Article

Halichondrin B: Synthesis of the C1-C22 Subunit

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Two efficient routes to the C1–C22 subunit of halichondrin B are described. The cage ketal 7, which contains 11 asymmetric centers embedded within the ABCDEF-ring framework, was assembled from (+)-conduritol E (27) in 18 steps and 4% overall yield. In a separate route, 7 was also synthesized in 18 steps and 2% overall yield from a derivative of α -D-glucoheptonic acid γ -lactone (62). While the former route installs the fully elaborated C-ring endowed with the correct C12 stereochemistry early in the synthesis, the latter features a late-stage introduction of the C12 stereocenter during the ultimate one-pot Michael addition/ketalization cascade to form the CDE-ring system of the cage. The importance of the C12 stereocenter to the crucial ketalization event is discussed through comparison of these two strategies.

Introduction

Halichondria okadai is a common marine sponge found off the coast of Japan. In 1986, Hirata and Uemura reported that extracts from this black-colored animal displayed compelling in vivo antitumor activity.^{1a} Eight active compounds were separated from these extracts and purified and were subsequently named the halichondrins. The most potent of these was halichondrin B (Figure 1), isolated in low yield ($1.8 \times 10^{-6}\%$), which displayed an in vitro IC₅₀ value for B-16 melanoma of 0.093 ng/mL. It was also found to have significant in vivo activity against B-16 melanoma, P-388 leukemia, and L-1210 leukemia.

Halichondrin B features 32 asymmetric centers that decorate a contiguous 54-carbon backbone. Structural features of note include a macrolactone spanning C1–C30, *exo*-methylene groups at C19 and C26, spiroketals at C38 and C44, and a 2,6,9-trioxatricyclo[3.2.2.0^{3,7}]-decane ketal (hereafter referred to as the "cage") embed-



FIGURE 1. Halichondrin B.

ded within the C8–C14 region of the molecule. The latter is an interesting structural motif unique to the halichondrins and is thought to be largely responsible for its remarkable biological activity. From their original data, Uemura and Hirata noted a strong correlation between the lipophilicity of the cage portion of the halichondrins and antitumor potency. Indeed, a C1–C38 diol (Figure 2) prepared by the Kishi group during the course of their synthetic studies² was found to display the same activity profile as that of halichondrin B over the 60 cell lines screened by the National Cancer Institute (NCI) and gave IC₅₀ values within 1 order of magnitude of the natural product itself. However, the in vivo potency of this compound was substantially lower than that of halichon-

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FIGURE 2. Biologically active halichondrin B analogues.

drin B. Recent drug discovery efforts at Eisai Research Institute have yielded many different halichondrin B analogues;³ the two most highly active ones (ER-086526 and ER-076349, Figure 2) feature a truncated H-ring bearing a hydrophilic side chain. The replacement of the macrolactone functionality with a hydrolytically stable ketone unit at C1 was done in order to prolong the in vivo activity of these compounds. The primary amine ER-086526 (E7389), which displays greater aqueous solubility than ER-076349, is currently in phase I and phase II trials at the National Cancer Institute.⁴

The initial findings by Hirata and Uemura prompted further investigation of the biological activity displayed by halichondrin B. It was determined by the NCI that halichondrin B is a mitotic inhibitor which binds to the vinca domain of tubulin, resulting in inhibition of microtubule formation and tubulin-dependent GTP hydrolysis.⁵ Halichondrin B was found to be extremely effective in vivo against human solid tumors which had been xenografted into immune-deficient mice.⁵ It also exhibited an IC₅₀ value of 0.3 nM against L-1210 murine leukemia cells, thereby surpassing in potency the best previously known agents dolastatin 10 (0.5 nM), rhizoxin (1 nM), and vinblastine (20 nM).⁵

Halichondrin B is currently in preclinical development by the NCI based on the data detailing its remarkable biological activity.^{5,6} However, the extremely low yield of natural product obtained from the original sponge has

(4) Compound E7389 is involved in three clinical trials: National Cancer Institute-Clinical Trials Results at http://www.cancer.gov/search/ResultsClinicalTrialsAdvanced.aspx?protocol-search-id=1667610, accessed 10/7/05.

thwarted progress toward this end. Although halichondrin B has since been isolated from three other sponge genera, the yields from these sources, which include Axinella $(1.8 \times 10^{-6}\%)$, ^{1b,c} Phakellia $(3.5 \times 10^{-6}\%)$, ^{1d} and Lissodendoryx ($4.0 \times 10^{-5\%}$),^{1e} have also been disappointingly low. The isolation of the halichondrins from four different sources has led Munro et al. to suggest that the natural products are synthesized by a microbial sponge symbiont rather than by the sponges themselves.^{7a} There are reports detailing attempts at aquacultural production of the most prolific sponge, *Lissodendoryx* n. sp. 1,^{7a,b} but this work is in the very early stages of development, and the authors note that this would require a yearly output of nearly 5 million kg of sponge in order to satisfy the anticipated clinical demand of 5 kg per year of halichondrin B.

Chemical synthesis has proven to be a viable method by which scarce natural products of considerable structural complexity may be accessed in larger quantities.⁸ The exceedingly small amounts of halichondrin B available from natural sources provides a strong impetus for its total synthesis. Efforts toward this end have been published by four groups, including Kishi,⁹ Salomon,¹⁰ Yonemitsu,¹¹ and our own.¹² Although a total synthesis of halichondrin B was published in 1992 by the Kishi group,^{9a} its length (ca. 53 linear steps/ca. 115 total steps) limits its scalability. Among several reports describing halichondrin subunit syntheses,¹² we have previously published syntheses of a C1-C15 subunit,^{12g} requiring 16 steps and proceeding in 0.6% overall yield from α -Dglucoheptonic acid γ -lactone, as well as a C1-C14 subunit model,^{12j} which was obtained in 3% overall yield over 18 steps from (+)-conduritol E.

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A key issue in both of our reported syntheses of the ABCDE-ring system was the late-stage construction of the acid-sensitive caged ketal. As shown in Scheme 1, C8, C9, and C11 hydroxyl unmasking in 1 gave the enone triol 2, which then underwent Michael addition of the C9 hydroxyl to set the C12 stereocenter and ketalization of the C14 ketone with the C8 and C11 hydroxyl groups to provide the C1–C15 subunit model 3. We hypothesized that poor control over the C12 stereochemistry during the Michael addition step led to low and variable yields of 3 in this first-generation synthesis.^{12g} In our subse-

quent synthesis of the C1–C14 subunit model 6^{12j} we introduced the C12 stereochemistry in a controlled manner early in the sequence by constructing the densely functionalized C-ring at the outset. Subjection of the enol ether substrate 4 to the acidic hydrolysis conditions shown led to clean formation of 6, presumably proceeding through the intermediate ketone 5. This favorable result appeared to validate our assertion that the C12 stereochemistry played a key role in facilitating cage formation by enforcing proximity between the reactive functionality. Furthermore, we were able to employ milder conditions in this case (p-TsOH, CH₂Cl₂, MeOH, H₂O) due to the absence of any robust protecting groups that resisted hydrolysis. The presence of the C8–C9 pentylidene ketal and the C11 TBS ether in our first synthesis necessitated the use of more forcing conditions (52% ag HF, CH_3CN) to induce the deprotection of the C8 and C9 alcohols, which preceded the Michael addition/ketalization cascade. We sought to modify the protecting group strategy of our first-generation synthesis in order to allow for the use of a milder acid promoter, thereby enabling a better comparison of the two ketalization protocols. We also desired to incorporate the adjacent F-ring into our synthesis in order to access the C1-C22 fragment of halichondrin B, incorporating our results in recently reported syntheses of the C14-C22 subunit.^{12i,k} In this paper, we report the synthesis of the C1-C22 subunit 7 by two routes, which feature alternative protocols for setting the C12 stereocenter and for effecting the critical ketalization event. Our work leading to the synthesis of 6 is also discussed in full detail, including results not described in our earlier communication.^{12j}

Results and Discussion

The retrosynthesis for our two routes to the C1-C22 subunit 7 are shown in Scheme 2.

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SCHEME 2



In route A, acidic hydrolysis of the enol ether mixture **8** is expected to lead to directly to the targeted subunit. Subunit convergence to form 8 will be realized via Wittig coupling of the C1–C13 aldehyde 9 with the ylide derived from the C14–C22 α -methoxyphosphonium salt **12a**. As mentioned previously, a potential advantage of this route is the preset configuration at C12, which is expected to facilitate the conversion of 8 to 7. Our route B synthesis employs enone 10 as the penultimate intermediate, which is anticipated to arise via Horner-Wadsworth-Emmons coupling between C1-C12 aldehyde 11 and the C13-C22 β -ketophosphonate **12b**. As with our earlier C1–C15 subunit synthesis, this route relies upon the late-stage assembly of the C-ring via Michael addition of the C9hydroxyl with C12 of the enone moiety, with the attendant issue of control at the C12 stereocenter prior to ketal formation.

