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Total syntheses of (-)-emestrin H and (-)-asteroxepin

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1. Introduction

ABSTRACT

First total syntheses of (–)-emestrin H and (–)-asteroxepin are described. To find the appropriate protecting group on the amide nitrogen of the diketopiperazine core, we conducted model studies using a simple diketopiperazine derivative. As a result, allyloxymethyl (Allom) group was the most suitable protecting group, which tolerated Nicolaou's sulfenylation conditions, and was easily cleavable under the mild conditions using Pd(PPh₃)₄ and *N*,*N*-dimethylbarbituric acid leaving methylthioethers intact. The general utility of Allom group for protection of amides was studied using simple substrates. Finally, the effectiveness of Allom group was robust enough during installation of two methylthioethers to the diketopiperazine core and easily removed at the final step. The first total synthesis of (–)-asteroxepin was also completed by acylation of (–)-emestrin H.

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Dithiodiketopiperazine alkaloids bearing the sulfur-containing diketopiperazine as a core structure have attracted attention due to their structural diversity and fascinating biological activities [1,2]. Among them, compounds bearing the 4,5-dihydrooxepine ring, such as (-)-acetylaranotin (1) [3], (+)-MPC1001B (7) [4], and (-)-acetylapoaranotin (6) [5], have been challenging synthetic targets and a lot of effort has been made to synthesize these compounds [6,7]. Reisman et al. reported the first innovative total synthesis of (-)-acetylaranotin (1) in 2012 [8], followed by the first total synthesis of (-)-acetylapoaranotin (6) [10]. We also focused on this class of compounds, reporting the second total synthesis of 1 just after Reisman et al. in 2012 [9]. Using the synthetic protocol established during the synthetic studies on 1, which was based on the characteristic proline-fused 4,5dihydrooxepine ring, we successfully performed the first total synthesis of (+)-MPC1001B (7) [11] and determined the structure of (-)-SCH64874 (3) and hirsutellomycin (8) via semi-synthesis [12]. Despite these synthetic developments over the last decade, the total syntheses of (-)-asteroxepin (4) [13] and (-)-emestrin H (5) [14] bearing the NH-free diketopiperazine core have not been reported so far.

Our preliminary model studies toward 4 and 5 revealed that the enolate-mediated introduction of two methylthioethers reported by Nicolaou *et al.* [15] was unsuccessful for the unprotected diketopiperazine 13 due to the competitive deprotonation of the unprotected N–H proton (Scheme 1b). Therefore, the major challenge toward the synthesis of 4 and 5 was the selection of the appropriate protecting group for the

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diketopiperazine ring's NH group. The ideal protecting group (PG in Scheme 1a) should be robust enough to tolerate the strong basic conditions during the introduction of the twomethylthioethers. Moreover, it should be easily and chemoselectively cleaved under the particularly mild conditions in the final stage of the total synthesis without affecting the base-, nucleophile-, and oxidant-sensitive methylthioethers and the acid-sensitive dihydrooxepin structure. However, compared to the many available protecting groups for amines, there are limited choices for amides, especially for diketopiperazines. In



Fig. 1. Dithiodiketopiperazine alkaloids

Tetrahedron

functionalized NH-free diketopiperazines and found that the less common allyloxymethyl (Allom) group, which only have been used for a protection of the N-3 position in xanthine and the imidazole nitrogen in histidine was the most suitable protecting group [19]. Its efficiency was further demonstrated by the first total synthesis of (–)-asteroxepin (4) and (–)-emestrin H (5).



Scheme 1. Synthetic plan and preliminary results.

2. Results and discussion

To identify the most suitable protecting group that would fulfill the abovementioned requirements, we prepared a series of *N*-protected diketopiperazines (16a-e) following a conventional method [16,17], which were then subjected to the sulfenylation conditions established by Nicolaou *et al.* (Table 1) [15]. Among the protecting groups in substrates 16a-e, stable benzylic protecting groups, such as *p*-methoxybenzyl (PMB) in 16a or 2,4-dimethoxybenzyl (2,4-DMB) in 16b, were not affected by the sulfenylation conditions, providing dithiodiketopiperazines 17aand 17b, respectively, in moderate yields (Table 1, entries 1

Table 1

Protecting groups and introduction of the methylthioether.



^aC3 (*S*), C8a (*S*) isomer. ^bC3 (*S*), C8a (*R*) isomer. ^cIsolated yield. ^dObtained as single diastereomer.

16d with the trimethylsilylethoxymethyl (SEM) group and **16e** with the Allom group, afforded the corresponding dimethylthio compounds (**17d** and **17e**) in good yields (Table 1, entries 4 and 5). However, analogue **16c**, protected as urethane with an allyloxycarbonyl group (Alloc), afforded a complex mixture under the same sulfenylation conditions (Table 1, entry 3).

The deprotection of the bis(methylthio) products 17a, 17b, 17d, and 17e was examined under the most frequently used deprotection conditions for each protecting group (Table 2). Analogues 17a and 17b bearing PMB and 2,4-DMB groups were decomposed upon treatment with ceric ammonium nitrate (CAN) 2,3-dichloro-5,6-dicyano-1,4-benzoquinone and (DDO). respectively (Table 2, entries 1 and 2). Moreover, the SEM group in 17d was stable upon treatment with HF/pyridine and 17d was recovered (Table 2, entry 3), whereas the use of TBAF in THF/DMF decomposed 17d (Table 2, entry 4). In contrast to these unsuccessful entries, the Allom group in 17e could be smoothly removed by a combination of $Pd(PPh_3)_4$ and N,Ndimethylbarbituric acid (DMBA) in MeOH yielding 15 in 56% yield, while a substantial amount of the hemiaminal intermediate 18 was also obtained (Table 2, entry 5) [18,19]. The subsequent in situ treatment of the reaction mixture with aq. NH₃ was effective in reducing the amount of 18, affording 15 in 87% yield (Scheme 2). Furthermore, the addition of solid NaHCO₃ after the consumption of 17e improved the yield of 15 to up to 95%.

Table 2

Deprotection conditions.

<pre>N</pre>		$ \begin{array}{c} \mathbf{PG} \\ \mathbf{PG} \\ \mathbf{Ph} \end{array} \xrightarrow{\text{conditions}} \begin{array}{c} MeS \\ \mathbf{Ph} \\ Me \end{array} $		Ph MeS O	N OH Ph SMe
	17		15	18 (from	17f)
entry	17	reagents	temp.	Time (h) Yie	eld (%) ^a
1^{b}	17a	CAN, CH ₃ CN/H ₂ O	rt	2	0
2 ^b	17b	DDQ, CH ₃ CN/H ₂ O	60 °C	20	0
3 ^b	17d	TBAF, THF/DMF	60 °C	1.8	0
4 ^c	17d	HF/pyridine	60 °C	22	0
5 ^d	17e	Pd(PPh ₃) ₄ , DMBA, MeOH	30 °C	6	56

^aisolated yield. ^bDecomposition. ^c**17d** was completely recovered. ^dReaction conditions: **17** (1.0 eq), Pd(PPh₃)₄ (45 mol%), DMBA (5.5 eq), MeOH (0.1 M), 30 °C, 6 h.°The hemiaminal intermediate **18** was obtained in 38% yield.



Scheme 2. Removal of the Allom group.

The feasibility of the removal of the Allom protecting group from secondary amides was investigated using a series of *N*-Allom secondary amides (**20**) (see Experimental Section for the preparation and additional data) (Table 3 and Scheme 3). The treatment of the *N*-Allom benzamides **20a–e** and γ lactam **20f** with Pd(PPh₃)₄ and DMBA in MeOH smoothly provided the hemiaminal intermediate **21**, which was converted into amides **19a–e** and **19f**, respectively, in good yields using aq. NH₃ (Table 3, entries 1–6). In contrast, the reaction of *N*-Allom aliphatic amides **20g** under the Pd/*N*,*N*- **21g** (21%), and *N*-MOM amides **22g** (39%) due to the exchange of the OH group in intermediate **21g** with the methoxy group of methanol (Scheme 3). This side reaction was overcome by performing the deprotection of **20g** in THF and H₂O, providing the hemiaminal intermediates **21g**, which were then converted to amides **19g** in good yields using trimethylsilyl iodide (TMSI).

Table 3.

One-pot Removal of the Allom group.



Reaction conditions: **20** (1.0 eq), $Pd(PPh_3)_4$ (10 mol%), DMBA (2.0 eq), MeOH (0.3 M), 40 °C, 6 h, then aq. NH₃ (excess). The yields refer to the isolated products.





The applicability of the Allom protecting group was fully demonstrated by the first total syntheses of (–)-emestrin H (5) and (–)-asteroxepin (4). The proline-fused dihydrooxepine derivative 9, which was prepared according to our previously established protocol in the total synthesis of (–)-acetylaranotin (1) [9], was first condensed with the (S)-phenylalanine methyl

then removed under transfer hydrogenation conditions and the formation of the tricyclic diketopiperazine derivative **25** was promoted upon treatment of the resulting aminoester with ammonium hydroxide. Subsequently, the Allom group was introduced using a combination of the allyloxymethyl chloride (AllomCl) **26** [20] and lithium hexamethyldisilazide (LiHMDS), which afforded the desired product **27** in 84% yield. After desilylation, the stereochemistry of the C-10 hydroxyl group was inverted through a stepwise process [9], which included an oxidation of **28** to ketone using PhI(OAc)₂ and 9-azanoradamantane *N*-oxyl (nor-AZADO) [21] and a subsequent Luche reduction, which gave the desired alcohol **30** as the sole isomer.



Scheme 4. Preparation of diketopiperazine 30.

The final steps toward the synthesis of (–)-emestrin H (**5**) and (–)-asteroxepin (**4**) including the crucial introduction of the two methylthioethers and the removal of the Allom protecting group are depicted in Scheme 5. According to the model study (*vide supra*), diketopiperazine **30** was treated with a mixture of excess LiHMDS and S₈ to form an epipolysulfide. Its reduction to the corresponding dithiol with NaBH₄ and the subsequent methylation afforded the bis(methylthio) derivative **31** in 65% yield over two steps from **30**. Finally, the Allom group was removed under the optimized conditions affording (–)-emestrin H (**5**) in 78% yield, which was further acetylated to give (–)-asteroxepin (**4**). All properties of synthetic (–)-emestrin H (**5**) and (–)-asteroxepin (**4**) were identical with those reported except magnitude of specific rotations proving the reported absolute stereochemistry of **4** and **5** by this total synthesis [22].



Scheme 5. Final steps of the total syntheses of (–)-emestrin H (5) and (–)-asteroxepin (4).

4 3.

In conclusion, the total syntheses of (–)-emestrin H (**5**) and (–)-asteroxepin (**4**) have been accomplished for the first time. Their preparation was successful owing to the suitability of the Allom group as protecting group for the amide nitrogen of the highly functionalized dithio-diketopiperazine intermediate. This protecting group could be easily cleaved even in the presence of the acid-sensitive dihydrooxepin structure and the

the acid-sensitive dihydrooxepin structure and the dimethylthioethers, which are sensitive to bases, nucleophiles, and oxidants. Thus, this study demonstrated the application potential of the Allom group not only for the synthesis of other *N*-unprotected diketopiperazine alkaloids, but also for the preparation of various functionalized amides.

4. Experimental section

4.1 General method

Materials were obtained from commercial suppliers and used without further purification unless otherwise mentioned. Anhydrous THF, Et₂O, CH₂Cl₂, toluene, DMF, and MeCN were purchased from commercial suppliers. Anhydrous MeOH was dried and distilled according to the standard protocols. All reactions were carried out under Ar atmosphere unless otherwise mentioned. Flash column chromatography was performed on Silica Gel 60N (Kanto, spherical neutral, 40-50 µm). Preparative TLC and analytical TLC was performed on Merck 60 F254 glass plates precoated with a 0.25 mm thickness of silica gel. NMR spectra were recorded on a JNM-AL400 spectrometer and a JEOL ECA600 spectrometer. Chemical shifts for ¹H NMR are reported in parts per million (ppm) downfield from tetramethylsilane as the internal standard and coupling constants are in Hertz (Hz). The following abbreviations are used for spin multiplicity: s = singlet, d = doublet, t = triplet, q = quartet, m =multiplet, and br = broad. Chemical shifts for ${}^{13}C$ NMR are reported in ppm, relative to the central line of a triplet at 77.0 ppm for CDCl₃ or a septet at 39.5 ppm for DMSO-d₆. IR spectra were measured on a SHIMADZU FTIR-8300 spectrometer. Mass spectra were recorded on a Bruker micrOTOF (ESI). Optical rotations were measured on a Horiba SEPA-300 highly sensitive polarimeter. Melting point determinations were performed by using a Yanaco MP-500 instrument. Compounds 9 [9], 19a-e [25], 19f [26], and 19g [25] were prepared according to the procedures reported in references.

4.2 Diketopiperazine16a

To a solution of N-Cbz-L-proline (3.68 g, 14.8mmol) and N-PMB-L-phenylalanine methyl ester (4.24 g, 14.2 mmol) in Et₃N and CH₂Cl₂ (5 mL and 30 mL) was added BOP-Cl (3.74 g, 14.7 mmol) at room temperature. After stirring for 24.5 h at room temperature, additional BOP-Cl (1.96 g, 7.70 mmol) was added, and the reaction mixture was stirred for 2.5 h. The reaction was quenched with sat. aq. NaHCO3, and the aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic layers were washed with 1 M aq. HCl, dried over Na2SO4, filtered and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 1:1) to afford amide 32a (5.80 g, 10.9 mmol, 77%). To a suspension of amide 32a (5.80 g, 10.9 mmol) and Pd(OAc)₂ (983 mg, 4.38 mmol) in CH₂Cl₂ (55.0 mL) was added Et₃N (3.08 mL, 21.9 mmol) at room temperature. After stirring at reflux for 5 min, Et₃SiH (8.74 mL, 54.7 mmol) was added and the mixture was stirred at reflux for 3.5 h. The reaction mixture was diluted with CH₂Cl₂ and the reaction was quenched with sat. aq. NaHCO₃. The separated aqueous phase was extracted with anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (EtOAc) to afford diketopiperazine **16a** (3.00 g, 8.23 mmol, 75%). A white foam; The spectral data of **16a** were identical with those reported in the literature [23].

4.3 Diketopiperazine16b

To a solution of N-Cbz-L-proline (1.06 g, 4.25 mmol), N-2,4-DMB-L-phenylalanine methyl ester (1.15 g, 3.49 mmol) in Et₃N and CH₂Cl₂ (2.0 mL and 20.0 mL) was added BOP-Cl (1.87 g, 7.35 mmol) at room temperature. After stirring for 20 h at room temperature, the reaction was quenched with 1 M aq. HCl and the resulting mixture was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes: EtOAc = 3:2 to 6:5) to afford amide 32b (846 mg, 1.51 mmol, 47%). To a suspension of amide 32b (846 mg, 1.51 mmol) and Pd(OAc)₂ (135 mg, 601 µmol) in CH₂Cl₂ (10.0 mL) was added Et₃N (425 µL, 3.02 mmol) at room temperature. After stirring at reflux for 7 min, Et₃SiH (1.20 mL, 7.51 mmol) was added and the mixture was stirred at reflux for 3.5 h. The reaction mixture was diluted with CH₂Cl₂ and the reaction was quenched with sat. aq. NaHCO3. The separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude which was purified by silica gel column material, chromatography (EtOAc) to afford diketopiperazine 16b (540 mg, 1.37 mmol, 91%). A white foam; ¹H NMR (400 MHz, CDCl₃): δ 7.31-7.20 (4H, m), 7.12-7.06 (2H, m), 6.48-6.43 (2H, m), 5.40 (1H, d, J = 14.8 Hz), 4.32 (1H, m), 4.25 (1H, d, J =14.8 Hz), 3.84 (3H, s), 3.81 (3H, s), 3.72 (1H, dd, J = 12.0, 6.0 Hz), 3.68-3.61 (1H, m), 3.36-3.27 (2H, m), 3.14-3.07 (1H, m), 1.91–1.81 (1H, m), 1.60–1.42 (2H, m), 0.14–0.01 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 165.3, 163.9, 160.7, 158.7, 135.1, 131.7, 130.0, 128.3, 127.1, 115.9, 104.4, 98.3, 60.3, 59.0, 55.3, 55.3, 44.0, 40.6, 36.4, 28.4, 20.9; IR (film): 2992, 2952, 2944, 2891, 1656, 1612, 1587, 1508, 1456, 1298, 1289, 1261, 1208, 1184, 1157, 1033, 751, 703 cm⁻¹; $[\alpha]_D^{31} = -116$ (c = 0.66, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₃H₂₆N₂NaO₄ [M⁺+Na] 417.1785, Found 417.1769.

4.4 Amide 33

To a solution of N-Boc-L-proline (10.8 g, 50.0 mmol) and Lphenylalanine methyl ester (8.95 g, 50.0 mmol) in CH₂Cl₂ (200 mL) were added Et₃N (27.9 mL, 200 mmol), HOBt (6.99 g, 51.7 mmol), and EDCI/HCl (9.91 g, 51.7 mmol) at room temperature. After stirred for 29.5 h at room temperature, the organic phase was washed with 1 M aq. HCl (x3) and sat. aq. NaHCO₃ (x2). The resulting organic phase was dried over Na₂SO₄, filtered and concentrated under reduced pressure to give a crude material, which was used for the next reaction without further purification. To a solution of the crude material in CH₂Cl₂ (120 mL) was added TFA (15 mL) at room temperature. After stirred for 20 h at room temperature, the reaction mixture was diluted with CH₂Cl₂ (150 mL) and neutralized by sat. aq. NaHCO₃. The aqueous phase was extracted with $CH_2Cl_2(x3)$, and the combined organic layers were dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (CH_2Cl_2 :MeOH = 10:1) to afford amine amide 33 (6.76 g, 24.5 mmol, 49%, 2 steps from N-Boc-L-

with those reported in the literature [24].

4.5 Diketopiperazine trans-13

To a solution of amide 33 (6.76 g, 24.5 mmol) in MeOH (400 mL) was added NaHCO3 (20.0 g, 85.3 mmol) at room temperature. After stirring for 22.5 h at 65 °C, MeOH was removed under reduced pressure. The residue was diluted with CH₂Cl₂ and filtered through a pad of Celite[®]. The filtrate was washed with H₂O, and the aqueous phase was extracted with CH₂Cl₂ (x2). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by crystallization from EtOAc to afford diketopiperazine trans-13 (3.43 g, 14.1 mmol, 57%). The mother liquor was concentrated in vacuo and the second crop was crystallized from EtOAc to afford diketopiperazine trans- $1\overline{3}$ (643 mg, 2.64 mmol, 11%). A white solid.; fmp: 153.5–154.2 °C; $\ ^1H$ NMR (400 MHz, CDCl_3): δ 7.35–7.20 (5H, m), 6.31 (1H, s), 4.22 (1H, ddd, J = 6.6, 4.0, 3.8Hz), 3.63 (1H, ddd, J = 12.4, 8.5, 8.5 Hz), 3.40 (1H, ddd, J = 12.4, 9.4, 3.0 Hz), 3.15 (1H, dd, J = 13.9, 6.6 Hz), 3.08 (1H, dd, *J* = 13.9, 4.0 Hz), 2.97 (1H, dd, *J* = 10.4, 6.4 Hz), 2.23–2.15 (1H, m), 1.98–1.90 (1H, m), 1.86–1.64 (2H, m); ¹³C NMR (100 MHz, CDCl₃): δ 169.5, 164.8, 135.3, 129.9, 128.5, 127.3, 58.7, 57.6, 44.9, 40.3, 28.8, 21.5; IR (film): 3483, 3238, 3063, 3029, 2981, 2953, 2931, 2886, 1664, 1496, 1454, 1336, 1307, 1296, 1206, 1186, 1115, 1106, 921, 732, 702, 593 cm⁻¹; $[\alpha]_D^{26} = +93.0$ (c = 0.200, H₂O); HRMS (ESI) m/z: calcd. for $C_{14}H_{17}N_2O_2$ [M+H⁺] 245.1285, found 245.1277. The spectral data of trans-13 were identical with those reported in the literature [27].

4.6 Diketopiperazine16c

To a solution of diketopiperazine trans-13 (203 mg, 0.832 mmol) in THF (4.0 mL) was added LiHMDS (1.3 M solution in THF, 650 µL, 845 µmol) at 0 °C. After stirring for 4 min, Alloc-OSu (172 mg, 863 µmol) in THF (1.50 mL) was added at 0 °C. The resulting mixture was stirred for 35 min at room temperature. The reaction was quenched with sat. aq. NH₄Cl, and the resulting mixture was diluted by water. The separated aqueous phase was extracted with CH₂Cl₂ (x3), and the combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material. which was purified by silica gel column chromatography (CH₂Cl₂:EtOAc = 5:1 to 3:1) to afford diketopiperazine 16c (248 mg, 789 µmol, 95%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.31–7.26 (3H, m), 7.19–7.14 (2H, m), 5.99–5.87 (1H, m), 5.43 (1H, d, J = 17.2 Hz), 5.31 (1H, d, J = 10.8 Hz), 5.01 (1H, ddd, J = 5.0, 5.0, 1.4 Hz), 4.75 (1H, ddd, *J* = 13.3, 5.8, 1.0 Hz), 4.69 (1H, ddd, *J* = 13.3, 5.6, 1.2 Hz), 3.60-3.50 (1H, m), 3.45-3.37 (1H, m), 3.33-3.22 (2H, m), 2.64 (1H, dd, J = 9.6, 6.8 Hz), 2.17-2.07 (1H, m), 1.96-1.79 (2H, m),1.72–1.57 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 167.2, 164.0, 151.8, 134.9, 130.7, 129.9, 128.6, 127.5, 119.4, 67.9, 62.3, 58.6, 44.8, 38.2, 29.1, 21.8; IR (film): 3086, 3062, 3028, 2983, 2953, 2885, 1782, 1731, 1672, 1455, 1384, 1270, 1231, 980, 763, 747, 703 cm⁻¹; $[\alpha]_D^{26} = +143$ (*c* = 0.425, CHCl₃); HRMS (ESI) *m/z*: calcd. for C₁₈H₂₀N₂NaO₄ [M+Na⁺] 351.1315, Found 351.1319.

4.7 Diketopiperazine16d

To a solution of diketopiperazine *trans*-**13** (1.10 g, 4.51 mmol) in dry DMF (10.0 mL) was added NaH (60% dispersion in mineral oil, 248 mg, 6.20 mmol) at room temperature. After stirring for 15 min, SEMC1 (1.60 mL, 9.03 mmol) was added at room temperature, and the resulting mixture was stirred at 60 °C for 18 h. The reaction was quenched with sat. aq. NH₄Cl at room

and washed with water (x5) and brine (x1). The organic phase was dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 1:3) to afford diketopiperazine 16d (1.33 g, 3.55 mmol, 79%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.29–7.26 (3H, m), 7.18–7.15 (2H, m), 5.13 (1H, d, J = 10.4 Hz), 4.53 (1H, d, J = 10.4 Hz), 4.42 (1H, dd, J = 5.2, 4.9 Hz), 3.61-3.52 (3H, m), 3.35 (1H, ddd, J = 12.1, 9.3, 2.9 Hz), 3.24 (1H, dd, J = 14.3, 5.2 Hz), 3.19 (1H, dd, J = 14.3, 4.9 Hz), 2.52 (1H, dd, J= 10.6, 6.6 Hz), 2.14-2.06 (1H, m), 1.92-1.84 (1H, m), 1.79-1.68 (1H, m), 1.69-1.53 (1H, m), 1.01-0.87 (2H, m), 0.01 (9H, s); ¹³C NMR (100 MHz, CDCl₃): δ 168.3, 164.7, 135.2, 129.9, 128.5, 127.5, 73.3, 66.3, 62.1, 57.6, 44.8, 37.6, 29.1, 21.8, 17.9, -1.5; IR (film): 2952, 2890, 1671, 1454, 1440, 1296, 1260, 1249, 1207, 1075, 1044, 860, 837, 749, 703 cm⁻¹; $[\alpha]_D^{25} = +55.0$ (c = 2.71, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₀H₃₀NaN₂O₃Si [M⁺+Na] 397.1918, Found 397.1909.

4.8 Diketopiperazine16e

To a solution of diketopiperazine trans-13 (880 mg, 3.61 mmol) in dry THF (8.0 mL) was added NaH (60% dispersion in mineral oil, 181 mg, 4.53 mmol) at room temperature. After stirring for 50 min, AllomCl (26) [20] (750 µL, 7.23 mmol) was added at room temperature, and the mixture was stirred for 9.5 h at 50 °C. The reaction was quenched with sat. aq. NH4Cl solution at room temperature, and the aqueous phase was diluted by water, extracted with Et₂O (x1) and CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 2:1, then EtOAc only) to afford diketopiperazine 16e (946 mg, 3.01 mmol, 83%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.32–7.24 (3H, m), 7.19–7.14 (2H, m), 5.89 (1H, m), 5.29 (1H, ddd, J = 17.6, 1.2,1.2, Hz), 5.19 (1H, d, J = 10.8 Hz), 5.14 (1H, d, J = 10.6 Hz), 4.58 (1H, d, J = 10.6 Hz), 4.43 (1H, dd, J = 4.7, 4.8 Hz), 4.10-3.98 (2H, m), 3.57 (1H, ddd, J = 12.3, 8.5, 8.5 Hz), 3.36 (1H, ddd, J = 12.3, 9.3, 2.9 Hz), 3.25 (1H, dd, J = 14.3, 4.8 Hz), 3.20 (1H, dd, J = 14.3, 4.7 Hz), 2.53 (1H, dd, J = 10.6, 6.6 Hz), 2.09 (1H, ddd, J = 12.0, 6.2, 6.2 Hz), 1.94-1.84 (1H, m), 1.81-1.69(1H, m), 1.66–1.53 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 168.4, 164.7, 135.3, 133.8, 129.9, 128.6, 127.5, 117.4, 73.6, 70.1, 62.4, 57.7, 44.9, 37.7, 29.0, 21.8; IR (film): 3028, 2979, 2951, 2883, 1668, 1453, 1440, 1344, 1296, 1261, 1207, 1072, 1057, 923, 748, 703 cm⁻¹; $[\alpha]_D^{26} = +60.0$ (*c* = 4.41, CHCl₃); HRMS (ESI) m/z: calcd. for C₁₈H₂₂N₂NaO₃ [M⁺+Na] 337.1523, Found 337.1506.

