



 International Edition:
 DOI: 10.1002/anie.201906297

 German Edition:
 DOI: 10.1002/ange.201906297

Establishment of Guidelines for the Control of Glycosylation Reactions and Intermediates by Quantitative Assessment of Reactivity

Chun-Wei Chang, Chia-Hui Wu, Mei-Huei Lin, Pin-Hsuan Liao, Chun-Chi Chang, Hsiao-Han Chuang, Su-Ching Lin, Sarah Lam, Ved Prakash Verma, Chao-Ping Hsu,* and Cheng-Chung Wang*

Abstract: Stereocontrolled chemical glycosylation remains a major challenge despite vast efforts reported over many decades and so far still mainly relies on trial and error. Now it is shown that the relative reactivity value (RRV) of thioglycosides is an indicator for revealing stereoselectivities according to four types of acceptors. Mechanistic studies show that the reaction is dominated by two distinct intermediates: glycosyl triflates and glycosyl halides from N-halosuccinimide (NXS)/TfOH. The formation of glycosyl halide is highly correlated with the production of α -glycoside. These findings enable glycosylation reactions to be foreseen by using RRVs as an α/β -selectivity indicator and guidelines and rules to be developed for stereocontrolled glycosylation.

Carbohydrates are essential biomolecules in living organisms.^[1] However, to access carbohydrate-based products in large quantities with well-defined regio- and stereochemistry is still challenging.^[2] To date, no unifying strategy for obtaining 1,2-*cis* glycosides exists as the glycosylation reaction outcome is influenced by numerous factors and the detail reaction mechanism is still unclear.^[3] Therefore, an indicator and a general guideline that predicts, foretells, and summarizes the stereoselectivity would greatly simplify this indispensible reaction and would facilitate advanced methods that streamline oligosaccharide synthesis, such as solid-phase automated oligosaccharide synthesis and one-pot glycosylation.^[4]

| [*] CW. Chang, Dr. CH. Wu, MH. Lin, PH. Liao, CC. Chang, HH. Chuang, Dr. SC. Lin, Dr. S. Lam, Dr. V. P. Verma, Dr. CP. Hsu Dr. CC. Wang | ١, |
|---|----|
| Institute of Chemistry, Academia Sinica | |
| Taipei 115 (Taiwan) | |
| E-mail: cherri@chem.sinica.edu.tw | |
| wangcc@chem.sinica.edu.tw | |
| CW. Chang, Dr. CH. Wu, Dr. CC. Wang | |
| Chemical Biology and Molecular Biophysics | |
| Taiwan International Graduate Program, Academia Sinica | |
| Taipei 115 (Taiwan) | |
| CW. Chang, Dr. CH. Wu, HH. Chuang | |
| Department of Chemistry, National Taiwan University | |
| Taipei 106 (Taiwan) | |
| HH. Chuang | |
| Nanoscience and Technology Program, Taiwan International Grad- uate Program, Academia Sinica and National Taiwan University Tainei 115 (Taiwan) | |
| Supporting information and the OPCID identification number(s) for | |
| b the author(s) of this article can be found under: | |

https://doi.org/10.1002/anie.201906297.

Angew. Chem. Int. Ed. 2019, 58, 1-6

© 2019 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

Wiley Online Library

These are not the final page numbers!



Thioglycosides 1 (Figure 1) are the most commonly used

glycosyl donors and can be easily activated chemoselectively

by electrophilic promoter systems, among which N-iodosuc-

Figure 1. RRV as a scale meter to define α/β stereoselectivity in chemical glycosylation.

cinimide (NIS, **6-I**)/TfOH is the most widely used.^[5] Ye, Huang, and Yoshida have developed a preactivation strategy that allows iterative one-pot glycosylation.^[3a,6] However, similar to all other glycosylation reactions, the poorly controlled stereoselectivity remains a long-standing problem.^[3c,5a] The promotor systems are generally a combination of a stoichiometric amount of an iodonium agent and a catalytic amount of TfOH. The reactions are complicated by the presence of various reactants, and glycosyl triflate **2** is commonly believed as the main intermediate, which leads to the mixture of products **5-** α and **5-** β .^[3d,7]

To predict glycosylation stereoselectivity and systematically analyze the factors that influence the outcome, an objective comparison system is required. Ley was the first to introduce deactivation factor (DF) to define the donor reactivity.^[8] Later, the relative reactivity value (RRV), a comprehensive quantification system developed by Wong, provides a quantitative assessment for programmable one-pot oligosaccharide synthesis.^[9] Herein, we show that RRVs serve as a parameter/indicator to statistically predict stereoselectivity in glycosidic bond formation and for the first time chemical glycosylation reaction can be correlated with a general statistical approach. Our mechanistic experiments revealed that glycosylations proceeded via a glycosyl triflate



Figure 2. Glycosyl donors with defined RRV.