Route A: Synthesis of the C1–C22 Subunit 7 from (+)-Conduritol E (27). Our retrosynthetic analysis of the pivotal route A intermediate 9 is depicted in Scheme 3. Recognizing an element of local C_2 -symmetry about the C8–C11 region of the ABC-ring aldehyde 9, we sought to preserve this substructure in our retrosynthetic simplification. We arrived at the L-mannose-configured γ -lactone 13, which contains the C8–C11 stereocenters and features differentiated termini at C7 and C12 for further elaboration. The dissymmetric nature of 13 was



of some significance, as we had also considered the analogous C_2 -symmetric bis(lactone) as a potentially attractive intermediate. At issue was the point at which desymmetrization could most effectively be accomplished. Our decision to pursue 13 was based on the proposal that it could be prepared via a symmetry-breaking ozonolytic cleavage of a C_2 -symmetric cyclohexene diol endowed with the L-mannose configuration. Compound 14, a protected derivative of (+)-conduritol E,¹³ meets these criteria and was therefore seen as an attractive precursor to 13. Ozonolytic cleavage of the carbon-carbon double bond in 14 was expected to give rise to the dissymmetric intermediate 15, which should form upon retro-[3 + 2]cycloaddition of the initially formed primary ozonide. Intramolecular trapping of the carbonyl and carbonyl oxide moieties in 15 with the C9 and C10 hydroxyls should then lead to formation of the dioxabicyclo[3.3.0]octane 16. Although the carbonyl oxide and carbonyl groups normally undergo intramolecular [3 + 2] cycloaddition to afford a secondary ozonide, the carbonyl oxide residues can be trapped intermolecularly by methanol to form peroxyacetals. Schreiber et al. has used this strategy for the desymmetrization of simple cycloalkenes.¹⁴ Moreover, Criegee et al. has demonstrated the intramolecular trapping of carbonyl oxides with tethered oxygen nucleophiles.¹⁵ Our prediction that **16** would be formed upon ozonolysis of 14 was predicated on this body of work. We

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have found only one isolated example in which these cyclization and desymmetrization tactics have been applied in tandem.¹⁶ Dehydration of the cyclic peroxyacetal in **16** with Ac_2O/Et_3N in line with Schreiber's method would then give rise to lactone **13**. In this way, the C8–C11 stereochemical array of halichondrin B would be assembled from a readily available C_2 -symmetric precursor. By virtue of its symmetry, the C8 and C11 stereocenters in **14** are homotopic, as are the C9 and C10 stereocenters. This reduces the problem of its synthesis to one involving the formation of *two* asymmetric centers instead of *four*. Desymmetrization of **14** effectively produces two new stereocenters by breaking this degeneracy, resulting in a substantial increase in molecular complexity.¹⁷

The synthesis of an appropriate (+)-conduritol E derivative for this purpose is shown in Scheme 4. Subjection of dimethyl L-tartrate acetonide 17^{18} to a one-pot DIBAl-H reduction/Grignard addition sequence provided diol 18 as a 67:23:10 mixture of diastereomers in 80% yield.^{13a-c} Benzylation and acetonide hydrolysis was then followed by acetylation to provide a chromatographically separable mixture of diacetates, and the L-mannitol-configured diene 19 was isolated as the major product in 51% overall yield over the three steps. Slow addition of the Grubbs second-generation ruthenium benzylidene catalyst 20^{19} (3 mol %) to a refluxing solution of 19 in benzene induced ring-closing metathesis (RCM) to afford the (+)-conduritol E derivative 21 in 98% yield. Subse-

SCHEME 5



quent treatment of $\mathbf{21}$ with K_2CO_3 in MeOH provided the expected diol $\mathbf{14a}$ in 96% yield.

With 14a in hand, the stage was now set for ozonolytic desymmetrization. However, treatment of a solution of 14a in CH_2Cl_2 (-78 °C) with O_3 followed by quenching with Ac₂O and Et₃N provided none of the expected lactone. We reasoned that the desired γ -lactone product may suffer decomposition in the presence of Et₃N via a β -elimination pathway. Fortunately, the use of AcCl (TsCl and MsCl were also effective) in conjunction with a heterogeneous base (NaOAc) effected the desired dehydration reaction without inducing subsequent product decomposition, and lactone 13a was obtained in ca. 25% yield following treatment of the crude product mixture with Amberlist H⁺ resin in refluxing MeOH (Scheme 5). The methanolysis step was necessary since the methoxy acetals so obtained proved more stable to purification by silica gel chromatography than the intermediate hemiacetals. However, the bis(methoxy acetal) 22a was isolated as the major product of this reaction in ca. 50% yield (ca. 21% yield of the C_2 -symmetric isomer), which pointed to a deleterious reduction pathway. This side reaction temporarily foiled our goal of desymmetrization, since the termini at C7 and C12 were rendered equivalent with respect to oxidation state.

A plausible mechanistic rationale for the formation of 22a is depicted in Scheme 6. Since it is known that carbonyl oxides can undergo dimerization followed by extrusion of a molecule of O_2 ,²⁰ we reasoned that a related process might be occurring in our system. The peroxyacetal **15a**, which contains a highly nucleophilic hydroperoxide residue, could compete with intramolecular carbonyl oxide capture via an intermolecular process with the hemiacetal-carbonyl oxide species 23. Since the desired intramolecular closure results in the formation of a strained bicyclic system, the intermolecular process (which involves an inherently more potent nucleophile) may have a comparable rate, thus allowing it to compete effectively. This would result in the formation of the geminal bis(alkylperoxide) dimer 24, which can lose O_2 as shown to afford the hydroxy aldehyde 25 and the bis-(hemiacetal) 22b. Rapid closure of 25 to the hemiacetal form would then give another molecule of **22b**. Since O₂

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SCHEME 6



is lost, this fragmentation process does indeed represent a net reduction and would account for the formation of the bis(acetal) product **22a**. Conducting the ozonolysis under high dilution conditions in an attempt to thwart this dimerization/reductive fragmentation pathway did not noticeably improve the ratio of lactone (**13**) to bis-(acetal) (**22**) products.

PhH. reflux

55%

(+ 25% 26)

HO

ÖTBDPS

14b

Although disappointed by the interference of this unanticipated side reaction, we were nonetheless encouraged by the isolation of lactone 13a, which validated our proposal that the desymmetrization of C_2 -symmetric cyclohexene diols typified by 14a could be achieved by ozonolysis. We reasoned that an increase in steric bulk adjacent to the reactive functionality might retard intermolecular processes such as the dimerization/reductive fragmentation pathway that plagued our earlier studies. The easiest way to achieve this would be to modify the protecting groups at the C8 and C11 alcohols. Accordingly, protection of diol 18 (Scheme 7) as its bis(TBDPS ether) followed by acetonide cleavage with FeCl₃ adsorbed on silica gel²¹ provided the diastereomerically pure diene diol 26 in 23% yield after chromatography, with 47% of recovered acetonide isolated as well. Since the TBDPS ethers suffered cleavage with other acidic conditions



SCHEME 8



employed for the hydrolysis of the acetonide group, the use of the more chemoselective FeCl_3 reagent was required; however, the reaction could not be driven to completion in this case. Moreover, these yields were obtained only after recovering and resubjecting the acetonide-protected bis(TBDPS) ether mixture to FeCl_3 . SiO_2 for a total of three cycles. Subjection of **26** to RCM with 10 mol % of the Grubbs catalyst **20**¹⁹ in refluxing benzene provided diol **14b** in 55% yield along with 25% of recovered **26**. The increased steric bulk adjacent to the vinyl groups apparently hinders the RCM reaction, as a substantial portion of the starting material failed to react even at high catalyst loadings. This setback, coupled with the inefficiency of the acetonide cleavage step, ultimately led us to develop a more satisfactory synthesis of **14b**.

We were hopeful that the TBDPS groups could be introduced after RCM had been achieved, which suggested that (+)-conduritol E itself might be employed as an intermediate. Although several syntheses of this biologically active natural product have been reported,¹⁴ there has been much recent activity in the preparation of conduritol derivatives (including the E-subtype) via RCM.^{13a-c} Accordingly, (+)-conduction E (27) was prepared in four steps and 46% overall yield from diol 18 with only minor modification of the published procedures (Scheme 8). Treatment of 27 with 2.5 equiv of TBDPSCl in DMF in the presence of pyridine for 2 days at room temperature provided the desired diol 14b in 83% yield, accompanied by 16% of the undesired regioisomer 28, which could be recycled to 27 with TBAF. Interestingly, substitution of imidazole for pyridine greatly increased the reaction rate; however, 14b and 28 were formed in a ratio of ca. 1:1 in this case.