4.9 Dithiodiketopiperazine 17a (General Procedure A)

To a suspension of sulfur S_8 (264 mg, 1.03 mmol) in THF (5.0 mL) was added NaHMDS (0.6 M in toluene, 5.00 mL, 3.00 mmol) at room temperature. After stirring for 1 min, diketopiperazine **16a** (372 mg, 1.02 mmol) in THF (5.0 mL) was added, and the mixture was stirred for 1 min. Then, NaHMDS (0.6 M in toluene, 3.40 mL, 2.04 mmol) was added, and the mixture was stirred for 30 min at room temperature. The reaction was quenched with sat. aq. NH₄Cl, and the separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was used for the next reaction without further purification. To a solution of the crude material in THF (10.0 mL) and EtOH (10.0 mL) was added NaBH₄ (964 mg, 25.5 mmol) at 0 °C. After

sti added at 0 °C, and the resulting mixture was stirred for 19 h at room temperature. The reaction was quenched by sat. aq. NH₄Cl, and the separated aqueous phase was extracted with CH_2Cl_2 (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes:EtOAc = 12:5) to afford bis(metylthio) diketopiperazine 17a (204 mg, 0.449 mmol, 44%, 2 steps from **16a**). A white solid; mp: 136.7–137.0 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.53 (2H, d, J = 8.8 Hz), 7.31–7.22 (3H, m), 7.09 (2H, d, J = 6.0 Hz), 6.83 (2H, d, J = 8.8 Hz), 5.18 (1H, d, J = 14.4Hz), 4.54 (1H, d, J = 14.4 Hz), 3.79 (3H, s), 3.66–3.48 (2H, m), 3.54 (1H, d, J = 13.6 Hz), 3.25 (1H, d, J = 13.6 Hz), 2.05-1.91 (2H, m), 2.01 (3H, s), 1.99 (3H, s), 1.58-1.49 (1H, m), 0.81-0.71 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 165.9, 163.0, 158.6, 134.1, 130.2, 129.6, 128.4, 127.7, 113.3, 78.8, 69.9, 55.1, 46.7, 45.8, 44.1, 32.7, 18.5, 15.0, 14.0 (One signal is missing due to overlap.), IR (film): 2994, 2956, 2933, 2922, 1662, 1512, 1421, 1395, 1352, 1247, 1177, 1033, 756, 703 cm⁻¹; $[\alpha]_D^{25} = +0.89$ (c 0.58, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₄H₂₈N₂NaO₃S₂ [M+Na⁺] 479.1419, Found 479.1434.

4.10 Dithiodiketopiperazine 17b

According to the General Procedure A, diketopiperazine 16b (301 mg, 0.763 mmol) were converted to bis-(methylthio) diketopiperazine 17b (210 mg, 0.431 mmol, 57 %, 2 steps from 16b). A white solid; mp: 186.3-186.7 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.39 (1H, d, J = 8.5 Hz), 7.30–7.20 (3H, m), 7.11 (2H, d, J = 6.4 Hz), 6.48 (1H, d, J = 2.1 Hz), 6.41 (1H, d, J = 8.5, 2.1 Hz), 5.07 (1H, d, J = 15.8 Hz), 4.68 (1H, d, J = 15.8 Hz), 3.90 (3H, s), 3.79 (3H, s), 3.67–3.53 (2H, m), 3.55 (1H, d, J = 13.6 Hz), 3.33 (1H, d, J = 13.6 Hz), 2.14 (3H, s), 2.14 (3H, s), 2.10-1.94 (2H, m), 1.61–1.50 (1H, m), 0.86–0.73 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 165.7, 162.9, 159.5, 157.2, 134.2, 130.1, 128.3, 127.5, 127.0, 118.5, 103.5, 98.2, 78.5, 69.7, 55.3, 55.2, 45.8, 44.1, 41.1, 32.5, 18.4, 15.2, 14.2; IR (film): 2998, 2959, 2936, 2921, 2835, 1662, 1616, 1590, 1508, 1455, 1439, 1420, 1396, 1361, 1300, 1261, 1208, 1156, 1120, 1036, 755, 703 cm⁻¹; $[\alpha]_D^{32} = +21.3$ (c = 0.26, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₅H₃₀N₂NaO₄S₂ [M+Na⁺] 509.1539, Found 509.1526.

4.11 Dithiodiketopiperazine 17d

According to the General Procedure A, diketopiperazine 16e (456 mg, 1.22 mmol) were converted to bis-(methylthio) diketopiperazine 17d (386 mg, 0.828 mmol, 68%, 2 steps from 16d). A white solid; mp: 82.9-83.3 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.25–7.19 (3H, m), 7.12 (2H, dd, *J* = 7.8, 2.2 Hz), 5.48 (1H, d, J = 9.6 Hz), 5.06 (1H, d, J = 9.6 Hz), 3.79 (2H, t, J = 8.6 Hz), 3.50 (2H, dd, J = 9.2, 6.4 Hz), 3.46 (1H, d, J = 13.6 Hz), 3.18 (1H, d, J = 13.2 Hz), 2.26 (3H, s), 2.14 (3H, s), 2.12-2.04 (1H, m), 2.02-1.89 (1H, m), 1.55-1.46 (1H, m), 1.07-0.96 (2H, m), 0.93–0.84 (1H, m), 0.03 (9H, s); ¹³C NMR (100 MHz, CDCl₃): δ 166.8, 163.0, 134.3, 130.6, 128.2, 127.4, 77.3, 73.1, 69.5, 67.4, 45.7, 44.8, 32.7, 18.5, 18.4, 15.3, 14.0, -1.4; IR (film): 2952, 2920, 1669, 1454, 1438, 1416, 1385, 1362, 1248, 1083, 1057, 860, 836, 756, 703 cm⁻¹; $[\alpha]_D^{30} = -6.99$ (c = 2.76, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₂H₃₄N₂NaO₃S₂Si [M+Na⁺] 489.1672, Found 489.1649.

4.12 Dithiodiketopiperazine 17e

According to the General Procedure A, diketopiperazine **16e** (843 mg, 2.68 mmol) were converted to bis-(methylthio) diketopiperazine **17e** (539 mg, 1.33 mmol, 50% 2 steps from **16e**). A white solid; mp: 84.8–85.2 °C; ¹H NMR (400 MHz,

5.51 (1H, d, J = 9.8 Hz), 5.36 (1H, dd, J = 17.2, 1.6 Hz), 5.21 (1H, dd, J = 15.9, 1.8 Hz), 5.11 (1H, d, 9.8 Hz), 4.29 (2H, ddd, J = 5.5, 1.5, 1.5 Hz), 3.54–3.47 (2H, m), 3.47 (1H, d, J = 13.6 Hz), 3.18 (1H, d, J = 13.6 Hz), 2.26 (3H, s), 2.15 (3H, s), 2.14–2.04 (1H, m), 2.03–1.89 (1H, m), 1.58–1.46 (1H, m), 0.98–0.86 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 166.8, 162.9, 134.4, 134.2, 130.6, 128.2, 127.4, 117.0, 77.3, 73.3, 71.2, 69.6, 45.7, 44.9, 32.8, 18.5, 15.2, 13.9; IR (film): 2952, 2920, 2896, 1669, 1438, 1416, 1385, 1362, 1248, 1083, 1057, 860, 836, 756, 703 cm⁻¹; $[\alpha]_D^{30} = -9.65$ (c = 0.660, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₀H₂₆N₂NaO₃S₂ [M+Na⁺] 429.1277, Found 429.1271.

4.13 Dithiodiketopiperazine 15

To a mixture of bis-(methylthio) diketopiperazine 17e (52.9 mg, 130 µmol) and Pd(PPh₃)₄ (7.53 mg, 6.51 µmol) and 1,3dimethylbarbituric acid (40.7 mg, 260 µmol) was added MeOH (1.3 mL) at room temperature. The resulting suspension was stirred for 14 h at 40 °C. The reaction mixture was cooled to room temperature before addition of excess NaHCO₃ (130 mg) and further stirred for 40 min at room temperature. The reaction mixture was diluted by water (1.0 mL), and the separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried with anhydrous sodium sulfate, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes: EtOAc = 2:1 to 1:1) to afford bis-(methylthio) diketopiperazine 15 (41.6 mg, 124 µmol, 95%). A white solid; mp: 159.9–160.5 °C; ¹H NMR (400 MHz, DMSO-d₆): δ 8.85 (1H, s), 7.26–7.14 (5H, m), 3.48 (1H, d, J = 12.8 Hz), 3.46–3.26 (2H, m), 3.00 (1H, d, J = 12.8 Hz), 2.29 (3H, s), 2.11 (3H, s), 2.02 (1H, dd, J = 13.2, 7.2 Hz), 1.95–1.81 (1H, m), 1.63–1.52 (1H, m), 0.97 (1H, ddd, J = 9.3, 12.1, 12.1 Hz); ¹³C NMR (100 MHz, DMSO-d₆): δ 165.6, 163.2, 134.9, 130.3, 127.8, 126.9, 69.6, 67.4, 44.9, 44.6, 33.7, 18.6, 14.0, 13.2; IR (film): 3211, 3096, 3087, 3062, 2987, 2961, 2922, 2896, 1676, 1659, 1406, 1208, 755, 705 cm⁻¹; $[\alpha]_D^{28} = -17.5$ (*c* = 0.65, CHCl₃); HRMS (ESI) m/z: calcd. for $C_{16}H_{20}N_2NaO_2S_2$ [M+Na⁺] 359.0858, Found 359.0835.

4.14 amide 20a (General Procedure B)

To a solution of amide 19a (1.11 g, 5.26 mmol) and TBAI (195 mg, 0.527 mmol) in THF (21.0 mL) was added LiHMDS (1.3 M solution in THF, 4.46 mL, 5.80 mmol) at -40 °C. After stirring for 30 min, AllomCl (26) [20] (0.764 mL, 5.80 mmol) was added at -40 °C, and the mixture was stirred at -40 °C for 14 h. The reaction was quenched with sat. aq. NH₄Cl solution and the mixture was diluted by water. The aqueous phase was extracted with CH₂Cl₂ (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 4:1) to afford amide 20a (1.43 g, 5.08 mmol, 97%). A colorless oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 7.48–7.42 (5H, m), 7.36–7.26 (5H, m), 5.82–5.75 (1H, m), 5.14 (1H, d, J = 17.6 Hz), 5.07 (1H, d, J = 10.4 Hz), 4.86–4.67 (4H, m), 3.84 (br, 2H); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 171.0, 137.2, 135.4, 134.0, 129.4, 128.0, 127.9, 127.1, 126.6, 126.5, 116.3, 77.0, 68.0, 48.1; IR (film): 3062, 3030, 2942, 2862, 1651, 1496, 1445, 1417, 1267, 1071, 1050, 699 cm⁻¹; HRMS (ESI) m/z: calcd. for C₁₈H₁₉NNaO₂ [M+Na⁺] 304.1308, found 304.1296.

4.15 amide 20b

According to the General Procedure B, amide **19b** (55.5 mg, 191 µmol) was converted to amide **20b** (51.6 mg, 143 µmol,

δ 7.64–7.16 (2H, m), 7.47 (2H, d, J = 8.4 Hz), 7.34–7.26 (5H, m), 5.82–5.77 (1H, m), 5.16 (1H, d, J = 17.6 Hz), 5.09 (1H, d, J = 10.0 Hz), 3.86 (2H, br s); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 170.1, 137.0, 134.5, 133.9, 130.9, 128.7, 128.0, 127.2, 126.7, 122.9, 116.3, 77.0, 68.0, 48.3; IR (film): 3085, 3063, 3030, 2979, 2942, 2898, 2863, 1652, 1590, 1443, 1417, 1360, 1292, 1266, 1072, 1011, 932, 837, 754, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₈H₁₈BrNNaO₂ [M+Na⁺] 382.0413, Found 382.0403.

4.16 amide 20c

According to the General Procedure B, amide **19c** (62.4 mg, 2.43 µmol) was converted to amide **20c** (70.4 mg, 215mmol, 89%). A yellow oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 8.26 (2H, d, J = 8.0 Hz), 7.73 (2H, d, J = 8.0 Hz), 7.35–7.25 (5H, m), 5.84–5.71 (1H, m), 5.16 (1H, d, J = 17.6 Hz), 5.08 (1H, d, J = 10.8 Hz), 4.70 (2H, br), 4.64 (2H, br), 3.86 (2H, br); ¹³C NMR (100 MHz, CDCl₃): δ 169.4, 147.9, 141.5, 136.8, 133.9, 128.1, 127.9, 127.3, 126.8, 123.2, 116.5, 68.1, 54.3, 48.5; IR (film): 3080, 3066, 3031, 2945, 2862, 1651, 1602, 1521, 1425, 1349, 1072, 732, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₈H₁₈N₂NaO₄ [M+Na⁺] 349.1159, Found 349.1140.

4.17 amide 20d

According to the General Procedure B, amide **19d** (60.3 mg, 250 µmol) was converted to amide **20d** (68.0 mg, 218 µmol, 87%). A colorless oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 7.49–7.46 (2H, m), 7.35–7.25 (5H, m), 6.99–6.97 (2H, m), 5.86–5.76 (m, 1H), 5.19–5.14 (m, 1H), 5.11–5.08 (m, 1H), 4.69–4.68 (m, 4H), 3.89–3.86 (m, 2H), 3.80 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 170.8, 160.4, 137.4, 134.0, 128.6, 127.9, 127.4, 127.2, 126.6, 116.3, 113.4, 77.5, 67.9, 59.5, 48.2; IR (film) 1442, 1417, 1358, 1302, 1253, 1175, 1112, 1049, 933, 842, 766, 700, 598 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₉H₂₁NNaO₃ [M+Na⁺] 334.1414, Found 334.1408.

4.18 amide 20e

According to the General Procedure B, amide **19e** (74.0 mg, 331 µmol) was converted to amide **20e** (92.1 mg, 314 µmol, 95%). A yellow oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 7.45–7.42 (5H, m), 5.84–5.78 (1H, m), 5.18 (1H, d, *J* = 17.6 Hz), 5.09 (1H, d, *J* = 10.4 Hz), 4.69 (2H, br), 4.54 (2H, br), 3.86 (2H, br), 3.51 (4H, br), 3.26 (3H, br), 1.88–1.85 (m, 2H); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 170.9, 135.8, 134.1, 129.1, 127.8, 126.4, 116.1, 95.6, 67.4, 64.8, 54.2, 42.8, 27.9 (One signal is missing due to overlap.); IR (film): 2933, 2883, 1651, 1446, 1419, 1404, 1271, 1147, 1112, 1043, 919, 724, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₆H₂₃NNaO₄ [M+Na⁺] 316.1519, Found 316.1516.

4.19 amide 20f

According to the General Procedure B, amide **19f** (40.5 mg, 251 mmol) was converted to amide **20f** (49.5 mg, 214 µmol, 85%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.39–7.21 (5H, m), 5.88–5.81 (1H, m), 5.25 (1H, d, *J* = 16.8 Hz), 5.17–5.12 (2H, m), 4.82 (m, 1H), 4.06 (1H, d, *J* = 10.8 Hz), 3.97–3.94 (m, 2H), 2.64–2.50 (3H, m), 1.99–1.95 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 176.2, 140.5, 134.0, 128.8, 128.0, 126.5, 116.9, 70.0, 69.4, 60.4, 30.2, 28.1; IR (film): 3081, 3065, 3031, 2979, 2941, 2885, 1704, 1456, 1415, 1392, 1319, 1256, 1227, 1065, 932, 769, 703 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₄H₁₇NNaO₂ [M+Na⁺] 254.1151, Found 254.1147.

4.20 amide 20g

-proof 19g, 250 μmol) was converted to amide **20g** (71.7 mg, 236 μmol, 95%). A colorless oil; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.37–7.18 (5H, m), 5.94–5.82 (2H, m), 4.92 (0.7H, s), 4.67 (2H, s), 4.63 (1.3H, s), 4.31 (0.7, d, J = 5.2 Hz), 3.94 (1.3H, d, J = 6.0 Hz), 2.46 (1.3 H, t, J = 7.6 Hz), 2.33 (0.7H, 8.0 Hz), 1.71–1.63 (m, 2H), 1.32–1.35 (m, 8H), 0.88–0.84 (m, 3 H); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 174.8, 174.1, 137.8, 136.9, 134.3, 133.8, 128.8, 128.4, 128.1, 127.4, 127.2, 126.3, 117.6, 116.9, 76.8, 74.3, 69.5, 68.5, 49.2, 48.3, 33.32, 32.99, 31.63, 31.58, 29.3, 29.2, 29.00, 28.95, 25.3, 25.0, 22.53, 22.50, 14.0; IR (film): 3086, 3064, 3030, 2954, 2926, 2855, 1664, 1496, 1453, 1421, 1359, 1236, 1036, 991, 932, 732, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₉H₂₉NNaO₂ [M+Na⁺] 326.2091, Found 326.2082.

4.21 amide 19a (General Procedure C)

To a mixture of amide **20a** (56.0 mg, 199 µmol) and Pd(PPh₃)₄ (23.3 mg, 20.0 µmol) and 1,3-dimethylbarbituric acid (62.2 mg, 398 µmol) was added MeOH (0.67 mL) at room temperature. The resulting suspension was stirred for 6 h at 40 °C. The reaction mixture was cooled to room temperature before addition of excess aq. NH₃ (0.3 ml) and further stirred for 40 min at room temperature. Then, the reaction mixture was diluted by water (1.0 mL) and then the separated aqueous phase was extracted with CH_2Cl_2 (x3). The combined organic extracts were dried with anhydrous sodium sulfate and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 3:1) to afford amide **19a** (41.4 mg, 196 µmol, 98%).

4.22 amide 19b

According to the General Procedure C, amide **20b** (78.4 mg, 218 μ mol) was converted to amide **19b** (54.1 mg, 186 μ mol, 86%).

4.23 amide 19c

According to the General Procedure C, amide **20c** (37.8 mg, 116 μ mol) was converted to amide **19c** (13.9 mg, 54.2 μ mol, 47%).

4.24 amide 19d

According to the General Procedure C, amide 20c (65.5 mg, 210 μ mol) was converted to amide 19c (40.9 mg, 170 μ mol, 81%).

4.25 amide 19e

According to the General Procedure C, amide 20e (66.7 mg, 227 µmol) was converted to amide 19e (39.0 mg, 175 µmol, 78%).

4.26 amide 19f

According to the General Procedure C, amide **20f** (39.7 mg, 172 μ mol) were converted to amide **19f** (19.3 mg, 120 μ mol, 70%).

4.27 hemiaminal **21g**

To a mixture of amide **20g** (20.9 mg, 68.8 µmol) and Pd(PPh₃)₄ (8.0 mg, 6.87 µmol) and 1,3-dimethylbarbituric acid (21.5 mg, 134 µmol) were added THF (0.17 mL) and MeOH (0.06 mL) at room temperature. After stirred for 8 h at 40 °C, the reaction was quenched with sat. aq. Na₂CO₃, and the aqueous phase was extracted with EtOAc (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and

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co Journal F which was purified by silica gel column chromatography (hexanes:EtOAc = 5:1) to afford amide hemiaminal **21g** (16.6 mg, 63.2 μmol, 92%). A colorless oil; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.36–7.22 (5H, m), 4.86 (2H, br), 4.86–4.65 (2H, br), 3.69 (0.7H, br), 2.64–2.49 (0.9H, m), 2.31 (1.4H, t, *J* = 8.0 Hz), 1.71–1.61 (m, 2H), 1.25 (br, 8H), 0.88–0.84 (m, 3 H); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 175.9, 174.2, 138.0, 136.9, 128.9, 128.7, 128.1, 127.6, 127.5, 1 26.4, 72.2, 71.7, 51.5, 48.2, 33.4, 33.1, 31.7, 31.6, 29.4, 29.2, 29. 1, 29.0, 25.4, 24.9, 22.6, 14.0 (Two signals are missing due to overlap.); IR (film): 3371, 3087, 3063, 3030, 2954, 2926, 2870, 2855, 1636, 1629, 1453, 1423, 1041, 1029, 699 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₆H₂₆N₂O₂ [M+H⁺] 264.1958, Found 264.1946.

4.28 amide 19g

To a mixture of hemiaminal **21g** (19.1 mg, 72.7 µmol) and NaI (76.3 mg, 509 µmol) in MeCN (0.36 mL) was added TMSCI (46.0 µL, 495 µmol) at 0 °C. After stirred for 24 h at room temperature, the reaction was quenched with sat. aq. Na₂CO₃, and the aqueous phase was extracted with EtOAc (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 1:1) to afford amide **19g** (17.0 mg, 72.3 µmol, quant).

4.29 amide **22g**

To a mixture of amide 20g (17.3 mg, 57.0 µmol) and Pd(PPh₃)₄ (6.6 mg, 5.67 µmol) and 1,3-dimethylbarbituric acid (17.8 mg, 114 µmol) was added MeOH (0.2 mL) at room temperature. The resulting suspension was stirred for 16.5 h at 40 °C. Then, the reaction was quenched with sat. aq. Na₂CO₃, and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with 1 M aq. HCl and brine, dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes: EtOAc = 3:1) to afford amide **19g** (0.7 mg, 3.00 µmol, 5.3%), 21g (3.2 mg, 12.1 µmol, 21%), and 22g (6.1 mg, 22.0 µmol, 39%). A colorless oil; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.37–7.18 (5H, m), 4.85 (0.8H, s), 4.67 (1.2H, s), 4.61 (2H, s), 3.33 (1.2H, s), 3.28 (1.8H, s), 2.46 (1.2H, t, J = 7.6 Hz), 2.35 (0.8H, t, J = 7.6 Hz), 2.48-1.64 (2H, C)m), 1.32–1.25 (8H, m), 0.88–0.84 (3H, m); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 175.0, 174.2, 137.8, 136.9, 128.8, 128.5, 128.1, 127.5, 127.3, 126.3, 78.9, 76.0, 56.1, 55.3, 49.1, 48.3, 33.4, 33.0, 31.7, 31.6, 29.3, 29.2, 29.1, 29.0, 25.2, 25.1, 22.57, 22.55, 14.0 (One signal is missing due to overlap.); IR (film): 3087, 3063, 3031, 2953, 2927, 2871, 2855, 1663, 1453, 1420, 1389, 1095, 1077, 699 cm⁻¹; HRMS (ESI) m/z: calcd. for C₁₇H₂₇NNaO₂ [M+Na⁺] 300.1934, Found 300.1921.

4.30 amide 23

To a solution of carboxylic acid **9** [9] (491 mg, 1.10 mmol) and L-phenylalanine methyl ester **10** (497 mg, 2.78 mmol) in dry CH₂Cl₂ (15.0 mL) was added Et₃N (612 μ L, 4.40 mmol) at room temperature. After stirring for 10 min, BOP-Cl (576 mg, 2.26 mmol) was added, and the mixture was stirred for 2.7 h at room temperature. The reaction was quenched with sat. aq. NaHCO₃, and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with 1 M aq. HCl and brine, dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 2:1) to

NMR (400 MHz, CDCl₃, mixture of rotamers): H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.49–7.28 (8.8H, m), 7.12– 7.09 (1.2H, m), 6.48–6.37 (2.4H, m), 6.11 (0.6H, d, J = 8.4 Hz), 5.37–5.13 (3H, m), 5.05–4.76 (3H, m), 4.50 (1H, ddd, J = 8.4, 8.4, 2.8 Hz), 3.81 (1.2H, s), 3.72 (1.8H, s), 3.26-3.04 (3H, m), 2.76 (0.4H, d, J = 15.2 Hz), 2.64 (0.6H, d, J = 15.2 Hz), 0.940 (9H, s), 0.120 (3H, s), 0.054 (3H, s); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 171.6, 171.5, 171.4, 171.2, 154.7, 153.9, 145.5, 145.2, 136.1, 135.7, 135.4, 135.0, 129.4, 129.1, 128.5, 128.5, 128.4, 128.3, 128.1, 128.1, 127.9, 127.1, 127.0, 120.3, 118.7, 105.9, 105.5, 67.6, 67.3, 67.1, 65.9, 65.0, 64.4, 61.8, 61.4, 53.0, 52.5, 52.2, 52.1, 37.8, 37.7, 33.3, 32.0, 25.6, 17.8, -4.5, -4.6, -5.0, -5.1 (Five signals are missing due to overlap.); IR (film): 3325, 3031, 2953, 2928, 2895, 2855, 1746, 1713, 1639, 1526, 1437, 1408, 1350, 1318, 1252, 1210, 1142, 1113, 1085, 959, 836, 777, 748, 698 cm⁻¹; $[\alpha]_D^{27} = -175$ (c = 1.20, CHCl₃); HRMS (ESI) m/z: calcd. for C₃₃H₄₂N₂NaO₇Si [M+Na⁺] 629.2653, found 629.2624.

4.31 Amide 24

To a suspension of amide 23 (398 mg, 657 µmol) and Pd(OAc)₂ (57.8 mg, 257 µmol) in CH₂Cl₂ (12.0 mL) was added Et₃N (300 µL, 2.16 mmol) at room temperature. After stirring for 30 min at reflux, Et₃SiH (1.10 mL, 6.90 mmol) was added, and the mixture. was stirred for 40 min at reflux. The reaction mixture was diluted by CH2Cl2 and the reaction was quenched with sat. aq. NaHCO₃. The separated aqueous phase was extracted with CH₂Cl₂ (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 6:1 to 2:1) to afford amine 24 (347 mg, quant). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.90 (1H, d, J = 8.4 Hz), 7.22– 7.16 (3H, m), 7.03 (2H, d, J = 6.8 Hz), 6.25 (1H, d, J = 2.0 Hz), 6.22 (1H, d, J = 8.0 Hz), 4.80 (1H, ddd, J = 10.8, 3.2, 3.2 Hz), 4.71 (1H, dd, J = 8.0, 7.8 Hz), 4.27 (1H, dd, J = 7.8, 2.2 Hz), 3.79 (1H, dd, J = 7.2, 5.6 Hz), 3.70 (3H, s), 3.55 (1H, s), 3.19 (1H, dd, J = 14.0, 5.8 Hz), 3.01 (1H, dd, J = 14.0, 6.6 Hz), 2.73-2.71 (2H, m), 2.28 (1H, s), 0.82 (9H, s), 0.033 (3H, s), 0.022 (3H, s); ¹³C NMR (100 MHz, CDCl₃): δ 173.0, 171.9, 145.5, 135.8, 133.1, 129.1, 128.3, 126.9, 120.5, 105.6, 67.9, 63.9, 60.9, 52.5, 52.1, 37.6, 35.0, 25.6, 17.9, -3.8, -4.7; IR (film): 3349, 3029, 2952, 2928, 2893, 2855, 2360, 2341, 1745, 1677, 1644, 1508, 1437, 1360, 1344, 1254, 1200, 1079, 961, 837, 779, 702 cm⁻¹; $[\alpha]_D^{26} = -195$ (c = 1.65, CHCl₃); HRMS (ESI) m/z: calcd. for $C_{25}H_{37}N_2O_5Si^+$ [M+H⁺] 473.2466, found 473.2451.

4.32 Dithiodiketopiperazine 25

To a solution of amine 24 (447 mg, 945 µmol) in MeOH (250 mL) was added 25% aq. NH₃ (15.0 mL) at room temperature. After stirring for 2.7 days at room temperature, the reaction was concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes: EtOAc = 2:1) to afford diketopiperazine 25 (202 mg, 459 μ mol, 48%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.33–7.18 (5H, m), 6.41 (1H, s), 6.29 (1H, d, J = 8.0 Hz), 5.90 (1H, s), 5.00 (1H, d, J = 7.6 Hz), 4.88 (1H, dd, J = 8.0, 7.6 Hz), 4.77 (1H, s), 4.23–4.16 (2H, m), 3.51 (1H, dd, J = 14.0, 3.6 Hz), 2.89–2.79 (2H, m), 2.52 (1H, dd, J = 13.0, 13.0 Hz), 0.79 (9H, s), 0.011 (3H, s), 0.006 (3H, s); ¹³C NMR (100 MHz, CDCl₃): δ 168.5, 163.9, 145.0, 135.6, 135.1, 129.3, 128.9, 127.3, 112.4, 106.4, 66.9, 63.1, 57.9, 56.2, 37.3, 33.4, 25.7, 17.8, -4.3, -4.5; IR (film): 3381, 3228, 3220, 2952, 2928, 2895, 2855, 2359, 2341, 1684, 1663, 1444, 1422, 1294, 1256, 1202, 1139, 1083,

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CHCl₃); HRMS (EI) m/z: calcd. for $C_{24}H_{32}N_2O_4Si$ [M⁺] 440.2131, found 440.2134.

