



One pot synthesis and anticancer activity of dimeric phloroglucinols [☆]

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ABSTRACT

A series of dimeric phloroglucinol compounds were synthesized in a single step using commercially available phloroglucinol and methanesulfonic acid. Based on the reported anticancer activity of plant derived dimeric phloroglucinols, these synthesized compounds were evaluated for their in vitro anti-proliferative activities against various cancer cell lines. Several compounds demonstrated in vitro cytotoxic effects across a wide array of tumor cell types. The compound **29** with pyridin-3-yl group on linker methylene and two diisovaleryl phloroglucinol moieties was found to be the most active in all the five cancer cell lines having a low IC₅₀ of 5.5 μM in colon cancer cell lines (HCT116).

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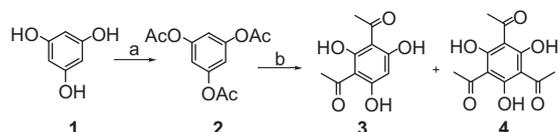
Cancer is a fatal disease and a leading cause of death worldwide with projected 12 million deaths in 2030.¹ Several classes of anti-cancer drugs have been developed and many of them are of natural origin. Natural products have been the mainstay of cancer chemotherapy for the past 30 years.² However, most of the currently used anticancer drugs cause undesirable side effects due to lack of tumor specificity and multidrug resistance. Therefore the search for potent, safe and selective anticancer compounds is crucial for new drug development in cancer research. Natural products, due to their structural diversity, provide excellent templates for the construction of novel compounds.²

Naturally occurring dimeric phloroglucinol compounds have shown a diverse range of biological activities including anticancer activity.³ Dimeric phloroglucinols comprise compounds having two phloroglucinol units joined either through a methylene linkage or by the formation of a chroman ring. Several dimeric phloroglucinol derivatives such as mallotojaponin, mallotochromene, mallotorin and mallotophenone have been isolated from *Mallotus japonicus* and evaluated for anticancer activity in various cancer cell lines.^{4a–e} Mallotojaponin was found to be active in all the cancer cell lines, it inhibited tumor promoter-enhanced phospholipid metabolism in cultured cells, and also suppressed the promoting effect of 12-*O*-tetradecanoylphorbol-13-acetate on skin tumor formation in mice.^{5,6} Several monomeric acylphloroglucinol derivatives from

Garcinia cowa and *Hypericum prolificum* have also exhibited potent anticancer activity.^{7,8}

As a part of our continuing efforts to synthesize naturally occurring phloroglucinol compounds and analogs to explore their biological potential,^{9–12} herein, we report a novel synthesis of diacyl phloroglucinols and one pot synthesis of dimeric phloroglucinol compounds containing various functionalities by varying substitution at the methylene bridge. All the synthesized compounds were tested for their in vitro anti-proliferative activities against various cancer cell lines.

Earlier reported methods for synthesis of monomeric acylated phloroglucinol units require toxic catalysts such as AlCl₃,⁹ ZnCl₂,¹³ BF₃-etherate,¹⁴ and toxic solvent nitrobenzene.⁹ Some reports used BF₃-etherate as a solvent for the reactions.¹⁴ Purification of final product required distillation of high boiling nitrobenzene. In this letter we report a simple and greener method for the synthesis of diacyl phloroglucinols and dimeric phloroglucinols using environment friendly methanesulfonic acid (MSA). Initially, solid catalysts such as basic alumina¹⁵ and perchloric acid-silica¹⁶ were



Scheme 1. Reagents and conditions: (a) Acetic anhydride, pyridine; (b) CH₃SO₂OH, 80 °C, 30 min.

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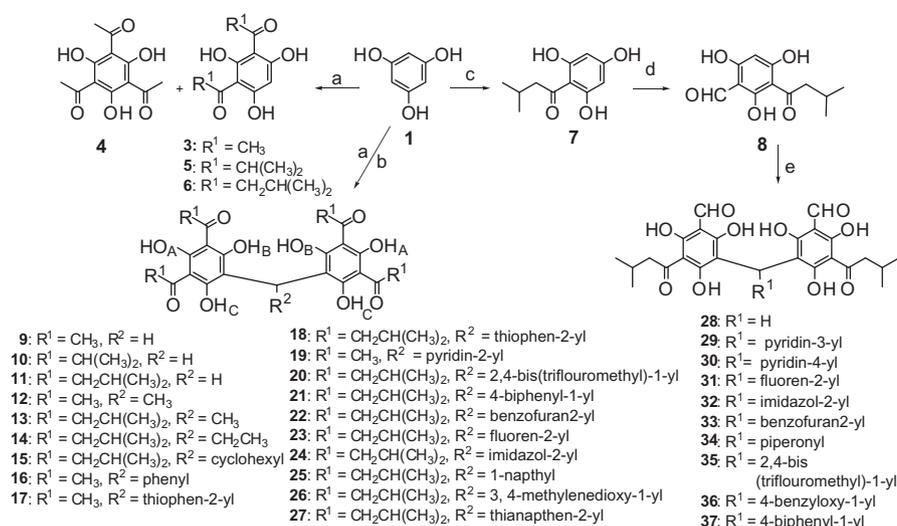
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Table 1
Reaction conditions for synthesis of 2,4-diacetylphloroglucinol (DAPG)

Catalyst	Reaction conditions	Yield ^a (%)
4 mol equiv MSA	3 mol equiv Ac ₂ O, THF, 80 °C, 0.5 h	0
4 mol equiv MSA	3 mol equiv Ac ₂ O, Methanol, 80 °C, 0.5 h	0
3 mol. equiv MSA and 1 mol equiv acetic acid	3 mol equiv Ac ₂ O, 80 °C, 0.5 h	94
3 mol equiv MSA	3 mol equiv Ac ₂ O, 80 °C, 0.5 h	90
3 mol equiv MSA	2 mol equiv Ac ₂ O, 80 °C, 0.5 h	64
3 mol equiv MSA	3 mol equiv CH ₃ COCl, 80 °C, 0.5 h	92
3 mol equiv HCl	3 mol equiv Ac ₂ O, 80 °C, 0.5 h	0
3 mol equiv CH ₃ COOH	4 mol equiv Ac ₂ O, 80 °C, 0.5 h	0
3 mol equiv HClO ₄	4 mol equiv Ac ₂ O, 80 °C, 0.5 h	0
3 mol equiv TFA	4 mol equiv Ac ₂ O, 80 °C, 0.5 h	50
3 mol equiv H ₂ SO ₄	4 mol equiv Ac ₂ O, 80 °C, 0.5 h	0
3 mol equiv HBr/Acetic acid	4 mol equiv Ac ₂ O, 80 °C, 0.5 h	94
MSA/Alumina	4 mol equiv Ac ₂ O, 80 °C, 0.5 h	0
MSA/Graphite	3 mol equiv CH ₃ COOH, 80 °C, 0.5 h	0
10 mol% of MSA and 1 mol equiv acetic acid	3 mol equiv Ac ₂ O, MW 700 W, 5 min	0
20 mol% of MSA and 1 mol equiv acetic acid	3 mol equiv Ac ₂ O, MW 700 W, 1 min	65
30 mol% of MSA and 1 mol equiv acetic acid	3 mol equiv Ac ₂ O, MW 700 W, 1 min	88

^a Isolated yields after crystallization.