With ample quantities of **14b** in hand, we turned our attention once again to the ozonolytic desymmetrization step. We were delighted to find that passing ozone through a -78 °C solution of **14b** in EtOAc followed by addition of Ac₂O/Et₃N and warming to 0 °C resulted in the formation of the desired lactone **13b** as a ca. 2:1 mixture of anomers as evidenced by ¹H NMR analysis of the crude product mixture (Scheme 9). Subsequent treatment with TESCl/imidazole provided lactone **13c** as a single diastereomer in 80% yield. A small amount of the bis(TES acetal) was also formed, indicating that the dimerization/reductive fragmentation pathway had not

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been suppressed entirely (cf. 22b, Scheme 6). Another noteworthy feature of this reaction is the stability of 13b and 13c to $\mathrm{Et}_3N,$ demonstrating that the bulky TBDPS groups effectively shield the base-sensitive γ -lactone from decomposition.

Having differentiated the two termini of the L-mannitol-configured backbone, we sought to construct the highly oxygenated C-ring of the natural product, thereby setting the stereochemical configuration at C12 at an early stage. We envisioned hydroboration-oxidation of an exocyclic enol ether^{12e,h,22} derived from the C12 carbonyl in **13c** as the most expedient method for achieving this goal. Engagement of the enol ether double bond by a borane reagent would be expected to occur on the convex face of the bicyclic system, thereby providing the desired 12R stereochemistry. The highly polarized nature of such olefins leads to exceptionally high levels of regioselectivity as well, leading to exclusive production of the anti-Markovnikov net hydration product.

Treatment of lactone 13c with a slight excess of dimethyltitanocene (Petasis' reagent)²³ in PhCH₃ at 85 °C cleanly afforded enol ether 29 in quantitative yield (Scheme 10). This compound could be purified on silica gel without decomposition or isomerization of the double bond to the endocyclic position, a process that plagued the isolation of a similar exocyclic enol ether in our work.^{12e,h} Although the steric bulk afforded by the adjacent TBDPS ethers may explain the exceptional stability of 29, another possible reason is that the adjacent oxygen substituent at C11 destabilizes any developing positive charge at C12, thereby attenuating the reactivity of the double bond. Although we had success with the Tebbe reagent²⁴ in this transformation as well, we found the preparation and handling of the Petasis reagent^{23b} more convenient for large-scale applications. Treatment of 29 with BH₃·SMe₂ followed by the addition of H₂O₂ and NaOH gave the desired C-ring alcohol 30 in 72% yield accompanied by 13% of its C12 epimer 31. We found the use of bulkier borane reagents unsuccessful, with catecholborane giving lower yields and 9-BBN failing to react with 29 altogether. The imposing

SCHEME 10





29 R = TBDPS

Мe

32 R = TBDPS

bulk of the TBDPS groups seems a likely cause of this sluggish reactivity, as the reaction is complete within 1 h at room temperature with the comparatively small BH₃ reagent. The undesired C12 epimer **31** was recycled to enol ether 29 by base-induced elimination of the derived iodide.

While **29** proved reasonably stable to acidic media such as silica gel, we encountered an acid decomposition pathway for this compound during the course of this work. Upon dissolution in CDCl₃ (of somewhat poor quality) that contained HCl, we observed the clean formation of the acyclic hemiacetal 33 as a single diastereomer by ¹H NMR analysis, with presumed intermediacy of the endocyclic enol ether 32 (Scheme 11). After concentration and chromatography on silica gel, the hemiacetal collapsed with loss of TESOH to afford aldehyde 34 in 96% yield. While acyclic hemiacetals are not normally sufficiently stable to be observed, 33 may be stabilized by an intramolecular hydrogen bond with the furan oxygen. The acidic medium in which 33 was formed may also prevent the dissociation of the hemiacetal. The fact that only one diastereomer of 33 was observed spectroscopically suggests that the hemiacetal stereochemistry was retained under these conditions. Exposure of **29** to an excess of PPTS in CH_2Cl_2 , on the

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SCHEME 12



other hand, gave only a mixture of enol ether products, of which **32** was a major component. This indicates that fragmentation of the bicyclic system to give furan byproducts requires more strongly acidic conditions.

The tentative C7 and C12 stereochemical assignments were verified by NOE studies of pivalate 35, obtained in 87% yield upon treatment of 30 with PivCl/DMAP (Scheme 12). Irradiation of the anomeric proton at C7 led to enhancement of the C12 methylene proton signals, an observation that is only possible if these protons reside within the concave face of the bicyclic framework. Since we sought to carry the C12 pivaloate ester throughout the synthesis, and the C7 hemiacetal had to be unmasked prior to further elaboration, we did not normally isolate 35 except for the purpose of the NOE study. A one-pot procedure was developed from 30 in which the pivalation reaction mixture was simply quenched with HCl in MeOH, thereby inducing chemoselective cleavage of the TES group to provide hemiacetal 36 in 98% yield.

Having confirmed the correct stereochemistry at C12, we turned our attention to the C7 hemiacetal moiety at the other end of the molecule, which would serve as the starting point for the assembly of the A- and B-rings. Kishi and co-workers have developed an efficient protocol for constructing the C1-C7 portion of halichondrin B from a C7 hemiacetal,^{9f,g} and we sought to employ a modification of this method for our purpose. Treatment of 36 with an excess of (methoxymethylene)triphenylphosphorane at 0 °C followed by stirring the crude product mixture in a biphasic mixture of THF/10% aq KOH provided 37 in 87% yield (Scheme 13), with only trace amounts of the corresponding Z isomer detected by ^{1}H NMR analysis. The aqueous base treatment was necessary due to the observation that silvl ether migration had occurred during the Wittig step. We found that KOH induced equilibration of the crude product mixture to give 37, with only a trace of its silvl ether migration regioisomer remaining. Treatment of 37 with a catalytic amount of OsO₄ in aqueous acetone in the presence of N-methylmorpholine-N-oxide (NMO)²⁵ yielded hemiacetal 38 in 73% yield, which features the desired 7R stereochemistry, along with 22% of its C7 epimer 39. The





stereoselectivity observed for this reaction is consistent with Kishi's empirical rule,²⁶ a rationale formulated to explain the high levels of diastereofacial selectivity exhibited by chiral allylic ethers in addition reactions. It was possible to recycle **39** by oxidative cleavage followed by hydrolysis of the intermediate formate ester with Amberlist-IRA-400 (OH) resin, affording 36 in 70% yield. The overall sequence from 36 to 38 represents a net ring expansion in which the C7 carbinol unit has been formally inserted in a stereoselective manner.

Attachment of a suitably functionalized C1–C5 chain at C6 would simultaneously complete construction of the B-ring and set the stage for A-ring closure. The Lewisacid-promoted C-glycosidation with an allylic silane nucleophile constitutes an appropriate tool for this task and has been employed by Kishi during his synthesis of this fragment of the halichondrins.^{9g,j} Accordingly, we sought to activate 38 as a glycosyl donor via acetylation of the two hydroxyl groups. Treatment of 38 with Ac₂O/ DMAP/py²⁷ at room temperature provided diacetate 40 in 68% yield as a single diastereomer (Scheme 14). Approximately 30% of a monoacetate product mixture was also isolated, which could not be converted to 40 upon resubjection to the reaction conditions. Fortunately, use of the more powerful combination of $Ac_2O/TMSOTf^{28}$ gave 40 in 65% yield²⁹ along with its C6 anomer, 41, in 35% yield after ca. 10 min at 0 °C. Apparently, the C7 hydroxyl of the β -hemiacetal **38** is quite difficult to acetylate, as 41 was not isolated from the milder reaction conditions.

With anomeric epimers 40 and 41 in hand, the stage was now set for the stereoselective installation of an A-ring handle at C6 via C-glycosidation followed by assembly of the complete AB-ring system. In the event, exposure of 40 to BF_3 ·OEt₂ in the presence of Kishi's

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⁽²⁹⁾ Recrystallization of 40 from heptane/CH₂Cl₂ provided crystals suitable for X-ray crystallographic analysis. See the Supporting Information.





allylic silane $\mathbf{42}^{\mathrm{9g}}$ provided the $\beta,\!\gamma\text{-unsaturated ester }\mathbf{43}$ in 81% yield as a ca. 3:1 (*E*)- to (*Z*)-mixture of isomers (Scheme 15). Although the analogous conversion of **41** to 43 proceeded sluggishly at room temperature, heating the reaction mixture to 60 °C resulted in complete conversion within 5 h. and 43 was isolated in 86% vield. The lower reactivity of **41** relative to **40** is probably stereoelectronic in nature, since the β -C6 acetate in **41** is equatorially disposed and hence more difficult to ionize due to the absence of an anomeric effect and the lack of anchimeric assistance by the C7 acetate. The construction of the B-ring had now been completed, and all of the carbons necessary for subsequent assembly of the fully elaborated A-ring were in place. Cleavage of the TBDPS ethers with TBAF buffered with AcOH followed by acetate cleavage with a basic resin provided triol 44 in 74% yield for two steps, with 8% yield of the pivalate migration product 45 also being isolated. Treatment of 44 with excess DBU in refluxing toluene resulted in isomerization of the C3–C4 double bond to the α,β unsaturated isomer, which then underwent Michael addition by the C7 hydroxyl. A mixture of 46 (desired) and its C3 epimer was observed by TLC analysis upon consumption of starting material (ca. 1 h). Equilibration



for another 16 h resulted in exclusive formation of **46** in 95% yield. A mixture (ca. 3:1) of **46** and the corresponding 3S isomer could be isolated in 94% yield when the reaction was conducted in refluxing benzene for 16 h, although no equilibration of the two was observed in this case. Apparently, the retro-Michael addition step that initiates the C3 epimerization requires the higher temperature afforded by the use of refluxing toluene. In like fashion, triol **45** was also converted to **46** in 95% yield, with migration of the pivalate group to the C13 hydroxyl occurring in this case. Although we had initially explored A-ring assembly directly from **43** using Triton-B methoxide, the C7 acetate proved resistant to cleavage in this case, prompting us to remove the bulky TBDPS ethers at C8 and C11 prior to cyclization.

