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Synthesis And Biological Activities of Three Sulfated Sialyl Le^x Ganglioside Analogues for Clarifying the Real Carbohydrate Ligand Structure of L-Selectin[†]

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Abstract—Sulfated sialyl Le^x ganglioside analogues at C-6 of p-galactose, N-acetyl-p-glucosamine, and of both p-galactose and N-acetyl-p-glucosamine residues have been synthesized, in order to clarify the structure of the real carbohydrate ligand of L-selectin. Coupling of the suitably protected N-acetyl-p-glucosaminyl-β(1→3)-lactose derivatives 13 and 16 with the sialyl α(2→3)-p-galactopyranosyl trichloroacetimidates 10 and 12 (glycosyl donors), via glycosylation of 2-(trimethylsilyl)ethyl 4,6-O-benzylidene-β-p-galactopyranoside (1) with the phenyl 2-thioglycoside derivative (2) of N-acetylneuraminic acid (Neu5Ac) using N-iodosuccinimide/TfOH, O-benzoylation, removal of the benzylidene group affording 5, selective 6-O-levulinoylation, O-benzoylation, removal of the 2-(trimethylsilyl)ethyl group, and imidate formation, or via O-acetylation of 5, removal of the 2-(trimethylsilyl)ethyl group, then imidate formation, gave the pentasaccharides 18–20. The glycosylation of the pentasaccharide acceptors (21–23) derived from 18–20 by removal of the 4-methoxybenzyl group, with phenyl 1-thioglycoside derivative 27 of L-fucose using dimethyl(methylthio)sulfonium triflate (DMTST) afforded the corresponding hexasaccharides 28–30, which were transformed in good yields, via reductive removal of their benzyl groups, O-acetylation, selective removal of the 2-(trimethylsilyl)ethyl group, imidate formation, coupling with (2S,3R,4E)-2-azido-O-benzoyl-4-octadecene-1,3-diol (35) in the presence of boron trifluoride etherate, selective reduction of the azido group, coupling with octadecanoic acid, selective removal of the levulinoyl groups, treatment with sulfur trioxide-pyridine complex, then removal of the protecting groups, into the desired sulfated sialyl Le^x ganglioside analogues 50–52. Copyright © 1996 Elsevier Science Ltd

Introduction

Selectins (E-, P-, and L-)²⁻⁸ are a class of recently discovered cell adhesion molecules which act to localize spatially and temporally the binding of specific leukocytes to certain vascular endothelial cells. All selectins possess a similar arrangement of their protein parts which are calcium-dependent lectin domains, an epidermal growth factor domain and a discrete number of sequence modules similar to those found in complement-binding proteins. The lectin domains of selectins mediate cell adhesion through recognition and binding of cell-specific oligosaccharide ligands on a variety of glycoconjugates such as glycoproteins, gangliosides and glycosaminoglycans. Although each selectin may have its own optimum carbohydrate ligand, there is now general agreement that all three selectins can efficiently recognize sialyl Lewis × (sLex), sialyl Lewis^a (sLe^a), and structurally related Lewis blood group oligosaccharides. It has been known 10-19 that L- and P-selectins can efficiently bind to sulfated carbohydrates such as fucoidan, sulfatides, sulfated glucuronic acid (HNK-1) epitope, heparin, and sulfo-Lex-like structures. In order to clarify (the structure of) the real carbohydrate ligands for each selectin, we have synthesized20-26 a variety of sialylated, sulfated and phosphated Lex and Lea oligosacharides and their analogues. L-selectin has been discovered as a lympho-

In view of these facts, and as a part of our continuing efforts on the chemical synthesis, biological functions, structural determination of carbohydrate ligands of cell-adhesion molecules, and for the development of the selectin blocker, we describe herein the synthesis of the sulfated sLe^x ganglioside analogues at C-6 of the Gal or GlcNAc, and of both Gal and GlcNAc residues, for clarifying the structure of the real carbohydrate ligand of L-selectin, and their binding activities not only to L-selectin but also to E- and P-selectins.

cyte homing receptor involved in the binding of lymphocytes to the high endothelial venules of limph nodes. One of the endothelial-derived ligands for L-selectin is GlyCAM-1, a mucin-like glycoprotein with sulfated, sialylated, and fucosylated oligosaccharides. Very recently, Rosen and co-workers^{27–30} have isolated the presence of sulfated Gal and GlcNAc residues at C-6, and Gal $\beta(1\rightarrow 4)$ 6-O-sulfo-GlcNAc and 6-O-sulfo-Gal $\beta(1\rightarrow 4)$ GlcNAc by a mild hydrolysis of GlyCAM-1, and suggested that sulfated sLex structure may comprise a recognition determinant on GlyCAM-1. On the other hand, Jacob and co-workers^{31a} have enzymatically synthesized 6-O-sulfo-Galß $(1\rightarrow 4)$ GlcNAc residue containing sLe^x-pentasaccharide and compared its inhibition ability with L-selectin binding with an sLex tetrasaccharide, and it displayed a stronger activity. Very recently, Matta and coworkers^{31b,c} have chemically synthesized 6-O-sulfo-Galβ(1→4)GlcNAc residue containing sLe^a and sLe^x oligosaccharides.

^{*}Synthetic Studies on Sialoglycoconjugates. Part 87. For Part 86, see ref. I.

Results and Discussion

For the synthesis of the three desired, sulfated sLe^x gangliosides **50–52**, the core oligosaccharides **21–23** were selected as the glycosyl acceptors. Compounds **21–23** each have a free hydroxy group at C-3 of the GlcNAc residue for α -fucosylation, and also levulinoyl groups at the desired positions and provide selectively the free hydroxy group at C-6 of the Gal (d) or GlcNAc, and of both Gal and GlcNAc residues for further sulfation.

The glycosyl acceptors **21–23** were prepared as follows: glycosylation³² of 2-(trimethylsilyl)ethyl 4,6-O-benzylidene- β -D-galactopyranoside (1)³³ with the phenyl 2-thioglycoside derivative (2)³⁴ (1.5 equiv., relative to the acceptor) of N-acetylneuraminic acid (NeuSAc) in acetonitrile for 3 h at $-30\,^{\circ}$ C in the presence of N-iodosuccinimide (NIS;^{32c,35} 2.3 equiv., relative to the donor) trifluoromethanesulfonic acid (TfOH), gave the expected α -glycoside 3 in 65% yield. The observed chemical shift and coupling constants [δ 2.75, $J_{gem} = 12.6$, $J_{3(eq),4} = 4.4$ Hz, 4.88 (m) and 5.35 ($J_{6,7} = 1.2$, $J_{7,8} = 8.0$ Hz] for H-3eq, H-4, and H-7 in the NeuSAc moiety are characteristic for α -glycosidic linkages.³⁶

Treatment of 3 with benzoic anhydride in the presence of 4-dimethylaminopyridine afforded the 2-benzoate 4 in 86% yield, which was converted via reductive removal (5, 94%) of the benzylidene group (10% Pd-C), selective 6-O-levulinovlation (6, 85%) with levulinic anhydride in pyridine in the presence of 4-dimethylaminopyridine at -50 °C, and 4-O-benzoylation into 2-(trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)-(2→3)-2,4-di-O-benzoyl-6-Olevulinoyl-β-D-galactopyranoside (7, 82%). Acetylation of 5 gave the 4,6-di-O-acetate 8 (91%), which was used for the synthesis of a sulfated sLex ganglioside at C-6 of the GlcNAc residue. Compounds 7 and 8 were transformed into the corresponding trichloroacetimidates 10 and 12 in good yields by selective removal^{20a,33} of the 2-(trimethylsilyl)ethyl group with trifluoroacetic acid and subsequent imidate formation.³⁷

The glycosylation of 13³⁸ or 16 derived from 14³⁸ by reductive removal (15, 82%) of the benzylidene group followed by selective levulinovlation in 81% yield, with 10 in dichloromethane in the presence of TMS-triflate for 12 h at 7 °C, afforded the corresponding pentasaccharides 18 (50%) and 20 (45%), respectively. In essentially the same way, coupling of 12 and 16 gave pentasaccharide 19 in 77% yield. The β-configurations of 18, 19, and 20 were assigned from their ¹H NMR data that showed the signals at δ 5.46 (t, 1H, $J_{1,2} = J_{2,3} = 8.1$ Hz, 18), 5.36 (t, 1H, $J_{1,2} = J_{2,3} = 8.4$ Hz, 19), and 5.46 (dd, 1H, $J_{1,2} = 9.9$, $J_{2,3} = 7.7$ Hz, **20**) for H-2d of the obtained pentasaccharides. Removal of the 4-methoxybenzyl group in 18-20 in acetonitrile-water in the presence of ceric ammonium nitrate (CAN)^{25d,39} for 1-3 h at room temperature afforded the desired glycosyl acceptors 21 (98%), 22 (90%), and 23 (68%), respectively. The glycosylation of 21, 22, or 23 was effected with phenyl 2,3,4-tri-O-benzyl-1-thio-β-L-fucopyranoside 27 which was newly prepared from tetra-O-acetyl-β-L-fucopyranose via replacement of the anomeric acetoxy group with a phenylthio group using thiophenol in the presence of boron trifluoride etherate, O-deacetylation with sodium methoxide in methanol, and subsequent O-benzylation with benzyl bromide in the presence of sodium hydride in N,N-dimethylformamide (DMF). Reaction in the presence of dimethyl(methylthio)sulfonium triflate (DMTST)^{32a,40} and molecular sieves 4 Å (MS-4 Å) in benzene for 48–72 h at 7 °C gave the desired hexasaccharides 28 (58%), 29 (90%), and 30 (50%), respectively, showing in their 'H NMR spectra, signals at δ 5.15 (d, $J_{1,2} = 3.6$ Hz, H-1f of **28**), 5.03 (d, $J_{1,2} = 3.3$ Hz, H-1f of **29**) and 5.17 (d, $J_{12}=3.5$ Hz, H-1f of **30**), characteristic of the α -fucopyranosyl unit.

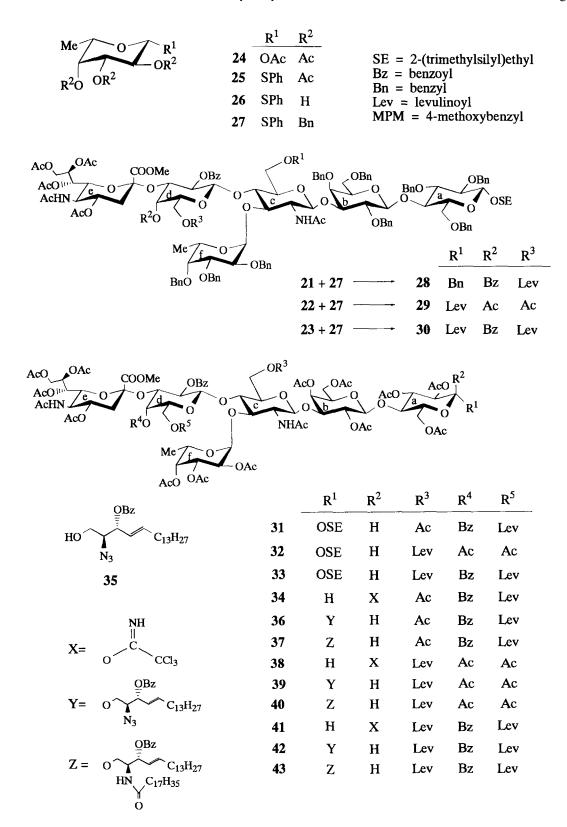
Removal of the benzyl groups from 28, 29, and 30 by catalytic hydrogenolysis over 10% Pd-C in ethanol:acetic acid (5:1) for 24–40 h at 40 °C, and subsequent acetylation gave the per-O-acylated hexasaccharide 31 (89%), 32 (94%), and 33 (82%), respectively. Selective removal of the 2-(trimethylsilyl)ethyl group from 31, 32, and 33 as described in the preparation of 9 gave the corresponding 1-hydroxy compounds which, on treatment³⁷ with trichloroacetonitrile in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) for 2 h at 0 °C, gave the corresponding α -trichloroacetimidates 34 (91%), 38 (98%), and 41 (89%), respectively.

The 'H NMR spectra of these compounds showed signals at δ 6.47 (d, 1H, $J_{1,2}=3.5-3.7$ Hz, H-1a) and 8.65-8.67 (s, 1H, C=NH), indicating the trichloroacetimidates to be α . The final glycosylation^{41,42a} of (2S,3R,4E)-2-azido-3-O-benzoyl-4-octadecene-1,3-diol (35)⁴² with 34, 38, or 41 thus obtained, in dichloromethane for 5 h at 0 °C in the presence of boron trifluoride etherate and MS-4 Å, gave only the expected β-glycosides 36 (63%), 39 (58%), and 42 (48%), respectively. The ¹H NMR spectra of 36, 39, and 42 showed a one-proton doublet at δ 4.43 ~ 4.49 $(J_{1,2}=7.0\sim7.7 \text{ Hz}, \text{ H-1a})$, showing the newly formed glycosidic linkages to be β. Selective reduction^{37b,43} of the azido group in 36, 39, or 42 with hydrogen sulfide in aq. 83% pyridine for 72 h at 0 °C, and subsequent condensation with octadecanoic acid 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (water-soluble carbodiimide, WSC) dichloromethane, furnished the corresponding ganglioside derivatives **37** (54%), **40** (60%), and **43** (60%). Selective removal^{25a,44} of the levulinoyl group from 37, 40, or 43 was carried out in ethanol with hydrazine monoacetate at room temperature to give 44 (70%), 46(50%), and 48 (49%), respectively. Treatment^{25a} of compounds 44, 46, or 48 with sulfur trioxide-pyridine complex in DMF for 2~4 h at room temperature afforded the corresponding sulfates 45 (98%), 47 (91%), and **49** (91%) as their pyridine salts. O-Deacylation of 45, 47, or 49 with sodium methoxide in methanol, with subsequent saponification of the sialic acid methyl ester group, yielded the desired sulfated sLe^x gangliosides 50-52 in almost quantitative yields as their sodium salts. FABMS spectra (negative ion mode) showed the base peaks at m/z (M-Na)⁻ 1794 and (M-2Na)²⁻ 1771 for **50** and **51**, and m/z (M-Na)⁻ 1896 for **52**.

