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# Synthesis and Biological Evaluation of Quinazolonethiazoles as New Potential Conquerors towards *Pseudomonas aeruginosa*

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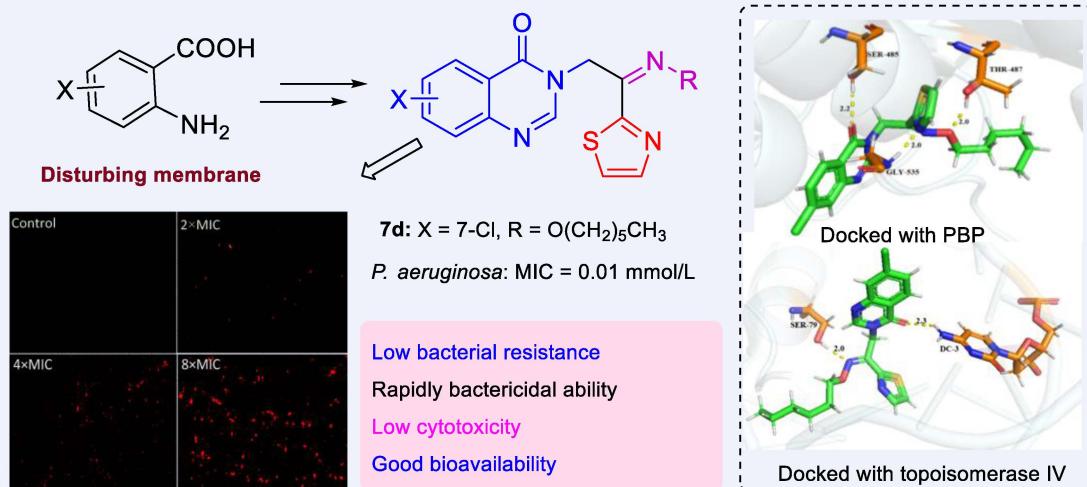
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**Keywords**

Thiazole | Schiff bases | Antimicrobial | Drug design | DNA

**Main observation and conclusion**

Novel quinazolonethiazoles were designed and synthesized as new potential antimicrobial agents by facile multi-step procedure from *o*-aminobenzoic acids and 2-acetylthiazole. A series of biological evaluation showed that compound **7d** was the most effective quinazolonethiazole with superior activity to reference drugs chloramphenicol and norfloxacin. This active molecule displayed unobvious bacterial resistance against *P. aeruginosa*, the low toxicity to normal hepatocytes, suitable pharmacokinetics and drug-likeness. The preliminary biological interaction suggested that quinazolonethiazole **7d** might induce bacterial death by disturbing the membrane permeability, whilst preventing bacteria from growth by integrating into DNA and binding with topoisomerase IV. These findings provided significant background for the further development of quinazolonethiazoles as new potential drugs in combating drug-resistant pathogens.

**Comprehensive Graphic Content**

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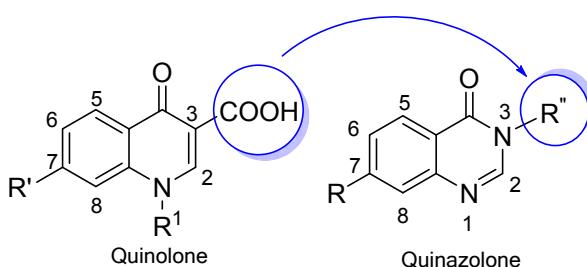
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Supporting Information

## Background and Originality Content

The alarmingly escalating microbial resistance continues to threaten the longevity and effectiveness of available antibiotics, causing the increased mortality of infected patients and aggravating the clinical challenge of tackling infections,<sup>[1]</sup> which could accelerate the process of approaching post-antibiotic era.<sup>[2]</sup> Accordingly, the research of novel antimicrobials is an immediate matter for discovery of new medicinal drugs to address causative diseases.

Quinazolone widely exists in many bioactive agents including naturally occurring alkaloids and synthetic compounds.<sup>[3-4]</sup> The unique structural feature of quinazolone backbone is identified as a new and important chemical structural framework, naturally its derivatives tremendously captivate attention on the synthesis, structural modification and biomedical effects.<sup>[5]</sup> Especially for the antibacterial properties, quinazolone does not contain  $\beta$ -lactam ring but exerts the similar mechanism of action to ceftaroline that could bind to the allosteric site of penicillin-binding protein (PBP).<sup>[6]</sup> More relevantly, quinazolone with benzopyrimidone skeleton is structurally different from the C-3 position at quinolone having benzopyridone core (Figure 1), and some studies revealed that the drug resistance of quinolones is primarily associated with the chelation of the carboxyl group at C-3 position.<sup>[7]</sup> These significant properties suggest that quinazolones could have potential to replace quinolones in exerting similar biological efficacy against drug-resistant strains.<sup>[8]</sup> In fact, some reported quinazolones showed higher activity than clinical drugs against dreadful pathogens.<sup>[9]</sup> Therefore, a promising approach to overcome resistance problem might be to develop innovative quinazolones by modifying the 3-position of quinazolones. This type of new derivatives maybe exert multiple modes of action.



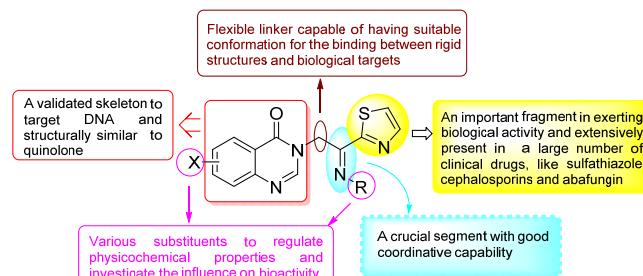
**Figure 1** The main structures of quinolone and quinazolone.

The five-member aromatic heterocyclic thiazole has been well-known for valuable contributions to the efficacy of clinically available antimicrobial drugs,<sup>[10]</sup> such as antibacterial ceftaroline fosamil acetate and aztreonam, antifungal ravuconazole and abafungin.<sup>[11]</sup> This could be attributed to the unique structure of thiazole ring with both electron-donating groups (NH or S) and electron-withdrawing group (C=N), which endows it to have the multi-binding ability to functional targets through diverse weak interactions.<sup>[12-13]</sup> The successful exploitation of numerous thiazole-based medications has been stimulating immense efforts to develop the conjugates of thiazole with various pharmacophores. This conjugation strategy has occupied a prominent place in discovery of new bioactive species.<sup>[14-15]</sup> Recently, the incorporation of thiazole ring into the antibacterial quinolone skeleton not only facilitates conjugates to possess the excellent bioactivities but also affects bacterial cellular normal processes including membrane permeability and DNA replication.<sup>[16]</sup> Thus, the introduction of thiazole in the C-3 position at quinazolone might construct more effective antimicrobial candidates.

Imine is a helpful functional group in exerting bioactivity because it easily performs hydrogen bonds, coordination and/or

polar interactions to regulate the lipid-water partition and to improve pharmacokinetics and biocompatibility, thereby presenting in the majority of drugs.<sup>[17-18]</sup> Particularly, many antibacterial drugs containing thiazole ring usually have the presence of imine bond like cefixime and cefdinir,<sup>[19-20]</sup> and extensive work has been devoted to the introduction of imine in the development of antimicrobials about quinazolone,<sup>[21]</sup> most of which display excellent antimicrobial activity.<sup>[22]</sup> This might mean that imine fragment is extremely important for the biological activity of compounds like thiazoles and quinazolones. As a result, these stimulating properties promote the synthesis of thiazole conjugated quinazolinone with different imine moieties including oximes, hydrazines and methylenimines for antimicrobials development.

In present work, the thiazole conjugated quinazolones as novel potential antimicrobial hybrids were designed and synthesized against bacterial and fungal strains with the hope of high safety, low microbial resistance and good bioavailability (Figure 2). For purpose of verifying the research value of this framework, the experiments involving bacterial resistance study, bactericidal kinetic study, cytotoxicity and ADME study were performed. In addition, antibacterial mechanisms including biofilm destruction, molecular docking study and the interaction with DNA were preliminarily explored to correlate them with their antimicrobial activities. This series of studies might provide valuable information for the design of new antimicrobial analogues with improved activity.