To access the targeted C13 aldehyde 9, we had planned to first mask the C8 and C11 alcohols in **46** followed by pivalate cleavage and oxidation of the resultant C13 alcohol. We encountered a procedure,^{30a,b} however, whereby a primary TMS or TES ether may be chemoselectively oxidized to the corresponding aldehyde in the presence of secondary TMS or TES ethers under Swern conditions.^{30c} This prompted us to modify our plans for oxidation at C13 accordingly. Cleavage of the pivalate ester in 46 was accomplished with a basic resin in refluxing methanol to provide triol 47 in 93% yield, along with 7% of recovered 46 (Scheme 16). It should be noted that 1.64 g (5.15 mmol) of 47 has been prepared by the present route. Subsequent treatment of 47 with an excess of TESCl afforded the tris(TES ether) 48 in 91% yield. Oxidation of ${\bf 48}$ under standard Swern conditions $^{\rm 30a,c}$ at -40 °C proceeded cleanly, providing aldehyde 9 in 89% yield, with recovery of unreacted 48 in 6% yield. No products from oxidation at either C8 or C11 were detected. Since 9 is an α -alkoxy aldehyde, and hence prone to epimerization at C12 to a presumably more stable product, it was purified by chromatography on Florisil and used in subsequent reactions without delay.

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SCHEME 17



We had now arrived at the point of investigating the Wittig reaction of **9** with a suitable (α -methoxyalkyl)triphenylphosphorane.³¹ This reaction would serve a dual purpose, as subunit convergence would occur simultaneously with the installation of a C14-ketone surrogate (in the form of a C13–C14 enol ether) as a prelude to the ultimate cage-forming event. The use of (α -methoxyalkyl)triphenylphosphoranes as Wittig partners for fragment coupling in complex natural products synthesis is, to the best of our knowledge, unprecedented. Therefore, we sought to explore its viability through the synthesis of a model system that would incorporate a simple alkyl chain at C14 in the cage product.

The preparation and use of a Wittig reagent appropriate for this task is shown in Scheme 17. Treatment of commercially available 1,1-dimethoxyhexane (49) with PPh_3 and $BF_3{\boldsymbol{\cdot}}OEt_2{}^{31a}$ provided the (\alpha-methoxyalkyl)triphenylphosphonium tetrafluoroborate salt 50 in 67% vield after recrystallization from THF. Since (α -methoxvalkvl)triphenvlphosphoranes are known to be unstable at temperatures above -40 °C,^{31b} their generation from the corresponding phosphonium salts must be conducted at or below this temperature. Accordingly, deprotonation of 50 with *n*-BuLi at -40 °C resulted in the formation of the blood-red (α-methoxyalkyl)triphenylphosphorane 51. Cooling this solution to -78 °C followed by addition of aldehyde 9 and warming to room temperature resulted in the isolation of enol ethers 4Eand 4Z in 41% and 37% yields, respectively. Since these sensitive compounds suffered decomposition on silica gel, Florisil was again employed for their chromatographic separation and purification.

All of the functionality required for formation of the caged ketal is present in 4E and 4Z. Since hydrolysis of the enol ether moiety will provide a C14 ketone, both 4E and 4Z are potentially viable precursors to the desired cage product. For this reason, the subsequent step in the synthetic sequence was most conveniently conducted on the mixture of geometrical isomers without chromato-

SCHEME 18



graphic purification. We were pleased to observe that treatment of the crude Wittig product mixture with *p*-TsOH in $CH_2Cl_2-H_2O$ for 1 h followed by the addition of MeOH provided the cage ketal 6 in 67% yield over two steps from aldehyde 9 via the intermediate ketone 52 (Scheme 18). The ¹H NMR data we obtained for **6** (vide infra) were in excellent agreement with the literature data for its enantiomer, ent-6, which was synthesized by Aicher and Kishi in 1987.9b It was determined (TLC analysis) that enol ether hydrolysis in the biphasic CH₂-Cl₂-H₂O solvent system was complete within 1 h, although silyl ether cleavage and subsequent ketalization were not realized until the mixture was homogenized with MeOH. The choice of conditions for this reaction was critical, as the C11-C14 furans 53a and 53b, epimeric at C10, were observed as side products with other acid promoters. For example, 53a was isolated in ca. 30% yield upon treatment of a 4Z/4E mixture with 52% ag HF in CH_3CN (5:95 v/v), while the use of a biphasic aq HCl-THF system with the same substrate resulted in formation of **53b** and **6** in 21% and 41% yields, respectively. In these cases, the desired ketal 6 was observed to form alongside 53a/53b, and TLC analysis of the reaction mixture revealed that the latter were slowly formed at the expense of the former. A likely mechanism for furan formation involves protonation of the C8 oxygen in 6 and

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ionization to form the oxocarbenium ion 54. Loss of a proton at C13 would give the endocyclic enol ether 55 which, upon protonation of the allylic C9 oxygen, would suffer C-ring rupture via elimination to give the delocalized oxocarbenium species 56. Loss of the C11 proton would then give furan 53a. Epimerization at C10 to give 53b, in which the furanyl side chain is deployed equatorially on the tetrahydropyranyl ring, most likely proceeds via the intermediacy of a furanylic C10 carbocation. This cage-to-furan pathway is suppressed under the conditions given above (p-TsOH, CH₂Cl₂-H₂O; MeOH), an observation initially noted by Hart et al. on their work detailing the acid lability of the halichondrins.³² They reported an analogous furan-forming process upon exposure of homohalichondrin B (which contains the same cage unit as halichondrin B) to various acids (TFA, CD₃-OD; PPTS, CDCl₃). Their observation that no furan was formed in the presence of a large excess of *p*-TsOH in either CH₂Cl₂ or CH₂Cl₂-H₂O prompted us to employ these conditions.

The cage-to-furan transformation deserves further comment. The C1–C15 cage model **3** (Scheme 1) does not give rise to any of the corresponding furan **57** (Scheme 19) upon exposure to a variety of acids, an observation made by both us^{12g} and Cooper and Salomon^{10b} in our respective syntheses of **3**. It is well-known that α -alkoxy substituents destabilize carbenium ion species through the inductive withdrawal of electron density, thereby rendering the oxocarbenium ion derived from **3** less stable than **54**. The acid lability of **6** relative to that of **3** strongly suggests a correlation between the facility of furan formation and C14-oxocarbenium ion stability.

Having established a reliable set of reaction conditions for the introduction of the sensitive cage functionality, we sought to apply this Wittig coupling/ketalization strategy toward the synthesis of a complete C1-C22 subunit. This would entail the preparation of a C14-C22 fragment endowed with (a-methoxyalkyl)triphenylphosphonium salt functionality at the C14 terminus. We have previously described the synthesis of the C14-C22 subunit 12c (Scheme 20), which bears an aldehyde at C14, via two different symmetry-based routes.^{12i,k} To install the requisite phosphonium salt at C14, 12c was first treated with PPTS in MeOH/HC(OMe)₃ to provide the dimethyl acetal 58 in 95% yield. Without further purification, 58 was treated with PPh₃/BF₃·OEt₂^{31a} as done previously with 49, affording the $(\alpha$ -methoxyalkyl)triphenylphosphonium tetrafluoroborate salt 12a in 80% yield as a mixture of C14 epimers. This compound proved somewhat moisture-sensitive, giving aldehyde 12c and triphenylphosphine as decomposition products. Additionally, 12a could not be rigorously purified either by silica



gel chromatography or recrystallization. Without delay, a THF solution of 12a in a base-washed round-bottom flask was treated with a slight deficiency of n-BuLi at -78 °C. We observed immediate formation of a bloodred solution of the $(\alpha$ -methoxyalkyl)triphenylphosphorane 59 under these conditions. Subsequent addition of aldehyde 9 in THF at -78 °C was followed by slow warming to room temperature. Since we were unsuccessful in isolating the intermediate enol ether 8 in pure form by chromatography on Florisil, the crude mixture was simply exposed to the hydrolysis/ketalization conditions we had developed earlier for 4E/Z. We were delighted to find that the expected C1–C22 subunit of halichondrin B 7, which contains the complete ABCDEF-ring system of the natural product, was formed to the exclusion of any furan products in a two-step yield of 29%. It should be noted that the transformation of 8 into cage ketal 7 requires four distinct reactions, including hydrolysis of the enol ether and two silvl ethers, and a dehydrative ketalization of the resulting keto diol. Thus, despite the poorer efficiency relative to our model system (possibly a consequence of the purity of **12a** relative to that of **50**), overall efficiency of subunit convergence and cage assembly was still acceptable. The overall yield of 7 from (+)-conduction E (27) was 4% over 18 linear steps.