Scheme 2. Reagents and conditions: (a) RCOCl, CH₃SO₂OH, 80 °C, 30 min/RCOCl, CH₃SO₂OH, MW, 1 min, 700 W, (b) R-CHO, 15 min, (c) AlCl₃, isovaleryl chloride, PhNO₂, 1.5 h, (d) DMF, POCl₃, EtOAc, 1 h, (e) CH₃SO₂OH, R-CHO, 80 °C, 15 min.

employed for direct Friedel–Crafts C-acylation of phloroglucinol (**1**) using acetic anhydride as an acylating agent. Several variations with the amount of catalyst, temperature and solvent mostly gave 1,3,5-tri-*O*-acetyl phloroglucinol (**2**) and phloroacetophenone⁹ as major and minor products, respectively. We have found that methanesulphonic acid (MSA), an environment friendly bronsted acid can also be used to synthesize the target compound through Fries rearrangement of 1,3,5-tri-*O*-acetyl phloroglucinol. First, 1,3,5-tri-*O*-acetyl phloroglucinol was reacted with 8 mol equiv of MSA at 80 °C for 30 min to yield a mixture of 2,4-diacetyl phloroglucinol (DAPG) (**3**) and 2,4,6-triacetyl phloroglucinol (**4**) in 8:2 ratio (Scheme 1).

Further, we have optimized the reaction in a single step for direct acylation of phloroglucinol (**1**) by varying the amount of MSA and acetic anhydride. Use of MSA less than 3 mol equiv resulted in an incomplete reaction. Use of solvent such as methanol or THF did not yield the desired products. The direct acylation of phloroglucinol (**1**) with 3 mol equiv of MSA, and 3 mol equiv Ac₂O at 80 °C for 0.5 h in neat condition gave 2,4-diacetylphloroglucinol (**3**) and 2,4,6-triacetyl phloroglucinol (**4**) in 8:2 ratio with 90% yield (Table 1). In order to minimize the formation of (**4**), we reduced the amount of acetic anhydride to 2 mol equiv, but it resulted

incomplete reaction and low yield of the product. Finally addition of 1 mol equiv of acetic acid along with 3 mol equiv MSA and 3 mol equiv Ac₂O at 80 °C for 0.5 h in neat condition gave DAPG (**3**) and 2,4,6-triacetyl phloroglucinol (**4**) in 95:5 ratio with 94% yield (Table 1). We also tried different bronsted acids like CH₃COOH, HCl, H₂SO₄, HBr in acetic acid, and HCOOH with acetic anhydride at 80 °C to observe the outcome of the reaction, except HBr in acetic acid none of the acid catalysts gave the desired products. MSA adsorbed on alumina or graphite with Ac₂O at 80 °C also failed to yield the desired product. The reaction conditions for synthesis of DAPG were optimized further to limit the use of MSA from stoichiometric to catalytic amount using microwave irradiation. Initially reaction of phloroglucinol (**1**) with 10 mol % of MSA, 1 mol equiv acetic acid and 3 mol equiv of acetic anhydride at 700 W power for 5 min showed incomplete reaction and a mixture of products. Increasing the concentration of MSA from 10 to 30 mol% gave the desired DAPG (**3**) and **4** in 9:1 ratio with 88% yield (Table 1). This improved synthetic methodology using microwave heating as well as conventional heating was applied for synthesis of various diacyl derivatives using different acyl chlorides and anhydrides (Scheme 2). Reaction of diisobutryl and diisovaleryl chloride with phloroglucinol (**1**) under similar reaction conditions gave

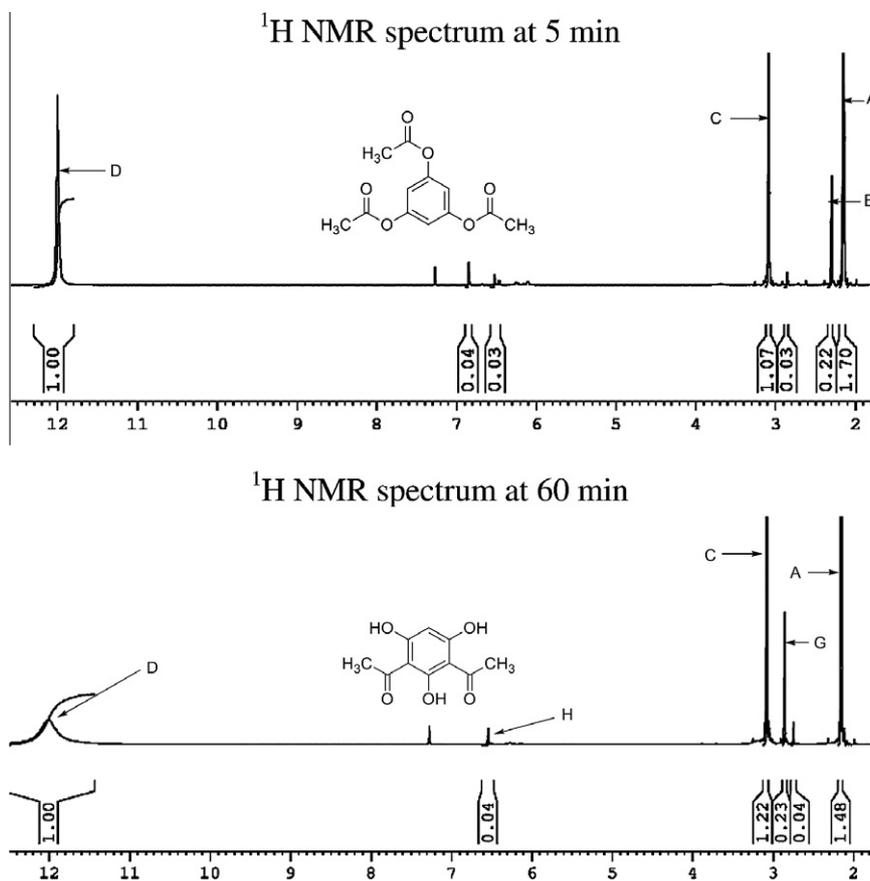


Figure 1. A = CH₃ of acetic acid, B = CH₃ of tri-*O*-acetyl phloroglucinol, C = CH₃ of MSA, D = OH of MSA, G = CH₃ of 2,4-diacetyl phloroglucinol (DAPG), H = Aromatic proton of DAPG.

Table 2

Cytotoxicity of dimeric phloroglucinol derivatives against various human cancer cell lines^a

Compd	ACHN	Panc1	Calu1	H460	HCT116	Compd	ACHN	Panc1	Calu1	H460	HCT116
3	44	0	0	25	0	21	59	54	52	54	55
5	33	0	15	22	0	22	7	0	56	37	20
6	30	0	0	46	55	23	17	18	0	15	0
8	52	50	59	58	57	24	0	0	11	15	0
9	46	22	39	0	8	25	6	17	15	26	0
10	30	12	16	25	6	26	67	69	62	63	60
11	35	0	7	3	1	27	54	59	54	54	65
12	19	0	38	0	3	28	35	15	29	0	9
13	11	0	12	12	25	29	71	67	66	68	69
14	0	0	10	10	22	30	21	5	17	0	0
15	14	0	18	29	0	31	50	57	58	54	52
16	29	0	47	29	0	32	38	0	0	22	18
17	57	64	0	54	55	33	6	0	2	5	11
18	50	60	62	54	57	34	53	55	54	51	50
19	12	0	24	0	11	35	0	0	6	5	14
20	57	60	65	64	61	36	44	6	30	49	48
Gemcitabine (500 nM)	77	83	74	81	86	37	0	0	0	28	5

^a The values given in the table is % death of cancer cells at 10 μM concentration. ACHN, renal cancer; Panc1, pancreatic cancer; Calu1, lung cancer, H460, non-small cell lung cancer; HCT116, colon cancer.