2 and/or iodide 3-I, which we found is highly associated with the formation of α -glycoside

5- α , as a function of RRV.

To understand the correlation between the stereoselectivity and the identity of glycosyl donors and acceptors, the reactivity of a series of glycosyl donors were quantified by their RRVs and the stereoselectivity of their reaction with a range of glycosyl acceptors were studied. In this study, the RRVs of donors 7-22 on different sugars without C2 participating group (Figure 2; Supporting Infor-Figures S1–S9), mation, including 1,2-trans thiotolyl glucosides 7, 10, 11, 13,^[9a] 16 and galactosides 17, 18, **19**,^[9a] mannosides **12**^[9a, 10] and 15,^[9c] 2-azido-2-deoxyglucosides 8, 9, β-thiotolyl 2dexoygalactoside 20, 21, and α-thiotolyl 2-deoxyglucoside 14, 22,^[11] were measured. These values were determined by using competition experiments against

a least reactive donor, tolyl tetra-*O*-acetylthiomannoside **23**, of which the RRV is defined as 1.0, as reported.^[9a,b]

Next, donors 7–22 were pre-mixed with acceptors 24–27, respectively. Secondary glycosyl alcohols 24 and 25, primary glycosyl alcohol 26, and alkyl alcohol 27 are four types of commonly used glycosyl acceptors in carbohydrate synthesis. After testing the reactions from -78 to 0°C, we found the changes in stereoselectivity were very limited. Therefore, the reaction condition was fixed at -40°C in DCM to eliminate the possible variables, and 1.0 equiv of NIS and 0.4 equiv of TfOH were introduced to activate the donor in a traditional (non-preactivated) manner. The reaction mixture was stirred for 3 h, and the α/β ratio of the reaction was determined by using HPLC.

After screening these combinations (Figure 3; Supporting Information, Table S6), we interestingly found the α -selectivity is in roughly a linear correlation with log(RRV). For 4-OH acceptor **24** (Figure 3A), the linear regression showed that the slope is 16.6, in which R^2 is 0.77 and Pearson's *r* is 0.89. Consistent trends were observed with the other acceptors **25–27**, but in different slopes. For the other secondary alcohols, namely, 3-OH acceptor **25** (Figure 3B), with the increasing log(RRV) from 0–6, α -selectivity also rose linearly with a steep slope of 15.7. For primary 6-OH acceptor **26** (Figure 3C), the linear regression showed that the slope was 13.3. When using sterically least-hindered methanol (**27**) as acceptor (Figure 3D), α -selectivity was enhanced the least.

These findings suggest that stereoselectivity is more closely related to inductive effects on donors and the



Figure 3. α-Selectivity vs. donor reactivity (RRV). Using A) glucoside 4-OH **24**; B) glucoside 3-OH **25**; C) glucoside 6-OH **26**; D) MeOH (**27**) as the acceptors.

www.angewandte.org

2

© 2019 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

These are not the final page numbers!

reactivity of acceptors.^[3c,12] For example, mannosides are commonly believed to produce α -major (1.2-trans) glycosides. Thiomannosides 12 (RRV = 315, log(RRV) = 2.50) and 15 (RRV = 5000, log(RRV) = 3.70), having moderate RRVs, in fact gave considerable amounts of β -products (1.2-cis) with the α/β ratio predictable by the 4 linear fits in Figure 3, For O6-actylated glucose donor 7 (dots in dash box), the selectivity deviated from the trends for secondary acceptors 24 and 25 as observed in previous reports;^[13] but not for primary acceptors 26 and 27. Based on Figure 3, we herein provide a reliable guideline to design glycosyl donors for both α - and β -selective glycosylation reactions by using RRVs as indicators (Supporting Information, Figure S29). Moreover, stereoselectivity showed no significant change between preactivation and the traditional approach (non-preactivation).^[14]

