

Synthesis and antitubercular activity of nucleoside analogs based on L-ascorbic acid and bases

R. P. Tripathi · Namrata Dwivedi · Nimisha Singh ·
Mridul Misra

Received: 22 October 2007 / Accepted: 31 October 2007 / Published online: 20 December 2007
© Birkhäuser Boston 2007

Abstract 5,6-*O*-isopropylidene-2,3-di-*O*-methyl ascorbic acid (**2**), obtained by reaction of acetone with ascorbic acid (**1**) followed by methylation with methyl iodide, on 1,8-Diazabicyclo[5.4.0]undec-7-ene-catalyzed elimination of acetone moiety led to the formation of respective 2,3-di-*O*-methyl didehydro-L-ascorbic acid (**4**) in good yield. The latter, on methanesulfonylation with methanesulfonyl chloride and subsequent reaction of the crude methanesulfonyloxy derivative with imidazole, benzimidazole, and adenine resulted in the corresponding tetronolactonyl nucleoside analogs **5**, **6**, and **7**. Compound **5** on reaction with benzyl amine led to the *N*-benzylated teramyl nucleoside analog, while compounds **6** and **7** did not react under similar conditions. All the synthesized compounds were evaluated for their antitubercular activity against *M. tuberculosis* H₃₇R_a and H₃₇R_v, exhibiting a minimum inhibitory concentration (MIC) of more than 12.5 µg/mL.

Keywords Ascorbic acid · Vitamin C · Nucleosides · Adenine · Tetramic acid

Introduction

Ascorbic acid, commonly known as vitamin C, has served as an excellent scaffold for the synthesis of many biologically active compounds (Cinatl *et al.*, 1995; Bram *et al.* 1980; Grdisa *et al.*, 1995). Its antioxidant activity and the immunomodulatory activities of many of its *O*-alkyl derivatives have been studied extensively (Olabisi and Wimalasena, 2004). Ascorbic acid is basically a tetranolactone and the two enols at the 2- and 3- positions play a crucial role both in organic synthesis and in eliciting the

R. P. Tripathi (✉) · N. Dwivedi · N. Singh · M. Misra
Medicinal and Process Chemistry Division, Central Drug Research Institute, Lucknow 226001,
India
e-mail: rpt.cdri@gmail.com

biological response. Replacement of the ring oxygen atom with sulfur in the tetranolactone results in thionotetranolactone, a pharmacophore in compounds exhibiting antimalarial and antitubercular activities via Fatty acid synthetase-II inhibition. One of such compounds possessing the thiotetranolactone moiety, thiolactomycin, is under preclinical development as a new antitubercular drug (Miyakawa *et al.*, 1982; Kremer *et al.*, 2000). However, replacement of the ring oxygen with nitrogen atom results in tetramates, another moiety found in a variety of antibiotics (Royles, 1995; Ley *et al.*, 1992; Wang *et al.*, 2003; Elbe *et al.*, 1956) and anti-HIV (Roggoet *et al.*, 1994), anticancer, and antitumor agents (Holtzel *et al.*, 2000; Iwata *et al.*, 2005). Very recently a few reports have come to light where ascorbic-based nucleosides were prepared as antiviral and antitumor agents (Malic *et al.*, 2000). Nucleoside analogs themselves are known for their diverse range of biological activities (Mishra *et al.*, 2005). Ascorbic acid has also been used in the synthesis of pyrano[3,4-b]indoles and a variety of other heterocycles by Preobrzhenskaya and co-workers (Larvrenov *et al.*, 2005; Preobrzhenskaya and Korolev 2000).

In our continuing program aimed towards the development of new antitubercular agents we were curious to synthesize certain teramic-acid-based nucleosides and investigate their antitubercular activity. Our curiosity arose due to the fact that many purines, nucleoside analogs, and thiolactomycins have displayed significant antitubercular activity (Miyakawa *et al.*, 1982; Kremer *et al.*, 2000; Somu, 2006). Thus herein we describe the methodology to synthesize tetranolactonyl and tetramyl nucleoside analogs. The synthesized compounds were evaluated for their antitubercular activities.

