

Total Syntheses of Bengamides B and E

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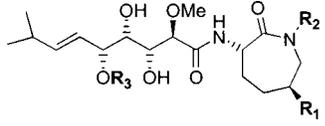
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Total syntheses of the cytotoxic marine natural products bengamides B and E are described. Both bengamides are prepared via amide coupling of a protected polyhydroxylated lactone intermediate **9** with a suitably substituted aminocaprolactam intermediate. Lactone **9** is prepared in five steps from commercially available α -D-glucoheptonic γ -lactone. The key reactions are a selective deprotection of a 1,2-acetonide in the presence of a 1,3-acetonide and an (*E*)-selective olefination of an unstable aldehyde using a *gem*-dichromium reagent. The bengamide B lactam intermediate **10** is prepared in seven steps from commercially available (*5R*)-5-hydroxy-L-lysine (**12**). The desired *S*-configuration at the γ -OH lactam position is established using the Mitsunobu reaction.

Introduction

The bengamides are marine natural products first isolated by Crews et al. from Jaspidae sponges indigenous to the coral reefs that surround the Fiji islands.¹ These cyclolysine derivatives are potent antiproliferative agents against both transformed and nontransformed cells.² Bengamides that possess a lipophilic ester substituent on the caprolactam (e.g., bengamides A, B, M, and O) inhibit tumor cell growth at 10–100 nM (IC₅₀ range) and are >100-fold more potent than their nonlactam ester-bearing counterparts (e.g., bengamides E, F, P, and Z).³ We recently reported that bengamides B, E, and Z significantly inhibit MDA-MB-435 human breast carcinoma cells implanted as xenografts in athymic mice at well-tolerated doses.^{2a} The mechanism of action of the bengamides is under investigation. To continue profiling the antitumor properties of the bengamides, additional supplies of these natural products were needed. Since a continued supply of bengamides from sponge extraction was not a practical option, we required an efficient synthesis capable of producing gram quantities of these compounds. Herein we report concise total syntheses of bengamides B and E that meet this need. Several syntheses of bengamides A, B, and E have been previously published.⁴ However, all of these syntheses are long (14–15 steps for bengamide E and 30–32 steps for bengamide B) which render them impractical for prepa-

Table 1. Representative Bengamides Isolated from Jaspidae Sponges



bengamide	R ₁	R ₂	R ₃
A (1)	O ₂ C(CH ₂) ₁₂ CH ₃	H	H
B (2)	O ₂ C(CH ₂) ₁₂ CH ₃	Me	H
E (3)	H	H	H
F (4)	H	Me	H
M (5)	O ₂ C(CH ₂) ₁₁ CH(CH ₃) ₂	Me	H
O (6)	O ₂ C(CH ₂) ₁₀ CH(CH ₃) ₂	Me	H
P (7)	H	H	O ₂ C(CH ₂) ₁₂ CH ₃
Z (8)	OH	Me	H

ration of grams of material.⁵ The present synthesis relies on the coupling of two key intermediates: lactone **9** and aminocaprolactam **10**. Although every reported synthesis takes advantage of a similar coupling strategy, the syntheses of intermediates corresponding to **9** and **10** require a prohibitive number of protection/deprotection steps. The utilization of two commercially available natural products, α -D-glucoheptonic γ -lactone (**11**) and (*5R*)-5-hydroxy-L-lysine (**12**), makes a much shorter bengamide synthesis possible. Lactone **11** contains the same

(1) (a) Quiñoà, E.; Adamczeski, M.; Crews, P.; Bakus G. J. *J. Org. Chem.* **1986**, *51*, 4494. (b) Adamczeski, M.; Quiñoà, E.; Crews, P. *J. Am. Chem. Soc.* **1989**, *111*, 647. (c) Adamczeski, M.; Quiñoà, E.; Crews, P. *J. Org. Chem.* **1990**, *55*, 240.

(2) (a) Kinder, F. R.; Bair, K. W.; Bontempo, J.; Crews, P.; Czuchta, A. M.; Nemzek, R. Thale, Z.; Vattay, A.; Versace, R. W.; Weltchek, S.; Wood, A.; Zabudoff, S. D.; Phillips, P. E. *Proc. Am. Assoc. Cancer Res.* **2000**, *41*, 600. (b) Phillips, P. E.; Bair, K. W.; Bontempo, J.; Crews, P.; Czuchta, A. M.; Kinder, F. R.; Vattay, A.; Versace, R. W.; Wang, B.; Wang, J.; Wood, A. *Proc. Am. Assoc. Cancer Res.* **2000**, *41*, 59.

(3) (a) Bengamide B was evaluated in the NCI 60 cell line screen. The data can be obtained at the NCI website (<http://dtp.nci.nih.gov/>). (b) Thale, Z.; Kinder, F. R.; Bair, K. W.; Czuchta, A. M.; Versace, R. W.; Phillips, P. E.; Sanders, M. L.; Wattanasin, S.; Crews, P., unpublished results.