2,4-diisobutyryl phloroglucinol (**5**)¹⁴ and 2,4-diisovaleryl phloroglucinol (**6**)¹⁴ in 93% and 95% yield. No triisovaleryl or triisobutyryl phloroglucinol was formed in conventional as well as microwave heating. Steric hindrance due to bulky isovaleryl and isobutyryl groups may have provided the selectivity. In case of conventional heating, reaction of the bulky acyl chlorides required more time compared to short chain acyl chlorides. Bronsted acids were studied for Friedel–Crafts C versus O-acylation of phenols. Direct Friedel–Crafts C-acylation has been reported with strong bronsted acids.¹⁷ Phloroglucinol nucleus is electron rich, therefore MSA

catalyzed acylation of phloroglucinol (**1**) may proceed either by direct Friedel–Crafts C-acylation or through Fries rearrangement of the intermediate *O*-acetyl derivatives (**2**) or both the mechanisms operating simultaneously (Fig. S1, Supplementary data). Reaction of 3 mol equiv of MSA and 3 mol equiv Ac₂O at room temperature for 1 h gave 1,3,5-tri-*O*-acetyl phloroglucinol (**2**) and phloroacetophenone as major and minor products, respectively. This interesting observation showed that the minor product phloroacetophenone could be formed from direct Friedel–Crafts C-acylation. Hence, 1D NMR spectroscopy was employed to elucidate the

reaction mechanism for formation of diacetyl phloroglucinol derivatives. Acetylation of phloroglucinol (**1**) was done as discussed above and NMR spectra were recorded at different time intervals starting from 5 min of the reaction till the completion of the reaction. The ¹H NMR spectrum at 5 min showed CH₃ protons at δ 2.3 indicating the presence of intermediate 1,3,5-tri-*O*-acetyl phloroglucinol (**2**) (Fig. 1). As the reaction progressed, the intermediate slowly converted to DAPG (**3**), NMR spectra at 60 min showed CH₃ protons at δ 2.85 indicating the complete transformation into the DAPG (**3**) (Fig. 1). The ¹³C NMR spectra at 5 and 60 min showed the characteristic ester and ketone carbonyl signals at 169.03 and 203.40, respectively.

These experiments concluded that the 2,4-diacetylphloroglucinol was formed through Fries migration (Figs. S2–S4, see Supplementary data).

3-Formyl phloroisovalerophenone moiety required for synthesis of dimeric phloroglucinol compounds (**28–37**) was synthesized in two steps.⁹ The Friedel–Crafts acylation of phloroglucinol (**1**) using isovaleryl chloride resulted in formation of phloroisovalerophenone (**7**) in 70% yield. Treatment of **7** with Vilsmeier–Haack reagent (POCl₃, DMF) in ethyl acetate resulted in formation of 3-formyl phloroisovalerophenone (**8**) in 70% yield.

The synthetic methodology described for DAPG using conventional heating was extended for the synthesis of naturally occurring dimeric phloroglucinol derivatives. We envisaged that the acidic hydrogen of the diacyl phloroglucinol derivatives could be trapped with aldehyde molecules to form the bioactive dimeric phloroglucinol in a single step. Synthesis of **9**¹⁷ was started with reaction of phloroglucinol (**1**), with 3 mol equiv MSA, 1 mol equiv acetic acid and 3 mol equiv Ac₂O at 80 °C for 0.5 h. After consumption of phloroglucinol in the reaction mixture, linker aldehyde was added to the reaction mixture and allowed to stir for another 15 min. TLC analysis in 50% ethyl acetate in hexane showed formation of the desired dimeric phloroglucinol. Similarly, other naturally occurring dimeric phloroglucinols with isovaleryl, isobutyryl and 3-formyl phloroisovalerophenone moieties viz. methylene-*bis*-(3,5-di-isobutanoyl-2,4,6-trihydroxybenzene) (**10**), methylene-*bis*-(3,5-di-isopentanoyl-2,4,6-trihydroxybenzene) (**11**), methylene-*bis*-(3-formyl-5-isopentanoyl-2,4,6-trihydroxybenzene) (**28**) and other dimeric phloroglucinols (**12–27** and **29–36**) were synthesized by varying the substitution on aromatic ring (changing acyl functionalities) and at methylene bridge (by using different aldehydes viz. saturated, aromatic and heteroaromatic) in a key condensation step as shown in Scheme 2. This methodology has wide applicability for one pot synthesis of naturally occurring dimeric phloroglucinol derivatives joined through the methylene linkage. The structures of synthesized compounds are shown in Scheme 2. The diacyl dimeric phloroglucinol compounds could be easily purified by crystallization. The dimers with formyl functionality were purified by RP-C₁₈ silica gel column chromatography. All the compounds were characterized by IR, MS, ¹H NMR and ¹³C NMR spectral data.

The single dose in vitro cytotoxicity screening assays of the analogs were carried out at 10 μM concentration. The human tumor cell line panel included non-small cell lung, colon, pancreas, and renal cancer cell lines. From the preliminary results (Table 2) dimeric phloroglucinols containing formyl substituents on the aromatic moiety exhibited good growth inhibition properties. The ten most active compounds from the preliminary screening were subsequently evaluated in four dose–response studies for their in vitro cytotoxic effects on growth parameters against each of human tumor cell lines. The dose response curves were created by plotting cytotoxic effect against the log₁₀ of the drug concentration for each cell line. IC₅₀ values are presented in Table 3. Structure–activity relationship study was established by varying the aromatic and heteroaromatic substituents on linker methylene carbon and the acyl groups on aromatic phloroglucinol moieties containing two

Table 3

IC₅₀ values of selected diacyl and dimeric phloroglucinol derivatives in μM concentration

Compd	Cancer cell lines (IC ₅₀ value) ^a					
	ACHN	Panc1	Calu1	H460	HCT116	MCF10A ^b
8	8.9	8.5	9.1	9.4	9.7	>27.3
17	9.2	9.4	9.7	8.9	9.9	>30
18	>10	8.5	8.9	9.1	8.9	>30
20	8.8	8.3	8.5	8.6	9.2	27.6
21	8.9	9.1	7.9	8.3	8.6	>26.3
26	6.7	7.2	6.5	6.6	7.1	>30
27	9.9	9.8	9.5	9.7	9.8	>30
29	5.7	6.1	6.3	5.8	5.5	>30
31	9.9	9.6	8.8	9.2	9.4	26.2
34	8.9	8.8	9.2	8.6	9.5	25.9
Gemcitabine	0.48	0.11	0.52	0.23	0.32	>10

^a The IC₅₀ values, defined as the drug concentration at which 50% of cells are viable, were calculated from the respective logarithmic cytotoxicity curves of the different cancer cells.