Results and discussion

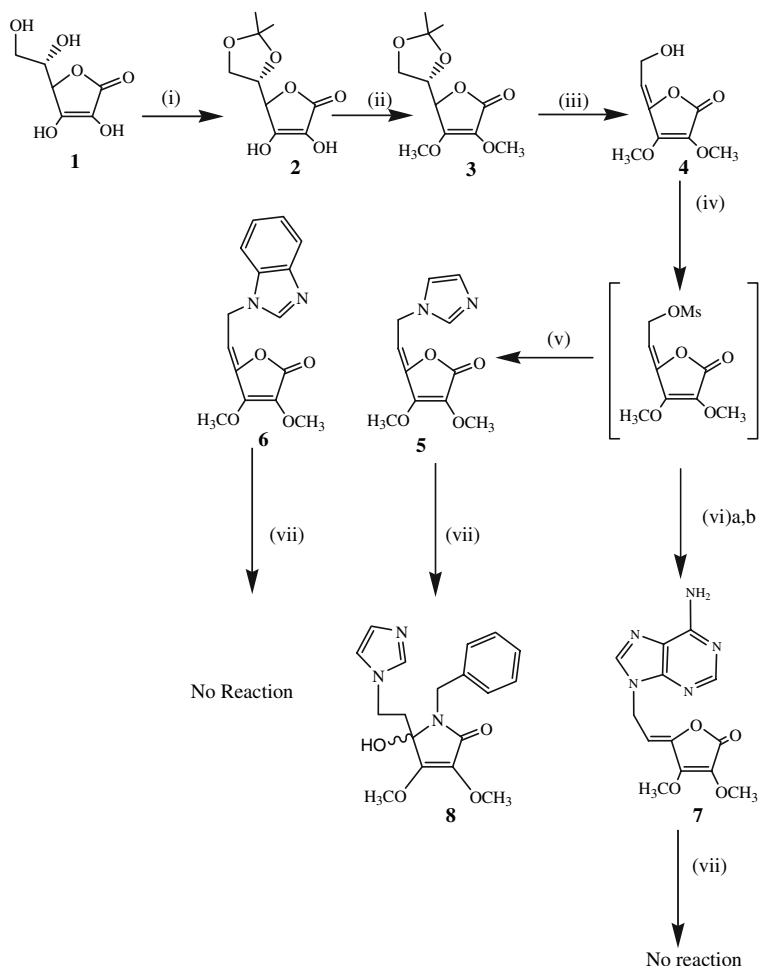
The synthetic strategy involves the reaction of ascorbic acid (**1**) with acetone in the presence of a catalytic amount of acetyl chloride, leading to selective protection of the 5,6-diol to give 5,6-*O*-isopropylidene-L-ascorbic acid (**2**) in quantitative yield. This compound (**2**) was methylated with methyl iodide in dimethyl sulfoxide (DMSO)/acetone (1:4) as the solvent in the presence of K₂CO₃ and tetrabutylammonium bromide (TBAB) to give 2,3-*O*-bis-methyl-5,6-*O*-isopropylidene ascorbic acid (**3**) (Khan and Adams, 1999). The latter was reacted with DBU (50 mol%) using Tetrahydrofuran as the solvent, leading to the elimination of a molecule of acetone to give 3,4-bis-methyl-5-(2-hydroxyethylidene)-5*H*-furan-2-one (**4**) (Singh *et al.*, 2006). Compound **4** was treated with methanesulfonyl chloride in dry CH₂Cl₂ in the presence of a catalytic compound of triethylamine, resulting in the intermediate methanesulfonyloxy derivative (**5**), which was used as such in the subsequent steps of the desired synthesis.

The intermediate methanesulfonyloxy derivative (**4**) was reacted with imidazole and a catalytic amount of DBU (25 mol%) in dry toluene at 120°C to give (*Z*)-5-[2-(imidazol-1-yl)ethylidene]-3,4-dimethoxy-5*H*-furan-2-one (**5**) at 65.7% yield.

A similar reaction of compound **5** with benzimidazole gave the corresponding (*Z*)-5-[2-(benzimidazol-1-yl) ethylidene]-3,4-dimethoxy-5*H*-furan-2-one (**6**) in good yield.