(4) (a) Chida, N.; Ogawa, N. *ChemComm.* **1997**, 807. (b) Mukai, C.; Hanaoka, M. *Synlett* **1996**, 11. (c) Mukai, C.; Moharram, S. M.; Kataoka, O.; Hanaoka, M. *J. Chem. Soc., Perkin Trans. 1* **1995**, 2849. (d) Mukai, C.; Kataoka, O.; Hanaoka, M.; *J. Org. Chem.* **1995**, *60*, 5910. (e) Chida, N.; Tobe, T.; Murai, K.; Yamazaki, K.; Ogawa, S. *Heterocycles* **1994**, *38*, 2383. (f) Mukai, C.; Kataoka, O.; Hanaoka, M. *Tetrahedron Lett.* **1994**, *35*, 6899. (g) Marshall, J. A.; Luke, G. P. *J. Org. Chem.* **1993**, *58*, 6229. (h) Marshall, J. A.; Luke, G. P. *Synlett* **1992**, 1007. (i) Chida, N.; Tobe, T.; Okada, S.; Ogawa, S. *J. Chem. Soc., Chem. Commun.* **1992**, 1064. (j) Kishimoto, H.; Ohru, H.; Meguro, H. *J. Org. Chem.* **1992**, *57*, 5042. (k) Broka, C. A.; Ehrler, J. *Tetrahedron Lett.* **1991**, *32*, 5907. (l) Chida, N.; Tobe, T.; Ogawa, S. *Tetrahedron Lett.* **1991**, *32*, 1063.

(5) A 10-step synthesis of bengamide E was recently reported at the 219th National ACS meeting: see Clark, T. J.; Boeckman, R. K.; Jr. *Book of Abstracts*; 219th American Chemical Society National Meeting, San Francisco, CA, Mar 26–31, 2000, Abstr ORGN-58.

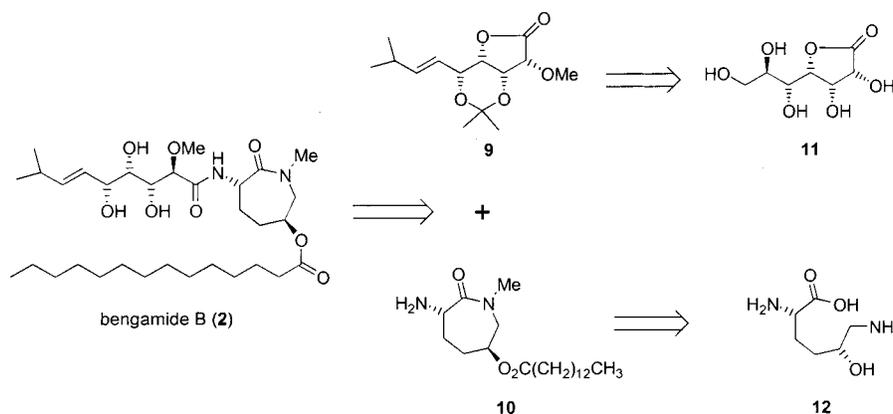
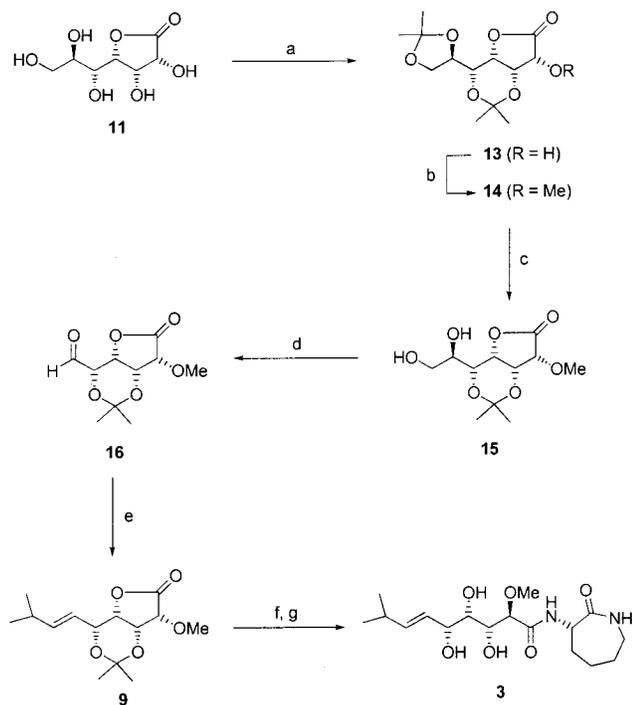


Figure 1. Retrosynthesis of bengamide B.

Scheme 1. Synthesis of Bengamide E (3)^a



^a Reagents: (a) acetone, cat. I_2 (86%); (b) MeI, Ag_2O (82%); (c) $HOAc$ (68%); (d) $NaIO_4$, MeOH (85%); (e) $(CH_3)_2CHI_2$, $CrCl_2$, THF, DMF (29%); (f) L-(-)- α -amino- ϵ -caprolactam, *i*-PrOH, reflux (85%); (g) TFA, THF, H_2O (54%).

sequence of chiral hydroxy groups present on the polyhydroxylated portion common to the bengamide class. Hydroxylysine **12** contains all the functionality required to prepare bengamides that bear a caprolactam oxygen substituent. However, **12** has the opposite configuration at the γ -OH position which made an inversion reaction necessary. The retrosynthetic analysis of bengamide B in Figure 1 is representative of the present bengamide synthesis strategy.

