

Efficient Synthesis of the Immunomodulating Compound KRP-203

Maiko Hamada,* Masatoshi Kiuchi, Kunitomo Adachi

Chemistry Laboratory Pharmaceuticals Research Division, Mitsubishi Pharma Corporation, 1000, Kamoshida-cho, Aoba-ku, Yokohama, Kanagawa 227-0033, Japan

Fax +81(45)9633529; E-mail: Hamada.Maiko@mc.m-pharma.co.jp

Received 6 March 2007; revised 27 April 2007

Abstract: A practical and concise synthesis of KRP-203 (**1**) was achieved by utilizing a palladium coupling reaction mediated by XantPhos and Pd₂(dba)₃ as the key step. The coupling reaction of aryl bromide **5** with 3-hydroxythiophenol (**6**) proceeded chemoselectively to give phenol **7** in spite of the presence of a chlorine substituent on **5** and a hydroxyl group on **6**. Phenol **7** was converted into KRP-203 by benzylation and removal of the protecting groups in high yield.

Key words: palladium, cross-coupling, synthesis, KRP-203, FTY720

Diaryl sulfides are common functionalities found in biologically active compounds.¹ The traditional methods for the construction of diaryl sulfides like Ullmann condensation often need harsh conditions.² Recently, mild and highly selective methods for the formation of diaryl sulfides using metal-catalyzed (Cu, Pd) coupling reactions were reported.³ These methods could shorten the synthetic schemes using traditional substitution reaction for compounds possessing a diaryl sulfide moiety.

KRP-203 (**1**, Figure 1) is a novel immunomodulating compound with a diaryl sulfide moiety.⁴ The hydrophilic part of **1**, 2-aminopropane-1,3-diol, is the same as that of FTY720 (**2**).⁵ Compound **2** is a synthetic immunomodulating agent with a sphingosine-like structure, and was efficacious against multiple sclerosis and kidney transplant rejection in clinical studies.⁶ For the immunomodulating mechanism of action is presumed that **2** is converted into the monophosphate ester of **2**, FTY720-P (**3**), in vivo, and then **3** interacts with sphingosine 1-phosphate (S1P) receptor type-1 (S1P₁), among five known S1P receptors (S1P₁₋₅), as an agonist.⁷ In the clinical trials of **2**, transient bradycardia was reported as a frequent adverse event.⁶ It

is considered that the effect of **2** on heart rate is also related to the agonism at S1P receptors, especially S1P₃ in rodents.⁸ Interestingly, while **1** showed potent immunomodulating effect comparable to **2**, the effect on heart rate of **1** was weak in guinea pigs.⁴ Furthermore, the potential of the monophosphate ester of **1**, KRP-203-P (**4**), causing bradycardia in rats was ten-fold weaker than that of **3**.⁹ These data show that **1** is a unique and useful compound to investigate the physiological role of S1P receptors. We present here a new concise synthetic approach to **1** using a Pd-catalyzed cross-coupling reaction.

In the previous preparations of **1**, the construction of the diaryl sulfide moiety was accomplished by a traditional substitution reaction using 2-chloro-4-fluorobenzaldehyde as the starting material.¹⁰ For the efficient proceeding of the substitution reaction, the aryl fluoride activated by the carbonyl group was required. However, the one-carbon elongation steps after the construction of the diaryl sulfide moiety were the main reason for the long synthetic schemes.

Our strategy for synthesis of **1** was to prepare the diaryl sulfide moiety by a Pd-catalyzed cross-coupling reaction of aryl bromide **5**, which was easily accessible by an established synthetic procedure¹¹ from diethyl 2-[N-(*tert*-butoxycarbonyl)amino]malonate and 4-bromo-2-chlorobenzoic acid, with commercially available thiol **6**. The synthesis of **1** is shown in Scheme 1.