This work reports the first stereocontrolled synthesis of the sulfated sLe^x ganglioside analogues which will be candidate structures for the L-selectin ligand, using the selectively levulinoylated pentasaccharide acceptors 21,

22, and **23**. The binding activities to E-, P-, and L-selectins of these sulfated sLe^x ganglioside analogues were examined⁴⁵ by H. Kondo and co-workers of Kanebo Ltd, Japan (unpublished report). Interestingly, the introduction of a sulfo group on the 6-OH of Gal, or on both the Gal and GlcNAc residues completely

abolished the binding to E-selectin. Compound 51 containing a sulfo group on the 6-OH of the GlcNAc residue bound to E-selectin almost as well as the sLe^x ganglioside. These results show that the introduction of the sulfo group (anionic substituent) on the 6-OH of the Gal residue abolishes E-selectin recognition. On



the contrary, for P-selectin, binding to **50–52** appeared to be equivalent to sLe^x ganglioside, indicating the introduction of sulfo group at the 6-OH positions did not affect for the recognition. Very interestingly, for L-selectin, binding to **52** was different from that of **50**, **51**, and the sLe^x ganglioside (hexasaccharide). Binding to **50** or **51** appeared to be equivalent to that of the sLe^x ganglioside, while binding to disulfated sLe^x **52** was effectively distinct from those of **50**, **51**, and sLe^x ganglioside, suggesting strongly that the real carbohydrate ligand structure of L-selectin is a disufated molecule. Details of this biological investigation will be appearing in the near future.

Experimental

General methods

Optical rotations were determined with a Union PM-201 polarimeter at 25 °C, and IR spectra were recorded with a Jasco IRA-100 spectrophotometer. ¹H NMR spectra were recorded at 270 MHz with a Jeol JNM-GX 270 spectrometer. FAB-MS spectra were

determined with a Jeol JMS-SX 102A mass spectrometer/JMA-DA 7000 data system. Each sample was mixed with diethylamine matrix on a target. The ion accelerating voltage was 8.0 KV, and the primary beam for the bombardment was 6.0 KeV of xenon. Preparative chromatography was performed on silica gel (Wako Chemical Co., 200 mesh) with the solvent systems specified. Concentrations were conducted in vacuo.

2-(Trimethylsilyl)ethyl *O*-(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)-(2→3)-4,6-*O*-benzylidene-β-D-galactopyranoside (3). To a solution of phenyl (methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-2-thio-D-glycero-D-galacto-2-nonulopyranosid)onate (2, 8.79 g, 15.1 mmol) and 2-(trimethylsilyl)ethyl 4,6-*O*-benzylidene-β-D-galactopyranoside (1, 3.61 g, 9.8 mmol) in dry acetonitrile (15 mL) was added molecular sieves 3 Å (MS-3 Å, 12.5 g), and the mixture was stirred for 5 h at room temperature, then cooled to −30 °C. To the cooled suspension were added, with stirring, NIS (7.7 g, 34.2 mmol) and TfOH (360 μL, 4.1 mmol), and the stirring was continued for 3 h at

-30 °C. The solids were filtered off and washed with CH₂Cl₂, and the filtrate and washings were combined and washed with $M Na_2CO_3$, $M Na_2S_2O_3$ and water, dried (Na₂SO₄) and concentrated. Column chromatography (toluene: MeOH 5:1) of the residue on silica gel afforded the α -glycoside 3 (5.26 g, 65%) as an amorphous mass; $[\alpha]_D + 4.2^{\circ}$ (c 0.8, CHCl₃); IR (KBr): 3500–3200, 1750, 1680, 1550, 1220, 860, 840, 700 cm⁻¹ ¹H NMR (270 MHz, CDCl₃): δ 1.08 (m, 2H, $Me_3SiCH_2CH_2$), 1.89 (s, 3H, AcN), 2.02, 2.05, 2.16, 2.20 (4s, 12H, 4AcO), 2.75 [dd, 1H, $J_{gem} = 12.6$, $J_{3b(eq).4} = 4.4$ Hz, H-3b(eq)], 3.59 (s, 3H, MeO), 4.47 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1a), 4.88 (m, 1H, H-4b), 5.25 (m, 1H, $J_{6,7}$ =2.1, $J_{7.8} = 8.0 \text{ Hz}, \text{ H-7b}$, 5.38 (s, 1H, PhC<u>H</u>), 5.43 (m, 1H, H-8b), 7.28-7.64 (m, 5H, Ph). Anal.: calcd for C₃₇H₅₃NO₁₈Si: C, 53.68; H, 6.45; N, 1.69; found: C, 53.43; H, 6.45; N, 1.65%.

2-(Trimethylsilvl)ethyl *O*-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-p-glycero-α-p-galacto-2nonulopyranosylonate)- $(2\rightarrow 3)$ -2-O-benzyl-4,6-O-benzylidene-\(\beta\)-p-galactopyranoside (4). To a solution of 3 (510 mg, 0.62 mmol) in pyridine (5 mL) were added benzoic anhydride (270 mg) and 4-dimethylaminopyridine (22 mg), and the mixture was stirred overnight at room temperature, concentrated and extracted with CH₂Cl₂. The extract was washed with 2 M HCl and water, dried (Na₂SO₄) and concentrated. Column chromatography (ethyl acetate:hexane 4:1) of the residue on silica gel gave 4 (500 mg, 86%) as an amorphous mass; $[\alpha]_D + 18.8 \degree$ (c 2.1, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.97 (m, 2H, Me₃SiCH₂CH₂), 1.82 [t, 1H, $J_{gem} = J_{3b(ax),4} = 12.6$ Hz, H-3b(ax)], 1.92, 1.95, 2.03, 2.16, 2.31 (5s, 15H, 4AcO, AcN), 2.69 [dd, 1H, $J_{3b(eq),4} = 4.4 \text{ Hz}, \text{ H-3b}(eq)$], 3.64 (s, 3H, MeO), 4.46 (dd, 1H, $J_{gen} = 12.1$, $J_{8.9'} = 2.4$ Hz, H-9'), 4.64 (dd, 1H, $J_{2,3} = 10.1$, $J_{3,4} = 3.7$ Hz, H-3a), 4.82 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1a), 5.35 (d, 1H, $J_{NH.5} = 9.3$ Hz, NH), 5.50 (s, 1H, PhCH), 5.55 (dd, 1H, $J_{2,3}$ = 10.1 Hz, H-2), 5.64 (m, 1H, H-8), 7.38-7.67 (m, 10H, 2Ph). Anal.: calcd for $C_{45}H_{59}NO_{19}Si: C, 57.13; H, 6.29; N, 1.48; found: C,$ 56.90; H, 6.00; N, 1.43%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2nonulopyranosylonate)-(2→3)-2-O-benzoyl-β-D-galactopyranoside (5). A solution of 4 (830 mg, 0.88 mmol) in AcOH (10 mL) was hydrogenolysed in the presence of 10% Pd-C (1 g) for 7 h at room temperature then filtered, and the solids were washed with CH₂Cl₂. The filtrate and washings were combined and concentrated. Column chromatography (toluene: MeOH 50:1) of the residue on silica gel gave 5 (710 mg, 94%) as an amorphous mass; $[\alpha]_D = 1.4^\circ$ (c 1.3, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.83 (m, 2H, Me₃SiCH₂CH₂), 1.65 (s, 3H, AcN), 1.82, 1.96, 2.06, 2.16 (4s, 12H, 4AcO), 2.51 [dd, 1H, $J_{gem} = 12.8$, $J_{3b(eq).4} = 4.6$ Hz, H-3b(eq)], 3.79 (s, 3H, MeO), 4.32 (dd, 1H, $J_{gem} = 12.5$, $J_{8,9'} = 2.4 \text{ Hz}, \text{ H-9'}$, 4.40 (dd, 1H, $J_{2,3} = 9.9, J_{3,4} = 3.3 \text{ Hz}$, H-3a), 4.67 (d, 1H, $J_{1,2}$ =7.9 Hz, H-1a), 4.79 (m, 1H, H-4b), 5.20 (dd, 1H, $J_{6,7}=1.7$, $J_{7,8}=9.0$ Hz, H-7), 5.31 (dd, 1H, H-2a), 5.36 (d, 1H, $J_{NH.5} = 8.4$ Hz, NH), 5.50 (m, 1H, H-8), 7.29–8.14 (m, 5H, Ph). Anal.: calcd for $C_{38}H_{55}NO_{19}Si$: C, 53.20; H, 6.46; N, 1.63; found: C, 53.18; H, 6.19; N, 1.39%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2nonulopyranosylonate)- $(2\rightarrow 3)$ -2-O-benzoyl-6-O-levulinoyl-β-D-galactopyranoside (6). To a solution of 5 (360 mg, 0.42 mmol) in pyridine (4.5 mL) and CH₂Cl₂ (1.5 mL), cooled to -50 °C, was added dropwise a solution of levulinic anhydride (110 mg) in CH₂Cl₂ (3 mL), and the mixture was stirred for 18 min at -50 °C. After completion of the reaction, MeOH (2 mL) was added to the mixture and concentrated then extracted with CH₂Cl₂. The extract was successively washed with 2 м HCl and water, dried (Na₂SO₄) and concentrated. Column chromatography (CH₂Cl₂:MeOH 25:1) of the residue on silica gel gave 6 (340 mg, 85%) as an amorphous mass; $[\alpha]_D + 6.6^\circ$ (c 0.9, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.85 (m, 2H, Me₃SiCH₂CH₂), 1.60 (s, 3H, AcN), 1.81, 1.97, 2.07, 2.17 (4s, 12H, 4AcO), 2.20 (s, 3H, MeCOCH₂CH₂), 2.66–2.85 (m, 4H, $MeCOCH_2CH_2$), 3.80 (s, 3H, MeO), 4.28 (dd, 1H, $J_{gem} = 12.5, J_{8.9} = 2.4 \text{ Hz}, \text{ H-9'}, 4.35 \text{ (m, 2H, H-6a)}, 4.42$ (dd, 1H, $J_{2,3}=9.9$, $J_{3,4}=3.1$ Hz, H-3a), 4.65 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1a), 4.78 (m, 1H, H-4b), 5.28 (dd, 1H, H-2a), 5.52 (m, 1H, H-8), 7.28-8.20 (m, 5H, Ph). Anal.: calcd for $C_{43}H_{61}NO_{21}Si$: C, 54.02; H, 6.43; N, 1.47; found: C, 53.79; H, 6.24; N, 1.32%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2nonulopyranosylonate) - $(2\rightarrow 3)$ - 2,4-di-O-benzoyl-6-Olevulinoyl-β-p-galactopyranoside (7). To a solution of **6** (580 mg, 0.61 mmol) in pyridine (5 mL) and 4-dimethylaminopyridine (22 mg) was added benzoic anhydride (270 mg, 1.2 mmol) and the mixture was stirred for 12 h at room temperature, MeOH (0.5 mL) was added to the mixture, and this was concentrated then extracted with CH₂Cl₂. The extract was successively washed with 2 M HCl and water, dried (Na₂SO₄) and concentrated. Column chromatography (toluene-:MeOH 50:1) of the residue on silica gel gave 7 (530 mg, 82%) as an amorphous mass; $[\alpha]_D + 40.5^\circ$ (c 1.6, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.87 (m, 2H, Me₃SiCH₂CH₂), 1.46 (s, 3H, AcN), 1.67 [t, 1H, $J_{gem} = J_{3b(ax),4} = 12.5$ Hz, H-3b(ax)], 1.78, 1.92, 2.09, 2.18 (4s, 12H, 4AcO), 2.24 (s, 3H, MeCOCH₂CH₂), 2.45 [dd, 1H, $J_{3b(eq),4} = 4.6$ Hz, H-3b(eq)], 2.51–2.79 (m, 4H, MeCOCH₂CH₂), 3.89 (s, 3H, MeO), 4.35 (dd, 1H, $J_{gen} = 12.5$, $J_{8.9'} = 2.4$ Hz, H-9'), 4.80 (dd, 1H, $J_{2.3} = 10.1$, $J_{3.4} = 3.3$ Hz, H-3a), 4.81 (m, 1H, H-4b), 4.83 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1a), 5.23 (d, 1H, H-4a), 5.39 (dd, 1H, H-2a), 5.63 (m, 1H, H-8), 7.28-8.21 (m, 10H, 2Ph). Anal.: calcd for C₅₀H₆₅NO₂₂Si: C, 56.65; H, 6.18; N, 1.32; found: C, 56.63; H, 6.08; N, 1.03%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate) - (2 \rightarrow 3) - 4,6 - di - O- acetyl - 2 - O-benzoyl- β -D-galactopyranoside (8). To a solution of 5 (930 mg, 1.1 mmol) in pyridine (10 mL) was added