To prepare the adenine derivative of thiotetranolactone, the adenine was persilylated by refluxing it with 1,1,1,3,3,3-Hexamethyldisilazane in anhydrous toluene in the presence of a catalytic amount of $(\text{NH}_4)_2\text{SO}_4$ under a nitrogen atmosphere for 3–4 h. The silylated adenine thus obtained was reacted with methanesulfonyloxy tetranolactone derivative **5** as above using DBU (25 mol%) in refluxing toluene to give (Z)-5-[2-(adenin-9-yl) ethylidene]-3,4-dimethoxy-5H-furan-2-one (**7**) at 10% yield (Scheme 1).

The reaction of the imidazolyl tetranolactone **6** with benzyl amine in ethanol gave the corresponding tetramate, 1-benzyl-5-hydroxy-5-(2-imidazol-1-yl-ethyl)-3,4-dimethoxy-1,5-dihydro-pyrrol-2-one (**8**) at very good yield. Similar reactions of



Scheme 1 (i) Acetone, Acetylchloride, 30°C, 4 h (ii) MeI, TBAB, DMSO:Acetone (1:4), K₂CO₃, 30°C, 18 h (iii) THF, DBU, 30°C, 7 h (iv) Methanesulfonyl chloride, CH₂Cl₂, Et₃N, 0–30°C, 5 h (v) imidazole or benzimidazole, Toluene, DBU, 120°C, 3–6 h. (vi) a. adenine, HMDS, (NH₄)₂SO₄, reflux, 3 h; b. DBU, 120°C, 3–4 h (vii) Ethanol, benzyl amine 3–4 h

compounds **6** and **7** did not afford the desired tetramates. The possible reason for the inertness of (Z)-5-[2-(benzimidazol-1-yl)ethylidene]-3,4-dimethoxy-5H-furan-2-one (**6**) and (Z)-5-[2-(adenin-9-yl)ethylidene]-3,4-dimethoxy-5H-furan-2-one (**7**) towards benzyl amine may be due the steric hindrance offered during reaction by the bulky benzimidazole and adenine moieties. Thus among the compounds **5**, **6**, and **7** described above only (Z)-5-[2-(imidazol-1-yl)ethylidene]-3,4-dimethoxy-5H-furan-2-one (**5**) undergoes a ring-opening reaction with the benzyl amine to give the enol-keto-amide, leading to compound **8**. The structures of all the products synthesized were established on the basis of their spectroscopic data and analysis (see the Experimental Section).

Biological activity

The ascorbic-acid-based nucleosides **5**, **6**, **7**, and **8** were evaluated for their antitubercular activity against *M. Tuberculosis* H₃₇R_a by Micro alamar blue assay technique (Collins and Franzblan, 1997); while the agar microdilution method (Saito *et al.*, 1991) was used against H₃₇R_v. As evident from Table 1 none of the compounds showed significant activity against *M. tuberculosis* H₃₇R_A and the virulent H₃₇R_A strains since the MIC values for them are greater than 25 µg/mL as compared to the standard drug Isoniazid. One of the possible reasons for their inactivity against mycobacterium may be their transport or instability in the cellular medium.

In summary, we have developed a simple method for the synthesis of tetranolctonyl nucleosides which can be manipulated to tetramyl nucleoside analogs. The compounds have been screened against *M. tuberculosis* and exhibited MIC values above 25 µg/mL.

Experimental section

5,6-*O*-isopropylidene-L-ascorbic acid (**2**)

To a magnetically stirred solution of ascorbic acid (30 g, 170.4 mmol) in acetone (120 mL), acetyl chloride (3 mL, 42.6 mmol) was added and the reaction mixture stirred for 2–3 h at ambient temperature. The reaction mixture was kept for 7–8 h in

Table 1 Antimycobacterial activity of synthesized compounds **5–8**

Sample number	Compound	H ₃₇ R _a MIC (µgmL ⁻¹)	H ₃₇ R _v MIC (µgmL ⁻¹)
1	5	>12.5	>25
2	6	>12.5	>25
3	7	>12.5	>25
4	8	>12.5	>25
INH	–	–	0.65

MIC = minimum inhibitory concentration

freezer, the solid separated was filtered and washed with cold acetone and was dried to give compound **2** as colorless granules (27 g, 73.7%), m.p. 195–198°C. lit m.p.

