



A novel type of acyclic nucleoside phosphonates derived from 2-(phosphonomethoxy)propanoic acid

Martin Maxmilian Kaiser, Petr Jansa, Martin Dračinský, Zlatko Janeba *

Institute of Organic Chemistry and Biochemistry, Academy of Sciences of the Czech Republic, v.v.i., Flemingovo nám. 2, CZ-16610 Prague 6, Czech Republic

ARTICLE INFO

Article history:

Received 10 October 2011

Received in revised form 27 February 2012

Accepted 19 March 2012

Available online 26 March 2012

Keywords:

Acyclic nucleoside phosphonates

CPMEA

HPMPA

PMEA

Oxidation

TEMPO

Microwave

Antiviral

ABSTRACT

A convenient and efficient synthesis of a novel class of acyclic nucleoside phosphonates derived from 2-(phosphonomethoxy)propanoic acid has been developed. The key step of the synthesis is the optimized oxidation of the 3-hydroxy-2-(phosphonomethoxy)propyl (HPMP) analogues to the corresponding 2'-carboxy-PME (CPME) derivatives using the TEMPO/NaClO₂/NaClO oxidizing system. Although (S)-3-(adenin-9-yl)-2-(phosphonomethoxy)propanoic acid ((S)-CPMEA) has been designed as a compound with potential anti-HIV activity, none of the newly prepared CPME analogues exhibited any antiviral activity.

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1. Introduction

Acyclic nucleoside phosphonates (ANPs) constitute a distinguished class of antiviral agents.¹ They also exhibit important cytostatic,² antiparasitic³ and immunomodulatory activities.⁴ The main advantages of ANPs are their resistance towards degradation by enzymes in vivo, and the ability to by-pass the first phosphorylation step necessary for the activation of the bioactive nucleosides.⁵ Three ANPs have been approved worldwide for clinical use:⁵ HPMPA (cidofovir, Vistide®) for the treatment of CMV retinitis in AIDS patients,^{5,6} PMEA (**1**, Fig. 1) (as its oral prodrug adefovir dipivoxil, Hepsera®) for the treatment of chronic HBV infections,⁵ and PMPA (**2**, Fig. 1) (as its oral prodrug tenofovir disoproxil fumarate, Viread®) for the treatment of HIV (itself or in combination with other antivirals) and chronic HBV infections.⁵ Many other ANPs were found to exhibit important antiviral properties, e.g., derivatives of 2,6-diaminopurine (HPMPDAP, PMEDAP, PMPDAP)^{1f} and 'open ring' derivatives of 2,4-diaminopyrimidine (HPMPO-DAPy, PMEO-DAPy, PMPO-DAPy).^{1f,7} Like nucleotides,⁸ due to their poor bioavailability, all ANPs have to be administered in the form of their prodrugs (or by injection in the case of cidofovir).⁹

Analogues containing adenine play a special role among ANPs for their potent antiviral properties. HPMPA (**3**, Fig. 1),^{1a} the

prototype of ANPs derived from (S)-DHPA (**5**), possesses broad-spectrum activity against DNA viruses (including herpes-, adeno-, pox- and iridoviruses), as well as against retroviruses. While only the (S)-HPMP enantiomer **3** is active in the adenine series, its 3'-O-phosphonomethyl regioisomer (iso-HPMPA)¹⁰ exhibits no antiviral activity.^{1a} The approved adenine analogues PMEA (**1**) and PMPA (**2**) are primarily active against hepadna- and retroviruses, while PMEA (**1**) also exhibits notable activity against herpesviruses.⁵ Finally, FPMPA (**4**), which proved to be less antivirally effective, but also less toxic than PMEA (**1**), was thoroughly studied for its antiretroviral activity but was not pursued further to the clinical use.¹¹

A number of other adenine derivatives proved to have important antiviral activities.^{1f} Acyclic nucleoside analogues like (S)-DHPA (**5**, Fig. 1), marketed in Czechoslovakia as Duvira® gel,¹² or structurally related D-eritadenine¹³ and 3-(adenin-9-yl)-2-hydroxypropanoic acid (**6**, AHPA)¹⁴ constitute another class of broad-spectrum antivirals. These acyclic analogues of adenosine are targeted at S-adenosyl-L-homocysteine hydrolase,¹⁵ and thus at the capping process (methylation of 5'-end-guanine of viral mRNA).¹⁶

Herein we describe the synthesis and biological properties of the novel type of acyclic nucleoside phosphonates structurally related to all above-mentioned biologically important compounds. The novel type of ANPs is derived from 2-(phosphonomethoxy)propanoic acid, with the adenine derivative (S)-2'-carboxy-PMEA (**7a**, (S)-CPMEA, Fig. 1) being the most promising candidate with potential biological properties.

* Corresponding author. Tel.: +420 220183143; fax: +420 220183560; e-mail addresses: janeba@uochb.cas.cz, zjaneba@yahoo.com (Z. Janeba).

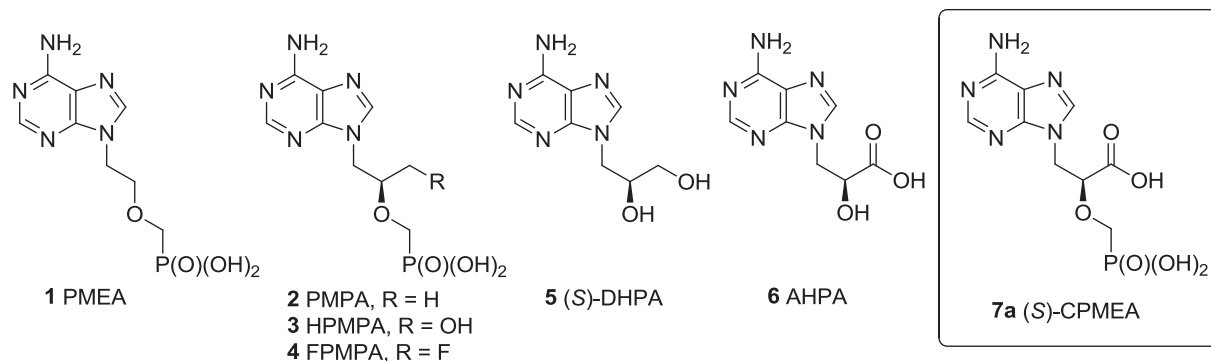


Fig. 1. Examples of biologically important acyclic nucleoside and nucleotide analogues **1–6** and structure of the target compound **7a**.

2. Results and discussion

The X-ray structure of HIV-1 RT–DNA complex after incorporation of tenofovir diphosphate is an ideal starting point for further rational and targeted drug design (Fig. 2a).¹⁷ In the complex, two amino acid residues (Arg 72 and Gln 151) in the binding site of the HIV-1 RT are oriented towards the C-2' methyl group of tenofovir diphosphate (Fig. 2a). Apparent incapability of the C-2' methyl group to substantially interact with the two polar amino acids of the HIV-1 RT led us to reason that replacement of the C-2' methyl with a polar group able to interact with Arg 72 and/or Gln 151 would increase the binding affinity of such an inhibitor to HIV-1 RT. Our docking studies indicated that replacement of the C-2' methyl group of tenofovir diphosphate (Fig. 2a) with a carboxyl group could, theoretically, lead to formation of up-to four new hydrogen bonds with arginine 72 and glutamine 151 (Fig. 2b).

The initial docking studies indicated that (S)-CPMEA (**7a**, Fig. 1), an analogue of PMEa (**1**) bearing a carboxyl group at the C-2' position of the aliphatic chain, potentially is a good candidate for strong binding affinity in the enzyme pocket of HIV-1 RT. Moreover, (S)-CPMEA (**7a**) can be considered to be a direct structural modification of PMPA (**2**) or HPMPA (**3**) with oxidized 2'-methyl or 2'-hydroxymethyl functions, respectively, or can also be looked upon as an O-phosphonomethyl derivative of AHPA (**6**, Fig. 1). Thus, compound **7a**, as well as the other CPME analogues, was expected to display a wide range of biological properties.

Generally, two strategies could be utilized for the synthesis of the target analogue, (S)-CPMEA (**7a**, Fig. 1): (a) direct oxidation of

the properly protected HPMPA followed by the removal of the protecting groups and (b) alkylation of adenine (or 6-chloropurine) with a suitable alkylating agent containing the protected carboxylic function, followed by deprotection (and ammonolysis in the case of 6-chloropurine).

The oxidation of the protected HPMPA derivative seems to be the more relevant and straightforward method for the synthesis of the compound **7a**, while the 'synthon' approach seems to be too laborious and less effective. Thus, diisopropyl (S)-9-[3-hydroxy-2-(phosphonomethoxy)propyl]adenine (**8a**, Scheme 1) was chosen as the suitable starting material. This can be prepared in sufficient amounts according to the previously described procedures.¹⁸

Nowadays, there are a considerable number of oxidizing agents, which can be used under relatively mild reaction conditions and which tolerate number of functional groups.¹⁹ Our goal was to develop a simple, mild and clean oxidation of the HPMPA derivative **8a**, which would tolerate the presence of the unprotected amino group at the C-6 position of the purine moiety.

The conversion of the 5'-hydroxyl group of 2',3'-protected purine nucleosides to the corresponding 5'-uronic acids is often accomplished by oxidation with an excess of KMnO_4 in aqueous KOH solution under conditions of high dilution.²⁰ An attempted oxidation of compound **8a** under analogous reaction conditions only led to complex reaction mixtures (TLC).