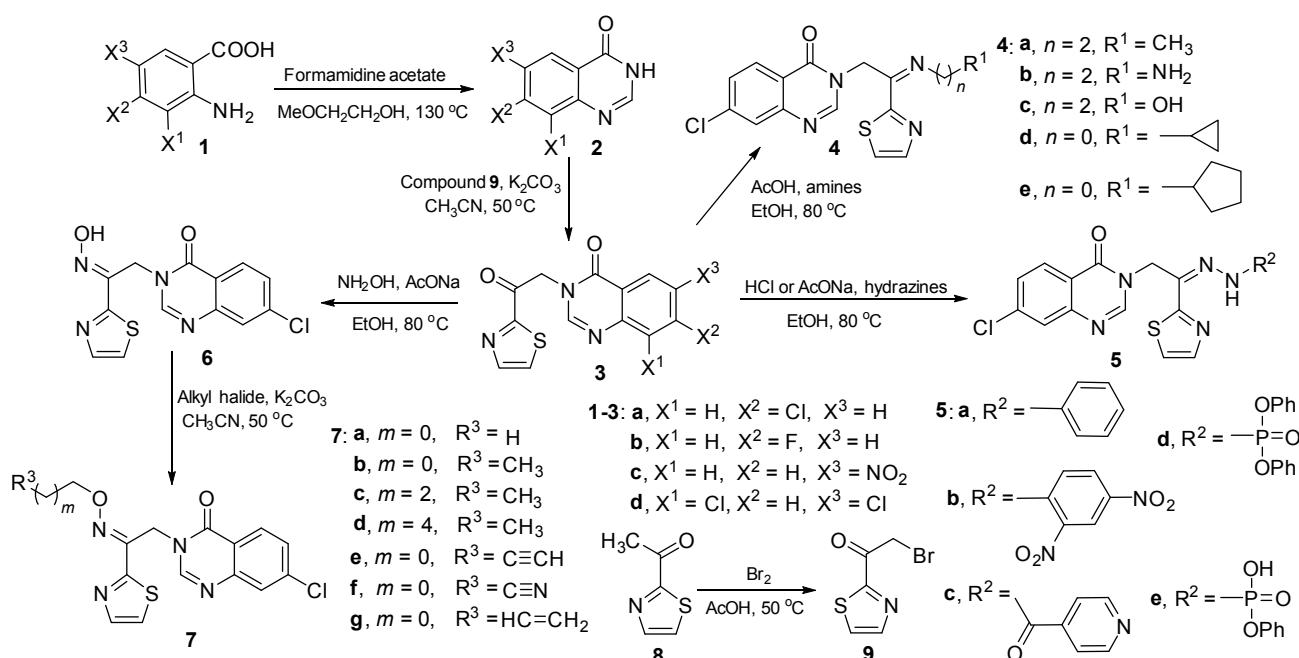


**Figure 2** Design of novel structural potential antimicrobial target quinazolonethiazoles.

## Results and Discussion

### Chemistry

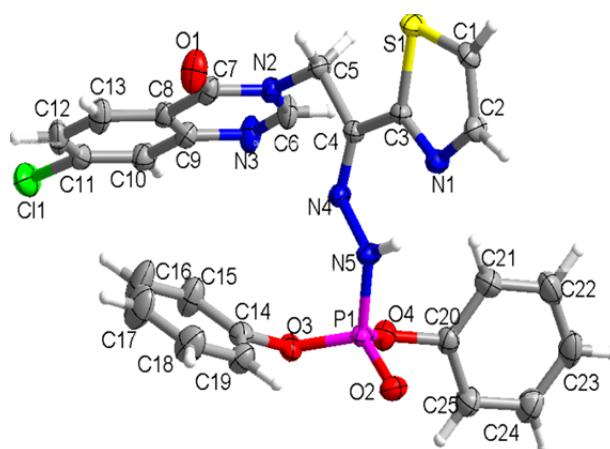
The preparation of target quinazolonethiazoles was carried out according to Scheme 1. The commercially available formamidine acetate and anthranilic acid with different substituents were used as starting materials to construct quinazolone skeleton **2**. Bromination of 2-acetylthiazole **8** gave 2-bromoacetylthiazole **9**, which was coupled with quinazolone skeleton **2** to afford the important precursors **3a–3d** with yields of 25.2%–49.9% in presence of potassium carbonate. Experiments revealed that quinazolones with electron-donating groups could not successfully react with 2-bromoacetylthiazole **9**, which may be attributed to the influences of electron density on reactivity of quinazolones, and the yield increased as the electron density in quinazolones decreased. The condensation reaction of compound **3a** with different primary amines resulted in the formation of methylenimine derivations **4a–4e** in yields of 32.8%–58.0% using acetic acid as catalyst and ethanol as solvent. Under similar conditions, hydrazone compounds **5a–5e** in good yields of 56.3%–82.7% were easily produced by reacting **3a** with different hydrazines. Moreover, the precursor **3a** was treated with hydroxylamine hydrochloride to furnish the oxime **6**, which was further modified by introducing alkyl halides at the O-atom of the oxime group in acetonitrile to produce the oxime derivations **7a–7g** in

**Scheme 1** Preparative process of target quinazolonethiazoles

yields ranging from 45.8% to 79.7%.

#### Analysis of configuration (*E* or *Z*)

The carbonyl group of quinazolonethiazole **3a** was condensed with the primary amine to form target imines **4a–4e** and **5a–5e** as well as oxime **6**. Theoretically, there should be two configurations for tautomeric equilibrium. Thus, the single crystal of hydrazone derivative **5d** was further cultured in order to understand its structure. X-ray diffraction analysis pointed out that the obtained single crystal of hydrazone derivative **5d** (Figure 3) showed the *Z* configuration, which could be ascribed to that the *E* configuration is general instability or easy conversion to the *Z* configuration.



**Figure 3** Crystal structure of quinazolonethiazole **5d**.

#### Antibacterial activity

To elucidate the structures of novel quinazolonethiazoles on effects of antimicrobial activity, the antibacterial evaluations were operated and the results were shown in Table 1. The first prepared quinazolonethiazoles **3a–3d** with different substituents on the benzene ring showed moderate antibacterial activity. In particular, compound **3a** containing chlorine substitution not only exhibited stronger anti-*E. faecalis* effect (MIC = 0.01 mmol/L) than

reference drugs chloramphenicol and norfloxacin, but also gave excellent antibacterial activity against Gram-positive stain *S. aureus* ATCC 29213 with MIC value of 0.003 mmol/L, which was 3-fold and 8-fold more potent than the clinical antibacterial chloramphenicol (MIC = 0.01 mmol/L) and norfloxacin (MIC = 0.025 mmol/L), respectively. Therefore, the structural modification of compound **3a** was carried out to construct novel quinazolone-thiazoles with methylenimines **4a–4e** showed moderate or weak biological activity with MIC values in the range of 0.05–0.74 mmol/L in comparison with reference drugs. However, they showed superior inhibition efficacy to precursor **3a** for most of the tested bacterial strains, especially aminoethyl and hydroxyethyl modified Schiff bases **4b** and **4c**, which were far stronger than compound **3a** (MIC = 0.84 mmol/L) towards drug-resistant *S. aureus* having MIC values of 0.05 mmol/L. The above information pointed out that the introduction of Schiff base on compound **3a** occupied a prominent place on antibacterial activity.

To better analyze the structure-activity relationship (SAR) and enrich the structural diversity, imine compounds **4b–4c** with the best activity were further modified to yield hydrazones **5a–5e** and oximes **7a–7g**. As for hydrazone-derived quinazolonethiazoles **5a–5e**, the antibacterial activities did not exhibit significant improvement in comparison to aminoethyl derivative **4b**, except for compound **5d**, which could inhibit the growth of MRSA and *E. faecalis* with a little lower MIC value of 0.06 mmol/L. For another, hexyl oxime derivative **7d** not only displayed good inhibitory activities against tested strains, but also was more effective than chloramphenicol and norfloxacin against the growth of *P. aeruginosa* and *P. aeruginosa* ATCC 27853, while the other oxime compounds including unsaturated alkyl chain apparently lost bioactivity compared to compound **7d**. However, in general, the introduction of oxime fragment was beneficial to antibacterial activity.