2,3-Dimethoxy-5, 6-*O*-isopropylidene-L-ascorbic acid (**3**)

A mixture of compound **2** (25 g, 115.7 mmol) and anhydrous K₂CO₃ (32 g, 231.4 mmol) in acetone:dimethyl sulfoxide (4:1, 250 mL) was magnetically stirred for 25 min. A solution of methyl iodide (14.9 mL, 231.4 mmol) was dissolved slowly in acetone (30 mL) for 30 min, followed by the addition of tetrabutylammonium bromide (2.0 g) and stirring overnight at ambient temperature. The result was filtered and the filtrate thus obtained was evaporated under reduced pressure to give the crude mass, which was dissolved in ethyl acetate and washed with water. The organic layer was dried (Na₂SO₄) and evaporated under reduced pressure to afford a crude solid, which was chromatographed over silica gel (230–400 mesh) using a gradient of hexane-EtOAc (17:3) as eluent to give the compound **3** as a colorless solid (22 g, 78%).

(*Z*)-3,4-Dimethoxy-5-(2-hydroxyethylidene)-5*H*-furan-2-one (**4**)

To a magnetically stirred solution of compound **3** (20 g, 81.96 mmol) in THF (80 mL), DBU (6.2 mL, 50 mol%) was added slowly and the reaction mixture was stirred for 18 h at room temperature. The solvent was evaporated under reduced pressure and the residue thus obtained was dissolved in ethyl acetate (100 mL) and washed with water (25 mL). The organic layer was dried (Na₂SO₄) and concentrated under reduced pressure to afford a crude mass, which was chromatographed over silica gel (230–400 mesh) using hexane:ethyl acetate (9:1) as eluent to give the above compound **4** as a colorless solid (10 g, 65.7%), m.p. 60°C, infrared(IR) (neat) 3391, 1688 cm⁻¹, MS (FAB) 187(M + H)⁺ ¹H NMR (200 MHz, CDCl₃) δ_H 1.39 (bs, 1H), 3.92 (s, 3H, OCH₃), 4.16 (s, 3H, -OCH₃), 4.41 (d, *J* = 7.0, 2H, OCH₂), 5.50 (t, *J* = 7.0 Hz, 1H, =CH–); ¹³C NMR (50 MHz, CDCl₃): δ 56.6, 59.9, 60.6, 108.1, 125.0, 142.3, 149.1, 164.7.

Preparation of the intermediate 5-(2-methanesulfonyloxy ethylidene)-3,4-dimethoxy-5*H*-furan-2-one

To a stirring solution of compound **4** (2 g, 10.65 mmol) and triethylamine (1 mL) in anhydrous dichloromethane (10 mL) a solution of methanesulfonyl chloride (1.11 mL, 10.65 mmol) in CH₂Cl₂ (2 mL) was added slowly at 0°C during 5 min. The reaction mixture was brought to 30°C and stirring continued for a further hour. The solvent was evaporated under reduced pressure and the residue thus obtained was dissolved in ethyl acetate (100 mL) and washed with water (25 mL). The organic layer was dried (Na₂SO₄) and concentrated under reduced pressure to give a crude mass of the title compound, which was used as such in the subsequent reactions.

(Z)-5-[2-(Imidazol-1-yl)ethylidene]-3, 4-dimethoxy-5*H*-furan-2-one (**5**)

A mixture of the above crude methanesulfonyloxy derivative (1 g, 3.78 mmol) imidazole (0.257 g, 3.78 mmol) and DBU (0.285 mL, 25 mol%) in anhydrous toluene was stirred at 120°C for 5 h. The solvent was evaporated under reduced pressure and the residue thus obtained was dissolved in ethyl acetate (100 mL) and washed with water (25 mL). The organic layer was dried (Na₂SO₄) and concentrated under reduced pressure to afford a crude mass, which was chromatographed over silica gel (230–400 mesh) using a gradient of hexane:ethylacetate (3:2) as eluent to give the above compound **6** as a light yellow foam (1 g, 65.7%), as foam IR (KBr) ν_{\max} 3210, 1775 ; FABMS 237(M + H)⁺; ¹H NMR (200Mz, CDCl₃) δ 3.95 (s, 3H, OCH₃), 4.14 (s, 3H, OCH₃), 4.81 (d, J = 7.4 Hz, 2H, CH₂), 5.73 (t, J = 7.4 Hz, 1H, = CH), 6.93, 7.07 (2s, 2H, imidazole CH), 7.51 (s, 1H, imidazolyl CH); analysis calculated for C₁₁H₁₂N₂O₄ C, 55.92; H, 5.12, N, 11.85; found: C, 55.87; H, 5.08; N, 11.93.