The first step was the formation of the diaryl sulfide. We adopted the condition using Pd₂(dba)₃ in combination with 4,5-bis(diphenylphosphino)-9,9-dimethylxanthene (XantPhos) as the phosphine ligand reported by Itoh.¹² Although nonactivated aryl chlorides showed poor reactivity under the condition, aryl-sulfur bonds were efficiently

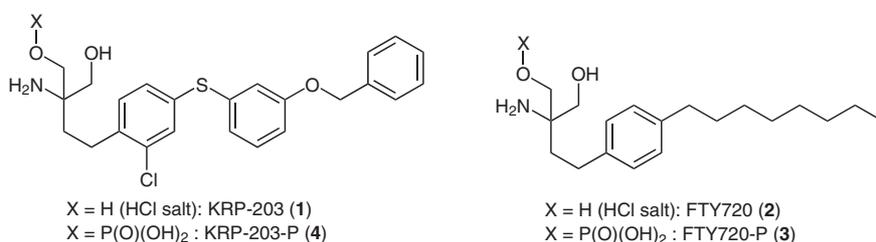


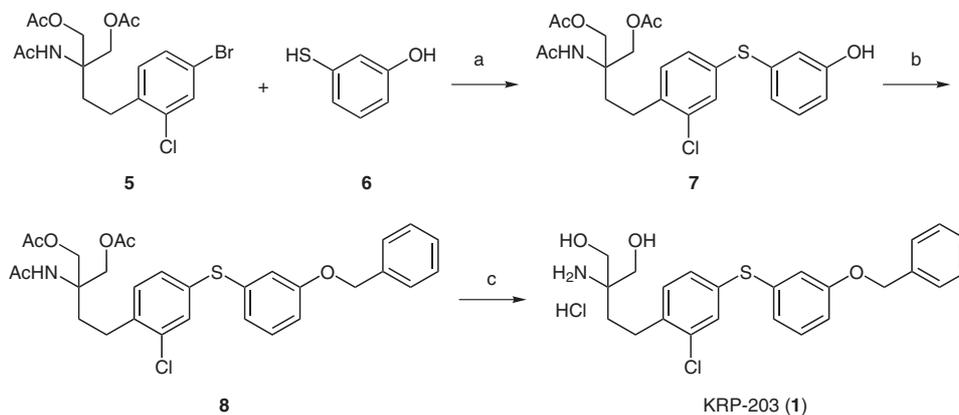
Figure 1

SYNTHESIS 2007, No. 13, pp 1927–1929

Advanced online publication: 18.06.2007

DOI: 10.1055/s-2007-983721; Art ID: F04807SS

© Georg Thieme Verlag Stuttgart · New York



Scheme 1 Reagents and conditions: (a) $\text{Pd}_2(\text{dba})_3$ (2.5 mol%), XantPhos (5 mol%), DIPEA (2.0 equiv), 1,4-dioxane, 120 °C; (b) BnBr (1.3 equiv), K_2CO_3 (3 equiv), DMF, r.t.; (c) 1. $\text{LiOH}\cdot\text{H}_2\text{O}$ (5 equiv), $\text{MeOH}\text{--}\text{H}_2\text{O}\text{--}\text{THF}$, 80 °C, 2. 2 M HCl/EtOH, EtOH.

formed from aryl bromides and thiols. This selectivity to bromide over chloride was favorable to our synthetic strategy using bromo- and chloro-substituted aryl compound **5** as the starting compound.

Under this condition we were able to obtain the desired product **7** in 86% yield. The carbon–sulfur bond formation proceeded chemoselectively in spite of the presence of a phenolic hydroxyl group.¹² In addition, the reaction of **5** with the thiol group of **6** occurred at the bromine atom exclusively. Triacetyl KRP-203 (**8**) was obtained by benzylation of the phenol group of **7** by a standard method using benzyl bromide and potassium carbonate. Removal of the acetyl groups was accomplished by treatment with LiOH in an aqueous medium to give free base of **1**. Finally, the free base of **1** was converted into its HCl salt **1** by treatment with 2 M HCl in EtOH. The three steps yield from **7** was 91%, and the spectroscopic data well supported the structure and high purity of compound **1** obtained by this scheme.