4.33 Dithiodiketopiperazine 27

To a solution of diketopiperazine 25 (63.4 mg, 144 µmol) in THF (630 $\mu L)$ was added LiHMDS (1.3 M solution in THF, 120 µL, 156 µmol) at -40 °C. After stirring for 8 min, AllomCl (26) [20] (120 μ L, 1.16 mmol) was added at -40 °C, and the mixture was stirred at -20 °C for 17 h. The reaction was quenched with sat. aq. NH₄Cl solution and the mixture was diluted by water. The separated aqueous phase was extracted with CH₂Cl₂ (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 4:1 to 2:1) to afford diketopiperazine 27 (66.3 mg, 120 µmol, 84%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.28–7.22 (3H, m), 7.04 (2H, d, J = 8.0 Hz), 6.28 (1H, d, J = 7.6 Hz), 6.27 (1H, s), 5.96–5.87 (1H, m), 5.51 (1H, d, J = 10.8 Hz), 5.31 (1H, dt, J = 17.2, 1.6 Hz), 5.24 (1H, dt, J = 10.8, 1.2 Hz), 4.87(1H, dd, J = 7.8, 7.6 Hz), 4.82 (1H, d, J = 7.8 Hz), 4.79 (1H, d, J = 10.8 Hz), 4.62 (1H, s), 4.57 (1H, m), 4.12 (1H, dd, J = 12.8, 6.0 Hz), 4.07–4.40 (2H, m), 3.42 (1H, dd, J = 14.0, 2.8 Hz), 3.21 (1H, dd, J = 14.0, 2.8 Hz), 2.44 (1H, dd, J = 13.2, 6.0 Hz), 0.94 (1H, dd, J = 13.2, 12.8 Hz), 0.79 (9H, s), 0.007 (3H, s), -0.009 (3H, s); ¹³C NMR (100 MHz, CDCl₃): § 166.2, 163.7, 144.9, 134.9, 134.7, 133.4, 130.1, 128.6, 127.4, 118.1, 112.8, 106.4, 72.5, 69.9, 68.6, 62.9, 59.1, 58.2, 36.5, 33.6, 25.8, 18.0, -4.3, -4.5; IR (film): 2952, 2928, 2856, 1662, 1452, 1441, 1295, 1200, 1138, 1092, 1051, 955, 906, 837, 777, 703 cm⁻¹; $[\alpha]_D^{25} = -333$ (*c* = 0.535, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₈H₃₈N₂NaO₅Si⁺[M+Na⁺] 533.2442, found 533.2417.

4.34 Dithiodiketopiperazine 28

To a solid of diketopiperazine 27 (66.3 mg, 120 µmol) was added HF·Py in pyridine (2.00 mL, 1:4, HF·Py containing ~70% hydrogen fluoride and ~30% pyridine) at room temperature. After stirring for 17 h at 45 °C, the reaction mixture was diluted by water at 0 °C. The separated aqueous phase was extracted with CH2Cl2 (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography ($CH_2Cl_2:EtOAc = 1:5$) to afford allyl alcohol 28 (46.6 mg, 118 µmol, 98%). A white solid; mp: 155.5–156.5 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.28–7.22 (3H, m), 7.07–7.04 (2H, m), 6.34 (1H, d, J = 8.0 Hz), 6.34 (1H, d, J = 2.0 Hz), 5.98–5.91 (1H, m), 5.41 (1H, d, J = 10.8 Hz), 5.35 (1H, ddd, J = 17.5, 2.9, 1.7 Hz), 5.24 (1H, ddd, J = 10.3, 2.9, 1.5 Hz), 5.05 (1H, dd, J = 8.2, 8.0 Hz), 4.82 (1H, d, J = 10.8 Hz), 4.79–4.76 (2H, m), 4.69 (1H, s), 4.58–4.56 (1H, m), 4.22 (1H, dd, J = 12.2, 6.0 Hz), 4.16–4.10 (2H, m), 3.42 (1H, dd, J = 14.2, 3.0 Hz), 3.23 (1H, dd, J = 14.2, 4.4 Hz), 2.48 (1H, dd, J = 14.4, 6.0 Hz), 0.94 (1H, dd, J = 14.4, 12.2 Hz); ¹³C NMR (100 MHz, CDCl₃): § 166.0, 163.8, 145.1, 135.2, 134.7, 133.6, 130.1, 128.4, 127.3, 117.8, 113.7, 105.7, 73.2, 70.3, 68.1, 62.6, 59.8, 58.3, 36.8, 33.3; $[\alpha]_D^{25} = -367$ (*c* = 0.260, CHCl₃); IR (film): 3425, 3419, 2921, 2359, 2340, 1694, 1659, 1454, 1339, 1296, 1200, 1137, 1074, 1047, 929, 857, 758, 736, 704 cm⁻¹; HRMS (ESI) m/z: calcd. for C₂₂H₂₄N₂NaO₅ [M+Na⁺] 419.1577, found 419.1558.

4.35 Dithiodiketopiperazine 29

To a solution of allyl alcohol $28~(74.3~mg,\,188~\mu mol)$ and nor-AZADO (8.50 mg, 61.5 $\mu mol)$ in dry $CH_2Cl_2~(1.40~mL)$ was

After stirring for 1.5 h at room temperature, the reaction was quenched with sat. aq. Na_2SO_3 solution and water (1:2). The separated aqueous phase was extracted with CH_2Cl_2 (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes: EtOAc = 1:3) to afford lactone 29 (71.0 mg, 180 µmol, 96%). A white foam; ¹H NMR (400 MHz, CDCl₃): δ 7.28–7.22 (3H, m), 7.09 (2H, d, *J* = 8.0 Hz), 7.00 (1H, d, J = 7.6 Hz), 6.59 (1H, d, J = 2.4 Hz), 6.01–5.91 (1H, m), 5.61 (1H, d, J = 7.6 Hz), 5.36 (1H, d, J = 18.4 Hz), 5.35 (1H, d, J = 10.4 Hz), 5.31 (1H, s), 5.27 (1H, d, J = 10.4 Hz), 4.90 (1H, d, J = 10.4 Hz), 4.68 (1H, dd, J = 4.0, 3.4 Hz), 4.20–4.10 (2H, m), 3.91 (1H, dd, J = 13.4, 6.0 Hz), 3.43 (1H, dd, J = 14.0, 3.4 Hz), 3.29(1H, dd, J = 14.0, 4.0 Hz), 2.53 (1H, dd, J = 15.2, 6.0 Hz), 1.01 (1H, dd, J = 13.4, 13.4 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 180.9, 164.9, 163.6, 153.3, 136.9, 134.5, 133.4, 130.0, 128.3, 127.1, 117.6, 115.4, 107.8, 73.5, 70.4, 66.3, 60.0, 57.0, 36.9, 31.8; IR (film): 3028, 2925, 2867, 1687, 1667, 1604, 1454, 1350, 1327, 1292, 1204, 1078, 1051, 940, 930, 864, 820, 758, 736 cm⁻¹; $[\alpha]_D^{26} = -279$ (*c* = 0.445, CHCl₃); HRMS (ESI) *m/z*: calcd. for C₂₂H₂₃N₂O₅ [M+H⁺] 395.1601, found 395.1605.

4.36 Dithiodiketopiperazine 30

To a solution of lactone 29 (138 mg, 350 µmol) in CH₂Cl₂ (750 µL) and EtOH (500 µL) was added CeCl₃· 7H₂O (381 mg) in EtOH (2.0 mL) at -78 °C. After stirring for 10 min, NaBH₄ in EtOH (22.0 mg, 582 μ mol in 550 μ L) was added at -78 °C, and the mixture was stirred at -78 °C for 1.6 h. The reaction was quenched with aq. NH₄Cl solution at room temperature. The separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 1:3) to afford allyl alcohol 30 (126 mg, 318 µmol, 91%). A white foam; ¹H NMR (400 MHz, CDCl₃): δ 7.31–7.23 (3H, m), 7.08 (2H, d, *J* = 6.4 Hz), 6.27 (1H, dd, J = 2.2, 2.2 Hz), 6.13 (1H, dd, J = 8.0, 2.4 Hz), 6.00-5.90 (1H, m), 5.42 (1H, d, J = 10.8 Hz), 5.36 (1H, d, J = 17.2 Hz), 5.26 (1H, d, J = 10.0 Hz), 5.07 (1H, d, J = 3.6 Hz), 4.88 (1H, s), 4.87 (1H, d, J = 10.8 Hz), 4.64 (1H, dd, J = 3.0, 4.2 Hz), 4.52 (1H, d, J = 7.6 Hz), 4.20–4.17 (1H, m), 4.14 (1H, d, J = 5.8 Hz), 4.13 (1H, d, J = 5.8 Hz), 3.91 (1H, dd, J = 13.0, 5.8 Hz), 3.40 (1H, dd, J = 13.8, 3.0 Hz), 3.27 (1H, dd, J = 13.8, 4.2 Hz), 2.36 (1H, dd, J = 13.6, 5.8 Hz), 0.80 (1H, dd, J = 13.6, 13.0 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 167.0, 165.1, 138.0, 137.0, 134.2, 133.4, 130.2, 128.7, 127.7, 118.1, 109.8, 109.7, 73.2, 70.5, 69.1, 64.1, 59.8, 56.3, 37.0, 33.7; IR (film): 3369, 3011, 2942, 2863, 1661, 1453, 1344, 1296, 1193, 1133, 1078, 1058, 930, 759, 756, 730, 704, 578 cm⁻¹; $[\alpha]_D^{25} = -229$ (c = 1.04, CHCl₃); HRMS (ESI) *m/z*: calcd. for C₂₂H₂₄N₂NaO₅ [M+Na⁺] 419.1577, found 419.1564.

4.37 Dithiodiketopiperazine 31

To a suspension of sulfur S₈ (63.6 mg, 247 µmol) in Et₂O (1.0 mL) was added LiHMDS (1.3 M in THF, 725 µL, 943 µmol) at room temperature. After stirring for 5 min, diketopiperazine **30** (18.6 mg, 47.0 µmol) in a mixture of THF (2.0 mL) and Et₂O (1.0 mL) was added at room temperature, and the resulting solution was stirred for 1 min. Then, LiHMDS (1.3 M in THF, 725 µL, 943 µmol) was added, and the mixture was stirred for 30 min at room temperature. The reaction was quenched with aq. NH₄Cl and diluted with water. The separated aqueous phase was extracted with Et₂O (x1) and CH₂Cl₂ (x2). The combined organic

and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 16:1x2, then hexanes: $CH_2Cl_2 = 3:5$) to afford the mixture of bridged polysulfides. To a solution of the polysulfides in a mixture of THF (3.2 mL) and EtOH (3.2 mL) was added NaBH₄ (36.5 mg, 965 $\mu mol)$ at 0 °C. After stirred for 8 min at room temperature, MeI (1.60 mL, 25.6 mmol) was added at 0 °C, and the resulting mixture was stirred at room temperature for 4 h. The reaction was quenched by aq. NH₄Cl, and the separated aqueous phase was extracted with Et₂O (x1) and CH₂Cl₂ (x2). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 16:1, then hexanes: EtOAc = 2:1) to afford bis-(methylthio) diketopiperazine **31** (18.3 mg, 30.3 µmol, 65% 2 steps from **30**). A white foam; ¹H NMR (400 MHz, CDCl₃): δ 7.29–7.24 (3H, m), 7.15–7.12 (2H, m), 6.31 (1H, m), 6.17 (1H, dd, J = 8.4, 2.4 Hz), 6.03–5.93 (1H, m), 5.40–5.34 (1H, m), 5.53 (1H, d, J = 9.6 Hz), 5.24–5.22 (2H, m), 5.12 (1H, d, *J* = 9.6 Hz), 4.91 (1H, dd, *J* = 8.2, 1.8 Hz), 4.58–4.55 (1H, m), 4.42 (1H, d, J = 8.0 Hz), 4.30– 4.28 (2H, m), 3.48 (1H, d, J = 13.2 Hz), 3.23 (1H, d, J = 13.2Hz), 2.55 (1H, d, J = 15.2 Hz), 2.29 (3H, s), 2.53 (3H, s), 1.59 (1H, d, J = 15.2 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 167.1, 166.1, 137.6, 137.2, 134.3, 133.6, 130.7, 128.6, 127.9, 117.2, 110.8, 107.1, 77.5, 73.4, 72.2, 71.3, 67.7, 65.4, 45.0, 38.1, 15.2, 14.0; IR (film): 3370, 3008, 2920, 2852, 1662, 1435, 1416, 1380, 1191, 1136, 1083, 1056, 1050, 772, 756, 704 cm⁻¹; $[\alpha]_D^{25} = -123$ $(c = 0.619, \text{CHCl}_3)$; HRMS (ESI) m/z: calcd. for $C_{24}H_{28}N_2NaO_5S_2$ [M+Na⁺] 511.1332, found 511.1303.

4.38 (-)-Emestrin H (5)

To a solid of bis-(methylthio) diketopiperazine **31** (6.65 mg, 13.6 µmol) was added Pd(PPh₃)₄ (27.2 mM solution in CH₂Cl₂, 25.0 µL 0.682 µmol). The mixture was concentrated under reduced pressure give a pale brown solid. To the solid was added 1,3-dimethylbarbituric acid (4.26 mg, 27.3 µmol) in MeOH (136 μ L), and the mixture was stirred for 16.5 h at 40 °C. The reaction mixture was diluted with MeOH (68 µL) and stirred for 2 h. Then, the mixture was cooled to room temperature and diluted with MeOH (200 μ L). To the mixture was added NaHCO₃ (40.0 mg), and the resulting suspension was stirred for 25 min at room temperature and diluted with CH₂Cl₂ and H₂O. The separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude by preparative material, which was purified TLC (hexanes:EtOAc = $3:1 \times 3$) to afford (-)-emestrin H (5) (4.46 mg, 10.6 µmol, 78%) and diketopiperazine 30 (1.15 mg, 17%). A white solid; mp: 86.0–87.2 °C; ¹H NMR (600 MHz, CDCl₃): δ 7.34–7.30 (3H, m), 7.19 (2H, d, J = 6.6 Hz), 6.36 (1H, s), 6.19 (1H, dd, J = 1.8, 8.4 Hz), 6.05 (1H, s), 5.12 (1H, d, J = 4.8 Hz),4.93 (1H, d, J = 8.4 Hz), 4.63–4.61 (1H, m), 4.55 (1H, d, J = 7.2 Hz), 3.64 (1H, d, J = 13.2 Hz), 3.02 (1H, d, J = 13.2 Hz), 2.63 (1H, d, J = 15.0 Hz), 2.41 (3H, s), 2.23 (3H, s), 1.88 (1H, d, J = 15.0 Hz); ¹³C NMR (150 MHz, CDCl₃): δ 167.2, 165.4, 137.7, 137.4, 133.2, 130.7, 128.8, 128.1, 110.9, 107.4, 72.3, 68.5, 68.4, 64.9, 46.3, 38.9, 14.7, 14.3; IR (film): 3361, 3224, 3062, 3008, 2918, 2848, 2360, 2341, 1675, 1638, 1409, 1340, 1193, 1139, 1123, 1047, 756, 730, 704, 581, 564 cm⁻¹; $[\alpha]_D^{28} = -203$ (c = 0.234, MeOH) (Lit. $[\alpha]_D^{25} = -97$ (c = 0.071, MeOH)); HRMS (ESI) m/z: calcd. for C₂₀H₂₂N₂NaO₄S₂ [M+Na⁺] 441.0913, found 441.0907. The spectral data were identical with those reported in the literature [14].

4.39 (-)-Asteroxepin (4)

DMAP (1.23 mg, 10.1 µmol) in pyridine (120 µL) was added Ac₂O in pyridine (0.127 M, 100 µL, 12.7 µmol) at room temperature. After stirring for 50 min at room temperature, additional Ac₂O (13.0 µL, 138 µmol) was added, and the mixture was stirred for 35 min. Then, additional DMAP (5.69 mg, 46.6 µmol) was added, and the mixture was stirred for 45 min at room temperature. The reaction was diluted with CH2Cl2, and quenched with aq. 1 M HCl. The separated aqueous phase was extracted with CH₂Cl₂ (x5), and the combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 7:11) to afford (-)-asteroxepin (4) (2.60 mg, 5.65 µmol, 89%). A white solid; mp: 189.6–190.0 °C; ¹H NMR (600 MHz, CDCl₃): δ = 7.32–7.27 (3H, m), 7.20–7.18 (2H, m), 6.41 (1H, dd, J = 2.4, 2.4 Hz), 6.29 (1H, dd, J = 8.4, 3.0 Hz), 6.00 (1H, br s), 5.88 (1H, ddd, *J* = 7.5, 2.1, 2.1), 4.89 (1H, br. d, *J* = 7.8 Hz), 4.68 (1H, dd, *J* = 8.4, 1.2 Hz), 3.66 (1H, d, *J* = 13.8 Hz), 2.97 (1H, d, *J* = 13.8 Hz), 2.64 (1H, d, *J* = 15.0 Hz), 2.36 (3H, s), 2.20 (3H, s), 2.13 (3H, s), 1.82 (1H, ddd, J = 15.3, 2.1, 2.1 Hz); ¹³C NMR (150 MHz, CDCl₃): $\delta = 170.1$, 165.6, 164.4, 139.7, 137.3, 133.5, 130.9, 128.7, 127.9, 109.1, 105.5, 71.4, 69.9, 68.0, 61.7, 46.4, 39.6, 21.2, 14.5, 14.0; $[\alpha]_D^{27} = -178$ (c = 0.556, CHCl₃) (Lit. $[\alpha]_D = -102$ (c = 0.4, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₂H₂₄N₂NaO₅S₂ [M+Na⁺] 483.1019, found 483.0996. The spectral data were identical with those reported in the literature [13].

4.40 AllomCl (26) [20]

To a mixture of allyl alcohol (40.0 mL, 0.588 mmol) and paraformaldehyde (19.0 g, 0.633 mol) was added anhydrous hydrogen chloride generated from H_2SO_4 and NaCl at room temperature. After stirred for 30 min at room temperature, the mixture was heated by oil bath at 150 °C for 2 h. The resulting mixture was diluted with hexanes, dried over magnesium sulfate, and the hexanes were removed under reduced pressure. The crude material was distilled from anhydrous calcium chloride (37.0 °C/67.5 mmHg) to afford AllomCl (26) (25.0 g, 0.234 mol, 40%) as colorless oil.

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16. Compounds **16a** and **16b** with C3 (*S*) and C8a (*S*) stereochemistries were synthesized by the stepwise condensation of *N*-Cbz-L-proline and *N*-PMB-L-phenylalanine methyl ester or *N*-2,4-DMB-phenylalanine methyl ester, respectively.



17. Compounds **16c–e** were synthesized from *trans*-**13** with C3 (*S*) and C8a (*R*) stereochemistries, which was prepared by the stepwise condensation of *N*-Boc-L-proline and L-phenylalanine methyl ester. During the intramolecular condensation step, the C8a (*S*) stereochemistry was completely isomerized to give *trans*-**13** with C8a (*R*) stereochemistry.



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16c-16e

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Total syntheses of (-)-emestrin H and (-)-asteroxepin

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ABSTRACT

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1. Introduction

Dithiodiketopiperazine alkaloids bearing the sulfur-containing diketopiperazine as a core structure have attracted attention due to their structural diversity and fascinating biological activities [1,2]. Among them, compounds bearing the 4,5-dihydrooxepine ring, such as (-)-acetylaranotin (1) [3], (+)-MPC1001B (7) [4], and (-)-acetylapoaranotin (6) [5], have been challenging synthetic targets and a lot of effort has been made to synthesize these compounds [6,7]. Reisman et al. reported the first innovative total synthesis of (-)-acetylaranotin (1) in 2012 [8], followed by the first total synthesis of (-)-acetylapoaranotin (6) [10]. We also focused on this class of compounds, reporting the second total synthesis of 1 just after Reisman et al. in 2012 [9]. Using the synthetic protocol established during the synthetic studies on 1, which was based on the characteristic proline-fused 4,5dihydrooxepine ring, we successfully performed the first total synthesis of (+)-MPC1001B (7) [11] and determined the structure of (-)-SCH64874 (3) and hirsutellomycin (8) via semi-synthesis [12]. Despite these synthetic developments over the last decade, the total syntheses of (-)-asteroxepin (4) [13] and (-)-emestrin H (5) [14] bearing the NH-free diketopiperazine core have not been reported so far.

Our preliminary model studies toward 4 and 5 revealed that the enolate-mediated introduction of two methylthioethers reported by Nicolaou *et al.* [15] was unsuccessful for the unprotected diketopiperazine 13 due to the competitive deprotonation of the unprotected N–H proton (Scheme 1b). Therefore, the major challenge toward the synthesis of 4 and 5 was the selection of the appropriate protecting group for the

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First total syntheses of (–)-emestrin H and (–)-asteroxepin are described. To find the appropriate protecting group on the amide nitrogen of the diketopiperazine core, we conducted model studies using a simple diketopiperazine derivative. As a result, allyloxymethyl (Allom) group was the most suitable protecting group, which tolerated Nicolaou's sulfenylation conditions, and was easily cleavable under the mild conditions using $Pd(PPh_3)_4$ and *N*,*N*-dimethylbarbituric acid leaving methylthioethers intact. The general utility of Allom group for protection of amides was studied using simple substrates. Finally, the effectiveness of Allom group was robust enough during installation of two methylthioethers to the diketopiperazine core and easily removed at the final step. The first total synthesis of (–)-asteroxepin was also completed by acylation of (–)-emestrin H.

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diketopiperazine ring's NH group. The ideal protecting group (PG in Scheme 1a) should be robust enough to tolerate the strong basic conditions during the introduction of the twomethylthioethers. Moreover, it should be easily and chemoselectively cleaved under the particularly mild conditions in the final stage of the total synthesis without affecting the base-, nucleophile-, and oxidant-sensitive methylthioethers and the acid-sensitive dihydrooxepin structure. However, compared to the many available protecting groups for amines, there are limited choices for amides, especially for diketopiperazines. In



Fig. 1. Dithiodiketopiperazine alkaloids

Tetrahedron

functionalized NH-free diketopiperazines and found that the less common allyloxymethyl (Allom) group, which only have been used for a protection of the N-3 position in xanthine and the imidazole nitrogen in histidine was the most suitable protecting group [19]. Its efficiency was further demonstrated by the first total synthesis of (–)-asteroxepin (4) and (–)-emestrin H (5).



Scheme 1. Synthetic plan and preliminary results.

2. Results and discussion

To identify the most suitable protecting group that would fulfill the abovementioned requirements, we prepared a series of *N*-protected diketopiperazines (16a-e) following a conventional method [16,17], which were then subjected to the sulfenylation conditions established by Nicolaou *et al.* (Table 1) [15]. Among the protecting groups in substrates 16a-e, stable benzylic protecting groups, such as *p*-methoxybenzyl (PMB) in 16a or 2,4-dimethoxybenzyl (2,4-DMB) in 16b, were not affected by the sulfenylation conditions, providing dithiodiketopiperazines 17aand 17b, respectively, in moderate yields (Table 1, entries 1

Table 1

Protecting groups and introduction of the methylthioether.



^aC3 (*S*), C8a (*S*) isomer. ^bC3 (*S*), C8a (*R*) isomer. ^cIsolated yield. ^dObtained as single diastereomer.

16d with the trimethylsilylethoxymethyl (SEM) group and **16e** with the Allom group, afforded the corresponding dimethylthio compounds (**17d** and **17e**) in good yields (Table 1, entries 4 and 5). However, analogue **16c**, protected as urethane with an allyloxycarbonyl group (Alloc), afforded a complex mixture under the same sulfenylation conditions (Table 1, entry 3).

The deprotection of the bis(methylthio) products 17a, 17b, 17d, and 17e was examined under the most frequently used deprotection conditions for each protecting group (Table 2). Analogues 17a and 17b bearing PMB and 2,4-DMB groups were decomposed upon treatment with ceric ammonium nitrate (CAN) 2,3-dichloro-5,6-dicyano-1,4-benzoquinone and (DDO). respectively (Table 2, entries 1 and 2). Moreover, the SEM group in 17d was stable upon treatment with HF/pyridine and 17d was recovered (Table 2, entry 3), whereas the use of TBAF in THF/DMF decomposed 17d (Table 2, entry 4). In contrast to these unsuccessful entries, the Allom group in 17e could be smoothly removed by a combination of $Pd(PPh_3)_4$ and N,Ndimethylbarbituric acid (DMBA) in MeOH yielding 15 in 56% yield, while a substantial amount of the hemiaminal intermediate 18 was also obtained (Table 2, entry 5) [18,19]. The subsequent in situ treatment of the reaction mixture with aq. NH₃ was effective in reducing the amount of 18, affording 15 in 87% yield (Scheme 2). Furthermore, the addition of solid NaHCO₃ after the consumption of 17e improved the yield of 15 to up to 95%.

Table 2

Deprotection conditions.

<pre>N</pre>		$ \begin{array}{c} \mathbf{PG} \\ \mathbf{PG} \\ \mathbf{Ph} \end{array} \xrightarrow{\text{conditions}} \begin{array}{c} MeS \\ \mathbf{Ph} \\ Me \end{array} $		Ph MeS O	N OH Ph SMe
	17		15	18 (from	17f)
entry	17	reagents	temp.	Time (h) Yie	eld (%) ^a
1^{b}	17a	CAN, CH ₃ CN/H ₂ O	rt	2	0
2 ^b	17b	DDQ, CH ₃ CN/H ₂ O	60 °C	20	0
3 ^b	17d	TBAF, THF/DMF	60 °C	1.8	0
4 ^c	17d	HF/pyridine	60 °C	22	0
5 ^d	17e	Pd(PPh ₃) ₄ , DMBA, MeOH	30 °C	6	56

^aisolated yield. ^bDecomposition. ^c**17d** was completely recovered. ^dReaction conditions: **17** (1.0 eq), Pd(PPh₃)₄ (45 mol%), DMBA (5.5 eq), MeOH (0.1 M), 30 °C, 6 h.°The hemiaminal intermediate **18** was obtained in 38% yield.



Scheme 2. Removal of the Allom group.

The feasibility of the removal of the Allom protecting group from secondary amides was investigated using a series of *N*-Allom secondary amides (**20**) (see Experimental Section for the preparation and additional data) (Table 3 and Scheme 3). The treatment of the *N*-Allom benzamides **20a–e** and γ lactam **20f** with Pd(PPh₃)₄ and DMBA in MeOH smoothly provided the hemiaminal intermediate **21**, which was converted into amides **19a–e** and **19f**, respectively, in good yields using aq. NH₃ (Table 3, entries 1–6). In contrast, the reaction of *N*-Allom aliphatic amides **20g** under the Pd/*N*,*N*- **21g** (21%), and *N*-MOM amides **22g** (39%) due to the exchange of the OH group in intermediate **21g** with the methoxy group of methanol (Scheme 3). This side reaction was overcome by performing the deprotection of **20g** in THF and H₂O, providing the hemiaminal intermediates **21g**, which were then converted to amides **19g** in good yields using trimethylsilyl iodide (TMSI).

Table 3.

One-pot Removal of the Allom group.



Reaction conditions: **20** (1.0 eq), $Pd(PPh_3)_4$ (10 mol%), DMBA (2.0 eq), MeOH (0.3 M), 40 °C, 6 h, then aq. NH₃ (excess). The yields refer to the isolated products.