(Z)-5-[2-(Adenin-9-yl)ethylidene]-3,4-dimethoxy-5*H*-furan-2-one (**7**)

Adenine (1.5 g, 11.1 mmol) was refluxed in anhydrous toluene with HMDS in the presence of a catalytic amount of (NH₄)₂SO₄ under an N₂ atmosphere for 3–4 h and the excess reagent and solvent were removed under reduced pressure to give the silylated adenine derivative. A mixture of this silylated adenine derivative, the above intermediate methanesulfonyloxy derivative (1 g, 3.78 mmol), and DBU (0.285 mL, 50 mol%) in anhydrous toluene (5 mL) was stirred at 120°C for 5 h. The solvent evaporated, and the residue thus obtained was dissolved in chloroform and washed with water and the organic layer dried (Na₂SO₄) and solvent-evaporated under reduced pressure to afford a crude mass. The latter was chromatographed over an SiO₂ column using CH₂Cl₂:MeOH (9:1) as eluent to give the title compound **7** as colorless granules (0.32 g) at 10% yield: m.p. 138–140°C; IR (KBr) ν_{\max} 3210, 1775 ; FABMS 304(M + H)⁺; ¹H NMR (200Mz, DMSO) δ 3.82 (s, 3H, OCH₃), 4.08 (s, 3H, OCH₃), 5.15 (d, J = 7.0 Hz, 2H, CH₂), 5.73 (t, J = 7.0 Hz, 1H, = CH), 7.76 (s, 1H, CH), 7.95 (br, 2H, NH₂), 8.14 (s, 1H, CH); analysis calculated for C₁₃H₁₃N₅O₄ C, 51.48; H, 4.29, N, 23.10; found: C, 51.40; H, 4.32; N, 23.18.

1-Benzyl-5-hydroxy-5-(2-imidazol-1-yl-ethyl)-3,4-dimethoxy-1,5-dihydro-pyrrol-one (**8**)

A solution of compound **5** (0.2 g, 0.847 mmol) and benzylamine (0.09 mL 0.847 mmol) in ethanol (1 mL) was stirred at 40°C for 5 h. The solvent was evaporated to a crude product which was chromatographed over silica gel (230–400 mesh) using a gradient of hexane:ethylacetate (3:2) as eluent to give compound **8** as a light yellow foam. Yield (174 mg, 60 %); IR (KBr) ν_{\max} 3210, 1775 cm⁻¹; FABMS 343(M + H)⁺; ¹H NMR (200Mz, CDCl₃) δ 2.00– 2.24 (m, 2H, H-6), 3.22– 3.40 (m, 2H, H-7), 3.85 (s, 3H, OCH₃), 4.00 (s, 3 H, OCH₃), 4.21 (d, J = 15.2 Hz, 1 H,

NCH_{2A}), 4.90 (d, $J = 15.2$ Hz, 1 H, NCH_{2B}), 6.30 (s, 1H, imidazole H), 6.85 (m, 2H, imidazole CH), 7.33–7.43 (m, 5H, Ar-H); analysis calculated for C₁₈H₂₁N₃O₄: C, 62.93; H, 6.16, N, 12.24; found: C, 62.90; H, 6.16; N, 12.24.

