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## Design, synthesis and biological evaluation of glycolamide, glycinamide, and $\beta$ -amino carbonyl 1,2,4-triazole derivatives as DPP-4 inhibitors

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#### ABSTRACT

Through modification of the skeleton of Sitagliptin and Vildagliptin, we successfully synthesized and built-up four series of 1,2,4-triazole derivatives, containing N,O-disubstituted glycolamide, N,N'-disubstituted glycinamide,  $\beta$ -amino ester, and  $\beta$ -amino amide as linkers, for the development of new dipeptidyl peptidase 4 (DPP-4) inhibitors. The synthetic strategy for glycolamides or glycinamides involved convenient two-steps reaction: functionalized transformation of 2-chloro-N-(2,4,5-triflurophenyl)acetamide 9 (hydroxylation or amination) and esterification or amidation of 1,2,4-triazole-3-carboxylic acid. On the other hand, the one-pot synthesis procedure, including substitution and deprotection, was developed for the preparation of  $\beta$ -amino carbonyl 1,2,4triazoles from (1H-1,2,4-triazol-3-yl)methanol 12 or (1H-1,2,4-triazol-3-yl)methanamine 13 and Boc-(R)-3amino-4-(2,4,5-trifluoro-phenyl)-butyric acid 14. All of glycolamides, glycinamides, and β-amino carbonyl 1,2,4triazoles were also evaluated against DPP-4 inhibitory activity. Based on the SAR study of DPP-4 inhibitory capacity,  $\beta$ -amino ester 5n and  $\beta$ -amino amide 1,2,4-triazoles 6d and 6p possessed the significant inhibition of DPP-4 (IC<sub>50</sub> < 51.0 nM), particularly for compound **6d** (IC<sub>50</sub> = 34.4 nM). The selectivity evaluation indicated compound 5n and 6p had excellent selectivity over QPP, DPP-8, and DPP-9. In addition, the docking results revealed compounds **5n** and **6p** provided stronger  $\pi$ - $\pi$  stacking interaction with residue Phe357 than 1,5-disubstituted 1,2,4-triazole 6d and Sitagliptin 1. In summary, compounds 5n and 6p could be promising lead compounds for further development of DPP-4 inhibitor.

minimal side effects.

inhibitors have been gained attention due to the well-tolerated and

followed by Vildagliptin, Saxagliptin, Linagliptin, and Alogliptin [13,14].

The chemical structure of Sitagliptin 1 is divided into three parts: 2,4,5-

triflorophenyl fragment pharmacophore, enantiomerically pure β-amino

carbonyl linker, and the 5,6,7,8-tetrahydro-[1,2,4]triazolo[4,3-a]pyri-

dine fragment (Fig. 1) [15]. Although numerous literatures on the syn-

thesis of Sitagliptin have appeared [16–18], the enantioselective enamine reduction via efficient catalytic route was widely utilized to

generate the pure  $\beta$ -amino carbonyl linker in Sitagliptin scaffold [19].

Sitagliptin 1 is the first marketed selective inhibitor of DPP-4 [12],

#### 1. Introduction

Dipeptidyl peptidase IV (DPP-4) inhibitors are a new type of oral hypoglycemic drugs that inhibit the enzyme DPP-4 for treating patients with type 2 diabetes [1–4]. DPP-4 is an abundant enzyme existing in most cell types. It can deactivate glucagon-like peptide 1 (GLP-1) by cleaving peptide bonds [5]. The GLP-1 has multiple physiological functions, including insulin stimulation, glucagon inhibition, and delayed gastric emptying [6]. Inhibition of DPP-4 can elevate the concentration of active GLP-1 to increase insulin secretion and reduction in HbAlc without weight gain and have a low risk of hypoglycemia [7–11]. Hence, DPP-4

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However, this method is an expensive and non-green procedure due to using the metal reagent-based catalysts. Herein, we revaluated that the  $\beta$ -amino carbonyl linker necessitated a simplified modification.

Sitagliptin 1 is structurally a non-substrate-like DPP-4 inhibitor, which is non-peptide mimetic compound that occupies the hydrophobic pocket of the enzyme binding site via the trifluorophenyl moiety. On the other hand, Vildagliptin 2 is a substrate-like DPP-4 inhibitor with a cvanopyrrolidine ring that mimic DPP-4 substrates to occupy the same hydrophobic pocket of the enzyme binding site [20]. Despite the diverse chemical structures of DPP-4 inhibitors, the  $\beta$ -amino carbonyl linker of Sitagliptin 1 and glycinamide linker of Vildagliptin 2 have common interactions with key residues that play an important role in the inhibitory activity through hydrogen bonds [20,21]. Based on the different structures but similar binding sites results of Sitagliptin 1 and Vildagliptin 2, we assumed that the linker structure and length of DPP-4 inhibitors might be a key factor in the binding mode. Moreover, functional glycinamide linkers also play a crucial role in the investigation of biological processes in modern pharmacology, biochemistry, and medicinal chemistry [22,23]. Thus, we designed four series of compounds 3-6with different linkers, including glycolamide, glycinamide, β-amino amide, and  $\beta$ -amino ester groups in a design of functional 1.2.4-triazole modules in which 2,4,5-triflorophenyl fragment was maintained into the architecture for modulating the heterocyclic moiety to structural variation (Fig. 2). Based on the considerable attracted academic interest, these designed 1,2,4-triazole compounds with various linkers were evaluated DPP-4 inhibitory activity compared with Sitagliptin 1 [24]. The selectivity of DPP-4 over QPP, DPP-8, and DPP-9 for each compound were also determined by the IC<sub>50</sub> values in a plaque assay to discovery novel DPP-4 inhibitors with increasing safety and durability for the treatment of T2DM.

#### 2. Result and discussion

## 2.1. Design of N,O-disubstituted glycolamides 3, N,N-disubstituted glycinamides 4, and $\beta$ -amino carbonyl 1,2,4-triazoles 5 and 6

To investigate the binding structural mode of Sitagliptin **1** and Vildagliptin **2** to DPP-4, the advanced docking program iGEMDOCK v2.1 was used to simulate the binding domain between inhibitors and DPP-4 enzyme [25]. The design approach was followed the rational Sitagliptin **1** and Vildagliptin **2** and carried out by structural optimization to pursue high in vitro activity, high selectivity and low cytotoxicity. To earn these benefits, we calculated the length of the linker and the binding mode of the kinase domain of DPP-4 to generate the new designed structures of *N*,*O*-disubstituted glycolamides **3**, *N*,*N*-disubstituted glycinamides **4**, and  $\beta$ -amino carbonyl 1,2,4-triazoles **5** and **6** grafting 2,4,5-triflorophenyl fragment (Fig. 2). The linker length is ranged between 4.6 Å and 8.5 Å from the standard drugs Sitagliptin **1** and Vildagliptin **2**.

Glycolamides **3** and glycinamides **4** were divided into two fragments, N-(2,4,5-trifluorophenyl)-2-hydroxyacetamide or N-(2,4,5trifluorophenyl)-2-aminoacetamide and 1,2,4-triazole heterocyclic moiety **7** (Scheme 1 and Fig. 2). At first, both 1,3-disubstituted 1,2,4triazoles **7a–g** [26a] and 1,3,5-trisubstituted 1,2,4-triazoles **7h–q** [26b] were synthesized by following our previous published literature procedure [26]. Their preliminary DPP-4 inhibition data were presented in Table 1. Based on the inhibitory activities against DPP-4, 1,3-disubstituted 1,2,4-triazoles **7a–e** and 1,3,5-trisubstituted 1,2,4-triazoles **71–q** were found possessed better inhibition rates (DPP-4 Inh@1mM  $\geq$ 35%). Therefore, compounds **7a–e** and **71–q** were further selected as promising targets for synthesis of *N*,O-disubstituted glycolamides **3**, *N*,N-disubstituted glycinamides **4**, and  $\beta$ -amino carbonyl 1,2,4-triazoles **5** and **6**.

#### 2.2. Chemistry

## 2.2.1. Synthesis of N,O-disubstituted glycolamides **3** and N,N-disubstituted glycinamides **4**

Because the triazolopiperazine moiety of Sitagliptin 1 was recognized as an important pharmacophore that contributed to its good pharmacokinetic profile, potency, and selectivity [12,27], a new series of heterocycle moieties were investigated. Preliminarily, we focused on the evaluation of the following variations on 1,2,4-triazole structure. As outlined retrosynthetically in Scheme 1, both product targets including 1,2,4-triazole moiety N,O-disubstituted glycolamides 3 and N,N-disubstituted glycinamides 4 could be accessed from key intermediates N-(2,4,5-trifluorophenyl)-2-hydroxyacetamide 10 or N-(2,4,5-trifluorophenyl)-2-aminoacetamide 11 treated with 1H-1,2,4triazole-3-carboxylic acids 8 [28], which were hydrolyzed from methyl 1H-1,2,4-triazole-3-carboxylates 7 [26]. Compounds 10 and 11 were obtained from their  $\alpha$ -chloro N-arylacetamide precursor 9 through hydrolysis with cesium formate [29] or amination with aqueous ammonium hydroxide [30].  $\alpha$ -Chloro *N*-arylacetamide **9** was prepared from 2.4.5-trifluorobenzenamine reacted towards 2-chloroacetvl chloride via our previous published procedure [31].

With an effective synthesis of intermediates 2-hydroxyacetamide 10 and 2-aminoacetamide 11, we examined the two-steps synthesies involving amidation and hydrolysis or amination from 2,4,5-trifluorobenzenamine (Scheme 2). At the outset, the reaction of 2,4,5-trifluorobenzenamine and 2-chloroacetyl chloride proceeded successfully in refluxing toluene for 3 h to give  $\alpha$ -chloro *N*-arylacetamide 9 in 92% high yield [31]. Subsequently, the hydrolysis of N-arylacetamide 9 which was performed at reflux for 6 h on treatment with 3.0 equivalents of cesium formate (HCO<sub>2</sub>Cs) [29] in EtOH solution to afford 2-hydroxyacetamide 10 efficiently in 86% yield. We then evaluated the scope of the amination using N-arylacetamide 9 at 60 °C for 3 h under the EtOH/aqueous ammonium hydroxide condition [30]. The desired 2-aminoacetamide product 11 was obtained in 88% yield (Scheme 2). Subsequently, a series of methyl 1H-1,2,4-triazole-3-carboxylates 7 were prepared by our reported literature procedures [26] and hydrolyzed in presence of DBU/MeOH at room temperature for 6 h to afford the corresponding 1H-1,2,4-triazole-3-carboxylic acids 8 in > 90% high yields [28]. Crude compounds 8 were extracted with  $CH_2Cl_2$  and washed with 10%  $HCl_{(aq)}$  without purification technique for further investigations.

To evaluate the activated forms of carboxylic acid in optimization study [32,33], compound **8a** (1.0 equiv) and 2-hydroxyacetamide **10** (1.1 equiv) was reacted in the presence of activating reagents (2.0 equiv), such as acid halides (Oxalyl chloride (COCl)<sub>2</sub> and thionyl chloride (SOCl<sub>2</sub>)) [34], activated amides (1,1'-carbonyldiimidazole, CDI) [35], activeesters (car bodiimides: *N*-(3-dimethylaminopropyl)-*N*-ethylcarbodiimide hydrochl oride (EDC) [36], and dicyclohexylcarbodiimide (DCC)) [37], and re



Fig. 1. The structure of Sitagliptin 1.

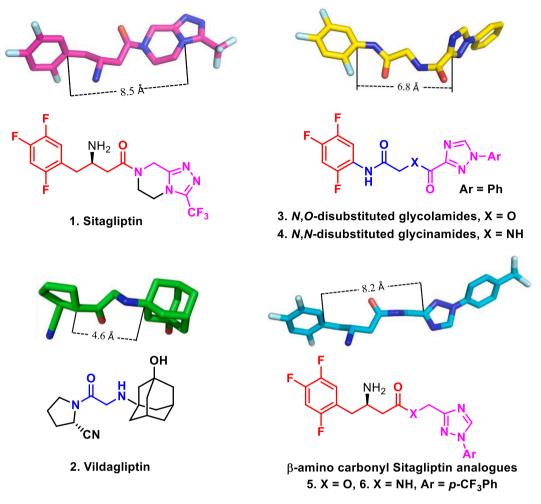
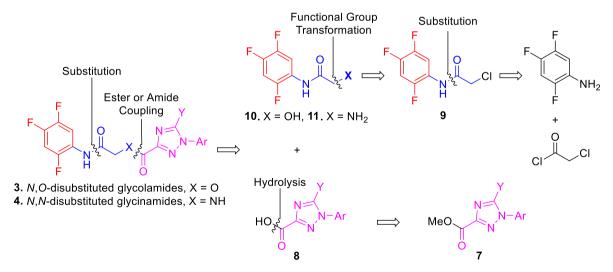


Fig. 2. The new designed structures of *N*,*O*-disubstituted glycolamides 3, *N*,*N*-disubstituted glycinamides 4, and  $\beta$ -amino carbonyl 1,2,4-triazoles 5 and 6 in comparison with Sitagliptin 1 and Vildagliptin 2.



Scheme 1. The retrosynthesis route of N,O-disubstituted glycolamide 1,2,4-triazoles 3 and N,N-disubstituted glycinamide 1,2,4-triazoles 4.

active phosphate agents (benzotriazol-1-yl-oxytripyrrolidinophosphonium hexafluorophosphate (PyBOP) [38], and *N*,*N*-bis[2-*oxo*-3-*oxazolidinyl*] phosphorodiamide chloride (BOP-Cl)) [39] in anhydrous THF or CH<sub>2</sub>Cl<sub>2</sub> solution under basic condition at 0 °C to room temperature for 6 h [33]. However, most of these reactions afforded *N*,*O*-disubstituted glycolamides product **3a** in moderate yields (<72%). Remarkably, significant and

successful reaction was allowed to stir with BOP-Cl activating reagent (2.0 equiv) and 2.0 equiv of NEt<sub>3</sub>, and the yield of product **3a** was significantly increased to 85% good yield. Since, *N*,*N*-bis[2-oxo-3-oxazolidinyl]phosphorodiamide chloride (BOP-Cl) could be served as the best convenient and efficient activating carboxyl groups reagent that has been employed in the further esterification and amidation transformation to prepare *N*,*O*-

The inhibitory activity results against DPP-4 of 1,3-disubstituted 1,2,4-triazoles

7a–g and 1,3,5-trisubstituted 1,2,4-triazoles 7h–q	MeO N= N-Ar
7a–g and 1,3,5-trisubstituted 1,2,4-triazoles 7h–q	. MeO N= N-Ar

		7a-q		
Compounds No.	Y	Ar	DPP-4% Inh@1mM <sup>a</sup>	
7a	Н	Ph	33.9	
7b	Н	p-Me-Ph	37.0	
7c	Н	p-Cl-Ph	34.6	
7d	Н	p-CF <sub>3</sub> -Ph	37.8	
7e	Н	m-CF3-Ph	40.1	
7f	Н	o-CF3-Ph	32.3	
7g	Н	<i>p</i> -F-Ph	10.1	
7h	Me	p-Cl-Ph	19.2	
7i	Me	<i>p</i> -Br-Ph	1.01	
7j	Et	p-Cl-Ph	4.13	
7k	<i>i</i> -Pr	p-Cl-Ph	24.4	
71	Furanyl	p-Cl-Ph	43.7	
7m	Cyclopentyl	p-Cl-Ph	85.1	
7n	Pyrrolyl	m-CF3-Ph	94.5	
70	<i>n</i> -Bu	p-Cl-Ph	100	
7p	Ph	Ph	100	
7q	Ph	p-Me-Ph	100	

<sup>a</sup> Concentration of each compound is 1.0 mmol  $L^{-1}$  in DMSO.

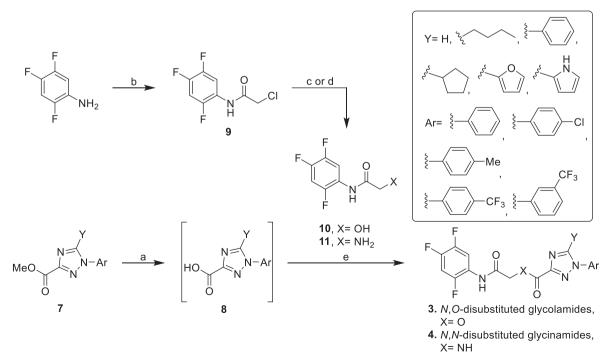
disubstituted glycolamides 3 and N,N-disubstituted glycinamides 4.

Following the above activated carboxylic acid optimization study, the 1,2,4-triazole-3-carboxylates **7a–e** and **7l–q** were preliminarily converted to the corresponding 1,2,4-triazole-3-carboxylic acids **8a–e** and **8l–q**. Without further purification, compounds **8** were directly reacted with 2-hydroxyacetamide **10** or 2-aminoacetamide **11** and 2.0 equiv of BOP-Cl in presence of 2.0 equiv of NEt<sub>3</sub> to give the corresponding glycolamides **3a–e** and **3m** and glycinamides products **4a–e** and **4l–q** in moderate to good yields (71–88% for **3** and 72–92% for **4**, Table 2). In general, 2-aminoacetamide **11** possessed better substitution reactivity with the activated acid **8** than 2-hydroxyacetamide **10**. All structures of *N*,*O*-disubstituted glycolamides **3** and *N*,*N*-disubstituted glycinamides **4** were fully characterized by spectroscopic methods, for example, compound **4m** presents one doublet peak at  $\delta$  4.39 ppm of  $\alpha$ -carbon methylene( $-CH_2-$ ) in <sup>1</sup>H NMR and characteristic absorption at  $\delta$  43.9 ppm for methylene carbon  $-^{13}CH_2-$ , at  $\delta$  167.3 ppm for glycinamide carbon N–( $^{13}C=O$ )– $CH_2$ , and at  $\delta$  161.9 ppm for amide carbon N–( $^{13}C=O$ ) in the <sup>13</sup>C NMR spectrum. Glycinamide **4m** was also further characterized by X-rays crystallographic analysis (as the single-crystal X-ray diffraction study, ORTEP) and shown in Fig. 3.

#### 2.2.2. Synthesis of $\beta$ -amino carbonyl 1,2,4-triazoles 5 and 6

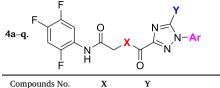
The enantiomerically β-amino carbonyl linker intensively plays an important role in examining the designed structure. Therefore, this result drove us to synthesize β-amino carbonyl Sitagliptin analogues containing 1,2,4-triazole moiety 5 and 6 (Scheme 3 and 4). Based on the preliminary DPP-4 inhibition data of substituted 1,2,4-triazole-3-carboxylates in Table 1, compounds 7d (DPP-4 Inh@1mM 37.8%) and 7m-q with good inhibition rates (DPP-4 Inh@1mM ≥85%) were selected as promising targets. Initially, we chose 1,2,4-triazole-3-carboxvlate 7d (1.0 equiv) as the model substrates to carry out the reduction reaction with NaBH<sub>4</sub> (1.5 equiv) in THF/MeOH at room temperature for 5 h. The desired 1H-1,2,4-triazol-3-ylmethanol product 12d was obtained in 91% yield (Scheme 3) [40]. The subsequent chlorination was performed using thionylchloride (SOCl<sub>2</sub>, 1.5 equiv) to yield the chloried intermediate [41]. Without further purification, the intermediate 3chloromethyl-1,2,4-triazole was treated with NH4OH(aa) in EtOH solution and smoothly converted to the amination (1H-1,2,4-triazol-3-yl) methanamine product 13d in 73% yield for two steps (Scheme 3) [42].

