cell surfaces, a critical initial step for initiation of endocytosis and delivery of linked agents. The absence of substantial cellular binding of the sitosterolamine analogue (**8**), differing from **4** by the presence of an additional ethyl group in the tail of the sterol, further supports a specific receptor-mediated uptake mechanism. This result is consistent with limited receptor-mediated[27, 28] uptake of phytosterols such as sitosterol by mammalian cells.

To investigate whether other cell types show differential effects, we examined rapid (5 min) binding of **4**, **5**, **8**, and **13** to three different human cell lines. Human Jurkat cells, grown in suspension were compared with the human cell lines HeLa and HEK-293, which were suspended in media prior to treatment. As shown in Figure 3, HeLa cells bound the fluorescent cholesterol mimics to the greatest extent. Comparison of HeLa with Jurkat and HEK-293 cells revealed 3.3-fold to 8.1-fold more specific binding to HeLa cell surfaces compared to the other cell lines. These results might be explained by higher expression by HeLa cells of a specific receptor that interacts with these compounds.

ChemComm Accepted Manuscript

Please do not adjust margins

View Article Online
DOI: 10.1039/C5CC06325F

Published on 13 August 2015. Downloaded by Old Dominion University on 13/08/2015 16:13:55.

For these assays, cellular fluorescence was converted to molecular equivalents of fluorescein (MEFL) using fluorescent bead standards. For the Jurkat cell line, this analysis indicated that treatment with **4** or **5** at a concentration of 2 μM for 5 minutes loads 0.5–1.5 × 10^6 molecules into the plasma membrane of individual cells. This rapid and massive increase in cellular fluorescence suggested that an enzyme may be actively inserting these compounds into the cellular plasma membrane. Further analysis of time-dependent fluorescence resulting from treatment with **4**, **5**, **8**, and **13** using the Mechaelis-Menten model of enzyme kinetics is shown in Figure 4. These studies revealed that the cellular uptake of **4**, **5**, and to a lesser extent **13**, is highly efficient, with 1.8–5.0 × 10^5 molecules incorporated per minute per cell, consistent with a catalytic process. Kinetic values of K_M and V_{max} from this analysis are shown in Table 1.

In conclusion, we identified novel structure-activity relationships that govern binding of fluorescent cholesterol mimics to the surface of living mammalian cells. New cholesterol-mimetic membrane anchor motifs of **4**, **5**, and **13** were identified that engage a rapid cellular uptake pathway, consistent with a receptor-mediated process, that catalytically inserts these compounds into the plasma membrane. Although the receptor or enzyme targeted by these compounds has not yet been identified, it is unlikely to be NPC1L1, the classical pharmacological target of ezetimibe, because this protein is not highly expressed outside of the liver and intestine,[29] and the active metabolite ezetimibe-glucuronide, prepared as previously reported,[30] does not inhibit binding of **4** to Jurkat cell surfaces (data shown in Figure S2 of the supporting information). Given that cholesterol trafficking and distribution involves dynamic receptor-mediated and vesicular processes that are not completely understood,[31] these compounds have potential as novel probes and tools for the delivery of impermeable molecules into mammalian cells.

This work was supported by the National Institutes of Health (R01-CA83831 and P20-GM103638) and the University of Kansas Cancer Center.