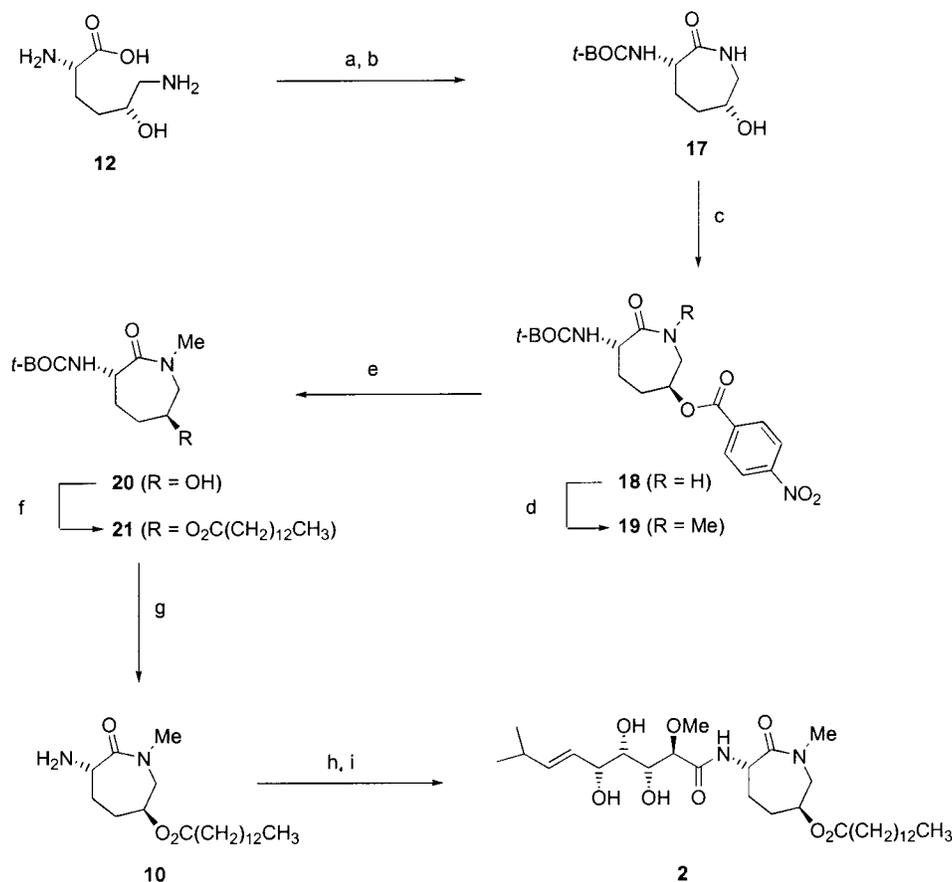
Discussion

The synthesis of bengamide E (**3**) is outlined in Scheme 1. α -D-Glucopyranose (**11**) was converted to the bis(acetonide) **13** by treatment of an acetone solution of **11** with catalytic I_2 .⁶ The reaction also produced approximately 10% of another bis(acetonide) that could be removed through crystallization. Methylation of the remaining unprotected hydroxyl group using Ag_2O with

methyl iodide provided methyl ether **14** in 82% yield.⁷ The key step in the transformation of **11** to the advanced intermediate **9** was the selective removal of the 1,2-acetonide in the presence of the 1,3-acetonide. This was accomplished by the treatment of bis(acetonide) **14** with acetic acid. The desired vicinal diol **15** was produced in 68% yield.⁸ Every step up to this point did not require chromatographic purification. Oxidative cleavage of the diol with $NaIO_4$ produced aldehyde **16**. This aldehyde was prone to hydration and required repeated rotary evaporation with $CHCl_3$ or toluene to remove all traces of water. Aldehyde **16** was then olefinated with the low-valent organochromium species generated in situ from 1,1-diodoisobutane (prepared in two steps from isobutyraldehyde⁹) and $Cr(II)Cl_2$ to produce a nearly 3:1 mixture of *E* and *Z* isomers in 39% yield.¹⁰ The desired *E* isomer, **9**, was isolated in 29% yield using preparative normal phase HPLC. None of aldehyde **16** was recoverable from the reaction mixture. Alternative olefination attempts including the Wittig reaction and the S. Julia olefination¹¹ produced <10% of the olefin **9**. Two steps remained in order to complete the synthesis of bengamide E. Commercially available L-(-)- α -amino- ϵ -caprolactam and lactone **9** were stirred at reflux in *i*-PrOH to give the protected bengamide E adduct in 85% yield. Finally, TFA-promoted acetonide hydrolysis produced bengamide E in 54% yield after reverse phase HPLC purification.

Once a feasible route to lactone **9** was established, the remaining challenge to produce bengamide B resided in the preparation of the substituted aminocaprolactam **10** (Scheme 2). Cyclization of (5*R*)-5-hydroxy-L-lysine (**12**) followed by *N*-Boc carbamoylation of the free amine was performed in one pot using standard peptide synthesis conditions (58% yield for two steps). The secondary alcohol was inverted using Mitsunobu conditions to give a 78% yield of *p*- NO_2 -benzoate **18** in the desired *S*-configuration.¹² Selective methylation of the lactam nitrogen with MeI/ $NaHMDS$ produced lactam intermediate **19** in 83% yield. LiOH-promoted ester hydrolysis of **19** generated the corresponding alcohol **20** in 97% yield.

- (6) Kartha, K. P. R. *Tetrahedron Lett.* **1986**, 27, 3415.
 (7) Gurjar, M. K.; Srinivas, N. R. *Tetrahedron Lett.* **1991**, 32, 3409.
 (8) Shing, T. K. M.; Zhou, Z. H.; Mak, C. W. *J. Chem. Soc., Perkin Trans. 1* **1992**, 1907.
 (9) Friedrich, E. C.; Falling, S. N.; Lyons, D. E. *Synth. Commun.* **1975**, 33.
 (10) Okazoe, T.; Takai, K.; Utimoto, K. *J. Am. Chem. Soc.* **1987**, 109, 951.
 (11) (a) Baudin, J. B.; Hareau, G.; Julia, S. A.; Ruel, O. *Tetrahedron Lett.* **1991**, 32, 1175. (b) Blakemore, P. R.; Cole, W. J.; Kocienski, P. J.; Morley, A. *Synlett* **1998**, 26.