Finally,  $\beta$ -amino carbonyl targets **5d** and **6d** were individually synthesized from Boc-(*R*)-3-amino-4-(2,4,5-trifluoro-phenyl)-butyric acid **14** with (1*H*-1,2,4-triazol-3-yl)methanol **12d** or (1*H*-1,2,4-triazol-3-yl) methanamine **13d** by employing *N*,*N*-bis[2-oxo-3-oxazolidinyl]phosphorodiamide chloride (BOP-Cl, 2.0 equiv) as activating agent in presence of 2.0 equiv of NEt<sub>3</sub> in CH<sub>2</sub>Cl<sub>2</sub> solution to afford the Boc-protected intermediates **15d** and **16d** (Scheme 4) [39]. In this key substitution



Scheme 2. Synthesis of *N*,*O*-disubstituted glycolamides 3 and *N*,*N*-disubstituted glycinamides 4. Reagents and conditions: (a) DBU, MeOH, r.t., 6 h, > 90%; (b) 2-chloroacetyl chloride, toluene, reflux, 3 h, 92%; (c) For 10: cesium formate, EtOH, reflux, 6 h, 86%; (d) For 11: NH<sub>4</sub>OH, EtOH, 40 °C, 4 h, 88%; (e) BOP-Cl, NEt<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 4 h, 71–92%.

The results of N,O-disubstituted glycolamide 1,2,4-triazoles **3a**–**q** and N,N-disubstituted glycinamide 1,2,4-triazoles



Compounds No.	х	Y	Ar	Yields (%) <sup>a</sup>	
3a	0	Н	Ph	85	
4a	NH			87	
3b	0	Н	p-Me-Ph	87	
4b	NH			92	
3c	0	Н	p-Cl-Ph	88	
4c	NH			89	
3d	0	Н	p-CF3-Ph	83	
4d	NH		-	84	
3e	0	Н	m-CF3-Ph	71	
4e	NH			72	
41	NH	Furanyl	p-Cl-Ph	83	
3m	0	Cyclopentyl	p-Cl-Ph	73	
4m	NH		-	74	
4n	NH	Pyrrolyl	m-CF3-Ph	73	
4o	NH	n-Bu	p-Cl-Ph	82	
4p	NH	Ph	Ph	82	
4q	NH	Ph	p-Me-Ph	84	

<sup>a</sup> The calculated isolation yield was based on the amount of starting materials 2-hydroxyacetamide **10** or 2-aminoacetamide **11**.

reaction, we found that the amino group of compound **13d** was more nucleophilic reactivity than the hydroxyl group of **12d**. Without further purification, the one-pot synthesis procedure was proceeded to prepare  $\beta$ -amino carbonyl target compounds **5d** and **6d**. The deprotection of Bocprotected group of intermediates **15d** and **16d** was carried out in TFA/

CH<sub>2</sub>Cl<sub>2</sub> acidic solution to obtain β-amino ester product **5d** and β-amino amide product **6d** in 76% and 79% yields for two steps, respectively (Scheme 4 and Table 3) [43]. On the other hand, Boc-protected β-amino carbonyl product **15d** was tried to isolate for further identification of structural skeleton (Scheme 4). Compound **15d** presents two singlet peaks at  $\delta$  1.33 ppm for the *tert*-Boc group and  $\delta$  8.62 ppm for the 1,2,4triazolic ring (N=CH–N=N) and one doublet peak  $\delta$  5.32 ppm of the ester (O–CH<sub>2</sub>—) in <sup>1</sup>H NMR. In <sup>13</sup>C NMR spectrum, compound **15d** possessed characteristic absorptions at  $\delta$  170.8 ppm for <sup>13</sup>C=O, at  $\delta$  59.2 ppm for O–<sup>13</sup>CH<sub>2</sub>—, and at  $\delta$  28.2 and 79.5 ppm for *tert*-Boc group O–<sup>13</sup>C(<sup>13</sup>CH<sub>3</sub>)<sub>3</sub>.

Employing the above reliable procedure for synthesis of **12d** and **13d**, we converted compounds **7m**–**q** to 1*H*-1,2,4-triazol-3-ylmethanols **12m**–**q** in 92–96% yields and 1*H*-1,2,4-triazol-3-ylmethanamines **13m**–**q** in 76–84% yields (Scheme 3) [40–42]. Subsequently, compound **14** was directly reacted with 1*H*-1,2,4-triazol-3-ylmethanols **12m**–**q** or 1*H*-1,2,4-triazol-3-ylmethanamines **13m**–**q** to carried out the substitution reaction for generation of intermediates **15m**–**q** and **16m**–**q** [39]. Without further purification, the deprotection of Bocprotected group of intermediates **15m**–**q** and **16m**–**q** was carried out in TFA/CH<sub>2</sub>Cl<sub>2</sub> acidic solution [43]. The β-amino ester products **5m**–**q** were provided from 74% to 84% yields and β-amino amide products **6m**–**q** were obtained from 74% to 88% yields in two steps, respectively (Table 3 and Scheme 4).

#### 2.3. Biological activity

2.3.1. Pharmacological evaluation in vitro DPP-4 inhibition studies and SAR analysis of N,O-disubstituted glycolamides **3**, N,N-disubstituted glycinamides **4**, and  $\beta$ -amino carbonyl 1,2,4-triazoles **5** and **6** 

Screening of glycolamides **3a**–**e** and glycinamides **4a**–**e** with 1,3disubstituted 1,2,4-triazole moiety were evaluated in vitro against a panel of standard DPP-4. The inhibitory activity was executed using the

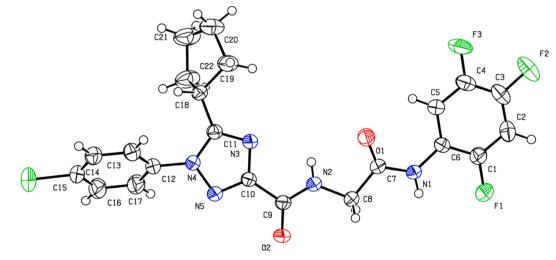
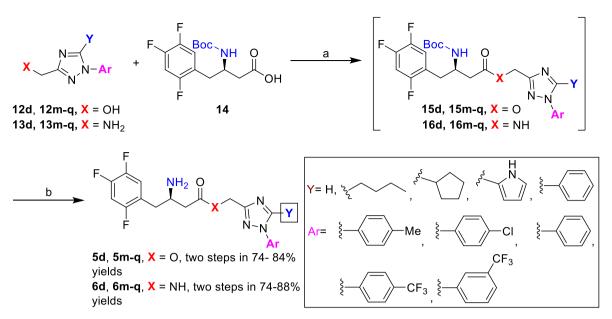


Fig. 3. The ORTEP diagram of 1-(4-chlorophenyl)-5-cyclopentyl-*N*-(2-oxo-2((2,4,5-trifluorophenyl)amino)ethyl)-1*H*-1,2,4-triazole-3-carboxamide 4m (CCDC No. 1973637).

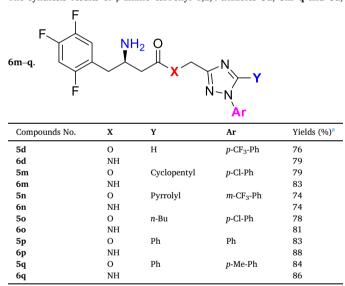


Scheme 3. Synthesis of (1*H*-1,2,4-triazol-3-yl)methanols 12d, 12m–q and (1*H*-1,2,4-triazol-3-yl)methanamine 13d, 13m–q. Reagents and conditions: (a) NaBH<sub>4</sub>, THF, MeOH, r.t., 4 h, 91–96%; (b) SOCl<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 2 h; (c) NH<sub>4</sub>OH, EtOH, 30 °C, 6 h, 73–84%.



Scheme 4. Synthesis of β-amino carbonyl target compounds containing 1,2,4-triazole analogues 5d, 5m–q and 6d, 6m–q. Reagents and conditions: (a) BOP-Cl, NEt<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 4 h; (b) CF<sub>3</sub>COOH, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 3 h, 74–88%.

**Table 3** The synthesis results of  $\beta$ -amino carbonyl 1,2,4-triazoles 5d, 5m–q and 6d,



<sup>a</sup> The calculated isolation yield was based on the amount of reactant Boc-(*R*)-3-amino-4-(2,4,5-trifluoro-phenyl)-butyric acid **14**.

disc diffusion method and the measurement of the zone of growth inhibition for a concentration 1.0 mmol/L (DMSO) for each glycolamides **3a–e** and glycinamides products **4a–e**. Sitagliptin **1** was used as comparison model for the inhibitory activity study (100%, Table 4). Noteworthily, compounds **3d** and **4d** with *p*-CF<sub>3</sub>Ph substituents on *N*-1 position of triazole ring possessed better inhibitory activity against DPP-4 (57.2% and 58.2%, Table 4). Moreover, the inhibitory activity tendency of DPP 4 was observed that most of glycinamides **4a–e** presented better inhibitory activity than glycolamides **3a–e**. Based on this result, glycinamides **4l–q** with 1,3,5-trisubstituted 1,2,4-triazole moiety bearing *n*-butyl, cyclopentyl, furanyl, and pyrrolyl on *C*-5 position and *p*-Cl-Ph, *m*-CF<sub>3</sub>-Ph, Phenyl, and *p*-Me-Ph on *N*-1 position of triazole ring were significantly synthesized for further investigation. However, the poor inhibitory data was obtained from glycinamides **4l–q** ( $\leq$ 48.8%, Table 4). The unsatisfactory inhibitory results led to the conclusion that glycolamides **3** and glycinamides **4** were unfavorable linkers related to the  $\beta$ -amino carbonyl of Sitagliptin **1**. To sum up, we observed that the enantiomerically  $\beta$ -amino carbonyl linker seems to intensively play an important role. Therefore, the results drove us to the discovery of  $\beta$ -amino carbonyl 1,2,4-triazoles **5** and **6**.

For further optimization, introducing  $\beta$ -amino carbonyl as a linker became a primary goal. Glycolamide 3d and glycinamide 4d possessed better inhibitory activity in Table 4. Thus,  $\beta$ -amino carbonyl 1,3-disubstituteds 1,2,4-triazoles 5d and 6d were selected to optimize linkage property for evaluating DPP-4 inhibitory capacity. Fortunately, β-amino amide **6d** showed superior inhibitory activity ( $IC_{50} = 34.4 \text{ nM}$ , Table 5). On the other hand, no significant improvement was observed for  $\beta$ -amino ester **5d** (IC<sub>50</sub> = 775 nM, Table 5). For further demonstration of bioactivity potency, two series of compounds including β-amino esters **5m**–**q** and  $\beta$ -amino amides **6m**–**q** were synthesized in parallel to modify the linker and 1,3,5-trisubstituted 1,2,4-triazoles (Table 5). Most of compounds 5m-q and 6m-q showed the moderate inhibitory activity and their  $IC_{50}$  values were ranging from 49.9 nM to 497 nM (Table 5). However, these results could not provide us conclusive potential inhibitory tendency order between  $\beta$ -amino esters **5m**–**q** and  $\beta$ -amino amides 6m-q (Table 5). To sum up, compounds 5n and 6p also exhibited the significant potent anti-DPP-4 activity with IC<sub>50</sub> value of 49.9 nM and 50.4 nM, respectively (Table 5).

## 2.3.2. Selectivity DPP-4 over DPP-7, DPP-8, and DPP-9 of $\beta$ -amino ester 5n and $\beta$ -amino amides 6d and 6p

The DPP-4 activity and/or structural homologue (DASH) family, including quiescent cell proline dipeptidase (QPP, also known as DPP-2 or DPP-7), DPP-8, DPP-9, and FAP [45]. As reported in previous study, the DPP-4 inhibitors containing  $\beta$ -amino acid fragment in the structure resulted in the QPP off-target activity [12,46]. In addition, selective inhibition of DPP-8/DPP-9 was associated with toxicity, such as alopecia, thrombocytopenia, anemia, and animal mortality [47]. Therefore, selectivity against QPP, DPP-8, DPP-9 was important for the safety of DPP-4 inhibitors. Based on the potency of DPP-4 inhibition, we selected  $\beta$ -amino ester **5n** and  $\beta$ -amino amides **6d** and **6p** (IC<sub>50</sub> < 51 nM) to evaluate selectivity across enzymes QPP, DPP-8, and DPP-9. As shown in Table 6, both  $\beta$ -amino carbonyl 1,3,5-trisubstituted 1,2,4-triazoles **5n** and **6p** showed excellent selectivity over QPP, DPP-8, and DPP-9 (>1950-fold), however, the selectivity of 1,5-disubstituted 1,2,4-triazole **6d** was lower (135-, 31-, and 105-fold selectivity over QPP, DPP-8,

The DPP-4 inhibitory activity results of *N*,*O*-disubstituted glycolamide 1,2,4-triazoles **3**, *N*,*N*-disubstituted glycinamide 1,2,4-triazoles **4**, and β-amino carbonyls **5** and

	X O N N N N	Ar		
F Compounds No.	X	Y	Ar	DPP-4% Inh@1mM <sup>a</sup>
3a 4a	O NH	Н	₹-	43.4 37.6
3b 4b	O NH	Н	ξ-√Me	28.4 34.1
3c 4c	O NH	Н	ξ-√_−Cι	39.4 47.2
3d 4d	O NH	Н	₹ <b>C</b> F <sub>3</sub>	57.2 58.2
3e 4e	O NH	н	CF <sub>3</sub>	37.0 42.5
41	NH	ers O	ş Ş CI	33.5
3m 4m	OH NH	rs -	ξ-√−Cι	39.3 48.8
4n	NH	<sup>₹</sup> <sup>5</sup> H N	CF3	34.0
40	NH	22	₹{CI	36.6
4p	NH	<b>ξ</b>		32.5
4q	NH		}Me	39.2
Sitagliptin 1				100

<sup>a</sup> Concentration of each used compound is 1.0 mmol  $L^{-1}$  (DMSO).

and DPP-9, respectively). This result indicated that an aryl group appeared to be essential for the 5-position of 1,2,4-triazole. Importantly, the selectivity over DPP8 of compounds **5n** and **6d** (>1984-fold) was better than Sitagliptin (1, 787-fold), making compounds **5n** and **6d** suitable for further study.

## 2.4. Docking study of $\beta$ -amino carbonyl target **5n**, **6d**, and **6p** into the active site of DPP-4

In order to gain an insight of the possible binding interactions of  $\beta$ -amino ester **5n** and  $\beta$ -amino amides **6d** and **6p** in the active site of DPP-4 enzyme, molecular modelling was performed. The X-ray structure of DPP-4 cocrystallized with Sitagliptin **1** (PDB ID 1X70, resolution 2.10 Å) was used to perform the docking study through docking program iGEMDOCK v2.1 [12,25]. Re-docking of the co-crystallized ligand Sitagliptin **1** into DPP-4 active site via iGEMDOCK was performed, and

the RMSD was calculated by using LigRMSD [48]. Sitagliptin 1 docked in the DPP-4 binding site almost at the same position (RMSD = 0.53 Å) as the co-crystallized data (PDB ID 1X70), which validated the iGEM-DOCK protocol. The DPP-4 enzyme was docked with compounds 5n, 6d, and **6p** in Sitagliptin **1** binding site. As depicted in Fig. 4, the docking model revealed that compounds 5n, 6d, and 6p were stabilized at DPP-4 active site similarly to Sitagliptin 1 through three hydrogen bond interactions with residues Tyr662, Glu205, and Glu206, and two  $\pi$ - $\pi$ stacking interaction with residues Tyr662 and Phe357. β-amino amide 6d with superior DPP-4 inhibition achieves lower docking scores of -150.4 kcal/mol (Table S2), because 4-(trifluoromethyl)phenyl group of compound 6d possesses additional van der Waals interaction with residue Arg358 in comparison with Sitagliptin 1 (Fig. 4A). On the other hand, it has been reported that neither DPP-8 nor DPP-9 has a residue corresponding to Phe357 of DPP-4 [49]. Based on the ducking result, we found that 1,3,5-trisubstituted 1,2,4-triazoles 5n (-26.4 kcal/mol) and

60

5p

6p

5q

6q

Sitagliptin 1

NH

0

0

NH

NF

$\begin{array}{c} F \\ 6. \\ F \\ F \\ \end{array} \\ \begin{array}{c} NH_2 \\ N \\ N \\ N \\ N \\ Ar \end{array}$						
Compounds No.	х	Y	Ar	DPP-4 IC <sub>50</sub> (nM) <sup>a</sup>		
5d 6d	O NH	Н	₹CF3	775 (nM) 34.4 (nM)		
5m 6m	O NH	rrs C	ξ-√_−CI	80.3 (nM) 131 (nM)		
5n 6n	O NH	rs <sup>2</sup> HN	₹	49.9 (nM) 99.8 (nM)		
50	0	5	, /=\	81.6 (nM)		

The DPP-4 inhibitory activity results of  $\beta\mbox{-amino}$  carbonyls  ${\bf 5}$  and  ${\bf F}$ 

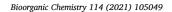
#### <sup>a</sup> $IC_{50}$ is the concentration (nM) needed to cause 50% inhibition human recombinant DPP-4 enzymatic activity. Values represent the mean of double determinations.

 $^{\rm b}$  The IC  $_{50}$  value of Sitagliptin was between 18 and 38 nM in reported literature [12,44].

**6p** (-23.8 kcal/mol) with additional aromatic ring on the triazole ring provided stronger  $\pi$ - $\pi$  stacking interaction with residue Phe357 than 1,5-disubstituted 1,2,4-triazole **6d** (-11.8 kcal/mol) and Sitagliptin **1** (-19.4 kcal/mol, Fig. 4B and Table S3), which might explain compounds **5n** and **6p** improved selectivity inhibition of DPP-4 over DPP-8 and DPP-9.

Selectivity of  $\beta$ -amino ester **5n**,  $\beta$ -amino amides **6d** and **6p**, Sitagliptin **1** against QPP, DPP-8, and DPP-9.

#### Table 6



#### 3. Conclusion

We have successfully synthesized and built-up four series of 1,2,4-triazole derivatives 3-6 containing N,O-disubstituted glycolamide, N,N'disubstituted glycinamides,  $\beta$ -amino ester, and  $\beta$ -amino amide as linkers through modifying Sitagliptin 1 and Vildagliptin 2. Different linkers grafted the same 2.4,5-triflorophenyl fragment into the architecture, and 1,2,4-triazole heterocyclic moiety was modulated for structural variation. The synthetic strategy for glycolamides or glycinamides involved functionalized transformation of  $\alpha$ -chloro N-arylacetamides (hydroxylation or amination) and esterification or amidation of 1,2,4triazole-3-carboxylic acid by activating with BOP-Cl reagent. We also developed one-pot synthesis procedure, including treating (1H-1,2,4triazol-3-yl)methanol or (1H-1,2,4-triazol-3-yl)methylamine with Boc-(R)-3-amino-4-(2,4,5-trifluoro-phenyl)-butyric acid and deprotection of *tert*-Butyloxycarbonyl (BOC) group, for the preparation of  $\beta$ -amino carbonyl 1,2,4-triazoles. For the preliminary screening in vitro, glycolamides 3 and glycinamides 4 showed poor dipeptidyl peptidase 4 inhibitory activity ( $\leq$ 58.2% DPP-4 Inh@1mM). Most of  $\beta$ -amino ester 5 and  $\beta$ -amino amide 1,2,4-triazoles 6 possessed the moderate inhibition of DPP 4 (IC<sub>50</sub> < 775 nM). Especially,  $\beta$ -amino carbonyl **5n**, **6d**, and **6p** had satisfactory DPP-4 inhibitory activity (50.4, 34.4, 49.9 nM, respectively). Docking study also revealed compounds 5n, 6d and 6p have favorable binding mode with DPP-4 enzyme. Selectivity evaluation indicated  $\beta$ -amino carbonyl **5n** and **6p** hold excellent selectivity over QPP, DPP-8, and DPP-9 and superior to reference Sitagliptin. Furthermore, compounds **5n** and **6p** provided stronger  $\pi$ - $\pi$  stacking interaction with residue Phe357 than 1,5-disubstituted 1,2,4-triazole 6d and Sitagliptin 1 based on the molecular docking results. In summary,  $\beta$ -amino carbonyl 5n and 6p exhibited good activity in vitro and excellent selectively and might be a promising lead for the treatment of diabetes mellitus.