acetic anhydride (4 mL) and the mixture was stirred for 3 h at room temperature and concentrated. Column chromatography (toluene: MeOH 50:1) of the residue on silica gel gave **8** (930 mg, 91%) as an amorphous mass; [α]_D +23.0° (c 1.1, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.87 (m, 2H, Me₃SiCH₂CH₂), 1.43 (s, 3H, AcN), 1.79, 1.97, 2.06, 2.08, 2.14, 2.22 (6s, 18H, 6AcO), 2.55 [dd, 1H, J_{gem} = 12.5, $J_{3b(eq),4}$ = 4.4 Hz, H-3b(eq)], 3.86 (s, 3H, MeO), 4.33 (dd, 1H, J_{gem} = 12.5, $J_{8,9'}$ = 2.2 Hz, H-9'), 4.69 (dd, 1H, $J_{2,3}$ = 10.1, $J_{3,4}$ = 3.2 Hz, H-3a), 4.76 (d, 1H, $J_{1,2}$ = 7.9 Hz, H-1a), 4.85 (m, 1H, H-4b), 4.99 (d, 1H, H-4a), 5.60 (m, 1H, H-8), 7.29–8.17 (m, 5H, Ph). Anal.: calcd for C₄₂H₅₉NO₂₁Si: C, 53.55; H, 6.31; N, 1.49; found: C, 53.46; H, 6.23; N, 1.38%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-p-glyceroα-p-galacto-2-nonulopyranosylonate)-($2\rightarrow 3$)-2,4-di-*O*-benzoyl-6-*O*-levulinoyl-p-galactopyranose (9). To a solution of 7 (1.0 g, 0.94 mmol) in CH₂Cl₂ (20 mL), cooled to 0 °C, was added CF₃CO₂H (10 mL) and the mixture was stirred for 30 min at room temperature and concentrated. Column chromatography (ethyl acetate:hexane 4:1) of the residue on silica gel gave 9 (810 mg, 90%) as an amorphous mass; IR (KBr): 3600–3300, 1750, 1670, 1550, 1225, 720 cm $^{-1}$. Anal.: calcd for C₄₅H₅₃NO₂₂: C, 56.31; H, 5.57; N, 1.46; found: C, 56.24; H, 5.53; N, 1.33%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -2.4-di-O-benzovl-6-O-levulinovl-p-galactopyranosyl trichloroacetimidate (10). To a solution of 9 (400 mg, 0.42 mmol) in CH₂Cl₂ (4 mL), cooled to 0 °C, were added trichloroacetonitrile (1.26 mL) and DBU (62 μL, 0.42 mmol), and the mixture was stirred for 1 h at 0 °C then concentrated. Column chromatography (ethyl acetate: hexane 4:1) of the residue on silica gel gave 10 (450 mg, 98%) as an amorphous mass; IR (KBr): $3450-3\overline{2}00$, 1740, 1680, 1540, 1220, 710 cm⁻¹. The anomeric ratio (α : β) was estimated as \sim 5:6 from the ratio of the intensities of the anomeric proton signals of galactose residue in the H NMR spectrum. Anal.: calcd for $C_{47}H_{53}N_2O_{22}Cl_3$: C, 51.12; H, 4.84; N, 2.54; found: C, 50.82; H, 4.69; N, 2.48%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxyp-glycero-α-p-galacto-2-nonulopyranosylonate)-($2\rightarrow 3$)-4,6-di-*O*-acetyl-2-*O*-benzoyl-p-galactopyranose (11). To a solution of **8** (880 mg, 0.93 mmol) in CH₂Cl₂ (10 mL), cooled to 0 °C, was added CF₃CO₂H (9 mL) and the mixture was stirred for 1 h at room temperature, then work up as described for **9** gave **11** (780 mg, 98%) as an amorphous mass; IR (KBr): 3600-3300, 1750, 1680, 1540, 1220, 710 cm⁻¹. Anal.: calcd for C₃₇H₄₇NO₂₁: C, 52.79; H, 5.63; N, 1.66; found: C, 52.61; H, 5.53; N, 1.37%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-2-nonulopyranosylonate)- $(2 \rightarrow 3)$ -4,6-di-O-acetyl-2-O-benzoyl-O-galactopyranosyl trichloroacetimidate (12). To a solution of 11 (250 mg, 0.3 mmol) in CH₂Cl₂ (3 mL), cooled to 0 °C, were added trichloroacetonitrile

(0.89 mL) and DBU (44 μ L, 0.3 mmol) and the mixture was stirred for 1.5 h at 0 °C, then work up as described for 10 gave 12 (233 mg, 80%) as an amorphous mass. The anomeric ratio (α : β) was estimated as \sim 3:4 from the ratio of the intensities of the anomeric proton signals of galactose residue in the 'H NMR spectrum. Anal.: calcd for $C_{39}H_{47}N_2O_{21}Cl_3$: C, 47.50; H, 4.80; N, 2.84; found: C, 47.79; H, 4.74; N, 2.82%.

2-(Trimethylsilyl)ethyl *O*-(2-acetamido-2-deoxy-3-*O*-4-methoxybenzyl-β-p-glucopyranosyl)-(1 \rightarrow 3)-*O*-(2,4,6-tri-*O*-benzyl-β-p-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl-β-p-glucopyranoside (15). A solution of 14 (100 mg, 72 μmol) in AcOH (8 mL) and water (2 mL) was heated for 36 h at 40 °C and concentrated. Column chromatography (ethyl acetate:hexane 3:2) of the residue on silica gel gave 15 (77.6 mg, 82%) as an amorphous mass; [α]_D –19.1° (*c* 1.6, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 1.49 (s, 3H, AcN), 3.75 (s, 3H, MeO), 6.80–7.38 (m, 34H, 6Ph, MeOPh). Anal.: calcd for C₇₅H₉₁NO₁₇Si: C, 68.94; H, 7.02; N, 1.07; found: C, 68.77; H, 7.01; N, 0.87%.

2-(Trimethylsilyl)ethyl O-(2-acetamido-2-deoxy-6-Olevulinoyl-3-O-methoxybenzyl-β-D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-benzyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -2,3,6-tri-O-benzyl- β -D-glucopyranoside To a solution of 15 (480 mg, 0.37 mmol) and 4-dimethylaminopyridine (45 mg, 0.37 mmol) pyridine (40 mL) and CH₂Cl₂ (10 mL), cooled to -50 °C, was dropwise added, with stirring, a solution of levulinic anhydride (90 mg) in CH₂Cl₂ (34 mL). The mixture was stirred for 2 h at -50 °C then MeOH (2 mL) was added to the mixture. The solution was stirred for 30 min and concentrated then extracted with CH₂Cl₂. The extract was successively washed with 2м HCl and water, dried (Na₂SO₄) and concentrated. Column chromatography (ethyl acetate: hexane 1:1) of the residue on silica gel gave 16 (420 mg, 81%) as an amorphous mass; $[\alpha]_D = 11.6^\circ$ (c 1.0, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 1.44 (s, 3H, AcN), 2.04 (s, 3H, MeCOCH₂CH₂), 2.53 (m, 4H, MeCOCH₂CH₂), 3.73 (s, 3H, MeO), 6.80–7.32 (m, 34H, 6Ph, MeOPh). Anal.: calcd for C₈₀H₉₇NO₁₉Si: C, 68.40; H, 6.96; N, 1.00; found: C, 68.39; H, 6.71; N, 0.97%.

2-(Trimethylsilyl)ethyl *O*-(2-acetamido-4-*O*-acetyl-2-deoxy-3-*O*-4-methoxybenzyl-β-D-glucopyranosyl)-(1→3)-*O*-(2,4,6-tri-*O*-benzyl-β-D-galactopyranosyl)-(1→4)-2,3,6-tri-*O*-benzyl-β-D-glucopyranoside (17). A solution of 16 (50.7 mg, 36 μmol) in pyridine (1 mL)-acetic anhydride (1 mL) was kept for 2 h at room temperature and concentrated. Column chromatography (ethyl acetate:hexane 1:2) of the residue on silica gel gave 17 (51 mg, 98%) as an amorphous mass; $[\alpha]_D + 2.0^\circ$ (*c* 1.0, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 1.42 (s, 3H, AcN), 2.01, 2.06 (2s, 6H, AcO, MeCOCH₂CH₂), 2.51 (m, 4H, MeCOCH₂CH₂), 3.77 (s, 3H, MeO), 5.01 (t, 1H, $J_{3,4} = J_{4,5} = 8.1$ Hz, H-4c),

1840 S. Комва et al.

6.81–7.36 (m, 34H, 6Ph, MeOPh). Anal.: calcd for $C_{82}H_{99}NO_{20}Si$: C, 68.08; H, 6.90; N, 0.97; found: C, 67.80; H, 6.79; N, 0.70%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-Olevulinoyl-β-D-galactopyranosyl)-(1→4)-O-(2-acetamido-6-O-benzyl-2-deoxy-3-O-4-methoxybenzyl-β-Dglucopyranosyl)-(1→3)-O-(2,4,6-tri-O-benzyl-β-D-galactopyranosyl)-(1→4)-2,3,6-tri-O-benzyl-β-p-glucopyra**noside** (18). To a solution of 13 (251 mg, 0.18 mmol) and 10 (297 mg, 0.27 mmol) in dry CH₂Cl₂ (1 mL) was added MS-4 Å (AW-300, 400 mg), and the mixture was stirred for 5 h at room temperature, then cooled to 0 °C. Trimethylsilyl trifluoromethanesulfonate (TMS• OTf, 16 µL, 83 µmol) was added to the mixture and this was stirred for 12 h at 7 °C, neutralized with Et₃N and filtered, and the residue was washed with CH₂Cl₂. The combined filtrate and washings were washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (ethyl acetate: hexane 2:1) of the residue on silica gel gave 18 (206 mg, 50%) as an amorphous mass; $[\alpha]_D + 13.5^\circ$ (c 1.3, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 1.44, 1.49 (2s, 6H, 2AcN), 1.78, 1.92, 1.97, 2.10, 2.21 (5s, 15H, 4AcO, MeCOCH₂CH₂), 2.39-2.63 (m, 4H, MeCOCH₂CH₂), 3.68 (s, 3H, MeOPh), 3.89 (s, 3H, MeO), 5.24 (d, 1H, $J_{3,4} = 2.9$ Hz, H-4d), 5.46 (t, 1H, $J_{1,2} = J_{2,3} = 8.1$ Hz, H-2d), 5.68 (m, 1H, H-8e), 6.62–8.24 49H, 9Ph, MeOPh). Anal.: calcd C₁₂₇H₁₄₈N₂O₃₈Si: C, 65.28; H, 6.38; N, 1.20; found: C, 65.04; H, 6.17; N, 1.02%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(4,6-di-O-acetyl-2-Obenzoyl- β -D-galactopyranosyl)- $(1 \rightarrow 4)$ -O-(2-acetamido-2-deoxy-6-*O*-levulinoyl-3-*O*-4-methoxybenzyl-β-D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-benzyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -2,3,6-tri-*O*-benzyl- β -D-glucopyrano**side (19).** Glycosylation of **16** (310.5 mg, 0.022 mmol) with 12 (326.5 mg, 0.33 mmol) in dry CH₂Cl₂ (1 mL) in the presence of TMS•OTf (19 µL, 98 µmol) and MS-4 Å (AW-300, 300 mg) for 12 h at 7 °C and workup as described for 18 gave 19 (377 mg, 77%) as an amorphous mass; $[\alpha]_D + 7.6^\circ$ (c 1.2, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 1.42, 1.50 (2s, 6H, 2AcN), 1.80, 1.96, 2.01, 2.05, 2.06 2.11, 2.19 (7s, 21H, 6AcO, MeCOCH2CH2), 2.47-2.56 (m, 4H, MeCOCH₂CH₂), 3.75 (s, 3H, MeOPh), 3.85 (s, 3H, MeO), 5.02 (d, 1H, $J_{3,4}$ =3.1 Hz, H-4d), 5.30 (t, 1H, $J_{1.2} = J_{2.3} = 8.4$ Hz, H-2d), 5.60 (m, 1H, H-8e), 6.76–8.19 39H, 7Ph, MeOPh). Anal.: calcd $C_{117}H_{142}N_2O_{39}Si: C, 63.06; H, 6.42; N, 1.26; found: C,$ 62.92; H, 6.17; N, 1.21%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-non-ulopyranosylonate)- $(2 \rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-O-levulinoyl- β -D-galactopyranosyl)- $(1 \rightarrow 4)$ -O-(2-acetamido-2-deoxy-6-O-levulinoyl-3-O-4-methoxybenzyl- β -D-

glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-benzyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -2,3,6-tri-O-benzyl- β -D-glucopyra**noside** (20). Glycosylation of 16 (420.5 mg, 0.3 mmol) with 10 (485.6 mg, 0.44 mmol) in dry CH₂Cl₂ (1 mL) in the presence of TMS•OTf (26 µL, 0.13 mmol) and MS-4 Å (AW-300, 300 mg) for 12 h at 7 °C and workup as described for **18** gave **20** (315.6 mg, 45%) as an amorphous mass; $[\alpha]_D + 25.5^{\circ}$ (c 1.0, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 1.41, 1.51 (2s, 6H, 2AcN), 1.79, 1.91, 2.07, 2.09, 2.13, 2.22 (6s, 18H, 4AcO, 2MeCOCH₂CH₂), 2.44-2.70 (m, 8H, 2MeCOCH₂CH₂), 3.68 (s, 3H, MeOPh), 3.89 (s, 3H, MeO), 5.25 (d, 1H, $J_{3,4}$ = 3.3 Hz, H-4d), 5.46 (dd, 1H, $J_{1,2}$ = 9.9 Hz, $J_{2,3}$ = 7.7 Hz, H-2d), 5.62 (m, 1H, H-8e), 6.63-8.21 (m, 44H, 8Ph, MeOPh). Anal.: calcd for $C_{125}H_{148}N_2O_{40}Si$: C, 63.98; H, 6.36; N, 1.19; found: C, 63.73; H, 6.27; N, 1.09%.