A simple and efficient oxidative conversion of primary alcohols to the corresponding methyl esters using $\text{I}_2/\text{K}_2\text{CO}_3$ in MeOH has been reported by Mori and Togo.²¹ Nevertheless, an attempt to prepare the methyl ester of derivative **9a** by the direct oxidative

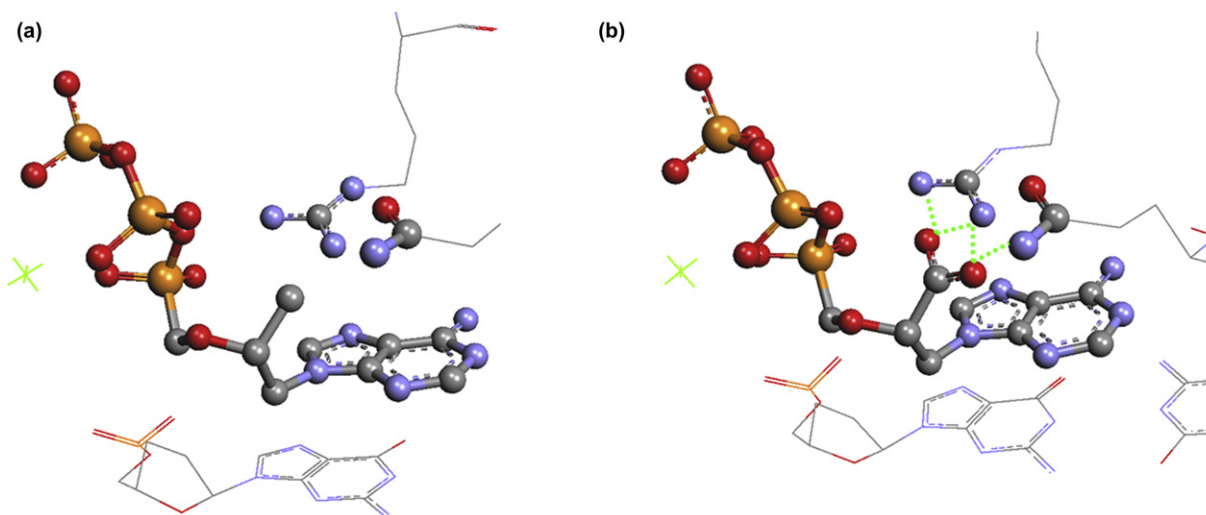
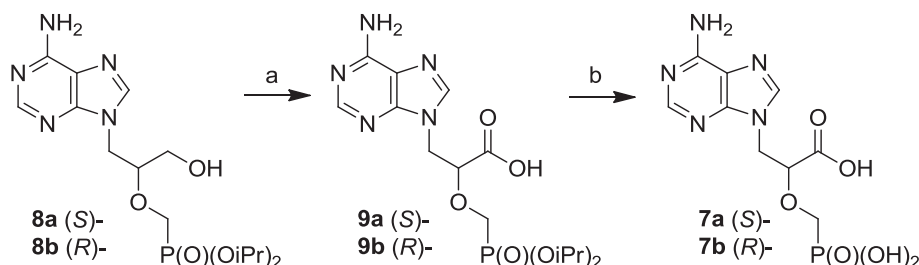


Fig. 2. (a) Immediate binding site of the complex of HIV-1 RT–DNA and tenofovir diphosphate. (b) Docking of the complex of HIV-1 RT–DNA and (S)-CPMEA (**7a**) showing possible hydrogen bonding (dotted green lines) between the carboxyl group of (S)-CPMEA diphosphate and Arg 72 and Gln 151.



Reagents and conditions: a) see table 1; b) Me₃SiBr, MeCN, rt, overnight.

Scheme 1. Synthesis of (S)-CPMEA (**7a**) and (R)-CPMEA (**7b**) through oxidation of the HPMPA derivatives **8a** and **8b**.

esterification of compound **8a** under the reported reaction conditions was not successful (Table 1).

Table 1
Oxidation of diisopropyl HPMPA (**8a**) to diisopropyl (S)-CPMEA (**9a**)

Oxidative agent; solvent	Desired product	Isolated yield
KMnO ₄ /KOH; H ₂ O	9a	Complex mixture
I ₂ /K ₂ CO ₃ ; MeOH	Methyl ester of 9a	Complex mixture
RuO ₂ /NaIO ₄ ; MeCN, CH ₂ Cl ₂ , H ₂ O	9a	63%
TEMPO/BAIB; MeCN, H ₂ O	9a	71%
TEMPO/NaClO ₂ /NaClO; MeCN, H ₂ O	9a	87%

Ruthenium tetroxide (RuO₄), another widely used oxidizing agent,¹⁹ is ideal when a very vigorous oxidizing agent is needed but mild reaction conditions must be maintained. Since RuO₄ can decompose explosively, it is mostly prepared in situ by oxidation of RuCl₃ or RuO₂. A mild oxidation of nucleosides to the corresponding 5'-carboxylic acids by use of RuCl₃/NaIO₄ and RuCl₃/K₂S₂O₈/KOH systems has been reported.²² Since RuCl₃ was found to be a very hygroscopic reagent, not easy to handle, RuO₂ was our oxidizing agent of choice.

Oxidation of diisopropyl HPMPA (**8a**) with RuO₂/NaIO₄ afforded the derivative **9a** in a satisfactory yield (63%, Table 1). The oxidation was carried out in a mixture of acetonitrile, chloroform, and water (1:1:2) according to the described procedure used for the preparation of adenosine 5'-uronamides,²³ with an addition of concentrated HCl to pH 2.5. Despite the good isolated yield, purification was tedious and column chromatography had to be repeated several times to obtain pure (S)-CPMEA derivative **9a**, probably owing to the fact that phosphonates strongly bind metal ions. Thus, a cleaner and less laborious procedure was still desirable.

Commercially available 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO) is a reagent of choice for oxidation of various organic substrates.²⁴ TEMPO belongs to the class of stable organic nitroxyl radicals commonly used for the oxidation of primary and secondary alcohols.^{19,24}

Mild and efficient oxidation of alcohols to ketones and aldehydes using catalytic amounts of TEMPO and stoichiometric amounts of [bis(acetoxy)iodo]benzene (BAIB) has been developed.²⁵ This method differs from most other TEMPO-mediated oxidations in that it generates acetic acid and iodobenzene as by-products and avoids inorganic contaminants.²⁵ This oxidizing system was successfully employed for the oxidation of 2',3'-protected nucleosides to give the corresponding 5'-carboxylic acids.²⁶ In our case, oxidation of HPMPA derivative **8a** with the TEMPO/BAIB oxidizing system afforded the expected product **9a** in a 71% yield (Table 1).

An even better yield (87%) of the (S)-CPMEA derivative **9a** was achieved when BAIB was replaced with NaClO₂/NaClO (sodium hypochlorite is a readily available and inexpensive oxidant, household bleach). This oxidizing system (TEMPO/NaClO₂/NaClO)²⁷

offers a clean reaction and easy work-up based on the isolation of the product by flash chromatography on silica gel followed by crystallization. This efficient procedure offered compound **9a** in very high purity.

Finally, the desired product (S)-CPMEA (**7a**) was obtained in a 76% yield by ester cleavage of compound **9a** under the standard conditions (Me₃SiBr in MeCN).²⁸

Analogously, oxidation of enantiomeric diisopropyl (R)-HPMPA (**8b**) using the TEMPO/NaClO₂/NaClO system afforded the derivative **9b** in a 70% yield. Subsequent deprotection of compound **9b** gave the free phosphonic acid **7b** in a 50% yield.

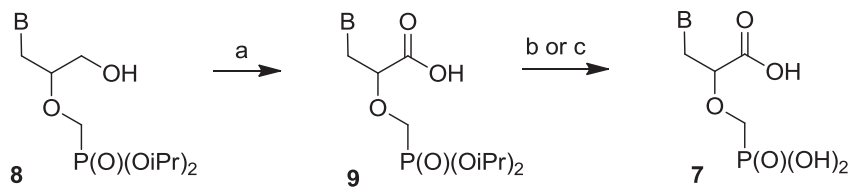
Unfortunately, none of the oxidizing systems listed in Table 1 afforded (S)-CPMEA (**7a**) by direct oxidation of free HPMPA (**3**, Fig. 1), confirming the importance of the diester protection of the phosphonate group during the procedure, and synthetic transformations in general.

Eventually, we decided to verify the more general applicability of our optimized oxidation protocol, and it was utilized for the preparation of CPME derivatives bearing other nucleobases (Scheme 2, Table 2). Thus, the expected diisopropyl CPME analogues **9** were obtained by the oxidation of the corresponding HPMPA derivatives **8** with TEMPO/NaClO₂/NaClO in high yields (76–87%).

The functional groups of the unprotected nucleobases are usually very well tolerated during the oxidation process. The only problem was observed with compounds bearing 2-aminopurine bases containing a guanidine motif (guanine and 2,6-diaminopurine). Treatment of the HPMPG derivative **8g**^{18b} and HPMPDAP derivative **8h**^{18b} with TEMPO/NaClO₂/NaClO gave complex and colourful reaction mixtures and no desired products were isolated (entries 7 and 8, respectively, Table 2).

To overcome the undesirable reactivity of the 2-aminopurine bases, we decided to protect the amino groups of compounds **8g** and **8h** with the benzoyl group. Thus, the N²-benzoylguanine derivative **8i**^{18b} was prepared in 49% yield from compound **8g** in pyridine by sequential treatment with Me₃SiCl, BzCl, water and aqueous ammonia (Scheme 3). Subsequent oxidation of the benzoyl derivative **8i** with TEMPO/NaClO₂/NaClO afforded the carboxy analogue **9i** in 78% yield (Table 2, entry 9). Direct deprotection of the phosphonate moiety of compound **9i** using Me₃SiBr in acetonitrile²⁸ afforded the N²-benzoylguanine derivative **7i** in 52% yield (Table 2, entry 9, Scheme 3). In order to obtain CPMEG (**7g**), the benzoyl group of derivative **9i** was removed first, using MeONa solution, followed by the removal of the isopropyl groups from the intermediate **9g** (Scheme 3).

Analogously, the HPMPDAP analogue **8h** was protected as its dibenzoyl derivative **8j** (in 47% yield), which was then efficiently oxidized using the TEMPO/NaClO₂/NaClO system to compound **9j** in 80% yield (Table 2, entry 10, Scheme 4). Finally, CPMEDAP (**7h**) was obtained in 38% yield from compound **9j** by simultaneous hydrolysis of both benzoyl groups and diisopropyl esters using 1 M



Reagents and conditions: a) TEMPO, NaClO₂, NaClO, MeCN, buffer, rt, ~24 h;

b) Me₃SiBr, MeCN, rt, overnight; c) HCl, H₂O, MW, 140 °C, 30 min.

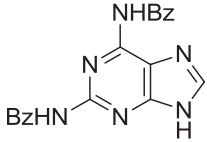
Scheme 2. Synthesis of CPME compounds **7** starting from the HPMP analogues **8**.