#### 4. Experimental section

#### 4.1. Chemistry

119 (nM)

91.3 (nM)

50.4 (nM)

497 (nM)

138 (nM)

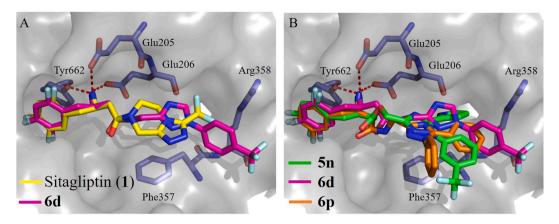
28 (18-38

 $nM)^{t}$ 

All chemicals were purchased with reagent grade. Both 1,3-disubstituted 1,2,4-triazoles **7a–g** [26a] and 1,3,5-trisubstituted 1,2,4-triazoles **7h–q** [26b] were prepared following our reported procedure [26]. All reactions were carried out under argon or nitrogen atmosphere and monitored by TLC. Flash column chromatography was carried out on

				F		N-N Ar	
Compounds No. X	Х	Y	Ar	IC <sub>50</sub> (nM) <sup>a</sup>			
				DPP-4	QPP	DPP-8	DPP-9
5n	0	rss H	CF <sub>3</sub>	49.9	>10 <sup>5</sup>	>10 <sup>5</sup>	>10 <sup>5</sup>
6d	NH	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	ş Ş CI	34.4	4630	1060	3610
6p	NH		ş	50.4	>10 <sup>5</sup>	>10 <sup>5</sup>	>10 <sup>5</sup>
Sitagliptin $1^{\mathrm{b}}$				28	>10 <sup>5</sup>	22030	>10 <sup>5</sup>

<sup>a</sup> IC<sub>50</sub> is the concentration (nM) needed to cause 50% inhibition human recombinant DPP-4 enzymatic activity. Values represent the mean of double determinations. <sup>b</sup> Sitagliptin was used as the reference drug in previous literature [12].



**Fig. 4.** Docked pose of the active site of DPP-4 enzyme with (A) Sitagliptin 1 (yellow) and β-amino amide **6d** (magenta) (B) β-amino ester **5n** (green), β-amino amides **6d** (magenta) and **6p** (orange). The red dashed lines indicate hydrogen-bonding interactions. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

silica gel (230–400 mesh). Analytical thin-layer chromatography (TLC) was performed on precoated plates (silica gel 60F-254) purchased from Merck Inc. Commercially available reagents were used without further purification unless otherwise noted. Dichloromethane, ethyl acetate, and hexane were purchased from Mallinckrodt Chemical Co. Dimethyl sulfone (DMSO<sub>2</sub>, purity = 100%) was purchased from Tokyo Chemical Industry Co. Proton NMR spectra were obtained on a Bruker (400 MHz or 500 MHz) spectrometer by use of CDCl<sub>3</sub> or DMSO-d<sub>6</sub> as solvent. Carbon-13 NMR spectra were obtained on a Bruker (100 MHz or 125 MHz) spectrometer by used of CDCl<sub>3</sub> or DMSO-d<sub>6</sub> as solvent. The following abbreviations are used for spin multiplicity: s, singlet; d, doublet; t, triplet; q, quartet; p, quintet/pentet; sept, septet; dd, doublet of doublets; dt, triplet of doublets; m, multiplet; J, coupling constant (Hz). The purities of the compounds **3–6** were determined by using the q<sup>1</sup>HNMR method (absolute q<sup>1</sup>HNMR with internal calibration) [50]. Most of purities were more than > 98%, for example, compound **6n** was 98.80% (Figure S91). The purity of compound 6n was calculated by following equation:

$$p(\%) = \frac{n_{ic}.int_t.MW_t.m_{ic}}{n_t.int_{ic}.MW_{ic}.m_s} p_{ic}$$

where MW is the molecular weight, *P* is the purity of internal calibrant,  $m_{\rm ic}$  is the amount of internal calibrant,  $m_{\rm s}$  is the amount of sample, int is integral, and *n* is the number of protons giving rise to a given NMR signal. The subscript ic and t represents the internal calibrant and target analyte, respectively. Infrared (IR) spectra were measured on a Bomem Michelson Series FT-IR spectrometer. The wavenumbers reported are referenced to the polystyrene absorption at 1601 cm<sup>-1</sup>. Flash column chromatography purification of compounds was carried out by gradient elution using hexanes in ethyl acetate (EA) unless otherwise stated. High-resolution mass spectra were obtained from a JEOL JMS-HX110 mass spectrometer.

## 4.1.1. Standard procedure for synthesis of N,O-disubstituted glycolamides 3a-e and 3m [26,28,39]

A series of methyl 1*H*-1,2,4-triazole-3-carboxylates **7a–e** and **7m** were prepared as the starting martials by following our previous published procedure [26]. 1*H*-1,2,4-triazole-3-carboxylates **7a–e** and **7m** (1.0 equiv, 5.5 mmol) was dissolved in MeOH (15 mL) and stirred at room temperature. 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU, 3.0 equiv) [28] was slowly added to the reaction mixture at room temperature. Then the mixture was stirred for 6 h. When the reaction was completed, the reaction mixture was added to water (15 mL) and extracted with ethyl acetate (15 mL × 3). The combined organic layers were washed sodium bicarbonate (15 mL × 3), dried over MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure to afford the

corresponding crude 1H-1,2,4-triazole-3-carboxylic acids  $\bf 8a-e$  and  $\bf 8m.$ 

A solution of crude 1*H*-1,2,4-triazole-3-carboxylic acids **8a–e** and **8m** (1.0 equiv from **7a–e** and **7m**) and 2-hydroxy-*N*-(2,4,5-tri-fluorophenyl)acetamide **10** (1.1 equiv) was treated with *N*,*N*-bis[2-*oxo*-3-*oxazolidinyl*]phosphorodiamide chloride (BOP-Cl, 2.0 equiv) in presence of NEt<sub>3</sub> (2.0 equiv) in CH<sub>2</sub>Cl<sub>2</sub> solution (20 mL) [39]. The reaction mixture was stirred at room temperature for 4 h. When the reaction was completed, the reaction mixture was washed with water (15 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (15 mL × 3). The combined organic layers were washed sodium bicarbonate (15 mL × 3), dried over MgSO<sub>4</sub> to give crude *N*,*O*-disubstituted glycolamides **3a–e** and **3m**. The crude products **3a–e** and **3m** were purified by column chromatography on silica gel to give the pure corresponding *N*,*O*-disubstituted glycolamides **3a–e** and **3m** in 71–88% yields.

4.1.1.1. 2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl 1-phenyl-1H-1,2,4-triazole-3- carboxylate (**3a**). Yield 85%, white solid, m.p. 184–186 °C, <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  5.08 (s, 2H, OCH<sub>2</sub>), 7.50 (t, J = 7.38 Hz, 1H, ArH), 7.62 (t, J = 7.86 Hz, 2H, ArH), 7.66–7.71 (m, 1H, ArH), 7.93 (d, J = 8.00 Hz, 2H, ArH), 8.03 (dt, J = 15.95, 6.15 Hz, 1H, ArH), 9.50 (s, 1H, ArH), 10.34 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, DMSO- $d_6$ )  $\delta$  63.12, 106.01 (dd, J = 25.63, 22.73 Hz), 111.60 (d, J = 24.08 Hz), 120.01 (2 × C), 122.37, 128.79, 129.93 (2 × C), 136.30, 144.27, 145.17 (d, J = 238.67 Hz), 147.55, 149.92, 154.11, 158.83, 165.75. IR (KBr): 3460, 3164, 1750, 1692, 1427, 1202, 883, 750 cm<sup>-1</sup>. EIMS *m/z*: 376 (M<sup>+</sup>, 5), 231 (25), 173 (20), 172 (100), 145 (22), 104 (11), 77 (13). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>17</sub>H<sub>11</sub>F<sub>3</sub>N<sub>4</sub>O<sub>3</sub>: 376.0783; Found 376.0785.

4.1.1.2. 2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl 1-(p-tolyl)-1H-1,2,4-triazole- 3-carboxylate (**3b**). Yield 87%, light yellow solid, m.p. 186–188 °C, <sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>)  $\delta$  2.39 (s, 3H, CH<sub>3</sub>), 5.06 (s, 2H, OCH<sub>2</sub>), 7.42 (d, J = 8.40 Hz, 2H, ArH), 7.68 (td, J = 10.72, 7.45 Hz, 1H, ArH), 7.80 (d, J = 8.40 Hz, 2H, ArH), 8.02 (dt, J = 15.96, 6.30 Hz, 1H, ArH), 9.44 (s, 1H, ArH), 10.34 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, DMSO-d<sub>6</sub>)  $\delta$  20.58, 63.12, 106.04 (t, J = 23.76 Hz), 111.65 (d, J = 23.52 Hz), 119.92 (2 × C), 120.19, 130.30 (2 × C), 134.08, 138.51, 144.06, 144.37, 146.04, 147.79, 153.97, 158.87, 165.78. IR (KBr): 3449, 3160, 2929, 1736, 1685, 1427, 1202, 883, 815 cm<sup>-1</sup>. EIMS *m/z*: 390 (M<sup>+</sup>, 6), 244 (22), 187 (20), 186 (100), 159 (30), 118 (15), 91 (12). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>18</sub>H<sub>14</sub>F<sub>3</sub>N<sub>4</sub>O<sub>3</sub>: 390.0940; Found 390.0945.

4.1.1.3. 2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl 1-(4-chlorophenyl)-1H-1,2,4- triazole-3-carboxylate (3c). Yield: 88%, light yellow solid; m. p. 182–183 °C, <sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>)  $\delta$  5.07 (s, 2H, OCH<sub>2</sub>), 7.65–7.67 (m, 1H, ArH), 7.69 (d, J = 8.91 Hz, 2H, ArH), 7.97 (d, J = 8.74 Hz, 2H, ArH), 8.02 (dt, J = 16.03, 6.19 Hz, 1H, ArH), 9.52 (s, 1H, ArH), 10.33 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, DMSO- $d_6$ )  $\delta$  63.14, 106.01 (dd, J = 25.91, 22.40 Hz), 111.59 (d, J = 23.85 Hz), 121.76 (2 × C), 122.34 (t, J = 11.36 Hz), 129.88 (2 × C), 130.05, 135.14, 144.47, 145.24 (dd, J = 240.59, 12.65 Hz), 147.75 (d, J = 8.48 Hz), 149.70 (d, J = 9.61 Hz), 154.17, 158.72, 165.69. IR (KBr): 3456, 3160, 2919, 2851, 1746, 1682, 1427, 1199, 883, 828 cm<sup>-1</sup>. EIMS m/z: 410 (M<sup>+</sup>, 6), 264 (26), 208 (35), 207 (10), 206 (100), 179 (20), 147 (11), 138 (11). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>17</sub>H<sub>10</sub>ClF<sub>3</sub>N<sub>4</sub>O<sub>3</sub>: 410.0394; Found 410.0390.

4.1.1.4. 2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl 1-(4-(trifluoromethyl) phenyl)- 1H- 1,2,4-triazole-3-carboxylate (**3d**). Yield 83%, yellow solid, m. p. 112–113 °C, <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  5.09 (s, 2H, OCH<sub>2</sub>), 7.69 (td, J = 10.65, 7.51 Hz, 1H, ArH), 8.00–8.07 (m, 3H, ArH), 8.19 (d, J = 8.46 Hz, 2H, ArH), 9.65 (s, 1H, ArH), 10.36 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, DMSO- $d_6$ )  $\delta$  63.23, 106.07 (dd, J = 25.58, 22.85 Hz), 111.59 (d, J = 24.22 Hz), 120.53(2 × C), 122.36, 123.82 (d, J = 272.31 Hz), 127.26, 127.30, 128.75 (q, J = 31.49 Hz), 139.20, 144.98, 145.28 (dd, J = 241.78, 12.67 Hz), 147.55, 149.94 (d, J = 7.37 Hz), 154.44, 158.70, 165.71. IR (KBr): 3439, 3221, 2953, 1746, 1678, 1430, 1321, 1127, 1066, 849 cm<sup>-1</sup>. EIMS *m/z*: 444 (M<sup>+</sup>, 2), 271 (15), 256 (13), 241 (13), 240 (100), 213 (26), 172 (16), 159 (22), 147 (14), 145 (18). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>18</sub>H<sub>10</sub>F<sub>6</sub>N<sub>4</sub>O<sub>3</sub>:444.0657; Found 444.0649.

4.1.1.5. 2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl 1-(3-(trifluoromethyl) phenyl)- 1H-1,2,4-triazole-3-carboxylate (**3e**). Yield: 71%, white solid, m. p. 163–164 °C, <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  5.08 (s, 2H, OCH<sub>2</sub>), 7.67 (td, J = 10.69, 7.44 Hz, 1H, ArH), 7.85–7.90 (m, 2H, ArH), 8.03 (dt, J = 15.98, 6.19 Hz, 1H, ArH), 8.25–8.27 (m, 1H, ArH), 8.31 (s, 1H, ArH), 9.65 (s, 1H, ArH), 10.35 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, DMSO- $d_6$ )  $\delta$  63.22, 106.05 (dd, J = 25.86, 22.09 Hz), 111.65 (d, J = 22.87 Hz), 116.76, 116.79, 123.59 (d, J = 272.83 Hz), 124.04, 125.34, 130.55 (q, J = 32.81 Hz), 131.42, 136.87, 144.91, 145.28 (dd, J = 241.05, 12.57 Hz), 147.85, 149.72, 154.31, 158.72, 165.71. IR (KBr): 3470, 3160, 1746, 1692, 1508, 1427, 1202, 1131, 883 cm<sup>-1</sup>. EIMS m/z: 444 (M<sup>+</sup>, 4), 298 (18), 241 (14), 240 (100), 213 (14), 172 (10), 147 (11), 145 (14). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>18</sub>H<sub>10</sub>F<sub>6</sub>N<sub>4</sub>O<sub>3</sub>: 444.0657; Found 444.0665.

4.1.1.6. 2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl 1-(4-chlorophenyl)-5- cyclopentyl-1H-1,2,4-triazole-3-carboxylate (3m). Yield 73%, white solid, m.p. 68–69 °C, <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.62–1.68 (m, 2H, CH<sub>2</sub>), 1.85–1.94 (m, 2H, CH<sub>2</sub>), 1.96–2.01 (m, 4H, CH<sub>2</sub>), 3.13 (quint, J = 8.28 Hz, 1H, CH), 5.00 (s, 2H, OCH<sub>2</sub>), 6.97 (td, J = 9.97, 6.99 Hz, 1H, ArH), 7.40 (d, J = 8.71 Hz, 2H, ArH), 7.53 (d, J = 8.72 Hz, 2H, ArH), 8.31 (dt, J = 15.73, 5.92 Hz, 1H, ArH), 8.46 (br, 1H, NH).<sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>) δ 25.77 (2 × C), 33.00 (2 × C), 36.26, 63.38, 104.93 (t, J = 23.47 Hz), 111.56 (d, J = 25.00 Hz), 121.64, 126.73 (2  $\times$  C), 129.88 (2 × C), 135.10, 136.05, 142.95, 145.05, 148.77, 152.91, 158.12, 162.87, 164.69. IR (KBr): 3303, 3072, 2960, 2875, 1756, 1715, 1549, 1501, 1426, 1192, 1097, 876, 839 cm<sup>-1</sup>. EIMS m/z: 480 (M<sup>+</sup> + 2, 5), 479 (M<sup>+</sup> + 1, 4), 478 (M<sup>+</sup>, 16), 305 (16), 277 (18), 276 (37), 275 (40), 274 (100), 249 (13), 248 (30), 247 (39), 246 (70), 232 (14), 206 (17), 160 (22), 153 (16), 151 (44), 147 (46), 146 (16), 138 (13), 129 (22), 127 (75), 125 (33), 119 (19), 113 (11), 111 (26), 99 (22). HRMS (EI) *m*/*z*: [M]<sup>+</sup> Calcd for C<sub>22</sub>H<sub>18</sub>ClF<sub>3</sub>N<sub>4</sub>O<sub>3</sub>: 478.1020; Found 478.1016.

## 4.1.2. Standard procedure for synthesis of N,N-disubstituted glycinamides 4a–e and 4l–q [26,28,39]

A series of methyl 1*H*-1,2,4-triazole-3-carboxylates **7a–e** and **7l–q** were prepared as the starting martials by following our previous published procedure [26]. 1*H*-1,2,4-Triazole-3-carboxylates **7a–e** and **7l–q** (1.0 equiv, 5.5 mmol) was dissolved in MeOH (15 mL) and stirred at room temperature. 1,8-diazabicyclo(5.4.0)undec-7-ene (DBU, 3.0 equiv) [28] was slowly added to the reaction mixture at room temperature. Then the mixture was stirred for 6 h. When the reaction was completed, the reaction mixture was added to water (15 mL) and extracted with ethyl acetate (15 mL  $\times$  3). The combined organic layers were washed sodium bicarbonate (15 mL  $\times$  3), dried over MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure to afford the corresponding crude 1*H*-1,2,4-triazole-3-carboxylic acids **8a–e** and **8–q**.

A solution of crude 1*H*-1,2,4-triazole-3-carboxylic acids **8a–e** and **8l–q** (1.0 equiv, from **7a–e** and **7l–q**) and 2-amino-*N*-(2,4,5-tri-fluorophenyl)acetamide **11** (1.1 equiv) was treated with *N*,*N*-bis[2-*oxo*-3-*oxazolidinyl*]phosphorodiamide chloride (BOP-Cl, 2.0 equiv) in presence of NEt<sub>3</sub> (2.0 equiv) in CH<sub>2</sub>Cl<sub>2</sub> solution (20 mL) [39]. The reaction mixture was stirred at room temperature for 4 h. When the reaction was completed, the reaction mixture was washed with water (15 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (15 mL × 3). The combined organic layers were washed sodium bicarbonate (15 mL × 3), dried over MgSO<sub>4</sub> to give crude *N*,*N*-disubstituted glycinamides **4a–e** and **4l–q**. The crude products **4a–e** and **4l–q** were purified by column chromatography on silica gel to give the pure corresponding *N*,*N*-disubstituted glycinamides **4a–e** and **4l–q** in 72–92% yields.