2-(Trimethylsilyl)ethyl *O*-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2.4-di-O-benzoyl-6-Olevulinovl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2-acetamido-6-O-benzyl-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-(2,4,6tri - O - benzyl - β - D - galactopyranosyl) - $(1\rightarrow 4)$ - 2,3,6 - tri -O-benzyl-β-D-glucopyranoside (21). To a solution of 18 (63.8 mg, 27 μmol) in CH₃CN (0.9 mL) and water (0.1 mL) was added ceric ammonium nitrate (CAN, 33 mg, 60 µmol), and the mixture was stirred for 3.5 h at room temperature and extracted with CH₂Cl₂. The extract was successively washed with M NaHCO3 and water, dried (Na₂SO₄) and concentrated. Column chromatography (ethyl acetate: hexane 4:1) of the residue on silica gel gave 21 (56.5 mg, 98%) as an amorphous mass; $[\alpha]_D + 22.2^\circ$ (c 1.1, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.92 (m, 2H, Me₃SiCH₂CH₂), 1.49, 1.56 (2s, 6H, 2AcN), 1.77, 1.90, 2.04, 2.11, 2.22 (5s, 15H, 4AcO, MeCOCH₂CH₂), 2.43 [dd, 1H, $J_{\text{gem}} = 12.6$, $J_{3c(eq),4} = 4.4$ Hz, H-3e(eq)], 2.54–2.72 (m, 4H, $MeCOCH_2CH_2$), 3.86 (s, 3H, MeO), 5.20 (d, 1H, $J_{3,4} = 3.1 \text{ Hz}$, H-4d), 5.50 (dd, 1H, $J_{1,2} = 9.7$, $J_{2,3} = 8.3 \text{ Hz}$, H-2d), 5.65 (m, 1H, H-8e), 7.05-8.27 (m, 45H, 9Ph). Anal.: calcd for $C_{119}H_{140}N_2O_{37}Si$: C, 64.43; H, 6.36; N, 1.26; found: C, 64.27; H, 6.16; N, 1.22%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(4,6-di-O-acetyl-2-Obenzoyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2-acetamido-2-deoxy-6-O-levulinoyl-β-D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-benzyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -2,3,6tri-O-benzyl-β-D-glucopyranoside (22). To a solution of 19 (253.2 mg, 0.11 mmol) in CH₃CN (2.7 mL) and water (0.3 mL) was added CAN (250 mg, 0.45 mmol), and the mixture was stirred for 1 h at room temperature. Workup as described for 21 gave 22 (216.5 mg, 90%) as an amorphous mass; $[\alpha]_D + 20.1^{\circ}$ (c 1.6, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.00 (m, 2H, Me₃SiCH₂CH₂), 1.49, 1.52 (2s, 6H, 2AcN), 1.78, 1.95, 2.01, 2.04, 2.07, 2.14, 2.18 (7s, 21H, 6AcO, MeCOCH₂CH₂), 2.34-2.51 (m, 4H, MeCOCH₂CH₂), 3.83 (s, 3H, MeO), 5.24 (dd, 1H, $J_{6,7} = 2.6$, $J_{7,8} = 9.4$ Hz, H-7e), 5.33 (dd, 1H, $J_{1,2}$ =9.9, $J_{2,3}$ =8.1 Hz, H-2d), 5.60 (m, 1H, H-8e), 7.05-8.20 (m, 35H, 7Ph). Anal.: calcd for $C_{109}H_{134}N_2O_{38}Si$: C, 62.10; H, 6.41; N, 1.33; found: C, 61.93; H, 6.26; N, 1.22%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9 $tetra-\textit{O}-acetyl-3,5-dideoxy-d-glycero-\alpha-d-galacto-2-non$ ulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-Olevulinoyl- β -D-galactopyranosyl)- $(1 \rightarrow 4)$ -O-(2-acetamido-2-deoxy-6-O-levulinoyl-β-D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-benzyl- β -D-galactopyranosyl)- $(1\rightarrow 4)-2,3,6$ -tri-*O*-benzyl- β -D-glucopyranoside Selective removal of the 4-methoxybenzyl group in 20 (190 mg, 81 µmol) with CAN (116 mg, 0.21 mmol) in CH₃CN (1.8 mL) and water (0.2 mL) for 3 h at room temperature and workup as described for 21 gave 23 (120 mg, 68%) as an amorphous mass; $[\alpha]_D + 22.6^{\circ}$ (c 1.6, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.00 (m, 2H, Me₃SiCH₂CH₂), 1.49, 1.54 (2s, 6H, 2AcN), 1.77, 2.03, 2.09, 2.13, 2.22 (6s, 18H, 2MeCOCH₂CH₂), 2.33-2.73 (m, 8H, 2MeCOCH₂CH₂), 3.86 (s, 3H, MeO), 5.21 (d, 1H, $J_{3,4}$ =3.3 Hz, H-4d), 5.48 (dd, 1H, $J_{1,2} = 9.7$, $J_{2,3} = 8.2$ Hz, H-2d), 5.62 (m, 1H, H-8e), 7.05-8.21 (m, 40H, 8Ph). Anal.: calcd for $C_{117}H_{140}N_2O_{39}Si: C, 63.12; H, 6.34; N, 1.26; found: C,$ 62.93; H, 6.25; N, 1.24%.

Phenyl 2,3,4-tri-O-acetyl-1-thio-β-L-fucopyranoside (25). To a solution of **24** (59.2 g, 178 mmol) in CH₂Cl₂ (620 mL), cooled to 0 °C, were added, with stirring, thiophenol (37.7 mL, 367 mmol) and boron trifluoride etherate (59 mL, 220 mmol), and the mixture was stirred for 2 h at room temperature and CH₂Cl₂ (300 mL) was added. The solution was successively washed with M Na₂CO₃ and water, dried (Na₂SO₄) and concentrated. Column chromatography (ethyl acetate:hexane 1:4) of the residue on silica gel gave 25 (58.5 g, 86%) as an amorphous mass; $[\alpha]_D = 3.0^\circ$ (c 0.9, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.24 (d, 3H, $J_{5.6}$ = 6.2 Hz, H-6), 1.97, 2.08, 2.14 (3s, 9H, 3AcO), 3.84 (q, 1H, $J_{5.6} = 6.4 \text{ Hz}, \text{ H-5}$, 4.71 (d, 1H, $J_{1.2} = 9.9 \text{ Hz}, \text{ H-1}$), 5.04 (dd, 1H, $J_{2,3} = 9.9$, $J_{3,4} = 3.3$ Hz, H-3), 5.23 (t, 1H, H-2), 5.27 (d, 1H, H-4), 7.28–7.53 (m, 5H, Ph). Anal.: calcd for $C_{18}H_{22}O_7S$: C, 56.53; H, 5.80; found: C, 56.42; H, 5.74%.

Phenyl 1-thio-β-L-fucopyranoside (26). To a solution of 25 (51 g, 0.13 mmol) in MeOH (250 mL) was added NaOMe (1.5 g, 28% NaOMe in MeOH), and the mixture was stirred for 5 h at room temperature and neutralized with Amberlite IR-120 (H $^+$) resin, then filtered and concentrated to a crystalline mass. Recrystallization from ether: hexane gave 26 (32.5 g, 95%) as needles; mp 91–92 °C; [α]_D +68° (c 0.6, MeOH); 1 H NMR (270 MHz, CDCl₃): δ 1.34 (d, 3H, $J_{5.6}$ =6.5 Hz, H-6), 7.30–7.54 (m, 5H, Ph). Anal.: calcd for C₁₂H₁₆O₄S: C, 56.23; H, 6.29; found: C, 56.18; H, 6.19%.

Phenyl 2,3,4-tri-*O***-benzyl-1-thio-** β **-L-fucopyranoside (27)**. To a solution of **26** (1.0 g, 3.9 mmol) in *N*, *N*-dimethylformamide (DMF, 30 mL), cooled to 0 °C, was added NaH in oil suspension (1.2 g, 60% NaH weight), the

mixture was stirred for 30 min at 0 °C, and then benzyl bromide (2 mL) was added. The stirring was continued for 1.5 h at room temperature, and MeOH (1 mL) was added and concentrated then extracted with ethyl acetate. The extract was washed with water, dried (Na₂SO₄) and concentrated to a crystalline mass. Recrystallization from ether: hexane gave **27** (1.7 g, 84%) as needles; mp 107–109 °C; [α]_D –14.0° (c 0.7, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.26 (d, 3H, $J_{5.6}$ =6.4 Hz, H-6), 3.52 (q, 1H, $J_{5.6}$ =6.4 Hz, H-5), 3.59 (dd, 1H, $J_{2.3}$ =11.7, $J_{3.4}$ =2.8 Hz, H-3), 3.93 (dd, 1H, $J_{1.2}$ =9.5 Hz, H-2), 4.60 (d, 1H, H-1), 4.73, 4.76, 4.83 (6H, 3CH₂Ph), 7.18–7.60 (m, 20H, 4Ph). Anal.: calcd for $C_{33}H_{30}O_4S$: C, 75.83; H, 5.79; found: C, 75.56; H, 5.53%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-D-glycero-\alpha-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-Olevulinoyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-benzyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-6-O-benzyl-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-(2,4,6-tri-O-benzyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -2,3,6tri-O-benzyl-β-D-glucopyranoside (28). To a solution of **21** (161.5 mg, 76 μmol) and **27** (45.2 mg, 87 μmol) in dry benzene (1 mL) was added powdered MS-4 Å (300 mg), and the mixture was stirred for 5 h at room temperature, then cooled to 0 °C. Dimethyl(methylthio)sulfonium triflate (DMTST, 68 mg, 0.26 mmol) and MS-4 Å (45 mg) were added to the mixture, and this was stirred for 48 h at 7 °C and neutralized with Et₃N. After dilution with CH₂Cl₂(50 mL), the solids were collected and washed with CH₂Cl₂, and the combined filtrate and washings was washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (ethyl acetate:hexane 3:1) of the residue on silica gel gave **28** (116 mg, 58%) as an amorphous mass; $[\alpha]_D$ -17.5° (c 1.8, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.02 (m, 2H, Me₃SiC \underline{H}_2 CH₂), 1.16 (d, 3H, $J_{5,6}$ =6.4 Hz, H-6f), 1.51, 1.79 (2s, 6H, 2AcN), 1.91, 1.97, 2.03, 2.09, 2.22 (5s, 15H, 4AcO, MeCOCH₂CH₂), 2.41-2.62 (m, 4H, MeCOCH₂CH₂), 3.83 (s, 3H, MeO), 5.15 (d, 1H, $J_{1,2} = 3.6$ Hz, H-1f), 5.42 (dd, 1H, $J_{1,2} = 9.9$, $J_{2,3} = 8.1$ Hz, H-2d), 5.67 (m, 1H, H-8e), 7.08–8.21 (m, 60H, 12Ph). Anal.: calcd for $C_{146}H_{168}N_2O_{41}Si$: C, 66.55; H, 6.43; N, 1.06; found: C, 66.50; H, 6.34; N, 0.91%.