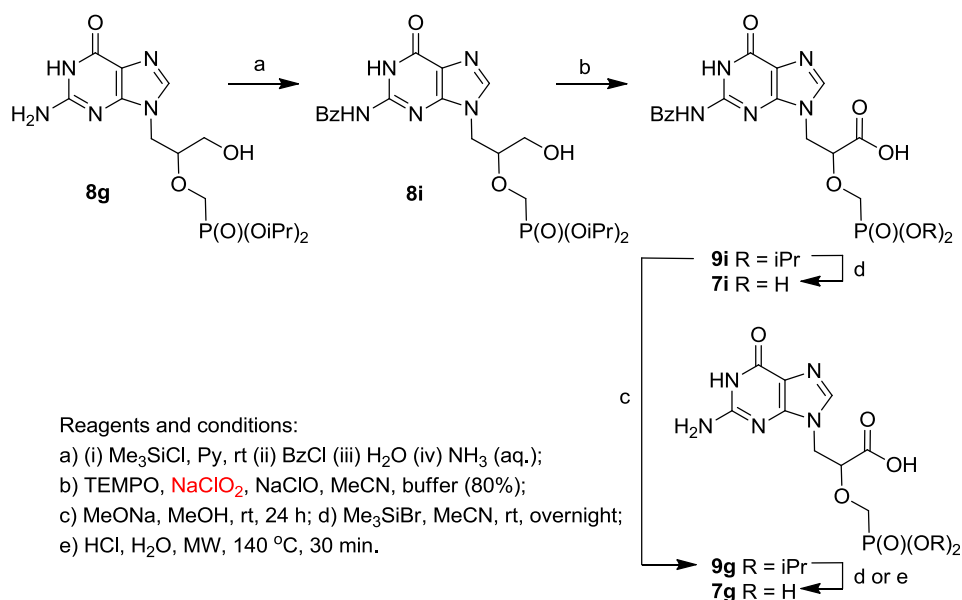
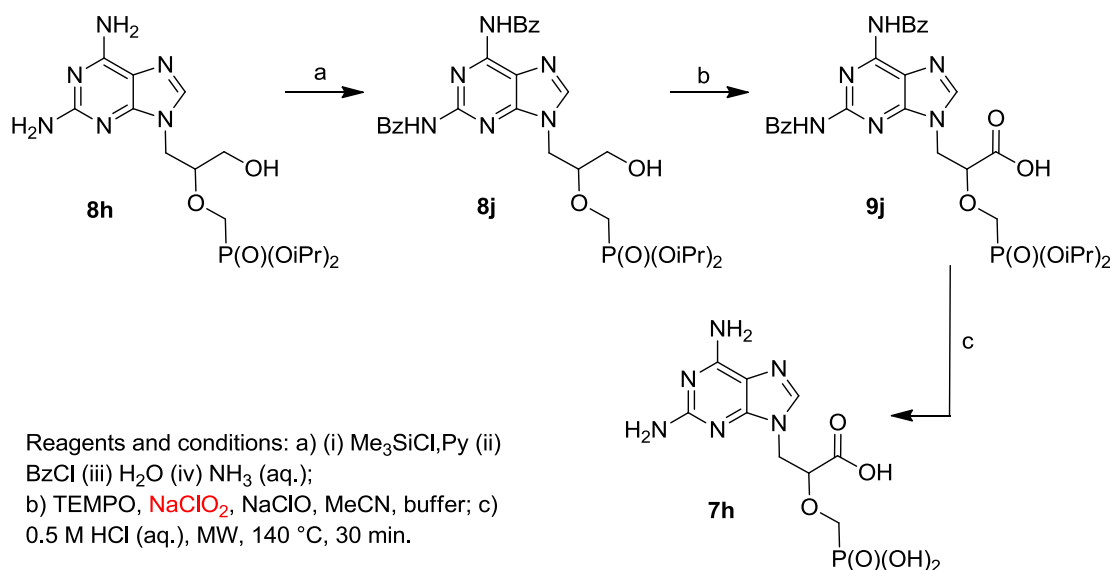
Table 2

Oxidation of the HPMP analogues **8** with TEMPO/NaClO₂/NaClO to give **9**, followed by diester removal to yield **7**

Entry	Nucleobase B	Starting compound	Product (yield) ^a of oxidation	Product (yield) ^a of the phosphonate deprotection
1		8a (S)-	9a (87%)	7a (68%), ^b (46%) ^c
2		8b (R)-	9b (76%)	7b (50%) ^b
3		8c (S)-	9c (83%)	7c (62%), ^b (39%) ^c
4		8d (S)-	9d (82%)	7d ^d (64%), ^b (44%) ^c
5		8e (S)-	9e (81%)	7e (42%) ^c
6		8f (R)-	9f (80%)	7f (48%) ^c
7		8g (S)-	Complex mixture (no 9g isolated) ^e	7g (61%), ^b (37%) ^c
8		8h (S)-	Complex mixture (no 9h isolated)	7h (38% from 9j) ^{c,f}
9		8i (S)-	9i (78%)	7i (52%) ^b

Table 2 (continued)

Entry	Nucleobase B	Starting compound	Product (yield) ^d of oxidation	Product (yield) ^d of the phosphonate deprotection
10		8j (S)-	9j (80%)	N.A. ^g

^a Isolated yield.^b Method A.^c Method B.^d Compound **7d** contains thymine as a base.^e Compound **9g** prepared from **9i** (Scheme 3).^f Compound **7h** prepared from **9j** (Scheme 4).^g N.A. not applicable.Scheme 3. Synthesis of CPMEG (**7g**) using TEMPO/ NaClO_2 / NaClO as a key step.Scheme 4. Synthesis of CPMEDAP (**7h**) using TEMPO/ NaClO_2 / NaClO as a key step.

aqueous HCl under microwave irradiation (Scheme 4), a general method for the convenient deprotection of phosphonate diesters recently developed in our laboratory.²⁹

All of the prepared diisopropyl CPME phosphonates **9** were deprotected to the final free phosphonic acids **7** (Table 2, Scheme 2). As already mentioned above, two general methods for the removal of alkyl esters from the phosphonate group can be employed: (a) commonly used deprotection with Me₃SiBr in acetonitrile followed by hydrolysis (method A),²⁸ (b) recently described microwave-assisted hydrolysis using aqueous HCl (method B).²⁹ Method A gave better yields (50–68%) of the free phosphonic acids compared to the method B (37–46%). Diisopropyl phosphonate **9d**, with the methoxy group in position C-4 of the pyrimidine ring, afforded under the deprotection conditions (by both methods A and B) the corresponding thymine derivative **7d** (Table 2, entry 4).

All newly prepared CPME analogues **7** were tested for their antiviral properties. Although completely nontoxic in all assays, none of the compounds (including the most promising (S)-CPMEA (**7a**)) exhibited any promising activity against the HIV-1 virus or any other viruses tested (HSV-1 (KOS), HSV-1 (KOS TK⁻), HSV-2 (G), RSV, Vaccinia virus, Vesicular stomatitis virus, Parainfluenza-3 virus, Reovirus-1, Sindbis virus, Coxsackie B4 virus, Punta Toro virus, Feline Corona virus and Feline Herpes virus). One of the obvious reasons for the lack of the biological properties of the CPME derivatives **7** may be their poor bioavailability caused by their high polarity (CPME derivatives are even more polar than the corresponding PME, PMP and HPMP analogues). Since the majority of the therapeutically successful ANPs have to be administered in the form of their prodrugs, the preparation of various prodrugs of the lead (S)-CPMEA (**7a**) is currently in progress and will be reported elsewhere.

3. Conclusion

A novel acyclic nucleoside analogue, (S)-3-(adenin-9-yl)-2-(phosphonomethoxy)propanoic acid (**7a**, (S)-CPMEA), was designed as an inhibitor of HIV-1 RT and its synthesis was developed and optimized. The key step of the (S)-CPMEA (**7a**) synthesis consists of oxidation of the (S)-HPMPA derivative **8a** with TEMPO/NaClO₂/NaClO to give compound **9a** in a high yield (87%). Subsequently, a whole series of CPME derivatives **9** was prepared in high yields from the corresponding HPMP analogues **8** by the optimized oxidative methodology using TEMPO. The oxidation process tolerates the functional groups present on the common nucleobases (A, C, T, U), with the exception of compounds containing the amino group in position C-2 of the purine ring (i.e., **8g** and **8h**). Compounds **8g** and **8h** had to be oxidized as their benzoyl derivatives **8i** and **8j**, respectively. Finally, two methods were employed for the deprotection of the phosphonate diesters **9**: (a) treatment with Me₃SiBr in acetonitrile followed by hydrolysis; (b) microwave-assisted hydrolysis with aqueous HCl. Unfortunately, none of the newly synthesized CPME compounds (including the most promising (S)-CPMEA) showed any interesting activity against the viruses tested.

4. Experimental section

4.1. General

Analytical TLC was performed on silica gel pre-coated aluminium plates with fluorescent indicator (Merck 5554, 60 F₂₅₄). Spots were visualized with UV light (254 nm) or by spraying with 4-nitrobenzylpyridine (2% solution in ethanol) followed by short heating to 600 °C and spraying with concentrated ammonia (26% solution in water). Column chromatography was carried out on silica gel (Sigma S-0507, 40–63 µm). Mass spectra were measured

on a Q-ToF micro (Waters) and HRMS were taken on an LTQ Orbitrap XL (Thermo Fisher Scientific) spectrometer. ¹H and ¹³C NMR spectra were recorded on a Bruker Avance 400 (¹H at 400 MHz and ¹³C 100.6 MHz), Bruker Avance 500 (¹H at 500 MHz and ¹³C 125.7 MHz) and Bruker Avance 600 (¹H at 600.1 MHz and ¹³C 150.9 MHz) in DMSO-*d*₆ (referenced to the solvent signal δ=2.50 and 39.70 ppm, respectively), or in D₂O (referenced to dioxane as an internal standard δ=ppm). Complete assignment is based on heteronuclear correlation experiments HSQC and H,C-HMBC. Chemical shifts (δ) are in parts per million and coupling constants (*J*) in hertz. UV spectra were taken on a Beckman DU-65 spectrophotometer in methanol or water solution. IR spectra were obtained on a FT IR Bruker Equinox IFS 55 spectrometer in KBr pellets.

The microwave-assisted reactions were carried out in CEM Discover (Explorer) microwave apparatus, 24-position system for 10-mL vessels sealed with a Teflon septum. It was operated at a frequency of 2.45 GHz with continuous irradiation power from 0 to 300 W. The solutions were steadily stirred during the reaction. The temperature was measured with an IR sensor on the outer of the process vessel. The vials were cooled to ambient temperature with gas jet cooling system. The pressure was measured with an inboard CEM Explorer pressure control system (0–21 bar).