Antitubercular screening

Activity against *M. tuberculosis* H₃₇R_a strain

All the compounds synthesized were evaluated for their efficacy against *M. tuberculosis* H₃₇R_a at concentrations ranging from 100 $\mu\text{g mL}^{-1}$ to 1.56 $\mu\text{g mL}^{-1}$ using twofold dilutions in the initial screen. Log phase culture of *M. tuberculosis* H₃₇R_a was diluted so as to give a final optical density (OD)_{550 nm} of 0.05 in Sauton's medium. In 96-well white plates, 190 μl of culture is dispensed in each well. A dimethyl sulfoxide (DMSO) solution of test compounds was dispensed into 96-well plates so as to make the final test concentration 25 $\mu\text{g mL}^{-1}$ (5 μg test compound dispensed in 10 μl of DMSO). Then the plate was incubated at 37°C in 5% CO₂ for 5 days. On fifth day 15 μl Alamar blue solution was added to the each well of plate. The plate was again incubated overnight at 37 °C in 5% CO₂. The fluorescence was read on a BMG Polar Star with an excitation frequency of 544 nm and emission frequency of 590 nm. The compounds that were found active to be (>90% inhibition compared with control) at this concentration were then tested at six serial dilutions from 50 to 1.56 $\mu\text{g mL}^{-1}$.

Activity against *M. tuberculosis* H₃₇R_v strain

Drug susceptibility and determination of MIC of the test compounds/drugs against *M. tuberculosis* H₃₇R_v was performed by the agar microdilution method where twofold dilutions of each test compound added to 7H10 agar supplemented with oleic acid-albumin-dextrose catalase and organism. A culture of *M. tuberculosis* H₃₇R_v growing on Lowenstein Jensen medium was harvested in 0.85% saline with 0.05% Tween-80. A 1 $\mu\text{g mL}^{-1}$ concentration suspension of the extracts/compounds was prepared in DMSO. This suspension was added to (in tubes) 7H10 middle Brook's medium (containing 1.7 ml medium and 0.2 ml OADC supplement) at different concentrations of compound, keeping the volume constant, i.e., 0.1 mL. The medium was allowed to cool with the tubes in a slanting position. These tubes were then incubated at 37°C for 24 h followed by streaking of *M. tuberculosis* H₃₇R_v (5×10^4 bacilli per tube). These tubes were then incubated at 37°C. Growth of bacilli was observed after 30 days of incubation. Tubes containing the compounds were compared with control tubes in which medium alone was incubated with H₃₇R_v. The concentration at which complete inhibition of colonies occurred was taken as the active concentration of test compound.

Acknowledgements The authors would like to thank the Director of the CDRI for his keen interest in this program. ND would like to thank CSIR New Delhi for the award of a SRF; NS and MM are thankful

for financial assistance from DRDO (ERIP/ER/050212/M/01/857) and ICMR, respectively. We would also like to thank the task force members involved in the tuberculosis screening program at our institute.