2-(Trimethylsilyl)ethyl *O*-(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -*O*-(4,6-di-*O*-acetyl-2-*O*-benzoyl-β-D-galactopyranosyl)- $(1\rightarrow 4)$ -*O*-[(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl)- $(1\rightarrow 3)$]-*O*-(2-acetamido-2-deoxy-6-*O*-levulinoyl-β-D-glucopyranosyl)- $(1\rightarrow 4)$ -2,3,6-tri-*O*-benzyl-β-D-glucopyranoside (29). Glycosylation of 22 (294.3 mg, 0.14 mmol) with 27 (87 mg, 0.17 mmol) in dry benzene (1 mL) in the presence of DMTST (130 mg, 0.5 mmol) and MS-4 Å (386 mg) for 72 h at 7 °C, then workup as described for 28 gave 29 (317 mg, 90%) as an amorphous mass; [α]_D -12.3° (*c* 1.5, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 1.22 (d, 3H, $J_{5.6}$ =6.4 Hz, H-6f),

1.34, 1.52 (2s, 6H, 2AcN), 1.79, 1.92, 1.95, 2.02, 2.04, 2.06, 2.18 (7s, 21H, 6AcO, MeCOCH₂CH₂), 2.47–2.55 (m, 4H, MeCOCH₂CH₂), 3.84 (s, 3H, MeO), 5.03 (d, 1H, $J_{1,2}$ =3.3 Hz, H-1f), 5.22 (dd, 1H, $J_{1,2}$ =10.1, $J_{2,3}$ =8.1 Hz, H-2d), 5.29 (dd, 1H, $J_{6,7}$ =2.8, $J_{7,8}$ =9.5 Hz, H-7e), 7.04–8.15 (m, 50H, 10Ph). Anal.: calcd for C₁₃₆H₁₆₂N₂O₄₂Si: C, 64.70; H, 6.47; N, 1.11; found: C, 64.54; H, 6.34; N, 0.82%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-O-levulinoyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-benzyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-6-O-levulinovl- β -D-glucopyranosyl)-(1→3)-O-(2,4,6-tri-O-benzyl- β -Dgalactopyranosyl)-(1→4)-2,3,6-tri-O-benzyl-β-Dglucopyranoside (30). Glycosylation of 23 (276 mg, 0.12 mmol) with **27** (77 mg, 0.15 mmol) in dry benzene (1 mL) in the presence of DMTST (116 mg, 0.45 mmol) and MS-4 Å (375 mg) for 72 h at 7 °C, then work up as described for 28 gave 30 (159 mg, 50%) as an amorphous mass; $[\alpha]_D = -14.3^\circ$ (c 1.7, CHCl₃); ¹H (270 MHz, $CDCl_3$): δ 1.01 (m, $Me_3SiCH_2CH_2$), 1.21 (d, 3H, $J_{5.6}=6.4$ Hz, H-6f), 1.41, 1.54 (2s, 6H, 2AcN), 1.79, 1.91, 2.06, 2.08, 2.11, 2.22 (6s, 18H, 4AcO, 2MeCOCH2CH2), 2.42-2.68 (m, 8H, 2MeCOCH₂CH₂), 3.84 (s, 3H, MeO), 5.17 (d, 1H, $J_{1,2} = 3.5$ Hz, H-1f), 5.24 (d, 1H, $J_{3,4} = 2.9$ Hz, H-4d), 5.29 (dd, 1H, $J_{6,7}=2.4$, $J_{7,8}=9.3$ Hz, H-7e), 5.43 (dd, 1H, $J_{1,2} = 10.1$, $J_{2,3} = 8.3$ Hz, H-2d), 5.62 (m, 1H, H-8e), 7.05-8.19 (m, 55H, 11Ph). Anal.: calcd C₁₄₄H₁₆₈N₂O₄₃Si: C, 65.44; H, 6.41; N, 1.06; found: C, 65.16; H, 6.27; N, 0.88%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-p-glycero-α-p-galacto-2nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-O-levulinovl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4)tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-6-O-acetyl-2-deoxy- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -2,3,6tri-O-acetyl-β-D-glucopyranoside (31). A solution of **28** (112.2 mg, 43 µmol) in EtOH (20 mL) and AcOH (4 mL) was hydrogenolysed in the presence of 10% Pd-C (120 mg) for 24 h at 40 °C, then filtered and concentrated. The residue was acetylated with Ac₂O (1 mL) in pyridine (5 mL) for 24 h at 40 °C. The product was dissolved in CH₂Cl₂ (50 mL) and the solution was washed with 2M HCl and water, dried (Na2SO4) and concentrated. Column chromatography (ethyl acetate: hexane 4:1) of the residue on silica gel gave 31 (81.5 mg, 89%) as an amorphous mass; $[\alpha]_D - 10.2^\circ$ (c 1.5, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.92 (m, 2H, Me₃SiCH₂CH₂), 1.19 (d, 3H, $J_{5.6}$ = 6.6 Hz, H-6f), 1.57, 1.78, 1.85, 1.90, 1.91, 2.00, 2.02, 2.03, 2.07, 2.09 (9H), 2.10, 2.11, 2.17, 2.22 2.24 (15s, 51H, 2AcN, 14AcO, MeCOCH₂CH₂), 2.62-2.82 (m, 4H, MeCOCH₂CH₂), 3.87 (s, 3H, MeO), 5.07 (d, 1H, $J_{1,2} = 2.6$ Hz, H-1f), 5.37 (dd, 1H, $J_{1,2}=9.7$, $J_{2,3}=8.6$ Hz, H-2d), 5.68 (m, 1H, H-8e), 7.16-8.17 (m, 10H, 2Ph). Anal.: calcd for $C_{96}H_{128}N_2O_{51}Si: C$, 53.53; H, 5.99; N, 1.30; found: C, 53.39; H, 5.94; N, 1.19%.

2-(Trimethylsilvl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(4,6-di-O-acetyl-2-Obenzoyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-Oacetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2deoxy-6-O-levulinoyl- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O- $(2.4.6 - tri - O - acetyl - \beta - D - galactopyranosyl) - (1 \rightarrow 4) -$ 2,3,6-tri-O-acetyl-β-D-glucopyranoside (**32**). Hydrogenolysis of 29 (317 mg, 0.13 mmol) in EtOH (40 mL) and AcOH (8 mL) in the presence of 10% Pd-C (320 mg) for 48 h at 40 °C, and subsequent acetylation with Ac₂O (2 mL) in pyridine (5 mL) as described for 31 gave 32 (247 mg, 94%) as an amorphous mass; $[\alpha]_D$ -18.0° (c 1.2, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.92 (m, 2H, Me₃SiCH₂CH₂), 1.24 (d, 3H, $J_{5.6} = 7.2$ Hz, H-6f), 1.49, 1.78 (2s, 6H, 2AcN), 1.90, 1.95, 1.96, 1.99, 2.02, 2.05, 2.08 (6H), 2.09, 2.10, 2.15, 2.17, 2.18 (9H), 2.20 (13s, 48H, 15AcO, MeCOCH₂CH₂), 2.51 (dd, 1H, $J_{gem} = 12.5$, $J_{3c(eq).4} = 4.6$ Hz, H-3e(eq)), 2.64–2.79 (m, 4H, McCOCH₂CH₂), 3.85 (s, 3H, MeO), 5.07 (d, 1H, $J_{1.2} = 3.1$ Hz, H-1f), 5.60 (m, 1H, H-8e), 7.32–8.14 (m, 5H, Ph). Anal.: calcd for $C_{91}H_{126}N_2O_{51}Si$: C, 52.25; H, 6.07; N, 1.34; found: C, 52.07; H, 5.77; N, 1.17%.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9tetra-O-acetyl-3,5-dideoxy-p-glycero-α-p-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-Olevulinoyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-6-O-levulinoyl- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O- $(2,4,6-\text{tri}-O-\text{acetyl}-\beta-\text{p-galactopyranosyl})-(1\rightarrow 4)$ **2,3,6-tri-O-acetyl-β-p-glucopyranoside** (33). Hydrogenolysis of 30 (159 mg, 60 µmol) in EtOH (20 mL) and AcOH (4 mL) in the presence of 10% Pd-C (160 mg) for 48 h at 40 °C, and subsequent acetylation with Ac₂O (2 mL) in pyridine (5 mL) as described for 31 gave 33 (109 mg, 82%) as an amorphous mass; $[\alpha]_D$ -11.0° (c 1.2, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.92 (m, 2H, Me₃SiCH₂CH₂), 1.19 (d, 3H, $J_{56} = 6.4$ Hz, H-6f), 1.57, 1.78 (2s, 6H, 2AcN), 1.85, 1.90, 1.91, 1.99, 2.02, 2.08 (12H), 2.09 (6H), 2.18 (6H), 2.20, 2.22 (10s, 45H, 13AcO, 2MeCOCH₂CH₂), 2.38 [dd, $J_{gem} = 13.0, J_{3e(eq).4} = 4.9 \text{ Hz}, \text{ H-3e}(eq), 2.61-2.86 \text{ (m, 8H,}$ 2MeCOCH₂CH₂), 3.87 (s, 3H, MeO), 5.66 (m, 1H, H-8e), 7.28-8.17 (m, 10H, 2Ph). Anal.: calcd for $C_{99}H_{132}N_2O_{52}Si: C, 53.80; H, 6.02; N, 1.27; found: C,$ 53.70; H, 5.93; N, 1.12%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxyn-glycero-α-d-death-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(2,4-di-O-benzoyl-6-O-levulinoyl-β-d-galactopyranosyl)-(1 \rightarrow 4)-O-[(2,3,4-tri-O-acetyl-α-L-fucopyranosyl)-(1 \rightarrow 3)]-O-(2-acetamido-6-O-acetyl-2-deoxy-β-d-glucopyranosyl)-(1 \rightarrow 3)-O-(2,4,6-tri-O-acetyl-β-d-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-acetyl-α-d-glucopyranosyl trichloroacetimidate (34). To a solution of 31 (72.6 mg, 34 μmol) in CH₂Cl₂ (4.5 mL), cooled to 0 °C, was added CF₃CO₂H (1.25 mL), and the mixture was stirred for 2 h at room temperature and concentrated. The product was purified by chromatography on a column of silica gel with ethyl acetate:hexane 4:1 to give the 1-hydroxy compound (63 mg). To a solution of

this in CH₂Cl₂ (0.8 mL), cooled to 0 °C, were added trichloroacetonitrile (92 μ L, 0.92 mmol) and DBU (5.5 μ L, 36 μ mol), the mixture was stirred for 2 h at 0 °C, and the progress of the reaction was monitored by TLC. The mixture was concentrated. Column chromatography (ethyl acetate:hexane 4:1) of the residue on silica gel gave **34** (67 mg, 91%) as an amorphous mass; [α]_D +6.6° (c 1.4, CHCl₃); 'H NMR (270 MHz, CDCl₃): δ 1.19 (d, 3H, $J_{5.6}$ =6.6 Hz, H-6f), 1.57–2.23 (13s, 51H, 14AcO, 2AcN, MeCOCH₂CH₂), 2.38 (dd, 1H, J_{gem} =12.5, $J_{3c(eq),4}$ =4.4 Hz, H-3e(eq)), 2.62–2.86 (m, 4H, MeCOCH₂CH₂), 3.87 (s, 3H, MeO), 6.47 (d, 1H, $J_{1.2}$ =3.5 Hz, H-1a), 7.28–8.17 (m, 10H, 2Ph), 8.66 (s, 1H, C=NH). Anal.: calcd for C₉₃H₁₁₆N₃O₅₁Cl₃: C, 50.81; H, 5.32; N, 1.91; found: C, 50.78; H, 5.13; N, 1.88%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-p-glycero-α-p-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzovl-6-O-levulinovl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-6-O-acetyl-2-deoxy- β -Dglucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2,3,6-tri-O-acetyl- β -D-glucopyranosyl)- $(1\rightarrow 1)$ -(2S,3R,4E)-2-azido-3-O-benzoyl-4octadecene-1,3-diol (36). To a solution of 34 (83.6 mg, 38 μ mol) and (2S,3R,4E)-2-azido-3-O-benzoyl-4-octadecene-1,3-diol (35, 32.5 mg, 76 μmol) in dry CH₂Cl₂ (1 mL) was added MS 4 Å (AW-300, 300 mg), and the mixture was stirred for 5 h at room temperature, then cooled to 0 °C. BF₃•OEt₂ (10 µL, 37 µmol) was added, and the mixture was stirred for a further 5 h at 0 °C. The solids were filtered off and washed with CH₂Cl₂. The combined filtrate and washings were washed with M Na₂CO₃ and water, dried (Na₂SO₄) and concentrated. Column chromatography (ethyl acetate: hexane 3:1) of the residue on silica gel gave 36 (59.5) mg, 63%) as an amorphous mass; $[\alpha]_D = 7.5^{\circ}$ (c 1.2, CHCl₃); IR (KBr): 3300, 2100, 1750, 1680, 1540, 1230, 710 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 0.88 (t, 3H, $J_{\text{Me,CH2}} = 6.8 \text{ Hz}, \underline{\text{Me}}_{\text{CH}_2}, 1.24 \text{ (s, 22H, 11CH}_2), 1.58,$ 1.78 (2s, 6H, 2AcN), 1.85-2.23 (15s, 45H, 14AcO, MeCOCH₂CH₂), 2.38 [dd, 1H, $J_{gem} = 12.6$, $J_{3c(eq),4} = 4.2$ Hz, H-3e(eq)], 2.62-2.81 (m, 4H, MeCOCH₂CH₂), 3.87(s, 3H, MeO), 4.46 (d, 1H, $J_{1,2}$ = 7.0 Hz, H-1a), 5.92 (dt, 1H, $J_{4,5} = 14.7$, $J_{5,6} = J_{5,6'} = 6.8$ Hz, H-5 of sphingosine), 7.28 - 8.17(m, 15H, 3Ph). Anal.: calcd $C_{116}H_{153}N_5O_{53}$: C, 56.51; H, 6.26; N, 2.84; found: C, 56.32; H, 5.96; N, 2.56%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dide-oxy-d-glycero-α-d-galacto-2-nonulopyranosylonate)-(2→3)-O-(2,4-di-O-benzoyl-6-O-levulinoyl-β-d-galacto-pyranosyl)-(1→4)-O-[(2,3,4-tri-O-acetyl-α-L-fucopyranosyl)-(1→3)]-O-(2-acetamido-6-O-acetyl-2-deoxy-β-d-glucopyranosyl)-(1→4)-O-(2,4,6-tri-O-acetyl-β-d-galacto-glucopyranosyl)-(1→4)-O-(2,3,6-tri-O-acetyl-β-d-glucopyranosyl)-(1→1)-(2S,3R,4E)-2-octadecanamido-4-octadecene-1,3-diol (37). Hydrogen sulfide was bubbled through a stirred solution of 36 (72.5 mg, 29 μmol) in aq 83% pyridine (10 mL) for 3 days at 0 °C, with the progress of the reaction being monitored by