Household bleach SAVO purchased from BOCHEMIE a.s. (Lidická 326, 735 95 Bohumín, Czech Republic) and the concentration of sodium hypochlorite was determined by iodometric titration (40.7 mg/mL). Starting HPMP analogues were prepared according to the published procedures.¹⁸

4.2. Computational details

The docking studies (Fig. 2) were performed with the Argus-Lab³⁰ program with the use of the 'ArgusDock' algorithm. An exhaustive search was performed by enabling the 'High precision' option in the Docking precision menu, 'Dock' was chosen as the calculation type, and 'flexible' for the ligand. Spacing of 0.2 Å between the grid points was used. The atomic co-ordinates for the X-ray structures of the HIV-1 RT–DNA complexes after incorporation of the anti-HIV drug tenofovir were downloaded from the Protein Data Bank (PDB code: 1T05).¹⁷ The final inhibitor–protein complex was visualized using ArgusLab.³⁰

4.3. Synthesis of the *N*-benzoyl derivatives **8i** and **8j**

Starting HPMP derivative **8g** or **8h** (1.5 mmol) was dissolved in dry pyridine (20 mL) in a round-bottomed flask with a drying tube, and the reaction mixture was cooled in an ice bath. Me₃SiBr (7.5 mmol) was added to the reaction mixture and after 30 min benzoyl chloride (7.5 mmol) was added. The reaction flask was removed from the ice bath and the mixture was stirred for 2 h at room temperature. Then the reaction mixture was cooled in ice bath and cold water (4 mL) was added, followed after 15 min of stirring by concentrated aqueous ammonia (4 mL). The mixture was stirred at room temperature for another 30 min and solvents were removed in vacuo to give crude oil. The chromatography on silica gel column (0–3% methanol in chloroform) afforded the product **8i** or **8j** as thick yellowish oil.

4.3.1. Diisopropyl (S)-N²-benzoyl-9-[3-hydroxy-2-(phosphonomethoxy)propyl]guanine (8i**).**^{18b} Yield 49%, yellowish solid (recryst EtOAc/hexane), mp 171–173 °C (mp 173–174 °C);^{18b} MS-ESI⁺ *m/z* (%) 508 (15, M+H⁺), 530 (100, M+Na⁺); HRMS-ESI⁺: *m/z* calcd for C₂₂H₃₁O₇N₅P (M+H⁺) 508.1956, found 508.1954. ¹H NMR (DMSO-*d*₆) spectrum is identical with the original spectroscopic data.^{18b}

4.3.2. Diisopropyl (S)-N²,N⁶-dibenzoyl-9-[3-hydroxy-2-(phosphonomethoxy)propyl]-2,6-diaminopurine (8j**).** Yield 47%, thick yellowish

oil; ^1H NMR (DMSO- d_6) δ : 11.16 (s, 1H, NH), 11.01 (s, 1H, NH), 8.31 (s, 1H, H-8), 8.06 (m, 2H, H-2''), 7.99 (m, 2H, H-2''), 7.50–7.66 (m, 6H, H-3', 4'), 5.04 (t, $J_{\text{OH}-\text{CH}_2}=5.7$ Hz, 1H, OH), 4.44–4.56 (m, 3H, CH₂Pr, H-1'a), 4.35 (dd, 1H, $J_{\text{gem}}=14.7$ Hz, $J_{1'b-2'}=6.4$ Hz, H-1'b), 3.89–3.94 (m, 3H, H-2', CH₂P), 3.50 (m, 1H, H-3'), 3.43 (m, 1H, H-3'), 1.20 (d, $J_{\text{CH}_3-\text{CH}}=6.2$ Hz, 3H, CH₃Pr), 1.17 (d, $J_{\text{CH}_3-\text{CH}}=6.2$ Hz, 3H, CH₃Pr), 1.16 (d, $J_{\text{CH}_3-\text{CH}}=6.2$ Hz, 3H, CH₃Pr), 1.13 (d, $J_{\text{CH}_3-\text{CH}}=6.2$ Hz, 3H, CH₃Pr); ^{13}C NMR (DMSO- d_6) δ : 165.86 (CON), 165.74 (CON), 153.90 (C-4), 152.30 (C-2), 150.68 (C-6), 145.28 (C-8), 134.46 (C-1''), 133.56 (C-1''), 132.64 (C-4''), 132.20 (C-4''), 128.70 (C-2'', 3''), 128.63 (C-2'', 3''), 128.59 (C-2'', 3''), 128.28 (C-2'', 3''), 123.08 (C-5), 79.91 (d, $J_{2'-\text{P}}=11.5$ Hz, C-2'), 70.48 (d, $J_{\text{C}-\text{O}-\text{P}}=6.3$ Hz, CH₂P), 63.57 (d, $J_{\text{C}-\text{P}}=165.1$ Hz, CH₂P), 59.97 (C-3'), 43.36 (C-1'), 23.78–24.00 (m, CH₃Pr); MS-ESI⁺ m/z (%) 611 (5, M+H⁺), 633 (100, M+Na⁺), 655 (12, M+2Na⁺); HRMS-ESI⁺: m/z calcd for C₂₉H₃₆O₇N₆P (M+H⁺) 611.2378, found 611.2376.

4.4. Oxidation of diisopropyl (S)-HPMPA (8a) with ruthenium tetroxide

Compound **8a** (1.0 mmol, 387 mg), RuO₂ (0.15 mmol, 20 mg) and NaIO₄ (5.0 mmol, 1.1 g) were vigorously stirred in a mixture of CH₃CN (15 mL), CHCl₃ (15 mL) and H₂O (30 mL). Concentrated HCl (eight drops) was added to ~pH 2.5. The heterogeneous mixture was vigorously stirred at room temperature for 72 h. The solid was removed by filtration of the reaction mixture through a silica gel layer and washed with methanol. The solvents were evaporated and the residue was purified by column chromatography on silica gel (CHCl₃/MeOH/AcOH, 88:6:6). The purification was repeated two more times to remove colourful by-products and inorganic salts. The desired product **9a** (253 mg, 63%) was obtained as a yellowish oil, which upon standing in EtOAc/MeOH mixture crystallized.

4.5. Oxidation of diisopropyl (S)-HPMPA (8a) with TEMPO/BAIB

Compound **8a** (1.0 mmol, 387 mg), TEMPO (0.2 mmol, 32 mg) and BAIB (2.2 mmol, 644 mg) in a mixture of CH₃CN (2 mL) and H₂O (2 mL) were stirred at room temperature for 24 h and the solvents were evaporated. Flash chromatography on silica gel (CHCl₃/MeOH, 4:1) afforded the product **9a** (285 mg, 71%) as a yellowish oil, which upon standing in EtOAc/MeOH mixture crystallized.

4.6. Oxidation of diisopropyl HPMP derivatives **8** with TEMPO/NaClO₂/NaClO. General procedure

The corresponding HPMP derivative **8** (1.0 mmol), TEMPO (0.3 mmol, 47 mg) and NaClO₂ (2.0 mmol, 226 mg) in a mixture of CH₃CN (10 mL) and 0.67 M aqueous sodium phosphate buffer (10 mL, ~pH 6.7) were vigorously stirred at room temperature. An aqueous solution of sodium hypochlorite (40.7 mg/mL), prepared by dilution of household bleach SAVO (1.0 mL) with H₂O (1.0 mL) was added. The mixture was stirred at room temperature until the reaction was complete (~24 h) and the solvents were evaporated. Flash chromatography on silica gel column (CHCl₃/MeOH, 4:1) afforded the desired CPME products **9** in good yields (76–87%) as yellowish oils, which upon standing in EtOAc/MeOH mixture gave yellowish crystals.

4.6.1. (S)-3-(6-Amino-9H-purin-9-yl)-2-[(diisopropoxyphosphono)methoxy]propanoic acid (9a). Yellowish crystals, dec >150 °C, yield 87%; ^1H NMR (DMSO- d_6) δ : 8.15 (s, 1H, H-2), 8.03 (s, 1H, H-8), 7.28 (br s, 2H, NH₂), 4.37–4.55 (m, 5H, H-1', H-2', CH₂P), 3.95 (dd, $J_{\text{gem}}=13.7$ Hz, $J_{\text{H}-\text{C}-\text{P}}=8.7$ Hz, 1H, CH₂Pb), 3.73 (dd, $J_{\text{gem}}=13.7$ Hz, $J_{\text{H}-\text{C}-\text{P}}=9.4$ Hz, 1H, CH₂Pa), 1.18 (d, $J=6.2$ Hz, 3H, CH₃Pr), 1.14 (d, $J=6.2$ Hz, 3H, CH₃Pr), 1.13 (d, $J=6.2$ Hz, 3H, CH₃Pr), 1.07 (d,

$J=6.2$ Hz, 3H, CH₃Pr); ^{13}C NMR (DMSO- d_6) δ : 170.61 (C-3'), 155.93 (C-6), 152.37 (C-2), 149.75 (C-4), 141.47 (C-8), 118.55 (C-5), 77.99 (d, $J_{2'-\text{P}}=12.7$ Hz, C-2'), 70.58 (d, $J_{\text{C}-\text{O}-\text{P}}=6.5$ Hz, CH₂P), 70.53 (d, $J_{\text{C}-\text{O}-\text{P}}=6.5$ Hz, CH₂P), 64.08 (d, $J_{\text{C}-\text{P}}=163.9$ Hz, CH₂P), 44.36 (C-1'), 23.92 (d, $J_{\text{C}-\text{C}-\text{O}-\text{P}}=4.0$ Hz, CH₃Pr), 23.88 (d, $J_{\text{C}-\text{C}-\text{O}-\text{P}}=3.9$ Hz, CH₃Pr), 23.78 (d, $J_{\text{C}-\text{C}-\text{O}-\text{P}}=4.5$ Hz, CH₃Pr), 23.67 (d, $J_{\text{C}-\text{C}-\text{O}-\text{P}}=4.5$ Hz, CH₃Pr); MS-ESI⁺ m/z (%) 402 (60, M+H⁺), 825 (100, 2M+Na⁺); HRMS-ESI⁺: m/z calcd for C₁₅H₂₅O₆N₅P (M+H⁺) 402.1537, found 402.1536; FTIR (KBr, cm⁻¹) ν : 3396, 3181, 2981, 1688, 1651, 1599, 1233, 996. [α]_D²⁰ –26.2 (c 0.302, MeOH).