4.1.2.1. 2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl-1-phenyl-1H-1,2,4-triazole-3- carboxylate (4a). Yield 87%, white solid, m.p. 245–246 °C, <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  4.17 (d, J = 3.34 Hz, 2H, —NHCH<sub>2</sub>—), 7.48 (t, J = 7.44 Hz, 1H, ArH), 7.61 (t, J = 7.97 Hz, 2H, ArH), 7.64–7.68 (m, 1H, ArH), 7.92 (d, J = 7.53 Hz, 2H, ArH), 8.02 (dt, J = 16.03, 6.25 Hz, 1H, ArH), 8.85 (t, J = 5.44 Hz, 1H, NH), 9.43 (s, 1H, ArH), 10.14 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, DMSO- $d_6$ )  $\delta$  42.60, 105.91 (dd, J = 26.21, 22.28 Hz), 111.58 (d, J = 22.53 Hz), 119.82 (2 × C), 122.88 (t, J = 11.12 Hz), 128.47, 129.87 (2 × C), 136.49, 143.48, 144.19, 146.09, 147.68, 157.08, 158.83, 168.04. IR (KBr): 3401, 3290, 2922, 1671, 1542, 1505, 1433, 1202, 873 cm<sup>-1</sup>. EIMS m/z: 375 (M<sup>+</sup>, 2), 230 (14), 229 (58), 202 (16), 201 (34), 173 (15), 172 (100), 147 (36), 145 (21), 104 (12), 77 (16). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>17</sub>H<sub>12</sub>F<sub>3</sub>N<sub>5</sub>O<sub>2</sub>: 375.0943; Found 375.0951.

4.1.2.2. N-(2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl)-1-(p-tolyl)-1H-1,2,4- triazole-3-carboxamide (**4b**). Yield 92%, light yellow solid, m.p. 251–252 °C, <sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>)  $\delta$  2.38 (s, 3H, *CH*<sub>3</sub>), 4.17 (d, J = 5.95 Hz, 2H, OCH<sub>2</sub>), 7.41 (d, J = 8.30 Hz, 2H, ArH), 7.64 (td, J = 15.98, 7.45 Hz, 1H, ArH), 7.80 (d, J = 8.45 Hz, 2H, ArH), 8.02 (dt, J = 16.01, 6.23 Hz, 1H, ArH), 8.19 (t, J = 5.90 Hz, 1H, NH), 9.37 (s, 1H, ArH), 10.11 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, DMSO-d<sub>6</sub>),  $\delta$  20.54, 42.58, 105.91 (t, J = 24.20 Hz), 111.56 (d, J = 24.48 Hz), 119.71 (2 × C), 122.82, 130.20 (2 × C), 134.22, 138.10, 143.23, 145.20 (dd, J = 237.34, 12.60 Hz), 147.73, 149.60, 156.92, 158.87, 168.05. IR (KBr): 3395, 3286, 3113, 2919, 2854, 1671, 1546, 1513, 1430, 1202, 866 cm<sup>-1</sup>. EIMS *m*/*z*: 389 (M<sup>+</sup>, 2), 243 (51), 187 (13), 186 (100), 159 (24), 118 (14), 91 (14). HRMS (EI) *m*/*z*: [M]<sup>+</sup> Calcd for C<sub>18</sub>H<sub>14</sub>F<sub>3</sub>N<sub>5</sub>O<sub>2</sub>: 389.1110; Found 389.1103.

4.1.2.3. 1-(4-Chlorophenyl)-N-(2-oxo-2-((2,4,5-trifluorophenyl)amino) ethyl)-1H- 1,2,4-triazole-3-carboxamide (**4**c). Yield 89%, white solid, m. p. 245–246 °C, <sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>)  $\delta$  4.17 (d, J = 5.99 Hz, 2H, NHCH<sub>2</sub>), 7.48 (t, J = 7.44 Hz, 1H, ArH), 7.63–7.67 (m, 1H, ArH), 7.69 (dd, J = 6.82, 2.12 Hz, ArH), 7.96 (dd, J = 6.82, 2.12 Hz, ArH), 8.02 (dt, J = 16.02, 6.23 Hz, 1H, ArH), 8.86 (t, J = 5.97 Hz, 1H, NH), 9.45 (s, 1H, ArH), 10.11 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, DMSO-d<sub>6</sub>)  $\delta$  42.58, 105.91 (dd, J = 25.86, 22.23 Hz), 111.54 (d, J = 24.94 Hz), 121.53 (2 × C), 122.79, 128.47, 129.82 (2 × C), 135.32, 143.70, 145.19 (dd, J = 240.40, 15.64 Hz), 147.72, 149.57, 157.14, 158.71, 168.00. IR (KBr): 3334, 3228, 3116, 2919, 1712, 1678, 1525, 1495, 825 cm<sup>-1</sup>. EIMS *m*/*z*: 412 (M<sup>+</sup> + 2, 2), 411 (M<sup>+</sup> + 1, 3), (M<sup>+</sup>, 4), 238 (13), 237 (22), 236 (26), 235 (38), 208 (41), 207 (20), 206 (100), 180 (10), 179 (20), 151 (11), 149 (12), 148 (47), 147 (68), 138 (21), 119 (12), 111 (18). HRMS (EI) *m*/*z*: [M]<sup>+</sup> Calcd for C<sub>17</sub>H<sub>11</sub>ClF<sub>3</sub>N<sub>5</sub>O<sub>2</sub>: 409.0553; Found 409.0545.

#### 4.1.2.4. N-(2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl)-1-(4-(tri-

fluoromethyl) phenyl)-1H-1,2,4-triazole-3-carboxamide (**4d**). Yield 84%, light yellow solid, m.p. 177–178 °C, <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  4.18 (d, J = 5.50 Hz, 2H, NHCH<sub>2</sub>), 7.65 (q, J = 9.61 Hz, 1H, ArH), 8.01 (d, J = 7.65 Hz, 2H, ArH), 8.03–8.04 (m, 1H, ArH), 8.17 (d, J = 7.90 Hz, 2H, ArH), 8.92 (t, J = 5.30 Hz, 1H, NH), 9.57 (s, 1H, ArH), 10.12 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, DMSO- $d_6$ )  $\delta$  42.63, 105.94 (dd, J = 25.98, 22.30 Hz), 111.62 (d, J = 23.94 Hz), 120.30 (2 × C), 122.78, 124.94, 127.21, 127.23, 128.49 (q, J = 32.50 Hz), 139.38, 144.19, 145.18 (dt, J = 243.33, 13.50 Hz), 147.78, 149.64, 157.42, 158.69, 168.01. IR (KBr): 3426, 3293, 3099, 2929, 1691, 1671, 1512, 1437, 1321, 1121, 1063, 842. cm<sup>-1</sup>. EIMS *m/z*: 443 (M<sup>+</sup>, 2), 297 (16), 269 (27), 241 (10), 240 (100), 213 (12), 172 (16), 147 (78), 145 (21). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>18</sub>H<sub>11</sub>F<sub>6</sub>N<sub>5</sub>O<sub>2</sub>: 443.0817; Found 443.0812.

#### 4.1.2.5. N-(2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl)-1-(3-(tri-

fluoromethyl) phenyl)-1H-1,2,4-triazole-3-carboxamide (**4e**). Yield 72%, white solid, m.p. 288–289 °C, <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$  4.18 (d, J = 6.05 Hz, 2H, NHC $H_2$ ), 7.66 (td, J = 10.71, 7.50 Hz, 1H, ArH), 7.86–7.87 (m, 2H, ArH), 8.03 (dt, J = 15.98, 6.24 Hz, 1H, ArH), 8.25–8.28 (m, 1H, ArH), 8.32 (s, 1H, ArH), 8.92 (t, J = 6.03 Hz, 1H, NH), 9.59 (s, 1H, ArH), 10.11 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, DMSO- $d_6$ )  $\delta$  42.57, 105.92 (dd, J = 25.85, 22.19 Hz), 111.54 (d, J = 23.37 Hz), 116.46, 116.49, 123.60 (d, J = 272.57 Hz), 123.67, 124.92, 130.50 (q, J = 32.79 Hz), 131.29, 137.02, 144.10, 145.21 (dt, J = 223.08, 14.88 Hz), 147.69, 149.64, 157.23, 158.65, 167.99. IR (KBr): 3402, 3230, 3106, 1671, 1560, 1505, 1433, 1321, 1131, 801 cm<sup>-1</sup>. EIMS m/z: 443 (M<sup>+</sup>, 2), 391 (21), 297 (15), 269 (23), 257 (50), 241 (10), 240 (87), 217 (92), 213 (14), 172 (26), 165 (95), 159 (68), 147 (100), 146 (18), 145 (21), 139 (11), 119 (21). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>18</sub>H<sub>11</sub>F<sub>6</sub>N<sub>5</sub>O<sub>2</sub>: 443.0817; Found 443.0812.

#### 4.1.2.6. 1-(4-Chlorophenyl)-5-(furan-2-yl)-N-(2-oxo-2-((2,4,5-tri-

fluorophenyl) amino) ethyl)-1H-1,2,4-triazole-3-carboxamide (4l). Yield 83%, white solid, m.p. 188–189 °C, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  4.32 (d, J = 5.91 Hz, 2H, NHCH<sub>2</sub>), 6.48 (d, J = 1.94 Hz, 1H, ArH), 6.77 (d, J = 2.97 Hz, 1H, ArH), 6.95 (td, J = 9.78, 6.90 Hz, 1H, ArH), 7.41 (d, J = 8.35 Hz, 1H, ArH), 7.46 (s, 1H, ArH), 7.49 (d, J = 8.62 Hz, 2H, ArH), 7.87 (br, 1H, NH), 8.22 (dt, J = 15.59, 5.85 Hz, 1H, ArH), 8.37 (br, 1H, NH), 8.22 (dt, J = 15.59, 5.85 Hz, 1H, ArH), 8.37 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  44.74, 105.91 (t, J = 24.23 Hz), 110.61 (d, J = 24.39 Hz), 112.05, 114.55, 122.14, 127.25 (2 × C), 129.60 (2 × C), 135.82, 136.23, 141.28, 145.14, 145.33, 146.90, 147.34, 147.38, 155.88, 159.94, 166.74. IR (KBr): 3405, 3123, 2926, 1708, 1678, 1613, 1549, 1491,1436, 1230, 1090, 835 cm<sup>-1</sup>. EIMS *m/z*: 475 (M<sup>+</sup>, 5), 331 (16), 329 (45), 304 (17), 303 (19), 302 (52), 301 (33), 274 (33), 273 (18), 272 (100), 247 (17), 245 (50), 206 (12), 204 (35), 151 (22), 147 (24), 111 (23). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>21</sub>H<sub>13</sub>ClF<sub>3</sub>N<sub>5</sub>O<sub>3</sub>: 475.0659; Found 475.0653.

#### 4.1.2.7. 1-(4-Chlorophenyl)-5-cyclopentyl-N-(2-oxo-2-((2,4,5-tri-

fluorophenyl) amino)ethyl)-1H-1,2,4-triazole-3-carboxamide (**4m**). Yield 74%, yellow solid, m.p. 159–160 °C, <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.55–1.57 (m, 2H, CH<sub>2</sub>), 1.77–1.92 (m, 6H, CH<sub>2</sub>), 3.05 (quint, J = 8.20 Hz, 1H, CH), 4.39 (d, J = 5.60 Hz, 2H, NHCH<sub>2</sub>), 6.84 (td, J = 9.96, 7.07 Hz, 1H, ArH), 7.31 (dd, J = 6.78, 1.90 Hz, 2H, ArH), 7.43 (dd, J = 6.72, 1.92 Hz, 2H, ArH), 8.06 (dt, J = 15.45, 6.12 Hz, 1H, ArH), 9.12 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  25.65 (2 × C), 32.80 (2 × C), 36.03, 43.93, 104.77 (t, J = 23.28 Hz), 111.06 (d, J = 24.19 Hz), 121.15, 126.62 (2 × C), 129.60 (2 × C), 135.18, 135.54, 145.60 (d, J = 250.82 Hz), 145.86 (dt, J = 247.30, 14.53 Hz), 149.26 (d, J = 10.65 Hz), 155.28, 160.21, 161.89, 167.33. IR (KBr): 3402, 3293, 2960, 1682, 1542, 1495, 1433, 1209, 835 cm<sup>-1</sup>. EIMS *m/z*: 477 (M<sup>+</sup>, 5), 333 (12), 331 (36), 306 (26), 305 (24), 304 (82), 303 (32), 291 (11), 276 (24), 275 (15), 274 (74), 252 (30), 251 (12), 250 (100), 248 (17), 247 (18), 246 (37), 232 (28), 206 (13), 151 (27), 147 (16), 141 (22), 138 (10), 129

(12), 127 (57), 125 (62), 111 (19). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for  $C_{22}H_{19}ClF_3N_5O_2$ : 477.1171; Found 477.1179.

4.1.2.8. N-(2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl)-5-(1H-pyrrol-2-vl)-1-(3-(trifluoromethyl)phenyl)-1H-1,2,4-triazole-3-carboxamide (4n). Yield 73%, light yellow solid, m.p. 181–182 °C, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  4.38 (d, J = 5.36 Hz, 2H, NHCH<sub>2</sub>), 5.83 (s, 1H, ArH), 6.01 (dd, J = 6.00, 2.55 Hz, 1H, ArH), 6.90–6.96 (m, 2H, ArH), 7.65–7.71 (m, 2H, ArH), 7.80–7.83 (m, 2H, ArH), 7.94 (t, J = 5.20 Hz, 1H, NH), 8.16 (dt, J = 16.03, 5.60 Hz, 1H, ArH), 8.60 (br, 1H, NH), 9.78 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  44.38, 104.95 (t, J = 23.49 Hz), 110.60, 110.82 (d, *J* = 23.88 Hz), 111.49, 117.77, 122.02, 122.31, 123. 63 (d, J = 3.43 Hz), 124.19, 127.07 (d, J = 3.81 Hz), 129.68, 130.36, 132.47 (q, J = 33.77 Hz), 137.88,145.28, 146.92, 148.80, 150.06, 155.47, 160.12, 166.93. IR (KBr): 3276, 3079, 2926, 1675, 1549, 1501, 1437, 1328, 1131 cm<sup>-1</sup>. EIMS *m/z*: 508 (M<sup>+</sup>, 10), 362 (34), 361 (24), 335 (40), 334 (28), 306 (18), 305 (100), 278 (32), 277 (25), 237 (42), 185 (13), 147 (15), 145 (22), 92 (13). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>22</sub>H<sub>14</sub>F<sub>6</sub>N<sub>6</sub>O<sub>2</sub>: 508.1082; Found 508.1088.

#### 4.1.2.9. 5-Butyl-1-(4-chlorophenyl)-N-(2-oxo-2-((2,4,5-trifluorophenyl) amino) ethyl)-1H-1,2,4-triazole-3-carboxamide (40). Yield 82%, white solid, m.p. 131–132 °C, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) $\delta$ 0.85 (t, J = 7.35Hz, 3H, $CH_3$ ), 1.30 (sext, J = 7.52 Hz, 2H, $CH_2$ ), 1.70 (quint, J = 7.64 Hz, 2H, CH<sub>2</sub>), 2.74 (t, J = 7.73 Hz, 2H, CH<sub>2</sub>), 4.35 (d, J = 5.80 Hz, 2H, NHCH<sub>2</sub>), 6.90 (td, J = 9.82, 7.21 Hz, 1H, ArH), 7.35 (d, J = 8.60 Hz, 2H, ArH), 7.47 (d, J = 8.60 Hz, 2H, ArH), 7.94 (t, J = 5.50 Hz, 1H, NH), 8.14 (dt, J = 15.86, 5.85 Hz), 8.77 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) $\delta$ 13.54, 22.17, 26.20, 29.51, 44.34, 104.85 (t, J = 23.61 Hz), 111.89 (d, J = 24.60 Hz), 122.18, 126.38 (2 $\times$ C), 129.75 (2 $\times$ C), 135.19, 135.67, 145.83 (dt, J = 249.22, 12.94 Hz), 146.14 (ddd, J = 243.81, 12.74, 2.93 Hz), 148.81, 155.36, 157.87, 160.26, 167.13. IR (KBr): 3313, 2963, 2933, 2871, 1729, 1678, 1546, 1495, 1433, 1216, 1090, 1015, 835 cm<sup>-1</sup>. EIMS *m/z*: 465 (M<sup>+</sup>, 6), 321 (18), 319 (51), 294 (26), 293 (27), 292 (76), 291 (47), 264 (33), 263 (20), 262 (100), 237 (13), 236 (11), 234 (29), 194 (13), 193 (19), 192 (12). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>21</sub>H<sub>19</sub>ClF<sub>3</sub>N<sub>5</sub>O<sub>2</sub>: 465.1179; Found 465.1176.

4.1.2.10. N-(2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl)-1,5-diphenyl-1H-1,2,4- triazole-3-carboxamide (**4p**). Yield 82%, white solid, m.p. 210–212 °C, <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  4.34 (d, J = 6.04 Hz, 2H, NHCH<sub>2</sub>), 6.94 (td, J = 9.99, 7.02 Hz, 1H, ArH), 7.32–7.38 (m, 4H, ArH), 7.41–7.46 (m, 4H, ArH), 7.47–7.50 (m, 2H, ArH), 7.96 (t, J = 5.92 Hz, 1H, NH), 8.21 (dt, J = 15.77, 5.98 Hz, 1H, ArH), 8.55 (br, 1H, NH). <sup>13</sup>C {<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  44.70, 104.89 (t, J = 23.29 Hz), 110.66 (d, J = 24.61 Hz), 122.18, 125.45 (2 × C), 126.72, 128.72 (2 × C), 128.96 (2 × C), 129.51 (2 × C), 129.64, 130.70, 137.55,145.82 (d, J = 249.18 Hz), 146.41, 148.93, 155.24, 155.50, 160.40, 166.98. IR (KBr): 3286, 3069, 1671, 1546, 1498, 1433, 1209, 692 cm<sup>-1</sup>. EIMS *m*/*z*: 451 (M<sup>+</sup>, 5), 305 (39), 279 (14), 278 (92), 277 (31), 249 (19), 248 (100), 222 (13), 221 (58), 180 (56), 147 (12), 117 (19), 77 (22). HRMS (EI) *m*/*z*: [M]<sup>+</sup> Calcd for C<sub>23</sub>H<sub>16</sub>F<sub>3</sub>N<sub>5</sub>O<sub>2</sub>: 451.1256; Found 451.1251.

# 4.1.2.11. N-(2-Oxo-2-((2,4,5-trifluorophenyl)amino)ethyl)-5-phenyl-1-(p-tolyl)-1H- 1,2,4- triazole-3-carboxamide (**4q**). Yield 84%, yellow solid, m.p. 121–122 °C, <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) $\delta$ 2.34 (s, 3H, CH<sub>3</sub>), 4.44 (d, J = 5.64 Hz, 2H, NHCH<sub>2</sub>), 6.80 (td, J = 9.91, 7.15 Hz, 1H, ArH), 7.14 (s, 4H, ArH), 7.26 (t, J = 7.52 Hz, 2H, ArH), 7.35 (t, J = 7.38 Hz, 1H, ArH), 7.42 (d, J = 7.80 Hz, 2H, ArH), 8.02 (dt, J = 15.79, 5.95 Hz, 1H, ArH), 8.21 (t, J = 5.62 Hz, 1H, NH), 9.22 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>) $\delta$ 21.07, 43.88, 104.69 (t, J = 23.40 Hz), 111.25 (d, J = 23.75 Hz), 122.13, 125.03 (2 × C), 126.68, 128.45 (2 × C), 128.75 (2 × C), 129.87 (2 × C), 130.36, 134.92, 139.64, 144.67, 146.70, 149.45, 154.85, 155.45, 160.18, 167.44. IR (KBr): 3286, 3058, 2922, 1739, 1695, 1549, 1512, 1430, 1209 cm<sup>-1</sup>. EIMS *m/z*: 465 (M<sup>+</sup>, 5), 447

(13), 391 (12), 319 (32), 293 (19), 292 (91), 291 (18), 279 (12), 263 (25), 262 (100), 236 (19), 235 (84), 195 (15), 194 (87), 147 (16), 132 (23), 131 (40), 129 (13), 105 (54), 104 (29), 103 (12). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>24</sub>H<sub>18</sub>F<sub>3</sub>N<sub>5</sub>O<sub>2</sub>: 465.1413; Found 465.1406.