The applicability of the Allom protecting group was fully demonstrated by the first total syntheses of (–)-emestrin H (5) and (–)-asteroxepin (4). The proline-fused dihydrooxepine derivative 9, which was prepared according to our previously established protocol in the total synthesis of (–)-acetylaranotin (1) [9], was first condensed with the (S)-phenylalanine methyl

then removed under transfer hydrogenation conditions and the formation of the tricyclic diketopiperazine derivative **25** was promoted upon treatment of the resulting aminoester with ammonium hydroxide. Subsequently, the Allom group was introduced using a combination of the allyloxymethyl chloride (AllomCl) **26** [20] and lithium hexamethyldisilazide (LiHMDS), which afforded the desired product **27** in 84% yield. After desilylation, the stereochemistry of the C-10 hydroxyl group was inverted through a stepwise process [9], which included an oxidation of **28** to ketone using PhI(OAc)₂ and 9-azanoradamantane *N*-oxyl (nor-AZADO) [21] and a subsequent Luche reduction, which gave the desired alcohol **30** as the sole isomer.



Scheme 4. Preparation of diketopiperazine 30.

The final steps toward the synthesis of (–)-emestrin H (**5**) and (–)-asteroxepin (**4**) including the crucial introduction of the two methylthioethers and the removal of the Allom protecting group are depicted in Scheme 5. According to the model study (*vide supra*), diketopiperazine **30** was treated with a mixture of excess LiHMDS and S₈ to form an epipolysulfide. Its reduction to the corresponding dithiol with NaBH₄ and the subsequent methylation afforded the bis(methylthio) derivative **31** in 65% yield over two steps from **30**. Finally, the Allom group was removed under the optimized conditions affording (–)-emestrin H (**5**) in 78% yield, which was further acetylated to give (–)-asteroxepin (**4**). All properties of synthetic (–)-emestrin H (**5**) and (–)-asteroxepin (**4**) were identical with those reported except magnitude of specific rotations proving the reported absolute stereochemistry of **4** and **5** by this total synthesis [22].



Scheme 5. Final steps of the total syntheses of (–)-emestrin H (5) and (–)-asteroxepin (4).

4 3.

In conclusion, the total syntheses of (–)-emestrin H (**5**) and (–)-asteroxepin (**4**) have been accomplished for the first time. Their preparation was successful owing to the suitability of the Allom group as protecting group for the amide nitrogen of the highly functionalized dithio-diketopiperazine intermediate. This protecting group could be easily cleaved even in the presence of the acid-sensitive dihydrooxepin structure and the

the acid-sensitive dihydrooxepin structure and the dimethylthioethers, which are sensitive to bases, nucleophiles, and oxidants. Thus, this study demonstrated the application potential of the Allom group not only for the synthesis of other *N*-unprotected diketopiperazine alkaloids, but also for the preparation of various functionalized amides.

4. Experimental section

4.1 General method

Materials were obtained from commercial suppliers and used without further purification unless otherwise mentioned. Anhydrous THF, Et₂O, CH₂Cl₂, toluene, DMF, and MeCN were purchased from commercial suppliers. Anhydrous MeOH was dried and distilled according to the standard protocols. All reactions were carried out under Ar atmosphere unless otherwise mentioned. Flash column chromatography was performed on Silica Gel 60N (Kanto, spherical neutral, 40-50 µm). Preparative TLC and analytical TLC was performed on Merck 60 F254 glass plates precoated with a 0.25 mm thickness of silica gel. NMR spectra were recorded on a JNM-AL400 spectrometer and a JEOL ECA600 spectrometer. Chemical shifts for ¹H NMR are reported in parts per million (ppm) downfield from tetramethylsilane as the internal standard and coupling constants are in Hertz (Hz). The following abbreviations are used for spin multiplicity: s = singlet, d = doublet, t = triplet, q = quartet, m =multiplet, and br = broad. Chemical shifts for ${}^{13}C$ NMR are reported in ppm, relative to the central line of a triplet at 77.0 ppm for CDCl₃ or a septet at 39.5 ppm for DMSO-d₆. IR spectra were measured on a SHIMADZU FTIR-8300 spectrometer. Mass spectra were recorded on a Bruker micrOTOF (ESI). Optical rotations were measured on a Horiba SEPA-300 highly sensitive polarimeter. Melting point determinations were performed by using a Yanaco MP-500 instrument. Compounds 9 [9], 19a-e [25], 19f [26], and 19g [25] were prepared according to the procedures reported in references.

4.2 Diketopiperazine16a

To a solution of N-Cbz-L-proline (3.68 g, 14.8mmol) and N-PMB-L-phenylalanine methyl ester (4.24 g, 14.2 mmol) in Et₃N and CH₂Cl₂ (5 mL and 30 mL) was added BOP-Cl (3.74 g, 14.7 mmol) at room temperature. After stirring for 24.5 h at room temperature, additional BOP-Cl (1.96 g, 7.70 mmol) was added, and the reaction mixture was stirred for 2.5 h. The reaction was quenched with sat. aq. NaHCO3, and the aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic layers were washed with 1 M aq. HCl, dried over Na2SO4, filtered and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 1:1) to afford amide 32a (5.80 g, 10.9 mmol, 77%). To a suspension of amide 32a (5.80 g, 10.9 mmol) and Pd(OAc)₂ (983 mg, 4.38 mmol) in CH₂Cl₂ (55.0 mL) was added Et₃N (3.08 mL, 21.9 mmol) at room temperature. After stirring at reflux for 5 min, Et₃SiH (8.74 mL, 54.7 mmol) was added and the mixture was stirred at reflux for 3.5 h. The reaction mixture was diluted with CH₂Cl₂ and the reaction was quenched with sat. aq. NaHCO₃. The separated aqueous phase was extracted with anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (EtOAc) to afford diketopiperazine **16a** (3.00 g, 8.23 mmol, 75%). A white foam; The spectral data of **16a** were identical with those reported in the literature [23].

4.3 Diketopiperazine16b

To a solution of N-Cbz-L-proline (1.06 g, 4.25 mmol), N-2,4-DMB-L-phenylalanine methyl ester (1.15 g, 3.49 mmol) in Et₃N and CH₂Cl₂ (2.0 mL and 20.0 mL) was added BOP-Cl (1.87 g, 7.35 mmol) at room temperature. After stirring for 20 h at room temperature, the reaction was quenched with 1 M aq. HCl and the resulting mixture was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes: EtOAc = 3:2 to 6:5) to afford amide 32b (846 mg, 1.51 mmol, 47%). To a suspension of amide 32b (846 mg, 1.51 mmol) and Pd(OAc)₂ (135 mg, 601 µmol) in CH₂Cl₂ (10.0 mL) was added Et₃N (425 µL, 3.02 mmol) at room temperature. After stirring at reflux for 7 min, Et₃SiH (1.20 mL, 7.51 mmol) was added and the mixture was stirred at reflux for 3.5 h. The reaction mixture was diluted with CH₂Cl₂ and the reaction was quenched with sat. aq. NaHCO3. The separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude which was purified by silica gel column material, chromatography (EtOAc) to afford diketopiperazine 16b (540 mg, 1.37 mmol, 91%). A white foam; ¹H NMR (400 MHz, CDCl₃): δ 7.31-7.20 (4H, m), 7.12-7.06 (2H, m), 6.48-6.43 (2H, m), 5.40 (1H, d, J = 14.8 Hz), 4.32 (1H, m), 4.25 (1H, d, J =14.8 Hz), 3.84 (3H, s), 3.81 (3H, s), 3.72 (1H, dd, J = 12.0, 6.0 Hz), 3.68-3.61 (1H, m), 3.36-3.27 (2H, m), 3.14-3.07 (1H, m), 1.91–1.81 (1H, m), 1.60–1.42 (2H, m), 0.14–0.01 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 165.3, 163.9, 160.7, 158.7, 135.1, 131.7, 130.0, 128.3, 127.1, 115.9, 104.4, 98.3, 60.3, 59.0, 55.3, 55.3, 44.0, 40.6, 36.4, 28.4, 20.9; IR (film): 2992, 2952, 2944, 2891, 1656, 1612, 1587, 1508, 1456, 1298, 1289, 1261, 1208, 1184, 1157, 1033, 751, 703 cm⁻¹; $[\alpha]_D^{31} = -116$ (c = 0.66, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₃H₂₆N₂NaO₄ [M⁺+Na] 417.1785, Found 417.1769.

4.4 Amide 33

To a solution of N-Boc-L-proline (10.8 g, 50.0 mmol) and Lphenylalanine methyl ester (8.95 g, 50.0 mmol) in CH₂Cl₂ (200 mL) were added Et₃N (27.9 mL, 200 mmol), HOBt (6.99 g, 51.7 mmol), and EDCI/HCl (9.91 g, 51.7 mmol) at room temperature. After stirred for 29.5 h at room temperature, the organic phase was washed with 1 M aq. HCl (x3) and sat. aq. NaHCO₃ (x2). The resulting organic phase was dried over Na₂SO₄, filtered and concentrated under reduced pressure to give a crude material, which was used for the next reaction without further purification. To a solution of the crude material in CH₂Cl₂ (120 mL) was added TFA (15 mL) at room temperature. After stirred for 20 h at room temperature, the reaction mixture was diluted with CH₂Cl₂ (150 mL) and neutralized by sat. aq. NaHCO₃. The aqueous phase was extracted with $CH_2Cl_2(x3)$, and the combined organic layers were dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (CH_2Cl_2 :MeOH = 10:1) to afford amine amide 33 (6.76 g, 24.5 mmol, 49%, 2 steps from N-Boc-L-

with those reported in the literature [24].

4.5 Diketopiperazine trans-13

To a solution of amide 33 (6.76 g, 24.5 mmol) in MeOH (400 mL) was added NaHCO3 (20.0 g, 85.3 mmol) at room temperature. After stirring for 22.5 h at 65 °C, MeOH was removed under reduced pressure. The residue was diluted with CH₂Cl₂ and filtered through a pad of Celite[®]. The filtrate was washed with H₂O, and the aqueous phase was extracted with CH₂Cl₂ (x2). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by crystallization from EtOAc to afford diketopiperazine trans-13 (3.43 g, 14.1 mmol, 57%). The mother liquor was concentrated in vacuo and the second crop was crystallized from EtOAc to afford diketopiperazine trans- $1\overline{3}$ (643 mg, 2.64 mmol, 11%). A white solid.; mp: 153.5–154.2 °C; $^{1}\mathrm{H}$ NMR (400 MHz, CDCl3): δ 7.35–7.20 (5H, m), 6.31 (1H, s), 4.22 (1H, ddd, J = 6.6, 4.0, 3.8Hz), 3.63 (1H, ddd, J = 12.4, 8.5, 8.5 Hz), 3.40 (1H, ddd, J = 12.4, 9.4, 3.0 Hz), 3.15 (1H, dd, J = 13.9, 6.6 Hz), 3.08 (1H, dd, *J* = 13.9, 4.0 Hz), 2.97 (1H, dd, *J* = 10.4, 6.4 Hz), 2.23–2.15 (1H, m), 1.98–1.90 (1H, m), 1.86–1.64 (2H, m); ¹³C NMR (100 MHz, CDCl₃): δ 169.5, 164.8, 135.3, 129.9, 128.5, 127.3, 58.7, 57.6, 44.9, 40.3, 28.8, 21.5; IR (film): 3483, 3238, 3063, 3029, 2981, 2953, 2931, 2886, 1664, 1496, 1454, 1336, 1307, 1296, 1206, 1186, 1115, 1106, 921, 732, 702, 593 cm⁻¹; $[\alpha]_D^{26} = +93.0$ (c = 0.200, H₂O); HRMS (ESI) m/z: calcd. for $C_{14}H_{17}N_2O_2$ [M+H⁺] 245.1285, found 245.1277. The spectral data of trans-13 were identical with those reported in the literature [27].

4.6 Diketopiperazine16c

To a solution of diketopiperazine trans-13 (203 mg, 0.832 mmol) in THF (4.0 mL) was added LiHMDS (1.3 M solution in THF, 650 µL, 845 µmol) at 0 °C. After stirring for 4 min, Alloc-OSu (172 mg, 863 µmol) in THF (1.50 mL) was added at 0 °C. The resulting mixture was stirred for 35 min at room temperature. The reaction was quenched with sat. aq. NH₄Cl, and the resulting mixture was diluted by water. The separated aqueous phase was extracted with CH₂Cl₂ (x3), and the combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material. which was purified by silica gel column chromatography (CH₂Cl₂:EtOAc = 5:1 to 3:1) to afford diketopiperazine 16c (248 mg, 789 µmol, 95%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.31–7.26 (3H, m), 7.19–7.14 (2H, m), 5.99–5.87 (1H, m), 5.43 (1H, d, J = 17.2 Hz), 5.31 (1H, d, J = 10.8 Hz), 5.01 (1H, ddd, J = 5.0, 5.0, 1.4 Hz), 4.75 (1H, ddd, *J* = 13.3, 5.8, 1.0 Hz), 4.69 (1H, ddd, *J* = 13.3, 5.6, 1.2 Hz), 3.60-3.50 (1H, m), 3.45-3.37 (1H, m), 3.33-3.22 (2H, m), 2.64 (1H, dd, J = 9.6, 6.8 Hz), 2.17-2.07 (1H, m), 1.96-1.79 (2H, m),1.72–1.57 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 167.2, 164.0, 151.8, 134.9, 130.7, 129.9, 128.6, 127.5, 119.4, 67.9, 62.3, 58.6, 44.8, 38.2, 29.1, 21.8; IR (film): 3086, 3062, 3028, 2983, 2953, 2885, 1782, 1731, 1672, 1455, 1384, 1270, 1231, 980, 763, 747, 703 cm⁻¹; $[\alpha]_D^{26} = +143$ (*c* = 0.425, CHCl₃); HRMS (ESI) *m/z*: calcd. for C₁₈H₂₀N₂NaO₄ [M+Na⁺] 351.1315, Found 351.1319.

4.7 Diketopiperazine16d

To a solution of diketopiperazine *trans*-**13** (1.10 g, 4.51 mmol) in dry DMF (10.0 mL) was added NaH (60% dispersion in mineral oil, 248 mg, 6.20 mmol) at room temperature. After stirring for 15 min, SEMC1 (1.60 mL, 9.03 mmol) was added at room temperature, and the resulting mixture was stirred at 60 °C for 18 h. The reaction was quenched with sat. aq. NH_4Cl at room

and washed with water (x5) and brine (x1). The organic phase was dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 1:3) to afford diketopiperazine 16d (1.33 g, 3.55 mmol, 79%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.29–7.26 (3H, m), 7.18–7.15 (2H, m), 5.13 (1H, d, J = 10.4 Hz), 4.53 (1H, d, J = 10.4 Hz), 4.42 (1H, dd, J = 5.2, 4.9 Hz), 3.61-3.52 (3H, m), 3.35 (1H, ddd, J = 12.1, 9.3, 2.9 Hz), 3.24 (1H, dd, J = 14.3, 5.2 Hz), 3.19 (1H, dd, J = 14.3, 4.9 Hz), 2.52 (1H, dd, J= 10.6, 6.6 Hz), 2.14-2.06 (1H, m), 1.92-1.84 (1H, m), 1.79-1.68 (1H, m), 1.69-1.53 (1H, m), 1.01-0.87 (2H, m), 0.01 (9H, s); ¹³C NMR (100 MHz, CDCl₃): δ 168.3, 164.7, 135.2, 129.9, 128.5, 127.5, 73.3, 66.3, 62.1, 57.6, 44.8, 37.6, 29.1, 21.8, 17.9, -1.5; IR (film): 2952, 2890, 1671, 1454, 1440, 1296, 1260, 1249, 1207, 1075, 1044, 860, 837, 749, 703 cm⁻¹; $[\alpha]_D^{25} = +55.0$ (c = 2.71, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₀H₃₀NaN₂O₃Si [M⁺+Na] 397.1918, Found 397.1909.

4.8 Diketopiperazine16e

To a solution of diketopiperazine trans-13 (880 mg, 3.61 mmol) in dry THF (8.0 mL) was added NaH (60% dispersion in mineral oil, 181 mg, 4.53 mmol) at room temperature. After stirring for 50 min, AllomCl (26) [20] (750 µL, 7.23 mmol) was added at room temperature, and the mixture was stirred for 9.5 h at 50 °C. The reaction was quenched with sat. aq. NH4Cl solution at room temperature, and the aqueous phase was diluted by water, extracted with Et₂O (x1) and CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 2:1, then EtOAc only) to afford diketopiperazine 16e (946 mg, 3.01 mmol, 83%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.32–7.24 (3H, m), 7.19–7.14 (2H, m), 5.89 (1H, m), 5.29 (1H, ddd, J = 17.6, 1.2,1.2, Hz), 5.19 (1H, d, J = 10.8 Hz), 5.14 (1H, d, J = 10.6 Hz), 4.58 (1H, d, J = 10.6 Hz), 4.43 (1H, dd, J = 4.7, 4.8 Hz), 4.10-3.98 (2H, m), 3.57 (1H, ddd, J = 12.3, 8.5, 8.5 Hz), 3.36 (1H, ddd, J = 12.3, 9.3, 2.9 Hz), 3.25 (1H, dd, J = 14.3, 4.8 Hz), 3.20 (1H, dd, J = 14.3, 4.7 Hz), 2.53 (1H, dd, J = 10.6, 6.6 Hz), 2.09 (1H, ddd, J = 12.0, 6.2, 6.2 Hz), 1.94-1.84 (1H, m), 1.81-1.69(1H, m), 1.66–1.53 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 168.4, 164.7, 135.3, 133.8, 129.9, 128.6, 127.5, 117.4, 73.6, 70.1, 62.4, 57.7, 44.9, 37.7, 29.0, 21.8; IR (film): 3028, 2979, 2951, 2883, 1668, 1453, 1440, 1344, 1296, 1261, 1207, 1072, 1057, 923, 748, 703 cm⁻¹; $[\alpha]_D^{26} = +60.0$ (*c* = 4.41, CHCl₃); HRMS (ESI) m/z: calcd. for C₁₈H₂₂N₂NaO₃ [M⁺+Na] 337.1523, Found 337.1506.

4.9 Dithiodiketopiperazine 17a (General Procedure A)

To a suspension of sulfur S_8 (264 mg, 1.03 mmol) in THF (5.0 mL) was added NaHMDS (0.6 M in toluene, 5.00 mL, 3.00 mmol) at room temperature. After stirring for 1 min, diketopiperazine **16a** (372 mg, 1.02 mmol) in THF (5.0 mL) was added, and the mixture was stirred for 1 min. Then, NaHMDS (0.6 M in toluene, 3.40 mL, 2.04 mmol) was added, and the mixture was stirred for 30 min at room temperature. The reaction was quenched with sat. aq. NH₄Cl, and the separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was used for the next reaction without further purification. To a solution of the crude material in THF (10.0 mL) and EtOH (10.0 mL) was added NaBH₄ (964 mg, 25.5 mmol) at 0 °C. After

sti added at 0 °C, and the resulting mixture was stirred for 19 h at room temperature. The reaction was quenched by sat. aq. NH₄Cl, and the separated aqueous phase was extracted with CH_2Cl_2 (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes:EtOAc = 12:5) to afford bis(metylthio) diketopiperazine 17a (204 mg, 0.449 mmol, 44%, 2 steps from **16a**). A white solid; mp: 136.7–137.0 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.53 (2H, d, J = 8.8 Hz), 7.31–7.22 (3H, m), 7.09 (2H, d, J = 6.0 Hz), 6.83 (2H, d, J = 8.8 Hz), 5.18 (1H, d, J = 14.4Hz), 4.54 (1H, d, J = 14.4 Hz), 3.79 (3H, s), 3.66–3.48 (2H, m), 3.54 (1H, d, J = 13.6 Hz), 3.25 (1H, d, J = 13.6 Hz), 2.05-1.91 (2H, m), 2.01 (3H, s), 1.99 (3H, s), 1.58-1.49 (1H, m), 0.81-0.71 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 165.9, 163.0, 158.6, 134.1, 130.2, 129.6, 128.4, 127.7, 113.3, 78.8, 69.9, 55.1, 46.7, 45.8, 44.1, 32.7, 18.5, 15.0, 14.0 (One signal is missing due to overlap.), IR (film): 2994, 2956, 2933, 2922, 1662, 1512, 1421, 1395, 1352, 1247, 1177, 1033, 756, 703 cm⁻¹; $[\alpha]_D^{25} = +0.89$ (c 0.58, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₄H₂₈N₂NaO₃S₂ [M+Na⁺] 479.1419, Found 479.1434.

4.10 Dithiodiketopiperazine 17b

According to the General Procedure A, diketopiperazine 16b (301 mg, 0.763 mmol) were converted to bis-(methylthio) diketopiperazine 17b (210 mg, 0.431 mmol, 57 %, 2 steps from 16b). A white solid; mp: 186.3-186.7 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.39 (1H, d, J = 8.5 Hz), 7.30–7.20 (3H, m), 7.11 (2H, d, J = 6.4 Hz), 6.48 (1H, d, J = 2.1 Hz), 6.41 (1H, d, J = 8.5, 2.1 Hz), 5.07 (1H, d, J = 15.8 Hz), 4.68 (1H, d, J = 15.8 Hz), 3.90 (3H, s), 3.79 (3H, s), 3.67–3.53 (2H, m), 3.55 (1H, d, J = 13.6 Hz), 3.33 (1H, d, J = 13.6 Hz), 2.14 (3H, s), 2.14 (3H, s), 2.10-1.94 (2H, m), 1.61–1.50 (1H, m), 0.86–0.73 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 165.7, 162.9, 159.5, 157.2, 134.2, 130.1, 128.3, 127.5, 127.0, 118.5, 103.5, 98.2, 78.5, 69.7, 55.3, 55.2, 45.8, 44.1, 41.1, 32.5, 18.4, 15.2, 14.2; IR (film): 2998, 2959, 2936, 2921, 2835, 1662, 1616, 1590, 1508, 1455, 1439, 1420, 1396, 1361, 1300, 1261, 1208, 1156, 1120, 1036, 755, 703 cm⁻¹; $[\alpha]_D^{32} = +21.3$ (c = 0.26, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₅H₃₀N₂NaO₄S₂ [M+Na⁺] 509.1539, Found 509.1526.

4.11 Dithiodiketopiperazine 17d

According to the General Procedure A, diketopiperazine 16e (456 mg, 1.22 mmol) were converted to bis-(methylthio) diketopiperazine 17d (386 mg, 0.828 mmol, 68%, 2 steps from 16d). A white solid; mp: 82.9-83.3 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.25–7.19 (3H, m), 7.12 (2H, dd, *J* = 7.8, 2.2 Hz), 5.48 (1H, d, J = 9.6 Hz), 5.06 (1H, d, J = 9.6 Hz), 3.79 (2H, t, J = 8.6 Hz), 3.50 (2H, dd, J = 9.2, 6.4 Hz), 3.46 (1H, d, J = 13.6 Hz), 3.18 (1H, d, J = 13.2 Hz), 2.26 (3H, s), 2.14 (3H, s), 2.12–2.04 (1H, m), 2.02-1.89 (1H, m), 1.55-1.46 (1H, m), 1.07-0.96 (2H, m), 0.93–0.84 (1H, m), 0.03 (9H, s); ¹³C NMR (100 MHz, CDCl₃): δ 166.8, 163.0, 134.3, 130.6, 128.2, 127.4, 77.3, 73.1, 69.5, 67.4, 45.7, 44.8, 32.7, 18.5, 18.4, 15.3, 14.0, -1.4; IR (film): 2952, 2920, 1669, 1454, 1438, 1416, 1385, 1362, 1248, 1083, 1057, 860, 836, 756, 703 cm⁻¹; $[\alpha]_D^{30} = -6.99$ (c = 2.76, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₂H₃₄N₂NaO₃S₂Si [M+Na⁺] 489.1672, Found 489.1649.

4.12 Dithiodiketopiperazine 17e

According to the General Procedure A, diketopiperazine **16e** (843 mg, 2.68 mmol) were converted to bis-(methylthio) diketopiperazine **17e** (539 mg, 1.33 mmol, 50% 2 steps from **16e**). A white solid; mp: 84.8–85.2 °C; ¹H NMR (400 MHz,

5.51 (1H, d, J = 9.8 Hz), 5.36 (1H, dd, J = 17.2, 1.6 Hz), 5.21 (1H, dd, J = 15.9, 1.8 Hz), 5.11 (1H, d, 9.8 Hz), 4.29 (2H, ddd, J = 5.5, 1.5, 1.5 Hz), 3.54–3.47 (2H, m), 3.47 (1H, d, J = 13.6 Hz), 3.18 (1H, d, J = 13.6 Hz), 2.26 (3H, s), 2.15 (3H, s), 2.14–2.04 (1H, m), 2.03–1.89 (1H, m), 1.58–1.46 (1H, m), 0.98–0.86 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 166.8, 162.9, 134.4, 134.2, 130.6, 128.2, 127.4, 117.0, 77.3, 73.3, 71.2, 69.6, 45.7, 44.9, 32.8, 18.5, 15.2, 13.9; IR (film): 2952, 2920, 2896, 1669, 1438, 1416, 1385, 1362, 1248, 1083, 1057, 860, 836, 756, 703 cm⁻¹; $[\alpha]_D^{30} = -9.65$ (c = 0.660, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₀H₂₆N₂NaO₃S₂ [M+Na⁺] 429.1277, Found 429.1271.

4.13 Dithiodiketopiperazine 15

To a mixture of bis-(methylthio) diketopiperazine 17e (52.9 mg, 130 µmol) and Pd(PPh₃)₄ (7.53 mg, 6.51 µmol) and 1,3dimethylbarbituric acid (40.7 mg, 260 µmol) was added MeOH (1.3 mL) at room temperature. The resulting suspension was stirred for 14 h at 40 °C. The reaction mixture was cooled to room temperature before addition of excess NaHCO₃ (130 mg) and further stirred for 40 min at room temperature. The reaction mixture was diluted by water (1.0 mL), and the separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried with anhydrous sodium sulfate, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes: EtOAc = 2:1 to 1:1) to afford bis-(methylthio) diketopiperazine 15 (41.6 mg, 124 µmol, 95%). A white solid; mp: 159.9–160.5 °C; ¹H NMR (400 MHz, DMSO-d₆): δ 8.85 (1H, s), 7.26–7.14 (5H, m), 3.48 (1H, d, J = 12.8 Hz), 3.46–3.26 (2H, m), 3.00 (1H, d, J = 12.8 Hz), 2.29 (3H, s), 2.11 (3H, s), 2.02 (1H, dd, J = 13.2, 7.2 Hz), 1.95–1.81 (1H, m), 1.63–1.52 (1H, m), 0.97 (1H, ddd, J = 9.3, 12.1, 12.1 Hz); ¹³C NMR (100 MHz, DMSO-d₆): δ 165.6, 163.2, 134.9, 130.3, 127.8, 126.9, 69.6, 67.4, 44.9, 44.6, 33.7, 18.6, 14.0, 13.2; IR (film): 3211, 3096, 3087, 3062, 2987, 2961, 2922, 2896, 1676, 1659, 1406, 1208, 755, 705 cm⁻¹; $[\alpha]_D^{28} = -17.5$ (*c* = 0.65, CHCl₃); HRMS (ESI) m/z: calcd. for $C_{16}H_{20}N_2NaO_2S_2$ [M+Na⁺] 359.0858, Found 359.0835.