Scheme 2. Synthesis of Bengamide B (**2**)^a

^a Reagents: (a) EDCI, HOBT, DMF; (b) (Boc)₂O (58% for two steps); (c) DEAD, PPh₃, *p*-nitrobenzoic acid (78%); (d) MeI, NaHMDS (83%); (e) LiOH (97%); (f) EDCI, DMAP, myristic acid (94%); (g) TFA, (99%); (h) **9**, *i*-PrOH, 80 °C; (i) TFA, THF, H₂O (61% for two steps).

Esterification of **20** with myristic acid in the presence of EDCI and DMAP produced the bengamide B lactam intermediate **21** in 94% yield. Treatment of **21** with TFA removed the BOC protecting group. Neutralization (NH₄-OH) of the crude TFA salt and chromatographic purification gave the amine **10** in 99% yield. Condensation of lactone **9** with aminoprolactam **10** in refluxing *i*-PrOH provided the corresponding amide without competing amine attack on the acyclic ester moiety. Subsequent acetone hydrolysis with TFA generated bengamide B (**2**) in 61% yield (two steps).

In conclusion, a synthetic route has been established that is capable of providing gram amounts of two members of the bengamide natural product class starting from two commercially available natural products. The benefits of the present synthesis are 2-fold: 1) multigram quantities of these relatively scarce natural products facilitate further biological testing in animal models, and 2) several series of bengamide analogues can be prepared through modification of both the lactone and aminoprolactam intermediates.¹³

Experimental Section

General Methods. All chemicals were obtained from commercial suppliers and used without further purification.

(12) (a) Mitsunobu, O. *Synthesis* **1981**, 1. (b) The Mitsunobu reaction was not attempted with myristic acid. *p*-NO₂-benzoate ester **18** was a convenient intermediate for the synthesis of bengamide B analogues.

(13) The synthesis and antitumor activity of bengamide analogues will be published elsewhere.

Flash column chromatography was performed with silica (Merck EM9385, 230–400 mesh). ¹H and ¹³C NMR spectra were recorded at 500 or 300 and at 125 and 75 MHz, respectively, in CDCl₃ unless otherwise mentioned. Proton and carbon chemical shifts are expressed in ppm relative to internal tetramethylsilane, coupling constants (*J*) are expressed in hertz.

Bengamide B (2). A solution consisting of **9** (1.0 g, 3.7 mmol), **10** (1.2 g, 3.4 mmol), and *i*-PrOH (5 mL) was stirred at 80 °C for 16 h. The *i*-PrOH was removed on a rotary evaporator, and the crude product was chromatographed (2% MeOH/CH₂Cl₂) to yield 1.82 g (83%) of acetone which was used directly in the next step. To the acetone (precooled in a 0 °C bath) was added a solution of 6 mL of TFA, 6 mL of THF, and 4 mL of H₂O precooled to ca. 0 °C. The reaction was stirred at 0 °C for 20 min. The solvents were removed using a rotary evaporator under high vacuum (<5 Torr) at 0 °C. The residue was dissolved in 20 mL of MeOH and neutralized with the dropwise addition of concentrated NH₄OH at 0 °C. The solution was evaporated to dryness and chromatographed (2% MeOH/CH₂Cl₂). The product was further purified by preparative reverse phase HPLC (C₁₈ column, isocratic 20% CH₃CN/H₂O) to yield 1.25 g (73%) of waxy white solid: α_D²⁰ +63.6° (c 2.9, CHCl₃) (lit.^{1a} α_D²⁰ +34.6° (c 0.075, MeOH)); ¹H NMR δ 8.11 (d, *J* = 6.2; 1H), 5.79 (ddd, *J* = 15.5, 6.5, 0.9, 1H), 5.46 (ddd, *J* = 15.6, 7.3, 1.3, 1H), 4.65 (m, 2H), 4.28 (s, 1H), 4.22 (t, *J* = 6.3, 1H), 3.80 (m, 2H), 3.67 (dd, *J* = 14.6, 10.1, 1H), 3.60 (s, 1H), 3.55 (s, 3H), 3.22 (m, 2H), 3.11 (s, 3H), 3.07 (s, 1H), 2.31 (t, *J* = 7.4, 2H), 2.15 (m, 2H), 1.97 (m, 1H), 1.63 (m, 4H), 1.26 (m, 20H), 1.00 (dd, *J* = 6.8, 2.7, 6H), 0.88 (t, *J* = 6.8, 3H); ¹³C NMR 173.02, 172.15, 171.78, 141.48, 125.40, 80.83, 74.32, 72.86, 72.33, 69.17, 60.06, 53.36, 51.33, 36.40, 34.35, 32.68, 31.92, 30.80, 29.68, 29.65, 29.60, 29.45, 29.35, 29.24, 29.10, 29.00, 24.78, 22.69, 22.22, 22.11, 14.12. Anal. Calcd for

$C_{32}H_{58}N_2O_8$: C, 64.19; H, 9.76; N, 4.68. Found: C, 63.91; H, 9.87; N, 4.50.

