Macromolecules

Hydrophobic, Hydrophilic, and Amphiphilic Polyglycocarbonates with Linear and Macrocyclic Architectures from Bicyclic Glycocarbonates Derived from CO₂ and Glucoside

Debasis Pati,[†] Xiaoshuang Feng,^{*,†} Nikos Hadjichristidis,[‡] and Yves Gnanou^{*,†}

[†]Physical Sciences and Engineering Division and [‡]KAUST Catalysis Center, Physical Sciences and Engineering Division, King Abdullah University of Science and Technology (KAUST), Thuwal 23955, Saudi Arabia

Supporting Information

ABSTRACT: Two bicyclic glycocarbonates were synthesized in five steps from α -methyl-D-glucoside without resorting to phosgene or to its derivatives for the first time. The 4- and 6positions of glucose were modified to introduce a six-membered carbonate ring, using CO₂ as the carbonylating reagent; the 2- and 3-positions of the same glucoside substrate were first transformed into either methyl or triethylene glycol monomethyl ether groups to protect these positions from undesirable reactions and also to impart hydrophobicity in the first case and hydrophilicity in the second. The polymerization behavior of these bicyclic glycocarbonates was then investigated under different conditions. On the one hand, through ring-opening polymerization of the above monomers, linear polyglycocarbonate homopolymers and diblock copolymers were obtained initiated by *p*-methylbenzyl alcohol



using 1,5,7-triazabicyclo[4.4.0]dec-5-ene (TBD) as catalyst; on the other hand, macrocyclic polyglycocarbonate homopolymers and diblock copolymers were grown using 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) which served as zwitterionic initiator. The various architectures derived were all thoroughly characterized by NMR, GPC, and MALDI-tof and shown to exhibit the expected structure. Finally, the self-assembly of linear and macrocyclic amphiphilic copolyglycocarbonates in water was investigated and characterized by cryo-TEM.

INTRODUCTION

Glycopolymers are synthetic polymer scaffolds that carry dangling sugar moieties. $^{1-6}$ Such sugar-modified synthetic polymers find applications as clinical diagnostics, bioassays, affinity separation, etc.,^{7,8} wherever different carbohydrates serving as multivalent ligands on the exterior of the membrane proteins and lipids can bring about biological activity through carbohydrate-protein interactions.^{9,10} Polysaccharides are another family of polymers that also comprise sugars in their structure. In contrast to the glycopolymers, sugars in polysaccharides are part of the main chain and serve as repeating units or building blocks.¹¹⁻¹⁴ In natural polysaccharides that originate from different sources (plants, bacteria, and animals), sugars are bound one to another by glycosidic linkages.^{15–19} But polysaccharides can also be synthetically produced, and besides glycosidic linkages other chemical functions have been contemplated to link sugars one to another.^{20,21} Carbonates are one such linkage that have been used to generate polycarbonates featuring sugar moieties in the main chain.^{22–28} Sugar-based polycarbonates are usually prepared through ring-opening polymerization (ROP) of either five- or six-membered, sugar-derived carbonate rings. For instance, Gross and collaborators^{22,24} reported the synthesis of D-xylofuranose-based polycarbonates and its copolymers

from the corresponding six-membered cyclic carbonate using for this purpose zinc or tin catalysts; Wooley and coworkers^{27,28} described the synthesis of D-glucose-based sixmembered cyclic carbonate that they successfully polymerized via an organocatalytic approach. Interestingly, the same sugar substrate, D-glucose, was modified by Endo and co-workers^{25,26} into a five-membered cyclic carbonate that they anionically ring-opened under mild conditions to obtain well-defined Dglucose-based polycarbonates of different structure from that prepared by Wooley.^{27,28} Polycondensation reactions involving sugar-based diols and phosgene were also tried as a means to generate sugar-containing linear polycarbonates.^{18,23,29,30} Whether the monomers were meant for ROP or polycondensation all of the carbonylation reactions used so to date for the generation of carbonate functions in the above works involved either phosgene or its derivatives: ethyl chloroformates or bis(pentafluorophenyl)carbonate.^{18,23-32}

The lack of studies on glucose-based polycarbonates or polyglycocarbonates and the limitations related to the use of phosgene and of its derivatives prompted us to explore new

Received: November 23, 2016 Revised: January 18, 2017 Scheme 1. General Scheme for the Synthesis of Six-Membered Bicyclic Glycocarbonate Monomers from α -Methyl D-Glucopyranoside^{α}



"Reagents and conditions: (i) PhCH(OMe)₂, PTSA, DMF, 1 h (yield, 69%); (ii) DMF/NaH, CH₃I (**3a**)/TEGM-OTs (**3b**), 0 °C for 1 h and overnight at RT (yield, 72%); (iii) NBS, CaCO₃, CCl₄, 1 h (yield, 58%); (iv) 2% NaOH in MeOH, 2 h (yield, 60%); (v) CO₂ (10 bar), DBU, DMF, RT, 5 h for monomer **6a** (yield, 40%) and 22 h for monomer **6b** (yield, 40%).

Table 1. ROP of Hydrophobic and Hydrophilic Monomers 6a and 6b under Different Initiating Conditions

entry	polymer	initiator/Cat.	М	$[M]_0/[I]_0$	time (h)	conv ^a (%)	yield (%)	${{M_{(\rm theo)}}}^b$ $(10^3 { m g/mol})$	${M_{ m n(NMR)}}^{c}$ (10 ³ g/mol)	${M_{n(GPC)}}^d$ (10 ³ g/mol)
01	LP ₁ -6a	PMBA/TBD	6a	25	1	98	90	6.20	6.08	6.5/1.2
02	LP ₂ -6a	PMBA/DBU	6a	30	45	90	75	6.82	5.33	4.0/1.3
03	LP-6b	PMBA/TBD	6b	17	5	94	83	8.18	7.80	5.0/1.2
04	CP-6a	DBU	6a	30	45	91	65	6.78	ND ^e	5.0/1.2
05	CP-6b	DBU/TU	6b	12	60	90	60	6.14	ND ^e	3.8/1.2
06	LP ₃ -6a	PEG-OH/TBD	6a	20	1	99	80	9.91	9.00	8.6/1.2
07	LP-6a- <i>b</i> -6b	PMBA/TBD	6a:6b	18:9	11	>99 (6a), >92 (6b)	75	8.86	8.61	8.1/1.2
08	CP-6a- <i>b</i> -6b	DBU/TU	6a:6b	15:7	96	>99 (6a), >90	75	6.90	ND^{e}	4.0/1.2

^{*a*}Aliquots of the reaction medium were analyzed by ¹H NMR to calculate the monomer conversion. ^{*b*}Expected molar mass deduced from monomer conversion (¹H NMR). ^{*c*}Molar mass was calculated from end-group analysis (¹H NMR) of the isolated polymer. ^{*d*}Calculated from GPC calibrated with narrow distributed polystyrene standard in THF as eluent (1 mL/min). ^{*e*}Not determined.

synthetic avenues. In this study, we demonstrate that CO_2 can be effectively resorted to for the carbonylation reaction of α methyl-D-glucoside instead of toxic phosgene or its derivatives, opening up the access to two bicyclic six-membered carbonate monomers (Scheme 1)—the first hydrophobic and the second water-soluble. From these two monomers a wealth of welldefined polyglycocarbonate architectures could be obtained, including hydrophobic and hydrophilic homopolymers with either a linear or a macrocyclic structure as well as linear and macrocyclic amphiphilic diblock copolymers. The ring-opening polymerization of these glucoside-based bicyclic carbonates was triggered by an organocatalyst: the latter gave rise to macrocyclic architectures, and in the presence of an alcohol serving as initiator it brought about the formation of linear chains.

RESULTS AND DISCUSSION

Monomer Synthesis. There are only very few reports on the synthesis of polymers featuring sugar units in their main chain linked by carbonate groups. As previously indicated,^{7,8} such polyglycocarbonates whether obtained either by polycocarbonates of sugar-based diols or by ring-opening polymerization of glycocarbonates required at one step of their synthesis the use of phosgene or its derivatives. In the course of this work Buchard and co-workers reported the synthesis of D-mannose-based six-membered glycocarbonate using CO_2 as carbonylation reagent.³³ Along the same line we show in this investigation (Scheme 1) that both hydrophobic (6a) and hydrophilic (6b) bicyclic glycocarbonates can be prepared from α -methyl-D-glucoside using CO₂ as carbonylation reagent. Both 6a and 6b monomers were synthesized in five steps starting from the commercially available α -methyl-D-glucoside: to the exception of step 2, all steps were carried out following the same experimental procedures for the two monomers. In step 1, the 4- and 6-dihydroxyl groups of α -methyl-D-glucoside were protected by the benzylidene group so as to obtain α -methyl-4,6-O-benzylidene D-glucopyranoside (2) (Supporting Information Figures S8 and S9), using for this purpose benzaldehyde dimethyl acetal in the presence of a catalytic amount of ptoluenesulfonic acid. The protection step of the two 2- and 3dihydroxyls groups of α -methyl-4,6-O-benzylidene D-glucopyranoside (2), or step 2, served to eventually generate either a hydrophobic bicyclic glycocarbonate (6a) or its hydrophilic equivalent (6b). In the first case, 3a (Figures S10 and S11), two methyl groups were introduced as protecting groups using methyl iodide and sodium hydride and in the second, 3b (Figures S24-S26), two molecules of triethylene glycol monomethyl ether were grafted at the 2- and 3-positions of the glucopyranosyl rings using tosylated triethylene glycol methyl ether and sodium hydride. Step 3 was identical for the two monomers: α -methyl-6-bromo-4-benzyl ester-2,3-di-Omethyl-d-glucopyranoside (4a) and α -methyl-6-bromo-4-benzoate-2,3-di-O-TEGM-D-glucopyranoside (4b) were obtained by bromination of α -methyl-2,3-dimethyl-4,6-O-benzylidene-D-



TU=Thiourea was used for CP-6b and CP-6a-b-6b



Figure 1. (A) Plot of monomer conversion (%) (¹H NMR analysis) vs M_n and PDI values (GPC traces), (B) GPC profiles of the reaction aliquots over time, and (C) MALDI-tof of LP₂-6a.

с

glucopyranoside (3a) and of α -methyl-2,3-di-O-TEGM-4,6-Obenzylidene-D-glucopyranoside (3b), respectively, using NBS/ CCl₄ in the presence of CaCO₃. The benzyl ester deprotection of 4a and 4b (Figures S27-S29) was carried out next (step 4) in 2% sodium hydroxide methanol solution at room temperature; these reactions yielded α -methyl-6-bromo-6-deoxy-2,3di-O-methyl-D-glucopyranoside (5a) (Figures S12–S17) and α methyl-6-bromo-6-deoxy-2,3-di-O-TEGM-D-glucopyranoside (5b) (Figures S30–S32), respectively. In the final step 5, 6a and 6b were obtained by carbonylation and cyclization of 5a and 5b, respectively, under a 10 bar CO₂ pressure at room temperature in the presence of 1,8-diazabicyclo[5.4.0]undec-7ene (DBU). The yields for the two monomers **6a** and **6b** were equal to 40%, a value comparable to that reported by Wooley and co-workers, who resorted to the use of bis(pentafluorophenyl)carbonate as carbonylation reagent.²⁷ All NMR (Figures S18-S23 for 6a and Figures S33-S38 for 6b) and HRMS (Figures S5 and S7) characterizations confirmed the purity and

the structure of monomers 6a and 6b synthesized and also those of the intermediate compounds.

Hydrophobic Linear Polyglycocarbonates. The polymerization of 6a was subsequently carried out in the presence of *p*-methylbenzyl alcohol as initiator and TBD as catalyst in anhydrous dichloromethane and at room temperature: the polymerization occurred in a "living" fashion.²⁷ The obtained polymer (LP1-6a) was thoroughly characterized by NMR, GPC, and MALDI-tof (Figure 4A and Figures S3, S39, and S40): the data collected confirmed the expected structure of the formed polymer (Table 1 and Scheme 2).

In contrast to the previous case which witnessed a rather fast propagation, with a weak base such as DBU, the polymerization rate was much slower (LP_2-6a) : the monomer conversion was indeed only 14% after 1 h, then 67% after 21 h, and it took 45 h of reaction to reach 90% of conversion (Figure 1 and Figure S41). As can be seen from Figure 1, the molar mass of the samples isolated over time increased linearly with the monomer

Article



Figure 2. (A) Plot of monomer conversion (%) (¹H NMR analysis) vs PDIs and M_n (GPC analysis) of reaction aliquots over time, (B) reaction kinetics plot: $\ln([M]_0/[M]_n)$ vs time, and (C) GPC profiles of reaction aliquots over same time for LP-6b.



Figure 3. (A) Plots of monomer conversion (%) (¹H NMR analysis) vs M_n and PDI (GPC analysis) of the reaction aliquots over time, (B) GPC profiles of the reaction mixture in same time, and (C) MALDI-tof of **CP-6a**.

conversion. Besides its characterization by NMR and GPC the final sample quenched with benzoic acid was also subjected to MALDI-tof analysis. As shown in Figure 1, MALDI-tof characterization confirms the incorporation of the initiator, *p*-methylbenzyl alcohol, at the chain end: the main population exactly matches the expected structure with a peak-to-peak mass difference corresponding to the molar mass of **6a** (248.23 g/mol). Another small population without any terminal group and thus corresponding to macrocyclic polycarbonates can also be seen; the latter are chains that underwent a ring closure reaction. In one of the next sections, we will discuss in detail how to obtain exclusively macrocyclic polyglycocarbonates from either **6a** or **6b**.

Hydrophilic and Amphiphilic Linear Polyglycocarbonates. After 6a the water-soluble monomer 6b was also subjected to polymerization using TBD as organic catalyst and p-methylbenzyl alcohol as initiator. The progress of the reaction was monitored by GPC, and aliquots of the reaction medium withdrawn over time were characterized by NMR. The monomer conversion was 54% after 30 min of reaction and 94% after 5 h of the reaction (Figure 2). In comparison to 6a, the polymerization of 6b was slower due to the hindering presence of the two bulky TEGM moieties. The isolated samples were lyophilized prior to their characterization by GPC, ¹H NMR, and ¹³C NMR spectroscopy (Figures S1A, S45, and \$46). The GPC traces showed single monodisperse profiles with no monomeric residues. From the signal due to the methyl group (at 2.4 ppm) in the ¹H NMR spectra to that of the initiator taken as reference M_{n(NMR)} values could be easily

deduced: as seen from the Table 1, they were in good agreement with the expected values.

One of our other endeavors was to synthesize an amphiphilic diblock copolymer containing both hydrophobic and hydrophilic polyglycocarbonate blocks. We first targeted the synthesis of a diblock copolymer with a composition prone to afford regular micelles in water. We thus used a hydrophilic poly(ethylene oxide) precursor (PEG-OH) as macroinitiator $(M_n = 5.0 \times 10^3 \text{ g/mol})$ and grew a hydrophobic polyglycocarbonate block of same weight fraction from it by polymerization of **6a**. The reaction was carried out in anhydrous dichloromethane at room temperature, and the catalyst used was TBD; the diblock copolymer synthesized was characterized by GPC and NMR (entry 06, Figures S1B and S42–S44) which confirmed the incorporation of PEG and the formation of the expected amphiphilic diblock copolymer.

We then tried to synthesize amphiphilic copolymers totally made of polyglycocarbonate blocks. Starting from the hydrophobic and hydrophilic monomers **6a** and **6b**, such amphiphilic copolymer could be generated by sequential polymerization of **6a** and **6b** in one pot as shown in Scheme 2. The first polymerization of **6a** was monitored by GPC and NMR. Then the second monomer **6b** was added after consumption of 90% of the first monomer **6a**. After 10 h of reaction, the polymerization of **6b** reached 92% conversion and that of **6a** was complete. The diblock copolyglycocarbonates obtained were purified by dialysis against Milli-Q water. Their GPC traces all exhibited a single monomodal peak clearly shifting to the high molar mass region with monomer conversion (Figure 4B and Figure S1C). The characteristic peaks due to the two monomers **6a** and **6b** *O*-methyl and *O*-TEG (3.6–3.3 ppm) could be well detected by ¹H NMR spectroscopy (Figure S47), the composition of the copolymer formed corresponding almost perfectly to the feeding ratio of monomers **6a** and **6b**.

Macrocyclic Polyglycocarbonates. As seen in Figure 1 in the presence of DBU used as catalyst initiation occurs by both *p*-methylbenzyl alcohol and DBU; the latter grows chains by a zwitterionic mechanism where both end-to-end ring closure reactions and propagation concomitantly occur (entry 02).²⁶ We thus used DBU alone as initiator to trigger the polymerization of 6a in anhydrous dimethylformamide (Scheme 2). The reaction proceeded very slowly, the monomer conversion calculated from NMR reaching only 5% after the first hour reaction and 13%, 51%, and 91% after 6, 21, and 45 h of reaction, respectively (Figure 3). As can be seen from Figures 3A and 3B the molar mass of the growing polymer (CP-6a) increased linearly with the increasing monomer conversion and PDIs values revolved around 1.2. The molar mass obtained from GPC was relatively low with respect to the value calculated from monomer conversion (¹H NMR analysis) and obtained from MALDI-tof (Figure 3C) likely due to the smaller hydrodynamic volume of the cyclic polymer formed (Figure 4A). The MALDI-tof spectrum of the sample isolated after 91%



Figure 4. GPC profiles of (A) LP₁-6a (linear) and CP-6a (macrocyclic) as well as (B) LP-6a-b-6b (linear) and CP-6a-b-6b (macrocyclic).

conversion contained indeed only one series of peaks corresponding exclusively to macrocyclic polycarbonates without end groups with a peak-to-peak mass difference corresponding perfectly to the monomer molar mass (248.5 g/mol). In addition, the average molar mass drawn from MALDI-tof (6700 g/mol) was in excellent agreement with the expected values. Further, the absence of DBU moieties or hydroxyl groups from NMR data also supports the formation of macrocyclic polycarbonates (Figure S49). Next, the polymerization behavior of water-soluble hydrophilic monomer 6b was also investigated. Because of the hindrance of TEGM groups in 6b, it took 5 days to reach 70% of monomer conversion with the same DBU organocatalyst. Upon adding in 1:1 molar ratio to DBU 1-(3,5-bis(trifluoromethyl)phenyl)-3-cyclohexylthiourea (TU), which is well-known for its ability to activate carbonyl groups in cyclic monomers (entry 05),³⁴⁻³⁸ the conversion of 6b reached 90% within 60 h without undesired back-biting side reactions. The obtained water-soluble macrocyclic polyglycocarbonate CP-6b was then subjected for detailed characterizations by NMR, GPC, and MALDI-tof. The MALDI-tof spectrum (Figure S4) of the polymer CP-6b exhibits peaks corresponding exclusively to macrocyclic polycarbonates without end groups and with a peak-to-peak mass difference corresponding perfectly to the monomer molar mass (512.5 g/mol). Further, the absence of DBU moieties or hydroxyl groups from NMR data also supports the formation of macrocyclic polycarbonates (Figures S50 and S51). PDI values obtained from GPC revolved around 1.2. The molar mass obtained from GPC was relatively low with respect to that given by MALDI-tof likely due to the smaller hydrodynamic volume of the cyclic polymer formed (Figure S1A).

Similarly, macrocyclic amphiphilic diblock polyglycocarbonates (**CP-6a-b-6b**) could also be prepared in a same manner as in the above case by subjecting sequentially **6a** and **6b** to ROP using DBU as initiator and TU as monomer activator. After consumption of 90% of the first monomer **6a** the second hydrophilic monomer **6b** was added to the reaction mixture. The polymerization was continued for another 3 days until reaching 90% conversion of **6b**. The GPC profile of this sample showed a monomodal signal, the value corresponding to 2/3 of the expected molar mass due to the lower apparent molar masses of macrocyclic structures (Figure 4B and Figure S1D). The ¹H NMR characterization confirmed the absence of any end group and the cyclic structure of the copolymer formed (Figures S52 and S53).

The mechanism (Scheme 3) involves first the ring-opening of the carbonate bond of monomer 6a or 6b by DBU, followed by repeated monomer addition by the zwitterionic species thus formed (zwitterionic intermediate). End-to-end ring closure reactions also occur but obviously at a much slower rate than propagation. The growing chains can thus undergo at any time a ring closure reaction and generate a macrocyclic poly-







Zwitterionic Intermediate



Figure 5. Cryo-TEM images of the nanostructures of the self-assemblies obtained from the amphiphilic polycarbonates (A) LP₃-6a, (B) LP-6a-*b*-6b, and (C) CP-6a-*b*-6b in water.

glycocarbonate, allowing the freed DBU to ring-open further monomers. Scheme 3 also features monomer activation by TU. A similar mechanism was proposed by Endo and co-workers in their work on the ROP of five-membered cyclic carbonates, but in their study they observed a mixture of macrocyclic species and linear chains.²⁶

Both linear and macrocyclic amphiphilic diblock copolymers were then subjected to self-assembling in water. The aggregates formed were then deposited on a carbon-coated copper-grid and analyzed by cryo-TEM.

All the amphiphilic samples exhibit micellar aggregation with an overall size varying from 20 to 40 nm depending upon the hydrophobic/hydrophilic balance and the overall macrostructure of the initial copolymers (Figure 5). Those with a macrocyclic architecture tended to self-assemble into micelles of bigger size due to entropic constrains. A paper focusing on the manipulation of the morphology of these self-assemblies as a function of the content in **6a** and **6b** and of the overall architecture is in preparation.

CONCLUSION

For the first time different types of bicyclic glycocarbonates both hydrophobic and hydrophilic could be prepared from synthetically modified glucose and CO2. Phosgene or its derivatives were purposely avoided in the chemistry described in this investigation which opens up the possibility to derive many more synthetically modified natural glycans. We also demonstrated the synthesis of both hydrophobic and hydrophilic linear polyglycocarbonates whose molar mass could be precisely controlled by using an organocatalyst combined with an initiator that mediated the ring-opening polymerization of the above-mentioned bicyclic glycocarbonates under "living" conditions. The chemistry described in this investigation was also versatile enough to allow the direct synthesis of macrocyclic polyglycocarbonates. Both linear and macrocyclic polyglycocarbonates were thoroughly characterized, and their structures were confirmed by GPC, MALDI-tof, ¹H NMR, and ¹³C NMR experiments. We also synthesized an amphiphilic diblock copolymer by using hydrophilic poly(ethylene glycol) (PEG-OH) as macroinitiator and amphiphilic diblock polyglycocarbonates by polymerizing first the hydrophobic monomer 6a followed by the hydrophilic monomer 6b in a very controlled manner. Both linear and macrocyclic amphiphilic structures could be obtained. In some instances TU was used to activate the cyclic glycocarbonate monomers and thus accelerate their ROP without provoking side reactions. Further, the self-assembling of the synthesized amphiphilic

polycarbonates were characterized by cryo-TEM which all showed micellar aggregation.

EXPERIMENTAL SECTION

Materials and Method. 1,8-Diazabicyclo[5.4.0]undec-7-ene (DBU), 1,5,7-triazabicyclo[4.4.0]dec-5-ene (TBD), and *p*-methylbenzyl alcohol (PMBA) were purchased from Sigma-Aldrich. DBU was dried with CaH₂ and stored in a glovebox after distillation. 1-(3,5-Bis(trifluoromethyl)phenyl)-3-cyclohexylthiourea (TU) was prepared following previously reported literature.³⁹ TBD was stirred in tetrahydrofuran with calcium hydride and filtered, and solvent was removed under vacuum and kept in a glovebox. Dichloromethane and dimethylformamide were dried with CaH₂ and stored on activated molecular sieves (4 Å) after distillation. CO₂ (99.995%) cylinder purchased from Abdullah Hashim Industrial & Gas Co. was further purified by flowing through the purifier of the VICI Co., USA. The synthesis and characterization of **5a** and **5b** are detailed in the Supporting Information.

FT-IR spectra were recorded on PerkinElmer FT-IR spectrum GX instrument. ¹H and ¹³C NMR spectra were recorded on Bruker spectrometers (400, 500, 600, and 950 MHz). Gel permeation chromatography (GPC) was performed on a VISKOTEK TDA 305-040 triple detector equipped with two columns (T6000M, GENERAL MIXED ORG 300X7.8 MM) using THF (1 mL/mL) as the eluent at 35 °C. MALDI-tof MS experiments were carried out by using trans-2-[3-(4-tert-butylphenyl)-2-methyl-2-propenylidene]malononitrile (DCTB) as the matrix in THF at a loading of 1:5 with Na-TFA as ionizing agent. The self-assembled morphology of amphiphilic polycarbonates in water was characterized by the transmission electron microscopy (cryo-TEM model Titan 80-300 Krios from FEI Company (Hillsboro, OR)) by operating it at the accelerating voltage of 300 keV. The specimens for cryo-TEM were prepared by using an automatic plunge-freezing system of model VitrobotTM from FEI Company (Hillsboro, OR). A small amount of sample (4 μ L) was placed on a holey-carbon coated hydrophilic copper grid and followed by blotting of the solution, and its plunge freezing into liquid ethane cryogen was done. Then, grids were loaded into the VitrobotTM. In this way, several specimens were prepared for each sample in order to realize a high quality data sets from superior specimens. Further, this microscope was equipped with the low-dose option to keep the sample free of electron beam damage and a charge coupled devices (CCD) of model US4000 from Gatan, Inc. to record the images in digital format. The polymer solution was prepared as following: The synthesized amphiphilic polyglycocarbonate was first dissolved in dimethyl sulfoxide, to which water was slowly added to get the final sample concentration of 1 mg/mL. Then the organic solvent was removed by continuous dialysis with Milli-Q water for 2 days.

Synthesis of α -Methyl-2,3-di-O-methyl-D-glucoside 4–6 Cyclic Carbonate (6a). α -Methyl-6-bromo-6-deoxy-2,3-di-O-methyl-D-glucopyranoside (5a, 0.5 g, 1.75 mmol), DBU (267 mg, 1.75 mmol), and 3 mL of anhydrous DMF were added into a 50 mL of dried autoclave with magnetic stirring bar inside a glovebox. Then the sealed autoclave was taken out and charged with CO₂ to a pressure of 10 bar. The reactor was kept stirring at room temperature for 5 h. Then, CO₂ was released; the reaction mixture was poured into HCl solution (1.0 M) and extracted by dichloromethane. The crude product was purified by silica gel flash chromatography eluted with a mixture of hexane/ethyl acetate (1:1–1:2) to give **6a** as a white solid (0.2 g 40% yield). FT-IR (dichloromethane) 1755 cm⁻¹ for sixmembered cyclic carbonate (ν_{co}). ¹H NMR (600 MHz, CD₂Cl₂): δ 4.93 (d, J = 3.6 Hz, 1H), 4.52 (dd, J = 9.8, 5.8 Hz, 1H), 4.28–4.22 (m, 1H), 4.13–4.02 (m, 2H), 3.66 (m, 4H), 3.54 (s, 3H), 3.49 (s, 3H), 3.31 (dd, J = 9.3, 3.6 Hz, 1H). ¹³C NMR (151 MHz, CD₂Cl₂): δ 147.27, 98.72, 81.06, 79.48, 79.13, 69.81, 60.92, 59.53, 59.05, 55.81. HRMS (M + H⁺) (ESI⁺) calcd for C₁₀H₁₆O₇H⁺ 249.09 g/mol; found 249.08 g/mol.

Synthesis of α -Methyl-2,3-di-O-TEGM-D-glucoside 4–6 Cyclic Carbonate (6b). α-Methyl-6-bromo-6-deoxy-2,3-di-O-TEGM-Dglucopyranoside (5b, 0.3 g, 546 µmol), DBU (91.44 mg, 600 mmol), and 4 mL of anhydrous DMF were added into a 50 mL of dried autoclave with magnetic stirring bar inside a glovebox. Then the sealed autoclave was taken out and charged with CO₂ to a pressure of 10 bar. The reactor was kept stirring at room temperature for 22 h. Then, CO₂ was released, and the reaction mixture was poured into HCl solution (1.0 M) and extracted by dichloromethane. The crude product was purified by silica gel flash chromatography eluted with a mixture of hexane/acetone (1:0.2-1:1) to give 6b as a colorless liquid (0.1 g, 40% yield). ¹H NMR (950 MHz, CDCl₃): δ 4.90 (d, J = 3.7 Hz, 1H), 4.46-4.44 m, 1H), 4.20-4.18 (m, 1H), 4.05-4.01 (m, 2H), 3.95-3.88 (m, 3H), 3.80 (ddd, J = 11.5, 7.6, 3.3 Hz, 1H), 3.75 (t, J = 9.0 Hz, 1H), 3.65-3.63 (m, 3H), 3.63-3.60 (m, 12 H), 3.59 (ddd, J = 10.9, 4.6, 3.4 Hz, 1H), 3.54–3.52 (m, 4H), 3.48 (dd, J = 9.4, 3.7 Hz, 1H), 3.45 (s, 3H), 3.37–3.34 (m, 6H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 147.36, 99.33, 79.97, 79.26, 78.21, 72.39, 71.85, 71.83, 71.73, 71.008, 70.54, 70.53, 70.48, 70.47, 70.45, 70.42, 70.33, 69.55, 59.42, 59.0, 58.98, 55.94 ppm. HRMS $(M + H^+)$ (ESI⁺) calcd for $C_{22}H_{40}O_{13}H^+$ 513.25 g/mol; found 513.25 g/mol.

Synthesis of Linear Polyglycocarbonates. Polymerization of 6 Using TBD and p-Methylbenzyl Alcohol. LP-6b is given as an example. Inside a glovebox, a solution of monomer 6b (71.32 mg, 139.15 μ mol) in 300 μ L of anhydrous dichloromethane was prepared in a Schlenk tube; a mixture of p-methyl benzyl alcohol (1 mg, 8.19 μ mol) and TBD (0.6 mg, 4.31 μ mol) in 100 μ L of anhydrous dichloromethane was then added to the monomer solution. After kept stirring inside the glovebox for 5 h, the polymerization was quenched with benzoic acid. The resulting suspension was dialyzed thoroughly against deionized water using 1 kDa cutoff cellulose membrane for 2 days to remove remaining monomers. The dialyzed solution was lyophilized, and 50 mg of LP-6b was obtained (yield 83%). FT-IR (dichloromethane) 1750 cm⁻¹ for carbonate (linear ν_{co}). ¹H NMR (400 MHz, CDCl₃): δ 4.90–4.81 (m, 1H), 4.79–4.52 (m, 1H), 4.50– 4.02 (m, 2H), 3.99-3.73 (m, 6H), 3.72-3.52 (m, 20H), 3.51-3.33 (m, 10H). Initiator incorporation was confirmed by the presence of 7.32-7.27 (m, 2H), 7.23-7.17 (m, 2H), 5.22-5.10 (m, 2H), 2.38 (s, 3H) ppm, $M_{n(NMR)}$ = 8300. ¹³C NMR (100 MHz, CDCl₃): δ 154.02, 154.20, 154.72, 97.98, 97.85, 79.97, 79.26, 78.21, 72.39, 71.85, 71.83, 71.73, 71.008, 70.54, 70.53, 70.48, 70.47, 70.45, 70.42, 70.33, 69.55, 59.42, 59.0, 58.98, 55.94 ppm. The incorporation of the initiator on the chain end was confirmed by the presence of the corresponding peaks; 128.50 (benzyl), 72.5 (CH2-benzyl) and 20.29 (Me-benzyl) ppm.

Synthesis of Linear Amphiphilic Block Polyglycocarbonate LP-6a-b-6b. Inside a glovebox, a solution of monomer 6a (36.57 mg, 147.34 μ mol) in 300 μ L of anhydrous dichloromethane was prepared in a Schlenk tube; a mixture of *p*-methylbenzyl alcohol (1 mg, 8.19 μ mol) and TBD (1.14 mg, 8.19 μ mol) in 100 μ L of anhydrous dichloromethane was then added to the monomer solution. The reaction was kept stirring inside the glovebox for 10 min; 90% of the monomer was consumed at this time. The second monomer, 6b (37.76 mg, 73.67 μ mol), was added to the reaction mixture. The polymerization continued for another 11 h and quenched by the addition of benzoic acid. The resulting suspension was dialyzed thoroughly against deionized water using 1 kDa cutoff cellulose

membrane for 2 days. Then the dialyzed solution was lyophilized to afford the amphiphilic block copolymer, LP-6a-b-6b as colorless liquid. (60 mg, yield 75%). FT-IR (dichloromethane) 1750 cm⁻¹ for carbonate (linear ν_{co}). ¹H NMR (400 MHz, CDCl₃): δ 4.88–4.76 (m, 1H), 4.75-4.49 (m, 1H), 4.46-3.98 (m, 2H), 3.96-3.67 (m, 3H), 3.66-3.45 (m, 10H), 3.45-3.30 (m, 5H), 3.30-3.16 (m, 1H). The initiator incorporation at the chain end was confirmed (pmethylbenzyl alcohol) by the presence of 7.28-7.22 (m, 2H), 7.15-7.12 (m, 2H), 5.17–5.05 (m, 2H), 2.32 (s, 3H) ppm and $M_{n(NMR)}$ = 8600. ¹³C NMR (100 MHz, CDCl₃): δ 154.02, 154.20, 154.72, 97.98, 97.85, 79.97, 79.26, 78.21, 72.39, 71.85, 71.83, 71.73, 71.008, 70.54, 70.53, 70.48, 70.47, 70.45, 70.42, 70.33, 69.55, 59.42, 59.0, 58.98, 55.94 ppm. Initiator incorporation was confirmed by the presence of the corresponding peaks (p-methylbenzyl alcohol); 129.19, 128.50 (benzyl), 72.5 (CH2-benzyl) and 20.29 (Me-benzyl) ppm.

Synthesis of Macrocyclic Polyglycocarbonate (CP-6a) by the Polymerization of 6a. Inside a glovebox, a solution of monomer 6a (195.06 mg, 786 μ mol) in 800 μ L of anhydrous DMF was prepared in a Schlenk tube; DBU (3.98 mg, 26.20 μ mol) was dissolved in 100 μ L of deuterated DMF and added to the monomer solution. The polymerization was kept stirring inside the glovebox for 48 h and then quenched with benzoic acid. Finally, the reaction mixture was precipitated in water; the collected crude product was dissolved in CH2Cl2 and precipitated again in diethyl ether to afford the macrocyclic polymer CP-6a (126 mg, yield 65%). ¹H NMR (400 MHz, CDCl₃): δ 4.73-4.61 (m, 1H), 4.61-4.27 (m, 1H), 4.17-3.93 (m, 2H), 3.80-3.62 (m, 1H), 3.50-3.40 (m, 1H), 3.39-3.28 (m, 6H), 3.27-3.18 (m, 3H), 3.16-3.05 (m, 1H) ppm. ¹³C NMR (151 MHz, CDCl₃): *δ* 155.1, 154.8, 154.46, 97.92, 81.7, 80.8, 78.1, 75.1, 67.8, 66.6, 61.33, 61.1, 59.1, 55.8 ppm. FT-IR (dichloromethane) 1750 cm⁻¹ for carbonate (linear $\nu_{\rm CO}$).

Synthesis of Macrocyclic Polyglycocarbonate (CP-6b) by the Polymerization of 6b. Inside a glovebox, a solution of monomer 6b (87 mg, 169.74 μ mol) in 400 μ L of anhydrous DMF was prepared in a Schlenk tube; DBU (2.15 mg, 14.14 µmol) and 1-(3,5-bis(trifluoromethyl)phenyl)-3-cyclohexylthiourea (TU; 5.24 mg, 14.14 μ mol) were dissolved in 100 μ L of deuterated DMF and added to the monomer solution. The polymerization was kept stirring inside the glovebox for 60 h and then quenched with benzoic acid. The obtained polymerization mixture was diluted with deionized water and dialyzed against water using 1 kDa cutoff cellulose membrane for 2 days. The dialyzed solution was lyophilized to give the cyclic polyglycocarbonate, CP-6b as colorless liquid (52 mg, yield 60%). ¹H NMR (400 MHz, CDCl₃): δ 4.90–4.781 (m, 1H), 4.79–4.50 (m, 1H), 4.50–4.0 (m, 2H), 3.99-3.70 (m, 6H), 3.72-3.50 (m, 20H), 3.51-3.30 (m, 10H). ¹³C NMR (100 MHz, CDCl₃): δ 154.02, 154.20, 154.72, 97.98, 97.85, 80.42, 79.44, 74.87, 72.39, 71.85, 71.83, 71.73, 71.008, 70.54, 70.53, 70.48, 70.47, 70.45, 70.42, 70.33, 69.55, 59.42, 59.0, 58.98, 55.94 ppm. FT-IR (dichloromethane) 1750 cm⁻¹ for carbonate (linear ν_{CO}).

Synthesis of Macrocyclic Amphiphilic Polyglycocarbonate (CP-6a-b-6b). Inside a glovebox, a solution of monomer 6a (42.80 mg, 172.80 μ mol) in 300 μ L of anhydrous DMF was prepared in a Schlenk tube; DBU (1.75 mg, 11.49 μ mol) dissolved in 100 μ L of deuterated dimethylformamide (DMF) was added to the monomer solution. The reaction was kept stirring inside the glovebox for 28 h; 90% of the monomer was consumed at this time. Then the second monomer, 6b (41.24 mg, 80.46 µmol), and 1-(3,5-bis(trifluoromethyl)phenyl)-3-cyclohexylthiourea (TU; 4.26 mg, 11.49 μ mol) were added to the reaction mixture. The polymerization continued for another 3 days before being quenched by a few drops of HCl (1 N). The obtained polymerization mixture was diluted with deionized water and dialyzed against water using 1 kDa cutoff cellulose membrane for 2 days. The dialyzed solution was lyophilized to give the amphiphilic cyclic block copolymer CP-6a-b-6b as colorless liquid (60 mg, yield 75%). ¹H NMR (400 MHz, CDCl₃): δ 4.90–4.78 (m, 1H), 4.77–4.51 (m, 1H), 4.50-4.09 (m, 2H), 4.01-3.67 (m, 2.18 H), 3.66-3.47 (m, 10H), 3.46-3.32 (m, 5H), 3.32-3.15 (m, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 154.02, 154.20, 154.72, 97.98, 97.85, 79.97, 79.26, 78.21, 72.39, 71.85, 71.83, 71.73, 71.008, 70.54, 70.53, 70.48, 70.47,

70.45, 70.42, 70.33, 69.55, 59.42, 59.0, 58.98, 55.94 ppm. FT-IR (dichloromethane) 1750 cm⁻¹ for carbonate (linear ν_{CO}).

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.macro-mol.6b02527.

Synthesis details of the precursors **5a** and **5b** for bicyclic monomers **6**, the characterizations of **2–5**, linear and cyclic polyglycocarbonates, Figures S1–S53 (PDF)

AUTHOR INFORMATION

Corresponding Authors

*E-mail: fxs101@gmail.com (X.F.).

*E-mail: yves.gnanou@kaust.edu.sa (Y.G.).

ORCID

Nikos Hadjichristidis: 0000-0003-1442-1714 Yves Gnanou: 0000-0001-6253-7856

Notes

The authors declare no competing financial interest.

REFERENCES

(1) Galbis, J. A.; García-Martín, M. d. G.; de Paz, M. V.; Galbis, E. Synthetic Polymers from Sugar-Based Monomers. *Chem. Rev.* 2016, *116*, 1600–1636.

(2) Ladmiral, V.; Mantovani, G.; Clarkson, G. J.; Cauet, S.; Irwin, J. L.; Haddleton, D. M. Synthesis of Neoglycopolymers by a Combination of "Click Chemistry" and Living Radical Polymerization. *J. Am. Chem. Soc.* **2006**, *128*, 4823–4830.

(3) Ladmiral, V.; Melia, E.; Haddleton, D. M. Synthetic glycopolymers: an overview. *Eur. Polym. J.* **2004**, *40*, 431–449.

(4) Okada, M. Molecular design and syntheses of glycopolymers. *Prog. Polym. Sci.* 2001, *26*, 67–104.

(5) Pati, D.; Shaikh, A. Y.; Das, S.; Nareddy, P. K.; Swamy, M. J.; Hotha, S.; Gupta, S. S. Controlled Synthesis of O-Glycopolypeptide Polymers and Their Molecular Recognition by Lectins. *Biomacromolecules* **2012**, *13*, 1287–1295.

(6) Spain, S. G.; Gibson, M. I.; Cameron, N. R. Recent advances in the synthesis of well-defined glycopolymers. J. Polym. Sci., Part A: Polym. Chem. 2007, 45, 2059–2072.

(7) Murrey, H. E.; Hsieh-Wilson, L. C. The Chemical Neurobiology of Carbohydrates. *Chem. Rev.* **2008**, *108*, 1708–1731.

(8) Pati, D.; Kalva, N.; Das, S.; Kumaraswamy, G.; Sen Gupta, S.; Ambade, A. V. Multiple Topologies from Glycopolypeptide–Dendron Conjugate Self-Assembly: Nanorods, Micelles, and Organogels. *J. Am. Chem. Soc.* **2012**, *134*, 7796–7802.

(9) Voit, B.; Appelhans, D. Glycopolymers of Various Architectures—More than Mimicking Nature. *Macromol. Chem. Phys.* 2010, 211, 727–735.

(10) Wells, L.; Vosseller, K.; Hart, G. W. Glycosylation of Nucleocytoplasmic Proteins: Signal Transduction and O-GlcNAc. *Science* **2001**, *291*, 2376–2378.

(11) Dane, E. L.; Grinstaff, M. W. Poly-amido-saccharides: Synthesis via Anionic Polymerization of a β -Lactam Sugar Monomer. J. Am. Chem. Soc. **2012**, 134, 16255–16264.

(12) Lavilla, C.; Alla, A.; Martínez de Ilarduya, A.; Benito, E.; Garcia-Martin, M. G.; Galbis, J. A.; Munoz-Guerra, S. Carbohydrate-based polyesters made from bicyclic acetalized galactaric acid. *Biomacromolecules* **2011**, *12*, 2642–2652.

(13) Lavilla, C.; Alla, A.; de Ilarduya, A. M.; Benito, E.; García-Martín, M. G.; Galbis, J. A.; Muñoz-Guerra, S. Carbohydrate-based copolyesters made from bicyclic acetalized galactaric acid. *J. Polym. Sci., Part A: Polym. Chem.* **2012**, *50*, 1591–1604.

(15) Klemm, D.; Heublein, B.; Fink, H.-P.; Bohn, A. Cellulose: Fascinating Biopolymer and Sustainable Raw Material. *Angew. Chem., Int. Ed.* **2005**, *44*, 3358–3393.

(16) Kobayashi, S.; Kashiwa, K.; Kawasaki, T.; Shoda, S. Novel method for polysaccharide synthesis using an enzyme: the first in vitro synthesis of cellulose via a nonbiosynthetic path utilizing cellulase as catalyst. *J. Am. Chem. Soc.* **1991**, *113*, 3079–3084.

(17) Kobayashi, S.; Kiyosada, T.; Shoda, S.-i. Synthesis of Artificial Chitin: Irreversible Catalytic Behavior of a Glycosyl Hydrolase through a Transition State Analogue Substrate. *J. Am. Chem. Soc.* **1996**, *118*, 13113–13114.

(18) Kricheldorf, H. R.; Sun, S.-J.; Chen, C.-P.; Chang, T.-C. Polymers of carbonic acid. XXIV. Photoreactive, nematic or cholesteric polycarbonates derived from hydroquinone-4-hydroxybenzoate 4,4'-dihydroxychalcone and isosorbide. *J. Polym. Sci., Part A: Polym. Chem.* **1997**, 35, 1611–1619.

(19) Scheible, W.-R.; Pauly, M. Glycosyltransferases and cell wall biosynthesis: novel players and insights. *Curr. Opin. Plant Biol.* 2004, 7, 285–295.

(20) Boltje, T. J.; Buskas, T.; Boons, G.-J. Opportunities and challenges in synthetic oligosaccharide and glycoconjugate research. *Nat. Chem.* **2009**, *1*, 611–622.

(21) Ting, S. R. S.; Chen, G.; Stenzel, M. H. Synthesis of glycopolymers and their multivalent recognitions with lectins. *Polym. Chem.* **2010**, *1*, 1392–1412.

(22) Chen, X.; Gross, R. A. Versatile Copolymers from [l]-Lactide and [d]-Xylofuranose. *Macromolecules* **1999**, *32*, 308–314.

(23) Acemoglu, M.; Bantle, S.; Mindt, T.; Nimmerfall, F. Novel bioerodible poly(hydroxyalkylene carbonates)s: a versatile class of polymers for medical and pharmaceutical applications. *Macromolecules* **1995**, *28*, 3030–3037.

(24) Shen, Y.; Chen, X.; Gross, R. A. Aliphatic Polycarbonates with Controlled Quantities of d-Xylofuranose in the Main Chain. *Macromolecules* **1999**, *32*, 3891–3897.

(25) Haba, O.; Tomizuka, H.; Endo, T. Anionic Ring-Opening Polymerization of Methyl 4,6-O-Benzylidene-2,3-O- carbonyl- α -dglucopyranoside: A First Example of Anionic Ring-Opening Polymerization of Five-Membered Cyclic Carbonate without Elimination of CO₂. *Macromolecules* **2005**, *38*, 3562–3563.

(26) Azechi, M.; Matsumoto, K.; Endo, T. Anionic ring-opening polymerization of a five-membered cyclic carbonate having a glucopyranoside structure. *J. Polym. Sci., Part A: Polym. Chem.* **2013**, *51*, 1651–1655.

(27) Mikami, K.; Lonnecker, A. T.; Gustafson, T. P.; Zinnel, N. F.; Pai, P. J.; Russell, D. H.; Wooley, K. L. Polycarbonates derived from glucose via an organocatalytic approach. *J. Am. Chem. Soc.* **2013**, *135*, 6826–6829.

(28) Gustafson, T. P.; Lonnecker, A. T.; Heo, G. S.; Zhang, S.; Dove, A. P.; Wooley, K. L. Poly(d-glucose carbonate) Block Copolymers: A Platform for Natural Product-Based Nanomaterials with Solvothermatic Characteristics. *Biomacromolecules* **2013**, *14*, 3346–3353.

(29) García-Martín, M. G.; Pérez, R. R.; Hernández, E. B.; Espartero, J. L.; Muñoz-Guerra, S.; Galbis, J. A. Carbohydrate-Based Polycarbonates. Synthesis, Structure, and Biodegradation Studies. *Macromolecules* **2005**, *38*, 8664–8670.

(30) Sapich, B.; Stumpe, J.; Krawinkel, T.; Kricheldorf, H. R. New Polymer Syntheses. 95. Photosetting Cholesteric Polyesters Derived from 4-Hydroxycinnamic Acid and Isosorbide. *Macromolecules* **1998**, *31*, 1016–1023.

(31) Haba, O.; Tomizuka, H.; Endo, T. Anionic ring-opening polymerization of methyl 4,6-O-benzylidene-2,3- ocarbonyl-a-D-glucopyranoside: A first example of anionic ring-opening polymerization of five-membered cyclic carbonate without elimination of CO 2. *Macromolecules* **2005**, *38*, 3562–3563.

(32) Suriano, F.; Pratt, R.; Tan, J. P.; Wiradharma, N.; Nelson, A.; Yang, Y. Y.; Dubois, P.; Hedrick, J. L. Synthesis of a family of

Macromolecules

amphiphilic glycopolymers via controlled ring-opening polymerization of functionalized cyclic carbonates and their application in drug delivery. *Biomaterials* **2010**, *31*, 2637–2645.

(33) Gregory, G. L.; Jenisch, L. M.; Charles, B.; Kociok-Kohn, G.; Buchard, A. Polymers from Sugars and CO_2 : Synthesis and Polymerization of a D-Mannose-Based Cyclic Carbonate. *Macromolecules* **2016**, *49*, 7165–7169.

(34) Kamber, N. E.; Jeong, W.; Waymouth, R. M.; Pratt, R. C.; Lohmeijer, B. G. G.; Hedrick, J. L. Organocatalytic Ring-Opening Polymerization. *Chem. Rev.* **200**7, *107*, 5813–5840.

(35) Ottou, W. N.; Sardon, H.; Mecerreyes, D.; Vignolle, J.; Taton, D. Update and challenges in organo-mediated polymerization reactions. *Prog. Polym. Sci.* 2016, *56*, 64–115.

(36) Fèvre, M.; Vignolle, J.; Gnanou, Y.; Taton, D. 4.06 -Organocatalyzed Ring-Opening Polymerizations. In *Polymer Science: A Comprehensive Reference;* Möller, K. M., Ed.; Elsevier: Amsterdam, 2012; pp 67–115.

(37) Thomas, C.; Bibal, B. Hydrogen-bonding organocatalysts for ring-opening polymerization. *Green Chem.* **2014**, *16*, 1687–1699.

(38) Kiesewetter, M. K.; Shin, E. J.; Hedrick, J. L.; Waymouth, R. M. Organocatalysis: Opportunities and Challenges for Polymer Synthesis. *Macromolecules* **2010**, *43*, 2093–2107.

(39) Pratt, R. C.; Lohmeijer, B. G. G.; Long, D. A.; Lundberg, P. N. P.; Dove, A. P.; Li, H.; Wade, C. G.; Waymouth, R. M.; Hedrick, J. L. Exploration, Optimization, and Application of Supramolecular Thiourea–Amine Catalysts for the Synthesis of Lactide (Co)polymers. *Macromolecules* **2006**, *39*, 7863–7871.