## 4.1.3. Standard procedure for synthesis of $\beta$ -amino esters **5d** and **5m**-q [39,43]

The 1*H*-1,2,4-triazol-3-ylmethanols **12d**, and **12m–q** (1.0 equiv, 3.0 mmol) were treated with Boc-(*R*)-3-amino-4-(2,4,5-trifluoro-phenyl)butyric acid **14** (1.1 equiv) and *N*,*N*-bis[2-oxo-3-oxazolidinyl]phosphorodiamide chloride (BOP-Cl, 2.0 equiv) in presence of NEt<sub>3</sub> (2.0 equiv) in CH<sub>2</sub>Cl<sub>2</sub> solution (20 mL) [39]. The reaction mixture was stirred at room temperature for 4 h. When the reaction was completed, the reaction mixture was washed with water (15 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (15 mL × 3). The combined organic layers were washed sodium bicarbonate (15 mL × 3), dried over MgSO<sub>4</sub> to give crude Boc-protected  $\beta$ -amino esters **15d** and **15m–q**. The reaction mixture of crude Bocprotected  $\beta$ -amino esters **15d** was purified by column chromatography on silica gel to give the pure corresponding  $\beta$ -amino esters **15d** in 84% yield. The crude intermediates **15m–q** were directly carried out the deprotection without the further purification.

Boc-Protected  $\beta$ -amino esters **15d** and **15m–q** (1.0 equiv) were added with trifluoroacetic acid (TFA, 1.5 equiv) in CH<sub>2</sub>Cl<sub>2</sub> solution (15 mL) [43]. The reaction mixture was stirred at room temperature for 3 h. When the reaction was completed, the reaction mixture was nertulized with sodium bicarbonate (15 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 30 mL). The combined organic layers were dried over MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure to afford the corresponding crude  $\beta$ -amino esters **5d** and **5m–q**. The crude products **5d** and **5m–q** were purified by column chromatography on silica gel to give the pure corresponding  $\beta$ -amino esters **5d** and **5m–q** in 74–84% yields.

4.1.3.1. (1-(4-(Trifluoromethyl)phenyl)-1H-1,2,4-triazol-3-yl)methyl (R)-3-((tertbutoxycarbonyl)amino)-4-(2,4,5-trifluorophenyl)butanoate (15d). Yield 84%, light yellow solid, m.p. 89–91 °C, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 1.33 (s, 9H, CH<sub>3</sub>), 2.60–2.69 (m, 2H, CH<sub>2</sub>), 2.86 (br, 2H,  $CH_2$ ), 4.16 (br, 1H, CH), 5.32 (d, J = 6.15 Hz, 2H,  $CH_2$ ), 6.84 (dd, J =15.36, 8.79 Hz, 1H, ArH), 7.03 (dd, J = 15.62, 8.73 Hz, 1H, ArH), 7.75 (d, J = 7.92 Hz, 2H, ArH), 7.80 (d, J = 8.30 Hz, 2H, ArH), 8.62 (s, 1H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 125 MHz):  $\delta$  28.18 (3 × C), 32.90, 37.92, 47.69, 59.16, 79.52, 105.27 (dd, J = 28.60, 20.84 Hz), 119.07 (d, J =18.93 Hz), 119.85 (2  $\times$  C), 121.32 (d, J = 18.72 Hz), 123.47 (d, J =272.17 Hz), 127.12, 127.14, 130.37 (q, J = 33.06 Hz), 139.14, 141.87, 145.57 (dd, *J* = 244.16, 12.51 Hz), 148.83 (dt, *J* = 251.26, 13.54 Hz), 155.14, 156.20 (dd, J = 244.02, 8.84 Hz), 160.78, 170.78. IR (KBr): 3354, 2977, 2929, 1739, 1705, 1692, 1522, 1328, 1168, 842 cm<sup>-1</sup>. ESIMS m/z: 581 ([M+Na]<sup>+</sup>, 15), 560 (25), 559 ([M+H]<sup>+</sup>, 100), 395 (11), 377 (14). HRMS (ESI) *m/z*: [M+H]<sup>+</sup> Calcd for C<sub>25</sub>H<sub>25</sub>F<sub>6</sub>N<sub>4</sub>O<sub>4</sub>: 559.1775; Found 559.1785.

4.1.3.2. (1-(4-(Trifluoromethyl)phenyl)-1H-1,2,4-triazol-3-yl)methyl (R)-3-amino-4- (2,4,5-trifluorophenyl)butanoate (5d). Yield: 76%, light yellow solid, m.p. 135–136 °C, <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta$  2.36 (dd, J = 15.78, 8.63 Hz, 1H, CH<sub>2</sub>), 2.51 (dd, J = 15.76, 4.10 Hz, 1H, CH<sub>2</sub>), 2.59 (dd, J = 13.71, 7.65 Hz, 1H, CH<sub>2</sub>), 2.67 (dd, J = 13.76, 6.00 Hz, 1H, CH<sub>2</sub>), 3.42 (quint, J = 6.53 Hz, 1H, CH<sub>2</sub>), 2.67 (dd, J = 16.04 Hz, 2H, OCH<sub>2</sub>), 6.78 (td, J = 9.52, 6.80 Hz, 1H, ArH), 6.98 (dd, J = 16.93, 8.98 Hz, 1H, ArH), 7.65 (d, J = 8.60 Hz, 2H, ArH), 7.72 (d, J = 8.50 Hz, 2H, ArH), 8.60 (s, 1H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 125 MHz):  $\delta$  35.14, 41.34, 48.58, 58.84, 105.18 (dd, J = 28.77, 20.72 Hz), 118.79 (dd, J = 18.88, 6.00 Hz), 119.48 (2 × C), 121.60 (dt, J = 18.17, 4.55 Hz), 123.33 (d, J = 272.19 Hz), 126.82, 126.85, 129.87 (q, J = 33.17 Hz), 138.97, 141.2, 146.33 (ddd, J = 244.11, 7.28 Hz), 160.65, 171.22. IR (KBr): 3221, 3116, 2922, 1729, 1617, 1549, 1522, 1335, 1179, 1138, 1049, 845 cm<sup>-1</sup>. ESIMS m/z: 460 (22), 459 (M + H<sup>+</sup>, 100), 317 (3), 258 (4), 244 (7), 173 (3). HRMS (ESI) m/z: [M+H]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>16</sub>F<sub>6</sub>N<sub>4</sub>O<sub>2</sub>: 459.1177; Found 459.1244.

4.1.3.3. (1-(4-Chlorophenyl)-5-cyclopentyl-1H-1,2,4-triazol-3-yl)methyl (R)-3- amino-4-(2,4,5-trifluorophenyl)butanoate (5m). Yield 79%, deep vellow liquid <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.57 (t, J = 5.50 Hz, 2H, CP), 1.64 (br, 2H, NH<sub>2</sub>), 1.80–1.93 (m, 6H, CP), 2.37 (dd, J = 15.69, 8.58 Hz, 1H,  $CH_2$ ), 2.54 (dd, J = 15.69, 4.04 Hz, 1H,  $CH_2$ ), 2.62 (dd, J = 13.66, 7.67 Hz, 1H, CH<sub>2</sub>), 2.71 (dd, J = 13.73, 5.87 Hz, 1H, CH<sub>2</sub>), 3.04 (quint, J = 8.19 Hz, 1H, CP), 3.45 (quint, J = 6.50 Hz, 1H, CH), 5.19 (d, J = 2.19 Hz, 2H, OCH<sub>2</sub>), 6.86 (dd, J = 9.42, 6.86 Hz, 1H, ArH), 7.02 (dd, J = 16.91, 8.99 Hz, 1H, ArH), 7.32 (d, J = 8.56 Hz, 2H, ArH), 7.44 (d, J = 8.55 Hz, 2H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  25.68 (2 × C), 32.90 (2 × C), 36.15, 36.20, 41.77, 48.74, 59.48, 105.37 (dd, *J* = 28.74, 20.68 Hz), 118.96 (dd, J = 18.86, 5.99 Hz), 121.88 (dt, J = 18.14, 4.49 Hz), 126 63 (2 × C), 129.59 (2 × C), 135.07, 135.70, 146.53 (ddd, J = 244.68, 12.43, 3.30 Hz), 148.70 (dt, J = 250.01, 13.44 Hz), 156.09 (dd, J = 244.24, 9.06 Hz), 158.53, 161.37, 171.41. IR (KBr): 3374, 3296, 3057, 2958, 2869, 1736, 1519, 1488, 1151, 1093, 836 cm  $^{-1}$ . ESIMS m/z: 492 ([M+H]<sup>+</sup> + 2, 34),494 (26), 493 ([M+H]<sup>+</sup>, 100), 381 (3), 292 (4). HRMS (ESI) *m/z*: [M+H]<sup>+</sup> Calcd for C<sub>24</sub>H<sub>25</sub>ClF<sub>3</sub>N<sub>4</sub>O<sub>2</sub>: 493.1613; Found 493.1601.

4.1.3.4. (5-(1H-Pyrrol-2-yl)-1-(3-(trifluoromethyl)phenyl)-1H-1,2,4-triazol-3-yl) methyl(R)-3-amino-4-(2,4,5-trifluorophenyl)butanoate (5n). Yield 74%, yellow liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  2.01 (br, 2H, NH<sub>2</sub>), 2.41 (dd, J = 15.61, 8.80 Hz, 1H, CH<sub>2</sub>), 2.59 (dd, J = 15.58, 3.98 Hz, 1H, CH<sub>2</sub>), 2.66 (dd, J = 13.73, 7.73 Hz, 1H, CH<sub>2</sub>), 2.74 (dd, J = 13.76, 5.90 Hz, 1H, CH<sub>2</sub>), 3.51 (quint, J = 6.58 Hz, 1H, CH), 5.25 (d, J = 6.55 Hz, 2H, OCH<sub>2</sub>), 5.81 (d, J = 3.35 Hz, 1H, ArH), 6.07 (t, J = 2.83 Hz, 1H, ArH), 6.84–6.89 (m, 2H, ArH), 7.02 (dd, J = 17.05, 8.85 Hz, 1H, ArH), 7.64 (t, J = 7.80 Hz, 1H, ArH), 7.70 (d, J = 8.10 Hz, 1H, ArH), 7.78 (d, J = 7.90 Hz, 1H, ArH), 7.80 (s, 1H, ArH), 10.20 (br, 1H, NH).<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  36.21, 41.85, 48.82, 59.07, 105.44 (dd, J =28.74, 20.71 Hz), 110.23, 110.87, 118.39, 118.96 (dd, J = 18.86, 5.87 Hz), 121.65, 121.83 (t, J = 4.51 Hz), 123.20 (d, J = 272.65 Hz), 123.58 (d, J = 3.52 Hz), 126.52 (d, J = 3.37 Hz), 129.68, 130.20, 132.29 (q, J = 3.52 Hz), 126.52 (d, J = 3.37 Hz), 129.68, 130.20, 132.29 (q, J = 3.37 Hz))33.46 Hz), 138.33, 146.58 (ddd, J = 244.74, 12.39, 3.26 Hz), 148.78 (dt, J = 250.09, 13.40 Hz), 149.50, 156.12 (dd, J = 243.94, 9.22 Hz), 158.96, 171.50. IR (KBr): 3360, 3289, 3071, 2926, 2855, 1738, 1599, 1520, 1329, 1175, 1134, 739 cm<sup>-1</sup>. EIMS *m/z*: 523 (M<sup>+</sup>, 5), 379 (17), 378 (100), 310 (14), 309 (95), 307 (20), 292 (21), 291 (95), 199 (13), 170 (23), 151 (41), 145 (24). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>24</sub>H<sub>19</sub>F<sub>6</sub>N<sub>5</sub>O<sub>2</sub>: 523.1443; Found 523.1448.

4.1.3.5. (5-Butyl-1-(4-chlorophenyl)-1H-1,2,4-triazol-3-yl)methyl (R)-3amino-4- (2,4,5-trifluorophenyl)butanoate (50). Yield: 78%, orange liquid, <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.79 (t, J = 7.36 Hz, 3H, CH<sub>3</sub>), 1.25 (sext, J = 7.44 Hz, 2H, CH<sub>2</sub>CH<sub>3</sub>), 1.63 (quint, J = 7.67 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.83 (br, 2H, NH<sub>2</sub>), 2.34 (dd, J = 15.75, 8.58 Hz, 1H, CH<sub>2</sub>), 2.51 (dd, J = 15.79, 3.90 Hz, 1H, CH<sub>2</sub>), 2.59 (dd, J = 13.56, 7.64 Hz, 1H, CH2), 2.65-2.70 (m, 3H, CH2CH2CH2CH3, CH2), 3.42 (br, 1H, CH), 5.15 (s, 2H, OCH<sub>2</sub>), 6.81 (td, J = 9.57, 6.69 Hz, 1H, ArH), 6.98 (dd, J = 17.05, 8.89 Hz, 1H, ArH), 7.28 (d, J = 8.70 Hz, 2H, ArH), 7.40 (d, J = 8.68 Hz, 2H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  13.40, 22.10, 26.06, 29.53, 36.03, 41.52, 48.58, 59.23, 105.25 (dd, *J* = 28.62, 20.78 Hz), 118.85 (dd, *J* = 18.71, 5.51 Hz), 121.73 (dt, *J* = 18.01, 4.22 Hz), 126 11 (2 × C), 129.50 (2 × C), 134.86, 135.45, 146.39 (dd, *J* = 244.75, 12.70 Hz), 148.55 (dt, *J* = 249.64, 13.41 Hz), 155.91 (d, *J* = 244.68 Hz), 157.21, 158.47, 171.30. IR (KBr): 3378, 3289, 3057, 2960, 2933, 1738, 1519, 1489, 1152, 1093, 837 cm<sup>-1</sup>. ESIMS *m/z*: 483 ([M+H]<sup>+</sup> + 2, 35), 482 (26), 481 ([M+H]<sup>+</sup>, 100), 289 (4), 280 (5). HRMS (ESI) m/z: [M+H]<sup>+</sup> Calcd for C<sub>23</sub>H<sub>25</sub>ClF<sub>3</sub>N<sub>4</sub>O<sub>2</sub>: 481.1613; Found 481.1601.

4.1.3.6. (1,5-Diphenyl-1H-1,2,4-triazol-3-yl)methyl (R)-3-amino-4-(2,4,5*trifluorophenyl)butanoate (5p).* Yield: 83%, orange liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  2.42 (dd, J = 15.76, 8.60 Hz, 1H, CH<sub>2</sub>), 2.58 (dd, J =15.78, 4.08 Hz, 1H, CH<sub>2</sub>), 2.64 (dd, J = 13.76, 7.65 Hz, 1H, CH<sub>2</sub>), 2.73 (dd, J = 13.76, 5.90 Hz, 1H, CH<sub>2</sub>), 3.48 (quint, J = 6.55 Hz, 1H, CH), 5.30 (d, J = 4.35 Hz, 2H, OCH<sub>2</sub>), 6.85 (td, J = 9.60, 6.65 Hz, 1H, ArH), 7.02 (dd, J = 17.11, 8.85 Hz, 1H, ArH), 7.29–7.31 (m, 4H, ArH), 7.36–7.39 (m, 4H, ArH), 7.43–7.45 (m, 2H, ArH).  $^{13}C{^{1}H}$  NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 36.20, 41.73, 48.71, 59.41, 105.36 (dd, *J* = 28.82, 20.59 Hz), 118.97 (dd, J = 18.81, 6.12 Hz), 121.86 (d, J = 18.45 Hz), 125.25 (2  $\times$  C), 127.36, 128.51 (2 × C), 128.82 (2 × C), 129.00, 129.35 (2 × C), 130.16, 137.84, 146.52 (dd, J = 244.77, 12.59 Hz), 148.68 (dt, J = 249.87, 13.35 Hz), 154.84, 156.06 (dd, J = 243.48, 9.19 Hz), 158.92, 163.24, 171.46. IR (KBr): 3370, 3299, 3064, 2929, 2858, 1738, 1519, 1424, 1152, 766, 694 cm<sup>-1</sup>. EsIMS *m/z*: 478 (27), 467 (M<sup>+</sup>, 100), 381 (2), 266 (6). HRMS (ESI) m/z:  $[M+H]^+$  Calcd for C<sub>25</sub>H<sub>22</sub>F<sub>3</sub>N<sub>4</sub>O<sub>2</sub>: 467.1689; Found 467.1680.

4.1.3.7. (5-Phenyl-1-(p-tolyl)-1H-1,2,4-triazol-3-yl)methyl (R)-3-amino-4-(2,4,5- trifluorophenyl)butanoate (5q). Yield: 84%, light yellow solid, m.p. 71–73 °C, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  2.34 (s, 3H, CH<sub>3</sub>), 2.40 (dd, J = 15.71, 8.60 Hz, 1H, CH<sub>2</sub>), 2.57 (dd, J = 15.68, 4.08 Hz, 1H, CH<sub>2</sub>), 2.62 (dd, J = 13.77, 7.70 Hz, 1H, CH<sub>2</sub>), 2.72 (dd, J = 13.76, 5.85 Hz, 1H, CH<sub>2</sub>), 3.47 (quint, J = 6.47 Hz, 1H, CH), 5.28 (d, J = 4.90 Hz, 2H, OCH<sub>2</sub>), 6.83 (td, J = 14.32, 6.69 Hz, 1H, ArH), 7.01 (dd, J = 16.98, 8.98 Hz, 1H, ArH), 7.16 (s, 4H, ArH), 7.27 (t, J = 7.50 Hz, 2H, ArH), 7.34 (t, J = 7.38 Hz, 1H, ArH), 7.43 (d, J = 7.30 Hz, 2H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz,  $CDCl_3$ )  $\delta$  20.91, 36.03, 41.65, 48.61, 59.27, 105.17 (dd, J = 28.78, 20.72 Hz), 118.83 (dd, J = 18.82, 6.02 Hz), 121.81 (dt, J = 18.11, 4.55 Hz), 124.93 (2 × C), 127.34, 128.31 (2 × C), 128.63 (2 × C), 129.76 (2  $\times$  C), 129.90, 135.26, 139.00, 146.33 (dd, J = 245.24, 12.39 Hz), 148.49 (dt, J = 249.95, 13.62 Hz), 154.56, 155.92 (dd, J = 241.75, 8.93 Hz), 158.63, 171.29. IR (KBr): 3449, 3374, 3299, 3050, 2954, 2926, 2858, 1738, 1519, 1423, 1151, 823, 732, 696 cm<sup>-1</sup>. ESIMS m/z: 482, (28), 481 ( $[M+H]^+$ , 100), 468 (2), 280 (6). HRMS (ESI) m/z:  $[M+H]^+$ Calcd for C<sub>26</sub>H<sub>24</sub>F<sub>3</sub>N<sub>4</sub>O<sub>2</sub>: 481.1846; Found 481.1845.