Bengamide E (3). A solution of **9** (98 mg, 0.37 mmol), L-(α -amino- ϵ -caprolactam (142 mg, 1.11 mmol), and *i*-PrOH (2 mL) was stirred at reflux for 18 h. The *i*-PrOH was removed on a rotary evaporator, and the crude product was chromatographed (3% MeOH/CH₂Cl₂) to yield 123.8 mg (85%) of acetone. To 150 mg of this acetone (precooled to 0 °C) was added a solution of TFA (3 mL), THF (3 mL), and H₂O (2 mL) precooled to 0 °C. The reaction was stirred at 0 °C for 20 min. The solvents were removed using a rotary evaporator under high vacuum (<5 Torr) at 0 °C. The residue was dissolved in MeOH (10 mL) and neutralized with concentrated NH₄OH at 0 °C. The solvents were removed using a rotary evaporator and chromatographed (3% MeOH/CH₂Cl₂). The product was further purified by preparative reverse phase HPLC (C₁₈ column, isocratic 30% CH₃CN/H₂O) to yield 73 mg (54%) of **3** as a white solid: $\alpha^{20}_D +57.9^\circ$ (c 0.75, CH₂Cl₂) (lit.^{1b} $\alpha^{20}_D +36.9^\circ$ (c 0.043, MeOH)); ¹H NMR δ 7.91 (d, *J* = 6.5, 1H), 6.17 (t, *J* = 5.7, 1H), 5.72 (ddd, *J* = 15.5, 6.5, and 0.8, 1H), 5.39 (ddd, *J* = 15.6, 7.3, and 1.4, 1H), 4.48 (ddd, *J* = 11.2, 6.5, and 1.3, 1H), 4.16 (t, *J* = 6.2, 1H), 3.76 (dd, *J* = 6.9 and 1.3, 1H), 3.72 (d, *J* = 6.9, 1H), 3.54 (d, *J* = 4.6, 1H), 3.47 (s, 3H), 3.22 (m, 2H), 2.25 (m, 1H), 1.99 (m, 2H), 1.78 (m, 2H), 1.51 (m, 1H), 1.36 (m, 1H), 0.94 (d, *J* = 2.5, 3H), 0.93 (d, *J* = 2.7, 3H); ¹³C NMR δ 172.18, 169.47, 139.22, 122.79, 78.42, 71.64, 70.14, 69.82, 57.31, 49.41, 39.50, 28.42, 28.20, 26.23, 25.34, 19.52. Anal. Calcd for C₁₇H₃₀N₂O₆: C, 56.97; H, 8.44; N, 7.82. Found: C, 56.61; H, 8.09; N, 7.59.

(6E)-6,7,8,9-Tetradecoxy-8-methyl-2-O-methyl-3,5-O-(1-methylethylidene)-gulonon-6-enoic Acid Lactone (9). **15** (100 g, 0.382 mol) was dissolved into a 1:1 mixture of methanol/H₂O (2 L). The stirred mixture was cooled in an ice-water bath to about 8 °C. Solid NaIO₄ (98 g, 0.458 mol) was added portionwise. The reaction was complete within 40 min as indicated by TLC (silica gel, 5% MeOH/15% EtOAc/CH₂Cl₂). Solid NaCl was added into the reaction mixture to saturate the methanolic solution. The solid was filtered and washed with 2 L CH₂Cl₂. The filtrate was extracted with 3 \times 200 mL CH₂Cl₂. The combined organic layers were dried over Na₂SO₄, filtered, and concentrated to a syrup, which formed a precipitate upon addition of hexane. The solid was filtered and rinsed with Et₂O. The crude product was chromatographed (3% MeOH/CH₂Cl₂) to give 75 g (85%) **16** as a white solid. This solid readily hydrated on standing. Prior to use in the next step, **16** was dissolved in CHCl₃ then concentrated in vacuo several times. Mp 125–127 °C; ¹H NMR δ 9.62 (s, 1H), 4.78 (t, *J* = 6, 1H), 4.46 (s, 2H), 4.15 (d, *J* = 6, 1H), 3.67 (s, 3H), 1.59 (s, 3H), 1.57 (s, 3H); ¹³C NMR δ 198.8, 171.9, 99.0, 78.4, 74.4, 72.9, 68.4, 67.4, 59.2, 28.7, 19.0. To a 1 L round-bottom flask, under N₂, was added anhydrous THF (300 mL) and anhydrous DMF (13 mL). Anhydrous CrCl₂ (20 g, 0.16 mol) was added and stirred for 1 h. A solution of **16** (4.6 g 20 mmol) and 1,1-diiodo-2-methylpropane (12.4 g 40 mmol) in THF (200 mL) and DMF (4 mL) was added over 5 min. The resulting slurry was stirred for 1.5 h then quenched with a 100 mL solution of saturated aq NH₄Cl containing 2 g Na₂S₂O₃ and 10 g NH₄HCO₃ followed by solid NH₄HCO₃ (30 g). The resulting slurry was filtered through a bed of Na₂SO₄ and silica gel and evaporated to give an oil. The oil was chromatographed on silica gel (25% EtOAc/CH₂Cl₂) to give 2.1 g (39%) of a mixture of **9** and the corresponding *Z* isomer. The mixture was purified by preparative normal phase HPLC (silica column, isocratic 50% EtOAc/hexane) to give 1.54 g (28.5%) of **9** as a white solid: $[\alpha]^{25}_D -132.7^\circ$ (c 1.036 CHCl₃); ¹H NMR δ 5.85 (dd, *J* = 15.6 and 6.2, 1H), 5.64 (ddd, *J* = 15.6, 7.5, and 1.3, 1H), 4.74 (dd, *J* = 3.8 and 2.1, 1H), 4.48 (dd, *J* = 7.5 and 1.8, 1H), 4.12 (d, *J* = 3.9, 1H), 4.02 (t, *J* = 2.0, 1H), 3.68 (s, 3H), 2.36 (m, 1H), 1.56 (s, 3H), 1.51 (s, 3H), 1.04 (d, *J* = 1.9, 3H), 1.03 (d, *J* = 1.9, 3H); ¹³C NMR δ 172.8, 143.2, 122.0, 98.7, 79.0, 71.7, 70.0, 67.6, 59.2, 30.7, 29.2, 21.9, 21.8, 19.2. HRMS calcd for C₁₄H₂₂O₅Na (M+Na)⁺ 293.1365, found 293.1355.