1. J. L. Goldstein and M. S. Brown, *Arterioscler. Thromb. Vasc. Biol.*, 2009, **29**, 431-438.
2. L. J. Wang and B. L. Song, *Biochim. Biophys. Acta*, 2012, **1821**, 964-972.
3. S. W. Altmann, H. R. Davis, Jr., L. J. Zhu, X. Yao, L. M. Hoos, G. Tetzloff, S. P. Iyer, M. Maguire, A. Golovko, M. Zeng, L. Wang, N. Murgolo and M. P. Graziano, *Science*, 2004, **303**, 1201-1204.
4. A. B. Weinglass, M. Kohler, U. Schulte, J. Liu, E. O. Nketiah, A. Thomas, W. Schmalhofer, B. Williams, W. Bildl, D. R. McMasters, K. Dai, L. Beers, M. E. McCann, G. J. Kaczorowski and M. L. Garcia, *Proc. Natl. Acad. Sci. U.S.A.*, 2008, **105**, 11140-11145.
5. L. Ge, J. Wang, W. Qi, H. H. Miao, J. Cao, Y. X. Qu, B. L. Li and B. L. Song, *Cell. Metab.*, 2008, **7**, 508-519.
6. M. Garcia-Calvo, J. Lisnock, H. G. Bull, B. E. Hawes, D. A. Burnett, M. P. Braun, J. H. Crona, H. R. Davis, Jr., D. C. Dean, P. A. Detmers, M. P. Graziano, M. Hughes, D. E. Macintyre, A. Ogawa, A. O'Neill K, S. P. Iyer, D. E. Shevell, M. M. Smith, Y. S. Tang, A. M. Makarewicz, F. Ujjainwalla, S. W. Altmann, K. T. Chapman and N. A. Thornberry, *Proc. Natl. Acad. Sci. U.S.A.*, 2005, **102**, 8132-8137.
7. E. D. Labonte, P. N. Howles, N. A. Granholm, J. C. Rojas, J. P. Davies, Y. A. Ioannou and D. Y. Hui, *Biochim. Biophys. Acta*, 2007, **1771**, 1132-1139.
8. J. J. Hulce, A. B. Cognetta, M. J. Niphakis, S. E. Tully and B. F. Cravatt, *Nat. Methods*, 2013, **10**, 259-264.
9. C. Wolfrum, S. Shi, K. N. Jayaprakash, M. Jayaraman, G. Wang, R. K. Pandey, K. G. Rajeev, T. Nakayama, K. Charrise, E. M. Ndungo, T. Zimmermann, V. Koteliansky, M. Manoharan and M. Stoffel, *Nat. Biotechnol.*, 2007, **25**, 1149-1157.
10. B. Pitard, N. Oudrhiri, J. P. Vigneron, M. Hauchecorne, O. Aguerre, R. Toury, M. Airiau, R. Ramasawmy, D. Scherman, J. Crouzet, J. M. Lehn and P. Lehn, *Proc. Natl. Acad. Sci. U.S.A.*, 1999, **96**, 2621-2626.
11. S. B. Sato, K. Ishii, A. Makino, K. Iwabuchi, A. Yamaji-Hasegawa, Y. Senoh, I. Nagaoka, H. Sakuraba and T. Kobayashi, *J. Biol. Chem.*, 2004, **279**, 23790-23796.
12. J. Zhang, S. Cai, B. R. Peterson, P. M. Kris-Etherton and J. P. Heuvel, *Assay Drug Dev. Tech.*, 2011, **9**, 136-146.
13. D. Wustner, *Chem. Phys. Lipids*, 2007, **146**, 1-25.
14. D. Wustner, L. Solanko, E. Sokol, O. Garvik, Z. G. Li, R. Bittman, T. Korte and A. Herrmann, *Chem. Phys. Lipids*, 2011, **164**, 221-235.
15. F. R. Maxfield and D. Wustner, *Methods Cell Biol.*, 2012, **108**, 367-393.
16. M. Holtta-Vuori, R. L. Uronen, J. Repakova, E. Salonen, I. Vattulainen, P. Panula, Z. G. Li, R. Bittman and E. Ikonen, *Traffic*, 2008, **9**, 1839-1849.
17. S. Sankaranarayanan, G. Kellner-Weibel, M. de la Llera-Moya, M. C. Phillips, B. F. Asztalos, R. Bittman and G. H. Rothblat, *J. Lipid Res.*, 2011, **52**, 2332-2340.
18. B. R. Peterson, *Org. Biomol. Chem.*, 2005, **3**, 3607-3612.
19. D. Hymel and B. R. Peterson, *Adv. Drug Deliv. Rev.*, 2012, **64**, 797-810.
20. Q. Sun, S. Cai and B. R. Peterson, *Org. Lett.*, 2009, **11**, 567-570.
21. Q. Sun, S. Cai and B. R. Peterson, *J. Am. Chem. Soc.*, 2008, **130**, 10064-10065.
22. Z. R. Woydziak, L. Fu and B. R. Peterson, *Synthesis-Stuttgart*, 2014, **46**, 158-164.
23. L. F. Mottram, E. Maddox, M. Schwab, F. Beaufils and B. R. Peterson, *Org. Lett.*, 2007, **9**, 3741-3744.
24. L. F. Mottram, S. Boonyarattanakalin, R. E. Kovel and B. R. Peterson, *Org. Lett.*, 2006, **8**, 581-584.
25. C. A. Strott and Y. Higashi, *J. Lipid. Res.*, 2003, **44**, 1268-1278.
26. A. P. Wade, *Clin. Chim. Acta*, 1970, **27**, 109-116.
27. R. G. Gould, R. J. Jones, G. V. LeRoy, R. W. Wissler and C. B. Taylor, *Metabolism*, 1969, **18**, 652-662.
28. L. J. Wang and B. L. Song, *Biochim. Biophys. Acta.*, 2012, **1821**, 964-972.
29. H. R. Davis, Jr. and S. W. Altmann, *Biochim. Biophys. Acta*, 2009, **1791**, 679-683.
30. L. Kvaerno, T. Ritter, M. Werder, H. Hauser and E. M. Carreira, *Angew. Chem. Int. Ed. Engl.*, 2004, **43**, 4653-4656.
31. D. B. Iaea and F. R. Maxfield, *Essays Biochem.*, 2015, **57**, 43-55.

ChemComm Accepted Manuscript

This journal is © The Royal Society of Chemistry 20xx