In conclusion, we have developed a practical and concise synthetic method for KRP-203 (**1**). The key step was the chemoselective coupling reaction of thiol **6** with aryl bromide **5**. This route shortened the previously reported synthetic schemes of **1**, and makes it easier to utilize **1** for pharmacological and biochemical studies.

Silica gel column chromatography was performed using Fuji Silysia PSQ100B. ^1H NMR spectra were recorded on a Bruker Avance 400 (400 MHz) spectrometer. Chemical shifts are expressed in ppm relative to TMS as an internal standard. IR spectra were recorded on a PerkinElmer FT-IR Spectrum One spectrometer. Mass spectra were measured in a combination with a Waters Acquity UPLC system (0.05% TFA in MeCN/0.05% TFA in H_2O) and a Micromass ZQ (ESI) spectrometer. Melting points were obtained on a Büchi 535 melting point apparatus and are uncorrected. Elemental analyses were performed on a Yonako CHN coder MT-5 analyzer.

***N*-{1,1-Bis(acetoxymethyl)-3-[2-chloro-4-(3-hydroxyphenylthio)phenyl]propyl}acetamide (7)**

A mixture of **5** (10.0 g, 23.0 mmol), 3-hydroxythiophenol (2.8 mL, 27.6 mmol), tris(dibenzylideneacetone)dipalladium(0)-chloroform adduct (600 mg, 0.58 mmol), 4,5-bis(diphenylphosphino)-9,9-di-

methylxanthene (665 mg, 1.2 mmol) and *N,N*-diisopropylethylamine (8.0 mL, 46.0 mmol) in 1,4-dioxane (46 mL) was stirred at 120 °C for 9 h. After pouring into aq 1 M HCl (100 mL), the mixture was extracted with EtOAc (3 \times 200 mL). The combined organic layers were washed with brine (2 \times 100 mL), dried (Na_2SO_4), and concentrated in vacuo. The residue was purified by silica gel column chromatography (hexane–EtOAc) to give **7** (9.48 g, 86%) as a white solid; mp 142–143 °C.

IR (ATR): 3468, 3271, 3072, 1740, 1714, 1641 cm^{-1} .

^1H NMR (400 MHz, $\text{DMSO}-d_6$): δ = 1.84 (s, 3 H), 1.92–1.97 (m, 2 H), 2.02 (s, 6 H), 2.62–2.66 (m, 2 H), 4.20 (d, J = 11.0 Hz, 2 H), 4.29 (d, J = 11.0 Hz, 2 H), 6.71–6.78 (m, 3 H), 7.18–7.33 (m, 4 H), 7.69 (s, 1 H), 9.70 (br s, 1 H).

MS (ESI): m/z = 480 [M + H].

Anal. Calcd for $\text{C}_{23}\text{H}_{26}\text{ClNO}_6\text{S}$: C, 57.55; H, 5.46; N, 2.92. Found: C, 57.46; H, 5.44; N, 2.86.

***N*-{3-[4-(3-Benzyloxyphenylthio)-2-chlorophenyl]-1,1-bis(acetoxymethyl)propyl}acetamide (8)**

To a solution of **7** (9.48 g, 19.8 mmol) in DMF (65 mL) were added K_2CO_3 (8.20 g, 59.3 mmol) and benzyl bromide (3.0 mL, 25.2 mmol). The mixture was stirred at r.t. overnight. After pouring into H_2O (150 mL), the mixture was extracted with EtOAc (3 \times 200 mL). The combined organic layers were washed with H_2O (3 \times 150 mL) and brine (2 \times 100 mL), dried (Na_2SO_4), and concentrated in vacuo. The residue was purified by silica gel column chromatography (hexane–EtOAc) to give **8** (10.9 g, 97%) as a white solid; mp 81–82 °C.

IR (ATR): 3269, 3097, 1744, 1732, 1650 cm^{-1} .

^1H NMR (400 MHz, $\text{DMSO}-d_6$): δ = 1.84 (s, 3 H), 1.93–1.97 (m, 2 H), 2.02 (s, 6 H), 2.62–2.66 (m, 2 H), 4.20 (d, J = 11.0 Hz, 2 H), 4.29 (d, J = 11.0 Hz, 2 H), 5.09 (s, 2 H), 6.89–7.00 (m, 3 H), 7.24 (dd, J = 8.0 Hz, 1.6 Hz, 1 H), 7.30–7.42 (m, 8 H), 7.69 (s, 1 H).