^b MCF10A: normal breast epithelium cells.

formyl groups. Monomeric diacyl phloroglucinol derivatives (**3, 5** and **6**) showed weak activity in the initial screening, while formyl phloroisovalerophenone (**8**) was active in all the cancer cell lines. Dimeric phloroglucinol nucleus with diisovaleryl, diisobutyryl, diacetyl and formyl-isovaleryl functionality (**9–11**) and **28** on the aromatic nuclei and devoid of substitution at methylene bridge were found weakly active in all the cancer cell lines. Next, different alkyl, aromatic, heteroaromatic were introduced on the linker methylene bridge. Results indicate that ten compounds (**8, 17, 18, 20, 21, 26, 27, 29, 31** and **34**) were active in all the five cancer cell lines. Moreover, the compounds were not toxic to normal cell lines (MCF10A, normal breast epithelium cell line). Among the ten active compounds **26** and **29** showed highest activity in all the cancer cell lines. Compound **29** with pyridin-3-yl group on linker methylene and two diisovaleryl phloroglucinol moieties was found to be the most active in all the five cancer cell lines having a lowest IC₅₀ of 5.5 μM in colon cancer cell lines (HCT116). Compounds (**12–14**) possessing small alkyl substituents on linker methylene carbon were inactive or less active as compared to compounds with bulky aromatic substituents. Replacing acyl functionality with formyl group resulted in increased activity as observed in **8, 29** and **34**. This SAR study suggested the basic pharmacophore responsible for activity would be a dimeric phloroglucinol nucleus with long chain acyl (isovaleryl) and formyl substituents and linker methylene carbon substituted with bulky aromatic groups.

In conclusion, a novel one pot synthesis has been developed for diacyl and dimeric phloroglucinol derivatives starting from commercially available phloroglucinol and biodegradable methanesulfonic acid catalyst. Twenty five novel dimeric phloroglucinols have been synthesized and screened for in vitro anticancer activity in various cell lines. The long chain acyl (isovaleryl) and formyl substituents and linker methylene carbon substituted with bulky aromatic group were found essential for anticancer activity. This new synthetic method could be applicable to bulk scale synthesis of variety of new diacyl and dimeric phloroglucinol derivatives. The active compound (**29**) could be used as a lead molecule for further development of new anticancer therapeutic molecules.

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2012.01.089.