#### Antifungal activity

The *in vitro* antifungal results in Table S1 showed that the inhibitory activities of the target compounds were generally inferior to that of fluconazole against the tested fungi apart from *A. fumigatus*. The antifungal activities of precursor **3a** showed twice

**Table 1** *In vitro* antibacterial data as MIC (mmol/L) for quinazolonethiazoles **3–7<sup>a,b</sup>**

Compds	Gram-positive bacteria <sup>b</sup>					Gram-negative bacteria <sup>b</sup>					
	MRSa	E. f.	S. a.	S. a. 25923	S. a. 29213	K. p.	E. c.	P. a.	A. b.	P. a. 27853	E. c. 25922
<b>3a</b>	0.21	0.01	0.84	0.84	0.003	0.84	0.84	0.84	0.84	0.84	1.67
<b>3b</b>	0.22	0.11	0.88	0.44	0.88	0.44	0.88	0.88	0.88	0.88	0.88
<b>3c</b>	0.81	0.40	0.40	0.40	0.81	0.40	0.81	0.81	0.40	0.81	0.81
<b>3d</b>	0.75	0.75	0.38	0.75	0.75	0.75	0.19	0.75	0.75	0.38	0.75
<b>4a</b>	0.74	0.37	0.74	0.74	0.19	0.74	0.74	0.74	0.74	0.74	0.37
<b>4b</b>	0.73	0.18	0.05	0.18	0.73	0.37	0.37	0.18	0.73	0.37	0.18
<b>4c</b>	0.09	0.18	0.05	0.37	0.74	0.37	0.37	0.37	0.18	0.37	0.05
<b>4d</b>	0.74	0.74	0.74	0.74	0.74	0.74	0.74	0.74	0.74	0.74	0.74
<b>4e</b>	0.69	0.34	0.69	0.34	0.69	0.69	0.69	0.69	0.69	0.69	0.69
<b>5a</b>	0.65	0.08	0.65	0.16	0.32	0.65	0.65	0.65	0.65	0.65	0.65
<b>5b</b>	0.53	0.26	0.53	0.53	0.53	0.53	0.53	0.26	0.53	0.03	0.53
<b>5c</b>	0.60	0.60	0.60	0.60	0.60	0.60	0.60	0.60	0.60	0.60	0.60
<b>5d</b>	0.06	0.06	0.46	0.46	0.23	0.23	0.23	0.46	0.46	0.46	0.46
<b>5e</b>	0.07	0.27	0.54	0.54	0.54	0.54	0.54	0.54	0.54	0.13	0.27
<b>6</b>	0.42	1.66	0.83	0.83	0.83	0.80	1.60	1.60	1.60	0.40	1.60
<b>7a</b>	0.38	0.19	0.76	0.38	0.76	0.38	0.38	0.76	0.76	0.19	0.76
<b>7b</b>	1.42	0.35	1.42	0.18	0.71	0.73	0.73	1.47	0.37	1.47	1.47
<b>7c</b>	0.09	0.73	0.73	0.37	0.73	0.17	0.68	1.36	0.68	0.34	0.68
<b>7d</b>	1.26	0.04	1.26	0.02	0.01	0.01	0.04	0.01	1.26	0.01	0.63
<b>7e</b>	1.60	0.80	1.60	0.40	1.60	1.43	0.71	1.43	1.43	1.43	0.71
<b>7f</b>	0.71	0.71	1.43	0.18	0.71	1.42	1.42	0.71	0.71	0.71	1.42
<b>7g</b>	0.71	1.42	1.42	0.71	1.42	0.71	1.42	0.71	1.42	1.42	1.42
<b>A<sup>c</sup></b>	0.05	0.02	0.02	0.02	0.01	0.02	0.10	0.10	0.05	0.02	0.05
<b>B<sup>d</sup></b>	0.003	0.013	0.006	0.013	0.025	0.002	0.001	0.025	0.006	0.025	0.050

<sup>a</sup> Minimal inhibitory concentrations (MIC, mmol/L) were determined by micro broth dilution method for microdilution plates. <sup>b</sup> MRSa, *Methicillin-Resistant Staphylococcus aureus*; S. a., *Staphylococcus aureus*; S. a. 25923, *Staphylococcus aureus* ATCC 25923; S. a. 29213, *Staphylococcus aureus* ATCC 29213; E. f., *Enterococcus faecalis*; K. p., *Klebsiella pneumonia*; E. c., *Escherichia coli*; E. c. 25922, *Escherichia coli* ATCC 25922; P. a., *Pseudomonas aeruginosa*; P. a. 27853, *Pseudomonas aeruginosa* ATCC 27853; A. b., *Acinetobacter baumanii*. <sup>c</sup> A = Chloramphenicol. <sup>d</sup> B = Norfloxacin.

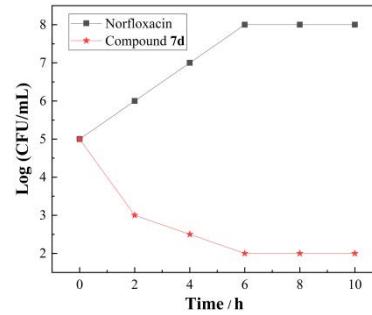
or equal to fluconazole against *A. fumigatus* and *C. albicans* ATCC 90023, respectively. For methylenimines **4a**–**4e** and hydrazone **5a**–**5e**, they showed moderate or good inhibitory efficacy to the growth of *A. fumigatus* in comparison to molecule **3a**, and all of them were superior to the reference drug fluconazole. In addition, aminoethyl compound **4b** and hydrazone derivative **5d** displayed promising antifungal activity against *C. albicans* with MIC values of 0.09 mmol/L and 0.03 mmol/L, respectively. Noticeably, oxime **7d** displayed excellent anti-*A. fumigatus* activity with MIC value of 0.02 mmol/L, which was 42-fold more potent than fluconazole, and it also possessed better antifungal activity than precursor **3a** against all tested fungal strains, indicating that the introduction of imine was beneficial to antifungal activity.

### Bactericidal kinetics

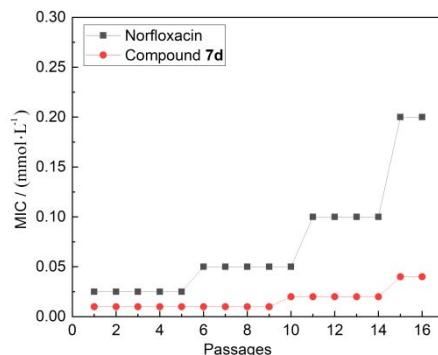
Bactericidal therapy might shorten the duration of bacterial infection and reduce the occurrence of bacterial resistant development.<sup>[23]</sup> So the bactericidal efficiency for quinazolonethiazole **7d** ( $6 \times$  MIC) was evaluated by a time-kill kinetic experiment against *P. aeruginosa*, and the results (Figure 4) revealed that bacterial concentration decreased by more than  $10^2$  CFU/mL in presence of potent compound **7d** within 2 h, which manifested quinazolonethiazole **7d** was effective in killing *P. aeruginosa*, having the possibility of alleviating bacterial resistance.

### Bacterial resistance study

The frequent evolution of drug-resistant pathogens imposes restrictions on the lifespan of antibiotic, thereby limiting the curative effect of infectious diseases, so the development of drug resistance is a critical criterion for evaluating antimicrobials.<sup>[24]</sup> The bacterial resistant results (Figure 5) of quinazolonethiazole **7d**



**Figure 4** Time-kill kinetics of quinazolonethiazole **7d** ( $6 \times$  MIC) against *P. aeruginosa*.



**Figure 5** Bacterial resistance test for quinazolonethiazole **7d** towards the *P. aeruginosa*.

showed that susceptibility of *P. aeruginosa* nearly was unaffected even after 9 passages, followed by slower resistant development compared to the clinical drug norfloxacin. This information indicated that quinazolonethiazole **7d** with low bacterial resistance might have future potential for further development as antibacterial agent.

### Cytotoxicity

Cytotoxicity is one of the most decisive factors in determining the fate of potential candidate molecules. Thiazole may undergo hepatotoxicity due to multi-step bioactivation to generate thioureas.<sup>[25]</sup> So the cell viability of LO2 cell (normal hepatocyte cell) was measured in the presence of highly active compound **7d** by MTT assay. The result in Figure 6 demonstrated that the LO2 cell viability treated by quinazolonethiazole **7d** remained around 100% at 0.01 mmol/L (concentration at which **7d** showed good inhibition against *P. aeruginosa*), while compound **7d** displayed low toxicity at 4 × MIC value. This manifested that quinazolonethiazole **7d** had no significant effect on the mammalian cell when inhibiting the growth of *P. aeruginosa*.

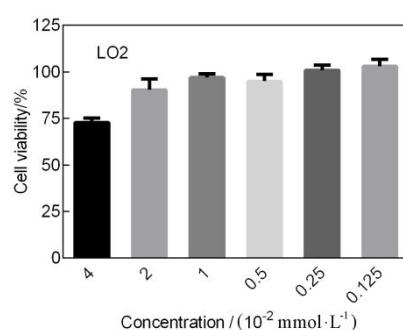


Figure 6 Cell viability of hexyl oxime derivative **7d** in LO2 cell.