(3S,6S)-3-Amino-6-hydroxy-6-(tridecylcarbonyloxy)-2H-azepin-2-one (10). A solution of **20** (3.6 g, 7.7 mmol), TFA (5 mL), and CH₂Cl₂ (10 mL) was stirred at 25 °C for 2 h. The

reaction mixture was concentrated in vacuo. The residue was dissolved in 100 mL of CH₃CN and neutralized with NH₄OH. The solution was concentrated using a rotary evaporator and chromatographed (2.5% MeOH/CH₂Cl₂) to yield 2.8 g (99%) of **10** as a white solid. ¹H NMR δ 5.69 (s, 2H), 4.61 (t, *J* = 9.4, 1H), 4.13 (d, *J* = 10.9, 1H), 3.64 (dd, *J* = 10.2 and 14.7, 1H), 3.20 (d, *J* = 14.7, 1H), 3.06 (s, 3H), 2.30 (t, *J* = 7.5, 2H), 2.14 (m, 2H), 1.83 (m, 2H), 1.61 (t, *J* = 6.8, 2H), 1.26 (m, 20H), 0.88 (t, *J* = 6.8, 3H); ¹³C NMR 179.87, 179.52, 76.06, 60.03, 59.68, 43.32, 41.26, 39.27, 38.85, 36.57, 36.37, 35.39, 31.77, 29.61, 21.06. HRMS calcd. for C₂₁H₄₀N₂O₃Na (M + Na)⁺ 391.2936, found 391.2926.

3,5,6,7-Bis-O-(1-methylethylidene)- α -D-glucoheptonic γ -Lactone (13). α -D-Glucoheptonic γ -lactone (**11**) (500 g, 2.4 mol) was added into 9 L of acetone in a 5 gal plastic drum. The mixture was agitated mechanically until most of the solid dissolved (15–20 min). Iodine (60 g, 0.236 mol) was added portionwise to the lactone solution over 5–10 min. The resulting mixture was stirred overnight. A saturated solution of Na₂S₂O₃ (1.3 L) was added to the iodine solution to quench the reaction. The resulting solution was concentrated to about half of its original volume in vacuo, and brine solution (5 L) was added. The resulting mixture was extracted with 3 \times 1.2 L EtOAc. All organic layers were combined and evaporated to dryness. The solid was slurried with a mixture of ether and hexane (3:7), and filtered. The filter cake was washed with Et₂O (50 mL) and air-dried to produce 599 g of the desired compound as a white powder (86.5%): mp 150–152 °C (lit.¹⁴ mp 153–154 °C), ¹H NMR δ 4.62 (m, 1H), 4.50 (m, 1H), 4.35 (m, 2H), 4.07 (m, 1H), 3.93 (m, 1H), 3.82 (dd, *J* = 4.0 and 8.9, 1H), 3.08 (dd, *J* = 2.0 and 8.5, 1H), 1.51 (s, 3H), 1.44 (s, 3H), 1.39 (s, 3H), 1.35 (s, 3H); ¹³C NMR δ 174.4, 109.4, 98.6, 72.8, 71.4, 69.3, 68.4, 67.8, 66.7, 28.6, 26.7, 24.6, 19.3.

2-O-Methyl-3,5,6,7-bis-O-(1-methylethylidene)- α -D-glucoheptonic γ -Lactone (14). **13** (719 g, 2.49 mol) was added into 4.5 L of CH₂Cl₂ in a 5 gal plastic drum. The mixture was stirred under N₂. Iodomethane (2500 g, 17.6 mol) was added immediately followed by addition of silver(I) oxide (1750 g, 7.58 mol). H₂O (30 mL) was added to the reaction mixture. An ice bath was used to maintain the reaction temperature at 15–30 °C. The reaction was stirred in the absence of light for 18 h. After diluting the reaction mixture with 1.5 L of CH₂Cl₂, the solid was filtered and washed with an additional 2.2 L of CH₂Cl₂. The undesired solid was discarded, and the filtrate was evaporated to dryness. The residue was slurried in Et₂O (1.5 L), filtered, and dried to give 618 g of product (82%): mp 139–141 °C; ¹H NMR δ 4.70 (dd, *J* = 5 and 6, 1H), 4.35 (m, 1H); 4.27 (s, 1H) 1.35 (s, 3H), 4.13 (d, *J* = 5, 1H), 4.10 (d, *J* = 6, 1 H), 3.92 (dd, *J* = 4.5 and 9, 1H), 3.82 (dd, *J* = 2 and 9, 1H), 3.65 (s, 3H), 1.48 (s, 3H), 1.42 (s, 6 H); ¹³C NMR δ 172.5, 109.6, 98.5, 79.0, 73.1, 69.5, 68.6, 67.5, 66.9, 59.1, 28.9, 26.9, 24.9, 19.4. HRMS calcd for C₁₄H₂₂O₇Na (M + Na)⁺ 325.1258, found 325.1264.