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- General method for synthesis of diacyl and dimeric phloroglucinols using conventional heating:* The mixture of phloroglucinol (**1**, 1 mmol), acid anhydride/acyl chloride (3 mmol), acetic acid (1 mmol) and methanesulfonic acid (3 mmol) was heated for 30 min at 80 °C. On cooling, water was added and the reaction mixture and was extracted with ethyl acetate. Crude diacyl phloroglucinol derivative were crystallised from methanol. In case of synthesis of dimeric phloroglucinol, after 30 min, the linker aldehyde was added to the reaction mixture and the reaction was continued for further 15 min. After completion of reaction (TLC), water was added to the reaction mixture and extracted with ethyl acetate. Organic layer was dried over sodium sulphate and the solvent was removed under vacuum and the crude product was purified by crystallisation from methanol. *General method for synthesis of diacyl phloroglucinol derivative using microwave (IFB, model: 30SC2 with functions to control heating) heating:* The mixture of phloroglucinol (**1**, 1 mmol), acid anhydride/acyl chloride (3 mmol), acetic acid (1 mmol) and methanesulfonic acid (30 mol %) was irradiated in microwave oven for 1 min. The reaction workup is similar to the conventional heating method. The yields are reported less than 50% as two equivalents of monomeric starting material produce 1 equiv of dimeric product. *Cell proliferation assay (CCK-8 assay):* All the cancer cells, Panc1, ACHN, H460, Calu1, HCT116, and MCF10A were obtained from the American Type Culture Collection (ATCC, Manassas, VA). Cells were maintained in Dulbecco's Modified Eagle Medium (DMEM, Sigma St. Louis, MO), supplemented with 10% fetal bovine serum Gibco (Paisley, Scotland), 100 U/ml penicillin (Sigma, St. Louis, MO) and 100 µg/ml streptomycin (Sigma St. Louis, MO). The cells were grown in 75-cm² culture flasks kept in a humidified (37 °C, 5% CO₂) incubator, and passaged on reaching 80% confluence. The assay is based on the reduction of water-soluble tetrazolium salt (WST-8; [2-(2-methoxy-4-nitrophenyl)-3-(4-nitrophenyl)-5-(2,4-disulfophenyl)]-2H-tetrazolium mono sodium salt) by dehydrogenase enzyme of live cells to give a yellow coloured product (formazan). 5 × 10³ cells were seeded per well in a 96 well tissue culture grade plate and the plate was kept overnight at 37 °C in 5% CO₂ incubator. After overnight incubation, cells were treated with different concentrations of compounds/standard. An equal concentration of vehicle (DMSO-never exceeding 0.1%) was used as a control. At 48 h after treatment, aliquots of 5 µl of CCK-8 reagent were added to each well and incubated for 4 h at 37 °C. After appropriate incubation, the absorbance was measured on Tecan Sapphire multi-fluorescence micro-plate reader (Tecan, Germany, GmbH) at a wavelength of 450 nm corrected to 650 nm and normalized to controls. DMSO treated control cells were considered to have a cell viability of 100%. The average number of viable cells at different compound/standard concentrations was expressed as a percentage of the control.
- Methyl-methylene-bis-(3,5-diacetyl-2,4,6-trihydroxybenzene) (12):* Yield: 32%; yellow solid; mp 206–208 °C; UV (CHCl₃): λ_{max} (log ε) 276 (4.37), 341 (3.92); IR (KBr): ν_{max} 3168, 1618, 1579, 1476, 1419, 1364, 1263, 1185, 1115, 1024 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ 17.65 (s, 2H, 2 × OH_C), 16.25 (s, 2H, 2 × OH_B), 10.71 (br s, 1H, OH_A), 10.22 (br s, 1H, OH_A), 4.80 (q, J = 7.2 Hz, 1H), 2.76 (s, 6H), 2.73 (s, 6H), 1.79 (d, J = 7.2 Hz, 3H); ¹³C NMR (CDCl₃, 75 MHz): δ 206.0, 205.7, 171.2, 168.2, 166.2, 108.9, 105.7, 104.6, 34.0, 32.9, 24.2, 17.6; CIMS: m/z 447 [M+1]⁺, 237 [M-C₁₀H₁₀O₅]; analysis for C₂₂H₂₂O₁₀ (446.1), calcd, C, 59.19; H, 4.97; found, C, 59.10; H, 5.04. *Methyl-methylene-bis-(3,5-diisopentanoyl-2,4,6-trihydroxybenzene) (13):* Yield: 37%; yellow sticky mass; UV (CHCl₃): λ_{max} (log ε) 285 (4.49), 340 (4.15); IR (Neat): ν_{max} 3403, 2949, 1615, 1581, 1459, 1399, 1459, 1399, 1195, 1044 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ 17.94 (br s, 2H, 2 × OH_C), 16.46 (s, 2H, 2 × OH_B), 10.70 (br s, 1H, OH_A), 10.22 (br s, 1H, OH_A), 4.80 (q, J = 7.2 Hz, 1H), 3.05 (d, J = 6.5 Hz, 8H), 2.27 (m, 4H), 1.79 (d, J = 7.2 Hz, 3H), 1.00 (d, J = 6.4 Hz, 24H); ¹³C NMR (CDCl₃, 75 MHz): δ 208.5, 208.0, 171.2, 167.2, 163.1, 109.2, 105.7, 104.7, 53.9, 52.9, 25.7, 23.3, 17.7; CIMS: m/z 321[M-C₁₀H₁₀O₅]; analysis for C₃₄H₄₆O₁₀ (614.3), calcd, C, 66.43; H, 7.54; found, C, 66.52; H, 7.49. *Ethyl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxybenzene) (14):* Yield: 41%; yellow oil; UV (CHCl₃): λ_{max} (log ε) 285 (4.48), 337 (4.13); IR (Neat): ν_{max} 3199, 2960, 2873, 1616, 1584, 1466, 1367, 1367, 1301, 1200, 1047 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ 18.00 (s, 1H, OH_C), 17.76 (s, 1H, OH_C), 16.45 (s, 2H, 2 × OH_B), 10.87 (s, 1H, OH_A), 10.06 (s, 1H, OH_A), 4.44 (t, J = 7.7 Hz, 1H), 3.06 (d, J = 6.5 Hz, 8H), 2.27 (m, 6H), 1.00 (d, J = 6.4 Hz, 24H), 0.87 (t, J = 7.2 Hz, 3H, overlapped); ¹³C NMR (CDCl₃, 75 MHz): δ 208.7, 208.1, 171.2, 169.6, 166.1, 107.9, 105.7, 104.6, 53.9, 52.9, 32.7, 25.7, 24.3, 23.3, 12.8; CIMS: m/z 335[M-C₁₀H₁₀O₅]; analysis for C₃₅H₄₈O₁₀ (628.3), calcd, C, 66.86; H, 7.89; found, C, 66.70; H, 7.82. *Cyclohexyl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxybenzene) (15):* Yield: 33%; reddish brown solid; mp 158–160 °C; UV (CHCl₃): λ_{max} (log ε) 282 (4.49), 341 (3.90); IR (KBr): ν_{max} 3199, 2957, 2930, 1618, 1579, 1467, 1365, 1300, 1197, 1052 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ 18.04 (s, 1H, OH_C), 17.74 (s, 1H, OH_C), 16.46 (s, 1H, OH_B), 16.42 (s, 1H, OH_B), 10.96 (s, 1H, OH_A), 10.07 (s, 1H, OH_A), 4.17 (d, J = 10.8 Hz, 1H), 3.01 (br s, 8H), 2.82 (m, 1H), 2.27 (m, 4H), 1.74–1.52 (m, 6H), 1.00 (d, J = 5.7 Hz, 24H), 0.80–0.74 (m, 4H); ¹³C NMR (CDCl₃, 75 MHz): δ 208.2, 208.0, 171.1, 169.1, 163.3, 166.4, 166.0, 107.4, 107.1, 105.6, 104.5, 53.9, 53.0, 43.8, 37.5, 36.6, 32.9, 30.3, 26.8, 26.6, 25.9, 25.7, 23.3; CIMS: m/z 683 [M+1]⁺, 389 [M-C₁₀H₁₀O₅]; analysis for C₃₉H₅₄O₁₀ (682.4), calcd, C, 68.60; H, 7.97; found, C, 68.49; H, 8.06. *Phenyl-methylene-bis-(3,5-diacetyl-2,4,6-trihydroxybenzene) (16):* Yield: 30%; cream colored solid; mp 109–111 °C; UV (CHCl₃): λ_{max} (log ε) 272 (4.22), 339 (3.39); IR (KBr): ν_{max} 2925, 2676, 2562, 1688, 1603, 1584, 1455, 1424, 1327, 1292, 1180, 1128, 1073, 1027 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ 17.65 (s, 2H, 2 × OH_C), 16.37 (s, 2H, 2 × OH_B), 10.21 (s, 2H, 2 × OH_A), 7.30–7.22 (m, 3H), 7.10 (d, J = 7.3 Hz, 1H), 6.17 (s, 1H), 2.78 (s, 6H), 2.72 (s, 6H); ¹³C NMR (CDCl₃, 75 MHz): δ 205.9, 205.6, 171.6, 169.2, 166.6, 137.5, 128.9, 127.1, 127.0, 106.5, 105.8, 104.9, 34.0, 33.7, 32.9, 30.2; CIMS: m/z 299 [M-C₁₀H₁₀O₅]; analysis for C₂₇H₂₄O₁₀ (508.1), calcd, C, 63.78; H, 4.76; found, C, 63.64; H, 4.65. *Thiophen-2-yl-methylene-bis-(3,5-diacetyl-2,4,6-trihydroxybenzene) (17):* Yield: 42%; yellowish green solid; mp 178–180 °C; UV (CHCl₃): λ_{max} (log ε) 272 (4.29), 334 (3.65); IR (KBr): ν_{max} 3179, 1616, 1579, 1481, 1409, 1365, 1262, 1186, 1116; ¹H NMR (CDCl₃, 300 MHz): δ 17.80 (s, 2H, 2 × OH_C), 16.39 (s, 2H, 2 × OH_B), 10.32 (s, 2H, 2 × OH_A), 7.19 (d, J = 5.0 Hz, 1H), 6.91 (t, J = 4.0 Hz, 1H), 6.70 (s, 1H), 6.26 (s, 1H), 2.78 (s, 6H), 2.73 (s, 6H); ¹³C NMR (CDCl₃, 75 MHz): δ 206.0, 205.7, 171.8, 169.2, 166.3, 142.9, 127.0, 125.4, 124.9, 106.9, 106.8, 105.4, 34.0, 32.8, 31.1; CIMS: m/z 305 [M-C₁₀H₁₀O₅]; analysis for C₂₅H₂₀O₁₀ (514.1), calcd, C, 58.36; H, 4.31; S, 6.23; found, C, 58.22; H, 4.44; S, 6.11. *Thiophen-2-yl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxybenzene) (18):* Yield: 51%; dark yellow solid; mp 170–172 °C; UV (CHCl₃): λ_{max} (log ε) 286 (4.56), 340 (3.99); IR (KBr): ν_{max} 3172, 2958, 2870, 1615, 1577, 1470, 1434, 1202, 1169, 1128, 1067 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ 18.06 (s, 2H, 2 × OH_C), 16.60 (s, 2H, 2 × OH_B), 10.33 (s, 2H, 2 × OH_A), 7.19 (d, J = 5.1 Hz, 1H), 6.92 (dd, J = 3.6, 5.0 Hz, 1H), 6.70 (t, J = 1.6 Hz, 1H), 6.25 (s, 1H), 3.05 (d, J = 6.1 Hz, 8H), 2.27 (m, 4H), 1.00 (d, J = 4.9 Hz, 24 H); ¹³C NMR (CDCl₃, 75 MHz): δ 207.9, 207.6, 171.3, 168.9, 165.4, 142.8, 126.4, 124.9, 124.3, 106.6, 105.2, 104.3, 53.3, 52.3, 31.2, 25.2, 23.3; CIMS: m/z 390 [M-C₁₀H₁₀O₅]; analysis for C₃₇H₄₆O₁₀ (682.3), calcd, C, 65.08; H, 6.79; S, 4.70; found, C, 65.19; H, 6.87; S, 4.59. *Pyridin-2-yl-methylene-bis-(3,5-diacetyl-2,4,6-trihydroxybenzene) (19):* Yield: 44%; brown solid; mp 230–232 °C; UV (CHCl₃): λ_{max} (log ε) 276 (4.32), 339 (3.53); IR (KBr): ν_{max} 3096, 2933, 1610, 1454, 1360, 1294, 1219, 1117, 1028 cm⁻¹; ¹H NMR (DMSO-d₆, 300 MHz): δ 17.09 (s, 1H, OH), 8.82 (d, J = 5.3 Hz, 1H), 8.56 (br s, 1H), 8.00 (s, 1H), 7.84 (d, J = 7.9 Hz, 1H), 6.94 (s, 1H), 2.86 (s, 6H), 2.81 (s, 6H); ¹³C NMR (DMSO-d₆, 75 MHz): δ 204.1, 172.7, 160.1, 145.2, 143.1, 125.2, 124.0, 106.0, 33.4, 32.8; CIMS: m/z 510 [M+1]⁺, 300 [M-C₁₀H₁₀O₅]; analysis for C₂₆H₂₃N₂O₁₀ (509.1), calcd, C, 61.30; H, 4.55; N, 2.75; found, C, 61.21; H, 4.39; N, 2.60. *2,4-bis-(trifluoromethyl)-1-yl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxybenzene) (20):* Yield: 40%; yellow crystals, mp 211–213 °C. ¹H NMR (CDCl₃, 300 MHz): δ 7.93 (s, 1H), 7.75 (d, J = 8.1 Hz, 1H), 7.48 (d, J = 8.1 Hz, 1H), 6.59 (s, 1H), 2.99–3.07 (Overlapped signals, 8H), 2.26 (m, 4H), 0.99 (d, J = 6.6 Hz, 24H); ¹³C NMR (CDCl₃, 75 MHz): δ 208.4, 208.1, 207.5, 207.2, 171.6, 168.5, 166.1, 165.6, 142.2, 130.2, 130.00, 129.7, 129.3, 129.08, 128.8, 128.4, 126.1, 125.7, 124.9, 122.4, 122.1, 106.7, 105.7, 104.9, 53.9, 53.5, 52.9, 30.1, 25.6, 23.2; MALDI-TOF: m/z 812.02. *4-Biphenyl-1-yl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxybenzene) (21):* Yield: 43%; yellow crystals, mp 227–229 °C. ¹H NMR (CDCl₃, 300 MHz): δ 17.95 (br s, 2H, 2 × OH_C), 16.57 (s, 2H, 2 × OH_B), 10.32 (br s, 2H, 2 × OH_A), 7.57 (d, J = 7.3 Hz, 2H), 7.49 (d, J = 8.1 Hz, 2H), 7.40 (t, 2H), 7.30 (d, J = 7.2 Hz, 1H), 7.15 (d, J = 8.0 Hz, 2H), 6.18 (s, 1H), 3.03 (d, J = 12 Hz, 8H), 2.27 (m, 4H), 0.99 (d, J = 5.5 Hz, 24 H); ¹³C NMR (CDCl₃, 75 MHz): δ 208.4, 208.2, 171.6, 169.4, 166.2, 141.1, 139.8, 136.8, 129.3, 127.7, 127.6, 127.5, 106.7, 105.8, 104.9, 53.8, 52.9, 33.7, 25.7, 23.3; MALDI-TOF: m/z 751.47. calcd, C, 71.79; H, 6.96; found, C, 71.75; H, 6.94. *Benzofuran-2-yl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxybenzene) (22):* Yield: 41%; yellow crystals, mp 186–188 °C, IR (KBr): ν_{max} 3435, 2958, 1614, 1580, 1466, 1301, 1199, 1108, ¹H NMR (CDCl₃, 300 MHz): δ 17.95 (br s, 2H, 2 × OH_C), 16.57 (s, 2H, 2 × OH_B), 10.32 (br s, 2H, 2 × OH_A), 7.48 (d, 1H), 7.33 (d, 1H), 7.16–7.21 (m, 2H), 6.42 (s, 1H), 6.17 (s, 1H), 2.92 (d, J = 12 Hz, 8H), 2.28 (m, 4H), 0.98 (d, J = 5.5 Hz, 24 H); ¹³C NMR (CDCl₃, 75 MHz): δ 208.4, 208.2, 172.1, 169.2, 165.8, 155.4, 129.0, 125.5, 124.2, 123.6, 120.9, 111.4, 110.1, 105.7, 105.4, 53.8, 52.7, 37.3, 25.4, 23.3; MALDI-TOF: m/z 717.4. *Flourene-2-yl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxybenzene) (23):* Yield: 40%; yellow crystals, mp 204–206 °C. ¹H NMR (CDCl₃, 300 MHz): δ 17.95 (br s, 2H, 2 × OH_C), 16.60 (s, 2H, 2 × OH_B), 10.32 (br s, 2H, 2 × OH_A), 7.72 (d, J = 7.2 Hz, 1H), 7.66 (d, J = 7.8 Hz, 1H), 7.49 (d, J = 7.2, 1H), 7.32–7.25 (m, 3H), 7.11 (d, J = 8.1, 1H), 6.24 (s, 1H), 3.81 (s, 2H), 3.06 (d,