### ADME study

The failures to develop many clinical candidates are attributed to poor pharmacokinetics and bioavailability in addition to efficacy and toxicity.<sup>[26]</sup> To better understand the pharmacokinetics and bioavailability of quinazolonethiazole **7d**, the ADME study was performed using online software SwissADME. The Lipinski rule is widely used in drug design and development on account of its important determinants in assessing the pharmacokinetic properties associated with oral bioavailability.<sup>[27]</sup> Table 2 showed that compound **7d** tended to have high oral bioavailability because it met the Lipinski rule and had the same oral bioavailability score with norfloxacin. Moreover, according to the Brain Or Intestinal EstimateD permeation method, the predictive results pointed out that compound **7d** displayed high gastrointestinal tract (GI) absorption and could not penetrate blood-brain barrier (BBB). All

the data suggested quinazolonethiazole **7d** *in silico* had excellent drug-likeness and appreciable pharmacokinetic profiles.

### Bacterial membrane permeabilization

Cell membranes can prevent antibiotics from entering bacteria or excrete antibiotics out of membrane to induce drug-resistance, so the development of membrane-targeted antimicrobial agents is expected to overcome bacterial resistance.<sup>[28]</sup> The estimation of membrane damage by quinazolonethiazole **7d** in different concentrations was performed with propidium iodide (PI), which is a common staining reagent that penetrates compromised membrane to produce fluorescence.<sup>[29]</sup> Figure 7 demonstrated the increase in fluorescence intensity of *P. aeruginosa* cells on treatment with increasing concentrations of compound **7d**, which further indicated the increased uptake of the dye by the membrane of compromised cells. The gradually increased number of red fluorescent cells under fluorescence microscopy provided additional support for the occurrence of bacterial membrane damage. In other words, quinazolonethiazole **7d** could damage the membrane of *P. aeruginosa* leading to cell death.

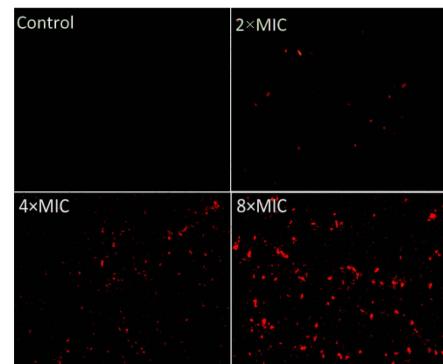
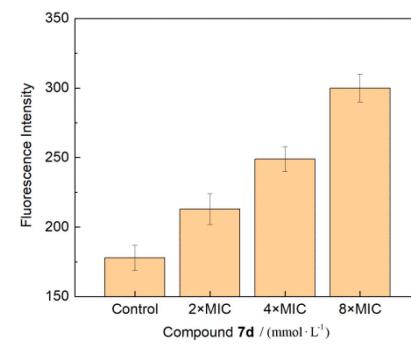


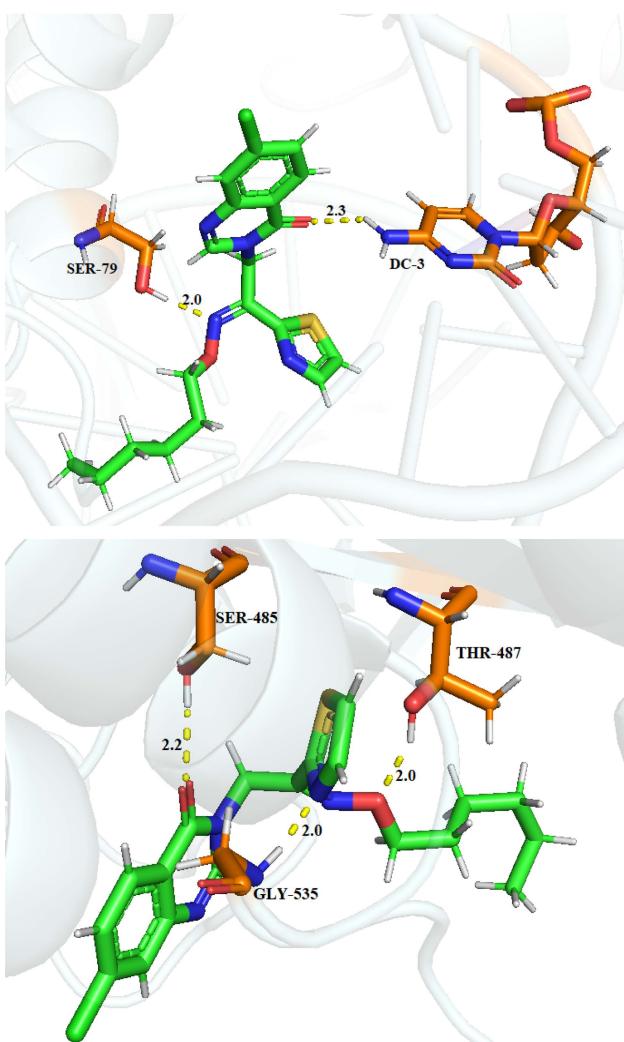
Figure 7 Estimation of membrane damage in *P. aeruginosa* treated with quinazolonethiazole **7d** by fluorescence and microscope.

### Interaction of quinazolonethiazole **7d** with topoisomerase IV and penicillin-binding protein (PBP)

In order to further provide appropriate antibacterial sights, the interactions between the highly bioactive molecule quinazolonethiazole **7d** and biomolecular enzymes topoisomerase IV or PBP (crystal data: protein data bank) were investigated by molecular docking.<sup>[30]</sup> Docking results for the topoisomerase IV (Figure 8A) demonstrated that the carbonyl group of quinazolone could form hydrogen bond with DC-3 of DNA at the distance of 2.3 Å. Noticeably, the nitrogen atom of imine fragment could bind with SER-79 residue through hydrogen bond, which further explained that the introduction of imine was beneficial to exert antibacterial activity. Besides, hydrogen bonds were formed between the carbonyl group of quinazolone and residue SER-485 of PBP, the oxygen atom of imine fragment and THR-487 residue (Figure 8B).

Table 2 The ADME data of quinazolonethiazole **7d**

	Quinazolonethiazole <b>7d</b>	Norfloxacin
MW (g/mol) < 500	404.91	319.33
Mlog <i>P</i> ≤ 4.15	2.21	1.04
H-bond acceptors < 10	5	4
H-bond donors < 5	0	2
Rotatable bonds < 10	9	3
Lipinski violations	0	0
Bioavailability Score	0.55	0.55
GI absorption	High	High
BBB permeant	No	No



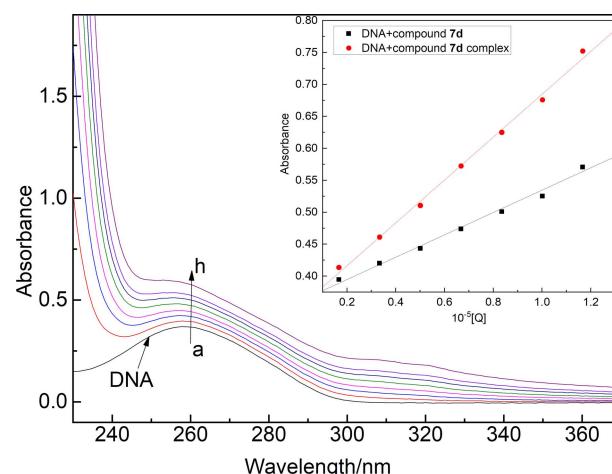
**Figure 8** Interaction view of quinazolonethiazole **7d** in the binding site of (A) DNA topoisomerase IV (PDB code: 3RAF) and (B) PBP (PDB code: 6VJE), respectively.

Furthermore, the nitrogen atom of thiazole was involved in the hydrogen bond with glycine on PBP, which revealed the importance of thiazole in antibacterial. All hydrogen bonds were conducive to stabilizing the complexes between enzyme and quinazolonethiazole, which might further provide clues regarding good antibacterial effect of quinazolonethiazole.

#### Interaction of quinazolonethiazole **7d** with DNA

As life-based biomacromolecule encoding genetic instructions, DNA is involved in the growth and development of organisms and also responsible for abnormality such as cancer and mutation.<sup>[31]</sup> Therefore, the study of the interaction between DNA and targeting compound can explain the action mechanism of some drugs, which is of great significance for finding antibacterial drugs with combating bacterial resistance, low cytotoxicity and good curative effect.<sup>[32]</sup> Calf thymus DNA is applied for DNA binding agent to perform *in vitro* interaction with quinazolonethiazole **7d**.