4.14 amide 20a (General Procedure B)

To a solution of amide 19a (1.11 g, 5.26 mmol) and TBAI (195 mg, 0.527 mmol) in THF (21.0 mL) was added LiHMDS (1.3 M solution in THF, 4.46 mL, 5.80 mmol) at -40 °C. After stirring for 30 min, AllomCl (26) [20] (0.764 mL, 5.80 mmol) was added at -40 °C, and the mixture was stirred at -40 °C for 14 h. The reaction was quenched with sat. aq. NH₄Cl solution and the mixture was diluted by water. The aqueous phase was extracted with CH₂Cl₂ (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 4:1) to afford amide 20a (1.43 g, 5.08 mmol, 97%). A colorless oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 7.48–7.42 (5H, m), 7.36–7.26 (5H, m), 5.82–5.75 (1H, m), 5.14 (1H, d, J = 17.6 Hz), 5.07 (1H, d, J = 10.4 Hz), 4.86–4.67 (4H, m), 3.84 (br, 2H); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 171.0, 137.2, 135.4, 134.0, 129.4, 128.0, 127.9, 127.1, 126.6, 126.5, 116.3, 77.0, 68.0, 48.1; IR (film): 3062, 3030, 2942, 2862, 1651, 1496, 1445, 1417, 1267, 1071, 1050, 699 cm⁻¹; HRMS (ESI) m/z: calcd. for C₁₈H₁₉NNaO₂ [M+Na⁺] 304.1308, found 304.1296.

4.15 amide 20b

According to the General Procedure B, amide **19b** (55.5 mg, 191 µmol) was converted to amide **20b** (51.6 mg, 143 µmol,

δ 7.64–7.16 (2H, m), 7.47 (2H, d, J = 8.4 Hz), 7.34–7.26 (5H, m), 5.82–5.77 (1H, m), 5.16 (1H, d, J = 17.6 Hz), 5.09 (1H, d, J = 10.0 Hz), 3.86 (2H, br s); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 170.1, 137.0, 134.5, 133.9, 130.9, 128.7, 128.0, 127.2, 126.7, 122.9, 116.3, 77.0, 68.0, 48.3; IR (film): 3085, 3063, 3030, 2979, 2942, 2898, 2863, 1652, 1590, 1443, 1417, 1360, 1292, 1266, 1072, 1011, 932, 837, 754, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₈H₁₈BrNNaO₂ [M+Na⁺] 382.0413, Found 382.0403.

4.16 amide 20c

According to the General Procedure B, amide **19c** (62.4 mg, 2.43 µmol) was converted to amide **20c** (70.4 mg, 215mmol, 89%). A yellow oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 8.26 (2H, d, J = 8.0 Hz), 7.73 (2H, d, J = 8.0 Hz), 7.35–7.25 (5H, m), 5.84–5.71 (1H, m), 5.16 (1H, d, J = 17.6 Hz), 5.08 (1H, d, J = 10.8 Hz), 4.70 (2H, br), 4.64 (2H, br), 3.86 (2H, br); ¹³C NMR (100 MHz, CDCl₃): δ 169.4, 147.9, 141.5, 136.8, 133.9, 128.1, 127.9, 127.3, 126.8, 123.2, 116.5, 68.1, 54.3, 48.5; IR (film): 3080, 3066, 3031, 2945, 2862, 1651, 1602, 1521, 1425, 1349, 1072, 732, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₈H₁₈N₂NaO₄ [M+Na⁺] 349.1159, Found 349.1140.

4.17 amide 20d

According to the General Procedure B, amide **19d** (60.3 mg, 250 µmol) was converted to amide **20d** (68.0 mg, 218 µmol, 87%). A colorless oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 7.49–7.46 (2H, m), 7.35–7.25 (5H, m), 6.99–6.97 (2H, m), 5.86–5.76 (m, 1H), 5.19–5.14 (m, 1H), 5.11–5.08 (m, 1H), 4.69–4.68 (m, 4H), 3.89–3.86 (m, 2H), 3.80 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 170.8, 160.4, 137.4, 134.0, 128.6, 127.9, 127.4, 127.2, 126.6, 116.3, 113.4, 77.5, 67.9, 59.5, 48.2; IR (film) 1442, 1417, 1358, 1302, 1253, 1175, 1112, 1049, 933, 842, 766, 700, 598 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₉H₂₁NNaO₃ [M+Na⁺] 334.1414, Found 334.1408.

4.18 amide 20e

According to the General Procedure B, amide **19e** (74.0 mg, 331 µmol) was converted to amide **20e** (92.1 mg, 314 µmol, 95%). A yellow oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 7.45–7.42 (5H, m), 5.84–5.78 (1H, m), 5.18 (1H, d, *J* = 17.6 Hz), 5.09 (1H, d, *J* = 10.4 Hz), 4.69 (2H, br), 4.54 (2H, br), 3.86 (2H, br), 3.51 (4H, br), 3.26 (3H, br), 1.88–1.85 (m, 2H); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 170.9, 135.8, 134.1, 129.1, 127.8, 126.4, 116.1, 95.6, 67.4, 64.8, 54.2, 42.8, 27.9 (One signal is missing due to overlap.); IR (film): 2933, 2883, 1651, 1446, 1419, 1404, 1271, 1147, 1112, 1043, 919, 724, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₆H₂₃NNaO₄ [M+Na⁺] 316.1519, Found 316.1516.

4.19 amide 20f

According to the General Procedure B, amide **19f** (40.5 mg, 251 mmol) was converted to amide **20f** (49.5 mg, 214 µmol, 85%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.39–7.21 (5H, m), 5.88–5.81 (1H, m), 5.25 (1H, d, *J* = 16.8 Hz), 5.17–5.12 (2H, m), 4.82 (m, 1H), 4.06 (1H, d, *J* = 10.8 Hz), 3.97–3.94 (m, 2H), 2.64–2.50 (3H, m), 1.99–1.95 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 176.2, 140.5, 134.0, 128.8, 128.0, 126.5, 116.9, 70.0, 69.4, 60.4, 30.2, 28.1; IR (film): 3081, 3065, 3031, 2979, 2941, 2885, 1704, 1456, 1415, 1392, 1319, 1256, 1227, 1065, 932, 769, 703 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₄H₁₇NNaO₂ [M+Na⁺] 254.1151, Found 254.1147.

4.20 amide 20g

-proof 19g, 250 μmol) was converted to amide **20g** (71.7 mg, 236 μmol, 95%). A colorless oil; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.37–7.18 (5H, m), 5.94–5.82 (2H, m), 4.92 (0.7H, s), 4.67 (2H, s), 4.63 (1.3H, s), 4.31 (0.7, d, J = 5.2 Hz), 3.94 (1.3H, d, J = 6.0 Hz), 2.46 (1.3 H, t, J = 7.6 Hz), 2.33 (0.7H, 8.0 Hz), 1.71–1.63 (m, 2H), 1.32–1.35 (m, 8H), 0.88–0.84 (m, 3 H); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 174.8, 174.1, 137.8, 136.9, 134.3, 133.8, 128.8, 128.4, 128.1, 127.4, 127.2, 126.3, 117.6, 116.9, 76.8, 74.3, 69.5, 68.5, 49.2, 48.3, 33.32, 32.99, 31.63, 31.58, 29.3, 29.2, 29.00, 28.95, 25.3, 25.0, 22.53, 22.50, 14.0; IR (film): 3086, 3064, 3030, 2954, 2926, 2855, 1664, 1496, 1453, 1421, 1359, 1236, 1036, 991, 932, 732, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₉H₂₉NNaO₂ [M+Na⁺] 326.2091, Found 326.2082.

4.21 amide 19a (General Procedure C)

To a mixture of amide **20a** (56.0 mg, 199 µmol) and Pd(PPh₃)₄ (23.3 mg, 20.0 µmol) and 1,3-dimethylbarbituric acid (62.2 mg, 398 µmol) was added MeOH (0.67 mL) at room temperature. The resulting suspension was stirred for 6 h at 40 °C. The reaction mixture was cooled to room temperature before addition of excess aq. NH₃ (0.3 ml) and further stirred for 40 min at room temperature. Then, the reaction mixture was diluted by water (1.0 mL) and then the separated aqueous phase was extracted with CH_2Cl_2 (x3). The combined organic extracts were dried with anhydrous sodium sulfate and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 3:1) to afford amide **19a** (41.4 mg, 196 µmol, 98%).

4.22 amide 19b

According to the General Procedure C, amide **20b** (78.4 mg, 218 μ mol) was converted to amide **19b** (54.1 mg, 186 μ mol, 86%).

4.23 amide 19c

According to the General Procedure C, amide **20c** (37.8 mg, 116 μ mol) was converted to amide **19c** (13.9 mg, 54.2 μ mol, 47%).

4.24 amide 19d

According to the General Procedure C, amide 20c (65.5 mg, 210 μ mol) was converted to amide 19c (40.9 mg, 170 μ mol, 81%).

4.25 amide 19e

According to the General Procedure C, amide 20e (66.7 mg, 227 µmol) was converted to amide 19e (39.0 mg, 175 µmol, 78%).

4.26 amide 19f

According to the General Procedure C, amide **20f** (39.7 mg, 172 μ mol) were converted to amide **19f** (19.3 mg, 120 μ mol, 70%).

4.27 hemiaminal **21g**

To a mixture of amide **20g** (20.9 mg, 68.8 µmol) and Pd(PPh₃)₄ (8.0 mg, 6.87 µmol) and 1,3-dimethylbarbituric acid (21.5 mg, 134 µmol) were added THF (0.17 mL) and MeOH (0.06 mL) at room temperature. After stirred for 8 h at 40 °C, the reaction was quenched with sat. aq. Na₂CO₃, and the aqueous phase was extracted with EtOAc (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and

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co Journal F which was purified by silica gel column chromatography (hexanes:EtOAc = 5:1) to afford amide hemiaminal **21g** (16.6 mg, 63.2 μmol, 92%). A colorless oil; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.36–7.22 (5H, m), 4.86 (2H, br), 4.86–4.65 (2H, br), 3.69 (0.7H, br), 2.64–2.49 (0.9H, m), 2.31 (1.4H, t, *J* = 8.0 Hz), 1.71–1.61 (m, 2H), 1.25 (br, 8H), 0.88–0.84 (m, 3 H); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 175.9, 174.2, 138.0, 136.9, 128.9, 128.7, 128.1, 127.6, 127.5, 1 26.4, 72.2, 71.7, 51.5, 48.2, 33.4, 33.1, 31.7, 31.6, 29.4, 29.2, 29. 1, 29.0, 25.4, 24.9, 22.6, 14.0 (Two signals are missing due to overlap.); IR (film): 3371, 3087, 3063, 3030, 2954, 2926, 2870, 2855, 1636, 1629, 1453, 1423, 1041, 1029, 699 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₆H₂₆N₂O₂ [M+H⁺] 264.1958, Found 264.1946.

4.28 amide 19g

To a mixture of hemiaminal **21g** (19.1 mg, 72.7 µmol) and NaI (76.3 mg, 509 µmol) in MeCN (0.36 mL) was added TMSCI (46.0 µL, 495 µmol) at 0 °C. After stirred for 24 h at room temperature, the reaction was quenched with sat. aq. Na₂CO₃, and the aqueous phase was extracted with EtOAc (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 1:1) to afford amide **19g** (17.0 mg, 72.3 µmol, quant).

4.29 amide **22g**

To a mixture of amide 20g (17.3 mg, 57.0 µmol) and Pd(PPh₃)₄ (6.6 mg, 5.67 µmol) and 1,3-dimethylbarbituric acid (17.8 mg, 114 µmol) was added MeOH (0.2 mL) at room temperature. The resulting suspension was stirred for 16.5 h at 40 °C. Then, the reaction was quenched with sat. aq. Na₂CO₃, and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with 1 M aq. HCl and brine, dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes: EtOAc = 3:1) to afford amide **19g** (0.7 mg, 3.00 µmol, 5.3%), 21g (3.2 mg, 12.1 µmol, 21%), and 22g (6.1 mg, 22.0 µmol, 39%). A colorless oil; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.37–7.18 (5H, m), 4.85 (0.8H, s), 4.67 (1.2H, s), 4.61 (2H, s), 3.33 (1.2H, s), 3.28 (1.8H, s), 2.46 (1.2H, t, J = 7.6 Hz), 2.35 (0.8H, t, J = 7.6 Hz), 2.48-1.64 (2H, C)m), 1.32–1.25 (8H, m), 0.88–0.84 (3H, m); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 175.0, 174.2, 137.8, 136.9, 128.8, 128.5, 128.1, 127.5, 127.3, 126.3, 78.9, 76.0, 56.1, 55.3, 49.1, 48.3, 33.4, 33.0, 31.7, 31.6, 29.3, 29.2, 29.1, 29.0, 25.2, 25.1, 22.57, 22.55, 14.0 (One signal is missing due to overlap.); IR (film): 3087, 3063, 3031, 2953, 2927, 2871, 2855, 1663, 1453, 1420, 1389, 1095, 1077, 699 cm⁻¹; HRMS (ESI) m/z: calcd. for C₁₇H₂₇NNaO₂ [M+Na⁺] 300.1934, Found 300.1921.

4.30 amide 23

To a solution of carboxylic acid **9** [9] (491 mg, 1.10 mmol) and L-phenylalanine methyl ester **10** (497 mg, 2.78 mmol) in dry CH₂Cl₂ (15.0 mL) was added Et₃N (612 μ L, 4.40 mmol) at room temperature. After stirring for 10 min, BOP-Cl (576 mg, 2.26 mmol) was added, and the mixture was stirred for 2.7 h at room temperature. The reaction was quenched with sat. aq. NaHCO₃, and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with 1 M aq. HCl and brine, dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 2:1) to

NMR (400 MHz, CDCl₃, mixture of rotamers): H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.49–7.28 (8.8H, m), 7.12– 7.09 (1.2H, m), 6.48–6.37 (2.4H, m), 6.11 (0.6H, d, J = 8.4 Hz), 5.37–5.13 (3H, m), 5.05–4.76 (3H, m), 4.50 (1H, ddd, J = 8.4, 8.4, 2.8 Hz), 3.81 (1.2H, s), 3.72 (1.8H, s), 3.26-3.04 (3H, m), 2.76 (0.4H, d, J = 15.2 Hz), 2.64 (0.6H, d, J = 15.2 Hz), 0.940 (9H, s), 0.120 (3H, s), 0.054 (3H, s); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 171.6, 171.5, 171.4, 171.2, 154.7, 153.9, 145.5, 145.2, 136.1, 135.7, 135.4, 135.0, 129.4, 129.1, 128.5, 128.5, 128.4, 128.3, 128.1, 128.1, 127.9, 127.1, 127.0, 120.3, 118.7, 105.9, 105.5, 67.6, 67.3, 67.1, 65.9, 65.0, 64.4, 61.8, 61.4, 53.0, 52.5, 52.2, 52.1, 37.8, 37.7, 33.3, 32.0, 25.6, 17.8, -4.5, -4.6, -5.0, -5.1 (Five signals are missing due to overlap.); IR (film): 3325, 3031, 2953, 2928, 2895, 2855, 1746, 1713, 1639, 1526, 1437, 1408, 1350, 1318, 1252, 1210, 1142, 1113, 1085, 959, 836, 777, 748, 698 cm⁻¹; $[\alpha]_D^{27} = -175$ (c = 1.20, CHCl₃); HRMS (ESI) m/z: calcd. for C₃₃H₄₂N₂NaO₇Si [M+Na⁺] 629.2653, found 629.2624.

4.31 Amide 24

To a suspension of amide 23 (398 mg, 657 µmol) and Pd(OAc)₂ (57.8 mg, 257 µmol) in CH₂Cl₂ (12.0 mL) was added Et₃N (300 µL, 2.16 mmol) at room temperature. After stirring for 30 min at reflux, Et₃SiH (1.10 mL, 6.90 mmol) was added, and the mixture. was stirred for 40 min at reflux. The reaction mixture was diluted by CH2Cl2 and the reaction was quenched with sat. aq. NaHCO₃. The separated aqueous phase was extracted with CH₂Cl₂ (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 6:1 to 2:1) to afford amine 24 (347 mg, quant). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.90 (1H, d, J = 8.4 Hz), 7.22– 7.16 (3H, m), 7.03 (2H, d, J = 6.8 Hz), 6.25 (1H, d, J = 2.0 Hz), 6.22 (1H, d, J = 8.0 Hz), 4.80 (1H, ddd, J = 10.8, 3.2, 3.2 Hz), 4.71 (1H, dd, J = 8.0, 7.8 Hz), 4.27 (1H, dd, J = 7.8, 2.2 Hz), 3.79 (1H, dd, J = 7.2, 5.6 Hz), 3.70 (3H, s), 3.55 (1H, s), 3.19 (1H, dd, J = 14.0, 5.8 Hz), 3.01 (1H, dd, J = 14.0, 6.6 Hz), 2.73-2.71 (2H, m), 2.28 (1H, s), 0.82 (9H, s), 0.033 (3H, s), 0.022 (3H, s); ¹³C NMR (100 MHz, CDCl₃): δ 173.0, 171.9, 145.5, 135.8, 133.1, 129.1, 128.3, 126.9, 120.5, 105.6, 67.9, 63.9, 60.9, 52.5, 52.1, 37.6, 35.0, 25.6, 17.9, -3.8, -4.7; IR (film): 3349, 3029, 2952, 2928, 2893, 2855, 2360, 2341, 1745, 1677, 1644, 1508, 1437, 1360, 1344, 1254, 1200, 1079, 961, 837, 779, 702 cm⁻¹; $[\alpha]_D^{26} = -195$ (c = 1.65, CHCl₃); HRMS (ESI) m/z: calcd. for $C_{25}H_{37}N_2O_5Si^+$ [M+H⁺] 473.2466, found 473.2451.

4.32 Dithiodiketopiperazine 25

To a solution of amine 24 (447 mg, 945 µmol) in MeOH (250 mL) was added 25% aq. NH₃ (15.0 mL) at room temperature. After stirring for 2.7 days at room temperature, the reaction was concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes: EtOAc = 2:1) to afford diketopiperazine 25 (202 mg, 459 μ mol, 48%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.33–7.18 (5H, m), 6.41 (1H, s), 6.29 (1H, d, J = 8.0 Hz), 5.90 (1H, s), 5.00 (1H, d, J = 7.6 Hz), 4.88 (1H, dd, J = 8.0, 7.6 Hz), 4.77 (1H, s), 4.23–4.16 (2H, m), 3.51 (1H, dd, J = 14.0, 3.6 Hz), 2.89–2.79 (2H, m), 2.52 (1H, dd, J = 13.0, 13.0 Hz), 0.79 (9H, s), 0.011 (3H, s), 0.006 (3H, s); ¹³C NMR (100 MHz, CDCl₃): δ 168.5, 163.9, 145.0, 135.6, 135.1, 129.3, 128.9, 127.3, 112.4, 106.4, 66.9, 63.1, 57.9, 56.2, 37.3, 33.4, 25.7, 17.8, -4.3, -4.5; IR (film): 3381, 3228, 3220, 2952, 2928, 2895, 2855, 2359, 2341, 1684, 1663, 1444, 1422, 1294, 1256, 1202, 1139, 1083,

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CHCl₃); HRMS (EI) m/z: calcd. for $C_{24}H_{32}N_2O_4Si$ [M⁺] 440.2131, found 440.2134.

4.33 Dithiodiketopiperazine 27

To a solution of diketopiperazine 25 (63.4 mg, 144 µmol) in THF (630 $\mu L)$ was added LiHMDS (1.3 M solution in THF, 120 µL, 156 µmol) at -40 °C. After stirring for 8 min, AllomCl (26) [20] (120 μ L, 1.16 mmol) was added at -40 °C, and the mixture was stirred at -20 °C for 17 h. The reaction was quenched with sat. aq. NH₄Cl solution and the mixture was diluted by water. The separated aqueous phase was extracted with CH₂Cl₂ (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 4:1 to 2:1) to afford diketopiperazine 27 (66.3 mg, 120 µmol, 84%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.28–7.22 (3H, m), 7.04 (2H, d, J = 8.0 Hz), 6.28 (1H, d, J = 7.6 Hz), 6.27 (1H, s), 5.96–5.87 (1H, m), 5.51 (1H, d, J = 10.8 Hz), 5.31 (1H, dt, J = 17.2, 1.6 Hz), 5.24 (1H, dt, J = 10.8, 1.2 Hz), 4.87(1H, dd, J = 7.8, 7.6 Hz), 4.82 (1H, d, J = 7.8 Hz), 4.79 (1H, d, J = 10.8 Hz), 4.62 (1H, s), 4.57 (1H, m), 4.12 (1H, dd, J = 12.8, 6.0 Hz), 4.07–4.40 (2H, m), 3.42 (1H, dd, J = 14.0, 2.8 Hz), 3.21 (1H, dd, J = 14.0, 2.8 Hz), 2.44 (1H, dd, J = 13.2, 6.0 Hz), 0.94 (1H, dd, J = 13.2, 12.8 Hz), 0.79 (9H, s), 0.007 (3H, s), -0.009 (3H, s); ¹³C NMR (100 MHz, CDCl₃): § 166.2, 163.7, 144.9, 134.9, 134.7, 133.4, 130.1, 128.6, 127.4, 118.1, 112.8, 106.4, 72.5, 69.9, 68.6, 62.9, 59.1, 58.2, 36.5, 33.6, 25.8, 18.0, -4.3, -4.5; IR (film): 2952, 2928, 2856, 1662, 1452, 1441, 1295, 1200, 1138, 1092, 1051, 955, 906, 837, 777, 703 cm⁻¹; $[\alpha]_D^{25} = -333$ (*c* = 0.535, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₈H₃₈N₂NaO₅Si⁺[M+Na⁺] 533.2442, found 533.2417.

4.34 Dithiodiketopiperazine 28

To a solid of diketopiperazine 27 (66.3 mg, 120 µmol) was added HF·Py in pyridine (2.00 mL, 1:4, HF·Py containing ~70% hydrogen fluoride and ~30% pyridine) at room temperature. After stirring for 17 h at 45 °C, the reaction mixture was diluted by water at 0 °C. The separated aqueous phase was extracted with CH2Cl2 (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography ($CH_2Cl_2:EtOAc = 1:5$) to afford allyl alcohol 28 (46.6 mg, 118 µmol, 98%). A white solid; mp: 155.5–156.5 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.28–7.22 (3H, m), 7.07–7.04 (2H, m), 6.34 (1H, d, J = 8.0 Hz), 6.34 (1H, d, J = 2.0 Hz), 5.98–5.91 (1H, m), 5.41 (1H, d, J = 10.8 Hz), 5.35 (1H, ddd, J = 17.5, 2.9, 1.7 Hz), 5.24 (1H, ddd, J = 10.3, 2.9, 1.5 Hz), 5.05 (1H, dd, J = 8.2, 8.0 Hz), 4.82 (1H, d, J = 10.8 Hz), 4.79–4.76 (2H, m), 4.69 (1H, s), 4.58–4.56 (1H, m), 4.22 (1H, dd, J = 12.2, 6.0 Hz), 4.16–4.10 (2H, m), 3.42 (1H, dd, J = 14.2, 3.0 Hz), 3.23 (1H, dd, J = 14.2, 4.4 Hz), 2.48 (1H, dd, J = 14.4, 6.0 Hz), 0.94 (1H, dd, J = 14.4, 12.2 Hz); ¹³C NMR (100 MHz, CDCl₃): § 166.0, 163.8, 145.1, 135.2, 134.7, 133.6, 130.1, 128.4, 127.3, 117.8, 113.7, 105.7, 73.2, 70.3, 68.1, 62.6, 59.8, 58.3, 36.8, 33.3; $[\alpha]_D^{25} = -367$ (*c* = 0.260, CHCl₃); IR (film): 3425, 3419, 2921, 2359, 2340, 1694, 1659, 1454, 1339, 1296, 1200, 1137, 1074, 1047, 929, 857, 758, 736, 704 cm⁻¹; HRMS (ESI) m/z: calcd. for C₂₂H₂₄N₂NaO₅ [M+Na⁺] 419.1577, found 419.1558.

4.35 Dithiodiketopiperazine 29

To a solution of allyl alcohol $28~(74.3~mg,\,188~\mu mol)$ and nor-AZADO (8.50 mg, 61.5 $\mu mol)$ in dry $CH_2Cl_2~(1.40~mL)$ was

After stirring for 1.5 h at room temperature, the reaction was quenched with sat. aq. Na_2SO_3 solution and water (1:2). The separated aqueous phase was extracted with CH_2Cl_2 (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes: EtOAc = 1:3) to afford lactone 29 (71.0 mg, 180 µmol, 96%). A white foam; ¹H NMR (400 MHz, CDCl₃): δ 7.28–7.22 (3H, m), 7.09 (2H, d, *J* = 8.0 Hz), 7.00 (1H, d, J = 7.6 Hz), 6.59 (1H, d, J = 2.4 Hz), 6.01–5.91 (1H, m), 5.61 (1H, d, J = 7.6 Hz), 5.36 (1H, d, J = 18.4 Hz), 5.35 (1H, d, J = 10.4 Hz), 5.31 (1H, s), 5.27 (1H, d, J = 10.4 Hz), 4.90 (1H, d, J = 10.4 Hz), 4.68 (1H, dd, J = 4.0, 3.4 Hz), 4.20–4.10 (2H, m), 3.91 (1H, dd, J = 13.4, 6.0 Hz), 3.43 (1H, dd, J = 14.0, 3.4 Hz), 3.29(1H, dd, J = 14.0, 4.0 Hz), 2.53 (1H, dd, J = 15.2, 6.0 Hz), 1.01 (1H, dd, J = 13.4, 13.4 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 180.9, 164.9, 163.6, 153.3, 136.9, 134.5, 133.4, 130.0, 128.3, 127.1, 117.6, 115.4, 107.8, 73.5, 70.4, 66.3, 60.0, 57.0, 36.9, 31.8; IR (film): 3028, 2925, 2867, 1687, 1667, 1604, 1454, 1350, 1327, 1292, 1204, 1078, 1051, 940, 930, 864, 820, 758, 736 cm⁻¹; $[\alpha]_D^{26} = -279$ (*c* = 0.445, CHCl₃); HRMS (ESI) *m/z*: calcd. for C₂₂H₂₃N₂O₅ [M+H⁺] 395.1601, found 395.1605.

4.36 Dithiodiketopiperazine 30

To a solution of lactone 29 (138 mg, 350 µmol) in CH₂Cl₂ (750 µL) and EtOH (500 µL) was added CeCl₃· 7H₂O (381 mg) in EtOH (2.0 mL) at -78 °C. After stirring for 10 min, NaBH₄ in EtOH (22.0 mg, 582 μ mol in 550 μ L) was added at -78 °C, and the mixture was stirred at -78 °C for 1.6 h. The reaction was quenched with aq. NH₄Cl solution at room temperature. The separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 1:3) to afford allyl alcohol 30 (126 mg, 318 µmol, 91%). A white foam; ¹H NMR (400 MHz, CDCl₃): δ 7.31–7.23 (3H, m), 7.08 (2H, d, *J* = 6.4 Hz), 6.27 (1H, dd, J = 2.2, 2.2 Hz), 6.13 (1H, dd, J = 8.0, 2.4 Hz), 6.00-5.90 (1H, m), 5.42 (1H, d, J = 10.8 Hz), 5.36 (1H, d, J = 17.2 Hz), 5.26 (1H, d, J = 10.0 Hz), 5.07 (1H, d, J = 3.6 Hz), 4.88 (1H, s), 4.87 (1H, d, J = 10.8 Hz), 4.64 (1H, dd, J = 3.0, 4.2 Hz), 4.52 (1H, d, J = 7.6 Hz), 4.20–4.17 (1H, m), 4.14 (1H, d, J = 5.8 Hz), 4.13 (1H, d, J = 5.8 Hz), 3.91 (1H, dd, J = 13.0, 5.8 Hz), 3.40 (1H, dd, J = 13.8, 3.0 Hz), 3.27 (1H, dd, J = 13.8, 4.2 Hz), 2.36 (1H, dd, J = 13.6, 5.8 Hz), 0.80 (1H, dd, J = 13.6, 13.0 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 167.0, 165.1, 138.0, 137.0, 134.2, 133.4, 130.2, 128.7, 127.7, 118.1, 109.8, 109.7, 73.2, 70.5, 69.1, 64.1, 59.8, 56.3, 37.0, 33.7; IR (film): 3369, 3011, 2942, 2863, 1661, 1453, 1344, 1296, 1193, 1133, 1078, 1058, 930, 759, 756, 730, 704, 578 cm⁻¹; $[\alpha]_D^{25} = -229$ (c = 1.04, CHCl₃); HRMS (ESI) *m/z*: calcd. for C₂₂H₂₄N₂NaO₅ [M+Na⁺] 419.1577, found 419.1564.