TLC. The mixture was concentrated, and the residue was stirred with octadecanoic acid (17 mg, 60 µmol) 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (WSC, 17 mg, 89 μmol) in dry CH₂Cl₂ (2 mL) for 12 h at room temperature. After completion of the reaction, CH₂Cl₂ (30 mL) was added to the mixture, and the solution was washed with water, dried (Na₂SO₄) and concentrated to a syrup that was chromatographed on a column of silica gel with ethyl acetate: hexane 4:1, to give 37 (43.2 mg, 54%) as an amorphous mass; $[\alpha]_D = 5.1^\circ$ (c 0.9, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.88 (t, 6H, $J_{\text{Me,CH2}} = 7.0$ Hz, 2MeCH₂), 1.26 (s, 52H, 26CH₂), 1.57, 1.78 (2s, 6H, 2AcN), 1.85–2.23 (15s, 45H, 14AcO, MeCOCH₂CH₂), 2.38 (dd, 1H, $J_{gem} = 12.5$, $J_{3c(eq),4} = 4.2$ Hz, H-3e(eq)), 2.62-2.81 (m, 4H, MeCOCH₂CH₂), 3.87 (s, 3H, MeO), 5.87 (dt, 1H, $J_{4,5} = 13.7$, $J_{5,6} = J_{5,6'} = 6.8$ Hz, H-5 of sphingosine), 7.27-8.17 (m, 15H, 3Ph). Anal.: calcd for $C_{134}H_{189}N_3O_{54}$: C, 59.48; H, 7.07; N, 1.50; found: C, 59.45; H, 6.88; N, 1.29%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(4,6-di-O-acetyl-2-O-benzoyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-6-O-levulinoyl- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- β -Dgalactopyranosyl)- $(1 \rightarrow 4)$ -2,3,6-tri-O-acetyl- α -Dglucopyranosyl trichloroacetimidate (38). Selective removal of the 2-(trimethylsilyl)ethyl group in 32 (220.5) mg, $0.11 \mu mol$) with CF₃CO₂H (4 mL) in CH₂Cl₂ (12 mL) for 2.5 h at room temperature, and subsequent reaction of the product with trichloroacetonitrile (0.32) μL, 3.2 mmol) in CH₂Cl₂ (1.8 mL) in the presence of DBU (19 μ L, 0.13 mmol) for 2 h at 0 °C as described for **34** gave **38** (218.7 mg, 98%) as an amorphous mass; [α]_D +8.3° (*c* 1.7, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.24 (d, 3H, $J_{5.6}$ =7.1 Hz, H-6f), 1.49, 1.78 (2s, 6H, 2AcN), 1.91-2.20 (16s, 48H, 15AcO, MeCOCH₂CH₂), 2.51 [dd, 1H, $J_{gem} = 12.8$, $J_{3c(eq),4} = 4.4$ Hz, H-3e(eq)], 2.69-2.79 (m, 4H, MeCOCH₂CH₂), 3.86(s, 3H, MeO), 6.47 (d, 1H, $J_{1,2}=3.7$ Hz, H-1a), 7.30-8.15 (m, 5H, Ph), 8.67 (s, 1H, C= NH). Anal.: calcd for $C_{88}H_{114}N_3O_{51}Cl_3$: C, 49.48; H, 5.38; N, 1.97; found: C, 49.41; H, 5.16; N, 1.71%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)-(2→3)-O-(4,6-di-O-acetyl-2-O-benzoyl-β-D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-6-O-levulinoyl- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2,3,6-tri-O-acetyl- β -p-glucopyranosyl)- $(1\rightarrow 1)$ -(2S,3R,4E)-2-azido-3-O-benzoyl-4-octadecene-1,3-diol (39). Coupling of 38 (218.7 mg, 0.1 mmol) with 35 (88 mg, 0.21 mmol) in CH₂Cl₂ (2 mL) in the presence of BF₃•OEt₂ (28 μL, 0.1 mmol) and MS-4 Å (AW-300, 600 mg) as described for **36** gave **39** (142 mg, 58%) as an amorphous mass; $[\alpha]_D - 18.6^{\circ}$ (c 1.8, CHCl₃); IR (KBr): 3350, 2100, 1750, 1680, 1550, 1220, 710 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 0.88 (t, 3H, $J_{\text{Me,CH2}} = 6.8$ Hz, MeCH₂), 1.24 (s, 22H, 11CH₂),

1.49, 1.78 (2s, 6H, 2AcN), 1.90–2.20 (16s, 48H, 15AcO, MeCOCH₂CH₂), 2.51 [dd, 1H, J_{gem} = 12.6, $J_{3c(eq).4}$ = 4.6 Hz, H-3e(eq)], 3.86 (s, 3H, MeO), 4.49 (d, 1H, $J_{1.2}$ = 7.7 Hz, H-1a), 5.91 (dt, 1H, $J_{4.5}$ = 14.1, $J_{5.6}$ = $J_{5.6'}$ = 7.0 Hz, H-5 of sphingosine), 7.30–8.15 (m, 10H, 2Ph). Anal.: calcd for $C_{111}H_{151}N_5O_{53}$: C, 55.47; H, 6.33; N, 2.91; found: C, 55.21; H, 6.24; N, 2.73%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(4,6-di-O-acetyl-2-O-benzoyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-6-O-levulinoyl- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-Oacetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2,3,6-tri-O-acetyl- β -p-glucopyranosyl)- $(1 \rightarrow 1)$ -(2S,3R,4E)-3-O-benzoyl-2-octadecanamido-4-octadecene-1,3-diol (40). Selective reduction of the azido group in 39 (142 mg, 59 μmol) with H₂S in aq 83% pyridine (20 mL), followed by coupling of the product with octadecanoic acid (34 mg, 0.12 mmol) in the presence of WSC (34 mg, 0.18 mmol) and workup as described for 37 gave 40 (91.5 mg, 60%) as an amorphous mass; $[\alpha]_D = 9.8^{\circ}$ (c 1.8, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.88 (t, 6H, $J_{\text{Me,CH2}} = 6.8 \text{ Hz}, 2 \underline{\text{Me}} \text{CH}_2$, 1.26 (s, 52H, 26CH₂), 1.49, 1.78 (2s, 6H, 2AcN), 1.89-2.20 (16s, 48H, 15AcO, <u>Me</u>COCH₂CH₂), 2.51 [dd, 1H, $J_{gem} = 12.3$, $J_{3e(eq).4} = 4.2$ Hz, H-3e(eq)], 2.63-2.81 (m, 4H, MeCOCH₂CH₂), 3.86(s, 3H, MeO), 5.07 (d, 1H, $J_{1,2}$ = 2.9 Hz, H-1f), 5.86 (dt, 1H, $J_{4.5} = 14.7$, $J_{5.6} = J_{5.6'} = 7.3$ Hz, H-5 of sphingosine), 7.30–8.14 (m, 10H, 2Ph). Anal.: calcd for $C_{129}H_{187}N_3O_{54}$: C, 58.60; H, 7.13; N, 1.59; found: C, 58.40; H, 6.93; N, 1.38%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-O-levulinoyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-6-O-levulinovl- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -2,3,6-tri-O-acetyl- α -Dglucopyranosyl trichloroacetimidate (41). Selective removal of the 2-(trimethylsilyl)ethyl group in 33 (109 mg, 50 μmol) with CF₃CO₂H (2 mL) in CH₂Cl₂ (6 mL) for 2.5 h at room temperature, followed by treatment of the 1-hydroxy compound with trichloroacetonitrile (0.143 mL, 1.4 mmol) in CH_2Cl_2 (0.9 mL) in the presence of DBU (8.5 µL, 57 µmol) for 2 h at 0 °C and work up as described for 34 gave 41 (99.5 mg, 89%) as an amorphous mass; $[\alpha]_D + 8.3^{\circ}$ (c 1.7, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 1.19 (d, 3H, $J_{5.6} = 6.4$ Hz, H-6f), 1.58, 1.78 (2s, 6H, 2AcN), 1.85-2.32 (15s, 45H, 2MeCOCH₂CH₂), 2.61 - 2.8513AcO, (m, 2MeCOCH₂CH₂), 3.87 (s, 3H, MeO), 5.64 (m, 1H, H-8e), 6.47 (d, 1H, $J_{1,2}$ =3.5 Hz, H-1a), 7.28-8.17 (m, 10H, 2Ph), 8.65 (s, 1H, C= NH). Anal.: calcd for C₉₆H₁₂₀N₃O₅₂Cl₃: C, 51.15; H, 5.37; N, 1.86; found: C, 51.02; H, 5.30; N, 1.62%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dide-oxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-O-levulinoyl- β -D-galacto-

pyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-6-O-levulinoyl- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2,3,6-tri-O-acetyl- β -Dglucopyranosyl)- $(1\rightarrow 1)$ -(2S,3R,4E)-2-azido-3-O-benzoyl-4-octadecene-1,3-diol (42). Coupling of 41 (99.5 mg, 44 μmol) with **35** (38 mg, 89 μmol) in CH₂Cl₂ (1 mL) in the presence of BF₃•OEt₂ (12 μL, 44 μmol) and MS-4 Å (AW-300, 300 mg) as described for **36** gave **42** (53 mg, 48%) as an amorphous mass; $[\alpha]_D = 12.8^\circ$ (c 1.1, CHCl₃); IR (KBr) 3350, 2100, 1750, 1690, 1540, 1220, 710 cm⁻¹; ¹H NMR (270 MHz, CDCl₃): δ 0.86 (t, 3H, $J_{\text{Me,CH2}} = 7.0$ Hz, MeCH₂), 1.24 (s, 22H, 11CH₂), 1.58, 1.78 (2s, 6H, 2AcN), 1.85-2.23 (15s, 45H, 13AcO, 2MeCOCH₂CH₂), 2.38 (dd, 1H, $J_{gem} = 12.3$, $J_{3c(eq),4} = 4.8$ Hz, H-3e(eq)), 2.63–2.78 (m, 8H, 2MeCOCH₂CH₂), 3.87 (s, 3H, MeO), 4.43 (d, 1H, $J_{1,2} = 7.6$ Hz, H-1a), 5.91 (dt, 1H, $J_{4.5} = 13.6$, $J_{5.6} = J_{5.6'} = 6.8$ Hz, H-5 of sphingosine), 7.28–8.17 (m, 15H, 3Ph). Anal.: calcd for $C_{119}H_{157}N_5O_{54}$: C, 56.68; H, 6.28; N, 2.78; found: C, 56.62; H, 6.14; N, 2.51%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-O-levulinoyl-β-D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-6-O-levulinoyl- β -Dglucopyranosyl)-(1→3)-O-(2,4,6-tri-O-acetyl-β-D-galactopyranosyl)- $(1\rightarrow 4)$ -(2,3,6-tri-O-acetyl- β -D-glucopyranosyl)- $(1\rightarrow 1)$ -(2S,3R,4E)-3-O-benzoyl-2-octadecanamido-**4-octadecene- 1,3-diol (43).** Selective reduction of the azido group in **42** (53 mg, 21 μ mol) with H_2S in aq 83% pyridine (10 mL), followed by coupling of the product with octadecanoic acid (12 mg, 42 µmol) in the presence of DBU (12 mg, 63 µmol) and workup as described for 37 gave 43 (34.5 g, 60%) as an amorphous mass; $[\alpha]_D - 6.0^\circ$ (c 0.7, CHCl₃); H NMR (270 MHz, CDCl₃): δ 0.88 (t, 6H, $J_{Me,CH2} = 6.8$ Hz, 2Me₂CH₂), 1.26 (s, 52H, 26CH₂), 1.58, 1.78 (2s, 6H, 2AcN), 1.85–2.24 (15s, 45H, 13AcO, 2MeCOCH₂CH₂), 2.38 [dd, 1H, $J_{gem} = 12.8$, $J_{3e(eq).4} = 4.8$ Hz, H-3e(eq)], 2.61-2.78 (m, ^{8}H , $^{2}MeCOCH_{2}CH_{2}$), $^{3.87}$ (s, ^{3}H , MeO), 5.86 (dt, 1H, $J_{4,5} = 14.4$, $J_{5,6} = J_{5,6} = 7.2$ Hz, H-5 of sphingosine), 7.27-8.17 (m, 15H, 3Ph). Anal.: calcd for C₁₃₇H₁₉₃N₃O₅₅: C, 59.58; H, 7.04; N, 1.52; found: C, 59.41; H, 6.80; N, 1.26%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dide-oxy-p-glycero-α-p-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-β-p-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl-α-L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-6-O-acetyl-2-deoxy-β-p-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl-β-p-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2,3,6-tri-O-acetyl-β-p-glucopyranosyl)- $(1\rightarrow 1)$ -(2S,3R,4E)-3-O-benzoyl-2-octadecanamido-4-octadecene-1,3-diol (44). To a solution of 37 (10.7 mg, 3.9 μmol) in EtOH (0.3 mL) was added hydrazine monoacetate (0.4 mg, 4.3 μmol), and the mixture was stirred for 30 min at room temperature then concentrated. Column chromatography (CH₂Cl₂:MeOH 20:1) of the residue on silica gel gave 44 (7.0 mg, 70%) as an amorphous mass; [α]_D -6.4° (c 0.5, CHCl₃); ¹H NMR