The ultraviolet-visible absorption in Figure 9 displayed that the absorbance peak of DNA (a constant concentration of  $7.44 \times 10^{-5}$  mol/L) progressively enhanced at 260 nm together with slightly blue shift as the proportionately improved concentration of **7d**. Meantime, the absorbance at 260 nm by adding up of quinazolonethiazole **7d** and free DNA was less than that of **7d**-DNA complex (inset of Figure 9). This proved a hyperchromic effect existing between DNA and **7d**, which might be due to DNA

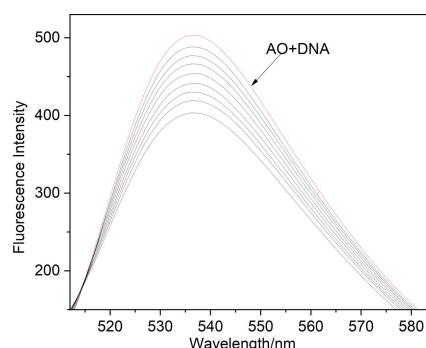


**Figure 9** Interaction between DNA and different concentrations of **7d** ( $0-1.75 \times 10^{-5}$  mol/L and in increment of  $1.75 \times 10^{-5}$  mol/L).

denaturation caused by interaction of DNA and molecule, leading to partial disintegration of DNA double helix and exposure of some DNA bases.<sup>[33-34]</sup> In a nutshell, the results suggested a strong interaction between DNA and quinazolonethiazole **7d**.

#### Competitive experiments of quinazolonethiazole **7d** and AO with DNA

To illuminate the binding model of DNA and quinazolonethiazole **7d**, further investigation was carried out by the fluorescent probe of acridine orange (AO), a nucleic acid binding dye that could intercalate into DNA to emit fluorescence.<sup>[35]</sup> The fluorescence intensity of the DNA-AO system in Figure 10 gradually decreased at around 540 nm as the amount of quinazolonethiazole **7d** increased, indicating that quinazolonethiazole **7d** could effectively intercalate DNA by substituting AO to exert powerful bioactivity.

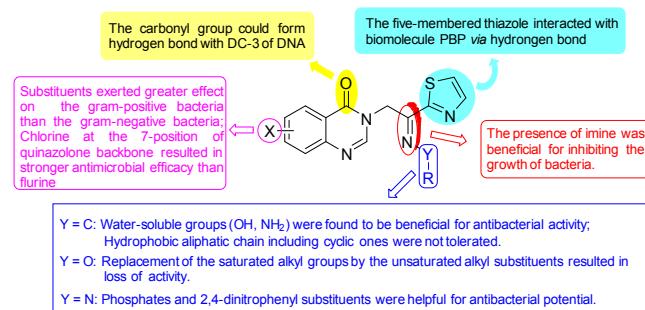


**Figure 10** Competitive experiments between compound **7d** and AO with DNA.

#### Conclusions

A type of new quinazolonethiazoles was developed via several easy synthetic steps comprising cyclization, bromination, nucleophilic substitution and condensation from aminobenzoic acids, and their medicinal chemical biological assay revealed that some target quinazolonethiazoles enjoyed good antibacterial activity toward the tested strains. Particularly, the hexyl oxime derivative **7d** showed the best antimicrobial activity, and also exhibited stronger activity with MIC value of 0.01 mmol/L than the reference drugs chloramphenicol and norfloxacin against *P. aeruginosa*. The SAR in Figure 11 suggested that the presence of imine was

beneficial for inhibiting the growth of bacteria, which was also demonstrated by the docking study of quinazolonethiazole **7d** with topoisomerase IV. In addition, the highly active molecule **7d** with low hepatotoxicity might damage bacterial cell by enhancing the permeability of membrane and disturb bacterial growth by intercalating into DNA, rendering unapparent resistant tendency and bactericidal activity. The *in silico* ADME investigation also revealed that quinazolonethiazole **7d** possessed excellent drug-likeness and considerable pharmacokinetic profiles. These results provided helpful information for further optimization of quinazolonethiazole as potential therapeutics for infectious diseases caused by bacteria.



**Figure 11** Brief biological summary of synthesized target quinazolonethiazoles.

## Experimental

**General methods.** All purchased reagents and solvents were used directly without further purification. Melting points: X–6 melting point apparatus; TLC analysis: pre-coated silica gel plates;  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectra: AVANCE III 600 MHz spectrometer with tetramethylsilane (TMS) as reference. Coupling constants ( $J$ ): hertz (Hz); Signals: singlet (s), doublet (d), triplet (t), quartet (q) as well as multiplet (m); Mass spectra: LCMS-2010A; High-resolution mass spectra (HRMS): IonSpec FTICR mass spectrometer with ESI resource. The compounds purity was determined to be >95% by HPLC (1 mL/min flow, 95%–60% linear gradient of solvent A (acetonitrile) in B (water) over 30 min).

**Quinazolone (2a–2d):** The synthesis of quinazolones **2a–2d** was done referring the reported literature.<sup>[36]</sup>

**Quinazolonethiazole (3a):** Compound **9** (8.81 g, 0.04 mol) and potassium carbonate (4.04 g, 0.03 mol) were added to acetonitrile with quinazolone **2a** (5.00 g, 0.03 mol). The mixture was stirred at 50 °C until the reaction was completed by TLC monitoring. The precipitated solid was filtered, washed with water and purified by silica gel column chromatography (eluent, dichloromethane/petroleum ether, 2/1, V/V) to afford desirable product **3a** (3.09 g) as yellow solid. Yield: 36.7%; m.p. 174–175 °C; Purity: 98%;  $^1\text{H}$  NMR (600 MHz, DMSO- $d_6$ )  $\delta$ : 8.45 (s, 1H, quinazolinone-2-H), 8.38 (d,  $J$  = 3.0 Hz, 1H, thiazole-4-H), 8.29 (d,  $J$  = 3.0 Hz, 1H, thiazole-5-H), 8.15 (d,  $J$  = 8.5 Hz, 1H, quinazolinone-5-H), 7.83 (d,  $J$  = 1.9 Hz, 1H, quinazolinone-8-H), 7.63 (dd,  $J$  = 8.5, 2.0 Hz, 1H, quinazolinone-6-H), 5.67 (s, 2H, CH<sub>2</sub>);  $^{13}\text{C}$  NMR (151 MHz, DMSO- $d_6$ )  $\delta$ : 186.58 (CH<sub>2</sub>CO), 163.97 (quinazolinone-4-C), 160.03, 150.11, 149.62, 146.03, 139.84, 129.41, 128.65, 128.14, 127.04, 120.60, 52.50 (CH<sub>2</sub>); HRMS (ESI) calcd. for C<sub>13</sub>H<sub>8</sub>ClN<sub>3</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 306.0104; found, 306.0100.

**Quinazolonethiazole (3b):** Yellow solid. Yield: 25.2%; m.p. 174–175 °C; Purity: 99%;  $^1\text{H}$  NMR (600 MHz, DMSO- $d_6$ )  $\delta$ : 8.45 (s, 1H, quinazolinone-2-H), 8.38 (d,  $J$  = 2.8 Hz, 1H, thiazole-4-H), 8.29 (d,  $J$  = 2.9 Hz, 1H, thiazole-5-H), 8.22 (dd,  $J$  = 8.7, 6.3 Hz, 1H, quinazolinone-5-H), 7.55 (dd,  $J$  = 9.9, 1.9 Hz, 1H, quinazolinone-8-H) 7.46 (td,  $J$  = 8.6, 2.1 Hz, 1H, quinazolinone-6-H), 5.67 (s, 2H, CH<sub>2</sub>);  $^{13}\text{C}$  NMR (151 MHz, DMSO- $d_6$ )  $\delta$ : 186.65, 167.09, 165.42,

163.98, 159.92, 150.77, 150.69, 150.05, 146.03, 129.79, 129.71, 129.40, 118.83, 116.51, 116.35, 113.10, 112.95, 52.43; HRMS (ESI) calcd. for C<sub>13</sub>H<sub>8</sub>FN<sub>3</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 290.0400; found, 290.0393.

**Quinazolonethiazole (3c):** Yellow solid. Yield: 49.9%; m.p. 214–215 °C; Purity: 99%;  $^1\text{H}$  NMR (600 MHz, DMSO- $d_6$ )  $\delta$ : 8.84 (d,  $J$  = 2.0 Hz, 1H, quinazolinone-5-H), 8.64–8.59 (m, 2H, quinazolinone-2-H, quinazolinone-7-H), 8.40 (d,  $J$  = 2.6 Hz, 1H, thiazole-4-H), 8.30 (d,  $J$  = 2.7 Hz, 1H, thiazole-5-H), 7.97 (d,  $J$  = 8.9 Hz, 1H, quinazolinone-8-H), 5.73 (s, 2H, CH<sub>2</sub>);  $^{13}\text{C}$  NMR (151 MHz, DMSO- $d_6$ )  $\delta$ : 186.25 (CH<sub>2</sub>CO), 163.83 (quinazolinone-4-C), 159.91, 152.54, 151.89, 146.08, 146.01, 129.80, 129.53, 129.18, 122.65, 121.81, 52.74 (CH<sub>2</sub>); HRMS (ESI) calcd. for C<sub>13</sub>H<sub>8</sub>N<sub>4</sub>O<sub>4</sub>S [M+H]<sup>+</sup>, 317.0345; found, 317.0340.