4.37 Dithiodiketopiperazine 31

To a suspension of sulfur S₈ (63.6 mg, 247 µmol) in Et₂O (1.0 mL) was added LiHMDS (1.3 M in THF, 725 µL, 943 µmol) at room temperature. After stirring for 5 min, diketopiperazine **30** (18.6 mg, 47.0 µmol) in a mixture of THF (2.0 mL) and Et₂O (1.0 mL) was added at room temperature, and the resulting solution was stirred for 1 min. Then, LiHMDS (1.3 M in THF, 725 µL, 943 µmol) was added, and the mixture was stirred for 30 min at room temperature. The reaction was quenched with aq. NH₄Cl and diluted with water. The separated aqueous phase was extracted with Et₂O (x1) and CH₂Cl₂ (x2). The combined organic

and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 16:1x2, then hexanes: $CH_2Cl_2 = 3:5$) to afford the mixture of bridged polysulfides. To a solution of the polysulfides in a mixture of THF (3.2 mL) and EtOH (3.2 mL) was added NaBH₄ (36.5 mg, 965 $\mu mol)$ at 0 °C. After stirred for 8 min at room temperature, MeI (1.60 mL, 25.6 mmol) was added at 0 °C, and the resulting mixture was stirred at room temperature for 4 h. The reaction was quenched by aq. NH₄Cl, and the separated aqueous phase was extracted with Et₂O (x1) and CH₂Cl₂ (x2). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 16:1, then hexanes: EtOAc = 2:1) to afford bis-(methylthio) diketopiperazine **31** (18.3 mg, 30.3 µmol, 65% 2 steps from **30**). A white foam; ¹H NMR (400 MHz, CDCl₃): δ 7.29–7.24 (3H, m), 7.15–7.12 (2H, m), 6.31 (1H, m), 6.17 (1H, dd, J = 8.4, 2.4 Hz), 6.03–5.93 (1H, m), 5.40–5.34 (1H, m), 5.53 (1H, d, J = 9.6 Hz), 5.24–5.22 (2H, m), 5.12 (1H, d, *J* = 9.6 Hz), 4.91 (1H, dd, *J* = 8.2, 1.8 Hz), 4.58–4.55 (1H, m), 4.42 (1H, d, J = 8.0 Hz), 4.30– 4.28 (2H, m), 3.48 (1H, d, J = 13.2 Hz), 3.23 (1H, d, J = 13.2Hz), 2.55 (1H, d, J = 15.2 Hz), 2.29 (3H, s), 2.53 (3H, s), 1.59 (1H, d, J = 15.2 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 167.1, 166.1, 137.6, 137.2, 134.3, 133.6, 130.7, 128.6, 127.9, 117.2, 110.8, 107.1, 77.5, 73.4, 72.2, 71.3, 67.7, 65.4, 45.0, 38.1, 15.2, 14.0; IR (film): 3370, 3008, 2920, 2852, 1662, 1435, 1416, 1380, 1191, 1136, 1083, 1056, 1050, 772, 756, 704 cm⁻¹; $[\alpha]_D^{25} = -123$ $(c = 0.619, \text{CHCl}_3)$; HRMS (ESI) m/z: calcd. for $C_{24}H_{28}N_2NaO_5S_2$ [M+Na⁺] 511.1332, found 511.1303.

4.38 (-)-Emestrin H (5)

To a solid of bis-(methylthio) diketopiperazine **31** (6.65 mg, 13.6 µmol) was added Pd(PPh₃)₄ (27.2 mM solution in CH₂Cl₂, 25.0 µL 0.682 µmol). The mixture was concentrated under reduced pressure give a pale brown solid. To the solid was added 1,3-dimethylbarbituric acid (4.26 mg, 27.3 µmol) in MeOH (136 μ L), and the mixture was stirred for 16.5 h at 40 °C. The reaction mixture was diluted with MeOH (68 µL) and stirred for 2 h. Then, the mixture was cooled to room temperature and diluted with MeOH (200 μ L). To the mixture was added NaHCO₃ (40.0 mg), and the resulting suspension was stirred for 25 min at room temperature and diluted with CH₂Cl₂ and H₂O. The separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude by preparative material, which was purified TLC (hexanes:EtOAc = $3:1 \times 3$) to afford (-)-emestrin H (5) (4.46 mg, 10.6 µmol, 78%) and diketopiperazine 30 (1.15 mg, 17%). A white solid; mp: 86.0–87.2 °C; ¹H NMR (600 MHz, CDCl₃): δ 7.34–7.30 (3H, m), 7.19 (2H, d, J = 6.6 Hz), 6.36 (1H, s), 6.19 (1H, dd, J = 1.8, 8.4 Hz), 6.05 (1H, s), 5.12 (1H, d, J = 4.8 Hz),4.93 (1H, d, J = 8.4 Hz), 4.63–4.61 (1H, m), 4.55 (1H, d, J = 7.2 Hz), 3.64 (1H, d, J = 13.2 Hz), 3.02 (1H, d, J = 13.2 Hz), 2.63 (1H, d, J = 15.0 Hz), 2.41 (3H, s), 2.23 (3H, s), 1.88 (1H, d, J = 15.0 Hz); ¹³C NMR (150 MHz, CDCl₃): δ 167.2, 165.4, 137.7, 137.4, 133.2, 130.7, 128.8, 128.1, 110.9, 107.4, 72.3, 68.5, 68.4, 64.9, 46.3, 38.9, 14.7, 14.3; IR (film): 3361, 3224, 3062, 3008, 2918, 2848, 2360, 2341, 1675, 1638, 1409, 1340, 1193, 1139, 1123, 1047, 756, 730, 704, 581, 564 cm⁻¹; $[\alpha]_D^{28} = -203$ (c = 0.234, MeOH) (Lit. $[\alpha]_D^{25} = -97$ (c = 0.071, MeOH)); HRMS (ESI) m/z: calcd. for C₂₀H₂₂N₂NaO₄S₂ [M+Na⁺] 441.0913, found 441.0907. The spectral data were identical with those reported in the literature [14].

4.39 (-)-Asteroxepin (4)

DMAP (1.23 mg, 10.1 µmol) in pyridine (120 µL) was added Ac₂O in pyridine (0.127 M, 100 µL, 12.7 µmol) at room temperature. After stirring for 50 min at room temperature, additional Ac₂O (13.0 µL, 138 µmol) was added, and the mixture was stirred for 35 min. Then, additional DMAP (5.69 mg, 46.6 µmol) was added, and the mixture was stirred for 45 min at room temperature. The reaction was diluted with CH2Cl2, and quenched with aq. 1 M HCl. The separated aqueous phase was extracted with CH₂Cl₂ (x5), and the combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 7:11) to afford (-)-asteroxepin (4) (2.60 mg, 5.65 µmol, 89%). A white solid; mp: 189.6–190.0 °C; ¹H NMR (600 MHz, CDCl₃): δ = 7.32–7.27 (3H, m), 7.20–7.18 (2H, m), 6.41 (1H, dd, J = 2.4, 2.4 Hz), 6.29 (1H, dd, J = 8.4, 3.0 Hz), 6.00 (1H, br s), 5.88 (1H, ddd, *J* = 7.5, 2.1, 2.1), 4.89 (1H, br. d, *J* = 7.8 Hz), 4.68 (1H, dd, *J* = 8.4, 1.2 Hz), 3.66 (1H, d, *J* = 13.8 Hz), 2.97 (1H, d, *J* = 13.8 Hz), 2.64 (1H, d, *J* = 15.0 Hz), 2.36 (3H, s), 2.20 (3H, s), 2.13 (3H, s), 1.82 (1H, ddd, J = 15.3, 2.1, 2.1 Hz); ¹³C NMR (150 MHz, CDCl₃): $\delta = 170.1$, 165.6, 164.4, 139.7, 137.3, 133.5, 130.9, 128.7, 127.9, 109.1, 105.5, 71.4, 69.9, 68.0, 61.7, 46.4, 39.6, 21.2, 14.5, 14.0; $[\alpha]_D^{27} = -178$ (c = 0.556, CHCl₃) (Lit. $[\alpha]_D = -102$ (c = 0.4, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₂H₂₄N₂NaO₅S₂ [M+Na⁺] 483.1019, found 483.0996. The spectral data were identical with those reported in the literature [13].

4.40 AllomCl (26) [20]

To a mixture of allyl alcohol (40.0 mL, 0.588 mmol) and paraformaldehyde (19.0 g, 0.633 mol) was added anhydrous hydrogen chloride generated from H_2SO_4 and NaCl at room temperature. After stirred for 30 min at room temperature, the mixture was heated by oil bath at 150 °C for 2 h. The resulting mixture was diluted with hexanes, dried over magnesium sulfate, and the hexanes were removed under reduced pressure. The crude material was distilled from anhydrous calcium chloride (37.0 °C/67.5 mmHg) to afford AllomCl (26) (25.0 g, 0.234 mol, 40%) as colorless oil.

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16. Compounds **16a** and **16b** with C3 (*S*) and C8a (*S*) stereochemistries were synthesized by the stepwise condensation of *N*-Cbz-L-proline and *N*-PMB-L-phenylalanine methyl ester or *N*-2,4-DMB-phenylalanine methyl ester, respectively.



17. Compounds **16c–e** were synthesized from *trans*-**13** with C3 (*S*) and C8a (*R*) stereochemistries, which was prepared by the stepwise condensation of *N*-Boc-L-proline and L-phenylalanine methyl ester. During the intramolecular condensation step, the C8a (*S*) stereochemistry was completely isomerized to give *trans*-**13** with C8a (*R*) stereochemistry.



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16c-16e

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Total syntheses of (-)-emestrin H and (-)-asteroxepin

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ABSTRACT

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Keywords: total synthesis alkaloids diketopiperazine amide protecting group First total syntheses of (–)-emestrin H and (–)-asteroxepin were accomplished. The allyloxymethyl (Allom) group, suitable protecting group for the amide nitrogen of the diketopiperazine which needs to be robust enough to tolerate Nicolaou's sulfenylation condition but also easily cleavable by the exceptionally mild allylic alkylation with Pd/N,N-dimethylbarbituric acid system, leaving acid, basic, oxidation labile methylthio groups intact was disclosed. The first total synthesis of (–)-emestrin H was accomplished via installation of two methylthio groups to the diketopiperazine core having Allom group, followed by removal of the Allom group at the final step. The total synthesis of (–)-asteroxepin was also completed by acylation of (–)-emestrin H.

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1. Introduction

Dithiodiketopiperazine alkaloids bearing the sulfur-containing diketopiperazine as core structure have attracted attention due to their structural diversity and fascinating biological activities [1,2]. Among them, compounds bearing the 4,5-dihydrooxepine ring, such as (-)-acetylaranotin (1) [3], (+)-MPC1001B (7) [4], and (-)-acetylapoaranotin (6) [5], have been challenging synthetic targets and a lot of effort has been made to synthesize these compounds [6,7]. Reisman et al. reported the first innovative total synthesis of (-)-acetylaranotin (1) in 2012 [8], followed by the first total synthesis of (-)-acetylapoaranotin (6) [10]. We also focused on this class of compounds, reporting the second total synthesis of 1 just after Reisman et al. in 2012 [9]. Using the synthetic protocol established during the synthetic studies on 1, which was based on the characteristic proline-fused 4,5-dihydrooxepine ring, we successfully performed the first total synthesis of (+)-MPC1001B (7) [11] and determined the structure of (-)-SCH64874 (3) and hirsutellomycin (8) via semi-synthesis [12]. Despite these synthetic developments over the last decade, the total syntheses of (-)-asteroxepin (4) [13] and (-)-emestrin H (5) [14] bearing the NH-free diketopiperazine core have not been reported so far.

Our preliminary model studies toward 4 and 5 (Scheme 1a) revealed that the enolate-mediated introduction of two methylthio groups reported by Nicolaou *et al.* [15] was unsuccessful for the unprotected diketopiperazine 13 due to the competitive deprotonation of the unprotected N–H proton (Scheme 1b). Therefore, the major challenge toward the synthesis of 4 and 5 was the selection of the appropriate protecting group for the

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diketopiperazine ring's NH group. The ideal protecting group (PG in Scheme 1) should be robust enough to tolerate the strong basic conditions during the introduction of the two-methylthio groups. Moreover, it should be easily and chemoselectively cleaved under the particularly mild conditions in the final stage of the total synthesis without affecting the base-, nucleophile-, and oxidant-sensitive methylthio groups and the acid-sensitive dihydrooxepin structure. However, compared to the many available protecting groups for amines, there are limited choices for amides, especially for diketopiperazines. In this study, we



Fig. 1. Dithiodiketopiperazine alkaloids.

Tetrahedron





Scheme 1. Synthetic plan and preliminary results.

2. Results and discussion

To identify the most suitable protecting group that would fulfill the abovementioned requirements, we prepared a series of *N*-protected diketopiperazines (**16a–e**) following a conventional method [16,17], which were then subjected to the sulfenylation conditions established by Nicolaou *et al.* (Table 1) [15]. Among the tested analogues **16a–e**, substrates **16a** and **16b** bearing a stable benzylic protecting group, such as *p*-methoxybenzyl (PMB) or 2,4-dimethoxybenzyl (2,4-DMB), were not affected by the sulfenylation conditions, providing dithiodiketopiperazines **17a** and **17b**, respectively, in moderate yields (Table 1, entries 1 and 2). Moreover, substrates protected as *N,O*-acetals, such as**16d** with the trimethylsilylethoxymethyl (SEM) group and **16e**

Table 1

Protecting groups and introduction of the methylthio group.



^aC3 (*S*), C8a (*S*) isomer. ^bC3 (*S*), C8a (*R*) isomer. ^cIsolated yield. ^dObtained as single diastereomer.

compounds (**17d** and **17e**) in good yields (Table 1, entries 4 and 5). However, analogue **16c**, protected as urethane with an allyloxycarbonyl group (Alloc), afforded a complex mixture under the same sulfenylation conditions (Table 1, entry 3).

The deprotection of the bis(methylthio) products 17a, 17b, 17d, and 17e was examined under the most frequently used deprotection conditions for each protecting group (Table 2). Analogues 17a and 17b bearing PMB and 2,4-DMB groups were decomposed upon treatment with ceric ammonium nitrate (CAN) and 2.3-dichloro-5.6-dicvano-1.4-benzoquinone (DDO). respectively (Table 2, entries 1 and 2). Moreover, the SEM group in 17d was stable upon treatment with HF/pyridine and 17d was recovered (Table 2, entry 4), whereas the use of TBAF in THF/DMF decomposed 17d (Table 2, entry 3). In contrast to these unsuccessful entries, the Allom group in 17e could be smoothly removed by a combination of $Pd(PPh_3)_4$ and N,Ndimethylbarbituric acid (DMBA) in MeOH yielding 15 in 56% yield, while a substantial amount of the hemiaminal intermediate 18 was also obtained (Table 2, entry 5) [18,19]. The subsequent in situ treatment of the reaction mixture with aq. NH₃ was effective in reducing the amount of 18, affording 15 in 87% yield (Scheme 2). Furthermore, the addition of solid NaHCO₃ after the consumption of 17e improved the yield of 15 to up to 95%.

Table 2

Deprotection conditions.



^aIsolated yield. ^bDecomposition. ^c**17d** was completely recovered. ^dReaction conditions: **17** (1.0 eq), Pd(PPh₃)₄ (45 mol%), DMBA (5.5 eq), MeOH (0.1 M), 30°C, 6 h. The hemiaminal intermediate **18** was obtained in 38% yield.



Scheme 2. Removal of the Allom group.

Feasibility of the deprotection condition of the Allom group from secondary amides was investigated using a series of *N*-Allom secondary amides (**20**) (see Experimental Section for the preparation and additional data) (Scheme 3). Treatment of *N*-Allom benzamides **20a**–**e** and γ -lactam **20f** with Pd(PPh₃)₄ and DMBA in MeOH smoothly provided the hemiaminal intermediate **21**, which was converted into amides **19a–e** and **19f**, respectively, in good yields using aq. NH₃ (condition A). In contrast, the reaction of *N*-Allom aliphatic amides **20g** under the Pd/*N*,*N*-dimethylbarbituric acid/MeOH condition provided a mixture of **19g** (5.3%), **21g** (21%), and *N*- due to an exchange of the OH group in intermediate 21g with methanol. This side reaction was eliminated by performing the reaction in THF and H₂O, providing the hemiaminal intermediates 21g, which were then converted to amides 19g in good yields using trimethylsilyl iodide (TMSI) (condition B).



Reaction conditions: Condition A: **20** (1.0 eq), Pd(PPh₃)₄ (10 mol%), DMBA (2.0 eq), MeOH (0.3 M), 40°C, 6 h, then aq. NH₃ (excess). Condition B: **20g** (1.0 eq), Pd(PPh₃)₄ (10 mol%) , DMBA (2.0 eq), THF/H₂O (3:1, 0.3 M), then **21g** (1.0 eq), TMSCI (7.0 eq), NaI (7.0 eq), CH₃CN (0.2 M). The yields refer to the isolated products.

Scheme 3. Removal of the Allom group under different conditions.

The applicability of the Allom protecting group was fully demonstrated by the first total syntheses of (–)-emestrin H (5) and (–)-asteroxepin (4). The proline-fused dihydrooxepine derivative 9, which was prepared according to our previously established protocol in the total synthesis of (–)-acetylaranotin (1) [9], was first condensed with the (S)-phenylalanine methyl ester (10) (Scheme 4). The benzyloxycarbonyl (Cbz) group was then removed under transfer hydrogenation conditions and the formation of the tricyclic diketopiperazine derivative 25 was promoted upon treatment of the resulting aminoester with ammonium hydroxide. Subsequently, the Allom group was introduced using a combination of the allyloxymethyl chloride



Scheme 4. Preparation of diketopiperazine 30.

(LiHMDS), which afforded the desired product **27** in 84% yield. After desilylation, the stereochemistry of the C-10 hydroxyl group was inverted through a stepwise process [9], which included an oxidation of **28** to ketone using PhI(OAc)₂ and 9-azanoradamantane *N*-oxyl (nor-AZADO) [21] and a subsequent Luche reduction, which gave the desired alcohol **30** as the sole isomer.

The final steps toward the synthesis of (–)-emestrin H (**5**) and (–)-asteroxepin (**4**) including the crucial introduction of the two methylthio groups and the removal of the Allom protecting group are depicted in Scheme 5. According to the model study (*vide supra*), diketopiperazine **30** was treated with a mixture of excess LiHMDS and S₈ to form an epipolysulfide. Its reduction to the corresponding dithiol with NaBH₄ and the subsequent methylation afforded the bis(methylthio) derivative **31** in 65% yield over two steps from **30**. Finally, the Allom group was removed under the optimized conditions affording (–)-emestrin H (**5**) in 78% yield, which was further acetylated to give (–)-asteroxepin (**4**). All properties of synthetic (–)-emestrin H (**5**) and (–)-asteroxepin (**4**) were identical with those reported except magnitude of specific rotations proving the reported absolute stereochemistry of **4** and **5** by this total synthesis [22].



Scheme 5. Final steps of the total syntheses of (–)-emestrin H (5) and (–)-asteroxepin (4).

3. Conclusion

In conclusion, the total syntheses of (–)-emestrin H (5) and (–)-asteroxepin (4) have been accomplished for the first time. Their preparation was successful owing to the suitability of the Allom group as protecting group for the amide nitrogen of the highly functionalized dithio-diketopiperazine intermediate. This protecting group could be easily cleaved even in the presence of the acid-sensitive dihydrooxepin structure and the dimethylthio groups, which are sensitive to bases, nucleophiles, and oxidants. Thus, this study demonstrated the application potential of the Allom group not only for the synthesis of other N-unprotected diketopiperazine alkaloids, but also for the preparation of various functionalized amides.

4. Experimental section

4.1 General method

Materials were obtained from commercial suppliers and used without further purification unless otherwise mentioned. Anhydrous THF, Et_2O , CH_2Cl_2 , toluene, DMF, and MeCN were purchased from commercial suppliers. Anhydrous MeOH was dried and distilled according to the standard protocols. All reactions were carried out under Ar atmosphere unless otherwise mentioned. Flash column chromatography was performed on Silica Gel 60N (Kanto, spherical neutral, 40–50 µm). Preparative 4

ΤI glass plates precoated with a 0.25 mm thickness of silica gel. NMR spectra were recorded on a JNM-AL400 spectrometer and a JEOL ECA600 spectrometer. Chemical shifts for ¹H NMR are reported in parts per million (ppm) downfield from tetramethylsilane as the internal standard and coupling constants are in Hertz (Hz). The following abbreviations are used for spin multiplicity: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, and br = broad. Chemical shifts for ${}^{13}C$ NMR are reported in ppm, relative to the central line of a triplet at 77.0 ppm for CDCl₃ or a septet at 39.5 ppm for DMSO-d₆. IR spectra were measured on a SHIMADZU FTIR-8300 spectrometer. Mass spectra were recorded on a Bruker micrOTOF (ESI). Optical rotations were measured on a Horiba SEPA-300 highly sensitive polarimeter. Compounds 9 [9], 19a-e [25], 19f [26], and 19g [25] were prepared according to the procedures reported in references.

4.2 Diketopiperazine16a

To a solution of N-Cbz-L-proline (3.68 g, 14.8mmol) and N-PMB-L-phenylalanine methyl ether (4.24 g, 14.2 mmol) in Et₃N and CH₂Cl₂ (5 mL and 30 mL) was added BOP-Cl (3.74 g, 14.7 mmol) at room temperature. After stirring for 24.5 h at room temperature, additional BOP-Cl (1.96 g, 7.70 mmol) was added, and the reaction mixture was stirred for 2.5 h. The reaction was quenched with sat. aq. NaHCO3, and the aqueous phase was extracted with CH2Cl2 (x3). The combined organic layers were washed with 1 M aq. HCl, dried over Na2SO4, filtered and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 1:1) to afford amide 32a (5.80 g, 10.9 mmol, 77%). To a suspension of amide 32a (5.80 g, 10.9 mmol) and Pd(OAc)₂ (983 mg, 4.38 mmol) in CH₂Cl₂ (55.0 mL) was added Et₃N (3.08 mL, 21.9 mmol) at room temperature. After stirring at reflux for 5 min, Et₃SiH (8.74 mL, 54.7 mmol) was added and the mixture was stirred at reflux for 3.5 h. The reaction mixture was diluted with CH₂Cl₂ and the reaction was quenched with sat. aq. NaHCO₃. The separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (EtOAc) to afford diketopiperazine 16a (3.00 g, 8.23 mmol, 75%). A white foam; The spectral data of 16a were identical with those reported in the literature [23].

4.3 Diketopiperazine16b

To a solution of N-Cbz-L-proline (1.06 g, 4.25 mmol), N-2,4-DMB-L-phenylalanine methyl ether (1.15 g, 3.49 mmol) in Et₃N and CH₂Cl₂ (2.0 mL and 20.0 mL) was added BOP-Cl (1.87 g, 7.35 mmol) at room temperature. After stirring for 20 h at room temperature, the reaction was quenched with 1 M aq. HCl and the resulting mixture was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material. which was purified by silica gel column chromatography (hexanes: EtOAc = 3:2 to 6:5) to afford amide **32b** (846 mg, 1.51 mmol, 47%). To a suspension of amide **32b** (846 mg, 1.51 mmol) and Pd(OAc)₂ (135 mg, 601 µmol) in CH₂Cl₂ (10.0 mL) was added Et₃N (425 µL, 3.02 mmol) at room temperature. After stirring at reflux for 7 min, Et₃SiH (1.20 mL, 7.51 mmol) was added and the mixture was stirred at reflux for 3.5 h. The reaction mixture was diluted with CH₂Cl₂ and the reaction was quenched with sat. aq. NaHCO₃. The separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate,

material, which was purified by silica gel column chromatography (EtOAc) to afford diketopiperazine 16b (540 mg, 1.37 mmol, 91%). A white foam; ¹H NMR (400 MHz, CDCl₃): δ 7.31-7.20 (4H, m), 7.12-7.06 (2H, m), 6.48-6.43 (2H, m), 5.40 (1H, d, J = 14.8 Hz), 4.32 (1H, m), 4.25 (1H, d, J =14.8 Hz), 3.84 (3H, s), 3.81 (3H, s), 3.72 (1H, dd, J = 12.0, 6.0 Hz), 3.68-3.61 (1H, m), 3.36-3.27 (2H, m), 3.14-3.07 (1H, m), 1.91–1.81 (1H, m), 1.60–1.42 (2H, m), 0.14–0.01 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 165.3, 163.9, 160.7, 158.7, 135.1, 131.7, 130.0, 128.3, 127.1, 115.9, 104.4, 98.3, 60.3, 59.0, 55.3, 55.3, 44.0, 40.6, 36.4, 28.4, 20.9; IR (film): 2992, 2952, 2944, 2891, 1656, 1612, 1587, 1508, 1456, 1298, 1289, 1261, 1208, 1184, 1157, 1033, 751, 703 cm⁻¹; $[\alpha]_D^{31} = -116$ (c = 0.66, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₃H₂₆N₂NaO₄ [M⁺+Na] 417.1785, Found 417.1769.

4.4 Amide 33

To a solution of N-Boc-L-proline (10.8 g, 50.0 mmol) and Lphenylalanine methyl ether (8.95 g, 50.0 mmol) in CH₂Cl₂ (200 mL) were added Et₃N (27.9 mL, 200 mmol), HOBt (6.99 g, 51.7 mmol), and EDCI/HCl (9.91 g, 51.7 mmol) at room temperature. After stirred for 29.5 h at room temperature, the organic phase was washed with 1 M aq. HCl (x3) and sat. aq. NaHCO₃ (x2). The resulting organic phase was dried over Na₂SO₄, filtered and concentrated under reduced pressure to give a crude material, which was used for the next reaction without further purification. To a solution of the crude material in CH₂Cl₂ (120 mL) was added TFA (15 mL) at room temperature. After stirred for 20 h at room temperature, the reaction mixture was diluted with CH₂Cl₂ (150 mL) and neutralized by sat. aq. NaHCO₃. The aqueous phase was extracted with CH₂Cl₂ (x3), and the combined organic layers were dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (CH_2Cl_2 :MeOH = 10:1) to afford amine amide 33 (6.76 g, 24.5 mmol, 49%, 2 steps from N-Boc-Lproline). A white solid. The spectral data of 33 were identical with those reported in the literature [24].