4.6.2. (R)-3-(6-Amino-9H-purin-9-yl)-2-[(diisopropoxyphosphono)methoxy]propanoic acid (9b). Yellowish crystals, dec >150 °C, yield 76%; ^1H NMR (DMSO- d_6) δ : 8.15 (s, 1H, H-2), 8.03 (s, 1H, H-8), 7.28 (br s, 2H, NH₂), 4.37–4.55 (m, 5H, H-1', H-2', CH₂P), 3.95 (dd, $J_{\text{gem}}=13.7$ Hz, $J_{\text{H}-\text{C}-\text{P}}=8.7$ Hz, 1H, CH₂Pb), 3.73 (dd, $J_{\text{gem}}=13.7$ Hz, $J_{\text{H}-\text{C}-\text{P}}=9.4$ Hz, 1H, CH₂Pa), 1.18 (d, $J=6.2$ Hz, 3H, CH₃Pr), 1.14 (d, $J=6.2$ Hz, 3H, CH₃Pr), 1.13 (d, $J=6.2$ Hz, 3H, CH₃Pr), 1.07 (d, $J=6.2$ Hz, 3H, CH₃Pr); ^{13}C NMR (DMSO- d_6) δ : 170.61 (C-3'), 155.93 (C-6), 152.37 (C-2), 149.75 (C-4), 141.47 (C-8), 118.55 (C-5), 77.99 (d, $J_{2'-\text{P}}=12.7$ Hz, C-2'), 70.58 (d, $J_{\text{C}-\text{O}-\text{P}}=6.5$ Hz, CH₂P), 70.53 (d, $J_{\text{C}-\text{O}-\text{P}}=6.5$ Hz, CH₂P), 64.08 (d, $J_{\text{C}-\text{P}}=163.9$ Hz, CH₂P), 44.36 (C-1'), 23.92 (d, $J_{\text{C}-\text{C}-\text{O}-\text{P}}=4.0$ Hz, CH₃Pr), 23.88 (d, $J_{\text{C}-\text{C}-\text{O}-\text{P}}=3.9$ Hz, CH₃Pr), 23.78 (d, $J_{\text{C}-\text{C}-\text{O}-\text{P}}=4.5$ Hz, CH₃Pr), 23.67 (d, $J_{\text{C}-\text{C}-\text{O}-\text{P}}=4.5$ Hz, CH₃Pr); MS-ESI⁺ m/z (%) 402 (60, M+H⁺), 825 (100, 2M+Na⁺); HRMS-ESI⁺: m/z calcd for C₁₅H₂₅O₆N₅P (M+H⁺) 402.1537, found 402.1536; FTIR (KBr, cm⁻¹) ν : 3396, 3181, 2981, 1688, 1651, 1599, 1233, 996. [α]_D²⁰ +28.0 (c 0.613, MeOH).

4.6.3. (S)-3-(4-Amino-2-oxopyrimidin-1(2H)-yl)-2-[(diisopropoxyphosphono)methoxy]propanoic acid (9c). Yellowish crystals, dec >150 °C, yield 83%; ^1H NMR (DMSO- d_6) δ : 7.43 (d, $J_{6-5}=7.3$ Hz, 1H, H-6), 7.34 (br s, 1H, NH₂), 7.19 (br s, 1H, NH₂), 5.66 (d, $J_{5-6}=7.3$ Hz, 1H, H-5), 4.49–4.58 (m, 2H, CH₂P), 4.26 (dd, $J_{\text{CH}-\text{CH}_2}=8.7$ and 3.7 Hz, 1H, 1-CH₂-CH), 4.15 (dd, $J_{\text{gem}}=13.8$ Hz, $J_{\text{CH}_2-\text{CH}}=3.7$ Hz, 1H, 1-CH₂a), 3.91 (dd, $J_{\text{gem}}=13.8$ Hz, $J_{\text{H}-\text{C}-\text{P}}=8.6$ Hz, 1H, CH₂Pa), 3.64–3.71 (m, 2H, CH₂Pb, 1-CH₂b), 1.17–1.23 (m, 12H, CH₃Pr); ^{13}C NMR (DMSO- d_6) δ : 171.04 (COOH), 165.84 (C-4), 155.22 (C-2), 147.29 (C-6), 93.24 (C-5), 77.77 (d, $J_{\text{C}-\text{O}-\text{C}-\text{P}}=12.8$ Hz, 1-CH₂-CH), 70.50 (d, $J_{\text{C}-\text{O}-\text{P}}=6.1$ Hz, CH₂P), 64.15 (d, $J_{\text{C}-\text{P}}=163.9$ Hz, CH₂P), 50.39 (N-CH₂), 23.77–23.98 (m, CH₃Pr); MS-ESI⁺ m/z (%) 378 (20, M+H⁺), 400 (85, M+Na⁺), 422 (100, M+2Na⁺); HRMS-ESI⁺: m/z calcd for C₁₄H₂₃O₇N₃P (M+H⁺) 376.1279, found 376.1280; FTIR (KBr, cm⁻¹) ν : 3357, 3183, 2979, 2901, 1723, 1652, 1376, 1238, 1002. [α]_D²⁰ –75.7 (c 0.264, MeOH).

4.6.4. (S)-2-[(Diisopropoxyphosphono)methoxy]-3-(4-methoxy-5-methyl-2-oxopyrimidin-1(2H)-yl)propanoic acid (9d). Yellowish crystals, mp 82–90 °C, dec at 130 °C, yield 82%; ^1H NMR (DMSO- d_6) δ : 7.63 (q, $J_{6-\text{CH}_3}=1.0$ Hz, 1H, H-6), 4.43–4.54 (m, 2H, CH₂P), 4.27 (dd, $J_{\text{gem}}=13.7$ Hz, $J_{1'a-2'}=3.3$ Hz, 1H, H-1'a), 4.03 (dd, $J_{\text{gem}}=13.8$ Hz, $J_{\text{H}-\text{C}-\text{P}}=7.8$ Hz, 1H, CH₂Pa), 3.96 (dd, $J_{2'-1'}=9.8$ and 3.3 Hz, 1H, H-2'), 3.82 (s, 3H, OCH₃), 3.62 (dd, $J_{\text{gem}}=13.7$ Hz, $J_{1'b-2'}=9.7$ Hz, 1H, H-1'b), 3.54 (dd, $J_{\text{H}-\text{C}-\text{P}}=9.1$ Hz, $J_{\text{gem}}=13.8$ Hz, 1H, CH₂Pb), 1.84 (d, $J_{\text{CH}_3-6}=1.0$ Hz, 3H, 5-CH₃), 1.20 (d, $J_{\text{CH}_3-\text{CH}}=6.2$ Hz, 3H, CH₃Pr), 1.17 (d, $J_{\text{CH}_3-\text{CH}}=6.2$ Hz, 3H, CH₃Pr), 1.15 (d, $J_{\text{CH}_3-\text{CH}}=6.2$ Hz, 3H, CH₃Pr), 1.12 (d, $J_{\text{CH}_3-\text{CH}}=6.2$ Hz, 3H, CH₃Pr); ^{13}C NMR (DMSO- d_6) δ : 171.38 (C-3'), 169.99 (C-4), 155.45 (C-2), 147.59 (C-6), 101.43 (C-5), 79.65 (d, $J_{2'-\text{P}}=12.5$ Hz, C-2'), 70.16 (m, CH₂P), 63.45 (d, $J_{\text{C}-\text{P}}=163.6$ Hz, CH₂P), 53.95 (OCH₃), 51.64 (C-1'), 23.66–23.90 (m, CH₃Pr), 11.81 (5-CH₃); MS-ESI⁺ m/z (%) 407 (5, M+H⁺), 429 (15, M+Na⁺), 451 (100, M+2Na⁺); HRMS-ESI⁺: m/z calcd for C₁₆H₂₇O₈N₂NaP (M+H⁺) 429.1397, found 429.1397; FTIR (KBr, cm⁻¹) ν : 3440, 2981, 2935, 1670, 1541, 1480, 1374, 1244, 1106, 994. [α]_D²⁰ –84.4 (c 0.399, MeOH).

4.6.5. (S)-2-[(Diisopropoxyphosphono)methoxy]-3-(2,4-dioxo-3,4-dihydropyrimidin-1(2H)-yl)propanoic acid (9e). Yellowish crystals,

dec >150 °C, yield 81%; ^1H NMR (DMSO- d_6) δ : 11.23 (br s, 1H, H-3), 7.49 (d, J_{6-5} =7.9 Hz, 1H, H-6), 5.48 (dd, J_{5-6} =7.9 Hz, J_{5-3} =2.1 Hz, 1H, H-5), 4.49–4.59 (m, 2H, CH_{ipr}), 4.14 (dd, J_{gem} =14.0 Hz, $J_{\text{CH}_2-\text{CH}}$ =3.5 Hz, 1H, 1-CH_{2a}), 4.08 (dd, $J_{\text{CH}-\text{CH}_2}$ =9.1 and 3.5 Hz, 1H, 1-CH₂-CH), 3.97 (dd, J_{gem} =13.9 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =8.3 Hz, 1H, CH₂Pa), 3.69 (dd, J_{gem} =14.0 Hz, $J_{\text{CH}_2-\text{CH}}$ =9.2 Hz, 1H, 1-CH_{2b}), 3.65 (dd, J_{gem} =13.9 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =8.5 Hz, 1H, CH₂Pb), 1.17–1.22 (m, 12H, CH_{3ipr}); ^{13}C NMR (DMSO- d_6) δ : 171.22 (COOH), 164.01 (C-4), 151.06 (C-2), 146.76 (C-6), 100.42 (C-5), 78.92 (d, $J_{\text{C}-\text{O}-\text{C}-\text{P}}$ =11.9 Hz, 1-CH₂-CH), 70.39 (m, CH_{ipr}), 63.72 (d, $J_{\text{C}-\text{P}}$ =163.7 Hz, CH₂P), 49.63 (1-CH₂), 23.79–23.99 (m, CH_{3ipr}); MS-ESI⁺ m/z (%) 379 (5, M+H⁺), 401 (60, M+Na⁺), 423 (100, M+2Na⁺); HRMS-ESI⁺ m/z calcd for C₁₄H₂₂O₈N₂P (M+H⁺): 377.1119, found: 377.1119; FTIR (KBr, cm⁻¹) ν : 3417, 2981, 1691, 1630, 1464, 1388, 1245, 1105, 1001. $[\alpha]_{\text{D}}^{20}$ –53.0 (c 0.322, MeOH).

4.6.6. (*R*)-2-[(Diisopropoxyphosphono)methoxy]-3-(2,4-dioxo-3,4-dihydropyrimidin-1(2H)-yl)propanoic acid (**9f**). Yellowish crystals, dec >150 °C, yield 80%; ^1H NMR (DMSO- d_6), ^{13}C NMR (DMSO- d_6), MS and FTIR spectra are identical with compound **9e**. $[\alpha]_{\text{D}}^{20}$ +68.1 (c 0.584, MeOH).