**Quinazolonethiazole (3d):** Yellow solid. Yield: 40.8%; m.p. 227–231 °C; Purity: 99%;  $^1\text{H}$  NMR (600 MHz, DMSO- $d_6$ )  $\delta$ : 8.55 (s, 1H, quinazolinone-2-H), 8.39 (d,  $J$  = 2.8 Hz, 1H, thiazole-4-H), 8.30 (d,  $J$  = 2.8 Hz, 1H, thiazole-5-H), 8.21 (d,  $J$  = 1.9 Hz, 1H, quinazolinone-7-H), 8.07 (d,  $J$  = 1.8 Hz, 1H, quinazolinone-5-H), 5.70 (s, 2H, CH<sub>2</sub>);  $^{13}\text{C}$  NMR (151 MHz, DMSO- $d_6$ )  $\delta$ : 186.25, 163.85, 159.18, 149.90, 146.07, 143.97, 134.93, 133.02, 131.93, 129.49, 124.85, 124.06, 52.75; HRMS (ESI) calcd. for C<sub>13</sub>H<sub>7</sub>Cl<sub>2</sub>N<sub>3</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 339.9714; found, 339.9707.

**Compound (4a):** Intermediate **3a** (100 mg, 0.33 mmol) and propylamine (39 mg, 0.66 mmol) in ethanol were condensed with acetic acid as catalyst at 80 °C. After the reaction was accomplished (monitored by TLC), the precipitated solid was filtered off, washed with ice water and recrystallized from ethanol to afford target compound **4a** (46 mg) as white solid. Yield: 40.7%; m.p. 200–201 °C; Purity: 97%;  $^1\text{H}$  NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.59 (s, 1H, quinazolinone-2-H), 8.18 (d,  $J$  = 8.6 Hz, 1H, quinazolinone-5-H), 7.80 (d,  $J$  = 3.1 Hz, 1H, thiazole-4-H), 7.67 (d,  $J$  = 1.4 Hz, 1H, quinazolinone-8-H), 7.42 (dd,  $J$  = 8.6, 1.6 Hz, 1H, quinazolinone-6-H), 7.33 (d,  $J$  = 3.1 Hz, 1H, thiazole-5-H), 5.19 (s, 2H, CH<sub>2</sub>), 3.91 (t,  $J$  = 6.9 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.83–1.76 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.03 (t,  $J$  = 7.4 Hz, 3H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>);  $^{13}\text{C}$  NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$ : 169.55 (C=N), 160.28 (quinazolinone-4-C), 154.79 (aromatic C), 149.32, 148.91, 143.34, 140.63, 128.15, 127.87, 127.07, 122.41, 120.29, 54.27, 42.08, 24.08, 12.01 (CH<sub>3</sub>); HRMS (ESI) calcd. for C<sub>16</sub>H<sub>15</sub>ClN<sub>4</sub>OS [M+H]<sup>+</sup>, 347.0733; found, 347.0723.

**Compound (4b):** White solid. Yield: 39.5%; m.p. 190–191 °C; Purity: 95%;  $^1\text{H}$  NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.23 (d,  $J$  = 8.5 Hz, 1H, quinazolinone-5-H), 8.03 (s, 1H, quinazolinone-2-H), 7.80 (d,  $J$  = 3.2 Hz, 1H, thiazole-4-H), 7.68 (d,  $J$  = 1.5 Hz, 1H, quinazolinone-8-H), 7.45 (dd,  $J$  = 8.5, 1.7 Hz, 1H, quinazolinone-6-H), 7.27 (d,  $J$  = 3.3 Hz, 1H, thiazole-5-H), 4.52 (s, 2H, CH<sub>2</sub>), 3.05 (s, 2H, NH<sub>2</sub>), 2.97 (dd,  $J$  = 10.1, 5.2 Hz, 2H, CH<sub>2</sub>NH<sub>2</sub>), 2.89 (dd,  $J$  = 10.1, 5.3 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>);  $^{13}\text{C}$  NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$ : 177.63 (C=N), 162.20 (quinazolinone-4-C), 149.09, 144.05, 140.82, 128.37, 127.83, 127.11, 120.85, 120.08, 81.85, 53.75 (CH<sub>2</sub>), 46.67; HRMS (ESI) calcd. for C<sub>14</sub>H<sub>15</sub>ClN<sub>5</sub>O<sub>4</sub>S [M+H]<sup>+</sup>, 348.0686; found, 348.0678.

**Compound (4c):** White solid. Yield: 35.2%; m.p. 190–191 °C; Purity: 99%;  $^1\text{H}$  NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.23 (d,  $J$  = 8.5 Hz, 1H, quinazolinone-5-H), 8.12 (s, 1H, quinazolinone-2-H), 7.85 (d,  $J$  = 2.4 Hz, 1H, thiazole-4-H), 7.70 (s, 1H, quinazolinone-8-H), 7.46 (d,  $J$  = 8.5 Hz, 1H, quinazolinone-6-H), 7.33 (d,  $J$  = 2.4 Hz, 1H, thiazole-5-H), 4.83 (d,  $J$  = 14.3 Hz, 1H, CH<sub>2</sub>), 4.46 (d,  $J$  = 14.3 Hz, 1H, CH<sub>2</sub>), 3.91 (dd,  $J$  = 12.7, 6.5 Hz, 1H, CH<sub>2</sub>OH), 3.73 (dd,  $J$  = 13.8, 6.8 Hz, 1H, CH<sub>2</sub>OH), 3.49 (s, 1H, OH), 3.14 (d,  $J$  = 4.8 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>OH);  $^{13}\text{C}$  NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$ : 172.80, 161.80, 149.09, 148.89, 144.30, 140.99, 128.57, 128.07, 127.28, 120.98, 120.32, 96.07, 67.72, 51.33, 46.20; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>13</sub>ClN<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 349.0526; found, 349.0528.

**Compound (4d):** White solid. Yield: 58.0%; m.p. 189–193 °C; Purity: 99%;  $^1\text{H}$  NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.61 (s, 1H, quinazolinone-2-H), 8.21 (d,  $J$  = 8.6 Hz, 1H, quinazolinone-5-H), 7.81 (d,  $J$  = 3.1 Hz, 1H, thiazole-4-H), 7.67 (d,  $J$  = 1.4 Hz, 1H, quinazolinone-8-H), 7.45–7.41 (m, 1H, quinazolinone-6-H), 7.31 (d,  $J$  = 3.1 Hz, 1H,

thiazole-5-*H*), 5.38 (s, 2H, *CH*<sub>2</sub>), 3.78–3.73 (m, 1H, *CH*), 1.15–1.10 (m, 2H, cyclopropylimino-*CH*<sub>2</sub>), 1.07 (d, *J* = 2.9 Hz, 2H, cyclopropylimino-*CH*<sub>2</sub>); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ: 169.94 (*C=N*), 160.41 (quinazolinone-4-*C*), 153.80, 148.98, 143.38, 140.74, 128.37, 128.00, 127.19, 122.12, 120.48, 50.92, 41.64, 35.59, 11.37; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>13</sub>CIN<sub>4</sub>OS [M+H]<sup>+</sup>, 345.0577; found, 345.0574.

Compound (**4e**): White solid. Yield: 32.8%; m.p. 193–196 °C; Purity: 99%; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ: 8.61 (s, 1H, quinazolinone-2-*H*), 8.21 (d, *J* = 8.6 Hz, 1H, quinazolinone-5-*H*), 7.81 (d, *J* = 3.1 Hz, 1H, thiazole-4-*H*), 7.67 (d, *J* = 1.4 Hz, 1H, quinazolinone-8-*H*), 7.45–7.41 (m, 1H, quinazolinone-6-*H*), 7.31 (d, *J* = 3.1 Hz, 1H, thiazole-5-*H*), 5.38 (s, 2H, *CH*<sub>2</sub>), 3.78–3.73 (m, 1H, *CH*), 1.15–1.10 (m, 2H, cyclopropylimino-*CH*<sub>2</sub>), 1.07 (d, *J* = 2.9 Hz, 2H, cyclopropylimino-*CH*<sub>2</sub>); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ: 170.01, 160.16, 152.83, 149.08, 148.86, 143.25, 140.59, 128.16, 127.85, 127.05, 122.30, 120.33, 62.40, 50.79, 41.87, 34.90, 34.78, 24.95, 24.56; HRMS (ESI) calcd. for C<sub>18</sub>H<sub>17</sub>CIN<sub>4</sub>OS [M+H]<sup>+</sup>, 373.0890; found, 373.0890.