$J = 10.6$ Hz, 8H), 2.28 (m, 4H), 0.99 (d, $J = 6.7$ Hz, 24 H); ^{13}C NMR (CDCl_3 , 75 MHz): δ 208.4, 208.2, 171.6, 169.5, 166.2, 144.0, 143.8, 141.9, 140.7, 136.3, 127.3, 127.1, 125.8, 125.5, 123.7, 120.3, 120.2, 107.0, 105.8, 104.9, 53.9, 52.9, 37.5, 34.1, 25.7, 23.3; calculated m/z for $\text{C}_{46}\text{H}_{52}\text{O}_{10}$ (764.36), found CIMS: m/z 471 [$\text{M}-\text{C}_{16}\text{H}_{21}\text{O}_5$], calcd, C, 72.23; H, 6.85 found, C, 72.28; H, 6.79. **Imidazole-2-yl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxy benzene) (24)**: Yield: 41%; white solid; mp 222–224 °C UV (CHCl_3): λ_{max} (log ϵ) 280 (4.51), 340 (3.72); IR (Neat): ν_{max} 3435, 2957, 1614, 1416, 1124, 917 cm^{-1} ; ^1H NMR (CDCl_3 , 300 MHz): δ 16.81 (s, 2H, $2 \times \text{OH}_C$), 15.31 (s, 2H, $2 \times \text{OH}_B$), 7.18 (s, 2H), 7.02 (s, 1H), 3.38 (d, $J = 8.7$ Hz, 2H), 3.11 (d, $J = 8.7$ Hz, 2H), 2.69 (d, $J = 7.8$ Hz, 2H), 2.36 (d, $J = 9.6$ Hz, 2H), 2.15 (m, 2H), 2.19 (m, 2H), 0.95 (d, $J = 6.7$ Hz, 24 H); ^{13}C NMR (CDCl_3 , 75 MHz): δ 208.1, 206.4, 174.4, 174.1, 170.3, 152.2, 118.3, 107.1, 104.6, 102.8, 53.3, 53.09, 27.07, 26.2, 25.8, 23.6, 23.4, 23.2, 23.1; MALDI-TOF: m/z 667.6. calcd, C, 64.85; H, 6.95; N, 4.20; found, C, 64.78; H, 6.86; N, 4.25. **1-Naphthyl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxy benzene) (25)**: Yield: 35%; brown crystals; mp 255–256 °C ^1H NMR (CDCl_3 , 300 MHz): δ 16.40 (s, 2H, $2 \times \text{OH}_C$), 10.42 (s, 2H, $2 \times \text{OH}_A$), 7.86 (d, $J = 8.1$ Hz, 1H), 7.80 (d, $J = 8.1$ Hz, 1H), 7.60–7.77 (m, 2H), 7.56 (d, $J = 8.7$ Hz, 1H), 7.44–7.28 (aromatic protons, 3H), 6.56 (s, 1H), 3.04 (d, $J = 10.6$ Hz, 8H), 2.27 (m, 4H), 0.98 (d, $J = 6.3$ Hz, 24 H); ^{13}C NMR (CDCl_3 , 75 MHz): δ 208.2, 171.1, 137.1, 135.8, 134.7, 133.3, 132.1, 129.7, 129.0, 128.8, 127.5, 126.6, 126.1, 125.6, 125.4, 123.4, 105.3, 53.5, 32.9, 25.7, 23.3, calculated m/z for $\text{C}_{43}\text{H}_{50}\text{O}_{10}$ (726.34), found CIMS: m/z 727.3 **3,4-methylenedioxy-1-yl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxybenzene) (26)**: Yield: 41%; yellow solid; mp 186–188 °C ^1H NMR (CDCl_3 , 300 MHz): δ 17.9 (br s, 2H, $2 \times \text{OH}_C$), 16.56 (s, 2H, $2 \times \text{OH}_B$), 10.24 (br s, 2H, $2 \times \text{OH}_A$), 6.71 (d, $J = 8.1$ Hz, 1H), 6.58 (s, 1H), 6.55 (d, $J = 8.7$ Hz, 1H), 6.07 (s, 1H), 5.94 (s, 1H), 3.04 (d, $J = 14.4$ Hz, 8H), 2.26 (m, 4H), 0.99 (d, $J = 6.5$ Hz, 24 H); ^{13}C NMR (CDCl_3 , 75 MHz): δ 208.4, 208.1, 171.5, 169.3, 166.1, 148.5, 146.6, 131.4, 119.9, 108.4, 107.9, 106.7, 105.7, 104.8, 101.6, 53.8, 52.8, 33.6, 25.7, 23.3, MALDI-TOF: m/z 720.471. **Thianaphene-2-yl-methylene-bis-(3,5-di-isopentanoyl-2,4,6-trihydroxybenzene) (27)**: Yield: 43%; yellow crystals; mp 172–174 °C ^1H NMR (CDCl_3 , 300 MHz): δ 16.48 (s, 2H, $2 \times \text{OH}_B$), 10.24 (br s, 1H, OH_A), 9.88 (br s, 1H, OH_A), 7.83 (d, $J = 7.1$ Hz, 1H), 7.17–7.35 (overlapped signals, 3H), 7.06 (s, 1H), 6.16 (s, 1H), 3.05 (d, $J = 10.6$ Hz, 8H), 2.27 (m, 4H), 0.98 (d, $J = 6.7$ Hz, 24 H); ^{13}C NMR (CDCl_3 , 75 MHz): δ 208.3, 171.4, 168.9, 141.05, 138.7, 132.0, 124.9, 124.6, 123.5, 121.9, 106.8, 105.6, 104.9, 53.8, 53.0, 31.0, 25.7, 23.3; calculated m/z for $\text{C}_{41}\text{H}_{48}\text{O}_{10}$ (732.88), found CIMS: m/z ; 732.3. calcd, C, 67.19; H, 6.60; found, C, 67.23; H, 6.58.