## 4.1.4. Standard procedure for synthesis of $\beta$ -amino amides **6d** and **6m**–q [39,43]

The 1*H*-1,2,4-triazol-3-ylmethanamines **13d** and **13m**–**q** (1.0 equiv, 3.0 mmol) were treated with Boc-(*R*)-3-amino-4-(2,4,5-trifluoro-phenyl)butyric acid **14** (1.1 equiv) and *N*,*N*-bis[2-*oxo*-3-*oxazolidinyl*]phosphorodiamide chloride (BOP-Cl, 2.0 equiv) in presence of NEt<sub>3</sub> (2.0 equiv) in CH<sub>2</sub>Cl<sub>2</sub> solution (20 mL) [39]. The reaction mixture was stirred at room temperature for 4 h. When the reaction was completed, the reaction mixture was washed with water (15 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (15 mL × 3). The combined organic layers were washed sodium bicarbonate (15 mL × 3), dried over MgSO<sub>4</sub> to give crude Boc-protected  $\beta$ -amino amides **16d** and **16m–q**. Without further purification, the crude intermediates **16d** and **16m–q** were directly carried out the deprotection.

Boc-Protected  $\beta$ -amino amides **16d** and **16m–q** (1.0 equiv) were added with trifluoroacetic acid (TFA, 1.5 equiv) in CH<sub>2</sub>Cl<sub>2</sub> solution (15 mL) [43]. The reaction mixture was stirred at room temperature for 3 h. When the reaction was completed, the reaction mixture was nertulized with sodium bicarbonate (15 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 30 mL). The combined organic layers were dried over MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure to afford the corresponding crude  $\beta$ -amino amides **6d** and **6m–q**. The crude products **6d** and **6m–q** were purified by column chromatography on silica gel to give the pure corresponding  $\beta$ -amino esters **6d** and **6m–q** in 74–88% yields.

4.1.4.1. (*R*)-3-*Amino-N-((1-(4-(trifluoromethyl)phenyl)-1H-1,2,4-triazol-3-yl) methyl)-4-(2,4,5-trifluorophenyl)butanamide (6d). Yield 79%, light yellow solid, m.p. 50–51 °C, <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): \delta 2.25 (dd, <i>J* = 15.09, 9.28 Hz, 1H, CH<sub>2</sub>), 2.46 (dd, *J* = 15.19, 1.90 Hz, 1H, CH<sub>2</sub>), 2.66 (dd, *J* = 13.41, 7.88 Hz, 1H, CH<sub>2</sub>), 2.76 (dd, *J* = 13.77, 5.35 Hz, 1H,

CH<sub>2</sub>), 3.44 (d, J = 6.71 Hz, CH), 4.63 (d, J = 5.15 Hz, 2H, NHCH<sub>2</sub>), 6.89 (q, J = 8.47 Hz, 1H, ArH), 7.03 (q, J = 8.55 Hz, 1H, ArH), 7.72–7.77 (m, 4H, ArH), 8.54 (s, 1H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  37.19 (2 × C), 42.66, 49.09, 105.55 (dd, J = 28.73, 20.45 Hz), 118.99 (dd, J = 18.82, 5.92 Hz), 119.60 (2 × C), 121.57 (d, J = 17.93 Hz), 123.50 (d, J = 273.01 Hz), 127.06, 127.10, 130.06 (d, J = 33.53 Hz), 139.21, 141.58, 146.54 (d, J = 243.56 Hz), 148.74 (dd, J = 249.08, 15.03 Hz), 156.06 (d, J = 245.32 Hz), 162.74, 171.46. IR (KBr): 3364, 3092, 2926, 1617, 1535, 1325, 1165, 1124, 841 cm<sup>-1</sup>. ESIMS *m/z*: 459 (23), 458 ([M+H]<sup>+</sup>, 100), 285 (17), 243 (20). HRMS (EI) *m/z*: [M+H]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>18</sub>F<sub>6</sub>N<sub>5</sub>O: 458.1410; Found 458.1414

4.1.4.2. (R)-3-Amino-N-((1-(4-chlorophenyl)-5-cyclopentyl-1H-1,2,4-triazol-3-yl) methyl)-4-(2,4,5-trifluorophenyl)butanamide (6m). Yield 83%, yellow liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.59–1.61 (m, 2H, CH<sub>2</sub>), 1.85–1.90 (m, 4H, CH<sub>2</sub>), 1.94–1.96 (m, 2H, CH<sub>2</sub>), 2.29 (dd, J = 14.73, 8.78 Hz, 1H, CH<sub>2</sub>), 2.46 (dd, J = 14.66, 2.25 Hz, 1H, CH<sub>2</sub>), 2.68-2.71 (m, 1H, CH<sub>2</sub>), 2.73 (br, 2H, NH<sub>2</sub>), 2.79 (dd, J = 13.53, 5.63 Hz, 1H, CH<sub>2</sub>), 3.06 (quint, J = 7.95 Hz, 1H, CH), 3.49 (br, 1H, CH), 4.55 (t, J = 4.13 Hz, 2H, NCH<sub>2</sub>), 6.90 (dt, J = 9.43, 6.74 Hz, 1H, ArH), 7.07 (dd, J = 16.73, 8.98 Hz, 1H, ArH), 7.34 (d, J = 8.45 Hz, 2H, ArH), 7.47 (d, J = 8.45 Hz, 2H, ArH), 7.65 (br, 1H, NH<sub>2</sub>).  ${}^{13}C{}^{1}H$  NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  25.54 (2 × C), 32.77 (2 × C), 36.01, 36.25, 37.25, 42.31, 48.98, 105.29 (dd, J = 28.66, 20.70 Hz), 118.94 (dd, J = 18.86, 5.89 Hz), 121.56 (d, J = 18.37 Hz), 126 49 (2  $\times$  C), 129.46 (2  $\times$  C), 134.84, 135.62, 146.44 (dd, J = 245, 12.10 Hz), 148.61 (dt, J = 250.07, 13.34 Hz), 155.97 (dd, J = 243.82, 7.98 Hz), 160.12, 161.03, 171.18. IR (KBr): 3434, 3349, 3289, 3054, 2957, 2872, 1659, 1518, 1424, 1210, 1093, 837 cm<sup>-1</sup>. EIMS *m/z*: 491 (M<sup>+</sup>, 1), 346 (10), 277 (21), 261 (18), 260 (21). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>24</sub>H<sub>25</sub>ClF<sub>3</sub>N<sub>5</sub>O: 491.1700; Found 491.1704.

#### 4.1.4.3. (R)-N-((5-(1H-Pyrrol-2-yl)-1-(3-(trifluoromethyl)phenyl)-1H-

1,2,4-triazol- 3-yl)methyl)-3-amino-4-(2,4,5-trifluorophenyl)butanamide (6n).. Yield 74%, light yellow liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$ 2.30 (dd, J = 14.86, 9.25 Hz, 1H, CH<sub>2</sub>), 2.45 (dd, J = 14.86, 3.10 Hz, 1H, CH<sub>2</sub>), 2.62–2.66 (m, 3H, CH<sub>2</sub> and NH<sub>2</sub>), 2.73 (dd, J = 13.73, 5.83 Hz, 1H, CH<sub>2</sub>), 3.49 (d, J = 6.75 Hz, 1H, CH), 4.56 (qd, J = 15.91, 5.32 Hz, 2H, NCH<sub>2</sub>), 5.76 (d, J = 3.10 Hz, 1H, ArH), 6.03 (t, J = 3.10 Hz, 1H, ArH), 6.81–6.86 (m, 2H, ArH), 6.97 (dd, J = 16.81, 8.90 Hz, 1H, ArH), 7.57–7.63 (m, 2H, ArH), 7.72–7.74 (m, 2H, ArH), 7.97 (t, J = 5.10 Hz, 1H, NH), 10.57 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) δ 36.48, 37.11, 42.52, 49.13, 105.45 (dd, *J* = 28.70, 20.68 Hz), 110.08, 110.89, 118.24, 118.94 (dd, J = 18.81, 5.86 Hz), 121.48 (d, J = 18.07 Hz), 121.71, 123.19 (d, J = 272.62 Hz), 123.43 (d, J = 3.52 Hz), 126.31 (d, J = 3.26 Hz), 129.57, 130.10, 132.14 (q, J = 33.41 Hz), 138.34, 146.57 (ddd, J = 244.91, 12.23, 3.43 Hz), 148.78 (dt, J = 250.27, 13.42 Hz),149.32, 156.08 (dd, J = 242.58, 8.84 Hz), 160.63, 171.76. IR (KBr): 3358, 3285, 3075, 2926, 2855, 1738, 1649, 1597, 1519, 1329, 1176, 1133, 1070, 738 cm<sup>-1</sup>. EIMS *m/z*: 523 (M<sup>+</sup>, 3), 378 (14), 377 (80), 309 (17), 308 (100), 306 (27), 292 (20), 291 (57), 171 (10), 151 (18), 145 (11). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>24</sub>H<sub>20</sub>F<sub>6</sub>N<sub>6</sub>O: 522.1603; Found 522.1612.

4.1.4.4. (*R*)-3-Amino-*N*-((5-butyl-1-(4-chlorophenyl)-1H-1,2,4-triazol-3yl)methyl)- 4-(2,4,5-trifluorophenyl)butanamide (**6o**). Yield: 81%, yellow liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  0.82 (t, *J* = 7.33 Hz, 3H, CH<sub>3</sub>), 1.28 (sext, *J* = 7.60 Hz, 2H, CH<sub>2</sub>CH<sub>3</sub>), 1.62 (quint, *J* = 7.63 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 2.41 (br, 1H, CH<sub>2</sub>), 2.50–2.52 (m, 1H, CH<sub>2</sub>), 2.68 (t, *J* = 7.67 Hz, 2H, CH<sub>2</sub>), 2.86 (br, 2H, CH<sub>2</sub>), 3.58 (br, 1H, CH), 4.11 (br, 2H, NH<sub>2</sub>), 4.52 (s, 2H, NCH<sub>2</sub>), 6.86 (td, *J* = 16.05, 9.32 Hz, 1H, ArH), 6.98 (dd, *J* = 15.14, 7.88 Hz, 1H, ArH), 7.30 (d, *J* = 8.41 Hz, 2H, ArH), 7.44 (d, *J* = 8.47 Hz, 2H, ArH) 7.60 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  13.53, 22.22, 26.15, 29.78, 36.14, 37.35, 42.04, 49.18, 105.53 (dd, *J* = 28.50, 20.65 Hz), 119.13 (dd, *J* = 18.82, 5.74 Hz), 121.35 (d, *J* = 20.24 Hz), 126 25 (2 × C), 129.67 (2 × C), 135.00, 135.67, 146.63 (d, J = 260.46 Hz, 148.88 (d, J = 250.23 Hz), 156.13 (d, J = 243.66 Hz), 157.12, 160.36, 171.15. IR (KBr): 3273, 3054, 2961, 2933, 2874, 1667, 1519, 1424, 1211, 1151, 1094, 838, 731 cm<sup>-1</sup>. EIMS m/z: 479 (M<sup>+</sup>, 1), 336 (14), 332 (42), 267 (30), 266 (16), 265 (100), 263 (16), 250 (11), 249 (18), 248 (26), 165 (15), 137 (13), 125 (10). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>23</sub>H<sub>25</sub>F<sub>3</sub>N<sub>5</sub>O: 479.1700; Found 479.1705.

4.1.4.5. (R)-3-Amino-N-((1,5-diphenyl-1H-1,2,4-triazol-3-yl)methyl)-4-(2,4,5- trifluorophenyl)butanamide (6p). Yield 88%, orange liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  2.25 (dd, J = 14.98, 8.93 Hz, 1H, CH<sub>2</sub>), 2.45 (d, 2.58 J = 14.91 Hz, 1H, CH<sub>2</sub>), 2.65 (dd, J = 15.53, 7.93 Hz, 1H, CH<sub>2</sub>), 2.76 (dd, J = 13.68, 5.34 Hz, 1H, CH<sub>2</sub>), 3.45 (br, 1H, CH), 4.64 (d, J = 5.20 Hz, 2H, NCH<sub>2</sub>), 6.84–6.89 (m, 1H, ArH), 7.01 (dd, J = 17.08, 8.78 Hz, 1H, ArH), 7.28-7.31 (m, 4H, ArH), 7.35-7.38 (m, 4H, ArH), 7.41–7.42 (m, 2H, ArH), 7.46 (br, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz,  $CDCl_3$ )  $\delta$  36.85, 37.42, 42.89, 49.07, 105.44 (dd, J = 28.77, 20.81 Hz), 119.03 (dd, J = 18.73, 6.30 Hz), 121.79 (d, J = 18.16 Hz), 125.26 (2  $\times$ C), 127.54, 128.57 (2  $\times$  C), 128.79 (2  $\times$  C), 128.93, 129.37 (2  $\times$  C), 130.15, 137.94, 146.63 (d, *J* = 241.14 Hz), 148.75 (d, *J* = 249.40 Hz), 154.63, 156.15 (dd, J = 244.04 Hz), 160.64, 171.35. IR (KBr): 3430, 3285, 3061, 2929, 1652, 1517, 1423, 1210, 1154, 765, 695 cm<sup>-1</sup>. EIMS *m/z*: 465 (M<sup>+</sup>, 2), 321 (13), 321 (74), 252 (17), 251 (100), 249 (19), 235 (21), 234 (37), 131 (12), 103 (20). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>25</sub>H<sub>22</sub>F<sub>3</sub>N<sub>5</sub>O: 465.1776; Found 465.1769.

#### 4.1.4.6. (R)-3-Amino-N-((5-phenyl-1-(p-tolyl)-1H-1,2,4-triazol-3-yl)

methyl)-4- (2,4,5-trifluorophenyl)butanamide (6q). Yield 86%, yellow liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  2.35 (dd, J = 14.88, 8.93 Hz, 1H, CH<sub>2</sub>), 2.47 (s, 3H, CH<sub>3</sub>), 2.54 (dd, J = 14.91, 3.30 Hz, 1H, CH<sub>2</sub>), 2.74 (dd, J = 13.71, 7.90 Hz, 1H, CH<sub>2</sub>), 2.85 (dd, J = 13.68, 5.73 Hz, 1H, CH<sub>2</sub>), 3.53–3.59 (m, 1H, CH), 4.74 (d, J = 5.30 Hz, 2H, NCH<sub>2</sub>), 6.97 (td, J = 9.42, 6.84 Hz, 1H, ArH), 7.12 (dd, J = 16.51, 9.35 Hz, 1H, ArH), 7.27 (s, 4H, ArH), 7.38–7.42 (m, 2H, ArH), 7.47 (t, J = 7.08 Hz, 1H, ArH), 7.53 (d, J = 7.75 Hz, 2H, ArH), 7.69 (t, J = 4.83 Hz, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  21.02, 36.71, 37.30, 42.85, 49.00, 105.30 (dd, J =28.69, 20.68 Hz), 118.93 (dd, *J* = 18.77, 6.01 Hz), 121.82 (dt, *J* = 18.20, 4.49 Hz), 124.99 (2  $\times$  C), 127.53, 128.43 (2  $\times$  C), 128.65 (2  $\times$  C), 129.84 (2 × C), 129.95, 135.38, 139.01, 146.44 (dd, J = 245.28, 12.82 Hz), 148.60 (dt, J = 250.12, 13.59 Hz), 154.41, 156.00 (dd, J = 244.24, 9.02 Hz), 160.41, 171.31. IR (KBr): 3424, 3286, 3050, 2926, 2865, 1651, 1518, 1423, 1210, 1151, 823, 698 cm<sup>-1</sup>. EIMS *m/z*: 479 (M<sup>+</sup>, 2), 335 (18), 334 (87), 266 (22), 265 (100), 263 (24), 249 (26), 248 (44), 145 (21), 117 (17), 105 (12). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>26</sub>H<sub>24</sub>F<sub>3</sub>N<sub>5</sub>O: 479.1933; Found 479.1941.

## 4.1.5. Standard procedure for synthesis of 2-chloro-N-(2,4,5-trifluorophenyl)acetamide **9** [31,51]

2,4,5-Trifluorobenzenamine (1.0 equiv, 5.0 mmol) was dissolved in toluene (20 mL) and stirred in an ice-bath. 2-Chloroacetyl chloride (1.1 equiv.) was slowly added to the reaction mixture under N2. Then the mixture was heated and stirred at refliux (ca. 108 °C) within 3 h. When the reaction was completed, the reaction mixture was added to water (15 mL) and extracted with ethyl acetate (15 mL  $\times$  3). The combined organic layers were washed sodium bicarbonate (15 mL  $\times$  3), dried over MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure to afford the corresponding crude 2-chloro-N-(2,4,5-trifluorophenyl)acetamide 9. The crude desired products 9 was recrystallized in EtOH/hexane (3/1) solution to obtain the pure 2-chloro-N-(2,4,5-trifluorophenyl)acetamide **9** in 92% yield; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta$  4.19 (s, CH<sub>2</sub>NH<sub>2</sub>), 6.99 (td, J = 9.96, 7.00 Hz, 1H, ArH), 8.22 (dt, J = 15.81, 8.78 Hz, 1H, ArH), 8.44 (br. 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  42.69, 105.01 (t, J =23.55 Hz), 110.30 (d, J = 24.81 Hz), 121.55 (t, J = 8.73 Hz), 146.07 (d, J = 249.56 Hz), 146.27 (dt, J = 247.84, 11.89 Hz), 147.73 (ddd, J = 243.43, 9.11, 2.73 Hz), 163.88. Physical and spectral data were consistent with those previously reported [51].

#### 4.1.6. Standard procedure for synthesis of 2-hydroxy-N-(2,4,5-

trifluorophenyl)acetamide 10 [29] 2-Chloro-N-(2,4,5-trifluorophenyl)acetamide 9 (1.0 equiv, 4.0 mmol) and cesium formate (HCO2Cs, 3.0 equiv) in dry EtOH (10 mL) was heated at reflux (ca. 77 °C) for 6 h. When the reaction was completed, the solution was filtered to remove the excess amount of HCO<sub>2</sub>Cs and the filtrate was concentrated under reduced pressure. corresponding crude 2-hydroxyacetamide **10**. The crude product **10** was recrystallized in EtOH/hexane (3/1) solution to obtain the pure 2hydroxyacetamide 10 in 86% yield; white solid; m.p. 89-90 °C, <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta$  2.93 (t, J = 5.15 Hz,1H, OH), 4.26 (d, J =5.15 Hz, 2H, CH<sub>2</sub>NH<sub>2</sub>), 6.98 (td, J = 10.03, 7.00 Hz, 1H, ArH), 8.29 (dt, J = 15.86, 5.95 Hz, 1H, ArH), 8.60 (br. 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125) MHz, CDCl<sub>3</sub>):  $\delta$  62.52, 104.95 (t, J = 23.53 Hz), 110.24 (d, J = 24.68Hz), 121.79 (t, J = 10.82 Hz), 145.84 (ddd, J = 249.12, 14.60, 11.75 Hz), 146.34 (ddd, J = 243.43, 12.72, 3.63 Hz), 147.57 (ddd, J = 243.43, 8.98, 2.70 Hz), 169.53. IR (KBr): 3429, 3374, 3286, 1688, 1552, 1437, 1199, 876 cm<sup>-1</sup>. EIMS m/z: 205 (M<sup>+</sup>, 32), 147 (100), 146 (18), 119 (21), 71 (19). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>8</sub>H<sub>6</sub>F<sub>3</sub>NO<sub>2</sub>: 205.0351; Found 205.0349.