References

- Bram S, Froussard P, Guichard M, Jasmin C, Augery Y, Sinoussi-Barre F, Wray W (1980) Vitamin C preferential toxicity for malignant melanoma cells. *Nature* 284:629–631
- Cinatl J, Weber B, Rabenau H, Gumbel H.O, Chenot J.F, Scholz M, Encke A, Doerr HW (1995) In vitro inhibition of human cytomegalovirus replication in human foreskin fibroblasts and endothelial cells by ascorbic acid 2-phosphate. *Antiviral Res* 27:405–418
- Collins LA, Franzblan SG (1997) Microplate alamar blue assay versus BACTEC 460 system for high-throughput screening of compounds against *Mycobacterium tuberculosis* and *Mycobacterium avium*. *Antimicrob Agents Chemother* 41:1004
- Elbe E, Large CM, De Vries WH, Crum GF, Shell JW (1956). In: Streptolydigin, a new antimicrobial antibiotic. II. Isolation and characterization. *Antibiotics Annual 1955–1956*, Medical Encyclopedia. New York, 893
- Grdisa M, Kralj M, Eckert-Maksic M, Maksic ZB, Pavelic K (1995) 6-amino-6-deoxyascorbic acid induces apoptosis in human tumor cells. *J Cancer Res Clin Oncol* 12:98–102
- Holtzel A, Ganzle MG, Nicholson GJ, Hammes WP, Jung G (2000) Characterization of Reutericyclin Produced by *Lactobacillus reuteri* LTH2584. *Angew Chem Int Ed* 39:2766–2768
- Iwata Y, Maekawara N, Tanino K, Miyashita M (2005) Tetramic acid antibiotics: stereoselective synthesis of streptolic acid and tirandalydigin. *Angew Chem Int Ed* 44: 1532–1536
- Khan MA, Adams H (1999) The formation of lactams from L-ascorbic acid. *Carbohydr Res* 322:279–283
- Kremer L, Douglas J.D, Baulard A.R, Morehouse C, Guy M.R, Alland D, Dover LG, Lakey JH, Jacobs WR, Brennan PJ, Minnikin DE, Besra GS (2000) Thiolactomycin and related analogues as novel anti-mycobacterial agents targeting KasA and KasB condensing enzymes in *Mycobacterium tuberculosis*. *J Biol Chem* 275:16857–16864
- Larvenov SN, Turchin KF, Korolev AM, Anisimova OS, Preobrzenskaya MN (2005) New pyrano[3,4-b]indoles from 2-hydroxymethylindole and L-dehydroascorbic acid. *Tetrahedron* 61:6610–6613
- Ley SV, Smith SC, Woodward PR (1992) Further reactions of t-butyl 3- oxobutanthioate and t-butyl 4-diethyl-phosphono-3-oxobutanthioate: carbonyl coupling reactions, amination, use in the preparation of 3-acyltetramic acids and application to the total synthesis of fulgorubin A. *Tetrahedron* 48:1145–1174
- Malic SR, Svedruzic D, Gazivoda T, Marunovic A, Brundic AR, Nagl A, Balzarini J, Clercq ED, Mintas M (2000) Synthesis and antitumor activities of novel pyrimidine derivatives of 2,3-O,O-dibenzyl-6-deoxy-L-ascorbic acid and 4,5-didehydro-5,6- dideoxy-L-ascorbic acid. *J Med Chem* 43:4806–4811
- Mishra RC, Dwivedi N, Bansal I, Saxena JK, Tripathi RP (2005) Synthesis and DNA topoisomerase-II inhibitory activity of unnatural nucleosides. *Nucleosides, Nucleotides Nucl Acids*, 24:15–35 and references cited therein
- Miyakawa SK, Suzuki Y, Harada H, Okazaki (1982) Thiolactomycin, a new antibiotic. IV. Biological properties and chemotherapeutic activity in mice. *J Antibiot* 35:411–419
- Preobrzenskaya MN, Korolev AM (2000) Indole derivatives in vegetables of the Cruciferae family. *Russ J Bioorg Chem* 26:85–97
- Olabisi A.O, Wimalasena K (2004) Rational approach to selective and direct 2-O-alkylation of 5,6-O-isopropylidene-L-ascorbic acid. *J Org Chem* 69:7026–7032
- Roggo BE, Petersen F, Delmendo R, Jenny HB, Peter HH, Rossel JJ (1994) 3- Alkanoyl-5-hydroxymethyl tetronic acid homologues and resistomycin: new inhibitors of HIV-1 protease. I. Fermentation, isolation and biological activity. *Antibiot* 47:136–142
- Royles BJL (1995) Naturally occurring tetramic acids: structure, isolation, and synthesis. *Chem Rev* 95:1981–2001
- Singh BK, Verma SS, Dwivedi N, Tripathi RP (2006) L-ascorbic acid in organic synthesis: DBU-catalysed one-pot synthesis of tetramic acid derivatives from 5,6-O-isopropylidene ascorbic acid. *Tetrahedron Lett* 47:2219–22
- Saito H, Tomioka H, Sato K, Emori M, Yamane T, Yamashita K (1991) In vitro antimycobacterial activities of newly synthesized benzoxazinorifamycins. *Antimicrob Agents Chemother* 35:542

- Somu RV, Wilson DJ, Bennett EM, Boshoff HI, Celia L, Beck BJ, Barry CE, Aldrich CC (2006) Antitubercular nucleosides that inhibit siderophore biosynthesis: SAR of the glycosyl domain. *J Med Chem* 49, 7623–35 and references cited therein
- Wang C-Y, Wang B-G, Wiryowidagdo S, Wray V, Van Soest R, Steube KG, Guan H.-S, Proksch P, Ebel RJ (2003) Melophlins C-O, thirteen novel tetramic acids from the marine sponge *Melophlus sarassinorum*. *Nat Prod* 66:51–56