4.6.7. (*S*)-3-(2-Benzamido-6-oxo-1H-purin-9(6H)-yl)-2-[(diisopropoxyphosphono)methoxy]propanoic acid (**9i**). Colourless oil, yield 78%; ^1H NMR (DMSO- d_6) δ : 10.59 (br s, 1H, NH), 8.01 (s, 1H, H-8), 7.95 (m, 2H, H-2''), 7.58 (m, 1H, H-4''), 7.49 (m, 2H, H-3''), 7.43 (br s, 2H, NH₂), 4.35–4.54 (m, 5H, H-2', CH_{ipr}, H-1'), 3.97 (dd, J_{gem} =13.7 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =8.9 Hz, 1H, CH₂Pa), 3.77 (dd, J_{gem} =13.7 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =9.7 Hz, 1H, CH₂Pb), 1.09–1.20 (m, 12H, CH_{3ipr}); ^{13}C NMR (DMSO- d_6) δ : 170.59 (C-3'), 165.88 (CON), 152.42 (C-2 or C-6), 150.61 (C-4), 141.37 (C-8), 134.65 (C-1''), 131.99 (C-4''), 128.50 (C-3''), 128.18 (C-2''), 116.23 (C-5), 77.98 (d, $J_{2'-\text{P}}$ =13.5 Hz, C-2'), 70.54–70.63 (m, CH_{ipr}), 64.16 (d, $J_{\text{C}-\text{P}}$ =164.4 Hz, CH₂P), 44.25 (C-1'), 23.68–23.95 (m, CH_{3ipr}); MS-ESI⁺ m/z (%) 522 (75, M+H⁺), 544 (100, M+Na⁺); HRMS-ESI⁺ m/z calcd for C₂₂H₂₉O₈N₅P (M+H⁺): 522.1748, found: 522.1747; FTIR (KBr, cm⁻¹) ν : 3318, 3143, 2981, 2500, 1674, 1602, 1529, 1453, 1443, 1387, 1255, 955. $[\alpha]_{\text{D}}^{20}$ –40.7 (c 0.194, MeOH).

4.6.8. (*S*)-3-(N²,N⁶-Dibenzoyl-2,6-diamino-9H-purin-9-yl)-2-[(diisopropoxyphosphono)methoxy]propanoic acid (**9j**). Colourless oil, yield 80%; MS-ESI⁺ m/z (%) 625 (7, M+H⁺), 647 (15, M+Na⁺) 669 (100, M+2Na⁺); HRMS-ESI⁺ m/z calcd for C₂₉H₃₄O₈N₆P (M+H⁺): 625.2170, found: 625.2171.

4.7. (*S*)-3-(2-Amino-6-oxo-1H-purin-9(6H)-yl)-2-[(diisopropoxyphosphoryl)methoxy]propanoic acid (**9g**)

To a solution of compound **9i** (1 mmol, 521 mg) in MeOH (10 mL) was added 1.0 M MeONa (2 mmol, 2 mL). After stirring at room temperature for 24 h, the reaction mixture was neutralized with glacial AcOH. The solvents were evaporated and the residue was codistilled with toluene (2×10 mL) and ethanol (2×10 mL). Chromatography on silica gel column (gradient 5–20% MeOH in chloroform) followed by crystallization from EtOAc afforded 296 mg (71%) of compound **9g** as a yellowish oil: ^1H NMR (DMSO- d_6) δ : 12.32 (br s, 1H, H-1), 7.56 (s, 1H, H-8), 7.22 (br s, 2H, NH₂), 4.41–4.53 (m, 2H, CH_{ipr}), 4.33 (dd, J_{gem} =13.5 Hz, $J_{1'a-2'}$ =1.9 Hz, 1H, H-1'a), 4.10 (dd, J_{gem} =13.7 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =8.1 Hz, 1H, CH₂Pa), 3.88–3.97 (m, 2H, H-1'b, H-2'), 3.56 (dd, J_{gem} =13.7 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =9.0 Hz, 1H, CH₂Pb), 1.19 (d, $J_{\text{CH}_3-\text{CH}}$ =6.2 Hz, 3H, CH_{3ipr}), 1.15 (d, $J_{\text{CH}_3-\text{CH}}$ =6.2 Hz, 3H, CH_{3ipr}), 1.14 (d, $J_{\text{CH}_3-\text{CH}}$ =6.2 Hz, 3H, CH_{3ipr}), 1.11 (d, $J_{\text{CH}_3-\text{CH}}$ =6.2 Hz, 3H, CH_{3ipr}); ^{13}C NMR (DMSO- d_6) δ : 171.53 (C-3'), 157.95 (C-6), 154.61 (C-2), 151.56 (C-4), 137.78 (C-8), 116.26 (C-5), 81.36 (d, $J_{2'-\text{P}}$ =13.0 Hz, C-2'), 70.24–70.38 (m, CH_{ipr}), 63.28 (d, $J_{\text{C}-\text{P}}$ =163.7 Hz, CH₂P), 45.54 (C-1'), 23.74–23.98 (m, CH_{3ipr}); MS-ESI⁺ m/z (%) 416 (100, M+H⁺), 438 (10, M+Na⁺); HRMS-ESI⁺ m/z calcd for C₁₅H₂₃O₇N₅P (M+H⁺): 416.1340, found 416.1339; FTIR (KBr,

cm⁻¹) ν : 3423, 3163, 1686, 1630, 1579, 1485, 1410, 1242, 1178, 1105, 997. $[\alpha]_{\text{D}}^{20}$ –35.6 (c 0.407, MeOH).

4.8. Deprotection of diisopropyl esters **9** to free phosphonic acids **7**

4.8.1. Method A.²⁸ General procedure. Diisopropyl esters **9** (0.5 mmol) in CH₃CN (10 mL) were treated with Me₃SiBr (2 mL) at room temperature overnight. Volatiles were evaporated, and the reaction mixture codistilled with dry acetonitrile (3×10 mL) and water (1×10 mL). Crystallization from MeOH gave the products **7** (50–68%) as white or yellowish crystals.

4.8.2. Method B.²⁹ General procedure. A solution of diisopropyl esters **9** (0.5 mmol) in 0.5 M aqueous HCl (2 mL) was heated in a microwave at 140 °C for 30 min. The volatiles were evaporated, the residue was dissolved in H₂O (0.5–1.0 mL) and the product was isolated by HPLC (gradient: 2% aqueous MeOH to 80% aqueous MeOH). Crystallization from a water/ethanol mixture afforded the products **7** (37–46%) as white or yellowish crystals.

4.8.3. (*S*)-3-(6-Amino-9H-purin-9-yl)-2-(phosphonomethoxy)propanoic acid (**7a**). Method A: white solid (68%), dec >250 °C; method B: white crystals (46%), dec >250 °C; ^1H NMR (D₂O) δ : 8.17 (s, 1H, H-2), 8.16 (s, 1H, H-8), 4.55 (dd, J_{gem} =14.8 Hz, $J_{1'a-2'}$ =4.0 Hz, 1H, H-1'a), 4.48 (dd, J_{gem} =14.8 Hz, $J_{1'b-2'}$ =6.0 Hz, 1H, H-1'b), 4.20 (dd, $J_{2'-1'b}$ =6.0 Hz, $J_{2'-1'a}$ =4.0 Hz, 1H, H-2'), 3.71 (dd, J_{gem} =12.8 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =8.6 Hz, 1H, CH₂Pa), 3.45 (dd, J_{gem} =12.8 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =9.8 Hz, 1H, CH₂Pb); ^{13}C NMR (D₂O) δ : 177.20 (C-3'), 155.86 (C-6), 149.62 (C-4), 143.74 (C-8), 118.57 (C-5), 81.75 (d, $J_{2'-\text{P}}$ =12.4 Hz, C-2'), 67.33 (d, $J_{\text{C}-\text{P}}$ =155.0 Hz, CH₂P), 46.12 (C-1'); MS-ESI⁺ m/z (%) 318 (100, M+H⁺), 340 (83, M+Na⁺), 362 (65, M+2Na⁺); HRMS-ESI⁺ m/z calcd for C₉H₁₃O₆N₅P (M+H⁺): 318.0598, found 318.0598; FTIR (KBr, cm⁻¹) ν : 3178, 3033, 2838, 1647, 1598, 1474, 1417, 1056. Anal. Calcd for C₉H₁₂N₅O₆P·0.75H₂O: C, 32.69; H, 4.11; N, 21.18; P, 9.37. Found: C, 32.86; H, 3.99; N, 20.95; P, 9.61. $[\alpha]_{\text{D}}^{20}$ –6.1 (c 0.379, H₂O).

4.8.4. (*R*)-3-(6-Amino-9H-purin-9-yl)-2-(phosphonomethoxy)propanoic acid (**7b**). Method A: white crystals (50%), dec >250 °C; ^1H NMR (D₂O) δ : 8.17 (s, 1H, H-2), 8.16 (s, 1H, H-8), 4.55 (dd, J_{gem} =14.8 Hz, $J_{1'a-2'}$ =4.0 Hz, 1H, H-1'a), 4.48 (dd, J_{gem} =14.8 Hz, $J_{1'b-2'}$ =6.0 Hz, 1H, H-1'b), 4.20 (dd, $J_{2'-1'b}$ =6.0 Hz, $J_{2'-1'a}$ =4.0 Hz, 1H, H-2'), 3.71 (dd, J_{gem} =12.8 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =8.6 Hz, 1H, CH₂Pa), 3.45 (dd, J_{gem} =12.8 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =9.8 Hz, 1H, CH₂Pb); ^{13}C NMR (D₂O) δ : 177.20 (C-3'), 155.86 (C-6), 149.62 (C-4), 143.74 (C-8), 118.57 (C-5), 81.75 (d, $J_{2'-\text{P}}$ =12.4 Hz, C-2'), 67.33 (d, $J_{\text{C}-\text{P}}$ =155.0 Hz, CH₂P), 46.12 (C-1'); MS-ESI⁺ m/z (%) 318 (100, M+H⁺), 340 (83, M+Na⁺), 362 (65, M+2Na⁺); HRMS-ESI⁺ m/z calcd for C₉H₁₃O₆N₅P (M+H⁺): 318.0598, found 318.0598; FTIR (KBr, cm⁻¹) ν : 3178, 3033, 2838, 1647, 1598, 1474, 1417, 1056. Anal. Calcd for C₉H₁₂N₅O₆P·0.75H₂O: C, 32.69; H, 4.11; N, 21.18; P, 9.37. Found: C, 32.86; H, 3.99; N, 20.97; P, 9.63. $[\alpha]_{\text{D}}^{20}$ +6.2 (c 0.324, H₂O).