## 4.1.7. Standard procedure for synthesis of 2-amino-N-(2,4,5-trifluorophenyl)acetamide 11 [30]

2-Chloro-N-(2,4,5-trifluorophenyl)acetamide 9 (1.0 equiv, 4.0 mmol) was added with ammonium hydroxide solution (NH<sub>4</sub>OH, 15 mL) in ethanol solution (15 mL). The mixture was heated at 60  $^\circ$ C under sealed conditions for 3 h. When the reaction was completed, the reaction mixture was concentrated and nertulized with 3% HCl(aq) and extracted with  $CH_2Cl_2$  (3  $\times$  30 mL). The organic extracts were dried over MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure to afford the corresponding crude 2-amino-N-(2,4,5-trifluorophenyl)acetamide 11. The crude product 11 was recrystallized in EtOH/hexane (3/1) solution to obtain the pure 2-aminoacetamide 11 in 88% yield; white solid; m.p. 69–70 °C, <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz): δ 3.44 (s, 2H, CH<sub>2</sub>NH<sub>2</sub>), 6.92 (q, J = 9.07 Hz, 1H, ArH), 8.30 (dt, J = 16.38, 5.89 Hz, 1H, ArH), 9.73 (s, 2H, NH<sub>2</sub>). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  44.98, 104.65 (t, J = 23.52Hz), 109.53 (d, J = 24.65 Hz), 122.47 (t, J = 8.89 Hz), 145.22 (dt, J = 247.25, 12.96 Hz), 146.17 (dd, J = 242.46, 9.04 Hz), 147.42 (dd, J = 245.87, 8.97 Hz), 171.00. IR (KBr): 3503, 3364, 1678, 1569, 1501, 1426, 1206, 862 cm<sup>-1</sup>. EIMS m/z: 205 (11), 204 (M<sup>+</sup>, 46), 173 (26), 148 (16), 147 (100), 146 (18), 145 (15), 81 (11), 69 (16). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>8</sub>H<sub>7</sub>F<sub>3</sub>N<sub>2</sub>O: 204.0510; Found 287.1163.

## 4.1.8. Standard procedure for synthesis of (1H-1,2,4-triazol-3-yl)methanol (12d, and 12m-q) [40]

The reliable procedure involved the treatment of 1*H*-1,2,4-triazole-3carboxylates **7d** and **7m–q** (1.0 equiv, 5.0 mmol) with NaBH<sub>4</sub> (1.5 equiv, 7.5 mmol) in THF/MeOH (1/3, 12 mL) [40]. The reaction mixture was stirred at room temperature for 5 h. When the reaction was completed, the reaction mixture was concentrated under reduced pressure to afford the crude residue. The residue was added water (15 mL) and extracted with ethyl acetate (15 mL × 3). The combined organic layers were washed sodium bicarbonate (15 mL × 3), dried over MgSO<sub>4</sub> to give crude (1*H*-1,2,4-triazol-3-yl)methanol product **12d** and **12m–q**. The crude desired products **12d** and **12m–q** were recrystallized in *n*-hexane/ methanol (1/4) solution to obtain the pure (1*H*-1,2,4-triazol-3-yl) methanol product **12d** and **12m–q** in 91–96% yields.

#### 4.1.8.1. (1-(4-(Trifluoromethyl)phenyl)-1H-1,2,4-triazol-3-yl)methanol

(12d). Yield 91%, white solid, m.p. 111–113 °C, <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  2.84 (t, J = 5.97 Hz, 1H, OH), 4.85 (d, J = 5.55 Hz, 2H, CH<sub>2</sub>OH), 7.76 (d, J = 8.79 Hz, 2H, ArH), 7.81 (d, J = 8.70 Hz, 2H, ArH), 8.59 (s, 1H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  58.44, 119.70 (2 × C), 123.17 (d, J = 272.03 Hz), 127.13, 127.17, 130.17 (q, J = 33.02 Hz), 139.27, 141.59, 164.97. IR (KBr): 3210, 3114, 2922, 2851, 1335, 1176,

1133, 1111, 1047, 848 cm<sup>-1</sup>. EIMS m/z: 243 (M<sup>+</sup>, 70), 242 (85), 224 (18), 214 (58), 187 (10), 172 (21), 159 (100), 145 (42), 140 (15), 139 (30), 109 (15). HRMS (EI) m/z: [M]<sup>+</sup> Calcd for C<sub>10</sub>H<sub>8</sub>F<sub>3</sub>N<sub>3</sub>O: 243.0619; Found 243.0625.

#### 4.1.8.2. (1-(4-Chlorophenyl)-5-cyclopentyl-1H-1,2,4-triazol-3-yl)meth-

anol (12m). Yield 94%, light yellow solid, m.p. 100–102 °C, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.59 (t, J = 5.35 Hz, 2H, CP), 1.83–1.95 (m, 6H, CP), 3.05 (quint, J = 8.18 Hz, 1H, CP), 3.41 (br, 1H, OH), 4.74 (s, 2H, CH<sub>2</sub>OH), 7.34 (d, J = 8.47 Hz, 2H, ArH), 7.46 (d, J = 8.50 Hz, 2H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  25.75 (2 × C), 32.96 (2 × C), 36.19, 58.29, 126.71 (2 × C), 129.64 (2 × C), 135.05, 135.82, 161.19, 162.83. IR (KBr): 3264, 2955, 2869, 1519, 1486, 1092, 1059, 1011, 838 cm<sup>-1</sup>. EIMS *m/z*: 279 (M<sup>+</sup> + 2, 5), 277 (M<sup>+</sup>, 14), 276 (14), 238 (24), 237 (10), 236 (77), 182 (18), 127 (32), 126 (13), 125 (100), 90 (22). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>14</sub>H<sub>16</sub>ClN<sub>3</sub>O: 277.0982; Found 277.0975.

4.1.8.3. (5-(1*H*-*Pyrrol*-2-*y*l)-1-(3-(trifluoromethyl)phenyl)-1*H*-1,2,4-triazol-3-*y*l) methanol (**12n**). Yield 92%, white solid, m.p. 183–184 °C, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  4.59 (br, 1H, OH), 4.83 (s, 1H, CH<sub>2</sub>OH), 5.81 (s, 1H, ArH), 6.09–6.11 (m, 1H, ArH), 6.92–6.93 (m, 1H, ArH), 7.65–7.71 (m, 2H, ArH), 7.80 (br, 2H, ArH), 10.01 (br, 2H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  57.99, 110.36, 111.07, 118.49, 121.74, 123.25 (d, *J* = 272.61 Hz), 123.73, 126.59, 129.84, 130.28, 132.43 (d, *J* = 33.19 Hz), 138.49, 149.38, 162.88. IR (KBr): 3317, 3128, 3114, 3054, 1499, 1327, 1161, 1112, 1070, 806, 753, 698 cm<sup>-1</sup>. EIMS *m/z*: 309 (19), 308 (M<sup>+</sup>, 93), 307 (40), 289 (53), 159 (100), 145 (14), 139 (11). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>14</sub>H<sub>11</sub>F<sub>3</sub>N<sub>4</sub>O: 308.0885; Found 308.8092.

#### 4.1.8.4. (5-Butyl-1-(4-chlorophenyl)-1H-1,2,4-triazol-3-yl)methanol

(120). Yield 93%, orange solid, m.p. 48–50 °C, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  0.85 (t, J = 7.35 Hz, 3H, CH<sub>3</sub>), 1.30 (sext, J = 7.44 Hz, 2H, CH<sub>2</sub>CH<sub>3</sub>), 1.69 (q, J = 7.68 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 2.72 (t, J = 7.78 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 3.97 (br, 1H, OH), 4.74 (s, 2H, CH<sub>2</sub>OH), 7.33 (dt, J = 9.06, 2.60 Hz, 2H, ArH), 7.45 (dt, J = 9.45, 2.35 Hz, 2H, ArH). <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  13.56, 22.24, 26.15, 29.72, 57.99, 126.32 (2 × C), 129.66 (2 × C), 134.98, 135.70, 157.14, 162.98. IR (KBr): 3292, 2958, 2931, 1520, 1487, 1092, 1055, 1012, 837 cm<sup>-1</sup>. EIMS *m/z*: 265 (M<sup>+</sup>, 5), 236 (18), 225 (32), 224 (12), 223 (100), 182 (13), 170 (35), 127 (22), 125 (54), 90 (19). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>13</sub>H<sub>16</sub>ClN<sub>3</sub>O: 265.0982; Found 265.0989.

4.1.8.5. (1,5-Diphenyl-1H-1,2,4-triazol-3-yl)methanol (12p). Yield 96%, light yellow solid, m.p. 137–139 °C, <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.70 (br, 1H, OH), 4.84 (s, 2H, CH<sub>2</sub>OH), 7.29–7.33 (m, 4H, ArH), 7.36–7.40 (m, 4H, ArH), 7.45–7.47 (m, 2H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  58.22, 125.33 (2 × C), 127.45, 128.59 (2 × C), 128.85 (2 × C), 128.96, 129.40 (2 × C), 130.17, 137.93, 154.65, 163.24. IR (KBr): 3307, 3061, 2922, 1595, 1511, 1449, 1050, 990, 774, 694 cm<sup>-1</sup>. EIMS *m/z*: 251 (M<sup>+</sup>, 46), 148 (36), 91 (100), 64 (11). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>15</sub>H<sub>13</sub>N<sub>3</sub>O: 251.1059; Found 251.1056.

4.1.8.6. (5-Phenyl-1-(p-tolyl)-1H-1,2,4-triazol-3-yl)methanol (12q). Yield 96%, light yellow solid, m.p. 112–114 °C, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  2.38 (s, 3H, CH<sub>3</sub>), 4.83 (s, 2H, CH<sub>2</sub>OH), 7.20 (s, 4H, ArH), 7.32 (t, J = 7.43 Hz, 2H, ArH), 7.38 (t, J = 7.35 Hz, 1H, ArH), 7.46–7.48 (m, 2H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  21.18, 58.55, 125.18 (2 × C), 127.67, 128.56 (2 × C), 128.83 (2 × C), 129.99 (2 × C), 130.07, 135.55, 139.13, 154.57, 162.99. IR (KBr): 3209, 3039, 2922, 2858, 1519, 1453, 1050, 992, 825, 778, 696 cm<sup>-1</sup>. EIMS *m/z*: 266 (19), 265 (M<sup>+</sup>, 91), 162 (59), 106 (12), 105 (100), 104 (29). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>16</sub>H<sub>15</sub>N<sub>3</sub>O: 265.1215; Found 265.1210.

## 4.1.9. Standard procedure for synthesis of (1H-1,2,4-triazol-3-yl) methanamine (13d, and 13m-q) [41,42]

The reliable procedure involved the treatment of 1H-1,2,4-triazol-3ylmethanol 12d, and 12m-q (1.0 equiv, 5.0 mmol) with thionyl chloride (SOCl<sub>2</sub>, 1.5 equiv, 7.5 mmol) in CH<sub>2</sub>Cl<sub>2</sub> solution (15 mL) [41]. The reaction mixture was stirred at room temperature for 2 h under N2. When the reaction was completed, the reaction mixture was concentrated under reduced pressure, added with CH<sub>2</sub>Cl<sub>2</sub> (15 mL), neutralized with saturated sodium bicarbonate solution (15 mL  $\times$  3). The organic layer was dried over MgSO<sub>4</sub>, filtered, then concentrated under reduced pressure to afford crude residue. The residue was added with ammonium hydroxide solution (NH<sub>4</sub>OH, 15 mL) in methanol solution (15 mL). The mixture was heated at 60 °C under sealed conditions for 6.0 h [42]. When the amination reaction was completed, the reaction mixture was concentrated and nertulized with 3% HCl<sub>(aq)</sub> and extracted with CH<sub>2</sub>Cl<sub>2</sub>  $(3 \times 30 \text{ mL})$ . The organic extracts were dried over MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure to afford the corresponding crude 1H-1,2,4-triazol-3-ylmethanamines 13d and 13m-q. The crude products 13d and 13m-q were purified by column chromatography on silica gel to give the pure corresponding  $\beta$ -amino esters **13d** and **13m–q** in 73-84% vields.

#### 4.1.9.1. (1-(4-(Trifluoromethyl)phenyl)-1H-1,2,4-triazol-3-yl)methan-

*amine* (13*d*). Yield 73%, light yellow solid, m.p. 75–77 °C <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.98 (s, 2H, NH<sub>2</sub>), 3.98 (s, 2H, CH<sub>2</sub>NH<sub>2</sub>), 7.69 (d, J = 7.90 Hz, 2H, ArH), 7.75 (d, J = 7.85 Hz, 2H, ArH), 8.53 (s, 1H, ArH). <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  39.76, 119.42 (2 × C), 123.42 (d, J = 272.15 Hz), 126.94, 126.97, 129.74 (q, J = 33.40 Hz), 139.37, 141.39, 166.82. IR (KBr): 3370, 3306, 3093, 2926, 2858, 1618, 1534, 1324, 1125, 983, 844 cm<sup>-1</sup>. EIMS *m/z*: 242 (M<sup>+</sup>, 54), 241 (100), 214 (77), 172 (12), 145 (14). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>10</sub>H<sub>9</sub>F<sub>3</sub>N<sub>4</sub>: 242.0779; Found 242.0770.

4.1.9.2. (1-(4-Chlorophenyl)-5-cyclopentyl-1H-1,2,4-triazol-3-yl)methanamine (13m). Yield 79%, orange liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.37–1.42 (m, 2H, CP), 1.64–1.77 (m, 6H, CP), 2.09 (br, 2H, NH<sub>2</sub>), 2.89 (quint, J = 8.24 Hz, 1H, CP), 3.76 (br, 2H, CH<sub>2</sub>NH<sub>2</sub>), 7.18 (d, J = 8.79 Hz, 2H, ArH), 7.27 (d, J = 8.65 Hz, 2H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  25.31 (2 × C), 32.51 (2 × C), 35.76, 39.37, 126.22 (2 × C), 129.12 (2 × C), 134.28, 135.60, 160.42, 163.98; IR (KBr): 3372, 3302, 3050, 2957, 2870, 1513, 1487, 1092, 1012, 835 cm<sup>-1</sup>. EIMS *m*/*z*: 278 (M<sup>+</sup> + 2, 26), 277 (30), 276 (M<sup>+</sup>, 92), 275 (57), 250 (15), 248 (53), 237 (27), 235 (100), 224 (12), 218 (20), 182 (14), 180 (45), 152 (22), 127 (20), 125 (62), 111 (13), 90 (15). HRMS (EI) *m*/*z*: [M]<sup>+</sup> Calcd for C<sub>14</sub>H<sub>17</sub>ClN<sub>4</sub>: 276.1142; Found 2776.1147.

4.1.9.3. (5-(1*H*-*Pyrrol*-2-*y*])-1-(3-(trifluoromethyl)phenyl)-1*H*-1,2,4-triazol-3-*y*]) methanamine (**13n**). Yield 76%, light yellow liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  2.65 (br, 2H, NH<sub>2</sub>), 4.00 (br, 2H, CH<sub>2</sub>NH<sub>2</sub>), 5.80 (d, J = 2.90 Hz, 1H, ArH), 6.05 (s, 1H, ArH), 6.87 (s, 1H, ArH), 7.60 (t, J =7.85 Hz, 1H, ArH), 7.67 (d, J = 7.85 Hz, 1H, ArH), 7.75 (d, J = 7.70 Hz, 1H, ArH), 7.79 (br, 1H, ArH), 11.39 (s, 1H, NH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  39.18, 19.84, 110.72, 118.62, 121.55, 123.18 (d, J =272.61 Hz), 123.53, 126.18, 129.67, 130.05, 132.10 (q, J = 33.37 Hz), 138.55, 149.37, 164.59. IR (KBr): 3356, 3292, 2922, 2855, 1597, 1508, 1328, 1126, 1069, 748, 697 cm<sup>-1</sup>. EIMS *m*/*z*: 308 (19), 307 (M<sup>+</sup>, 100), 306 (53), 289 (26), 279 (20), 237 (12), 222 (18), 221 (13), 145 (12). HRMS (EI) *m*/*x*: [M]<sup>+</sup> Calcd for C<sub>14</sub>H<sub>12</sub>F<sub>3</sub>N<sub>5</sub>: 307.1045; Found 307.1039.

4.1.9.4. (5-Butyl-1-(4-chlorophenyl)-1H-1,2,4-triazol-3-yl)methanamine (130). Yield: 81%, orange liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  0.80 (t, *J* = 7.35 Hz, 3H, CH<sub>3</sub>), 1.25 (sext, *J* = 7.40 Hz, 2H, CH<sub>2</sub>CH<sub>3</sub>), 1.63 (q, *J* = 7.67 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 2.65 (t, *J* = 7.83 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 3.05 (br, 1H, NH<sub>2</sub>), 3.92 (br, 2H, CH<sub>2</sub>NH<sub>2</sub>), 7.29 (d, *J* = 8.70 Hz, 2H, ArH), 7.39 (d, *J* = 8.65 Hz, 2H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  13.46, 22.16, 26.10, 29.68, 39.28, 126.14 (2 × C), 129.48 (2 × C), 134.61, 135.75, 156.82, 163.61. IR (KBr): 3406, 3296, 3046, 2959, 2930, 2630, 1504, 1488, 1092, 1012, 836 cm<sup>-1</sup>. EIMS *m/z*: 266 (M<sup>+</sup> + 2, 19), 265 (26), 264 (57), 263 (53), 249 (19), 238 (19), 237 (15), 236 (59), 235 (21), 224 (33), 223 (25), 222 (100), 205 (25), 191 (32), 182 (15), 180 (40), 170 (63), 152 (28), 130 (21), 129 (18), 128 (64), 127 (62), 125 (94), 111 (25), 90 (31). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for  $C_{13}H_{17}ClN_4$ : 264.1142; Found 264.1134.

4.1.9.5. (1,5-Diphenyl-1H-1,2,4-triazol-3-yl)methanamine (**13p**).. Yield 82%, yellow liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.94 (s, 2H, NH<sub>2</sub>), 3.99 (s, 2H, CH<sub>2</sub>NH<sub>2</sub>), 7.25–7.30 (m, 4H, ArH), 7.31–7.36 (m, 4H, ArH), 7.41–7.43 (m, 2H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  39.62, 124.93 (2 × C), 127.53, 128.18(2 × C), 128.39, 128.44 (2 × C), 128.99 (2 × C), 129.61, 137.81, 154.06, 164.78. IR (KBr): 3371, 3303, 3064, 2923, 2851, 1596, 1508, 1448, 989, 770, 694 cm<sup>-1</sup>. EIMS *m/z*: 251 (19), 250 (M<sup>+</sup>, 100), 249 (69), 235 (35). 222 (53), 180 (30), 146 (31), 118 (19), 104 (14), 103 (14), 92 (12), 91 (62). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>15</sub>H<sub>14</sub>N<sub>4</sub>: 250.1218; Found 250.1223.

4.1.9.6. (5-Phenyl-1-(p-tolyl)-1H-1,2,4-triazol-3-yl)methanamine (**13**q). Yield 84%, yellow liquid, <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  2.09 (br, 2H, NH<sub>2</sub>), 2.37 (s, 3H, CH<sub>3</sub>), 4.03 (s, 2H, CH<sub>2</sub>NH<sub>2</sub>), 7.18 (d, J = 8.43 Hz, 2H, ArH), 7.21 (d, J = 8.52 Hz, 2H, ArH), 7.31 (t, J = 7.39 Hz, 2H, ArH), 7.36 (t, J = 7.33 Hz, 1H, ArH), 749 (dd, J = 8.28, 1.10 Hz, 2H, ArH). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  20.82, 39.58, 124.80 (2 × C), 127.61, 128.17 (2 × C), 128.45 (2 × C), 129.57, 129.60 (2 × C), 135.36, 138.53, 154.00, 164.54. IR (KBr): 3367, 3292, 3068, 2923, 2853, 1517, 1451, 990, 821, 696 cm<sup>-1</sup>. EIMS *m/z*: 265 (24), 264 (M<sup>+</sup>, 100), 263 (56), 249 (31), 236 (49), 194 (24), 160 (29), 132 (27), 106(13), 105 (87), 104 (32). HRMS (EI) *m/z*: [M]<sup>+</sup> Calcd for C<sub>16</sub>H<sub>16</sub>N<sub>4</sub>: 264.1375; Found 264.1385.