4.5 Diketopiperazine trans-13

To a solution of amie 33 (6.76 g, 24.5 mmol) in MeOH (400 mL) was added NaHCO3 (20.0 g, 85.3 mmol) at room temperature. After stirring for 22.5 h at 65 °C, MeOH was removed under reduced pressure. The residue was diluted with CH_2Cl_2 and filtered through a pad of $Celite^{\circledast}$. The filtrate was washed with H₂O, and the aqueous phase was extracted with CH₂Cl₂ (x2). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by crystallization from EtOAc to afford diketopiperazine trans-13 (3.43 g, 14.1 mmol, 57%). The mother liquor was concentrated in vacuo and the second crop was crystallized from EtOAc to afford diketopiperazine trans-13 (643 mg, 2.64 mmol, 11%). A white solid.; ¹H NMR (400 MHz, CDCl₃): δ 7.35-7.20 (5H, m), 6.31 (1H, s), 4.22 (1H, ddd, J = 6.6, 4.0, 3.8 Hz), 3.63 (1H, ddd, J =12.4, 8.5, 8.5 Hz), 3.40 (1H, ddd, J = 12.4, 9.4, 3.0 Hz), 3.15 (1H, dd, J = 13.9, 6.6 Hz), 3.08 (1H, dd, J = 13.9, 4.0 Hz), 2.97(1H, dd, J = 10.4, 6.4 Hz), 2.23-2.15 (1H, m), 1.98-1.90 (1H, m)m), 1.86–1.64 (2H, m); ¹³C NMR (100 MHz, CDCl₃): δ 169.5, 164.8, 135.3, 129.9, 128.5, 127.3, 58.7, 57.6, 44.9, 40.3, 28.8, 21.5; IR (film): 3483, 3238, 3063, 3029, 2981, 2953, 2931, 2886, 1664, 1496, 1454, 1336, 1307, 1296, 1206, 1186, 1115, 1106, 921, 732, 702, 593 cm⁻¹; $[\alpha]_D^{26} = +93.0$ (*c* = 0.200, H₂O); HRMS (ESI) m/z: calcd. for C₁₄H₁₇N₂O₂ [M+H⁺] 245.1285, found

reported in the literature [27].

4.6 Diketopiperazine16c

To a solution of diketopiperazine trans-13 (203 mg, 0.832 mmol) in THF (4.0 mL) was added LiHMDS (1.3 M solution in THF, 650 µL, 845 µmol) at 0 °C. After stirring for 4 min, Alloc-OSu (172 mg, 863 µmol) in THF (1.50 mL) was added at 0 °C. The resulting mixture was stirred for 35 min at room temperature. The reaction was quenched with sat. aq. NH₄Cl, and the resulting mixture was diluted by water. The separated aqueous phase was extracted with CH₂Cl₂ (x3), and the combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (CH₂Cl₂:EtOAc = 5:1 to 3:1) to afford diketopiperazine 16c (248 mg, 789 µmol, 95%). A colorless solid; ¹H NMR (400 MHz, CDCl₃): δ 7.31-7.26 (3H, m), 7.19-7.14 (2H, m), 5.99–5.87 (1H, m), 5.43 (1H, d, J = 17.2 Hz), 5.31 (1H, d, J = 10.8 Hz), 5.01 (1H, ddd, J = 5.0, 5.0, 1.4 Hz), 4.75(1H, ddd, J = 13.3, 5.8, 1.0 Hz), 4.69 (1H, ddd, J = 13.3, 5.6, 1.2 Hz), 3.60-3.50 (1H, m), 3.45-3.37 (1H, m), 3.33-3.22 (2H, m), 2.64 (1H, dd, J = 9.6, 6.8 Hz), 2.17–2.07 (1H, m), 1.96–1.79 (2H, m), 1.72–1.57 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 167.2, 164.0, 151.8, 134.9, 130.7, 129.9, 128.6, 127.5, 119.4, 67.9, 62.3, 58.6, 44.8, 38.2, 29.1, 21.8; IR (film): 3086, 3062, 3028, 2983, 2953, 2885, 1782, 1731, 1672, 1455, 1384, 1270, 1231, 980, 763, 747, 703 cm⁻¹; $[\alpha]_D^{26} = +143$ (*c* = 0.425, CHCl₃); HRMS (ESI) *m/z*: calcd. for C₁₈H₂₁N₂NaO₄ [M+Na⁺] 351.1315, Found 351.1319.

4.7 Diketopiperazine16d

To a solution of diketopiperazine trans-13 (1.10 g, 4.51 mmol) in dry DMF (10.0 mL) was added NaH (60% dispersion in mineral oil, 248 mg, 6.20 mmol) at room temperature. After stirring for 15 min, SEMCl (1.60 mL, 9.03 mmol) was added at room temperature, and the resulting mixture was stirred at 60 °C for 18 h. The reaction was quenched with sat. aq. NH₄Cl at room temperature and the resulting mixture was diluted by EtOAc and washed with water (x5) and brine (x1). The organic phase was dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 1:3) to afford diketopiperazine 16d (1.33 g, 3.55 mmol, 79%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.29–7.26 (3H, m), 7.18–7.15 (2H, m), 5.13 (1H, d, J = 10.4 Hz), 4.53 (1H, d, J =10.4 Hz), 4.42 (1H, dd, J = 5.2, 4.9 Hz), 3.61–3.52 (3H, m), 3.35 (1H, ddd, J = 12.1, 9.3, 2.9 Hz), 3.24 (1H, dd, J = 14.3, 5.2 Hz),3.19 (1H, dd, J = 14.3, 4.9 Hz), 2.52 (1H, dd, J = 10.6, 6.6 Hz), 2.14-2.06 (1H, m), 1.92-1.84 (1H, m), 1.79-1.68 (1H, m), 1.69-1.53 (1H, m), 1.01–0.87 (2H, m), 0.01 (9H, s); ¹³C NMR (100 MHz, CDCl₃): δ 168.3, 164.7, 135.2, 129.9, 128.5, 127.5, 73.3, 66.3, 62.1, 57.6, 44.8, 37.6, 29.1, 21.8, 17.9, -1.5; IR (film): 2952, 2890, 1671, 1454, 1440, 1296, 1260, 1249, 1207, 1075, 1044, 860, 837, 749, 703 cm⁻¹; $[\alpha]_D^{25} = +55.0$ (c = 2.71, CHCl₃); HRMS (ESI) *m/z*: calcd. for C₂₀H₃₀NaN₂O₃Si [M⁺+Na] 397.1918, Found 397.1909.

4.8 Diketopiperazine16e

To a solution of diketopiperazine *cis*-**13** (880 mg, 3.61 mmol) in dry THF (8.0 mL) was added NaH (60% dispersion in mineral oil, 181 mg, 4.53 mmol) at room temperature. After stirring for 50 min, AllomCl (**26**) [20] (750 μ L, 7.23 mmol) was added at room temperature, and the mixture was stirred for 9.5 h at 50 °C. The reaction was quenched with sat. aq. NH4Cl solution at room

extracted with $Et_2O(x1)$ and $CH_2Cl_2(x3)$. The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 2:1, then EtOAc only) to afford diketopiperazine 16e (946 mg, 3.01 mmol, 83%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.32–7.24 (3H, m), 7.19–7.14 (2H, m), 5.89 (1H, m), 5.29 (1H, ddd, J = 17.6, 1.2, 1.2, Hz), 5.19 (1H, d, J = 10.8 Hz), 5.14 (1H, d, J = 10.6 Hz), 4.58 (1H, d, J = 10.6 Hz), 4.43 (1H, dd, J = 4.7, 4.8 Hz), 4.10–3.98 (2H, m), 3.57 (1H, ddd, J = 12.3, 8.5, 8.5 Hz), 3.36 (1H, ddd, J = 12.3, 9.3, 2.9 Hz), 3.25 (1H, dd, J = 14.3, 4.8 Hz), 3.20 (1H, dd, J = 14.3, 4.7 Hz), 2.53 (1H, dd, J = 10.6, 6.6 Hz), 2.09 (1H, ddd, J = 12.0, 6.2, 6.2 Hz), 1.94-1.84 (1H, m), 1.81-1.69 (1H, m), 1.66-1.53 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 168.4, 164.7, 135.3, 133.8, 129.9, 128.6, 127.5, 117.4, 73.6, 70.1, 62.4, 57.7, 44.9, 37.7, 29.0, 21.8; IR (film): 3028, 2979, 2951, 2883, 1668, 1453, 1440, 1344, 1296, 1261, 1207, 1072, 1057, 923, 748, 703 cm⁻¹; $[\alpha]_D^{26} = +60.0$ (*c* = 4.41, CHCl₃); HRMS (ESI) *m/z*: calcd. for $C_{18}H_{22}N_2NaO_3$ [M⁺+Na] 337.1523, Found 337.1506.

4.9 Dithiodiketopiperazine 17a (General Procedure A)

To a suspension of sulfur S₈ (264 mg, 1.03 mmol) in THF (5.0 mL) was added NaHMDS (0.6 M in toluene, 5.00 mL, 3.00 mmol) at room temperature. After stirring for 1 min, diketopiperazine 16a (372 mg, 1.02 mmol) in THF (5.0 mL) was added, and the mixture was stirred for 1 min. Then, NaHMDS (0.6 M in toluene, 3.40 mL, 2.04 mmol) was added, and the mixture was stirred for 30 min at room temperature. The reaction was quenched with sat. aq. NH₄Cl, and the separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was used for the next reaction without further purification. To a solution of the crude material in THF (10.0 mL) and EtOH (10.0 mL) was added NaBH₄ (964 mg, 25.5 mmol) at 0 °C. After stirring for 45 min at 0 °C, MeI (3.18 mL, 51.0 mmol) was added at 0 °C, and the resulting mixture was stirred for 19 h at room temperature. The reaction was quenched by sat. aq. NH₄Cl, and the separated aqueous phase was extracted with CH_2Cl_2 (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes: EtOAc = 12:5) to afford bis(metylthio) diketopiperazine 17a (204 mg, 0.449 mmol, 44%, 2 steps from **16a**). A white solid; ¹H NMR (400 MHz, CDCl₃): δ 7.53 (2H, d, J = 8.8 Hz), 7.31–7.22 (3H, m), 7.09 (2H, d, J = 6.0 Hz), 6.83 (2H, d, J = 8.8 Hz), 5.18 (1H, d, J = 14.4 Hz), 4.54 (1H, d, J = 14.4 Hz), 3.79 (3H, s), 3.66–3.48 (2H, m), 3.54 (1H, d, J = 13.6 Hz), 3.25 (1H, d, J = 13.6 Hz), 2.05-1.91 (2H, m), 2.01 (3H, s), 1.99 (3H, s), 1.58–1.49 (1H, m), 0.81–0.71 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 165.9, 163.0, 158.6, 134.1, 130.2, 129.6, 128.4, 127.7, 113.3, 78.8, 69.9, 55.1, 46.7, 45.8, 44.1, 32.7, 18.5, 15.0, 14.0 (One signal is missing due to overlap.), IR (film): 2994, 2956, 2933, 2922, 1662, 1512, 1421, 1395, 1352, 1247, 1177, 1033, 756, 703 cm⁻¹; $[\alpha]_D^{25} = +0.89$ (*c* 0.58, CHCl₃); HRMS (ESI) *m/z*: calcd. for C₂₄H₂₈N₂NaO₃S₂ [M+Na⁺] 479.1419, Found 479.1434.

4.10 Dithiodiketopiperazine 17b

According to the General Procedure A, diketopiperazine **16b** (301 mg, 0.763 mmol) were converted to bis-(methylthio) diketopiperazine **17b** (210 mg, 0.431 mmol, 57 %, 2 steps from **16b**). A white solid; ¹H NMR (400 MHz, CDCl₃): δ 7.39 (1H, d,

(1H, d, J = 2.1 Hz), 6.41 (1H, d, J = 8.5, 2.1 Hz), 5.07 (1H, d, J = 15.8 Hz), 4.68 (1H, d, J = 15.8 Hz), 3.90 (3H, s), 3.79 (3H, s), 3.67–3.53 (2H, m), 3.55 (1H, d, J = 13.6 Hz), 3.33 (1H, d, J = 13.6 Hz), 2.14 (3H, s), 2.14 (3H, s), 2.10–1.94 (2H, m), 1.61–1.50 (1H, m), 0.86–0.73 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 165.7, 162.9, 159.5, 157.2, 134.2, 130.1, 128.3, 127.5, 127.0, 118.5, 103.5, 98.2, 78.5, 69.7, 55.3, 55.2, 45.8, 44.1, 41.1, 32.5, 18.4, 15.2, 14.2; IR (film): 2998, 2959, 2936, 2921, 2835, 1662, 1616, 1590, 1508, 1455, 1439, 1420, 1396, 1361, 1300, 1261, 1208, 1156, 1120, 1036, 755, 703 cm⁻¹; $[\alpha]_D^{32} = +21.3$ (c = 0.26, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₅H₃₀N₂NaO₄S₂ [M+Na⁺] 509.1539, Found 509.1526.

4.11 Dithiodiketopiperazine 17d

According to the General Procedure A, diketopiperazine 16e (456 mg, 1.22 mmol) were converted to bis-(methylthio) diketopiperazine 17d (386 mg, 0.828 mmol, 68%, 2 steps from **16d**). A white solid; ¹H NMR (400 MHz, CDCl₃): δ 7.25–7.19 (3H, m), 7.12 (2H, dd, *J* = 7.8, 2.2 Hz), 5.48 (1H, d, *J* = 9.6 Hz), 5.06 (1H, d, J = 9.6 Hz), 3.79 (2H, t, J = 8.6 Hz), 3.50 (2H, dd, J = 9.2, 6.4 Hz), 3.46 (1H, d, J = 13.6 Hz), 3.18 (1H, d, J = 13.2 Hz), 2.26 (3H, s), 2.14 (3H, s), 2.12-2.04 (1H, m), 2.02-1.89 (1H, m), 1.55-1.46 (1H, m), 1.07-0.96 (2H, m), 0.93-0.84 (1H, m), 0.03 (9H, s); ¹³C NMR (100 MHz, CDCl₃): δ 166.8, 163.0, 134.3, 130.6, 128.2, 127.4, 77.3, 73.1, 69.5, 67.4, 45.7, 44.8, 32.7, 18.5, 18.4, 15.3, 14.0, -1.4; IR (film): 2952, 2920, 1669, 1454, 1438, 1416, 1385, 1362, 1248, 1083, 1057, 860, 836, 756, 703 cm⁻¹; $[\alpha]_D^{30} = -6.99$ (*c* = 2.76, CHCl₃); HRMS (ESI) *m/z*: calcd. for $C_{22}H_{34}N_2NaO_3S_2Si$ [M+Na⁺] 489.1672, Found 489.1649.

4.12 Dithiodiketopiperazine 17e

According to the General Procedure A, diketopiperazine 16e (843 mg, 2.68 mmol) were converted to bis-(methylthio) diketopiperazine 17e (539 mg, 1.33 mmol, 50% 2 steps from **16e**). A white solid; ¹H NMR (400 MHz, CDCl₃): δ 7.29–7.18 (3H, m), 7.17–7.10 (2H, m), 5.97 (1H, m), 5.51 (1H, d, J = 9.8)Hz), 5.36 (1H, dd, J = 17.2, 1.6 Hz), 5.21 (1H, dd, J = 15.9, 1.8 Hz), 5.11 (1H, d, 9.8 Hz), 4.29 (2H, ddd, J = 5.5, 1.5, 1.5 Hz), 3.54–3.47 (2H, m), 3.47 (1H, d, J = 13.6 Hz), 3.18 (1H, d, J = 13.6 Hz), 2.26 (3H, s), 2.15 (3H, s), 2.14-2.04 (1H, m), 2.03-1.89 (1H, m), 1.58–1.46 (1H, m), 0.98–0.86 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 166.8, 162.9, 134.4, 134.2, 130.6, 128.2, 127.4, 117.0, 77.3, 73.3, 71.2, 69.6, 45.7, 44.9, 32.8, 18.5, 15.2, 13.9; IR (film): 2952, 2920, 2896, 1669, 1438, 1416, 1385, 1362, 1248, 1083, 1057, 860, 836, 756, 703 cm⁻¹; $[\alpha]_D^{30} = -9.65$ (c = 0.660, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₀H₂₆N₂NaO₃S₂ [M+Na⁺] 429.1277, Found 429.1271.

4.13 Dithiodiketopiperazine 15

To a mixture of bis-(methylthio) diketopiperazine **17e** (52.9 mg, 130 µmol) and Pd(PPh₃)₄ (7.53 mg, 6.51 µmol) and 1,3dimethylbarbituric acid (40.7 mg, 260 µmol) was added MeOH (1.3 mL) at room temperature. The resulting suspension was stirred for 14 h at 40 °C. The reaction mixture was cooled to room temperature before addition of excess NaHCO₃ (130 mg) and further stirred for 40 min at room temperature. The reaction mixture was diluted by water (1.0 mL), and the separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried with anhydrous sodium sulfate, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 2:1 to 1:1) to afford bis-(methylthio) diketopiperazine **15** (41.6 mg, 124 µmol, 95%). A white solid; ¹H -17.5 (c = 0.65, CHCl₃); HRMS (ESI) m/z: calcd. for

C₁₆H₁₂N₂NaO₂S₂ [M+Na⁺] 351.0232, Found 351.0207.

4.14 amide 20a (General Procedure B)

To a solution of amide 19a (1.11 g, 5.26 mmol) and TBAI (195 mg, 0.527 mmol) in THF (21.0 mL) was added LiHMDS (1.3 M solution in THF, 4.46 mL, 5.80 mmol) at -40 °C. After stirring for 30 min, AllomCl (26) [20] (0.764 mL, 5.80 mmol) was added at -40 °C, and the mixture was stirred at -40 °C for 14 h. The reaction was quenched with sat. aq. NH₄Cl solution and the mixture was diluted by water. The aqueous phase was extracted with CH₂Cl₂ (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 4:1) to afford amide 20a (1.43 g, 5.08 mmol, 97%). A colorless oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 7.48–7.42 (5H, m), 7.36–7.26 (5H, m), 5.82–5.75 (1H, m), 5.14 (1H, d, J = 17.6 Hz), 5.07 (1H, d, *J* = 10.4 Hz), 4.86–4.67 (4H, m), 3.84 (br, 2H); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 171.0, 137.2, 135.4, 134.0, 129.4, 128.0, 127.9, 127.1, 126.6, 126.5, 116.3, 77.0, 68.0, 48.1; IR (film): 3062, 3030, 2942, 2862, 1651, 1496, 1445, 1417, 1267, 1071, 1050, 699 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₈H₁₉NNaO₂ [M+Na⁺] 304.1308, found 304.1296.

4.15 amide 20b

According to the General Procedure B, amide **19b** (55.5 mg, 191 µmol) was converted to amide **20b** (51.6 mg, 143 µmol, 75%). A yellow oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 7.64–7.16 (2H, m), 7.47 (2H, d, *J* = 8.4 Hz), 7.34–7.26 (5H, m), 5.82–5.77 (1H, m), 5.16 (1H, d, *J* = 17.6 Hz), 5.09 (1H, d, *J* = 10.0 Hz), 3.86 (2H, br s); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 170.1, 137.0, 134.5, 133.9, 130.9, 128.7, 128.0, 127.2, 126.7, 122.9, 116.3, 77.0, 68.0, 48.3; IR (film): 3085, 3063, 3030, 2979, 2942, 2898, 2863, 1652, 1590, 1443, 1417, 1360, 1292, 1266, 1072, 1011, 932, 837, 754, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₈H₁₈BrNNaO₂ [M+Na⁺] 382.0413, Found 382.0403.

4.16 amide 20c

According to the General Procedure B, amide **19c** (62.4 mg, 2.43 µmol) was converted to amide **20c** (70.4 mg, 215mmol, 89%). A yellow oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 8.26 (2H, d, J = 8.0 Hz), 7.73 (2H, d, J = 8.0 Hz), 7.35–7.25 (5H, m), 5.84–5.71 (1H, m), 5.16 (1H, d, J = 17.6 Hz), 5.08 (1H, d, J = 10.8 Hz), 4.70 (2H, br), 4.64 (2H, br), 3.86 (2H, br); ¹³C NMR (100 MHz, CDCl₃): δ 169.4, 147.9, 141.5, 136.8, 133.9, 128.1, 127.9, 127.3, 126.8, 123.2, 116.5, 68.1, 54.3, 48.5; IR (film): 3080, 3066, 3031, 2945, 2862, 1651, 1602, 1521, 1425, 1349, 1072, 732, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₈H₁₈N₂NaO₄ [M+Na⁺] 349.1159, Found 349.1140.

4.17 amide 20d

According to the General Procedure B, amide **19d** (60.3 mg, 250 μ mol) was converted to amide **20d** (68.0 mg, 218 μ mol, 87%). A colorless oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 7.49–7.46 (2H, m), 7.35–7.25 (5H, m), 6.99–6.97 (2H, m),

4.68 (m, 4H), 3.89–3.86 (m, 2H), 3.80 (s, 1H); ¹⁻C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 170.8, 160.4, 137.4, 134.0, 128.6, 127.9, 127.4, 127.2, 126.6, 116.3, 113.4, 77.5, 67.9, 59.5, 48.2; IR (film) 1442, 1417, 1358, 1302, 1253, 1175, 1112, 1049, 933, 842, 766, 700, 598 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₉H₂₁NNaO₃ [M+Na⁺] 334.1414, Found 334.1408.

4.18 amide **20e**

According to the General Procedure B, amide **19e** (74.0 mg, 331 µmol) was converted to amide **20e** (92.1 mg, 314 µmol, 95%). A yellow oil; ¹H NMR (400 MHz, DMSO-d₆, VT 80 °C): δ 7.45–7.42 (5H, m), 5.84–5.78 (1H, m), 5.18 (1H, d, *J* = 17.6 Hz), 5.09 (1H, d, *J* = 10.4 Hz), 4.69 (2H, br), 4.54 (2H, br), 3.86 (2H, br), 3.51 (4H, br), 3.26 (3H, br), 1.88–1.85 (m, 2H); ¹³C NMR (100 MHz, DMSO-d₆, VT 80 °C): δ 170.9, 135.8, 134.1, 129.1, 127.8, 126.4, 116.1, 95.6, 67.4, 64.8, 54.2, 42.8, 27.9 (One signal is missing due to overlap.); IR (film): 2933, 2883, 1651, 1446, 1419, 1404, 1271, 1147, 1112, 1043, 919, 724, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₆H₂₃NNaO₄ [M+Na⁺] 316.1519, Found 316.1516.

4.19 amide 20f

According to the General Procedure B, amide **19f** (40.5 mg, 251 mmol) was converted to amide **20f** (49.5 mg, 214 µmol, 85%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.39–7.21 (5H, m), 5.88–5.81 (1H, m), 5.25 (1H, d, *J* = 16.8 Hz), 5.17–5.12 (2H, m), 4.82 (m, 1H), 4.06 (1H, d, *J* = 10.8 Hz), 3.97–3.94 (m, 2H), 2.64–2.50 (3H, m), 1.99–1.95 (1H, m); ¹³C NMR (100 MHz, CDCl₃): δ 176.2, 140.5, 134.0, 128.8, 128.0, 126.5, 116.9, 70.0, 69.4, 60.4, 30.2, 28.1; IR (film): 3081, 3065, 3031, 2979, 2941, 2885, 1704, 1456, 1415, 1392, 1319, 1256, 1227, 1065, 932, 769, 703 cm⁻¹; HRMS (ESI) *m*/*z*: calcd. for C₁₄H₁₇NNaO₂ [M+Na⁺] 254.1151, Found 254.1147.

4.20 amide 20g

According to the General Procedure B, amide **19g** (58.3 mg, 250 µmol) was converted to amide **20g** (71.7 mg, 236 µmol, 95%). A colorless oil; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.37–7.18 (5H, m), 5.94–5.82 (2H, m), 4.92 (0.7H, s), 4.67 (2H, s), 4.63 (1.3H, s), 4.31 (0.7, d, J = 5.2 Hz), 3.94 (1.3H, d, J = 6.0 Hz), 2.46 (1.3 H, t, J = 7.6 Hz), 2.33 (0.7H, 8.0 Hz), 1.71–1.63 (m, 2H), 1.32–1.35 (m, 8H), 0.88–0.84 (m, 3 H); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 174.8, 174.1, 137.8, 136.9, 134.3, 133.8, 128.8, 128.4, 128.1, 127.4, 127.2, 126.3, 117.6, 116.9, 76.8, 74.3, 69.5, 68.5, 49.2, 48.3, 33.32, 32.99, 31.63, 31.58, 29.3, 29.2, 29.00, 28.95, 25.3, 25.0, 22.53, 22.50, 14.0; IR (film): 3086, 3064, 3030, 2954, 2926, 2855, 1664, 1496, 1453, 1421, 1359, 1236, 1036, 991, 932, 732, 700 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₁₉H₂₉NNaO₂ [M+Na⁺] 326.2091, Found 326.2082.

4.21 amide 19a (General Procedure C)

To a mixture of amide **20a** (56.0 mg, 199 μ mol) and Pd(PPh₃)₄ (23.3 mg, 20.0 μ mol) and 1,3-dimethylbarbituric acid (62.2 mg, 398 μ mol) was added MeOH (0.67 mL) at room temperature. The resulting suspension was stirred for 6 h at 40 °C. The reaction mixture was cooled to room temperature before addition of excess aq. NH₃ (0.3 ml) and further stirred for 40 min at room temperature. Then, the reaction mixture was diluted by water (1.0 mL) and then the separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried with anhydrous sodium sulfate and concentrated under reduced pressure to give a crude material, which was purified by silica gel

amide **19a** (41.4 mg, 196 µmol, 98%).

4.22 amide 19b

According to the General Procedure C, amide **20b** (78.4 mg, 218 µmol) was converted to amide **19b** (54.1 mg, 186 µmol, 86%).

4.23 amide 19c

According to the General Procedure C, amide **20c** (37.8 mg, 116 μ mol) was converted to amide **19c** (13.9 mg, 54.2 μ mol, 47%).

4.24 amide 19d

According to the General Procedure C, amide 20c (65.5 mg, 210 µmol) was converted to amide 19c (40.9 mg, 170 µmol, 81%).

4.25 amide **19e**

According to the General Procedure C, amide **20e** (66.7 mg, 227 μ mol) was converted to amide **19e** (39.0 mg, 175 μ mol, 78%).

4.26 amide 19f

According to the General Procedure C, amide **20f** (39.7 mg, 172 μ mol) were converted to amide **19f** (19.3 mg, 120 μ mol, 70%).

4.27 hemiaminal 21g

To a mixture of amide 20g (20.9 mg, 68.8 µmol) and Pd(PPh₃)₄ (8.0 mg, 6.87 µmol) and 1,3-dimethylbarbituric acid (21.5 mg, 134 µmol) were added THF (0.17 mL) and MeOH (0.06 mL) at room temperature. After stirred for 8 h at 40 °C, the reaction was quenched with sat. aq. Na₂CO₃, and the aqueous phase was extracted with EtOAc (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 5:1) to afford amide hemiaminal 21g (16.6) mg, 63.2 µmol, 92%). A colorless oil; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.36–7.22 (5H, m), 4.86 (2H, br), 4.86-4.65 (2H, br), 3.69 (0.7H, br), 2.64-2.49 (0.9H, m), 2.31 (1.4H, t, J = 8.0 Hz), 1.71-1.61 (m, 2H), 1.25 (br, 8H), 0.88-0.84(m, 3 H); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 175.9, 174.2, 138.0, 136.9, 128.9, 128.7, 128.1, 127.6, 127.5, 1 26.4, 72.2, 71.7, 51.5, 48.2, 33.4, 33.1, 31.7, 31.6, 29.4, 29.2, 29. 1, 29.0, 25.4, 24.9, 22.6, 14.0 (Two signals are missing due to overlap.); IR (film): 3371, 3087, 3063, 3030, 2954, 2926, 2870, 2855, 1636, 1629, 1453, 1423, 1041, 1029, 699 cm⁻¹; HRMS (ESI) m/z: calcd. for C₁₆H₂₆N₂O₂ [M+H⁺] 264.1958, Found 264.1946.