Compound (**5a**): To a mixture of intermediate **3a** (100 mg, 0.33 mmol) and phenylhydrazine (71 mg, 0.66 mmol) in ethanol was added hydrochloric acid. The system was stirred at 80 °C until the reaction was completed (monitored by TLC, dichloromethane). The precipitated solid was filtered off, washed with ice water and ethanol to afford target compound (79 mg) as yellow solid. Yield: 61.0%; m.p. 228–229 °C; Purity: 98%; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ: 13.31 (s, 1H, NH), 8.36 (s, 1H, quinazolinone-2-*H*), 8.25 (d, *J* = 8.5 Hz, 1H, quinazolinone-5-*H*), 7.99 (d, *J* = 3.1 Hz, 1H, thiazole-4-*H*), 7.72 (d, *J* = 1.5 Hz, 1H, quinazolinone-8-*H*), 7.45 (dd, *J* = 8.5, 1.6 Hz, 1H, quinazolinone-6-*H*), 7.38 (d, *J* = 3.2 Hz, 1H, thiazole-5-*H*), 7.29–7.26 (m, 2H, Ph-3,5-*H*), 7.16 (d, *J* = 8.0 Hz, 2H, Ph-2,6-*H*), 6.94 (t, *J* = 7.2 Hz, 1H, Ph-4-*H*), 5.27 (s, 2H, *CH*<sub>2</sub>); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ: 161.19, 160.50, 149.20, 147.68, 143.62, 143.24, 140.78, 129.49, 128.56, 128.08, 127.34, 124.44, 122.11, 120.63, 118.65, 113.97, 48.46; HRMS (ESI) calcd. for C<sub>19</sub>H<sub>14</sub>CIN<sub>5</sub>OS [M+H]<sup>+</sup>, 396.0686; found, 396.0686.

Compound (**5b**): Yellow solid. Yield: 60.7%; m.p. > 250 °C; Purity: 98%; <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ: 12.39 (s, 1H, NH), 8.96 (d, *J* = 2.4 Hz, 1H, (2,4-dinitrophenyl)hydrazineylidene-3-*H*), 8.81 (s, 1H, quinazolinone-2-*H*), 8.56 (dd, *J* = 9.4, 2.3 Hz, 1H, (2,4-dinitrophenyl)hydrazineylidene-5-*H*), 8.13 (d, *J* = 8.6 Hz, 1H, (2,4-dinitrophenyl)hydrazineylidene-6-*H*), 8.01–7.98 (m, 2H, quinazolinone-5-*H*, thiazole-4-*H*), 7.88 (d, *J* = 3.0 Hz, 1H, quinazolinone-6-*H*), 7.78 (s, 1H, quinazolinone-8-*H*), 7.58 (d, *J* = 8.6 Hz, 1H, thiazole-5-*H*), 5.38 (s, 2H, *CH*<sub>2</sub>). HRMS (ESI) calcd. for C<sub>19</sub>H<sub>12</sub>CIN<sub>5</sub>O<sub>5</sub> [M+H]<sup>+</sup>, 486.0387; found, 486.0379.

Compound (**5c**): Yellow solid. Yield: 82.7%. m.p. > 250 °C; Purity: 97%; <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ: 12.22 (s, 1H, NH), 8.88 (s, 2H, isonicotinohydrazide-2,6-*H*), 8.78 (s, 1H, quinazolinone-2-*H*), 8.18 (d, *J* = 8.2 Hz, 1H, quinazolinone-5-*H*), 7.98 (s, 3H, isonicotinohydrazide-3,5-*H*, quinazolinone-8-*H*), 7.85 (s, 1H, thiazole-4-*H*), 7.77 (s, 1H, quinazolinone-6-*H*), 7.60 (d, *J* = 8.5 Hz, 1H, thiazole-5-*H*), 5.52 (s, 2H, *CH*<sub>2</sub>). HRMS (ESI) calcd. for C<sub>19</sub>H<sub>13</sub>CIN<sub>6</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 425.0587; found, 425.0593.

Compound (**5d**): White solid. Yield: 56.3%; m.p. 223–225 °C; Purity: 99%; <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ: 12.20 (d, *J* = 36.5 Hz, 1H, NH), 8.54 (s, 1H, quinazolinone-2-*H*), 8.22 (d, *J* = 2.6 Hz, 1H, quinazolinone-5-*H*), 8.19 (d, *J* = 8.5 Hz, 1H, thiazole-4-*H*), 8.16 (d, *J* = 2.7 Hz, 1H, thiazole-5-*H*), 7.87 (s, 1H, quinazolinone-8-*H*), 7.63 (d, *J* = 8.5 Hz, 1H, quinazolinone-6-*H*), 7.26 (t, *J* = 7.6 Hz, 4H, diphenyl-3,5-*H*), 7.17 (t, *J* = 7.3 Hz, 2H, diphenyl-4-*H*), 6.95 (d, *J* = 7.8 Hz, 4H, diphenyl-2,6-*H*), 5.36 (s, 2H, *CH*<sub>2</sub>); <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>) δ: 159.47, 158.72, 149.77, 149.52, 149.48, 149.16, 143.40, 139.24, 129.74, 128.22, 127.48, 126.52, 125.41, 122.82, 120.34, 120.14, 120.11, 47.63; HRMS (ESI) calcd. for C<sub>25</sub>H<sub>19</sub>CIN<sub>5</sub>O<sub>4</sub>S [M+H]<sup>+</sup>, 552.0662; found, 552.0656.

Compound (**5e**): To a mixture of intermediate **3a** (100 mg,

0.33 mmol) and diphenyl hydrazinylphosphonate (165 mg, 0.66 mmol) in ethanol was added sodium acetate (53 mg, 0.66 mmol). The system was stirred at 80 °C until the reaction was completed (monitored by TLC, dichloromethane). The precipitated solid was filtered off, washed with ice water and ethanol to afford target compound (112 mg) as white solid. Yield: 71.3%; m.p. 234–235 °C; Purity: 96%; <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ: 11.38 (d, *J* = 26.9 Hz, 1H, NH), 8.37 (s, 1H, quinazolinone-2-*H*), 8.16 (d, *J* = 8.5 Hz, 1H, quinazolinone-5-*H*), 8.05 (d, *J* = 3.1 Hz, 1H, thiazole-4-*H*), 7.85 (d, *J* = 3.1 Hz, 1H, thiazole-5-*H*), 7.79 (d, *J* = 1.5 Hz, 1H, quinazolinone-8-*H*), 7.61 (dd, *J* = 8.5, 1.7 Hz, 1H, quinazolinone-6-*H*), 6.98 (t, *J* = 7.7 Hz, 2H, phenyl-3,5-*H*), 6.92 (d, *J* = 7.8 Hz, 2H, phenyl-2,6-*H*), 6.80 (t, *J* = 7.1 Hz, 1H, phenyl-4-*H*), 5.16 (s, 2H, *CH*<sub>2</sub>); <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>) δ: 160.14, 159.39, 153.84, 148.95, 142.89, 139.03, 128.36, 128.29, 127.33, 126.41, 126.30, 121.44, 120.31, 120.28, 119.74, 48.53; HRMS (ESI) calcd. for C<sub>14</sub>H<sub>15</sub>CIN<sub>5</sub>O<sub>4</sub>S [M+H]<sup>+</sup>, 476.0349; found, 476.0345.

Compound (**6**): White solid. Yield: 76.3%; m.p. > 250 °C; Purity: 98%; <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ: 12.87 (s, 1H, OH), 8.55 (s, 1H, quinazolinone-2-*H*), 8.15 (d, *J* = 8.4 Hz, 1H, quinazolinone-5-*H*), 8.13 (d, *J* = 3.2 Hz, 1H, thiazole-4-*H*), 8.09–8.07 (m, 1H, thiazole-5-*H*), 7.77 (s, 1H, quinazolinone-8-*H*), 7.59 (dd, *J* = 8.5, 2.0 Hz, 1H, quinazolinone-6-*H*), 5.47 (s, 2H, *CH*<sub>2</sub>); <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>) δ: 163.69, 159.99, 153.23, 150.73, 144.37, 142.75, 139.58, 128.65, 127.89, 126.85, 124.98, 120.92, 46.82; HRMS (ESI) calcd. for C<sub>13</sub>H<sub>9</sub>CIN<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 321.0213; found, 321.0204.