We then tried to identify the intermediates before the addition of glycosyl acceptor by using low-temperature NMR experiments at -40 °C (Figure 4). Donors **7–22** were treated with 1.0 equiv of NIS and 0.4 equiv of TfOH in CD₂Cl₂ at -40 °C for 15 min individually. We initially expected to observe the signals of glycosyl triflate **2**, which was the most common believed intermediate from previous reports.^[3b, 15]



Figure 4. Intermediate distribution vs. donor reactivity (RRV). Using A) NIS/TfOH; B) NBS/TfOH; C) NCS/TfOH; D) ToISCI/AgOTf as the promotor.

Angew. Chem. Int. Ed. 2019, 58, 1-6

© 2019 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

www.angewandte.org

However, our NMR spectrum in turn showed the evidence of glycosyl iodide **3-I** intermediate formation for the first time in the TfOH promoted system (Supporting Information, Table S1, Figures S10–24). Furthermore, the existence of **3-I** in the reaction was confirmed by comparing the ¹H NMR spectrum obtained with that of the suspected intermediates synthesized independently through relevant procedures under TMSI conditions (Supporting Information, Figures S10–S24, Table S1).^[16] Moreover, **3-I** can be further identified by high-resolution ESI-MASS (Supporting Information, Figure S11).

The intermediate distribution of **3-I** (Figure 4A) is also inductive-effect-dependent and was highly correlated with α selectivity (Figure 3). RRV provides a universal system to predict the intermediate change in the scaffold of log(RRV) between 1.27 and 2.50 (or RRV between 18.6 and 315). Consistent results of the intermediate change were also observed in other halide-containing promotor systems such as NBS/TfOH (Figure 4B), NCS/TfOH (Figure 4C), and TolSCl/AgOTf (Figure 4D) with their glycosyl bromide **3-Br** and chloride **3-Cl** being observed as well (Supporting Information, Figures S10–S24, Table S1).

To further clarify the intermediate transformation, we performed a theoretical computational study. The transition states **TS** consisted of an oxocarbenium cation with a certain degree of association with both triflate anion (OTf⁻) and halide anion (X⁻). The conversion mechanism is most accurately described as an S_N 2-like process. Computational experiments all yielded consistent results in each NXS/TfOH system (Supporting Information, Figures S25–S28, Tables S2–S5). The relatively low free-energy barriers were 4.17 and 5.48 kcal mol⁻¹, respectively, when calculated with the temperature set at -40 °C (Supporting Information, Figure S25). These address the fact that facile intermediate transformation could happen to produce the glycosyl halide intermediate in situ.

Although the relevance between glycosyl intermediates and stereoselectivity is hard to define owing to the participation of solvent-separated ion pairs (SSIPs),[3d,17] and clearly, the electronic nature of the protecting groups bound to the glycosyl donors play a key role in shifting the $S_N 1-S_N 2$ reaction paradigm.^[17] We have found the correlation between donor reactivity (RRV) and the intermediate glycosyl halide/ triflate ratio, which in turn influences the α/β selectivity upon the addition of an acceptor. This is important, because the reaction stereoselectivity is highly dependent on the identity of the reactive intermediate as reported by Gervay-Hague, Field, and Codee.^[12,16,17b,c,18] The glycosyl iodide **3-I** can lead to distinct α -selectivity, and **3-I** can then be converted into an even more reactive β -counterpart **3-I-\beta** via a Lemieux anomerization pathway,^[19] which undergoes an S_N 2-like reaction with a nucleophilic acceptor from the bottom side for selective α -glycosyl bond formation.^[16,18] In contrast, the α -triflate intermediate **2** favors the S_N2 reaction and β selective reaction with the acceptor in the cases of the unreactive donors.^[12, 17b,c] Since both preactivation and nonpreactivation showed very similar stereoselectivity (Figure 3), it implied that glycosides formation occurs through glycosyl iodide **3-I** after facile intermediate transformation at a low temperature within the donor activation timeframe.