MS (ESI): m/z = 570 [M + H].

Anal. Calcd for $\text{C}_{30}\text{H}_{32}\text{ClNO}_6\text{S}$: C, 63.20; H, 5.66; N, 2.46. Found: C, 63.16; H, 5.61; N, 2.42.

2-Amino-2-[2-[4-(3-benzyloxyphenylthio)-2-chlorophenyl]ethyl]propan-1,3-diol Hydrochloride (KRP-203, 1)

To a solution of **8** (10.9 g, 19.1 mmol) in MeOH (90 mL), THF (30 mL), and H_2O (90 mL) was added $\text{LiOH}\cdot\text{H}_2\text{O}$ (4.14 g, 100 mmol). After stirring at 80 °C for 2 h, the organic solvents were removed in vacuo. The solid obtained was collected by filtration to give the free base of **1**. The solid was dissolved in 2 M HCl in EtOH (30 mL) and

EtOH (50 mL). After stirring, the precipitate formed was collected by filtration to give **1** (8.6 g, 94%); white solid; mp 199–200 °C (dec.).

IR (ATR): 3238, 3034, 2943 cm⁻¹.

¹H NMR (400 MHz, DMSO-*d*₆): δ = 1.76–1.80 (m, 2 H), 2.71–2.76 (m, 2 H), 3.55 (d, *J* = 4.8 Hz, 4 H), 5.10 (s, 2 H), 5.42 (t, *J* = 4.8 Hz, 2 H), 6.90–7.01 (m, 3 H), 7.25 (dd, *J* = 7.9, 1.2 Hz, 1 H), 7.30–7.42 (m, 8 H), 7.91 (br s, 3 H).

MS (ESI): *m/z* = 444 [M + H].

Anal. Calcd for C₂₄H₂₇Cl₂NO₃S: C, 60.00; H, 5.66; N, 2.92. Found: C, 59.83; H, 5.59; N, 2.90.