2-O-Methyl-3,5-O-(1-methylethylidene)- α -D-glucoheptonic γ -Lactone (15). **14** (618 g, 2.05 mol) was dissolved in 8 L of a mixture of HOAc and H₂O (1:1) over 30 min. The solution was stirred at ambient temperature overnight. The solution was evaporated to dryness in vacuo. The solid was slurried in 3–5 L of hot acetone and filtered. After oven drying at 20–30 °C, 363 g of the desired compound was obtained (67.6%): mp 177–178 °C; ¹H NMR δ 4.92 (bt, 1H), 4.80 (dd, *J* = 2 and 4, 1H), 4.50 (d, *J* = 5.5, 1H), 4.42 (bd, 1H), 4.38 (s, 1H), 3.95 (dd, *J* = 2 and 9, 1H), 3.55 (bdd, *J* = 2 and 9, 2H), 3.42 (s, 3H), 3.38 (m, 1H), 1.42 (s, 3H), 1.22 (s, 3H); ¹³C NMR δ 173.3, 97.5, 78.3, 68.8, 68.5, 67.2, 67.1, 62.2, 57.4, 28.9, 19.2. Anal. Calcd for C₁₁H₁₈O₇: C, 50.38; H, 6.92. Found: C, 50.14; H, 6.91.

(3S,6R)-3-(tert-Butoxycarbonyl)amino-6-hydroxy-2H-azepin-2-one (17). (5R)-5-Hydroxy-L-lysine (**12**) (10 g, 0.040 mol), 1-hydroxybenzotriazole hydrate (8.2 g, 0.060 mol), and 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide-HCl (EDCI) (11.6 g, 0.060 mol) were added sequentially to 500 mL of DMF with stirring. After 0.5 h, Et₃N (16.8 mL, 0.120

mol) was added. The reaction was stirred at 25 °C for 48 h. Di-*tert*-butyl dicarbonate (17.6 g, 0.080 mol) and Et₃N (16.8 mL, 0.120 mol) were then sequentially added. Stirring was continued for 16 h. The reaction mixture was filtered to remove Et₃N-HCl, and the solvent was removed by rotary evaporation under high vacuum to give a thick oil. The oil was dissolved in 150 mL of CH₂Cl₂ and applied to a silica gel column (150 g, 40 × 250 mm). The column was eluted with 3% MeOH/CH₂-Cl₂ to give the crude product as a solid. The crude solid was dissolved in 120 mL of CH₂Cl₂ with heating and then cooled to -20 °C for 1 h. The resulting solid was filtered and washed with 50 mL of CH₂Cl₂. The combined filtrates were evaporated to dryness. CH₂Cl₂ (40 mL) was added to this residue, and the resulting slurry was stirred for 0.5 h at 25 °C. The slurry was filtered and the solid washed with 25 mL of CH₂Cl₂. The solids were combined to give 5.57 g (58%) of **17**: ¹H NMR (*d*₆-DMSO) δ 7.42 (t, *J* = 5.1, 1H), 6.38 (d, *J* = 6.6, 1H), 4.60 (d, *J* = 4.2, 1H), 4.07 (m, 1H), 3.74 (m, 1H), 3.32 (m, 1H), 3.03 (m, 1H), 1.8–1.5 (m, 4H), 1.39 (s, 9H); ¹³C NMR (*d*₆-DMSO) δ 173.8, 154.5, 77.9, 63.7, 52.4, 45.1, 34.2, 28.1, 24.9. Anal. Calcd for C₁₁H₂₀N₂O₄: C, 54.08; H, 8.25; N, 11.47. Found: C, 54.06; H, 8.11; N, 11.41.

(3S,6S)-3-(tert-Butoxycarbonyl)aminohexahydro-6-(4-nitrophenylcarbonyl)oxy-2H-azepin-2-one (18). To a 500 mL flask were added **17** (6 g, 0.025 mol), 4-nitrobenzoic acid (8.25 g, 0.05 mol), and triphenylphosphine (12.9 g, 0.05 mol). The flask was purged with N₂, and 200 mL of freshly distilled THF (Na metal) was added. The mixture was cooled in a -20 °C bath, and diethyl azodicarboxylate (7.8 mL, 0.05 mol) was added at a rate to maintain ≤10 °C. The reaction was allowed to warm to room temperature (~2 h) and stirred for 14 h. The solvent was evaporated, and the residue dissolved in 200 mL of EtOAc. This was washed with 5% NaHCO₃ (3 × 150 mL) and brine (1 × 150 mL) and then dried with Na₂SO₄ and evaporated to give an oil. To this oil was added 100 mL of ether. The resulting solid was filtered and washed with ether (3 × 50 mL), acetone (2 × 50 mL), and MeOH (2 × 50 mL) to give 6.2 g of **18**. The combined filtrate and washes were evaporated, and the residue was chromatographed on a silica gel column with CH₂Cl₂:hexane (1:1) and then ether to give 1.34 g of **18** for a combined yield of 7.54 g (78%): ¹H NMR δ 8.29 (d, *J* = 8.4, 1H), 8.18 (d, *J* = 8.4, 2H), 6.44 (m, 1H), 5.90 (d, *J* = 5.7, 1H), 4.91 (m, 1H), 4.42 (m, 1H), 3.58–3.41 (m, 2H), 2.36–2.25 (m, 2H), 2.16–2.03 (m, 1H), 1.82–1.65 (m, 1H), 1.45 (s, 9H); ¹³C NMR δ 174.9, 163.6, 155.2, 150.8, 134.9, 130.8, 123.6, 79.8, 72.8, 52.8, 45.0, 33.0, 29.8, 28.4. Anal. Calcd for C₁₈H₂₃N₃O₇: C, 54.96; H, 5.86; N, 10.68. Found: 54.97; H, 5.88; N, 10.60.