Route B: Synthesis of the C1–C22 Subunit 7 from α -D-Glucoheptonic γ -Lactone (62). The fully elaborated C1–C22 subunit (7) was also constructed as depicted through Route B in Scheme 2. The retrosynthesis for the formation of **11** is shown in Scheme 21.

Tris(TES)-protected aldehyde **11** is available from β , γ unsaturated ester **60**, through construction of the A-ring, followed by inversion at C11 and protecting group manipulations. β , γ -Unsaturated ester **60** comes from **61**, available in two steps from commercially available and relatively inexpensive α -D-glucoheptonic γ -lactone (**62**)

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10

OAc

SCHEME 22 1. 3-pentanone, H₂SO₄, 55% OTBS TBSCI, Imidazole, DMF 89% 62 61 DIBAI-H, PhCH₃, -78 °C O3, EtOAc, -78 °C / MgBr 1. в THF, 0 °C to rt Ac₂O, py, rt 2. 81% (2 steps) 3. TBAF, THF, rt 85% (3 steps) 63 64



OTBS

(\$0.07/g, Aldrich) through a ring expansion from the γ -lactone to form the B-ring. This starting material is extremely attractive for this synthesis, as it contains the correct stereochemistry for the C8, C9, and C10 stereocenters and the opposite stereochemistry for C11. This is also the same starting material that was used in our initial synthesis of the C1–C15 subunit.^{12g} Several problems arose in the previous synthesis using this starting material, and modifications to address these issues will be elaborated below.

Lactone **62** was regioselectively protected as the bis-(pentylidene) ketal, ^{12b,33} which left the C11 hydroxyl open for orthogonal protection as TBS ether **61** (Scheme 22). This fully protected lactone was then treated with DIBAl-H at -78 °C to yield the crude hemiacetal, which was treated with an excess of vinylmagnesium bromide³⁴ to give triol **63** after cleavage of the TBS group with TBAF (85% yield over the three steps). The selective formation of the C7 stereocenter is consistent with observations on related reactions of carbohydrate derivatives.³⁵

Ring expansion to form the B-ring was achieved through ozonolysis of **63** to yield the corresponding aldehyde, which was intramolecularly trapped with the C10 hydroxyl to form the B-ring pyran.³⁶ This was followed by protection of the remaining hydroxyls as acetates by treatment with acetic anhydride in pyridine to yield glycoside triacetate **64** as the major diastereomer by ¹H NMR in 81% yield over the two steps.

In the early stages of our previously published synthesis of the C1–C15 subunit from **61**,^{12g} an α -alkoxy-organolithium reagent was added to the γ -lactone, fol-

lowed by pinacol ring expansion of the corresponding diol to form the B-ring. This method suffered from lack of stereoselectivity and regioselectivity in the formation and rearrangement of the diol, which thus resulted in low yield of the target product. Additionally, the instability of the α -alkoxyorganostannane and its α -hydroxystannane precursor made that route unsuitable for supplying enough material for further chemical studies. This current sequence avoids those problems and forms the B-ring in high yield and high stereocontrol.

With the B-ring now installed, attention turned to the construction of the A-ring of halichondrin B. As predicted, glycoside triacetate **64** undergoes axial *C*-allylation³⁷ at the C6 anomeric center with Kishi's allylic silane 42^{9g} and an excess of BF₃·OEt₂ in acetonitrile to yield β , γ unsaturated ester 60, as a mixture of E and Z isomers in 70% yield (Scheme 23). The ratio of the E and Zisomers could not be determined, but proved to be inconsequential, as treatment of with benzyltrimethylammonium (Triton B) methoxide9b leads to concomitant acetate cleavage, double bond migration, and intramolecular Michael addition from the unprotected C7 hydroxyl group to the enoate moiety. This initially produces a mixture of C3 epimers, which then equilibrates to the more stable equatorial isomer 65, thus completing the A-ring in excellent yield.

Once the A-ring had been assembled, the next step was to invert the C11 hydroxyl to the desired halichondrin stereochemistry (Scheme 24). The oxidation/reduction protocol utilized by us developed previously was revisited.^{12g} Oxidation of the C11 hydroxyl with TPAP/NMO³⁸

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^{(35) (}a) Lygo, B.; Swiatyj, M.; Trabsa, H.; Voyle, M. Tetrahedron Lett. **1994**, 35, 4197–4200. (b) Shing, T. K. M.; Elsley, D. A.; Gillhouley, J. G. J. Chem. Soc., Chem. Commun. **1989**, 1280–1282.

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^{(37) (}a) Giannis, A.; Sandhoff, K. Tetrahedron Lett. 1985, 26, 1479–1482.
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(c) Kozikowski, A. P.; Sorgi, K. L. Tetrahedron Lett. 1982, 23, 2281–2284.



TESCI

Imidazole

quant

DMF

H

TESC

73

TESO

SCHEME 25



MeOH/H₂O

reflux

70% (after one resubjection of 71) HC

72

The next step required selective cleavage of the terminal pentylidene ketal, which was achieved utilizing FeCl₃ on SiO₂ in CHCl₃²¹ and optimized to yield exclusively 1,2 diol 69 in 90% yield after one recycle of 68.39

Because of what we had learned about the acid-induced decomposition of the CDE-ring cage ketal to a furan (Scheme 18), it was clear that the protecting groups for the C8, C9, and C11 hydroxyls in 69 were too robust. To address this issue and to establish a C12 aldehyde for subunit coupling, the conversion of **69** to **73** (Scheme 25) was executed.

Installation of a terminal alkene as a C12 aldehyde precursor preceded the C8, C9, C11 protecting group switch and was synthesized from diol 69 using the Corey-Winter protocol⁴⁰ to yield alkene **71** via thionocarbonate 70 (Scheme 25).41

TES

Protecting group removal was facilitated with acidic DOWEX 50WX4-400 resin in refluxing MeOH yielding 70% of triol **72** after one resubjection of starting material. This triol was then protected as the tris-TES ether (73) in quantitative yield.

Ozonolysis of 73 at -78 °C in CH₂Cl₂ and quenching with PPh₃ yielded aldehyde **11**, which was concentrated, redissolved in DME, and combined with the anion formed from commercially available dimethyl-2-oxoheptyl phosphonate to form α,β -unsaturated ketone **74** in 64% yield over the two steps (Scheme 26). Several other Horner-Wadsworth-Emmons conditions were also attempted⁴² but were either low yielding or led to decomposition.

⁽³⁸⁾ Griffith, W. P.; Ley, S. V.; Whitcombe, G. P.; White, A. D. J. Chem. Soc., Chem. Commun. 1987, 1625–1627.
 (39) Use of 80% aqueous AcOH^{12g} led to extended reaction times

⁽²⁰⁻²⁴ h), some TBS cleavage, and lower yields (maximum yields were 75%, also after one recycle of recovered SM).

⁽⁴⁰⁾ Winter, R. A. E.; Corey, E. J. J. Am. Chem. Soc. 1963, 85, 2677-2678

⁽⁴¹⁾ Gratifyingly, thionocarbonate $\mathbf{70}$ produced crystals suitable for X-ray crystallography, which allowed us to unambiguously determine the C11 stereochemistry. See the Supporting Information.

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JOC Article

SCHEME 26



SCHEME 28

The final step for the production of the ABCDE-ring model involved treatment of **74** under mildly acidic conditions (*p*-TsOH in CH₂Cl₂/MeOH/H₂O) to afford cage product **6** in 47% yield. All spectral data matched that of **6** synthesized as shown in Scheme 18, and Kishi's subunit model *ent-6*.^{9b}

Once the model study was complete, coupling of the ABC-ring and F-ring subunits followed by caged ketal formation to complete the C1–C22 subunit was explored (Scheme 27). Oxidation of F-ring alcohol **12d**^{12k} with freshly prepared Jones' reagent⁴³ followed by esterification with TMSCHN₂ yielded methyl ester **75** in 73% over the two steps. β -Ketophosphonate **12b** was synthesized in 93% yield by addition of the lithium anion of dimethyl methylphosphonate to **75** in THF at -78 °C.