(270 MHz, CDCl₃): δ 0.88 (t, 6H, $J_{\text{Me,CH2}}$ =7.0 Hz, 2MeCH₂), 1.26 (s, 52H, 26CH₂), 1.54, 1.78 (2s, 6H, 2AcN), 1.86–2.22 (14s, 42H, 14AcO), 2.38 [dd, 1H, J_{gem} = 12.1, $J_{3c(eq),4}$ = 4.2 Hz, H-3e(eq)], 3.77 (s, 3H, MeO), 5.86 (dt, 1H, $J_{4.5}$ = 13.4, $J_{5.6}$ = $J_{5.6'}$ = 6.7 Hz, H-5 of sphingosine), 7.27–8.18 (m, 15H, 3Ph). Anal.: calcd for C₁₂₇H₁₈₃N₃O₅₂: C, 59.41; H, 7.07; N, 1.61; found: C, 59.32; H, 6.99; N, 1.47%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl-6-O-sulfo- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-6-O-acetyl-2-deoxy- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -(2,3,6-tri-O-acetyl- β -D-glucopyranosyl)- $(1\rightarrow 1)$ -(2S,3R,4E)-3-O-benzoyl-2-octadecanamido-4-octadecene-1,3-diol pyridine salt (45). To a solution of 44 (23.5 mg, 8.9 µmol) in DMF (0.5 mL) was added sulfur trioxide-pyridine complex (7 mg, 44 µmol) and the mixture was stirred for 4 h at room temperature. MeOH (2 mL) was added, and the mixture was concentrated. Column chromatography (CHCl₃: MeOH 5:1) of the residue on Sephadex LH-20 (20 g) gave the crude product, and this was purified by column chromatography on silica gel (20 CH₂Cl₂: MeOH 15:1, to give 45 (24.2 mg, 98%) as an amorphous mass; $[\alpha]_D$ -3.3° (c 0.5, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.88 (t, 6H, $J_{Mc,CH2} = 7.0$ Hz, 2MeCH₂), 1.26 (s, 52H, 26CH₂), 1.55, 1.78 (2s, 6H, 2AcN), 1.88-2.24 (14s, 42H, 14AcO), 3.82 (s, 3H, MeO), 5.86 (dt, 1H, $J_{4.5} = 14.7$, $J_{5.6} = J_{5.6} = 7.3$ Hz, H-5 of sphingosine), 7.27-8.15 (m, 20H, 3Ph, C₅H₅N). Anal.: calcd for $C_{134}H_{187}N_4O_{55}S$: C, 58.19; H, 6.81; N, 2.03; found: C, 58.10; H, 6.65; N, 1.98%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(4,6-di-O-acetyl-2-O-benzoyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2,3,6-tri-O-acetyl- β -D-glucopyranosyl)- $(1\rightarrow 1)$ -(2S,3R,4E)-3-O-benzoyl-2-octadecanamido-4octadecene-1,3-diol (46). Selective hydrolysis of the levulinoyl group in 40 (91.1 mg, 34 μmol) with hydrazine monoacetate (3.8 mg, 41 µmol) in EtOH (2.1 mL) and workup as described for 44 gave 46 (44.5 mg, 50%) as an amorphous mass; $[\alpha]_D - 13.0^\circ$ (c 0.6, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.88 (t, 6H, $J_{\text{Me,CH2}} = 6.8$ Hz, 2MeCH₂), 1.26 (s, 52H, 26CH₂), 1.48, 1.78 (2s, 6H, 2AcN), 1.88-2.21 (15s, 45H, 15AcO), 2.53 [dd, 1H, $J_{gem} = 12.3$, $J_{3e(eq),4} = 4.4$ Hz, H-3e(eq)], 3.87 (s, 3H, MeO), 5.87 (dt, 1H, $J_{4,5} = 13.4$, $J_{5,6} = J_{5,6'} = 6.7$ Hz, H-5 of sphingosine), 7.27-8.18 (m, 10H, 2Ph). Anal.: calcd for $C_{124}H_{181}N_3O_{52}$: C, 58.50; H, 7.17; N, 1.65; found: C, 58.34; H, 7.12; N, 1.64%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dide-oxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)- $(2 \rightarrow 3)$ -O-(4,6-di-O-acetyl-2-O-benzoyl- β -D-galacto-pyranosyl)- $(1 \rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- α -L-fucopyra-

nosyl)- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-6-O-sulfo- β -Dglucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)-(1→4)-(2,3,6-tri-O-acetyl-β-D-glucopyranosyl)- $(1\rightarrow 1)$ -(2S,3R,4E)-3-O-benzoyl-2-octadecanamido-4-octadecene-1,3-diol pyridine salt (47). To a solution of 46 (29 mg, 11 µmol) in DMF (0.5 mL) was added sulfur trioxide-pyridine complex (9 mg, 57 µmol), and the mixture was stirred for 2 h at room temperature and work up as described for 45 gave 47 (28 mg, 91%) as an amorphous mass; $[\alpha]_D = 18.7^\circ$ (c 0.6, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.88 (t, 6H, $J_{\text{Me,CH}2} = 7.0 \text{ Hz}$, $2\underline{\text{Me}}\text{CH}_2$), 1.26 (s, 52H, 26CH₂), 1.48, 1.80 (2s, 6H, 2AcN), 1.85–2.18 (15s, 45H, 15AcO), 3.80 (s, 3H, MeO), 5.86 (dt, 1H, $J_{4.5}$ =13.2, $J_{5.6} = J_{5.6'} = 6.6$ Hz, H-5 of sphingosine), 7.27–8.24 (m, 15H, 2Ph, C_5H_5N). Anal.: calcd for $C_{129}H_{185}N_4O_{55}S$: C, 57.30; H, 6.90; N, 2.07; found: C, 57.04; H, 6.87; N, 1.80%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4-di-O-benzoyl- β -D-galactopyranosyl)- $(1\rightarrow 4)-O-[(2,3,4-\text{tri}-O-\text{acetyl}-\alpha-\text{L-fucopyranosyl})-(1\rightarrow$ 3)]-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 3)-O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)- $O-(2,3,6-\text{tri}-O-\text{acetyl}-\beta-\text{D-glucopyranosyl})-(1\rightarrow 1)$ (2S, 3R, 4E)-3-O-benzoyl-2-octadecanamido-4-octadecene-1,3-diol (48). Selective hydrolysis of the levulinoyl groups in 43 (34.2 mg, 12 µmol) with hydrazine monoacetate (2.7 mg, 29 µmol) in EtOH (0.7 mL) for 3 h at room temperature and workup as described for 44 gave 48 (16 mg, 49%) as an amorphous mass; $[\alpha]_D$ -6.3° (c 0.3, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.88 (t, 6H, $J_{Mc,CH_2} = 7.0$ Hz, 2MeCH₂), 1.26 (s, 52H, 26CH₂), 1.55, 1.78 (2s, 6H, 2AcN), 1.82–2.24 (13s, 39H, 13AcO), 3.77 (s, 3H, MeO), 5.86 (dt, 1H, $J_{4,5} = 14.8$, $J_{5,6} = J_{5,6'} = 7.4$ Hz, H-5 of sphingosine), 7.32–8.30 (m, 15H, 3Ph). Anal.: calcd for $C_{127}H_{181}N_3O_{51}$: C, 59.45; H, 7.11; N, 1.64; found: C, 59.43; H, 7.07; N, 1.50%.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2.4-di-O-benzoyl-6-O-sulfo-β-D-galactopyranosyl)- $(1\rightarrow 4)-O-[(2,3,4-\text{tri}-O-\text{acetyl}-\alpha-\text{L-fucopyranosyl})-(1\rightarrow 3)]-$ O-(2-acetamido-2-deoxy-6-O-sulfo- β -D-glucopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ - $O-(2,3,6-\text{tri-}O-\text{acetyl-}\beta-\text{d-glucopyranosyl})-(1\rightarrow 1)-$ (2S,3R,4E)-3-O-benzoyl-2-octadecanamido-4-octadecene-1,3-diol dipyridine salt (49). To a solution of 48 (10.5 mg, 4 µmol) in DMF (0.5 mL) was added sulfur trioxide-pyridine complex (6.4 mg, 40 µmol) and the mixture was stirred for 2 h at room temperature and work up as described for 45 gave 49 (10.6 mg, 91%) as an amorphous mass; $[\alpha]_D = -8.0^\circ$ (c 0.4, CHCl₃); ¹H NMR (270 MHz, CDCl₃): δ 0.88 (t, 6H, $J_{Mc,CH_2} = 6.8$ Hz, 2MeCH₂), 1.26 (s, 52H, 26CH₂), 1.60-2.24 (15s, 45H, 13AcO, 2AcN), 3.78 (s, 3H, MeO), 5.86 (m, 1H, H-5 of sphingosine), 7.26-8.25 (m, 25H, 3Ph, $2C_5H_5N$). Anal.: calcd for $C_{137}H_{189}N_5O_{57}S_2$: C, 57.09; H, 6.61; N, 2.43; found: C, 57.02; H, 6.53; N, 2.23%.

O-(5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2nonulopyranosylonic acid)-(2→3)-O-(6-O-sulfo-β-Dgalactopyranosyl)- $(1\rightarrow 4)$ -O- $[\alpha$ -L-fucopyranosyl- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-β-D-glucopyranosyl)-(1 \rightarrow 3)-O-β-D-galactopyranosyl- $(1 \rightarrow 4)$ -O- β -D-glucopyranosyl- $(1 \rightarrow 1)$ -(2S,3R,4E)-2-octadecanamido-1,3-diol disodium salt (50). To a solution of 45 (24.2 mg, 8.7 μ mol) in MeOH (2 mL) and 1,4-dioxane (1 mL) was added NaOMe (600 mg, 28% NaOMe in MeOH), and the mixture was stirred for 48 h at 20 °C then water (1 mL) was added. After completion of the reaction (24 h), the mixture was concentrated. Column chromatography $(CHCl_3: MeOH: H_2O: Et_3N 5: 4: 0.7: 0.07)$ of the residue on Sephadex (40 g) gave **50** (15.5 mg, 98%) as -5.7° (c an amorphous mass; $[\alpha]_D -5.7^\circ$ (c 0.5, CHCl₃:MeOH:H₂O 5:4:0.7); ¹H NMR (270 MHz, CDCl₃:CD₃OD 1:1): δ 0.89 (t, 6H, $J_{Me,CH2} = 7.0$ Hz, 2MeCH₂), 1.27 (s, 52H, 26CH₂), 2.31 [dd, 1H, $J_{gem} = 11.0$, $J_{3e(eq).4} = 4.0$ Hz, H-3e(eq)], 5.46 (dd, 1H, $J_{3.4} = 6.8$, $J_{4.5} = 15.9$ Hz, H-4 of sphingosine), 5.69 (dt, 1H, $J_{5,6} = J_{5,6'} = 7.7$ Hz, H-5 of sphingosine); IR (KBr) 3600–3200, 2920, 2850, 1700, 1660, 1550 cm⁻¹; The mass spectrum of 50 (negative ion mode) showed the base peak at m/z 1794 (M-Na) and 1771 (M-2Na)².

O-(5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2nonulopyranosylonicacid) - (2 → 3) - O-β-D-galactopyranosyl- $(1\rightarrow 4)$ -O-[α -L-fucopyranosyl- $(1\rightarrow 3)$]-O-(2-acetamido-2-deoxy-6-*O*-sulfo-β-D-glucopyranosyl)-(1→3)-*O*- β -D-galactopyranosyl- $(1 \rightarrow 4)$ -O- β -D-glucopyranosyl- $(1\rightarrow 1)$ -(2S,3R,4E)-2-octadecanamido-1,3-diol disodium salt (51). To a solution of 47 (28 mg, 10 μ mol) in MeOH (2 mL) and 1,4-dioxane (1 mL) was added NaOMe (600 mg, 28% NaOMe in MeOH), and work up as described for 50 gave 51 (18.2 mg, 98%) as an amorphous mass; $[\alpha]_D$ -8.7° (c 0.6, CHCl₃: MeOH: H₂O 5:7:2); IR (KBr) 3600-3200, 2920, 2850, 1700, 1660, 1550 cm⁻¹; ¹H NMR (270 MHz, CDCl₃:CD₃OD 1:1): δ 0.89 (t, 6H, $J_{\text{Me,CH}_2}$ =6.8 Hz, 2MeCH₂), 1.28 (s, 52H, 26CH₂), 2.31 [m, 1H, H-3e(eq)], 5.46 (m, 1H, H-4 of sphingosine), 5.69 (dt, 1H, $J_{vem} = 15.5$, $J_{5.6} = J_{5.6} = 7.0$ Hz, H-5 of sphingosine). The mass spectrum of 51 (negative ion mode) showed the base peaks at m/z 1794 (M-Na) and 1771 $(M-2Na)^{2-}$.