We targeted numerous disaccharides or glycoconjugates (Scheme 1; Supporting Information, Table S6). The coupling between glycan and linkers **34** and **35** is useful and common in studies on glycosciences.^[4a] Clearly indicated by our RRV



Scheme 1. Stereoselectivity-predictable glycosylation using RRV as the indicator.

indicating system to synthesize alkyl glycoside **31** (Figures 3D), for **18** (RRV of 7180, $\log(\text{RRV}) = 3.86$), product **36** was obtained only with poor α/β selectivity (α -selectivity = 33%) when the linker acceptor **35** was treated. According to Figures 3D, we then adopted a partially protected donor **33** with lower RRV of 104 ($\log(\text{RRV}) = 2.02$) (Scheme 1A). Without the electron-donating effect on *C*2 and *C*3, the unreactive **33** resulted in high β -selectivity with linkers **34** and **35** individually, in which **37** (α -selectivity = 15%) and **38** (α -selectivity = 0%) were isolated.

A prediction based on secondary alcohols in Figures 3 B was noted for donors **22**, **13**, and **39** with the secondary threonine **40** as the acceptor (Scheme 1 B). As pinpointed by the linear fit, 80% α -selectivity was observed on 2-deoxy-glucosyl donor **22** (log(RRV) = 6). Next, for tetra-*O*-benzy-lated thioglucoside **13** (log(RRV) = 3.42), the selectivity of **42** decreased to 66% α -selectivity. Eventually, for 2-azido-2-deoxy thioglucoside **39** (log(RRV) = 2.44, RRV = 270),^[9e] product **43** was obtained with the poorest selectivity (50% α -selectivity) in 75%.

Finally, we aimed at an α -Gal-(1 \rightarrow 4)- β -GlcNA disaccharide motif **48** (Scheme 1 C), equipped with a β -pentyl linker at the reducing end. First, a semi-protected glucosamine donor **44** with a low RRV of 151 was designed for β -selective glycosylation. Predicted well by our system (α selectivity = 21% in Figures 3D), a majority of β -glycoside **46** (α selectivity = 22%) was furnished in 81%. Next, a Gal donor **47** with higher RRV of 9153 gave a high α -selectivity and produced **48** in 70%.

Stereocontrolled glycosylation was the main challenge in carbohydrate chemistry owning to the lack of a clear mechanism and too many control variables.^[3c] We herein first demonstrated that stereoselectivity can be predicted using reactivity of donors (RRVs) on different acceptors in a NIS/ TfOH system. Our studies also revealed that the reaction was dominated by two distinct intermediates, including glycosyl triflate and iodide. A further understanding of mechanistic pathways is in progress. We believe the behavior of donors can be further clarified, and our indicator and guideline can provide a new solution for chemists to simplify glycosylation reaction and design building blocks in a quantifiable manner. This discovery gives new insight to stereocontrolled glycosylation for carbohydrate/organic chemistry and presents a multidisciplinary direction that combines statistics. Since anomeric selectivity is simultaneously influenced by remote functional group participation,^[3a,20] acceptor nucleophilicity, solvent polarity, temperature, and activation system.^[3b,11,12,17b,c,21] we are studying these permanent and environmental variables by using RRV as the analytic tools. Similar trends can also be observed when using TolSCI/ AgOTf and BSP/Tf₂O combinations as the promoters. More types of building blocks, such as L-6-deoxy sugars and glycuronic acids, will be included in our future studies. The incorporation of this system into artificial intelligence for carbohydrate synthesis is now undergoing.^[22]

Acknowledgements

We thank Dr. Tsyr-Yan Yu, Dr. Shang-Te Danny Hsu (Academia Sinica), Prof. Yasuhiro Kajihara, and Yuta Maki (Osaka University) for helpful discussion and Ping-Yu Lin (Academia Sinica) for Mass measurement; This work was supported by the Ministry of Science and Technology, Taiwan (MOST 108-2133-M-001-019-; 106–2113-M-001-009-MY2) and Academia Sinica (MOST 108-3114-Y-001-002; AS-SUMMIT-108).

Conflict of interest

The authors declare no conflict of interest.

Keywords: carbohydrates · diastereoselectivity · glycosylation

© 2019 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

These are not the final page numbers!

^[1] C.-Y. Wu, C.-H. Wong, Chem. Commun. 2011, 47, 6201-6207.