Aldehyde 11 was formed in the same manner as before and was subjected to a quick purification on silica gel to remove triphenylphosphine oxide, which we discovered was interfering with recovery of excess 12b after the Horner–Wadsworth–Emmons coupling step. This aldehyde was then coupled to F-ring β -ketophosphonate 12b through a Horner–Wadsworth–Emmons reaction in refluxing DME to yield enone 10 (Scheme 28). Unfortunately, coupling of the C13–C22 subunit to the C1–C12 subunit proved to be problematic using the conditions employed for the synthesis of enone 74.

The use of sodium hydride proved not to be feasible due to the uncertainties in weighing out such a small



amount; therefore, a more easily handled base, KHMDS, was employed. Enone **10** was also found to be extremely difficult to isolate, as an unknown byproduct constantly coeluted with **10** during chromatography. Thus, enone **10** and this byproduct were carried on crude through the Michael addition/ketalization cascade to form ketal **7** in 10% yield over the three steps. Again, the complexity of this transformation deserves comment, in that three silyl ether cleavages, a Michael addition, and a ketalization constitute the conversion of **10** into **7**. This sample of **7** was identical in all respects to the one synthesized via the less problematic Route A as shown in Scheme 20.⁴⁴

Efforts are underway to complete the total synthesis of halichondrin B.

⁽⁴³⁾ Guanti, G.; Banfi, L.; Riva, R. Tetrahedron **1995**, 51, 10343–10360.

Experimental Section

Hemiacetal 13b. A solution of diol 14b (11.6 g, 18.7 mmol) in EtOAc (746 mL, 0.025 M) was cooled to -78 °C in a threenecked round-bottom flask fitted with a gas inlet tube. The solution was purged with ozone until a blue color was seen (40 min) that persisted for 2-3 min. At this point the solution was purged with N₂ until the blue color dissipated. To the resulting colorless solution was added Ac₂O (2.64 mL, 28.0 mmol, 1.5 equiv) and Et₃N (5.16 mL, 37.3 mmol, 2 equiv), and the solution was warmed to 0 °C and stirred for 2 h. The solution was quenched with water (300 mL) and warmed to room temperature, where it stirred for 1 h. After extraction with EtOAc $(3\times)$, the combined organic layers were washed sequentially with 5% aq HCl and brine, dried over MgSO₄, filtered, and concentrated in vacuo. The crude hemiacetal 13b (ca. 2:1 mixture of anomers) was isolated as a white foam and was used directly in the next reaction without further purification. Data for **13b**: $R_f 0.27 (20\% \text{ EtOAc in hexanes}); [\alpha]^{23} - 85$ (c 1.8, CHCl₃); IR (thin film) 3631-3134 (br), 1803 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) see S48; ¹³C NMR (75 MHz, CHCl₃) see S49; HRMS calculated for $C_{38}H_{44}O_6Si_2 (M + Na)^+ 675.2574$, found 675.2588 (2.1 ppm).

Aldehyde 9. To a solution of oxalyl chloride (0.263 mL of a 2~M solution in $CH_2Cl_2,\,0.526$ mmol, 2.2~equiv) in $CH_2Cl_2\,(1.2$ mL) was added DMSO (0.075 mL, 1.1 mmol, 4.4 equiv) at -50 °C. After stirring for 10 min at this temperature, 48 (158 mg, 0.239 mmol) in CH₂Cl₂ (1.2 mL) was added via cannula. The mixture was warmed to -40 °C and stirred for 1 h, at which point Et₃N (0.291 mL, 2.10 mmol, 8.8 equiv) was added. After stirring at -40 °C for 5 min, the reaction mixture was warmed to room temperature. The mixture was diluted with water (8 mL) and extracted with CH_2Cl_2 . The combined organic extracts were washed with brine, dried over Na₂SO₄, and concentrated in vacuo. Chromatography of the crude product on Florisil (10% to 20% EtOAc in hexanes) provided aldehyde 9 (116 mg, 0.213 mmol, 89%) as a colorless oil along with recovered 48 (10 mg, 0.015 mmol, 6%). Data for **9**: R_f 0.18 (20% EtOAc in hexanes); $[\alpha]^{23}D - 124$ (c 1.38, CHCl₃); IR (thin film) 2732, 1734 cm^-1; ¹H NMR (300 MHz, CDCl_3) δ 0.56–0.69 (m, 12 H), 0.92– 1.00 (m, 18 H), 1.26-1.51 (m, 2 H), 1.70-1.80 (m, 1 H), 2.05-2.14 (m, 1 H), 2.41 (ABX, $J_{\rm ab}=$ 15.4 Hz, $J_{\rm ax}=$ 5.0 Hz, 1 H), 2.58 (ABX, $J_{\rm ab}=$ 15.4 Hz, $J_{\rm bx}=$ 8.25 Hz, 1 H), 3.01 (dd, J=9.6, 2.1 Hz, 1 H), 3.69 (s, 3 H), 3.81-3.92 (m, 1 H), 4.06-4.15 (m, 2 H), 4.20-4.30 (m, 3 H), 4.45-4.50 (m, 1 H), 9.75 (d, J =2.8 Hz, 1 H); ¹³C NMR (75 MHz, CHCl₃) δ 4.6 (CH₂), 5.0 (CH₂), $6.8 (CH_3), \, 7.0 (CH_3), \, 29.5 (CH_2), \, 30.1 (CH_2), \, 40.5 (CH_2), \, 51.6$ (CH₃), 65.2 (CH), 67.3 (CH), 74.3 (CH), 74.4 (CH), 75.2 (CH), 76.1 (CH), 77.1 (CH), 85.4 (CH), 171.7 (C), 203.7 (CH); HRMS calculated for $C_{26}H_{48}O_8Si_2 (M + Na)^+ 567.2785$, found 567.2773 (2.1 ppm).

Enol ethers 4Z and 4E. A slurry of phosphonium salt 50 (112 mg, 0.242 mmol, 2.4 equiv) in THF (1 mL) was cooled to -40 °C for the dropwise addition of *n*-BuLi (0.125 mL of a 1.8 M solution in hexanes, 0.222 mmol, 2.2 equiv). The resulting blood-red solution was stirred at this temperature for 10 min before being cooled to -78 °C, and aldehyde 9 (55 mg, 0.10 mmol) in THF (1 mL) was added via cannula. After stirring at -78 °C for another 5 min, the mixture was warmed to room temperature and quenched with brine (4 mL). The layers were separated, and the aqueous later was extracted with Et₂O. The combined ether layers were then dried over Na₂SO₄ and concentrated in vacuo. The residue was triturated with hexanes to separate the product from excess phosphonium salt and triphenylphosphine oxide, and the combined hexane washings were concentrated in vacuo. Although the crude product could be used directly in the next step, analytically pure samples of enol ethers 4Z (24 mg, 0.037 mmol, 37%) and