Compound (**7a**): Compound **6** (200 mg, 0.62 mmol) and potassium carbonate (171 mg, 1.24 mmol) were stirred in acetonitrile at 50 °C for 0.5 h, then cooled to room temperature for adding methyl iodide (176 mg, 1.24 mmol) and continued to reaction at 50 °C. After the reaction was completed (monitored by TLC, dichloromethane), acetonitrile was removed and the residue was extracted with ethyl acetate (3 × 20 mL), dried over anhydrous sodium sulfate and purified by silica gel column chromatography (eluent, dichloromethane/petroleum ether, 2/1, V/V) to afford desirable product (137 mg) as white solid. Yield: 65.6%; m.p. 177–178 °C; Purity: 97%; <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>) δ: 8.55 (s, 1H, quinazolinone-2-*H*), 8.15 (d, *J* = 7.9 Hz, 3H, quinazolinone-5-*H*, thiazole-4-*H*, thiazole-5-*H*), 7.79 (s, 1H, quinazolinone-8-*H*), 7.60 (d, *J* = 8.5 Hz, 1H, quinazolinone-6-*H*), 5.47 (s, 2H, *CH*<sub>2</sub>), 3.95 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>) δ: 159.38, 152.37, 150.07, 149.01, 144.65, 142.58, 139.08, 128.18, 127.33, 126.38, 125.28, 120.26, 63.12, 46.16; HRMS (ESI) calcd. for C<sub>14</sub>H<sub>11</sub>CIN<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 335.0369; found, 335.0363.

Compound (**7b**): White solid. Yield: 74.5%; m.p. 153–154 °C; Purity: 99%; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ: 8.34 (s, 1H, quinazolinone-2-*H*), 8.26 (d, *J* = 5.7 Hz, 1H, quinazolinone-5-*H*), 7.99 (d, *J* = 3.2 Hz, 1H, thiazole-4-*H*), 7.70 (d, *J* = 1.9 Hz, 1H, quinazolinone-8-*H*), 7.59 (d, *J* = 3.2 Hz, 1H, thiazole-5-*H*), 7.43 (dd, *J* = 3.6, 2.0 Hz, 1H, quinazolinone-6-*H*), 5.45 (s, 2H, *CH*<sub>2</sub>), 4.36 (q, *J* = 7.1 Hz, 2H, OCH<sub>2</sub>), 1.36 (t, *J* = 7.1 Hz, 3H, CH<sub>3</sub>); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ: 160.33, 153.40, 148.76, 144.05, 143.33, 142.42, 140.45, 128.41, 127.80, 127.73, 126.93, 123.51, 120.68, 71.99, 46.86, 14.50; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>14</sub>CIN<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 349.0526; found, 349.05203.

Compound (**7c**): White solid. Yield: 54.0%; m.p. 172–173 °C; Purity: 96%; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ: 8.26 (s, 1H, quinazolinone-2-*H*), 8.21 (d, *J* = 8.6 Hz, 1H, quinazolinone-5-*H*), 7.85 (d, *J* = 3.0 Hz, 1H, quinazolinone-8-*H*), 7.70 (d, *J* = 1.5 Hz, 1H, thiazole-4-*H*), 7.44 (dd, *J* = 8.6, 1.7 Hz, 1H, quinazolinone-6-*H*), 7.33 (d, *J* = 3.2 Hz, 1H, thiazole-5-*H*), 5.35 (s, 2H, *CH*<sub>2</sub>), 4.27 (t, *J* = 6.6 Hz, 2H, OCH<sub>2</sub>), 1.68–1.62 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.50–1.42 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 0.88 (t, *J* = 7.4 Hz, 3H, CH<sub>3</sub>); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ: 162.74, 160.10, 148.83, 148.79, 148.53, 147.58, 143.31, 140.49, 128.25, 128.13, 128.06, 127.82, 126.91, 126.88, 120.57, 76.08, 50.78, 31.00, 30.74, 18.91, 13.68; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>17</sub>CIN<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 377.0839; found, 377.0839.

**Compound (7d):** White solid. Yield: 79.7%; m.p. 172–173 °C; Purity: 99%; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ: 8.33 (s, 1H, quinazoline-2-H), 8.25 (d, J = 2.8 Hz, 1H, quinazolinone-5-H), 7.99 (d, J = 3.2 Hz, 1H, thiazole-4-H), 7.70 (d, J = 1.8 Hz, 1H, quinazolinone-8-H), 7.59 (d, J = 3.2 Hz, 1H, thiazole-5-H), 7.44–7.41 (m, 1H, quinazolinone-6-H), 5.45 (s, 2H, CH<sub>2</sub>), 4.31 (t, J = 6.6 Hz, 2H, OCH<sub>2</sub>), 1.75–1.70 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.39 (dd, J = 10.3, 4.5 Hz, 2H, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 1.28 (dd, J = 7.1, 3.5 Hz, 4H, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 0.86 (t, J = 6.9 Hz, 3H, CH<sub>3</sub>); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ: 159.93, 159.86, 152.96, 150.63, 149.58, 144.75, 143.14, 139.58, 128.70, 127.83, 127.80, 126.91, 126.81, 125.71, 120.81, 75.97, 46.73, 31.26, 28.87, 25.45, 22.29, 14.26; HRMS (ESI) calcd. for C<sub>19</sub>H<sub>21</sub>ClN<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 405.1152; found, 405.1145.

**Compound (7e):** Yellow solid. Yield: 63.0%; m.p. 177–178 °C; Purity: 99%; <sup>1</sup>H NMR (600 MHz, DMSO-d<sub>6</sub>) δ: 8.54 (s, 1H, quinazolinone-2-H), 8.19 (d, J = 3.2 Hz, 1H, quinazolinone-5-H), 8.18 (d, J = 3.2 Hz, 1H, thiazole-4-H), 8.15 (d, J = 8.5 Hz, 1H, quinazolinone-8-H), 7.78 (d, J = 1.9 Hz, 1H, thiazole-5-H), 7.60 (dd, J = 8.5, 2.0 Hz, 1H, quinazolinone-6-H), 5.50 (s, 2H, CH<sub>2</sub>), 4.84 (d, J = 2.3 Hz, 2H, OCH<sub>2</sub>), 3.53 (t, J = 2.2 Hz, 1H, CH). <sup>13</sup>C NMR (151 MHz, DMSO-d<sub>6</sub>) δ: 174.75 (C=N), 162.93 (s, quinazolinone-4-C), 150.51 (aromatic C), 142.97, 138.00, 130.30, 120.34, 117.84 (C≡CH), 114.04 (C≡CH), 69.89 (OCH<sub>2</sub>), 60.76 (CH<sub>2</sub>). HRMS (ESI) calcd. for C<sub>16</sub>H<sub>11</sub>ClN<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 359.0369; found, 359.03662.

**Compound (7f):** Yellow solid. Yield: 56.6%; m.p. 178–179 °C; Purity: 99%; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ: 8.33 (s, 1H, quinazolinone-2-H), 8.23 (d, J = 8.5 Hz, 1H, quinazolinone-5-H), 8.06 (d, J = 3.1 Hz, 1H, thiazole-4-H), 7.72 (d, J = 2.8 Hz, 2H, quinazolinone-8-H, thiazole-5-H), 7.45 (dd, J = 8.5, 1.8 Hz, 1H, quinazolinone-6-H), 5.49 (s, 2H, CH<sub>2</sub>), 4.95 (s, 2H, OCH<sub>2</sub>). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ: 160.34 (C=N), 152.37, 148.43, 147.74, 143.72, 142.98, 140.69, 128.33, 127.93, 127.12, 125.08, 120.53, 114.69, 60.07, 46.97. HRMS (ESI) calcd. for C<sub>15</sub>H<sub>10</sub>ClN<sub>5</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 360.0322; found, 360.03198.

**Compound (7g):** White solid. Yield: 45.8%; m.p. 155–157 °C; Purity: 99%; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ: 8.31 (s, 1H, quinazolinone-2-H), 8.24 (dd, J = 8.5, 4.0 Hz, 1H, quinazolinone-5-H), 7.99 (dd, J = 4.4, 2.4 Hz, 1H, thiazole-4-H), 7.71–7.68 (m, 1H, quinazolinone-8-H), 7.60 (t, J = 2.6 Hz, 1H, thiazole-5-H), 7.44–7.40 (m, 1H, quinazolinone-6-H), 6.03–5.96 (m, 1H, OCH<sub>2</sub>CH), 5.45 (s, 2H, CH<sub>2</sub>), 5.32–5.28 (m, 1H, OCH<sub>2</sub>CH=CH<sub>2</sub>), 5.22 (d, J = 9.2 Hz, 1H, OCH<sub>2</sub>CH=CH<sub>2</sub>), 4.79 (d, J = 5.0 Hz, 2H, OCH<sub>2</sub>); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ: 160.34, 153.27, 148.69, 144.51, 143.37, 142.49, 140.44, 132.76, 132.71, 128.39, 128.24, 127.79, 127.72, 126.96, 123.76, 120.69, 119.08, 118.94, 46.83, 30.87; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>13</sub>ClN<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>, 361.0526; found, 361.05298.

## Supporting Information

The supporting information for this article is available on the WWW under <https://doi.org/10.1002/cjoc.202000627>.

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