(3S,6S)-1-Methyl-3-(tert-Butoxycarbonyl)aminohexahydro-6-(4-nitrophenylcarbonyl)oxy-2H-azepin-2-one (19). To a solution of **17** (5.0 g, 12.7 mmol) and THF (50 mL) at -7 °C was added KHMDS (15.3 mL, 15.3 mmol, 1 M solution in THF) within 5 min. After the mixture was stirred at -78 °C for 30 min, MeI (3.6 g, 25 mmol) was added dropwise to the reaction mixture. The reaction was stirred at 25 °C for 18 h.

The reaction was quenched with 50 mL of H₂O. The mixture was extracted with 2 × 100 mL of EtOAc. The organic layers were dried over Na₂SO₄, concentrated, and chromatographed (40% EtOAc/hexane) to yield 4.3 g (83.0%) of **19** as a white solid: ¹H NMR δ 8.31 (d, *J* = 9.1, 2H), 8.20 (d, *J* = 8.7, 2H), 6.00 (d, *J* = 5.7, 1H), 4.88 (t, *J* = 10.6, 1H), 4.50 (dd, *J* = 6.0 and 10.2, 1H), 3.82 (dd, *J* = 9.8 and 17.7, 1H), 3.38 (d, *J* = 14.7, 1H), 3.15 (s, 3H), 2.21 (m, 3H), 1.68 (m, 1H), 1.46 (s, 9H); ¹³C NMR δ 172.94, 164.16, 155.59, 151.18, 135.37, 131.19, 124.05, 80.06, 71.59, 53.66, 52.99, 36.76, 33.15, 30.38, 28.78. HRMS calcd for C₁₉H₂₅N₃O₇Na (M + Na)⁺ 430.1585, found 430.1597.

(3S,6S)-1-Methyl-3-(tert-butoxycarbonyl)aminohexahydro-6-hydroxy-2H-azepin-2-one (20). To a solution of **19** (4.1 g, 9.8 mmol), MeOH (40 mL), and H₂O (10 mL) was added LiOH/H₂O (0.83 g, 20 mmol) at 25 °C. After being stirred at this temperature for 1h, the reaction was quenched with dry ice (CO₂) and evaporated to dryness. To the residue was added a solution consisting of CH₂Cl₂ (45 mL) and MeOH (5 mL). The solution was filtered through Celite. The filtrate was evaporated to dryness and chromatographed (10% EtOAc/CH₂-Cl₂) to yield 2.52 g (97.0%) of **20** as a white solid: ¹H NMR δ 6.00 (d, *J* = 5.6, 1H), 4.38 (dd, *J* = 6.4 and 10.2, 1H), 3.61 (m, 2H), 3.21 (dd, *J* = 1.9 and 12.4, 1H), 3.06 (s, 3H), 2.66 (d, *J* = 4.5, 1H), 2.21 (d, *J* = 12.4, 1H), 2.08 (d, *J* = 14.3, 1H), 1.82 (m, 1H), 1.60 (td, *J* = 2.3 and 13.9, 1H), 1.44 (s, 9H); ¹³C NMR δ 174.53, 157.04, 81.46, 69.80, 58.37, 54.63, 38.78, 38.41, 32.14, 30.21. HRMS calcd for C₁₂H₂₂N₂O₄Na (M+Na)⁺ 281.1472, found 281.1475.

(3S,6S)-1-Methyl-3-(tert-butoxycarbonyl)aminohexahydro-6-(tridecylcarbonyl)oxy-2H-azepin-2-one (21). To a solution of myristic acid (2.97 g, 13.0 mmol) and CH₂Cl₂ (50 mL) were added EDCI (2.62 g, 13.65 mmol) and DMAP (1.67 g, 13.65 mmol) at 25 °C. After being stirred at 25 °C for 30 min, a solution of **19** (2.4 g, 9.3 mmol) and CH₂Cl₂ (50 mL) was added portionwise. The reaction was stirred for 16 h at 25 °C. After being stirred, the reaction was quenched with 50 mL of H₂O. The mixture was extracted with EtOAc (2 × 100 mL). The organic layers were dried (Na₂SO₄), concentrated, and chromatographed (2.5% EtOAc/CH₂Cl₂) to yield 4.1 g (94.2%) of **21** as a white solid: ¹H NMR δ 5.97 (d, *J* = 5.6, 1H), 4.60 (tt, *J* = 3.4 and 10.6, 1H), 4.43 (dd, *J* = 6.0 and 10.6, 1H), 3.63 (dd, *J* = 10.2 and 14.7, 1H), 3.20 (d, *J* = 14.7, 1H), 3.24 (s, 3H), 2.30 (t, *J* = 7.5, 2H), 2.13 (d, *J* = 10.6, 2H), 1.92 (m, 1H), 1.61 (m, 3H), 1.45 (s, 9H), 0.93 (m, 20H), 0.88 (t, *J* = 6.4, 3H); ¹³C NMR (CDCl₃) δ 173.42, 172.98, 155.58, 79.94, 69.75, 53.79, 53.00, 36.68, 34.73, 33.23, 32.30, 30.49, 30.02, 29.98, 29.83, 29.73, 29.62, 29.47, 28.78, 25.25, 23.07, 14.50. HRMS calcd for C₂₆H₄₈N₂O₅Na (M + Na)⁺ 491.3455, found 491.3470.

Supporting Information Available: Copies of ¹H and ¹³C NMR spectra of **9**, **14**, **19**, **20**, and **21** are available free of charge via the Internet at <http://pubs.acs.org>.

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