General experimental procedures for compound 29–37

The mixture of 3-formyl phloroisovalerophenone (**8**) (1 mmol), methanesulfonic acid (3 mmol) and aldehyde (0.5 mmol) in was heated at 80 °C for 30 min. After completion of reaction (TLC), water was added to the reaction mixture and extracted with ethyl acetate. Organic layer was dried over sodium sulphate and the solvent was removed under vacuum and the crude product was purified by reverse phase silica gel column chromatography using methanol: water as eluent.

Pyridine-3-yl-methylene-bis-(3-isopentanoyl-5-formyl-2,4,6-trihydroxybenzene) (29): Yield: 44%; yellow solid; mp 156–157 °C; UV (CHCl_3): λ_{max} (log ϵ) 268 (3.40); IR (KBr): ν_{max} 3470, 2930, 1630, 1545, 1470, 1366, 1302, 1045 cm^{-1} ; ^1H NMR ($\text{DMSO}-d_6$, 300 MHz): δ 9.90 (s, 2H), 8.48 (d, $J = 5.4$ Hz, 1H), 8.27 (s, 1H), 7.69 (d, $J = 7.5$ Hz, 1H), 7.54 (t, $J = 4.8$ Hz, 1H), 6.56 (s, 1H), 2.88 (d, $J = 5.4$ Hz, 4H), 2.10 (m, 2H), 0.83 (d, $J = 6.6$ Hz, 12H); ^{13}C NMR ($\text{DMSO}-d_6$, 75 MHz): δ 209.0, 207.0, 194.9, 192.1, 173.6, 171.4, 146.2, 142.0, 140.0, 127.9, 107.8, 103.1, 53.9, 32.0, 26.9, 23.5; Calcd m/z for $\text{C}_{30}\text{H}_{31}\text{NO}_{10}$ (565.57): found CIMS: m/z 566.1, analysis for $\text{C}_{30}\text{H}_{31}\text{NO}_{10}$, C, 67.71; H, 5.52; N, 2.48. Found: C, 67.62; H, 5.56; N, 2.43. **Pyridine-4-yl-methylene-bis-(3-isopentanoyl-5-formyl-2,4,6-trihydroxybenzene) (30)**: Yield: 40%; yellow solid; mp 158–159 °C ^1H NMR (CD_3OD , 300 MHz): δ 9.89 (s, 2H), 8.38 (d, $J = 5.3$ Hz, 2H), 7.78 (d, $J = 5.3$ Hz, 2H), 2.86 (d, $J = 5.4$ Hz, 4H), 2.14 (m, 2H), 0.87 (d, $J = 6.6$ Hz, 12H); ^{13}C NMR (CD_3OD , 75 MHz): δ 206.9, 194.9, 191.8, 173.4, 164.4, 145.3, 126.1, 108.3, 53.9, 34.6, 31.1, 26.6, 23.5; calculated m/z for $\text{C}_{30}\text{H}_{31}\text{NO}_{10}$ (565.57), found CIMS: m/z 566.0. calcd,