4E (27 mg, 0.042 mmol, 42%) were obtained as oils by FCC on Florisil (5% to 10% to 20% EtOAc in hexanes). Data for 4Z: $R_f 0.33 (20\% \text{ EtOAc in hexanes}); [\alpha]^{23} - 62 (c 1.1, \text{ CHCl}_3); \text{ IR}$ (thin film) 1743, 1653 cm^-1; ¹H NMR (300 MHz, CDCl₃) δ 0.58-0.69 (m, 12 H), 0.87 (t, J = 6.8 Hz, 3 H), 0.94-1.02 (m,18 H), 1.15-1.55 (m, 8 H), 1.69-1.78 (m, 1 H), 2.03-2.28 (m, 3 H), 2.38 (ABX, $J_{\rm ab} = 15.2$ Hz, $J_{\rm ax} = 4.9$ Hz, 1 H), 2.55 (ABX, $J_{\rm ab}=15.2$ Hz, $J_{\rm ax}=8.2$ Hz, 1 H), 2.96 (dd, $J=9.4,\,2.2$ Hz, 1 H), 3.56 (s, 3 H), 3.68 (s, 3 H), 3.78–3.79 (m, 1 H), 3.93 (dd, J= 5.2, 3.6, 1 H), 4.00 (dd, J = 9.8, 5.3 Hz, 1 H), 4.06–4.17 (m, 2 H), 4.30 (dd, J = 9.9, 5.2 Hz, 1 H), 4.45 (dd, J = 9.3, 3.5 Hz, 1 H), 4.74 (d, J=9.4 Hz, 1 H); $^{13}\mathrm{C}$ NMR (75 MHz, CHCl_3) δ $5.1 (CH_2), 5.2 (CH_2), 7.1 (CH_3), 7.2 (CH_3), 14.0 (CH_3), 22.5$ (CH₂), 27.8 (CH₂), 29.8 (CH₂), 30.2 (CH₂), 31.0 (CH₂), 31.6 (CH₂), 40.7 (CH₂), 51.6 (CH₃), 54.2 (CH₃), 64.8 (CH), 67.1 (CH), 73.6 (CH), 74.0 (CH), 74.9 (CH), 76.1 (CH), 77.4 (CH), 78.2 (CH), 93.8 (CH), 160.5 (C), 171.9 (C); HRMS calculated for $C_{33}H_{62}O_8Si_2 (M + Na)^+$ 665.3881, found 665.3911 (5.0 ppm). Data for 4*E*: $R_f 0.18$ (20% EtOAc in hexanes); $[\alpha]^{23}$ _D -63 (*c* 1.1, CHCl₃); IR (thin film) 1743, 1675 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) & 0.57-0.68 (m, 12 H); 0.75-1.06 (m, 21 H), 1.24-1.53 (m, 8 H), 1.68–1.80 (m, 1 H), 2.03–2.20 (m, 3 H), 2.38 (ABX, $J_{\rm ab} = 15.4$ Hz, $J_{\rm ax} = 5.0$ Hz, 1 H), 2.55 (ABX, $J_{\rm ab} = 15.2$ Hz, $J_{ax} = 8.2$ Hz, 1 H), 2.94 (dd, J = 9.3, 1.9 Hz, 1 H), 3.54 (s, 3 H), 3.68 (s, 3 H), 3.75–3.89 (m, 1 H), 3.97 (dd, $J=5.3,\,1.2$ Hz, 1 H), 3.99 (dd, J = 7.6, 5.4 Hz, 1 H), 4.04–4.17 (m, 2 H), 4.27 (dd, J = 9.5, 5.5 Hz, 1 H), 4.78 (dd, J = 9.3, 3.4 Hz, 1 H), 4.88 (d, J = 8.9 Hz, 1 H); $^{13}\mathrm{C}$ NMR (75 MHz, CHCl_3) δ 5.0 (CH₂), 5.2 (CH₂), 7.0 (CH₃), 7.2 (CH₃), 13.9 (CH₃), 22.6 (CH₂), 26.7 (CH₂), 29.7 (CH₂), 30.2 (CH₂), 31.2 (CH₂), 31.7 (CH₂), 40.7 (CH₂), 51.6 (CH₃), 56.0 (CH₃), 64.8 (CH), 67.1 (CH), 74.0 (CH), 74.4 (CH), 75.2 (CH), 76.1 (CH), 77.5 (CH), 104.8 (CH), 158.2 (C), 171.9 (C); HRMS calculated for $C_{33}H_{62}O_8Si_2$ (M + Na)⁺ 665.3881, found 665.3911 (5.0 ppm).

ABCDE Cage Ketal 6, Route A. A crude mixture of 4Z and 4E (ca. 1:1) was prepared as described above from aldehyde 9 (35 mg, 0.064 mmol) and dissolved in CH₂Cl₂ (2.1 mL). Water (0.5 mL) was added, followed by p-TsOH·H₂O (12 mg, 0.063 mmol, 1 equiv). The biphasic mixture was stirred vigorously at room temperature for 1 h before MeOH (1.5 mL) was added. The mixture became homogeneous, and stirring was continued at room temperature for another 2 h. The reaction was quenched by the addition of saturated aqueous NaHCO₃, and the reaction mixture was extracted with CH₂-Cl₂. The combined organic layers were washed with brine, dried over Na₂SO₄, and concentrated in vacuo. FCC (20% to 30% EtOAc in hexanes) provided the desired ketal 6 (17 mg, 0.043 mmol, 67%) as a yellow oil. Data for **6**: R_f 0.25 (50%) EtOAc in hexanes); $[\alpha]^{23}$ _D -46 (*c* 1.0, CHCl₃); IR (thin film) 1740, 1437, 1189, 1140, 1082 cm⁻¹; ¹H NMR (500 MHz, CD₃-OD) δ 0.92 (t, J=7.3 Hz, 3 H), 1.25–1.45 (m, 7 H), 1.46–1.55 (m, 1 H), 1.55-1.65 (m, 1 H), 1.69 (ddd, J = 13.5, 11.2, 5.1Hz, 1 H), 1.75–1.85 (m, 2 H), 1.98 (dd, J = 13.2, 5.0 Hz, 1 H), 2.04 (d, $J = 13.2~{\rm Hz},\,1~{\rm H}$), 2.45 (ABX, $J_{\rm ab} = 15.7~{\rm Hz},\,J_{\rm ax} = 5.0$ Hz, 1 H), 2.52 (ABX, $J_{\rm ab}=$ 15.7 Hz, $J_{\rm bx}=$ 7.8 Hz, 1 H), 2.96 (dd, J = 9.5, 1.9 Hz, 1 H), 3.66 (s, 3 H), 3.79–3.86 (m, 1 H), 4.11 (dd, J = 6.6, 4.3 Hz, 1 H), 4.17 (dd, J = 6.6, 4.6 Hz, 1 H),4.29 (app td, J = 10.0, 4.8 Hz, 1 H), 4.33 (dd, J = 4.0, 2.0 Hz,1 H), 4.58 (app t, J = 4.5 Hz, 1 H), 4.68 (app t, J = 4.6 Hz, 1 H); ¹³C NMR (75 MHz, CD₃OD) δ 14.5 (CH₃), 24.0 (CH₂), 25.1 (CH₂), 31.3 (CH₂), 31.8 (CH₂), 33.3 (CH₂), 40.0 (CH₂), 41.4 (CH₂), 48.2 (CH₂), 52.3 (CH₃), 69.7 (CH), 75.4 (CH), 76.1 (CH), 76.2 (CH), 78.1 (CH), 79.6 (CH), 82.4 (CH), 83.8 (CH), 112.1 (C), 173.5 (C); HRMS calculated for $C_{20}H_{30}O_7$ (M + Na)⁺ 405.1889, found 405.1896 (1.728 ppm).

ABCDEF Cage Ketal 7, Route A. A solution of phosphonium salt **12a** (29 mg, 0.037 mmol, 2 equiv) in THF (0.4 mL) was cooled to -78 °C for the dropwise addition of *n*-BuLi (15 μ L of a 2.3 M solution in hexanes, 0.035 mmol, 1.9 equiv). The resulting blood-red solution was stirred at this temperature for 10 min before aldehyde **9** (10 mg, 0.018 mmol, 1 equiv) in THF (0.3 mL) was added via cannula. After stirring at -78

⁽⁴⁴⁾ The ¹H and ¹³C NMR data for **6** and **7** prepared via both routes A and B described here (Schemes 18, 20, 26, and 28) were in excellent agreement with that for halichondrin B itself.^{1a} See the Supporting Information.

°C for another 5 min, the mixture was warmed to room temperature and quenched with brine (4 mL). The layers were separated and the aqueous later was extracted with Et₂O. The combined ether layers were then dried over Na₂SO₄ and concentrated in vacuo. The resultant residue was passed through a plug of Florisil to give a crude enol ether mixture (ca. 11 mg, ca. 0.011 mmol, ca. 62%). As further purification was not feasible, this material was carried directly into the subsequent acid hydrolysis procedure. To this mixture was added water (0.1 mL) and CH₂Cl₂ (0.4 mL) followed by p-TsOH·H₂O (ca. 2 mg, ca. 0.010 mmol). The biphasic mixture was stirred vigorously at room temperature for 1 h before MeOH (0.3 mL) was added. The mixture became homogeneous, and stirring was continued at room temperature for another 3 h. The reaction was quenched by the addition of saturated aqueous NaHCO₃, and the reaction mixture was extracted with CH₂Cl₂. The combined organic layers were washed with brine, dried over Na₂SO₄, and concentrated in vacuo. FCC (40% to 50% EtOAc in hexanes) provided the desired C1-C22 ketal 7 (3.7 mg, 0.0053 mmol, 29% from 9) as a yellow oil. Data for 7: $R_f 0.25$ (50% EtOAc in hexanes); $[\alpha]^{23}_{D}$ -36 (c 0.50, CHCl₃); IR (thin film) 3070, 2928, 2855, 1739, 1428, 1360, 1177, 1111, 1078, 996, 918 cm⁻¹; ¹H NMR (500 MHz, CD_3OD) δ 1.04 (s, 9 H), 1.32-1.42 (m, 2 H), 1.58-1.77 (m, 4 H), 1.80-1.92 (m, 4 H), 1.96–2.03 (m, 2 H), 1.97 (dd, J = 13.3, 4.8 Hz, 1 H), 2.05 (d, J = 13.0 Hz, 1 H), 2.24–2.31 (m, 1 H), 2.40 (dd, J = 15.8, 5.1 Hz, 1 H), 2.51 (dd, J = 15.8, 7.7 Hz, 1 H), 2.64–2.70 (m, 1 H), 2.95 (dd, J = 9.6, 1.9 Hz, 1 H), 3.62 (s, 3 H), 3.75-3.83 (m, 2 H), 3.83-3.91 (m, 2 H), 4.01-4.07 (m, 1 H), 4.11 (J = 6.3, 3.8 Hz, 1 H), 4.17 (dd, J = 6.7, 4.3 Hz, 1 H), 4.28 (ddd, J = 9.7, 9.7, 4.4, 1 H), 4.33 (dd, J = 3.7, 2.0 Hz, 1 H), 4.53-4.62 (m, 2 H), 4.59 (app t, J = 4.6 Hz, 1 H), 4.67 (app t, J = 4.6 Hz, 1 H), 4.89 (aq, J = 1.8 Hz, 1 H), 4.99 (aq, J = 2.1 Hz, 1 H), 7.38-7.46 (m, 6 H), 7.66-7.71 (m, 4 H); ¹³C NMR (125 MHz, CD₃OD) & 20.5 (C), 27.9 (CH₃ × 3), 31.3 (CH₂), 31.6 (CH₂), 32.1 (CH₂), 36.6 (CH₂), 39.9 (CH₂), 40.0 (CH₂), 41.8 (CH₂), 48.8 (CH₂), 52.6 (CH₃), 62.4 (CH₂), 70.1 (CH), 75.7 (CH), 76.3 (CH), 76.5 (CH), 78.0 (CH), 78.5 (CH), 79.0 (CH), 79.8 (CH), 82.8 (CH), 84.2 (CH), 105.8 (CH_2), 112.1 (C), 129.3 (CH \times 4), 131.31 (CH), 131.33 (CH), 135.5 (C), 135.6 (C), 137.18 (CH × 2), 137.20 $(CH \times 2)$, 153.8 (C), 173.4 (C); HRMS calculated for $C_{40}H_{52}O_9$ -Si (M + Na)⁺ 727.3278, found 727.3281 (0.41 ppm).