O-(5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→3)-*O*-(6-*O*-sulfo-β-D-galactopyranosyl)-(1→4)-*O*-[α-L-fucopyranosyl-(1→3)]-*O*-(2-acetamido-2-deoxy-6-*O*-sulfo-β-D-glucopyranosyl-(1→3)-*O*-β-D-galactopyranosyl-(1→4)-*O*-β-D-glucopyranosyl-(1→1)-(2*S*,3*R*,4*E*)-2-octadecanamido-4-octadecene-1,3-diol trisodium salt (52). To a solution of 49 (10.6 mg, 3.6 μmol) in MeOH (2 mL) and 1,4-dioxanc (1 mL) was added NaOMe (600 mg, 28% NaOMe in MeOH), and work up as described for 50 gave 52 (6.8 mg, 98%) as an amorphous mass; [α]_D −10.3° (*c* 0.2, CHCl₃:MeOH:H₂O 5:4:0.7); IR (KBr): 3600–3250, 2920, 2850, 1700, 1660, 1550 cm⁻¹; ¹H NMR (270 MHz, CDCl₃:CD₃OD 1:1): δ 0.89 (t, 6H, $J_{\text{Me.CH}_2}$ =6.8 Hz, 2MeCH₂), 1.27 (s, 52H, 26CH₂), 1.97, 2.03 (2s, 6H, 2AcN), 2.30 [m, 1H, H-3e(eq)], 5.45 (m, 1H, H-4 of

sphingosine), 5.70 (dt, 1H, $J_{4.5} = 14.5$, $J_{5.6} = J_{5.6}$; = 6.8 Hz, H-5 of sphingosine). The MS of **52** (negative ion mode) showed the base peak at m/z 1895 (M-Na).

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References

- 1. Komba, S.; Ishida, H.; Kiso, M.; Hasegawa, A. Carbohydr. Res. 1996, 285, c1.
- 2. Berg, E. L.; Robinson, M. R.; Mansson, O.; Butcher, E. C.; Magnani, J. L. *J. Biol. Chem.* **1991**, *265*, 14869.
- 3. Lowe, J. B.; Stoolman, L. M.; Nair, R. P.; Larsen, R. D.; Berhend, T. L.; Mark, M. Cell **1990**, *63*, 475.
- 4. Phillips, M. L.; Nudelman, E.; Gaeta, F. C. A.; Perez, M.; Singhal, A. K.; Hakomori, S.; Paulson, J. C. *Science* **1990**, 250, 1130.
- 5. Ticmeyer, M.; Swiedler, S. J.; Ishihara, M.; Moreland, M.; Schweigruber, H.; Hirtzer, P.; Bandley, B. K. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 1138.
- 6. Tyrrell, D.; James, P.; Rao, N.; Foxall, C.; Abbas, S.; Dasgupta, F.; Nashed, M.; Hasegawa, A.; Kiso, M.; Asa, D.; Kidd, J.; Brandley, B. K. *Proc. Natl Acad. Sci. USA* **1991**, *88*, 10372.
- 7. Walz, G.; Aruffo, A.; Kolanus, W.; Bevilacqua, M.; Seed, B. *Science* **1990**, *250*, 1132.
- 8. (a) Larsen, E.; Palabrica, T.; Sajer, S.; Gilbert, G. E.; Wagner, D. D.; Furie, B. C.; Furie, B. Cell 1990, 63, 467; (b) Polley, M. J.; Phillips, M. L.; Wayner, E.; Nudelman, E.; Singhal, A. K.; Hakomori, S.; Paulson, J. C. Proc. Natl Acad. Sci. USA 1991, 88, 6224.
- 9. Cooling, L. L. W.; Zhang, D.-S.; Waker, K. E.; Koerner, A. W. *Glycobiology* **1995**, *5*, 571.
- 10. Imai, Y.; True, D. D.; Singer, M. S.; Rosen, S. D. J. Cell Biol. 1990, 111, 1225.
- 11. Aruffo, A.; Kolanus, W.; Walz, G.; Fredman, P.; Seed, B. *Cell* **1991**, *67*, 35.
- 12. Asa, D.; Gant, T.; Oda, Y.; Brandley, B. K. Glycobiology 1992, 2, 395.
- 13. Needham, L. K.; Schnaar, R. L. *Proc. Natl Acad. Sci. USA* **1993**, *90*, 1359.
- 14. Green, P. J.; Tamatani, T.; Watanabe, T.; Miyasaka, M.; Hasegawa, A.; Kiso, M.; Yuen, C.-T.; Stoll, M. S.; Feizi, T. *Biochem. Biophys. Res. Commun.* 1992, 188, 244.
- 15. Brandley, B. K.; Kiso, M.; Abbas, S.; Nikrad, P.; Srivasatava, O.; Foxall, C.; Oda, Y.; Hasegawa, A. *Glycobiology* **1993**, *3*, 633.
- 16. Green, P. J.; Yuen, C.-T.; Childs, R. A.; Chai, W.; Miyasaka, M.; Lemoine, R.; Lubineau, A.; Smith, B.; Veno, H.; Nicolaou, K. C.; Feizi, T. *Glycobiology* **1995**, *5*, 29.
- 17. Ohmoto, H.; Nakamura, K.; Inoue, T.; Kondo, N.; Inoue, Y.; Yoshino, K.; Kondo, H.; Ishida, H.; Kiso, M.; Hasegawa, A. J. Med. Chem. 1996, 39, 1339.

- 18. Yuen, C.-T.; Lawson, A. M.; Chai, W.; Larkin, M.; Stoll, M. S.; Stuart, A. C.; Sullivan, X.; Ahern, T. J.; Feizi, T. *Biochemistry* **1992**, *31*, 9126.
- 19. Yuen, C.-T.; Bezouska, K.; O'Brien, J.; Stoll, M.; Lemonie, R.; Lubineau, A.; Kiso, M.; Hasegawa, A.; Bockovich, J.; Nicolaou, K. C.; Feizi, T. *J. Biol. Chem.* **1994**, 269, 1595.
- 20. (a) Kameyama, A.; Ishida, H.; Kiso, M.; Hasegawa, A. *Carbohydr. Res.* **1991**, 209, c1; (b) Kameyama, A.; Ishida, H.; Kiso, M.; Hasegawa, A. *J. Carbohydr. Chem.* **1991**, 10, 549 (c) Kameyama, A.; Ishida, H.; Kiso, M.; Hasegawa, A. *J. Carbohydr. Chem.* **1991**, 10, 729.
- 21. Hasegawa, A.; Ando, T.; Kameyama, A.; Kiso, M. Carbohydr. Res. 1992, 230, c1.
- 22. (a) Kameyama, A.; Ishida, H.; Kiso, M.; Hasegawa, A. *J. Carbohydr. Chem.* **1994**, *13*, 641; (b) Hotta, K.; Ito, K.; Kameyama, A.; Ishida, H.; Kiso, M.; Hasegawa, A. *J. Carbohydr. Chem.* **1995**, *14*, 115.
- 23. (a) Yoshida, M.; Uchimura, A.; Kiso, M.; Hasegawa, A. *Glycoconjugate J.* **1993**, *10*, 3; (b) Kiso, M.; Furui, H.; Ando, K.; Ishida, H.; Hasegawa, A. *Bioorg. Med. Chem.* **1994**, 2, 1295.
- 24. Hasegawa, A.; Uchimura, A.; Ishida, H.; Kiso, M. *Biosci. Biotech. Biochem.* 1995, 59, 1091.
- 25. (a) Hasegawa, A.; Ito, K.; Ishida, H.; Kiso, M. J. Carbohydr. Chem. 1995, 14, 353; (b) Maeda, H.; Ishida, H.; Kiso, M.; Hasegawa, A. J. Carbohydr. Chem. 1995, 14, 369; (c) Maeda, H.; Ito, K.; Ishida, H.; Kiso, M.; Hasegawa, A. J. Carbohydr. Chem. 1995, 14, 387; (d) Kameyama, A.; Ehara, T.; Yamada, Y.; Ishida, H.; Kiso, M.; Hasegawa, A. J. Carbohydr. Chem. 1995, 14, 507; (e) Hasegawa, A.; Kato, M.; Ando, T.; Ishida, H.; Kiso, M. Carbohydr. Res. 1995, 274, 165; (f) Terada, T.; Kiso, M.; Hasegawa, A. Carbohydr. Res. 1994, 259, 201.
- 26. (a) Komba, S.; Ishida, H.; Kiso, M.; Hasegawa, A. *Glycoconjugate J.* **1996**, *13*, 241; (b) Ehara, T.; Kameyama, A.; Yamada, Y.; Ishida, H.; Kiso, M.; Hasegawa, A. *Carbohydr. Res.* **1995**, *366*, **1996**, *281*, 237.
- 27. Imai, Y.; Lasky, L. A.; Rosen, S. D. Nature 1993, 361, 555.
- 28. Hemmerich, S.; Bertzzi, C. R.; Leffler, H.; Rosen, S. D. *Biochemistry* **1994**, *33*, 4820.
- 29. Hemmerich, S.; Rosen, S. D. Biochemistry 1994, 33, 4830.
- 30. Hemmerich, S.; Leffler, H.; Rosen, S. D. J. Biol. Chem. **1995**, 270, 12635.

- 31. (a) Scudder, P. R.; Shailubhai, K.; Duffin, K. L.; Streeter, P. R.; Jacob, G. S. *Glycobiology* **1994**, *4*, 929; (b) Vig, R.; Jain, R. K.; Piskorz, C. F.; Matta, K. L. *J. Chem. Soc., Chem. Commun.* **1995**, 2073; (c) Jain, R. K.; Vig, R.; Rampal, R.; Chandrasekaran, E. V.; Matta, K. L. *J. Am. Chem. Soc.* **1994**, *116*, 12123.
- 32. (a) Murase, T.; Ishida, H.; Kiso, M.; Hasegawa, A. *Carbohydr. Res.* **1988**, *184*, c1; (b) Hasegawa, A.; Ohki, H.; Nagahama, T.; Ishida, H.; Kiso, M. *Carbohydr. Res.* **1991**, *212*, 277; (c) Hasegawa, A.; Nagahama, T.; Ohki, H.; Hotta, K.; Ishida, H.; Kiso, M. *J. Carbohydr. Chem.* **1991**, *10*, 493; (d) Hasegawa, A.; Fushimi, K.; Ishida, H.; Kiso, M. *J. Carbohydr. Chem.* **1993**, *12*, 1203.
- 33. Jansson, K.; Ahlfors, S.; Frejd, T.; Kihlberg, J.; Magnusson, G.; Dahmen, J.; Noori, G.; Stenvall, K. J. Org. Chem. 1988, 53, 5629.
- 34. Marra, A; Sinaÿ, P. Carbohydr. Res. 1989, 187, 35.
- 35. (a) Veeneman, G. H.; van Leeuwen, S. H.; van Boom, J. H. *Tetrahedron Lett.* **1990**, *31*, 1131; (b) Konradsson, P.; Mootoo, D. R.; McDevitt, R. E.; Fraser-Reid, B. *J. Chem. Soc. Commun.* **1990**, *270*.
- 36. (a) Kanie, O.; Kiso, M.; Hasegawa, A. *J. Carbohydr. Chem.* **1988**, *7*, 501; (b) Okamoto, K.; Goto, T. *Tetrahedron* **1990**, *46*, 5835.
- 37. (a) Schmidt, R. R.; Michel, J. *Angew. Chem. Int. Ed. Engl.* **1980**, *19*, 732; (b) Murase, T.; Ishida, H.; Kiso, M.; Hasegawa, A. *Carbohydr. Res.* **1989**, *188*, 71.
- 38. Kameyama, A.; Ishida, H.; Kiso, M.; Hasegawa, A. *Carbohydr. Res.* **1990**, *200*, 269.
- 39. Classon, B.; Garegg, P. J.; Samuelsson, B. *Acta Chem. Scand. B* **1984**, *38*, 419; (b) Nilsson, M.; Norberg, T. *J. Carbohydr. Chem.* **1990**, *9*, 1.
- 40. Fügedi, P.; Garegg, P. J. Carbohydr. Res. 1986, 149, c9.
- 41. Schmidt, R. R.; Grundler, G. Synthesis 1981, 885.
- 42. (a) Ito, Y.; Kiso, M.; Hasegawa, A. J. Carbohydr. Chem. 1989, 8, 285; (b) Schmidt, R. R.; Zimmermann, P. Angew. Chem. Int. Ed. Engl. 1986, 25, 725.
- 43. Adachi, T.; Yamada, Y.; Inoue, I.; Saneyoshi, M. Synthesis 1977, 45.
- 44. Koeners, H. J.; Verhoeven, J.; van Boom, J. H. Rec. Trav. Chim. Pays-Bas 1981, 100, 65.
- 45. Foxall, C.; Watson, S. R.; Dowbenko, D.; Fennie, C.; Lasky, L. A.; Kiso, M.; Hasegawa, A.; Asa, D.; Brandley, B. K. J. Cell Biol. 1992, 117, 895.

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