4.8.5. (*S*)-3-(4-Amino-2-oxypyrimidin-1(2H)-yl)-2-(phosphonomethoxy)propanoic acid (**7c**). Method A: yellowish crystals (62%), dec >250 °C; method B: yellowish crystals (39%), dec >250 °C; ^1H NMR (D₂O) δ : 7.64 (d, J_{6-5} =7.4 Hz, 1H, H-6), 5.98 (d, J_{5-6} =7.3 Hz, 1H, H-5), 4.21 (dd, J_{gem} =14.0 Hz, $J_{1'a-2'}$ =3.9 Hz, 1H, H-1'a), 4.05 (dd, $J_{2'-1'b}$ =7.7 Hz, $J_{2'-1'a}$ =3.9 Hz, 1H, H-2'), 3.89 (dd, J_{gem} =14.0 Hz, $J_{1'b-2'}$ =7.7 Hz, 1H, H-1'b), 3.71 (dd, J_{gem} =12.9 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =8.7 Hz, 1H, CH₂Pa), 3.42 (dd, J_{gem} =12.8 Hz, $J_{\text{H}-\text{C}-\text{P}}$ =10.2 Hz, 1H, CH₂Pb); ^{13}C NMR (D₂O) δ : 177.57 (C-3'), 166.01 (C-4), 157.42 (C-2), 148.94 (C-6), 95.69 (C-5), 81.62 (d, $J_{2'-\text{P}}$ =13.1 Hz, C-2'), 67.07 (d, $J_{\text{C}-\text{P}}$ =156.4 Hz, CH₂P), 51.87 (C-1'); MS-ESI⁺ m/z (%) 292 (100, M+H⁺), 314 (50, M+Na⁺), 336 (25, M+2Na⁺); HRMS-ESI⁺ m/z calcd for C₈H₁₁O₇N₃P

(M–H⁺) 292.0340, found 292.0340; FTIR (KBr, cm^{−1}) ν : 3385, 3171, 2795, 1650, 1613, 1494, 1400, 1051. [α]_D²⁰ –7.0 (c 0.339, H₂O).

4.8.6. (S)-3-(5-Methyl-2,4-dioxo-3,4-dihydropyrimidin-1(2H)-yl)-2-(phosphonomethoxy)propanoic acid (**7d**). Method A: white crystals (64%), dec >250 °C; method B: white crystals (44%), dec >250 °C; ¹H NMR (D₂O) δ : 7.50 (q, J_{6-CH3}=1.2 Hz, 1H, H-6), 4.10 (dd, J_{gem}=13.9 Hz, J_{1'a-2'}=4.1 Hz, 1H, H-1'a), 4.05 (dd, J_{2'-1'}=6.6 and 4.1 Hz, 1H, H-2'), 3.96 (dd, J_{gem}=13.9 Hz, J_{1'b-2'}=6.6 Hz, 1H, H-1'b), 3.69 (dd, J_{gem}=12.8 Hz, J_{H-C-P}=8.7 Hz, 1H, CH₂Pa), 3.42 (dd, J_{gem}=12.8 Hz, J_{H-C-P}=9.9 Hz, 1H, CH₂Pb), 1.87 (d, J_{CH3-6}=1.2 Hz, 3H, 5-CH₃); ¹³C NMR (D₂O) δ : 177.59 (COOH), 167.78 (C-4), 152.84 (C-2), 144.62 (C-6), 110.71 (C-5), 81.61 (d, J_{2'-P}=12.6 Hz, C-2'), 67.24 (d, J_{C-P}=157.1 Hz, CH₂P), 50.34 (C-1'), 11.93 (5-CH₃); MS-ESI⁺ m/z (%) 307 (100, M–H⁺), 329 (60, M+Na⁺), 351 (15, M+2Na⁺); HRMS-ESI⁺ m/z calcd for C₉H₁₂O₈N₂P (M–H⁺) 307.0326, found 307.0327; FTIR (KBr, cm^{−1}) ν : 3191, 3036, 2833, 1691, 1601, 1475, 1432, 1359, 1110, 1062. [α]_D²⁰ –2.7 (c 0.401, H₂O).

4.8.7. (S)-3-(2,4-Dioxo-3,4-dihydropyrimidin-1(2H)-yl)-2-(phosphonomethoxy)propanoic acid (**7e**). Method B: white crystals (42%), dec >250 °C; ¹H NMR (D₂O+NaOD) δ : 7.59 (d, J₆₋₅=7.5 Hz, 1H, H-6), 5.74 (d, J₅₋₆=7.5 Hz, 1H, H-5), 3.97–4.05 (m, 3H, H-1', 2'), 3.53 (dd, J_{gem}=11.8 Hz, J_{H-C-P}=9.1 Hz, 1H, CH₂P), 3.30 (dd, J_{gem}=11.8 Hz, J_{H-C-P}=10.2 Hz, 1H, CH₂P); ¹³C NMR (D₂O+NaOD) δ : 178.47 (C-3'), 174.43 (C-4), 158.01 (C-2), 147.85 (C-6), 102.05 (C-5), 81.99 (d, J_{2'-P}=13.6 Hz, C-2'), 68.91 (d, J_{C-P}=150.7 Hz, CH₂P), 51.03 (C-1'); MS-ESI[−] m/z (%) 293 (100, M–H⁺); HRMS-ESI[−] m/z calcd for C₈H₁₀O₈N₂P (M–H⁺): 293.0180, found: 293.0180; FTIR (KBr, cm^{−1}) ν : 3428, 3168, 2976, 1692, 1617, 1402, 1356, 1070, 1050. [α]_D²⁰ –50.0 (c 0.346, H₂O).

4.8.8. (R)-3-(2,4-Dioxo-3,4-dihydropyrimidin-1(2H)-yl)-2-(phosphonomethoxy)propanoic acid (**7f**). Method B: white crystals (48%), dec >250 °C; ¹H NMR (D₂O+NaOD), ¹³C NMR (D₂O+NaOD), MS and FTIR spectra are identical with compound **7e**. [α]_D²⁰ +46.0 (c 0.336, H₂O).

4.8.9. (S)-3-(2-Amino-6-oxo-1H-purin-9(6H)-yl)-2-(phosphonomethoxy)propanoic acid (**7g**). Method A: yellowish crystals (61%), dec >250 °C; method B: white solid (37%), dec >250 °C; ¹H NMR (D₂O) δ : 7.84 (s, 1H, H-8), 4.40 (dd, J_{gem}=14.7 Hz, J_{1'a-2'}=4.1 Hz, 1H, H-1'a), 4.32 (dd, J_{gem}=14.7 Hz, J_{1'b-2'}=6.1 Hz, 1H, H-1'b), 4.17 (dd, J_{2'-1'}=6.1 and 4.1 Hz, 1H, H-2'), 3.73 (dd, J_{gem}=13.0 Hz, J_{H-C-P}=8.7 Hz, 1H, CH₂Pa), 3.51 (dd, J_{gem}=13.0 Hz, J_{H-C-P}=9.8 Hz, 1H, CH₂Pb); ¹³C NMR (D₂O) δ : 177.21 (C-3'), 159.57 (C-6), 154.24 (C-2), 152.37 (C-4), 141.38 (C-8), 116.08 (C-5), 81.73 (d, J_{2'-P}=12.5 Hz, C-2'), 66.85 (d, J_{C-P}=156.1 Hz, CH₂P), 45.82 (C-1'); MS-ESI[−] m/z (%) 332 (100, M–H⁺), 354 (50, M+Na⁺), 376 (15, M+2Na⁺); HRMS-ESI[−] m/z calcd for C₉H₁₁O₇N₅P (M–H⁺): 332.0402, found: 332.0404; FTIR (KBr, cm^{−1}) ν : 3405, 3128, 1693, 1607, 1540, 1481, 1409, 1108, 1069, 914. [α]_D²⁰ –20.2 (c 0.316, H₂O).

4.8.10. (S)-3-(2,6-Diamino-9H-purin-9-yl)-2-(phosphonomethoxy)propanoic acid (**7h**). Method B (from **9j**): white crystals (38%), dec >250 °C; ¹H NMR (D₂O+NaOD) δ : 7.94 (s, 1H, H-8), 4.33–4.39 (m, 2H, H-1'), 4.13 (t, J_{2'-1'}=5.1 Hz, 1H, H-2'), 3.58 (dd, J_{gem}=12.1 Hz, J_{H-C-P}=8.9 Hz, 1H, CH₂P), 3.32 (dd, J_{gem}=12.1 Hz, J_{H-C-P}=9.8 Hz, 1H, CH₂P); ¹³C NMR (D₂O+NaOD) δ : 178.02 (C-3'), 160.60 (C-2), 456.66 (C-6), 151.92 (C-4), 141.68 (C-8), 113.20 (C-5), 81.90 (d, J_{2'-P}=12.7 Hz, C-2'), 68.82 (d, J_{C-P}=150.5 Hz, CH₂P), 45.75 (C-1'); MS-ESI⁺ m/z (%) 333 (100, M+H⁺), 355 (25, M+Na⁺), 377 (15, M+2Na⁺); HRMS-ESI⁺ m/z calcd for C₉H₁₄O₆N₆P (M+H⁺) 333.0707, found 333.0707; FTIR (KBr, cm^{−1}) ν : 3194, 2470, 1605, 1560, 1551, 1482, 1407, 1119, 1087, 916. [α]_D²⁰ –17.4 (c 0.259, H₂O).

4.8.11. (S)-3-(2-Benzamido-6-oxo-1H-purin-9(6H)-yl)-2-(phosphonomethoxy)propanoic acid (**7i**). Method A: white crystals (52%), dec

>250 °C; ¹H NMR (D₂O) δ : 8.11 (s, 1H, H-8), 7.97 (m, 2H, H-2''), 7.62 (m, 1H, H-4''), 7.53 (m, 2H, H-3''), 4.45–4.52 (m, 2H, H-1'), 4.16 (m, 1H, H-2'), 3.59 (m, 1H, CH₂P), 3.40 (m, 1H, CH₂P); ¹³C NMR (D₂O) δ : 177.97 (C-3'), 172.77 (CON), 162.48 (C-6), 153.12 (C-2), 151.42 (C-4), 142.76 (C-8), 135.04 (C-1''), 133.13 (C-4'), 129.28 (C-3'), 128.54 (C-2'), 119.73 (C-5), 81.83 (d, J_{2'-P}=12.9 Hz, C-2'), 68.76 (d, J_{C-P}=150.8 Hz, CH₂P), 45.76 (C-1'); MS-ESI⁺ m/z (%) 438 (100, M+H⁺), 460 (78, M+Na⁺); HRMS-ESI⁺ m/z calcd for C₁₆H₁₅O₈N₅P (M–H⁺) 436.0664, found 436.0664; FTIR (KBr, cm^{−1}) ν : 3408, 3135, 3033, 1665, 1604, 1402, 1269, 1154, 1061. [α]_D²⁰ –31.9 (c 0.244, H₂O).