C, 63.71; H, 5.52; N, 2.48; found, C, 63.77; H, 5.49; N, 2.43. **Flourene-2-yl-methylene-bis-(3-isopentanoyl-5-formyl-2,4,6-trihydroxybenzene) (31)**: Yield: 41%; yellow solid; mp 216–218 °C IR (Neat): ν_{max} 3369, 1618, 1404, 1344, 1275, 1261, 1126, 1053 cm^{-1} ; ^1H NMR (CDCl_3 , 300 MHz): δ 17.90 (br s, 2H, $2 \times \text{OH}$), 14.68 (s, 2H, $2 \times \text{OH}$), 10.16 (s, 1H) 7.72 (d, $J = 7.2$ Hz, 1H), 7.66 (d, $J = 7.8$ Hz, 1H), 7.49 (d, $J = 7.2$, 1H), 7.32–7.25 (m, 3H), 7.11 (d, $J = 8.1$, 1H), 6.20 (s, 1H), 3.82 (s, 2H), 3.03 (d, $J = 5.7$ Hz, 4H), 2.26 (m, 2H), 1.00 (d, $J = 6.3$ Hz, 12H); ^{13}C NMR (CDCl_3 , 75 MHz): 208.11, 194.0, 171.4, 169.5, 166.1, 144.2, 143.8, 141.7, 140.9, 135.4, 127.3, 127.2, 125.7, 125.5, 123.6, 120.3, 107.3, 106.4, 105.7, 104.4, 52.6, 37.5, 33.4, 32.4, 30.2, 25.6, 23.3; MALDI-TOF: m/z 652.08. calcd, C, 69.93; H, 5.56; found, C, 69.87; H, 5.45. **Imidazole-2-yl-methylene-bis-(3-isopentanoyl-5-formyl-2,4,6-trihydroxybenzene) (32)**: Yield: 43%; white solid; IR (Neat): ν_{max} 3431, 2919, 1614, 1404, 1275, 1260, 1109 cm^{-1} ; ^1H NMR ($\text{DMSO}-d_6$, 300 MHz): δ 16.05 (br s, 1H, OH_C), 15.92 (s, 1H, OH_C), 15.12 (s, 1H, OH_B), 13.68 (br s, 1H, OH_A), 13.46 (s, 1H, OH_A), 9.94 (s, 2H), 7.47 (s, 2H), 6.55 (s, 1H), 2.93 (d, $J = 5.7$ Hz, 4H), 2.15 (m, 2H), 0.92 (d, $J = 6.3$ Hz, 12H); ^{13}C NMR ($\text{DMSO}-d_6$, 75 MHz): δ 207.0, 204.9, 193.6, 190.2, 173.3, 171.7, 170.4, 167.3, 148.8, 119.0, 106.5, 104.3, 101.3, 51.9, 25.7, 25.1, 22.9; MALDI-TOF: m/z 555.13. calcd, C, 60.64; H, 5.45; N, 5.05; found, C, 60.61; H, 5.36; N, 4.96. **Benzofuran-2-yl-methylene-bis-(3-isopentanoyl-5-formyl-2,4,6-trihydroxybenzene) (33)**: Yield: 39%; cream colored solid; IR (Neat): ν_{max} 3369, 2958, 2869, 1620, 1275, 1454, 1425, 1187, 1123 cm^{-1} ; ^1H NMR (CD_3OD , 300 MHz): δ 10.04 (s, 2H) 7.38 (d, 1H), 7.29 (d, 1H), 7.08 (m, 2H), 6.55 (s, 1H), 6.29 (s, 1H), 2.94 (d, $J = 5.7$ Hz, 4H), 2.18 (m, 2H), 0.93 (d, $J = 6.3$ Hz, 12 H); ^{13}C NMR (CD_3OD , 75 MHz): δ 205.2, 192.4, 171.0, 168.7, 168.1, 157.3, 154.2, 128.0, 122.1, 121.7, 119.1, 113.0, 109.4, 105.0, 104.0, 101.5, 51.5, 28.5, 24.3, 21.02; MALDI-TOF: m/z 621.36 ($\text{M}+\text{NH}_3$). calcd, C, 65.56; H, 5.33; found, C, 65.63; H, 5.39. **3,4-Methylenedioxy-1-yl-methylene-bis-(3-isopentanoyl-5-formyl-2,4,6-trihydroxybenzene) (34)**: Yield: 46%; yellow solid; mp 190–192 °C; UV (CHCl_3): λ_{max} (log ϵ) 282 (4.53), 340 (3.95); IR (KBr): ν_{max} 3429, 2924, 1620, 1487, 1422, 1122, 1041 ^1H NMR (CD_3OD , 300 MHz): δ 10.16 (s, 2H), 6.72 (d, $J = 7.8$ Hz, 1H), 6.57 (s, 1H), 6.55 (d, $J = 7.8$ Hz, 1H), 6.03 (s, 1H), 5.94 (s, 2H), 3.08–2.99 (m, 4H), 2.21 (m, 2H), 0.97 (d, $J = 6.6$ Hz, 12H); ^{13}C NMR (CD_3OD , 75 MHz): δ 206.2, 194.5, 170.1, 168.3, 167.8, 148.8, 146.3, 120.3, 111.3, 110.6, 109.4, 108.3, 101.7, 53.5, 32.6, 26.9, 26.7, 23.2; calculated m/z for $\text{C}_{38}\text{H}_{47}\text{NO}_{10}$ (608), found CIMS: m/z 609.2. calcd, C, 63.15; H, 5.30; found, C, 63.20; H, 5.36. **2,4-bis-(trifluoromethyl)-1-yl-methylene-bis-(3-isopentanoyl-5-formyl-2,4,6-trihydroxybenzene) (35)**: Yield: 35% cream colour solid; mp 162–164 °C ^1H NMR (CD_3OD , 300 MHz): δ 9.98 (s, 2H), 7.88 (s, 1H), 7.78 (d, $J = 8.4$ Hz, 1H), 7.61 (d, $J = 8.4$ Hz, 1H), 6.84 (s, 1H), 2.96 (d, $J = 6.6$, 4H), 2.15–2.24 (m, 2H), 0.94 (d, $J = 6.6$ Hz, 12H); ^{13}C NMR (CD_3OD , 75 MHz): δ 205.6, 191.2, 170.0, 167.4, 165.3, 130.7, 127.1, 125.0, 124.7, 122.6, 121.4, 121.1, 107.4, 104.1, 103.1, 51.7, 31.1, 24.1, 20.9; calculated m/z for $\text{C}_{33}\text{H}_{30}\text{F}_6\text{O}_{10}$ (700.58), found CIMS: m/z 701.4. **4-Benzyloxy-1-yl-methylene-bis-(3-isopentanoyl-5-formyl-2,4,6-trihydroxybenzene) (36)**: Yield: 33%; cream colour solid; mp 195–197 °C; ^1H NMR (CDCl_3 , 300 MHz): δ 17.84 (br s, 2H, $2 \times \text{OH}_C$), 14.6 (s, 2H, $2 \times \text{OH}_B$), 10.16 (s, 2H), 9.63 (s, 2H, $2 \times \text{OH}_A$), 7.42–7.30 (m, 5H), 7.00 (d, $J = 8.7$ Hz, 2H), 6.89 (d, $J = 8.7$ Hz, 2H), 6.06 (s, 1H), 5.04 (s, 2H), 3.05–2.96 (m, 4H), 2.22–2.31 (m, 2H), 1.00 (d, $J = 6.6$ Hz, 12H); ^{13}C NMR (CDCl_3 , 75 MHz): δ 208.11, 194.0, 171.4, 169.4, 166.0, 158.0, 137.4, 129.1, 128.7, 128.5, 128.2, 128.0, 115.3, 107.2, 105.6, 104.3, 70.6, 53.9, 52.6, 32.5, 25.6, 23.3; calculated m/z for $\text{C}_{38}\text{H}_{38}\text{O}_{11}$ (670.24) found CIMS: m/z 671.20. calcd, C, 68.05; H, 5.71; found, C, 68.11; H, 5.66. **4-Biphenyl-1-yl-methylene-bis-(3-isopentanoyl-5-formyl-2,4,6-trihydroxybenzene) (37)**: Yield: 35%; white solid; mp 247–249 °C; ^1H NMR (CD_3OD , 300 MHz): δ 10.01 (s, 2H), 7.55 (d, $J = 7.5$ Hz, 2H), 7.45 (d, $J = 7.8$ Hz, 2H), 7.36 (t, $J = 7.2$ Hz, 2H), 7.26 (d, $J = 7.2$ Hz, 1H), 7.12 (d, $J = 7.2$ Hz, 2H), 6.55 (s, 1H), 2.96 (d, $J = 6.0$ Hz, 4H), 2.15–2.23 (m, 2H), 0.95 (d, $J = 6.0$ Hz, 12H); ^{13}C NMR (CDCl_3 , 75 MHz): δ 207.7, 193.7, 174.2, 171.5, 142.9, 139.7, 130, 128.6, 128.4, 128.1, 127.9, 110.3, 106.6, 105.9, 54.0, 33.4, 27.0, 23.5; calculated m/z for $\text{C}_{37}\text{H}_{36}\text{O}_{10}$ (640.23), found CIMS: m/z 403.4 [$\text{M}-\text{C}_{12}\text{H}_{13}\text{O}_5$]. calcd, C, 69.36; H, 5.66; found, C, 69.42; H, 5.60.