References

- (1) (a) Liu, G.; Link, J. T.; Pei, Z.; Reilly, E. B.; Leitza, S.; Nguyen, B.; Marsh, K. C.; Okasinski, G. F.; von Geldern, T. W.; Ormes, M.; Fowler, K.; Gallatin, M. *J. Med. Chem.* **2000**, *43*, 4025. (b) Wang, Y.; Chackalamannil, S.; Chang, W.; Greenlee, W.; Ruperto, V.; Duffy, R. A.; McQuade, R.; Lachowicz, J. E. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 891. (c) Bonnet, B.; Soullez, D.; Girault, S.; Maes, L.; Landry, V.; Davioud-Charvet, E.; Sergheraert, C. *Bioorg. Med. Chem.* **2000**, *8*, 95.
- (2) Pinchart, A.; Dallaire, C.; Gingras, M. *Tetrahedron Lett.* **1998**, *39*, 543.
- (3) (a) Herradura, P. S.; Pendola, K. A.; Guy, R. K. *Org. Lett.* **2000**, *2*, 2019. (b) Savarin, C.; Srogl, J.; Liebeskind, L. S. *Org. Lett.* **2002**, *4*, 4309. (c) Bates, C. G.; Gujadhur, R. K.; Venkataraman, D. *Org. Lett.* **2002**, *4*, 2803. (d) Paloma, C.; Oiarbide, M.; López, R.; Gómez-Bengoa, E. *Tetrahedron Lett.* **2000**, *41*, 1283. (e) Kwong, F. Y.; Buchwald, S. L. *Org. Lett.* **2002**, *4*, 3517. (f) Schopfer, U.; Schlapbach, A. *Tetrahedron* **2001**, *57*, 3069.
- (4) Shimizu, H.; Takahashi, M.; Kaneko, T.; Murakami, T.; Hakamata, Y.; Kudou, S.; Kishi, T.; Fukuchi, K.; Iwanami, S.; Kuriyama, K.; Yasue, T.; Enosawa, S.; Matsumoto, K.; Takeyoshi, I.; Morishita, Y.; Kobayashi, E. *Circulation* **2005**, *111*, 222.
- (5) Kiuchi, M.; Adachi, K.; Kohara, T.; Minoguchi, M.; Hanano, T.; Aoki, Y.; Mishina, T.; Arita, M.; Nakao, N.; Ohtsuki, M.; Hoshino, Y.; Teshima, K.; Chiba, K.; Sasaki, S.; Fujita, T. *J. Med. Chem.* **2000**, *43*, 2946.
- (6) (a) Kahan, B. D.; Karlix, J. L.; Ferguson, R. M.; Leichtman, A. B.; Mulgaonkar, S.; Gonwa, T. A.; Skerjanec, A.; Schmuuder, R. L.; Chodoff, L. *Transplantation* **2003**, *76*, 1079. (b) Tedesco-Silva, H.; Mourad, G.; Kahan, B. D.; Boira, J. G.; Weimar, W.; Mulgaonkar, S.; Nashan, B.; Madsen, S.; Charpentier, B.; Pellet, P.; Vanrenterghem, Y. *Transplantation* **2004**, *77*, 1826. (c) Kappos, L.; Antel, J.; Comi, G.; Montalban, X.; O'Connor, P.; Polman, C. H.; Haas, T.; Korn, A. A.; Karlsson, G.; Radue, E. W. *N. Engl. J. Med.* **2006**, *355*, 1124.
- (7) (a) Mandala, S.; Hadju, R.; Bergstrom, J.; Quackenbush, E.; Xie, J.; Milligan, J.; Thornton, R.; Shei, G.; Card, D.; Keohane, C.; Rosenbach, M.; Hale, J.; Lynch, C. L.; Rupprecht, K.; Parsons, W.; Rosen, H. *Science* **2002**, *296*, 346. (b) Brinkmann, V.; Davis, M. D.; Heise, C. E.; Albert, R.; Cottens, S.; Hof, R.; Bruns, C.; Prieschl, E.; Baumruker, T.; Hiestand, P.; Foster, C. A.; Zollinger, M.; Lynch, K. R. *J. Biol. Chem.* **2002**, *277*, 21453.
- (8) Sanna, M. G.; Liao, J.; Jo, E.; Alfonso, C.; Ahn, M.; Peterson, M. S.; Webb, B.; Lefebvre, S.; Chun, J.; Gray, N.; Rosen, H. *J. Biol. Chem.* **2004**, *279*, 13839.
- (9) Fujishiro, J.; Kudou, S.; Iwai, S.; Takahashi, M.; Hakamata, Y.; Kinoshita, M.; Iwanami, S.; Izawa, S.; Yasue, T.; Hashizume, K.; Murakami, T.; Kobayashi, E. *Transplantation* **2006**, *82*, 804.
- (10) (a) Konishi, Y.; Ando, N.; Kuriyama, K.; Iwanami, S.; Kudou, S. PCT Int. Appl. WO 03/029205, **2003**; *Chem. Abstr.* **2003**, *138*, 304049. (b) Tsubuki, T.; Kobayashi, K.; Komatsu, H. PCT Int. Appl. WO 2006/041019, **2006**; *Chem. Abstr.* **2006**, *144*, 412416.
- (11) (a) Fan, Y.; Gao, W.; Gray, N.; Terding, K.; Lefebvre, S.; Mi, Y.; Nussbaumer, P.; Pan, S.; Wang, W.; Zecri, F.; Perez, L. B.; La Montagne, K. R.; Eitmayer, P. PCT Int. Appl. WO 03/099192, **2003**; *Chem. Abstr.* **2003**, *140*, 4842. (b) Kiuchi, M.; Kobayashi, N.; Sugahara, K.; Nakamura, M. PCT Int. Appl. WO 2006/129688, **2006**; *Chem. Abstr.* **2006**, *146*, 45290.
- (12) Itoh, T.; Mase, T. *Org. Lett.* **2004**, *6*, 4587.