- [2] a) M. M. Nielsen, C. M. Pedersen, *Chem. Rev.* 2018, *118*, 8285–8358; b) T. J. Wadzinski, A. Steinauer, L. Hie, G. Pelletier, A. Schepartz, S. J. Miller, *Nat. Chem.* 2018, *10*, 644–652; c) Y. Yang, X. Zhang, B. Yu, *Nat. Prod. Rep.* 2015, *32*, 1331–1355.
- [3] a) W.-L. Leng, H. Yao, J.-X. He, X.-W. Liu, Acc. Chem. Res. 2018, 51, 628–639; b) S. K. Mulani, W.-C. Hung, A. B. Ingle, K.-S. Shiau, K.-K. T. Mong, Org. Biomol. Chem. 2014, 12, 1184–1197; c) S. Chatterjee, S. Moon, F. Hentschel, K. Gilmore, P. H. Seeberger, J. Am. Chem. Soc. 2018, 140, 11942–11953; d) P. O. Adero, H. Amarasekara, P. Wen, L. Bohé, D. Crich, Chem. Rev. 2018, 118, 8242–8284.
- [4] a) M. Panza, S. G. Pistorio, K. J. Stine, A. V. Demchenko, *Chem. Rev.* 2018, *118*, 8105–8150; b) S. S. Kulkarni, C.-C. Wang, N. M. Sabbavarapu, A. R. Podilapu, P.-H. Liao, S.-C. Hung, *Chem. Rev.* 2018, *118*, 8025–8104.
- [5] a) G. Lian, X. Zhang, B. Yu, *Carbohydr. Res.* 2015, 403, 13–22;
 b) R. Das, B. Mukhopadhyay, *ChemistryOpen* 2016, 5, 401–433.
- [6] a) S. Yamago, T. Yamada, T. Maruyama, J. Yoshida, Angew. Chem. Int. Ed. 2004, 43, 2145–2148; Angew. Chem. 2004, 116, 2197–2200; b) B. Sun, B. Srinivasan, X. Huang, Chem. Eur. J. 2008, 14, 7072–7081.
- [7] M. Huang, G. E. Garrett, N. Birlirakis, L. Bohé, D. A. Pratt, D. Crich, *Nat. Chem.* **2012**, *4*, 663–667.
- [8] N. L. Douglas, S. V. Ley, U. Lücking, S. L. Warriner, J. Chem. Soc. Perkin Trans. 1 1998, 51–66.
- [9] a) Z. Zhang, I. R. Ollmann, X.-S. Ye, R. Wischnat, T. Baasov, C.-H. Wong, J. Am. Chem. Soc. 1999, 121, 734–753; b) C.-W. Cheng, Y. Zhou, W.-H. Pan, S. Dey, C.-Y. Wu, W.-L. Hsu, C.-H. Wong, Nat. Commun. 2018, 9, 5202; c) X.-S. Ye, C.-H. Wong, J. Org. Chem. 2000, 65, 2410–2431; d) K.-K. T. Mong, C.-H. Wong, Angew. Chem. Int. Ed. 2002, 41, 4087–4090; Angew. Chem. 2002, 114, 4261–4264; e) T. K. Ritter, K.-K. T. Mong, H. Liu, T. Nakatani, C.-H. Wong, Angew. Chem. Int. Ed. 2003, 42, 4657–4660; Angew. Chem. 2003, 115, 4805–4808.
- [10] K. M. Koeller, C.-H. Wong, Chem. Rev. 2000, 100, 4465-4494.
- [11] J.-H. Chen, J.-H. Ruei, K.-K. T. Mong, Eur. J. Org. Chem. 2014, 1827–1831.
- [12] S. van der Vorm, J. M. A. van Hengst, M. Bakker, H. S. Overkleeft, G. A. van der Marel, J. D. C. Codée, *Angew. Chem. Int. Ed.* 2018, *57*, 8240–8244; *Angew. Chem.* 2018, *130*, 8372–8376.
- [13] a) J. Kalikanda, Z. Li, J. Org. Chem. 2011, 76, 5207–5218; b) G. Ngoje, Z. Li, Org. Biomol. Chem. 2013, 11, 1879–1886.
- [14] a) J. D. C. Codée, R. E. J. N. Litjens, L. J. van den Bos, H. S. Overkleeft, G. A. van der Marel, *Chem. Soc. Rev.* 2005, *34*, 769 –

782; b) W. Yang, B. Yang, S. Ramadan, X. Huang, *Beilstein J. Org. Chem.* **2017**, *13*, 2094–2114.

- [15] a) P. Konradsson, U. E. Udodong, B. Fraser-Reid, *Tetrahedron Lett.* 1990, *31*, 4313–4316; b) M. Heuckendorff, P. S. Bols, C. B. Barry, T. G. Frihed, C. M. Pedersen, M. Bols, *Chem. Commun.* 2015, *51*, 13283–13285; c) M. Mazur, B. Barycza, H. Andriamboavonjy, S. Lavoie, M. Tamigney Kenfack, A. Laroussarie, Y. Blériot, C. Gauthier, *J. Org. Chem.* 2016, *81*, 10585–10599.
- [16] J. Gervay-Hague, Acc. Chem. Res. 2016, 49, 35-47.
- [17] a) T. Hansen, L. Lebedel, W. A. Remmerswaal, S. van der Vorm, D. P. A. Wander, M. Somers, H. S. Overkleeft, D. V. Filippov, J. Desire, A. Mingot, Y. Bleriot, G. A. van der Marel, S. Thibaudeau, J. D. C. Codee, ACS Cent. Sci. 2019, 5, 781–788; b) S. van der Vorm, T. Hansen, H. S. Overkleeft, G. A. van der Marel, J. D. C. Codée, Chem. Sci. 2017, 8, 1867–1875; c) S. van der Vorm, H. S. Overkleeft, G. A. van der Marel, J. D. C. Codée, J. Org. Chem. 2017, 82, 4793–4811.
- [18] a) W. Du, J. Gervay-Hague, Org. Lett. 2005, 7, 2063–2065; b) R. van Well, K. Ravindranathan Kartha, R. Field, J. Carbohydr. Chem. 2005, 24, 463–474; c) H. W. Hsieh, M. W. Schombs, J. Gervay-Hague, J. Org. Chem. 2014, 79, 1736–1748; d) J.-C. Hu, A.-F. W. Feng, B.-Y. Chang, C.-H. Lin, K.-K. T. Mong, Org. Biomol. Chem. 2017, 15, 5345–5356; e) L. Wang, H. S. Overkleeft, G. A. van der Marel, J. D. C. Codée, J. Am. Chem. Soc. 2018, 140, 4632–4638.
- [19] R. U. Lemieux, J.-I. Hayami, Can. J. Chem. 1965, 43, 2162-2173.
- [20] a) D. Yao, Y. Liu, S. Yan, Y. Li, C. Hu, N. Ding, *Chem. Commun.* 2017, *53*, 2986–2989; b) D. Crich, T. Hu, F. Cai, *J. Org. Chem.* 2008, *73*, 8942–8953; c) R. A. Mensink, T. J. Boltje, *Chem. Eur. J.* 2017, *23*, 17637–17653.
- [21] A. B. Ingle, C.-S. Chao, W.-C. Hung, K.-K. T. Mong, Org. Lett. 2013, 15, 5290-5293.
- [22] a) J. M. Granda, L. Donina, V. Dragone, D. L. Long, L. Cronin, *Nature* 2018, 559, 377–381; b) J. W. Lehmann, D. J. Blair, M. D.
 Burke, *Nat. Rev. Chem.* 2018, 2, 0115; c) M. H. S. Segler, M.
 Preuss, M. P. Waller, *Nature* 2018, 555, 604–610.

Manuscript received: May 21, 2019 Revised manuscript received: August 23, 2019 Accepted manuscript online: September 13, 2019 Version of record online:







Communications

Glycosylation

C.-W. Chang, C.-H. Wu, M.-H. Lin, P.-H. Liao, C.-C. Chang, H.-H. Chuang, S.-C. Lin, S. Lam, V. P. Verma, C.-P. Hsu,* C.-C. Wang*

Establishment of Guidelines for the Control of Glycosylation Reactions and Intermediates by Quantitative Assessment of Reactivity



Selectivity rev-counter: The relative reactivity value (RRV) of thioglycosides is an indicator for revealing stereoselectivities for chemical glycosylation.

6 www.angewandte.org

@ 2019 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

Angew. Chem. Int. Ed. 2019, 58, 1-6

These are not the final page numbers!