Acknowledgements

This study was performed as a part of research project RVO: 61388963 of the Institute of Organic Chemistry and Biochemistry, v.v.i. This study was supported by Ministry of the Interior of the Czech Republic (VG20102015046), and by Gilead Sciences and IOCB Research Centre.

Supplementary data

Supplementary data associated with this article can be found in the online version, at doi:10.1016/j.tet.2012.03.066. These data include MOL files and InChIKeys of the most important compounds described in this article.

References and notes

- (a) De Clercq, E.; Holý, A.; Rosenberg, I.; Sakuma, T.; Balzarini, J.; Maudgal, P. C. *Nature* **1986**, 323, 464; (b) De Clercq, E. *J. Med. Microbiol.* **1998**, 47, 1; (c) Holý, A. *Curr. Pharm. Des.* **2003**, 9, 2567; (d) De Clercq, E.; Holý, A. *Nat. Rev. Drug Discov.* **2005**, 4, 928; (e) Holý, A. *Antiviral Res.* **2006**, 71, 248; (f) De Clercq, E. *Antiviral Res.* **2007**, 75, 1; (g) De Clercq, E. *Biochem. Pharmacol.* **2007**, 73, 911.
- (a) Holý, A.; Votruba, I.; Tloušťová, E.; Masojídková, M. *Collect. Czech. Chem. Commun.* **2001**, 66, 1545; (b) Zidek, Z.; Potměšil, P.; Holý, A. *Toxicol. Appl. Pharmacol.* **2003**, 192, 246; (c) Reiser, H.; Wang, J.; Chong, L.; Watkins, W. J.; Ray, A. S.; Shibata, R.; Birkus, G.; Cihlar, T.; Wu, S.; Li, B.; Liu, X.; Henne, I. N.; Wolfgang, G. H. I.; Desai, M.; Rhodes, G. R.; Fridland, A.; Lee, W. A.; Plunkett, W.; Vail, D.; Thamm, D. H.; Jeraj, R.; Tumas, D. B. *Clin. Cancer Res.* **2008**, 14, 2824.
- (a) Keough, D. T.; Hocková, D.; Holý, A.; Naesens, L. M. J.; Skinner-Adams, T. S.; de Jersey, J.; Guddat, L. W. *J. Med. Chem.* **2009**, 52, 4391; (b) Hocková, D.; Holý, A.; Masojídková, M.; Keough, D. T.; de Jersey, J.; Guddat, L. W. *Bioorg. Med. Chem.* **2009**, 17, 6218.
- (a) Zidek, Z.; Franková, D.; Holý, A. *Int. J. Immunopharmacol.* **2000**, 22, 1121; (b) Zidek, Z.; Potměšil, P.; Kmoníčková, E.; Holý, A. *Eur. J. Pharmacol.* **2003**, 475, 149; (c) Potměšil, P.; Krečmerová, M.; Kmoníčková, E.; Holý, A.; Zidek, Z. *Eur. J. Pharmacol.* **2006**, 540, 191.
- (a) De Clercq, E. *Intervirology* **1997**, 40, 295; (b) De Clercq, E. *Clin. Microbiol. Rev.* **2003**, 16, 569; (c) De Clercq, E. *Expert Rev. Anti-infect. Ther.* **2003**, 1, 21.
- (a) De Clercq, E.; Holý, A. *Antimicrob. Agents Chemother.* **1991**, 35, 701; (b) Neyts, J.; Snoeck, R.; Balzarini, J.; De Clercq, E. *Antiviral Res.* **1991**, 16, 41; (c) De Clercq, E. *Rev. Med. Virol.* **1993**, 3, 85; (d) Hitchcock, M. J. M.; Jaffe, H. S.; Martin, J. C.; Stagg, R. J. *Antiviral Chem. Chemother.* **1996**, 7, 115.
- (a) Holý, A.; Votruba, I.; Masojídková, M.; Andrei, G.; Snoeck, R.; Naesens, L.; De Clercq, E.; Balzarini, J. *J. Med. Chem.* **2002**, 45, 1918; (b) Balzarini, J.; Pan-necouque, C.; De Clercq, E.; Aquaro, S.; Perno, C.-F.; Egberink, H.; Holý, A. *Antimicrob. Agents Chemother.* **2002**, 46, 2185; (c) Hocková, D.; Holý, A.; Masojídková, M.; Andrei, G.; Snoeck, R.; De Clercq, E.; Balzarini, J. *J. Med. Chem.* **2003**, 46, 5064; (d) Hocková, D.; Holý, A.; Masojídková, M.; Andrei, G.; Snoeck, R.; De Clercq, E.; Balzarini, J. *Bioorg. Med. Chem.* **2004**, 12, 3197; (e) Ying, C.; Holý, A.; Hocková, D.; Havlas, Z.; De Clercq, E.; Neyts, J. *Antimicrob. Agents Chemother.* **2005**, 49, 1177.
- Wagner, C. R.; Iyer, V. V.; McIntee, E. *J. Med. Res. Rev.* **2000**, 20, 417.
- Hecker, S. J.; Erion, M. D. *J. Med. Chem.* **2008**, 51, 2328.
- (a) Holý, A.; Rosenberg, I. *Collect. Czech. Chem. Commun.* **1987**, 52, 2775; (b) Rosenberg, I.; Holý, A. *Collect. Czech. Chem. Commun.* **1987**, 52, 2792; (c) Krečmerová, M.; Masojídková, M.; Holý, A. *Collect. Czech. Chem. Commun.* **2004**, 69, 1889; (d) Janeba, Z.; Masojídková, M.; Holý, A. *Collect. Czech. Chem. Commun.* **2010**, 75, 371.
- (a) Jindřich, J.; Holý, A.; Dvořáková, H. *Collect. Czech. Chem. Commun.* **1993**, 58, 1645; (b) Balzarini, J.; Holý, A.; Jindřich, J.; Dvořáková, H.; Hao, Z.; Snoeck, R.; Herdewijn, P.; John, D. G.; De Clercq, E. *Proc. Natl. Acad. Sci. U.S.A.* **1991**, 88, 4961; (c) Balzarini, J.; Holý, A.; Jindřich, J.; Naesens, L.; Snoeck, R.; Schols, D.; De Clercq, E. *Antimicrob. Agents Chemother.* **1993**, 37, 332.
- (a) De Clercq, E.; Descamps, J.; De Somer, P.; Holý, A. *Science* **1978**, 200, 563; (b) De Clercq, E.; Holý, A. *J. Med. Chem.* **1979**, 22, 510.
- Holý, A.; Votruba, I.; De Clercq, E. *Collect. Czech. Chem. Commun.* **1982**, 47, 1392.

14. (a) Holý, A.; Votruba, I.; De Clercq, E. *Collect. Czech. Chem. Commun.* **1985**, *50*, 262; (b) De Clercq, E.; Holý, A. *J. Med. Chem.* **1985**, *28*, 282.
15. De Clercq, E. *Nucleosides Nucleotides Nucleic Acids* **2005**, *24*, 1395.
16. Votruba, I.; Hasobe, M.; Holý, A.; Borchardt, R. T. *Biochem. Pharmacol.* **1990**, *39*, 1573.
17. Tuske, S.; Sarafianos, S. G.; Clark, A. D., Jr.; Ding, J.; Naeger, L. K.; White, K. L.; Miller, M. D.; Gibbs, C. S.; Boyer, P. L.; Clark, P.; Wang, G.; Gaffney, B. L.; Jones, R. A.; Jerina, D. M.; Hughes, S. H.; Arnold, E. *Nat. Struct. Mol. Biol.* **2004**, *11*, 469.
18. (a) Webb, R. R.; Martin, J. C. *Tetrahedron Lett.* **1987**, *28*, 4963; (b) Holý, A. *Collect. Czech. Chem. Commun.* **1993**, *58*, 649.
19. *Modern Oxidation Methods*; Bäckvall, J.-E., Ed.; WILEY-VCH: Weinheim, 2004.
20. (a) Hutchison, A. J.; Williams, M.; De Jesus, R.; Yokoyama, R.; Oei, H. H.; Ghai, G. R.; Webb, R. L.; Zoganas, H. C.; Stone, G. A.; Jarvis, M. F. *J. Med. Chem.* **1990**, *33*, 1919; (b) Ha, S. B.; Nair, V. *Tetrahedron Lett.* **1996**, *37*, 1567.
21. Mori, N.; Togo, H. *Tetrahedron* **2005**, *61*, 5915.
22. (a) Singh, A. K.; Varma, R. S. *Tetrahedron Lett.* **1992**, *33*, 2307; (b) Varma, R. S.; Hogan, M. E. *Tetrahedron Lett.* **1992**, *33*, 7719.
23. Homma, H.; Watanabe, Y.; Abiru, T.; Murayama, T.; Nomura, Y.; Matsuda, A. *J. Med. Chem.* **1992**, *35*, 2881.
24. (a) Vogler, T.; Studer, A. *Synthesis* **2008**, 1979; (b) De Souza, M. V. N. *Mini-Rev. Org. Chem.* **2006**, *3*, 155; (c) Nooy, A. E. J.; Besemer, A. C.; Van Bekkum, H. *Synthesis* **1996**, 1153.
25. De Mico, A.; Margarita, R.; Parlanti, L.; Vescovi, A.; Piancatelli, G. *J. Org. Chem.* **1997**, *62*, 6974.
26. Epp, J. B.; Widlanski, T. S. *J. Org. Chem.* **1999**, *64*, 293.
27. Zhao, M.; Li, J.; Mano, E.; Song, Z.; Tschaen, D. M.; Grabowski, E. J. J.; Reider, P. J. *J. Org. Chem.* **1999**, *64*, 2564.
28. Holý, A.; Rosenberg, I. *Collect. Czech. Chem. Commun.* **1982**, *47*, 3447.
29. (a) Jansa, P.; Holý, A.; Dračinský, M.; Baszczyński, O.; Česnek, M.; Janeba, Z. *Collect. Symp. Ser.* **2011**, *12*, 347; (b) Jansa, P.; et al., *Green. Chem.*, in press.
30. ArgusLab 4.0, M. A. Thompson, Planaria Software LLC, Seattle, <http://www.ArgusLab.com>.