4.28 amide **19**g

To a mixture of hemiaminal **21g** (19.1 mg, 72.7 µmol) and NaI (76.3 mg, 509 µmol) in MeCN (0.36 mL) was added TMSCI (46.0 µL, 495 µmol) at 0 °C. After stirred for 24 h at room temperature, the reaction was quenched with sat. aq. Na₂CO₃, and the aqueous phase was extracted with EtOAc (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 1:1) to afford amide **19g** (17.0 mg, 72.3 µmol, quant).

4.29 amide 22g

Pd(PPh₃)₄ (6.6 mg, 5.67 µmol) and 1,3-dimethylbarbituric acid (17.8 mg, 114 µmol) was added MeOH (0.2 mL) at room temperature. The resulting suspension was stirred for 16.5 h at 40 °C. Then, the reaction was quenched with sat. aq. Na₂CO₃, and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with 1 M aq. HCl and brine, dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes: EtOAc = 3:1) to afford amide **19g** (0.7 mg, 3.00 µmol, 5.3%), 21g (3.2 mg, 12.1 µmol, 21%), and 22g (6.1 mg, 22.0 µmol, 39%). A colorless oil; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.37–7.18 (5H, m), 4.85 (0.8H, s), 4.67 (1.2H, s), 4.61 (2H, s), 3.33 (1.2H, s), 3.28 (1.8H, s), 2.46 (1.2H, t, J = 7.6 Hz), 2.35 (0.8H, t, J = 7.6 Hz), 2.48-1.64 (2H, t)m), 1.32–1.25 (8H, m), 0.88–0.84 (3H, m); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 175.0, 174.2, 137.8, 136.9, 128.8, 128.5, 128.1, 127.5, 127.3, 126.3, 78.9, 76.0, 56.1, 55.3, 49.1, 48.3, 33.4, 33.0, 31.7, 31.6, 29.3, 29.2, 29.1, 29.0, 25.2, 25.1, 22.57, 22.55, 14.0 (One signal is missing due to overlap.); IR (film): 3087, 3063, 3031, 2953, 2927, 2871, 2855, 1663, 1453, 1420, 1389, 1095, 1077, 699 cm⁻¹; HRMS (ESI) *m/z*: calcd. for $C_{17}H_{27}NNaO_2$ [M+Na⁺] 300.1934, Found 300.1921.

4.30 amide 23

To a solution of carboxylic acid 9 [9] (491 mg, 1.10 mmol) and L-phenylalanine methyl ether 10 (497 mg, 2.78 mmol) in dry CH2Cl2 (15.0 mL) was added Et3N (612 µL, 4.40 mmol) at room temperature. After stirring for 10 min, BOP-Cl (576 mg, 2.26 mmol) was added, and the mixture was stirred for 2.7 h at room temperature. The reaction was quenched with sat. aq. NaHCO₃, and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with 1 M aq. HCl and brine, dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 2:1) to afford amide 23 (582 mg, 906 µmol, 87%). A white solid; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers): δ 7.56–7.32 (8H, m), 7.32-7.25 (0.70H, m), 7.15-7.05 (1.30H, m), 6.48-6.34 (1.05H+1.30H, m), 6.06 (1.30H, d, J = 7.6 Hz), 5.36 (0.35H, d, J = 12.4 Hz), 5.29–5.11 (0.35H+1.95H, m), 5.04 (0.35H, ddd, J = 7.8, 5.6, 5.6 Hz), 4.96-4.84 (0.70H+1.95H, m), 4.78 (0.35H, dd, J = 7.8, 7.8 Hz), 4.53–4.46 (1H, m), 3.83 (0.35H, s), 3.77 (1.95H, s), 3.29–3.03 (3H, m), 2.76 (0.35H, d, J = 15.0 Hz), 2.64 (0.65H, d, J = 15.0 Hz), 0.940 (9H, s), 0.131 (1.05H, s), 0.122 (1.95H, s), 0.054 (3H, s); ¹³C NMR (100 MHz, CDCl₃, mixture of rotamers): δ 171.6, 171.5, 171.4, 171.2, 154.7, 153.9, 145.5, 145.2, 136.1, 135.7, 135.4, 135.0, 129.4, 129.1, 128.5, 128.5, 128.4, 128.3, 128.1, 128.1, 127.9, 127.1, 127.0, 120.3, 118.7, 105.9, 105.5, 67.6, 67.3, 67.1, 65.9, 65.0, 64.4, 61.8, 61.4, 53.0, 52.5, 52.2, 52.1, 37.8, 37.7, 33.3, 32.0, 25.6, 17.8, -4.5, -4.6, -5.0, -5.1 (Five signals are missing due to overlap.); IR (film): 3325, 3031, 2953, 2928, 2895, 2855, 1746, 1713, 1639, 1526, 1437, 1408, 1350, 1318, 1252, 1210, 1142, 1113, 1085, 959, 836, 777, 748, 698 cm⁻¹; $[\alpha]_D^{27} = -175$ (c = 1.20, CHCl₃); HRMS (ESI) m/z: calcd. for $C_{33}H_{42}N_2NaO_7Si$ [M+Na⁺] 629.2653, found 629.2624.

4.31 Amide 24

To a suspension of amide **23** (398 mg, 657 μ mol) and Pd(OAc)₂ (57.8 mg, 257 μ mol) in CH₂Cl₂ (12.0 mL) was added Et₃N (300 μ L, 2.16 mmol) at room temperature. After stirring for 30 min at reflux, Et₃SiH (1.10 mL, 6.90 mmol) was added, and the mixture was stirred for 40 min at reflux. The reaction mixture was diluted by CH₂Cl₂ and the reaction was quenched with sat. aq. NaHCO₃. The separated aqueous phase was

dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 6:1 to 2:1) to afford amine 24 (347 mg, quant). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.90 (1H, d, J = 8.4 Hz), 7.22– 7.16 (3H, m), 7.03 (2H, d, J = 6.8 Hz), 6.25 (1H, d, J = 2.0 Hz), 6.22 (1H, d, J = 8.0 Hz), 4.80 (1H, ddd, J = 10.8, 3.2, 3.2 Hz), 4.71 (1H, dd, J = 8.0, 7.8 Hz), 4.27 (1H, dd, J = 7.8, 2.2 Hz), 3.79 (1H, dd, J = 7.2, 5.6 Hz), 3.70 (3H, s), 3.55 (1H, s), 3.19 (1H, dd, J = 14.0, 5.8 Hz), 3.01 (1H, dd, J = 14.0, 6.6 Hz), 2.73-2.71 (2H, m), 2.28 (1H, s), 0.82 (9H, s), 0.033 (3H, s), 0.022 (3H, s); ¹³C NMR (100 MHz, CDCl₃): δ 173.0, 171.9, 145.5, 135.8, 133.1, 129.1, 128.3, 126.9, 120.5, 105.6, 67.9, 63.9, 60.9, 52.5, 52.1, 37.6, 35.0, 25.6, 17.9, -3.8, -4.7; IR (film): 3349, 3029, 2952, 2928, 2893, 2855, 2360, 2341, 1745, 1677, 1644, 1508, 1437, 1360, 1344, 1254, 1200, 1079, 961, 837, 779, 702 cm⁻¹; $[\alpha]_D^{26} = -195$ (c = 1.65, CHCl₃); HRMS (ESI) m/z: calcd. for $C_{25}H_{37}N_2O_5Si^+$ [M+H⁺] 473.2466, found 473.2451.

4.32 Dithiodiketopiperazine 25

To a solution of amine 24 (447 mg, 945 µmol) in MeOH (250 mL) was added 25% aq. NH₃ (15.0 mL) at room temperature. After stirring for 2.7 days at room temperature, the reaction was concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes: EtOAc = 2:1) to afford diketopiperazine 25 (202 mg, 459 μ mol, 48%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.33-7.18 (5H, m), 6.41 (1H, s), 6.29 (1H, d, J = 8.0 Hz), 5.90 (1H, s), 5.00 (1H, d, J = 7.6 Hz), 4.88 (1H, dd, J = 8.0, 7.6 Hz), 4.77 (1H, s), 4.23–4.16 (2H, m), 3.51 (1H, dd, *J* = 14.0, 3.6 Hz), 2.89–2.79 (2H, m), 2.52 (1H, dd, J = 13.0, 13.0 Hz), 0.79 (9H, s), 0.011 (3H, s), 0.006 (3H, s); ¹³C NMR (100 MHz, CDCl₃): δ 168.5, 163.9, 145.0, 135.6, 135.1, 129.3, 128.9, 127.3, 112.4, 106.4, 66.9, 63.1, 57.9, 56.2, 37.3, 33.4, 25.7, 17.8, -4.3, -4.5; IR (film): 3381, 3228, 3220, 2952, 2928, 2895, 2855, 2359, 2341, 1684, 1663, 1444, 1422, 1294, 1256, 1202, 1139, 1083, 957, 905, 837, 777, 746, 702 cm⁻¹; $[\alpha]_D^{25} = -356$ (*c* = 0.270, CHCl₃); HRMS (EI) *m/z*: calcd. for C₂₄H₃₂N₂O₄Si [M⁺] 440.2131, found 440.2134.

4.33 Dithiodiketopiperazine 27

To a solution of diketopiperazine 25 (63.4 mg, 144 µmol) in THF (630 µL) was added LiHMDS (1.3 M solution in THF, 120 µL, 156 µmol) at -40 °C. After stirring for 8 min, AllomCl (26) [20] (120 μ L, 1.16 mmol) was added at -40 °C, and the mixture was stirred at -20 °C for 17 h. The reaction was quenched with sat. aq. NH₄Cl solution and the mixture was diluted by water. The separated aqueous phase was extracted with CH_2Cl_2 (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 4:1 to 2:1) to afford diketopiperazine 27 (66.3 mg, 120 µmol, 84%). A colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.28–7.22 (3H, m), 7.04 (2H, d, J = 8.0 Hz), 6.28 (1H, d, J = 7.6 Hz), 6.27 (1H, s), 5.96–5.87 (1H, m), 5.51 (1H, d, J = 10.8 Hz), 5.31 (1H, dt, J = 17.2, 1.6 Hz), 5.24 (1H, dt, J = 10.8, 1.2 Hz), 4.87(1H, dd, J = 7.8, 7.6 Hz), 4.82 (1H, d, J = 7.8 Hz), 4.79 (1H, d, J = 10.8 Hz), 4.62 (1H, s), 4.57 (1H, m), 4.12 (1H, dd, J = 12.8, 6.0 Hz), 4.07–4.40 (2H, m), 3.42 (1H, dd, J = 14.0, 2.8 Hz), 3.21 (1H, dd, J = 14.0, 2.8 Hz), 2.44 (1H, dd, J = 13.2, 6.0 Hz), 0.94 (1H, dd, J = 13.2, 12.8 Hz), 0.79 (9H, s), 0.007 (3H, s), -0.009 (3H, s); ¹³C NMR (100 MHz, CDCl₃): δ 166.2, 163.7, 144.9, 134.9, 134.7, 133.4, 130.1, 128.6, 127.4, 118.1, 112.8, 106.4, 72.5, 69.9, 68.6, 62.9, 59.1, 58.2,

1662, 1452, 1441, 1295, 1200, 1138, 1092, 1051, 955, 906, 837, 777, 703 cm⁻¹; $[\alpha]_D^{25} = -333$ (*c* = 0.535, CHCl₃); HRMS (ESI) *m/z*: calcd. for C₂₈H₃₈N₂NaO₅Si⁺[M+Na⁺] 533.2442, found 533.2417.

4.34 Dithiodiketopiperazine 28

To a solid of diketopiperazine 27 (66.3 mg, 120 µmol) was added HF·Py in pyridine (2.00 mL, 1:4, HF·Py containing ~70% hydrogen fluoride and ~30% pyridine) at room temperature. After stirring for 17 h at 45 °C, the reaction mixture was diluted by water at 0 °C. The separated aqueous phase was extracted with CH2Cl2 (x4). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (CH_2Cl_2 :EtOAc = 1:5) to afford allyl alcohol 28 (46.6 mg, 118 µmol, 98%). A white solid; ¹H NMR (400 MHz, CDCl₃): δ 7.28–7.22 (3H, m), 7.07–7.04 (2H, m), 6.34 (1H, d, J = 8.0 Hz), 6.34 (1H, d, J = 2.0 Hz), 5.98-5.91 (1H, m), 5.41 (1H, d, J = 10.8 Hz), 5.35 (1H, dt, J = 17.5, 2.9, 1.7 Hz), 5.24 (1H, dt, J = 10.3, 2.9, 1.5 Hz), 5.05 (1H, dd, J = 8.2, 8.0 Hz), 4.82 (1H, d, J = 10.8 Hz), 4.79–4.76 (2H, m), 4.69 (1H, s), 4.58–4.56 (1H, m), 4.22 (1H, dd, J = 12.2, 6.0 Hz), 4.16– 4.10 (2H, m), 3.42 (1H, dd, J = 14.2, 3.0 Hz), 3.23 (1H, dd, J = 14.2, 4.4 Hz), 2.48 (1H, dd, J = 14.4, 6.0 Hz), 0.94 (1H, dd, J = 14.4, 12.2 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 166.0, 163.8, 145.1, 135.2, 134.7, 133.6, 130.1, 128.4, 127.3, 117.8, 113.7, $105.7, 73.2, 70.3, 68.1, 62.6, 59.8, 58.3, 36.8, 33.3; [\alpha]_D^{25} = -367$ (c = 0.260, CHCl₃); IR (film): 3425, 3419, 2921, 2359, 2340, 1694, 1659, 1454, 1339, 1296, 1200, 1137, 1074, 1047, 929, 857, 758, 736, 704 cm⁻¹; HRMS (ESI) *m/z*: calcd. for C₂₂H₂₄N₂NaO₅ [M+Na⁺] 419.1577, found 419.1558.

4.35 Dithiodiketopiperazine 29

To a solution of allyl alcohol 28 (74.3 mg, 188 µmol) and nor-AZADO (8.50 mg, 61.5 µmol) in dry CH₂Cl₂ (1.40 mL) was added PhI(OAc)₂ (64.2 mg, 200 µmol) at room temperature. After stirring for 1.5 h at room temperature, the reaction was quenched with sat. aq. Na₂SO₃ solution and water (1:2). The separated aqueous phase was extracted with CH_2Cl_2 (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes: EtOAc = 1:3) to afford lactone 29 (71.0 mg, 180 µmol, 96%). A white solid; ¹H NMR (400 MHz, CDCl₃): δ 7.28–7.22 (3H, m), 7.09 (2H, d, J = 8.0 Hz), 7.00 (1H, d, J = 7.6 Hz), 6.59 (1H, d, J = 2.4 Hz), 6.01–5.91 (1H, m), 5.61 (1H, d, J = 7.6 Hz), 5.36 (1H, d, J = 18.4 Hz), 5.35 (1H, d, J = 10.4 Hz), 5.31 (1H, s), 5.27 (1H, d, J = 10.4 Hz), 4.90 (1H, d, J = 10.4 Hz), 4.68 (1H, dd, J = 4.0, 3.4 Hz), 4.20–4.10 (2H, m), 3.91 (1H, dd, J = 13.4, 6.0 Hz), 3.43 (1H, dd, J = 14.0, 3.4 Hz), 3.29 (1H, dd, J = 14.0, 4.0 Hz), 2.53 (1H, dd, J = 15.2, 6.0 Hz), 1.01 (1H, dd, J = 13.4, 13.4 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 180.9, 164.9, 163.6, 153.3, 136.9, 134.5, 133.4, 130.0, 128.3, 127.1, 117.6, 115.4, 107.8, 73.5, 70.4, 66.3, 60.0, 57.0, 36.9, 31.8; IR (film): 3028, 2925, 2867, 1687, 1667, 1604, 1454, 1350, 1327, 1292, 1204, 1078, 1051, 940, 930, 864, 820, 758, 736 cm⁻¹; $[\alpha]_D^{26} = -279$ (c = 0.445, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₂H₂₃N₂O₅ [M+H⁺] 395.1601, found 395.1605.

4.36 Dithiodiketopiperazine 30

To a solution of lactone **29** (138 mg, 350 μ mol) in CH₂Cl₂ (750 μ L) and EtOH (500 μ L) was added CeCl₃· 7H₂O (381 mg) in EtOH (2.0 mL) at -78 °C. After stirring for 10 min, NaBH₄ in EtOH (22.0 mg, 582 μ mol in 550 μ L) was added at -78 °C, and

quenched with aq. NH₄Cl solution at room temperature. The separated aqueous phase was extracted with CH_2Cl_2 (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes:EtOAc = 1:3) to afford allyl alcohol 30 (126 mg, 318 μ mol, 91%). A white solid; ¹H NMR (400 MHz, CDCl₃): δ 7.31–7.23 (3H, m), 7.08 (2H, d, *J* = 6.4 Hz), 6.27 (1H, dd, J = 2.2, 2.2 Hz), 6.13 (1H, dd, J = 8.0, 2.4 Hz), 6.00–5.90 (1H, m), 5.42 (1H, d, J = 10.8 Hz), 5.36 (1H, d, J = 17.2 Hz), 5.26 (1H, d, J = 10.0 Hz), 5.07 (1H, d, J = 3.6 Hz), 4.88 (1H, s), 4.87 (1H, d, J = 10.8 Hz), 4.64 (1H, dd, J = 3.0, 4.2 Hz), 4.52 (1H, d, *J* = 7.6 Hz), 4.20–4.17 (1H, m), 4.14 (1H, d, *J* = 5.8 Hz), 4.13 (1H, d, J = 5.8 Hz), 3.91 (1H, dd, J = 13.0, 5.8 Hz), 3.40 (1H, dd, J = 13.8, 3.0 Hz), 3.27 (1H, dd, J = 13.8, 4.2 Hz), 2.36 (1H, dd, J = 13.6, 5.8 Hz), 0.80 (1H, dd, J = 13.6, 13.0 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 167.0, 165.1, 138.0, 137.0, 134.2, 133.4, 130.2, 128.7, 127.7, 118.1, 109.8, 109.7, 73.2, 70.5, 69.1, 64.1, 59.8, 56.3, 37.0, 33.7; IR (film): 3369, 3011, 2942, 2863, 1661, 1453, 1344, 1296, 1193, 1133, 1078, 1058, 930, 759, 756, 730, 704, 578 cm⁻¹; $[\alpha]_D^{25} = -229$ (c = 1.04, CHCl₃); HRMS (ESI) m/z: calcd. for C₂₂H₂₄N₂NaO₅ [M+Na⁺] 419.1577, found 419.1564.

4.37 Dithiodiketopiperazine 31

To a suspension of sulfur S₈ (63.6 mg, 247 µmol) in Et₂O (1.0 mL) was added LiHMDS (1.3 M in THF, 725 µL, 943 µmol) at room temperature. After stirring for 5 min, diketopiperazine 30 (18.6 mg, 47.0 μ mol) in a mixture of THF (2.0 mL) and Et₂O (1.0 mL) was added at room temperature, and the resulting solution was stirred for 1 min. Then, LiHMDS (1.3 M in THF, 725 µL, 943 µmol) was added, and the mixture was stirred for 30 min at room temperature. The reaction was quenched with aq. NH₄Cl and diluted with water. The separated aqueous phase was extracted with Et₂O (x1) and CH₂Cl₂ (x2). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 16:1x2, then hexanes: $CH_2Cl_2 = 3:5$) to afford the mixture of bridged polysulfides. To a solution of the polysulfides in a mixture of THF (3.2 mL) and EtOH (3.2 mL) was added NaBH₄ (36.5 mg, 965 µmol) at 0 °C. After stirred for 8 min at room temperature, MeI (1.60 mL, 25.6 mmol) was added at 0 °C, and the resulting mixture was stirred at room temperature for 4 h. The reaction was quenched by aq. NH₄Cl, and the separated aqueous phase was extracted with Et₂O (x1) and CH₂Cl₂ (x2). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes: EtOAc = 16:1, then hexanes: EtOAc = 2:1) to afford bis-(methylthio) diketopiperazine **31** (18.3 mg, 30.3 µmol, 65% 2 steps from **30**). A white foam; ¹H NMR (400 MHz, CDCl₃): δ 7.29–7.24 (3H, m), 7.15–7.12 (2H, m), 6.31 (1H, m), 6.17 (1H, dd, J = 8.4, 2.4Hz), 6.03–5.93 (1H, m), 5.40–5.34 (1H, m), 5.53 (1H, d, J = 9.6 Hz), 5.24–5.22 (2H, m), 5.12 (1H, d, J = 9.6 Hz), 4.91 (1H, dd, J = 8.2, 1.8 Hz), 4.58–4.55 (1H, m), 4.42 (1H, d, J = 8.0 Hz), 4.30– 4.28 (2H, m), 3.48 (1H, d, J = 13.2 Hz), 3.23 (1H, d, J = 13.2Hz), 2.55 (1H, d, J = 15.2 Hz), 2.29 (3H, s), 2.53 (3H, s), 1.59 (1H, d, J = 15.2 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 167.1, 166.1, 137.6, 137.2, 134.3, 133.6, 130.7, 128.6, 127.9, 117.2, 110.8, 107.1, 77.5, 73.4, 72.2, 71.3, 67.7, 65.4, 45.0, 38.1, 15.2, 14.0; IR (film): 3370, 3008, 2920, 2852, 1662, 1435, 1416, 1380, 1191, 1136, 1083, 1056, 1050, 772, 756, 704 cm⁻¹; $[\alpha]_D^{25} = -123$ $(c = 0.619, \text{CHCl}_3)$; HRMS (ESI) m/z: calcd. for $C_{24}H_{28}N_2NaO_5S_2$ [M+Na⁺] 511.1332, found 511.1303.

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and

To a solid of bis-(methylthio) diketopiperazine 31 (6.65 mg, 13.6 $\mu mol)$ was added $Pd(PPh_3)_4$ (0.788 mg, 0.682 $\mu mol)$ in CH₂Cl₂ (25.0 µL). The mixture was concentrated under reduced pressure give a pale brown solid. To the solid was added 1,3dimethylbarbituric acid (4.26 mg, 27.3 µmol) in MeOH (136 µL), and the mixture was stirred for 16.5 h at 40 °C. The reaction mixture was diluted with MeOH (68 µL) and stirred for 2 h. Then, the mixture was cooled to room temperature and diluted with MeOH (200 μ L). To the mixture was added NaHCO₃ (40.0 mg), and the resulting suspension was stirred for 25 min at room temperature and diluted with CH₂Cl₂ and H₂O. The separated aqueous phase was extracted with CH₂Cl₂ (x3). The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude purified material. which was by preparative TLC (hexanes:EtOAc = $3:1 \times 3$) to afford emestrin H (5) (4.46 mg, 10.6 µmol, 78%) and diketopiperazine 30 (1.15 mg, 17%). A white solid; ¹H NMR (600 MHz, CDCl₃): δ 7.34–7.30 (3H, m), 7.19 (2H, d, J = 6.6 Hz), 6.36 (1H, s), 6.19 (1H, dd, J = 1.8, 8.4 Hz), 6.05 (1H, s), 5.12 (1H, d, J = 4.8 Hz), 4.93 (1H, d, J = 8.4 Hz), 4.63–4.61 (1H, m), 4.55 (1H, d, *J* = 7.2 Hz), 3.64 (1H, d, *J* = 13.2 Hz), 3.02 (1H, d, J = 13.2 Hz), 2.63 (1H, d, J = 15.0 Hz), 2.41 (3H, s), 2.23 (3H, s), 1.88 (1H, d, J = 15.0 Hz); ¹³C NMR (150 MHz, CDCl₃): δ 167.2, 165.4, 137.7, 137.4, 133.2, 130.7, 128.8, 128.1, 110.9, 107.4, 72.3, 68.5, 68.4, 64.9, 46.3, 38.9, 14.7, 14.3; IR (film): 3361, 3224, 3062, 3008, 2918, 2848, 2360, 2341, 1675, 1638, 1409, 1340, 1193, 1139, 1123, 1047, 756, 730, 704, 581, 564 cm⁻¹; $[\alpha]_{D}^{28} = -203$ (*c* = 0.234, MeOH) (Lit. $[\alpha]_{D}^{25}$ = -97 (*c* = 0.071, MeOH)); HRMS (ESI) *m/z*: calcd. for $C_{20}H_{22}N_2NaO_4S_2$ [M+Na⁺] 441.0913, found 441.0907. The spectral data were identical with those reported in the literature [14].

4.39 (-)-Asteroxepin (4)

To a solution of (-)-emestrin H (5) (2.64 mg, 6.32 µmol) and DMAP (1.23 mg, 10.1 µmol) in pyridine (120 µL) was added Ac₂O in pyridine (0.127 M, 100 µL, 12.7 µmol) at room temperature. After stirring for 50 min at room temperature, additional Ac_2O (13.0 $\mu L,$ 138 $\mu mol)$ was added, and the mixture was stirred for 35 min. Then, additional DMAP (5.69 mg, 46.6 µmol) was added, and the mixture was stirred for 45 min at room temperature. The reaction was diluted with CH₂Cl₂, and quenched with aq. 1 M HCl. The separated aqueous phase was extracted with CH₂Cl₂ (x5), and the combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes:EtOAc = 7:11) to afford asteroxepin (4) (2.60 mg, 5.65 µmol, 89%). A white solid; ¹H NMR (600 MHz, CDCl₃): δ = 7.32–7.27 (3H, m), 7.20– 7.18 (2H, m), 6.41 (1H, dd, J = 2.4, 2.4 Hz), 6.29 (1H, dd, J = 8.4, 3.0 Hz), 6.00 (1H, br s), 5.88 (1H, ddd, J = 7.5, 2.1, 2.1), 4.89 (1H, br. d, J = 7.8 Hz), 4.68 (1H, dd, J = 8.4, 1.2 Hz), 3.66 (1H, d, *J* = 13.8 Hz), 2.97 (1H, d, *J* = 13.8 Hz), 2.64 (1H, d, *J* = 15.0 Hz), 2.36 (3H, s), 2.20 (3H, s), 2.13 (3H, s), 1.82 (1H, ddd, J = 15.3, 2.1, 2.1 Hz); ¹³C NMR (150 MHz, CDCl₃): $\delta = 170.1$, 165.6, 164.4, 139.7, 137.3, 133.5, 130.9, 128.7, 127.9, 109.1, 105.5, 71.4, 69.9, 68.0, 61.7, 46.4, 39.6, 21.2, 14.5, 14.0; $[\alpha]_D^{27} = -178$ $(c = 0.556, CHCl_3)$ (Lit. $[\alpha]_D = -102$ ($c = 0.4, CHCl_3$)); HRMS (ESI) *m/z*: calcd. for C₂₂H₂₄N₂NaO₅S₂ [M+Na⁺] 483.1019, found 483.0996. The spectral data were identical with those reported in the literature [13].

4.40 AllomCl (26) [20]

paraformaldehyde (19.0 g, 0.633 mol) was added anhydrous hydrogen chloride generated from H_2SO_4 and NaCl at room temperature. After stirred for 30 min at room temperature, the mixture was heated by oil bath at 150 °C for 2 h. The resulting mixture was diluted with hexanes, dried over magnesium sulfate, and the hexanes were removed under reduced pressure. The crude material was distilled from anhydrous calcium chloride to afford AllomCl (**26**) (25.0 g, 0.234 mol, 40%) as colorless oil.

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17. Compounds **16c–e** were synthesized from *trans*-**13** with C3 (*S*) and C8a (*R*) stereochemistries, which was prepared by the stepwise condensation of *N*-Boc-L-proline and L-phenylalanine methyl ester. During the intramolecular condensation step, the C8a (*S*) stereochemistry was completely isomerized to give *trans*-**13** with C8a (*R*) stereochemistry.



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Declaration of interests

 \square The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: