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Palladium-Catalyzed Solvent-Controlled Selective Synthesis of Acyl Isoureas and Imides from Amides, Isocyanides, Alcohols and Carboxylates

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Abstract: A highly selective synthesis of acyl isoureas and imides from readily accessible amides, isocyanides, alcohols and carboxylates based on reaction solvent selection is described. In the presence of a catalytic amount of [1,1]bis(diphenylphosphino)ferrocene]dichloropalladium(II) and cupric acetate, treatment of amides and isocyanides in alcohols at 60 °C provided acyl isoureas in high yields. Interestingly, when other solvents such as acetonitrile was used instead of alcohols, imides were exclusively produced in good to excellent yields *via* direct *N*-acylation of amides with carboxylates as the acyl sources. This protocol offers an attractive alternative approach toward isoureas and imides.

Introduction

Isourea moiety is prevalent functional group widely existing in not only agrochemicals but also pharmaceuticals, functional materials, and other versatile intermediates.^[1] Owing to their wide occurrence and importance, the methods for the synthesis of isoureas are of high interest. To date, the reported methods for the synthesis of isoureas are highly concentrated on intermolecular addition of alcohols to carbodiimides catalyzed by copper salts,^[2] zinc salts,^[3] or actinide complexes^[4] (Scheme 1a). Although these methods have the advantages of being atom-efficient, there are limitations, such as limited substrate availability, comparatively complicated catalyst species and a relatively long reaction time. In addition, intramolecular amino-oxylation of alkenes also provided access to bicyclic isoureas by the use of a catalytic amount of Pt,^[5] Ag,^[6] or excess amount of Lewis acids (Scheme 1b).^[7] Moreover, Shi and coworkers reported the Cu-catalyzed intramolecular amino- versus oxy-acetoxylation of N-allyl ureas for the efficient synthesis of cyclic isoureas (Scheme 1c).^[8] However, in most of these methods, a isocyanides; acyl isoureas; imides

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stoichiometric or excess amount of strong Lewis acids, oxidants or additives, or special preparation of the substrates is required. Hence, development of new and efficient strategies for the synthesis of isoureas is still highly attractive, especially when their direct access from simple substrates could be secured under mild conditions. Recently, Ji et al. developed a Co(II)-catalyzed synthesis of sulfonyl isoureas by the reactions of sulfonyl azides with isocyanides in alcohols via nitrene intermediate, which provided an alternative route to isoureas.^[9] Herein, we would like to report a novel and efficient methodology for the synthesis of acyl isoureas from readily available amides, isocyanides and alcohols. In the presence of a catalytic amount of $Pd(dppf)Cl_2$ and $Cu(OAc)_2$, acyl isoureas were synthesized in high yields by treating primary amides with isocyanides in alcohols at 60 °C for 2 h. Interestingly, when other solvents such as CH₃CN, was used instead of alcohols, imides were exclusively produced in good to excellent yields via N-acylation of primary amides. Despite numerous methods including acylation of amides with carboxylic acids derivatives,^[10] oxidation of *N*alkylamides or oxazoles,^[11] coupling of amides with

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Scheme 1. Comparison of previous study and our work.

Results and Discussion

We selected benzamide (1a) as the model compound to examine its behavior under different conditions (Table 1). After extensive screenings, the optimized reaction conditions for the formation of acyl isourea (2a) was obtained when a mixture of 1a and tertbutyl isocyanide (1:1 mol ratio) was treated with 10 mol % Pd(dppf)Cl₂ and 40 mol % Cu(OAc)₂ in ethanol at 60 °C for 2 h (Table 1, entry 1). It was found that both Pd catalyst and Cu(OAc)₂ were necessary for the successful transformation, since in the absence of Pd(dppf)Cl₂ or Cu(OAc)₂, no desired product 2a was detected (Table 1, entries 2-3). Other Pd salts also showed good catalytic activity (Table 1, entries 4-7), whereas other Cu salts, such as CuCl₂, CuSO₄, CuI or CuO, were not effective for this conversion (Table 1, entries 8-11). 10 mol % of the Pd(dppf)Cl₂ and 40 mol % of Cu(OAc)₂ were sufficient to promote the reaction effectively, and the vield of 2a was not improved when 20 mol % of the $Pd(dppf)Cl_2$ and 1 equiv of $Cu(OAc)_2$ were used (Table 1, entry 12). However, decreasing the loading amount of Cu(OAc)₂ reduced the yield to 56% (Table 1, entry 13). It was noted that the reaction could also lead to 2a in 87% yield when the 1:1 ratio of ethanol and CH₃CN solvent mixtures was used instead of ethanol (Table 1, entry 14). Interestingly, when 1a, tert-butyl isocyanide and stoichiometric ethanol were subjected to CH₃CN, along with 50% yield of 2a, a white solid was unexpectedly generated in 35%

isolated yield, which was characterized as N-acetylbenzamide **3a** on the basis of its spectral and analytical data (Table 1, entry 15).

Table 1. Optimization of reaction conditions.[a]

$NH_2 + Bu'-N=C + OH \frac{Catalyst, Additive}{Solvent 60, 95, 2b} + H + H + H + H + H + H + H + H + H + $					
\checkmark	1a	00110111,000 0,211	2a	\checkmark	3a
Entr y	Pd Catalyst (mol%)	Additive (mol%)	Solvent	Yields of 2a (%) ^b	Yields of 3a (%) ^b
1	Pd(dppf)Cl ₂ (10)	Cu(OAc) ₂ (40)	EtOH	92	0
2	$Pd(dppf)Cl_2(10)$	none	EtOH	0	0
3	none	Cu(OAc) ₂ (40)	EtOH	0	0
4	Pd(OAc) ₂ (10)	Cu(OAc) ₂ (40)	EtOH	89	0
5	$PdCl_{2}(10)$	$Cu(OAc)_2(40)$	EtOH	87	0
6	$Pd(CF_3COO)_2(10)$	$Cu(OAc)_2(40)$	EtOH	84	0 _
7	$Pd(PPh_3)_2Cl_2(10)$	$Cu(OAc)_2(40)$	EtOH	82	С
8	Pd(dppf)Cl ₂ (10)	CuCl ₂ (40)	EtOH	0	0
9	$Pd(dppf)Cl_2(10)$	$CuSO_4(40)$	EtOH	<5%	0
10	$Pd(dppf)Cl_2(10)$	CuI (40)	EtOH	0	0
11	$Pd(dppf)Cl_2(10)$	CuO (40)	EtOH	0	^
12	$Pd(dppf)Cl_2(20)$	$Cu(OAc)_2(100)$	EtOH	92	0
13	Pd(dppf)Cl ₂ (10)	Cu(OAc) ₂ (20)	EtOH	56	0
14	Pd(dppf)Cl ₂ (10)	Cu(OAc) ₂ (40)	CH ₃ CN/ EtOH (1:1)	87	<5
15 ^c	$Pd(dppf)Cl_2(10)$	$Cu(OAc)_2(40)$	CH ₃ CN	50	35

^[a] *Reactions conditions*: benzamide **1a** (0.5 mmol), *tert*-butyl isocyanide (0.5 mmol), Pd catalyst (0.05 mmol, 10 mol%), additive (0.2 mmol, 40 mol%), solvent (2.0 mL), 60 °C, 2 h (except for entries 2-3 and 12-13). Entry in bold highlights optimized reaction conditions.
^[b] Isolated yield of pure product based on **1a**.

Having identified the optimal conditions for the synthesis of acyl isoureas 2a, a range of reactions of amides 1 and isocyanides in alcohols were carried out aiming to determine the scope of the acyl isoureas synthesis. As summarized in Table 2, wide ranges of benzamides with different substituents or substitution patterns on the phenyl ring worked effectively to afford the corresponding acyl isoureas (2a-j) in excellent yields (81-95%), and the structure of 2j was further confirmed by X-ray crystallography analysis, as described in Figure 1. The 1-naphthamide and thiophene-2-carboxamide gave the pruducts 2k and 2l in 92% and 85% yields, respectively. Besides, cinnamamide and 2-phenylacetamide also exhibited good reactivities in this transformation, delivering the target products 2m and 2n in 75% and 78% yields, respectively. Unfortunately, only trace amount of product could be detected when acrylamide was applied in the reaction (20). To further explore the diversity of products, the scope of alcohols and isocyanides were also investigated. The reactions of 1a with *tert*-butyl isocyanide in different alcohols, such as butan-1-ol, 3-methylbutan-1-ol, cyclohexanol and benzyl alcohol, proceeded readily to produce the desired acyl isoureas 2p-s in high yields. However, when tert-butanol was used as the solvent, along with 30% yield of imide 3a, only trace amount of isourea 2t could be detected, other compounds, such as phenol and trifluoroethanol did not give any products. In case of isocyanides, the reaction of **1a** with cyclohexyl isocyanide, 1,1,3,3-tetramethylbutyl isocyanide, benzyl isocyanide and 2,6dimethylphenyl isocyanide in ethanol performed well, affording the corresponding acyl isoureas **2u-x** in excellent yields.

Table 2. Substrate scope for the reaction of amides 1 and isocyanides in alcohols.^[a,b]



^[a] *Reactions conditions*: 0.5 mmol of **1** and 0.5 mmol of isocyanides in the presence of Pd(dppf)Cl₂ (10 mol %) and Cu(OAc)₂ (40 mol %) in 2.0 mL of alcohols at 60 °C for 2 h.

^[b] Isolated yield of pure product based on **1**.



Figure 1. X-ray crystal structure of acyl isourea 2j (CCDC 1868295).

ubiquitous various Imide bond is in pharmaceuticals, natural products, and polymers,^[15] and the above findings led us to envisage that the selective synthesis of imides via direct N-acylation of amides using carboxylates as acyl sources may be realized. Thus, the optimization of the imides synthesis was also investigated. After a series of experiments, the optimized reaction conditions for the formation of imide 3a was obtained when a mixture of **1a**, Cu(OAc)₂ and *tert*-butyl isocyanide (1:1:1 mol ratio) was treated with 10 mol % Pd(dppf)Cl₂ in CH₃CN at 60 °C for 2 h, whereby N-acetylbenzamide **3a** was obtained in 82% isolated yield (see Table S1, entry 1 in the Supporting Information).

Subsequently, the substrate scope of this Nacylation of primary amides with carboxylates was explored and the results are summarized in Table 3. A variety of benzamides bearing several substituents (-Me, -Cl, -F, -I, -CF₃, -OMe, -OEt, -NH₂) at different positions of the benzene ring fairly tolerated the reaction conditions, giving the desired imides **3a-m** in good to high yields. In general, primary amides possessing electron-withdrawing groups on the phenyl ring provided higher yields than those bearing electron-donating groups (3c-e vs. 3f-g). Notably, sensitive functionalities such as $-NH_2(3f)$ and -I(3i)were unaffected under the present reaction conditions. addition, 1-naphthamide and thiophene-2-In carboxamide also proceeded smoothly, leading to imides **3n** and **3o** in 87% and 74% yields, respectively. Cinnamamide also reacted with $Cu(OAc)_2$, delivering the target product **3p** in 68% yield. Unfortunately, the simple alkyl-substituted primary amides failed to generate the desired the current conditions. products under The carboxylate substrates were further investigated, and to our delight, various carboxylates, such as sodium benzoate, copper(II) methacrylate and copper bis(2ethylhexanoate), reacted successfully and afforded the corresponding products (**3q-s**) in good yields.

Table 3. Substrate scope for the reaction of amides 1 and carboxylates in CH_3CN .^[a,b]



- ^[a] *Reactions conditions*: 0.5 mmol of 1 and 0.5 mmol of Cu(OAc)₂ in the presence of Pd(dppf)Cl₂ (10 mol %) and *tert*-butyl isocyanide (1.0 equiv.) in 2.0 mL of CH₃CN at 60 °C for 2 h.
- ^[b] Isolated yield of pure product based on **1**.
- ^[c] Sodium benzoate was used as the carboxylate.
- ^[d] Copper(II) methacrylate was used as the carboxylate.
- ^[e] Copper bis(2-ethylhexanoate) was used as the carboxylate.

To test the practicality of the method, gram-scale (10 mmol-scale) synthesis of acyl isourea **2a** and imide **3a** were carried out. The reaction of benzamide (**1a**) and *tert*-butyl isocyanide in 20 mL of ethanol on the 10 mmol scale gave the desired product **2a** in 84% yield (Scheme 2a). When **1a**, cupric acetate and

tert-butyl isocyanide (1:1:1 mol ratio) in 20 mL of acetonitrile were used as starting materials, the reaction proceeded efficiently to obtain 1.63 g of **3a** (77% isolated yield) (Scheme 2b).



Scheme 2. Gram-scale synthesis of acyl isourea 2a and imide 3a.

Some control experiments were carried out in order to explore the possible reaction pathway. The reaction of **1a** with *tert*-butyl isocyanide in ultradry EtOH under Argon atmosphere provided 2a in 91% yield (Scheme 3a). The reaction of 1a and $Cu(OAc)_2$ in the presence of 10 mol % Pd(dppf)Cl₂ and 1.0 equivalent of tert-butyl isocyanide in ultradry CH₃CN under Argon atmosphere generated 3a in 83% yield (Scheme 3b). In addition, no ¹⁸O-labeled product $[^{18}O]$ -2a or $[^{(18}O]$ -3a were detected in the presence of $H_2^{18}O$ under the standard conditions (Scheme 3c and 3d). The above results indicated that the oxygen atom of the acyl isourea and imide product was not originated from H₂O or molecular oxygen. Furthermore, the yields of acyl isourea 2a and imide **3a** were nearly unaffected in reactions conducted in the presence of radical scavengers TEMPO or BHT (2 equiv), which argues against a radical pathway (Scheme 3e and 3f).

On the basis of the above results, a proposed mechanism is listed in Scheme 4. The isocyanideligated Pd species **A** is first formed,^[16] then reacts with amide **1** to give complex **B**. In alcohols, the reaction of the complex **B** with alcohol affords acyl isourea **2** and the resultant Pd(0) species is reoxidized to Pd(II) by Cu(II) in the system to complete the cycle.^[17] Whereas in CH₃CN, the complex **B** reacts with carboxylates to generate the Pd(0) species and intermediate **C**, which undergoes rearrangement to form intermediate **D**. Immediately, the unstable intermediate **D** conducts decomposition to furnish the final imide product **3**.





Scheme 4. Proposed mechanism for the formation of acyl isoureas and imides.

Conclusion

In summary, we have developed a facile and efficient synthesis of imides and acyl isoureas from readily accessible amides, carboxylates and isocyanides in a highly selective manner by variation of the reaction solvent. In addition, this transformation also features good yields, good functional group tolerance and experimentally convenient catalytic process. Further, study on extension of the scope of the present protocol to construct the biologically active molecules is ongoing in our laboratory.

Experimental Section

General Experimental Details

Proton nuclear magnetic resonance spectra (¹H NMR) and carbon nuclear magnetic resonance spectra (¹³C NMR) were recorded at 400 MHz and 100 MHz, respectively. HRMS (ion trap) were recorded using ESI. Melting points are uncorrected. Precoated silica gel plates F-254 were

used for thin-layer analytical chromatography. Column chromatography was performed on silica gel (300-400 mesh). Unless otherwise noted, all reagents were obtained commercially and used without further purification.

General experimental procedure for synthesis of acyl isoureas 2.

A mixture of amides **1** (1.0 equiv. 0.5 mmol), isocyanides (1.0 equiv, 0.5 mmol), $Pd(dppf)Cl_2$ (10 mol %, 0.05 mmol), $Cu(OAc)_2$ (40 mol %, 0.2 mmol) and 2 mL of alcohols was stirred at 60 °C for 2 h. The progress of the reaction was monitored by thin-layer chromatography. The mixture was then cooled and evaporated under reduced pressure, and the residue was purified by flash column chromatography (petroleum ether/ethyl acetate) to afford acyl isoureas 2.

(Z)-1-benzoyl-3-tert-butyl-2-ethylisourea (2a):

(Z)-1-benzoyl-3-tert-butyl-2-ethylisourea (2a): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.22 (s, 1H), 8.33–8.12 (m, 2H), 7.50–7.35 (m, 3H), 4.56 (q, J = 7.1 Hz, 2H), 1.44–1.39 (m, 12H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 177.3, 162.6, 138.1, 131.4, 129.1, 128.3, 127.9, 63.4, 51.8, 29.5, 14.6 ppm; HRMS (*m*/z) (ESI): calcd for C₁₄H₂₁O₂N₂249.1598 [M+H]⁺; found 249.1586. (Z)-1-tert-butyl-2-ethyl-3-(4-fluorobenzoyl)isourea (2b): colorless oil; ¹H NMR (400 MHz, DMSO): δ 10.12 (s, 1H), 8.21–8.05 (m, 2H), 7.05 (s, 2H), 4.48 (q, J = 7.1 Hz, 2H), 1.39–1.33 (m, 12H) ppm; ¹³C NMR (100 MHz, DMSO): δ 175.4, 164.8 (d, ¹J_{CF} = 248 Hz), 162.5, 134.5, 131.5 (d, ³J_{CF} = 8.0 Hz), 114.90, 114.8 (d, ⁴J_{CF} = 2.0 Hz), 63.5, 51.7, 29.5, 14.6 ppm; HRMS (*m*/z) (ESI): calcd for C₁₄H₂₀O₂N₂F 267.1503 [M+H]⁺; found 267.1487. (Z)-1-tert-butyl-2-ethyl-3-(4-(trifluoromethyl)benzoyl)isourea (2c): colorless oil; ¹H

(**z**)-1-*tetr*-bill(*y*-2-eth*y*)-5-(4-(**trifluoromethyl**)**benzoyl**)**isourea** (**2c**): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.19 (s, 1H), 8.30 (d, *J* = 8.1 Hz, 2H), 7.65 (d, *J* = 8.2 Hz, 2H), 4.56 (q, *J* = 7.1 Hz, 2H), 1.45–1.37 (m, 12H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 175.8, 162.8, 141.3, 132.8 (q), 129.9, 128.2, 124.9 (q), 124.1(q), 63.7, 52.0, 29.4, 14.5 ppm; HRMS (*m*/z) (ESI): calcd for C₁₅H₂₀O₂N₂F₃ 317.1471 [M+H]⁺; found 317.1455 317.1455.

^{317.1455.} (**Z**)-1-tert-butyl-2-ethyl-3-(4-nitrobenzoyl)isourea (2d): yellow solid, m.p. 76.7-78.8 °C; ¹H NMR (400 MHz, CDCl₃): δ 10.19 (s, 1H), 8.36–8.32 (m, 2H), 8.24–8.21 (m, 2H), 4.58 (q, J = 7.1 Hz, 2H), 1.46–1.43 (m, 12H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 174.9, 162.9, 149.6, 143.7, 129.9, 123.1, 63.9, 52.1, 29.3, 14.5 ppm; HRMS (*m*/z) (ESI): calcd for C₁₄H₂₀O₄N₃ 294.1454 [M+H]⁺; found 294.1432 294.1432

(291): outcut for Clarify 2041(3) 2041(4) 2041(4) (114) (2)-1-tert-butyl-2-ethyl-3-(4-methoxybenzoyl)isourea (2e): white solid, m.p. 69.8-72.2 °C; ¹H NMR (400 MHz, CDCl₃): δ 10.18 (s, 1H), 8.28–8.03 (m, 2H), 6.89 (dd, J =6.7, 4.8 Hz, 2H), 4.53 (q, J = 7.1 Hz, 2H), 3.83 (s, 3H), 1.44–1.35 (m, 12H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 176.9, 162.4, 162.3, 131.0, 130.8, 113.1, 63.2, 55.3, 51.6, 29.5, 14.5 ppm; HRMS (m/z) (ESI): calcd for C₁₅H₂₃O₃N₂ 279.1709 [M+H]⁺; found 279.1688. (Z)-1-tert-butyl-2-ethyl-3-(2-fluorobenzoyl)isourea (2f): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.14 (s, 1H), 8.05 (td, J = 7.8, 1.8 Hz, 1H), 7.42–7.36 (m, 1H), 7.18–7.02 (m, 2H), 4.50 (q, J = 7.1 Hz, 2H), 1.43–1.36 (m, 12H); ¹³C NMR (100 MHz, CDCl₃): δ 175.7, 162.5, 162.0 (d, ¹ $J_{CF} =$ 256 Hz), 132.4 (d, ³ $J_{CF} =$ 9.0 Hz), 131.9, 126.7, 123.4 (d, ⁴ $J_{CF} =$ 4.0 Hz), 116.7 (d, ² $J_{CF} =$ 23.0 Hz), 63.6 51.9, 29.4, 14.5 ppm; HRMS (m/z) (ESI): calcd for C₁₄H₂₀O₂N₂F 267.1509 [M+H]⁺; found 267.1490. (Z)-1-tert-butyl-2-ethyl-3-(2-

(Z)-1-tert-butyl-2-ethyl-3-(2-(Z)-1-tert-buty1-2-etny1-3-(2-(trifluoromethyl)benzoyl)isourea (2g): colorless oil ; ¹H NMR (400 MHz, DMSO): δ 9.92 (s, 1H), 7.90–7.33 (m, 4H), 4.59–4.12 (m, 2H), 1.37 (t, J = 7.1 Hz, 9H), 1.30 (t, J = 7.1 Hz, 3H) ppm; ¹³C NMR (100 MHz, DMSO): δ 178.4, 162.4, 139.8, 131.8, 129.9, 129.9, 129.8, 126.3, 63.8, 52.0, 29.4, 14.5 ppm; HRMS (m/z) (ESI): calcd for C₁₅H₂₀O₂N₂F₃ 317.1477 [M+H]⁺; found 317.1453. (7)-1-tert-buty1-3-(2-ethoyybenzoyl)-2-ethylisourea

C₁₅H₂₀O₂N₂P₃ 317.1477 [M+H]⁺; found 317.1435. (Z)-1-tert-butyl-3-(2-ethoxybenzoyl)-2-ethylisourea (**2h**): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.14 (s, 1H), 7.90 (dd, J = 7.9, 1.8 Hz, 1H), 7.38–7.29 (m, 1H), 6.92 (dd, J = 11.4, 4.3 Hz, 2H), 4.47 (q, J = 7.1 Hz, 2H), 4.14 (q, J = 7.0 Hz, 2H), 1.44 (t, J = 7.0 Hz, 3H), 1.41– 1.34 (m, 12H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 178.4,

162.3, 158.0, 131.5, 131.4, 128.8, 119.8, 113.4, 64.4, 63.2, 51.7, 29.5, 14.9, 14.5 ppm; HRMS (m/z) (ESI): calcd for C₁₆H₂₅O₃N₂ 293.1865 [M+H]⁺; found 293.1841. (**Z)-1-tert-butyl-3-(2,6-difluorobenzoyl)-2-ethylisourea (2i)**: colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.00 (s, 1H), 7.29–7.21 (m, 1H), 6.97–6.80 (m, 2H), 4.42 (q, J = 7.1 Hz, 2H), 1.42 (s, 9H), 1.34 (t, J = 7.1 Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 173.0, 162.5, 161.5 (d, ¹ $J_{CF} = 251$ Hz), 160.1 (d, ¹ $J_{CF} = 250$ Hz), 130.1, 130.0 (d, ⁴ $J_{CF} = 3.0$ Hz), 111.7 (d, ² $J_{CF} = 19.0$ Hz), 111.6 (d, ² $J_{CF} = 19.0$ Hz), 64.1, 52.2, 29.3, 14.4 ppm; HRMS (m/z) (ESI): calcd for C₁₄H₁₉O₂N₂F₂ 285.1415 [M+H]⁺; found 285.1394. 285.1394.

(Z)-1-tert-butyl-3-(3,5-dimethoxybenzoyl)-2-

(Z)-1-tert-butyl-3-(3,5-dimethoxybenzoyl)-2-ethylisourea (2j): light yellow solid, m.p. 92.5–95.7 °C; ¹H NMR (400 MHz, CDCl₃): δ 10.21 (s, 1H), 7.40 (d, J = 2.4 Hz, 2H), 6.59 (t, J = 2.4 Hz, 1H), 4.55 (q, J = 7.1 Hz, 2H), 3.83 (s, 6H), 1.46–1.38 (m, 12H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 176.8, 162.6, 160.4, 140.2, 106.7, 104.2, 63.5, 55.5, 51.8, 29.4, 14.5 ppm; HRMS (*m*/z) (ESI): calcd for C₁₆H₂₅O₄N₂ 309.1814 [M+H]⁺; found 309.1791 309.1791

255.1151.

(25).1 calc for $C_{12}H_{19}O_{21}A_{20}$ 255.1167 [1111], four (25).151. (Z)-1-tert-butyl-3-cinnamoyl-2-ethylisourea (2m): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.17 (s, 1H) 7.68 (d, J = 15.9 Hz, 1H), 7.54 (d, J = 6.7 Hz, 2H), 7.40– 7.28 (m, 3H), 6.63 (d, J = 15.9 Hz, 1H), 4.47 (q, J = 7.1Hz, 2H), 1.42–1.35 (m, 12H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 178.0, 162.4, 140.8, 135.7, 129.3, 128.7, 128.1, 127.9, 63.2, 51.7, 29.5, 14.5 ppm; HRMS (m/z) (ESI): calcd for C₁₆H₂₃O₂N₂ 275.1760 [M+H]⁺; found 275.1737. (Z)-1-tert-butyl-2-ethyl-3-(2-phenylacetyl)isourea (2n): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 9.87 (s, 1H), 7.35–7.18 (m, 5H), 4.35 (q, J = 7.1 Hz, 2H), 3.64 (s, 2H), 1.34–1.27 (m, 12H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 184.6, 162.1, 137.1, 129.6, 128.2, 126.2, 63.3, 51.6, 48.1, 29.4, 14.4 ppm; HRMS (m/z) (ESI): calcd for C₁₅H₂₃N₂O₂ 263.1760 [M+H]⁺; found 263.1742. (Z)-1-benzoyl-3-tert-butyl-2-butylisourea (2p): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.24 (s, 1H),

(Z)-1-benzoyl-3-tert-butyl-2-butylisourea (2p): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.24 (s, 1H), 8.22 (d, J = 7.4 Hz, 2H), 7.48–7.38 (m, 3H), 4.51 (t, J = 6.5 Hz, 2H), 1.83–1.73 (m, 2H), 1.52 (d, J = 7.5 Hz, 2H), 1.41 (s, 9H), 1.00 (t, J = 7.4 Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 177.3, 162.8, 138.0, 131.4, 129.1, 127.9, 67.3, 51.7, 31.0, 29.5, 19.4, 13.8 ppm; HRMS (*m/z*) (ESI): calcd for C₁₆H₂₅O₂N₂ 277.1916 [M+H]⁺; found 277.1897. (Z)-1-benzoyl-3-tert-butyl-2-isopentylisourea (2q). colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.23 (s 1H)

(Z)-1-benzoyl-3-*tert*-butyl-2-isopentylisourea (2q). colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.23 (s, 1H), 8.25–8.15 (m, 2H), 7.48–7.37 (m, 3H), 4.54 (t, *J* = 6.8 Hz, 2H), 1.84–1.76 (m, 1H), 1.71–1.64 (m, 2H), 1.41 (s, 9H), 0.99 (d, *J* = 6.6 Hz, 6H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 177.3, 162.7, 138.1, 131.4, 129.1, 127.9, 66.0, 51.7, 37.7, 29.6, 29.5, 25.1, 22.5 ppm; HRMS (*m*/*z*) (ESI): calcd for C₁₇H₂₇O₂N₂291.2073 [M+H]⁺; found 291.2049. (Z)-1-benzoyl-3-*tert*-butyl-2-cyclohexylisourea (2r): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.25 (s, 1H), 8.21–8.14 (m, 2H), 7.49–7.43 (m, 1H), 7.43–7.36 (m, 2H), 5.39–5.29 (m, 1H), 2.05–1.97 (m, 2H), 1.81–1.75 (m, 2H), 1.69–1.64 (m, 2H), 1.61–1.55 (m, 2H), 1.56–1.47(m, 2H), 1.42 (s, 9H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 177.2, 162.2, 138.2, 131.3, 129.0, 127.9, 75.7, 51.7, 31.7, 29.5,

25.6, 23.6 ppm; HRMS (m/z) (ESI): calcd for C₁₈H₂₇O₂N₂ 303.2067 [M+H]⁺; found 303.2051. (**Z**)-1-benzoyl-2-benzyl-3-*tert*-butylisourea (2s):

303.2067 [M+H]⁺; found 303.2051. (**Z**)-1-benzoyl-2-benzyl-3-*tert*-butylisourea (2s): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.31 (s, 1H), 8.30–8.21 (m, 2H), 7.52–7.33 (m, 8H), 5.57 (s, 2H), 1.41 (s, 9H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 177.4, 162.5, 137.9, 136.2, 131.6, 129.1, 128.6, 128.3, 128.2, 128.0, 52.0, 30.0 ppm; HRMS (*m*/z) (ESI): calcd for C₁₉H₂₃O₂N₂ 311.1760 [M+H]⁺; found 311.1740. (**Z**)-1-benzoyl-3-cyclohexyl-2-ethylisourea (2u): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.04 (s, 1H), 8.27–8.16 (m, 2H), 7.48–7.44 (m, 1H), 7.43–7.37 (m, 2H), 4.53 (q, *J* = 7.1 Hz, 2H), 3.75–3.61 (m, 1H), 2.00–1.94 (m, 2H), 1.77–1.71 (m, 2H), 1.69–1.51 (m, 2H), 1.41–1.32 (m, 7H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 177.4, 162.2, 138.0, 131.5, 129.1, 127.9, 63.5, 49.9, 32.9, 25.5, 24.5, 14.7 ppm; HRMS (*m*/z) (ESI): calcd for C₁₆H₂₃O₂N₂ 275.1760 [M+H]⁺; found 275.1740. (**Z**)-1-benzoyl-2-ethyl-3-(2,4,4-trimethylpentan-2-yl)isourea (2v): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 10.29 (s, 1H), 8.36–8.04 (m, 2H), 7.46 (dd, *J* = 6.2, 3.6, 1.3 Hz, 1H), 7.40 (t, *J* = 7.3 Hz, 2H), 4.56 (q, *J* = 7.1 Hz, 2H), 1.75 (s, 2H), 1.47–1.40 (m, 9H), 1.00 (s, 9H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 177.2, 162.4, 138.0, 131.4, 129.1, 127.9, 63.3, 55.4, 52.1, 31.7, 31.5, 30.2, 14.7 ppm; HRMS (*m*/z) (ESI): calcd for C₁₈H₂₉O₂N₂ 305.2229 [M+H]⁺; found 305.2208. (**Z**)-1-benzoyl-3-benzyl-2-ethylisourea (2w): white solid, mp, 5.86–62.3 °C: ¹H NMR (400 MHz, CDCl₃): δ

[M+H]⁺; found 305.2208. (Z)-1-benzoyl-3-benzyl-2-ethylisourea (2w): white solid, m.p. 58.6–62.3 °C; ¹H NMR (400 MHz, CDCl₃): δ 10.36 (s, 1H), 8.24–8.21 (m, 2H), 7.50–7.44 (m, 1H), 7.42–7.26 (m, 7H), 4.59–4.47 (m, 4H), 1.38 (t, J = 7.1 Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 177.7, 162.9, 137.8, 137.5, 131.6, 129.2, 128.7, 127.9, 127.6, 127.5, 63.8, 45.0, 14.6 ppm; HRMS (m/z) (ESI): calcd for C₁₇H₁₉O₂N₂283.1447 [M+H]⁺; found 283.1426. (Z)-1-benzoyl-3-(2,6-dimethylphenyl)-2-ethylisourea (2x): colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 11.43 (s, 1H), 8.39–8.27 (m, 2H), 7.57–7.42 (m, 3H), 7.18–7.08 (m, 3H), 4.55 (q, J = 7.1 Hz, 2H), 2.27 (s, 6H), 1.33 (t, J = 7.1Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 178.2, 162.2, 137.6, 135.2, 133.8, 131.9, 129.4, 128.1, 127.4, 63.9, 18.5, 14.7 ppm; HRMS (m/z) (ESI): calcd for C₁₈H₂₁O₂N₂ 297.1603 [M+H]⁺; found 297.1591. General experimental procedure for synthesis of imides 3. A mixture of amides 1 (1.0 equiv, 0.5 mmol),

imides 3. A mixture of amides 1 (1.0 equiv, 0.5 mmol), *tert*-butyl isocyanide (1.0 equiv. 0.5 mmol), carboxylates (1.0 equiv, 0.5 mmol), Pd(dppf)Cl₂ (10 mol %, 0.05 mmol) and 2 mL of CH₃CN was stirred at 60 °C for 2 h. The progress of the reaction was monitored by thin-layer chromatography. The mixture was monitored by thin layer evaporated under reduced pressure, and the residue was purified by flash column chromatography (petroleum ether/ethyl acetate) to afford imides **3**.

euler/enlyl acetale) to allord infides 5. **N-acetylbenzamide (3a)**: white solid, m.p. 107.9–109.8 °C (lit.^[18] 109–110°C); ¹H NMR (400 MHz, CDCl₃): δ 9.20 (s, 1H), 7.94–7.85 (m, 2H), 7.59 (t, J = 7.4 Hz, 1H), 7.49 (t, J = 7.7 Hz, 2H), 2.60 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 174.0, 165.9, 133.2, 132.7, 128.9, 127.9, 25.7 ppm; HRMS (m/z) (ESI): calcd for C₉H₉O₂NNa 186.0531 [M+Na]⁺; found 186.0518. **N-acetyl-4-methylbenzamide (3b)**: white solid map

186.0531 [M+Na]⁺; found 186.0518. **N-acetyl-4-methylbenzamide** (**3b**): white solid, m.p. 101.9–103.2 °C (litt.^[119] 103–104°C); ¹H NMR (400 MHz, CDCl₃): δ 8.79 (s, 1H), 7.76 (d, J = 8.3 Hz, 2H), 7.30 (d, J = 8.0 Hz, 2H), 2.61 (s, 3H), 2.43 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 173.6, 165.6, 144.2, 129.8, 129.7, 127.7, 25.6, 21.7 ppm; HRMS (m/z) (ESI): calcd for C₁₀H₁₁O₂NNa 200.0687 [M+Na]⁺; found 200.0674. **N-acetyl-4-chlorobenzamide** (**3c**): white solid, m.p. 136.9–138.4 °C (litt.^[20] 140–142°C); ¹H NMR (400 MHz, CDCl₃): δ 9.36 (s, 1H), 7.86 (dd, J = 8.9, 2.0 Hz, 2H), 7.62–7.35 (m, 2H), 2.60 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 174.2, 165.0, 139.8, 131.0, 129.4, 129.3, 25.8 ppm; HRMS (m/z) (ESI): calcd for C₉H₈O₂ClNNa 220.0141 [M+Na]⁺; found 220.0133. **N-acetyl-4-fluorobenzamide (3d):** white needle crystal,

N-acetyl-4-fluorobenzamide (3d): white needle crystal, m.p. 108.2–109.9°C (lit.^[21]111–112°C); ¹H NMR (400 MHz, CDCl₃): δ 9.52 (s, 1H), 8.13–7.84 (m, 2H), 7.23– 7.03 (m, 2H), 2.61 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 174.4, 165.8 (d, ¹J_{CF} = 254 Hz), 164.9, 130.5 (d, ³J_{CF} = 9.0 Hz), 128.8 (d, ⁴J_{CF} = 3.0 Hz), 116.1 (d, ²J_{CF} = 22.0 Hz),

25.7 ppm; HRMS (m/z) (ESI): calcd for C₉H₈O₂NFNa 204.0437 [M+Na]⁺; found 204.0423.

204.0437 [M+Na]'; found 204.0423. **N-acetyl-4-(trifluoromethyl)benzamide** (**3e**): white solid, m.p. 105.7–107.1°C (lit.^[22]111–112°C); ¹H NMR (400 MHz, CDCl₃): δ 9.89 (d, J = 5.2 Hz, 1H), 8.09 (d, J = 8.2 Hz, 2H), 7.77 (d, J = 8.3 Hz, 2H), 2.63 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 174.4, 165.2, 136.0, 134.6 (q), 128.6, 125.8 (q), 123.5(q), 29.6 ppm; HRMS (*m*/*z*) (ESI): calcd for C₁₀H₈O₂NF₃Na 254.0405 [M+Na]⁺; found 254.0388 254.0388.

N-acetyl-4-aminobenzamide (3f): yellow solid, m.p. 97.5–101.4°C; ¹H NMR (400 MHz, CDCl₃): δ 8.53 (s, 1H), 7.74–7.62 (m, 2H), 6.79–6.58 (m, 2H), 4.17 (s, 2H), 2.59 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 173.5, 164.9, 151.3, 129.9, 121.6, 114.2, 25.4 ppm; HRMS (*m*/*z*) (ESI): calcd for C₉H₁₀O₂N₂Na 201.0640 [M+Na]⁺; found 201.0626.

cured 10 C911[002172174 20110040 [[N1174]], found 201.0626. **N-acetyl-4-methoxybenzamide (3g)**: light yellow solid, m.p. 108.3–111.5°C (lit.^[23]119–119.5°C); ¹H NMR (400 MHz, CDCl₃): δ 9.10 (s, 1H), 7.94–7.81 (m, 2H), 6.98 (d, J = 8.9 Hz, 2H), 3.88 (s, 3H), 2.60 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 174.0, 165.2, 163.6, 129.9, 124.8, 114.2, 55.6, 25.6 ppm; HRMS (m/z) (ESI): calcd for C₁₀H₁₁O₃NNa 216.0637 [M+Na]⁺; found 216.0623. **N-acetyl-2-fluorobenzamide (3h)**: light yellow solid, m.p. 40.5–43.8 °C; ¹H NMR (400 MHz, CDCl₃): δ 9.01 (s, 1H), 8.00 (td, J = 7.8, 1.7 Hz, 1H), 7.16 (dd, J = 12.0, 8.4 Hz, 1H), 2.56 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 172.6, 162.1, 160.5 (d, ¹J_{CF} = 248 Hz), 135.1 (d, ³J_{CF} = 10.0 Hz), 132.1, 125.2 (d, ⁴J_{CF} = 3.0 Hz), 120.2 (d, ³J_{CF} = 11.0 Hz), 116.5 (d, ²J_{CF} = 24.0 Hz), 25.9 ppm; HRMS (m/z) (ESI): calcd for C₉H₈O₂NFNa 204.0437 [M+Na]⁺; found 204.0423. **N-acetyl-2-iodobenzamide (3i)**: yellow solid, m.p.

found 204.0423. **N-acetyl-2-iodobenzamide** (**3i**): yellow solid, m.p. 69.8–73.2 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.58 (s, 1H), 7.92 (d, J = 8.0 Hz, 1H), 7.45 (t, J = 4.9 Hz, 2H), 7.23– 7.14 (m, 1H), 2.56 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 172.5, 167.7, 140.4, 140.1, 132.3, 128.4, 128.3, 92.0, 25.4 ppm; HRMS (*m*/*z*) (ESI): calcd for C₉H₈O₂NINa 311.9497 [M+Na]⁺; found 311.9471. **N-acetyl-2-(trifluoromethyl)benzamide** (**3j**): white solid m p. 89 8–92 9°C: ¹H NMR (400 MHz, CDCl₃): ⁵

N-acetyi-2-(**ITILIOOTOMETRYI)DERZAMIGE** (3J): White solid, m.p. 89.8–92.9°C; ¹H NMR (400 MHz, CDCl₃): ⁵ 8.83 (s, 1H), 7.73 (d, J = 7.9 Hz, 1H), 7.65–7.58 (m, 2H), 7.51 (d, J = 6.8 Hz, 1H), 2.49 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 172.2, 166.9, 134.1, 132.1, 130.9, 127.7, 127.4(q), 126.8 (q), 123.3 (q), 25.2 ppm; HRMS (*m*/*z*) (ESI): calcd for C₁₀H₈O₂NF₃Na 254.0405 [M+Na]⁺; found 254.0389 254.0389

254.0389. **N-acetyl-2-ethoxybenzamide** (3k): white solid, m.p. 84.2–86.9°C; ¹H NMR (400 MHz, CDCl₃): δ 10.45 (s, 1H), 8.25–8.02 (m, 1H), 7.53–7.46 (m, 1H), 7.12–7.03 (m, 1H), 6.98 (d, J = 8.1 Hz, 1H), 4.22 (t, J = 7.0 Hz, 2H), 2.55 (s, 3H), 1.55 (t, J = 7.0 Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 173.2, 164.0, 157.2, 134.7, 132.8, 121.6, 120.2, 112.7, 65.3, 29.6, 14.6 ppm; HRMS (m/z) (ESI): calcd for C₁₁H₁₃O₃NFNa 230.0793 [M+Na]⁺; found 230.0779. **N-acetyl-2 6-difluorobenzamide** (31): white needle

112.7, 65.3, 29.6, 14.6 ppm; HRMS (*m*/*z*) (ESI): calcd for C₁₁H₁₃O₃NFNa 230.0793 [M+Na]⁺; found 230.0779. *N*-acetyl-2,6-difluorobenzamide (31): white needle crystal, m.p. 138.5–140.3°C (lit.^[24] 141.5°C); ¹H NMR (400 MHz, CDCl₃): δ 8.82 (s, 1H), 7.48–7.28 (m, 1H), 6.92 (td, *J* = 8.4, 2.0 Hz, 2H), 2.54–2.33 (m, 3H) ppm;¹³C NMR (100 MHz, CDCl₃): δ 172.0, 160.5 (d, ¹*J*_{CF} = 253.0 Hz), 159.9 (d, ¹*J*_{CF} = 253 Hz), 159.8, 133.2 (d, ³*J*_{CF} = 10.0 Hz), 133.1 (d, ³*J*_{CF} = 11.0 Hz), 112.2 (d, ²*J*_{CF} = 22.0 Hz). 25.5 ppm; HRMS (*m*/*z*) (ESI): calcd for C₉H₇O₂NF₂Na 222.0343 [M+Na]⁺; found 222.0329. *N*-acetyl-3,5-dimethoxybenzamide (3m): white solid, m.p. 157.2–159.5°C (lit.^[25] 161–163°C); ¹H NMR (400 MHz, CDCl₃): δ 8.93 (s, 1H), 6.99 (d, *J* = 2.2 Hz, 2H), 6.66 (t, *J* = 2.2 Hz, 1H), 3.84 (s, 6H), 2.60 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 173.6, 165.6, 161.2, 134.7, 105.6, 105.5, 55.7, 25.6 ppm; HRMS (*m*/*z*) (ESI): calcd for C₁₁H₁₃O₄NNa 246.0742 [M+Na]⁺; found 246.0727. *N*-acetyl-1-naphthamide (3n): light yellow solid, m.p. 109.2–112.1°C; ¹H NMR (400 MHz, CDCl₃): δ 8.94 (s, 1H), 7.93 (d, *J* = 8.1 Hz, 1H), 7.84 (d, *J* = 7.7 Hz, 1H), 7.66 (d, *J* = 7.0 Hz, 1H), 7.58–7.48 (m, 2H), 7.41 (dd, *J* = 10.9, 4.3 Hz, 1H), 2.57 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 173.2, 167.0, 133.8, 132.5, 131.8, 130.2, 128.6, 127.8, 126.8, 126.2, 124.9,

124.5, 25.6 ppm; HRMS (m/z) (ESI): calcd for $C_{13}H_{11}O_2NNa$ 236.0687[M+Na]⁺; found 236.0673.

N-acetylthiophene-2-carboxamide (30): light yellow solid, m.p. 108.3–110.5°C (lit.^[21] 134–135°C); ¹H NMR (400 MHz, CDCl₃): δ 8.79 (s, 1H), 7.71 (d, J = 3.8 Hz, 1H), 7.67 (d, J = 5.0 Hz, 1H), 7.16 (t, J = 4.3 Hz, 1H), 2.60 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 173.2, 159.9, 127.5 123.6 120.5 128.2 5 ppm; UDNG (w/z) (CDN) 137.5, 133.6, 130.5, 128.3, 25.5 ppm; HRMS (m/z) (ESI): calcd for C₇H₇O₂NNaS 192.0095 [M+Na]⁺; found 192.0080

N-acetylcinnamamide (**3**p): light yellow needle crystal, m.p. 109.2–112.8 °C (lit.^[23] 131–132°C); ¹H NMR (400 m.p. 109.2–112.8 °C (lit.^[23] 131–132°C); ¹H NMR (400 MHz, CDCl₃): δ 9.50 (s, 1H), 7.83 (d, J = 15.7 Hz, 1H), 7.58 (dt, J = 7.4, 3.6 Hz, 2H), 7.41 (dd, J = 6.3, 3.9 Hz, 3H), 6.97 (d, J = 15.6 Hz, 1H), 2.49 (s, 3H) ppm; ¹³C NMR (101 MHz, CDCl₃): δ 173.4, 165.8, 145.9, 134.2, 130.8, 129.0, 128.5, 119.5, 25.4 ppm; HRMS (m/z) (ESI): calcd for C₁₁H₁₁O₂NNa 212.0687 [M+Na]⁺; found 212.0674. **N-Benzoylbenzamide (3q)**: white solid, m.p. 143.6–145.2°C (lit.^[18] 144–145°C); ¹H NMR (400 MHz, CDCl₃): δ 9.06 (s, 1H), 7.87 (d, J = 7.9 Hz, 4H), 7.60 (t, J = 7.4 Hz, 2H), 7.50 (t, J = 7.5 Hz, 4H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 166.5, 133.4, 133.1, 128.9, 128.0 ppm; HRMS (m/z) (ESI): calcd for C₁₄H₁₁O₂NNa 248.0687 [M+Na]⁺; found 248.0672.

found 248.0672

found 248.0672. *N***-methacryloylbenzamide** (**3r**): light yellow solid, m.p. 68.6–72.4°C (lit.^[26] 71–72°C); ¹H NMR (400 MHz, CDCl₃): δ 8.92 (s, 1H), 7.79 (d, J = 8.0 Hz, 2H), 7.56 (dd, J = 8.9, 4.5 Hz, 1H), 7.45 (dd, J = 14.3, 7.0 Hz, 2H), 5.82 (d, J = 0.5 Hz, 1H), 5.59 (s, 1H), 2.03 (s, 3H) ppm; ¹³C NMR (100MHz, CDCl₃): δ 166.4, 165.4, 139.3, 132.3, 132.0, 127.8, 126.9, 121.0, 17.4 ppm; HRMS (m/z) (ESI): calcd for C₁₁H₁₁O₂NNa 212.0687 [M+Na]⁺; found 212.0672 212.0672

N-(2-ethylhexanoyl)benzamide (3s): White solid, m.p. **N-(2-ethylhexanoyl)benzamide** (3s): White solid, m.p. 94.1–98.3°C; ¹H NMR (400 MHz, CDCl₃): δ 8.94 (s, 1H), 7.95–7.84 (m, 2H), 7.59 (t, J = 7.4 Hz, 1H), 7.48 (t, J = 7.7 Hz, 2H), 3.58–3.42 (m, 1H), 1.83–1.70 (m, 2H), 1.65–1.49 (m, 2H), 1.36–1.26 (m, 4H), 0.96 (t, J = 7.4 Hz, 3H), 0.87 (t, J = 7.0 Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 179.3, 165.6, 133.2, 133.1, 128.9, 127.8, 47.1, 31.2, 29.5, 25.0, 22.9, 14.0, 11.7 ppm; HRMS (m/z) (ESI): calcd for C₁₅H₂₁O₂NNa 270.1470 [M+Na]⁺; found 270.1454.

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FULL PAPER

Palladium-Catalyzed Solvent-Controlled Selective Synthesis of Acyl Isoureas and Imides from Amides, Isocyanides, Alcohols and Carboxylates

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