Aldehyde 11. Alkene 73 (60 mg, 0.093 mmol for synthesis of enone 74; 28.5 mg, 0.0462 mmol for synthesis of enone 10) was dissolved in CH_2Cl_2 (3 mL) and cooled to -78 °C. Ozone was bubbled through the reaction mixture for 5 min, and then the mixture was purged with N₂ for 3 min. Triphenylphosphine (24 mg, 1.5 equiv for synthesis of enone 74; 12 mg, 1.5 equiv for synthesis of enone 74; 12 mg, 1.5 equiv for synthesis of enone 74; 12 mg, 1.5 equiv for synthesis of enone 10) was added to the reaction mixture at -78 °C, and then, the bath was removed and reaction mixture was stirred at room temperature for 30 min. The contents of the flask were concentrated under a stream of N₂ and then were put under high vacuum for 1 h. The resulting aldehyde was used without further purification.

 α , β -Unsaturated Enone 74. To a suspension of dry NaH (8.7 mg, 0.3627 mmol, 3.9 equiv), in DME (1 mL) was added dimethyl-2-oxoheptyl phosphonate (0.077 mL, 0.372 mmol, 4 equiv). The reaction mixture bubbled slightly, and then went clear. This was stirred at room temperature for 5 min with gentle heating by a heating mantle. Crude aldehyde 11 (based on 60 mg, or 0.093 mmol of alkene 73) from the previous reaction was dissolved in DME (1 mL) and transferred via cannula with an additional 1 mL of DME to assist in the transfer. The reaction mixture was then heated at reflux for 3 h. After this time, the reaction was cooled and quenched by addition of water, and the reaction mixture was extracted with ether. The combined organic extracts were dried over MgSO₄, filtered, and concentrated in vacuo. FCC (9% Et₂O in hexanes) yielded enone **74** as an oil (44.2 mg, 64%). Data for **74**: R_f 0.62 (50% hexanes in Et₂O); $[\alpha]^{23}_{D}$ –34.9 (c 0.00235, CHCl₃); IR (thin film) 2954, 2876, 1743, 1676, 1630, 1081 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 0.57 (q, J = 7.7 Hz, 6 H), 0.65 (q, J

= 8.1 Hz, 6 H), 0.66 (q, J = 8.2 Hz, 6 H), 0.92 (t, J = 8.0 Hz, 9 H), 0.98 (t, J = 8.1 Hz, 9 H), 0.99 (t, J = 7.9 Hz, 9 H), 1.21-1.34 (m, 9 H), 1.61-1.68 (m, 3 H), 1.80-1.85 (m, 1 H), 2.35 $(ABX, J_{ab} = 15.3 \text{ Hz}, J_{ax} = 8.4 \text{ Hz}, 1 \text{ H}), 2.50 (ABX, J_{ab} = 15.3 \text{ Hz})$ Hz, $J_{bx} = 4.9 Hz$, 1 H), 2.56 (dt, J = 7.4, 5.8 Hz, 2 H), 2.91 (dd, J = 9.6, 2.2 Hz, 1 H), 3.38 (app dt, J = 9.9, 4.1 Hz, 1 H), 3.68 (s, 3 H), 3.81 (dd, J = 7.1, 2.5 Hz, 2 H), 3.94 (dd, J = 6.9, 3.5Hz, 1 H), 4.00 (m, 1 H), 5.07 (dd, J = 7.3, 3.0 Hz, 1 H), 6.28 (dd, J = 16.1, 0.8 Hz, 1 H), 7.09 (dd, J = 16.2, 7.5 Hz, 1 H); 13 C NMR (75 MHz, CDCl₃) δ 4.7 (CH₂ × 3), 5.1 (CH₂ × 3), 5.3 $(CH_2 \times 3)$, 6.9 $(CH_3 \times 3)$, 6.99 $(CH_3 \times 3)$, 7.04 $(CH_3 \times 3)$, 13.9 $(CH_3), 22.5 (CH_2), 24.1 (CH_2), 28.7 (CH_2), 30.5 (CH_2), 31.5$ (CH₂), 39.8 (CH₂), 40.5 (CH₂), 51.6 (CH₃), 64.6 (CH), 69.9 (CH), 71.5 (CH₂), 72.2 (CH), 74.0 (CH), 78.7 (CH), 82.3 (CH), 130.3 (CH), 146.1 (CH), 171.7 (C), 200.8 (C); HRMS calculated for $C_{38}H_{74}O_8Si_3(M + Na)^+$ 765.4589, found 765.4626 (5 ppm).

ABCDE Cage Ketal 6, Route B. To a solution of enone 74 (35 mg, 0.047 mmol) in a mixture of CH_2Cl_2 /water (0.5 mL/ 0.25 mL) was added *p*-TsOH (8.96 mg, 0.047 mmol, 1 equiv), and this was stirred at room temperature for 30 min, followed by the addition of MeOH (0.25 mL). The reaction mixture was stirred for 5 min and was then quenched by addition of saturated aqueous NaHCO₃ (ca. 2 mL), followed by extraction with CH_2Cl_2 . The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated in vacuo. FCC (10% to 20% to 50% hexanes in EtOAc to 100% EtOAc) yielded desired ABCDE ketal 6 (8.5 mg, 47%) as an oil. Data for 6 are given above.

 α , β -Unsaturated Enone 10. To a cooled (0 °C) orangecolored solution of dry KHMDS (32 mg, 0.160 mmol, 3.6 equiv) in DME (1 mL) was added β -ketophosphonate **12b** (99.7 mg, 0.183 mmol, 4.1 equiv, dissolved in 1 mL of DME) via cannula. An additional 1 mL of DME was used to assist in the transfer. This mixture stirred at 0 °C for 30 min. In another flask, aldehyde 11 (based on 28.5 mg, or 0.0462 mmol of alkene 73) was dissolved in DME (0.5 mL) and transferred via cannula to the reaction flask. An additional 0.5 mL of DME was used to assist in the transfer. This was stirred at 0 °C for 10 min, warmed to room temperature, and then heated at reflux for 10 h. The reaction was then cooled and quenched with addition of water and extracted with EtOAc $(4 \times)$ and $CH_2Cl_2(2 \times)$. The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated in vacuo. The desired enone could not be separated from an impurity that coelutes, so FCC (10% to 20% Et₂O in hexanes followed by 50% EtOAc in hexanes) gave a mixture of enone 10 plus the unknown impurity (~35% pure), which was carried on without further purification.

ABCDEF Cage Ketal 7, Route B. To a solution of crude enone **10** from above in CH_2Cl_2 (2 mL) and 5 drops of water was added *p*-TsOH (4.8 mg, 0.025 mmol, 1 equiv), and the reaction mixture was stirred for 5 min. MeOH (2 mL) was added, and the reaction mixture was stirred at room temperature for 2 h. The reaction was quenched by addition of saturated aqueous NaHCO₃, followed by extraction with CH_2 - Cl_2 (4×). The combined organic extracts were dried over Na₂-SO₄, filtered, and concentrated in vacuo. FCC (50% EtOAc in hexanes) yielded desired ketal **7** (3.4 mg, 10% from **73**). Data for **7** are given above.

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Supporting Information Available: Description of general procedures, additional experimental data, ¹H and ¹³C NMR spectra for all new compounds, and crystallographic

information for **40** and **70**. This material is available free of charge via the Internet at http://pubs.acs.org. JO051479M