#### 4.2. In vitro dipeptidyl peptidase 4 assay

#### 4.2.1. Preparation of the DPP-4 enzyme

Human recombinant dipeptidyl peptidase IV and its substrate, Gly-Pro-7-amido-4-methylcoumarin (GP- AMC) and its natural inhibitor, Ile-Pro-Ile tripeptide (diprotin A), were purchased from R&D Systems (Minneapolis, MN). All other chemicals were purchased from Sigma-Aldrich (St. Louis, MO).

#### 4.2.2. Enzyme-based assay of DPP-4 [24]

The reaction was carried out at pH 7.5 in the presence of 50 mM HEPES, 140 mM NaCl, 10 mM KCl, and 15  $\mu$ M of G-P-AMC (the Km for substrate G-P-AMC is about 15  $\mu$ M). The reaction volume is 200  $\mu$ l with 20 ng of recombinant DPP-4 at 25 °C. The reaction rate was monitored by a fluorometer with 380-nm excitation and 460-nm emission filters, and determined by the linear slope of the fluorescence accumulation. For inhibition kinetic, substrate and inhibitor concentrations varied and encompassed the Km and Ki values, respectively, and the data were analyzed by the EZ-FIT program, which uses a nonlinear regression method to obtain kinetic parameters. Inhibition constants (IC<sub>50</sub>) were calculated from enzyme progress curves using standard mathematical models.

#### 4.3. In vitro QPP, DPP-8 and DPP-9 assay [52]

DPP-7, DPP-8 and DPP-9 inhibitions were measured using fluorogenic DPP-7 assay kit, fluorogenic DPP-8 assay kit, and fluorogenic DPP-9 assay kit (BPS Bioscience, San Diego, CA, USA) following the manufacturer's instructions. Briefly, solutions of test compounds in varying concentrations were prepared in dimethyl sulfoxide (DMSO) and then diluted into assay buffer. Human recombinant DPP-7, DPP-8, or DPP-9 was added to the dilutions and pre-incubated for 10 min at 25 °C before the reaction was initiated by the addition of fluorogenic DPP substrate (AMC). The reaction rate was monitored by a fluorometer with 380-nm excitation and 460-nm emission filters, and determined by the linear slope of the fluorescence accumulation. Inhibition constants (IC<sub>50</sub>) were calculated from enzyme progress curves using standard mathematical models.

#### 4.4. Molecular modeling [12,25]

Computational studies were performed with Sitagliptin 1 and  $\beta$ -amino carbonyl target 5n, 6d, and 6p. The lowest energy conformer for chemical structure was saved in mol2 files by Chemdraw before use in docking studies. The structure of DPP-4 enzyme, encoded PDB ID: 1X70 was downloaded from the protein data bank (PDB) before performing docking studies [12]. Molecular docking studies were performed using iGemdock 2.1 [25]. iGemdock docking calculations were performed at the drug screening docking accuracy setting with parameters set for population size, generation, and number of solutions as 800, 80, and 10, respectively, and a Gemdock score function of hydrophobic and electrostatic (1:1 preference). Residue interaction figure was produced by Pymol to demonstrate the binding pocket.

#### **Declaration of Competing Interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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#### Appendix A. Supplementary material

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#### References

- [1] A.G. Almagthali, E.H. Alkhaldi, A.S. Alzahrani, A.K. Alghamdi, W.Y. Alghamdi, A. M. Kabel, Dipeptidyl peptidase-4 inhibitors: Anti-diabetic drugs with potential effects on cancer, Diabetes Metab. Syndr. Clin. Res. Rev. 13 (2019) 36–39, https://doi.org/10.1016/j.dsx.2018.08.012.
- [2] L. Mendieta, T. Tarrago, E. Giralt, Recent patents of dipeptidyl peptidase IV inhibitors, Expert Opin. Ther. Pat. 21 (2011) 1693–1741, https://doi.org/10.1517/ 13543776.2011.627325.
- [3] Y. Liu, Y. Hu, T. Liu, Recent Advances in Non-Peptidomimetic Dipeptidyl Peptidase 4 Inhibitors: Medicinal Chemistry and Preclinical Aspects, Curr. Med. Chem. 19 (2012) 3982–3999, https://doi.org/10.2174/092986712802002491.
- [4] Y. Liu, M. Si, L. Tang, S. Shangguan, H. Wu, J. Li, P. Wu, X. Ma, T. Liu, Y. Hu, Synthesis and biological evaluation of novel benzyl-substituted (S)-phenylalanine derivatives as potent dipeptidyl peptidase 4 inhibitors, Bioorg. Med. Chem. 21 (2013) 5679–5687, https://doi.org/10.1016/j.bmc.2013.07.034.
- [5] A.M. Kabel, M.S. Omar, A. Alhadhrami, S.S. Alharthi, M.M. Alrobaian, Linagliptin potentiates the effect of I-dopa on the behavioural, biochemical and immunohistochemical changes in experimentally-induced Parkinsonism: Role of toll-like receptor 4, TGF-β1, NF-κB and glucagon-like peptide 1, Physiol. Behav. 188 (2018) 108–118, https://doi.org/10.1016/j.physbeh.2018.01.028.
- [6] T.J. Kieffer, J. Francis Habener, The Glucagon-Like Peptides, Endocr. Rev. 20 (1999) 876–913, https://doi.org/10.1210/edrv.20.6.0385.
- [7] R. Soni, S.S. Soman, Design and synthesis of aminocoumarin derivatives as DPP-IV inhibitors and anticancer agents, Bioorg. Chem. 79 (2018) 277–284, https://doi. org/10.1016/j.bioorg.2018.05.008.
- [8] S.S. Abd El-Karim, M.M. Anwar, Y.M. Syam, M.A. Nael, H.F. Ali, M.A. Motaleb, Rational design and synthesis of new tetralin-sulfonamide derivatives as potent anti-diabetics and DPP-4 inhibitors: 2D & 3D QSAR, in vivo radiolabeling and bio distribution studies, Bioorg. Chem. 81 (2018) 481–493, https://doi.org/10.1016/j. bioorg.2018.09.021.
- [9] M. Xie, M. Zhu, C.-M. Lu, Y. Jin, L.-H. Gao, L. Li, J. Zhou, F. Li, Q.H. Zhao, H.-K. Liu, P.J. Sadler, C. Sanchez-Cano, Synthesis and characterization of

oxidovanadium complexes as enzyme inhibitors targeting dipeptidyl peptidase IV, J. Inorg. Biochem. 175 (2017) 29–35, https://doi.org/10.1016/j. jinorgbio.2017.06.014.

- [10] X. Deng, J. Shen, H. Zhu, J. Xiao, R. Sun, F. Xie, C. Lam, J. Wang, Y. Qiao, M. S. Tavallaie, Y. Hu, Y. Du, J. Li, L. Fu, F. Jiang, Surrogating and redirection of pyrazolo[1,5-a]pyrimidin-7(4H)-one core, a novel class of potent and selective DPP-4 inhibitors, Bioorg. Med. Chem. 26 (2018) 903–912, https://doi.org/10.1016/j.bmc.2018.01.006.
- [11] M. Zhu, J. Zhou, Y. Jin, L.-H. Gao, L. Li, J.-R. Yang, C.-M. Lu, Q.H. Zhao, M. Xie, A manganese-salen complex as dipeptidyl peptidase IV inhibitor for the treatment of type 2 diabetes, Int. J. Biol. Macromol. 120 (2018) 1232–1239, https://doi.org/ 10.1016/j.ijbiomac.2018.08.089.
- [12] D. Kim, L. Wang, M. Beconi, G.J. Eiermann, M.H. Fisher, H. He, G.J. Hickey, J. E. Kowalchick, B. Leiting, K. Lyons, F. Marsilio, M.E. McCann, R.A. Patel, A. Petrov, G. Scapin, S.B. Patel, R.S. Roy, J.K. Wu, M.J. Wyvratt, B.B. Zhang, L. Zhu, N. A. Thornberry, A.E. Weber, (2R)-4-0xo-4-[3-(Trifluoromethyl)-5,6-dihydro[1,2,4] triazolo[4,3-a]pyrazin-7(8H)-yl]-1(2,4,5-trifluorophenyl)butan-2-amine: A Potent, Orally Active Dipeptidyl Peptidase IV Inhibitor for the Treatment of Type 2 Diabetes, J. Med. Chem. 48 (2005) 141–151, https://doi.org/10.1021/jm0493156.
- [13] B. Gallwitz, Small molecule dipeptidylpeptidase IV inhibitors under investigation for diabetes mellitus therapy, Expert Opin. Investig. Drugs. 20 (2011) 723–732, https://doi.org/10.1517/13543784.2011.576667.
- [14] T. Yoshida, F. Akahoshi, H. Sakashita, H. Kitajima, M. Nakamura, S. Sonda, M. Takeuchi, Y. Tanaka, N. Ueda, S. Sekiguchi, T. Ishige, K. Shima, M. Nabeno, Y. Abe, J. Anabuki, A. Soejima, K. Yoshida, Y. Takashina, S. Ishii, S. Kiuchi, S. Fukuda, R. Tsutsumiuchi, K. Kosaka, T. Murozono, Y. Nakamaru, H. Utsumi, N. Masutomi, H. Kishida, I. Miyaguchi, Y. Hayashi, Discovery and preclinical profile of teneligliptin (3-[(2S,4S)-4-[4-(3-methyl-1-phenyl-1*H*-pyrazol-5-yl) piperazin-1-yl]pyrrolidin-2-ylcarbonyl]thiazolidine): A highly potent, selective, long-lasting and orally active dipeptidyl peptidase IV inhibitor for t, Bioorg. Med. Chem. 20 (2012) 5705-5719, https://doi.org/10.1016/j.bmc.2012.08.012.
- [15] O. Gutierrez, D. Metil, N. Dwivedi, N. Gudimalla, E.R.R. Chandrashekar, V. H. Dahanukar, A. Bhattacharya, R. Bandichhor, M.C. Kozlowski, Practical, Asymmetric Route to Sitagliptin and Derivatives: Development and Origin of Diastereoselectivity, Org. Lett. 17 (2015) 1742–1745, https://doi.org/10.1021/ acs.orglett.5b00520.
- [16] (a) K.B. Hansen, Y. Hsiao, F. Xu, N. Rivera, A. Clausen, M. Kubryk, S. Krska, T. Rosner, B. Simmons, J. Balsells, N. Ikemoto, Y. Sun, F. Spindler, C. Malan, E.J.J. Grabowski, J.D. Armstrong, Highly Efficient Asymmetric Synthesis of Sitagliptin, J. Am. Chem. Soc. 131 (2009) 8798–8804. https://doi.org/10.1021/ja902462q. (b) Y. Xiao, J.D.III Armstrong, S.W. Krska, E. Njolito, N.R. Rivera, Y. Sun, T. Rosner, A. M. Clausen, Process to Chiral beta Amino Acid Derivatives by Asymmetric Hydrogenation, WO 2006/081151 A1.
- [17] (a) F. Liu, W. Yu, W. Ou, X. Wang, L. Ruan, Y. Li, X. Peng, X. Tao, X. Pan, The asymmetric synthesis of Sitagliptin, a selective dipeptidyl peptidase IV inhibitor for the treatment of type 2 diabetes, J. Chem. Res. 2010 (2010) 230–232, https://doi. org/10.3184/030823410X12709912414009;

(b) S.G. Davies, A.M. Fletcher, L. Lv, P.M. Roberts, J.E. Thomson, Asymmetric synthesis of (-)-(R)-sitagliptin, Tetrahedron Lett. 53 (2012) 3052–3055. https ://doi.org/10.1016/j.tetlet.2012.04.025;

(c) L.L. Zeng, Y.J. Ding, G.C. Zhang, H.R. Song, W.H. Hu, A practical synthesis of trifluorophenyl R-amino acid: The key precursor for the new anti-diabetic drug sitagliptin, Chinese Chem. Lett. 20 (2009) 1397–1399. https://doi.org/10.1016/j. cclet.2009.06.036.

- [18] A.A. Desai, Sitagliptin Manufacture: A Compelling Tale of Green Chemistry, Process Intensification, and Industrial Asymmetric Catalysis, Angew. Chemie Int. Ed. 50 (2011) 1974–1976, https://doi.org/10.1002/anie.201007051.
  [19] (a) C.K. Savile, J.M. Janey, E.C. Mundorff, J.C. Moore, S. Tam, W.R. Jarvis, J.C.
- [19] (a) C.K. Savile, J.M. Janey, E.C. Mundorff, J.C. Moore, S. Tam, W.R. Jarvis, J.C. Colbeck, A. Krebber, F.J. Fleitz, J. Brands, P.N. Devine, G.W. Huisman, G.J. Hughes, Biocatalytic Asymmetric Synthesis of Chiral Amines from Ketones Applied to Sitagliptin Manufacture, Science. 329 (2010) 305–309. https://doi.org/10.1126/science.1188934. (b) G. Hughes, P.N. Devine, F.J. Fleitz, B.T. Grau, J. Limanto, C. Savile, E. Mundorff, Transaminase Reactions, WO 2011/005477 A1.
- [20] (a) X. Deng, N. Wang, L. Meng, S. Zhou, J. Huang, J. Xing, L. He, W. Jiang, Q. Li, Optimization of the Benzamide Fragment Targeting the S2' Site Leads to Potent Dipeptidyl Peptidase-IV Inhibitors, Bioorg. Chem. 94 (2020), 103366, https://doi. org/10.1016/j.bioorg.2019.103366;

(b) G. Schnapp, T. Klein, Y. Hoevels, R.A. Bakker, H. Nar, Comparative Analysis of Binding Kinetics and Thermodynamics of Dipeptidyl Peptidase-4 Inhibitors and Their Relationship to Structure, J. Med. Chem. 59 (2016) 7466–7477. https://doi. org/10.1021/acs.jmedchem.6b00475.

- [21] M. Nabeno, F. Akahoshi, H. Kishida, I. Miyaguchi, Y. Tanaka, S. Ishii, T. Kadowaki, A comparative study of the binding modes of recently launched dipeptidyl peptidase IV inhibitors in the active site, Biochem. Biophys. Res. Commun. 434 (2013) 191–196, https://doi.org/10.1016/j.bbrc.2013.03.010.
- [22] (a) R. Sharma, S.S. Soman, Design and synthesis of novel diamide derivatives of glycine as antihyperglycemic agents, Synth. Commun. 46 (2016) 1307–1317, https://doi.org/10.1080/00397911.2016.1203435;

(b) Z.-P. Xiao, W. Wei, P.-F. Wang, W.-K. Shi, N. Zhu, M.-Q. Xie, Y.-W. Sun, L.-X. Li, Y.-X. Xie, L.-S. Zhu, N. Tang, H. Ouyang, X.-H. Li, G.-C. Wang, H.-L. Zhu, Synthesis and evaluation of new tyrosyl-tRNA synthetase inhibitors as antibacterial agents based on a N2-(arylacetyl)glycianilide scaffold, Eur. J. Med. Chem. 102 (2015) 631–638. https://doi.org/10.1016/j.ejmech.2015.08.025.

[23] (a) R. Soni, S.D. Durgapal, S.S. Soman, J.J. Georrge, Design, synthesis and antidiabetic activity of chromen-2-one derivatives, Arab. J. Chem. 12 (2019) 701–708, https://doi.org/10.1016/j.arabjc.2016.11.011; (b) L. Wu, K. Lu, M. Packiarajan, V. Jubian, G. Chandrasena, T.C. Wolinsky, M. W. Walker, Indolyl and dihydroindolyl N-glycinamides as potent and in vivo active NPY5 antagonists, Bioorg. Med. Chem. Lett. 22 (2012) 2167–2171. https://doi.org/10.1016/j.bmcl.2012.01.117.

- [24] S.-H. Lu, W.-P. Yen, H.J. Tsai, C.-S. Chen, F.F. Wong, Vilsmeier reagent initialed sequential one-pot multicomponent synthesis of N, O-disubstituted glycolamides as dipeptidyl peptidase 4 inhibitors, Tetrahedron. 71 (2015) 6749–6758, https://doi. org/10.1016/j.tet.2015.07.041.
- [25] J.-M. Yang, C.-C. Chen, GEMDOCK: A generic evolutionary method for molecular docking, Proteins Struct. Funct. Bioinforma. 55 (2004) 288–304, https://doi.org/ 10.1002/prot.20035.
- [26] (a) L.-Y. Wang, H.J. Tsai, H.-Y. Lin, K. Kaneko, F.-Y. Cheng, H.-S. Shih, F.F. Wong, J.-J. Huang, One-flask synthesis of 1,3,5-trisubstituted 1,2,4-triazoles from nitriles and hydrazonoyl chlorides via 1,3-dipolar cycloaddition, RSC Adv. 4 (2014) 14215–14220, https://doi.org/10.1039/C4RA00113C;
  (b) W.-C. Tseng, L.-Y. Wang, T.-S. Wu, F.F. Wong, 'One-flask' synthesis to 3,5-disubstituted 1,2,4-triazoles from aldehydes with hydrazonoyl hydrochlorides via 1,3-dipolar cycloaddition, Tetrahedron. 67 (2011) 5339–5345. https://doi.org/10.1016/j.tet.2011.05.003;
  (c) S.-E. Tsai, K.-H. Chiang, C.-C. Tseng, N.-W. Chen, C.-Y. Chern, F.F. Wong, Facile One-Pot Synthesis of Methyl 1-Aryl-1 H -1,2,4-triazole-3-carboxylates from Nitrilimines with Vilsmeier Reagent, European J. Org. Chem. 2019 (2019)
- 1754–1762. https://doi.org/10.1002/ejoc.201801808.
  [27] J.E. Kowalchick, B. Leiting, K.D. Pryor, F. Marsilio, J.K. Wu, H. He, K.A. Lyons, G. J. Eiermann, A. Petrov, G. Scapin, R.A. Patel, N.A. Thornberry, A.E. Weber, D. Kim, Design, synthesis, and biological evaluation of triazolopiperazine-based β-amino amides as potent, orally active dipeptidyl peptidase IV (DPP-4) inhibitors, Bioorg. Med. Chem. Lett. 17 (2007) 5934–5939, https://doi.org/10.1016/j. bmcl.2007.07.100.
- [28] (a) B. Dyck, V.S. Goodfellow, T. Phillips, J. Grey, M. Haddach, M. Rowbottom, G. S. Naeve, B. Brown, J. Saunders, Potent imidazole and triazole CB 1 receptor antagonists related to SR141716, Bioorg. Med. Chem. Lett. 14 (2004) 1151–1154, https://doi.org/10.1016/j.bmcl.2003.12.068;
  (b) H.W. Horn, G.O. Jones, D.S. Wei, K. Fukushima, J.M. Lecuyer, D.J. Coady, J. L. Hedrick, J.E. Rice, Mechanisms of Organocatalytic Amidation and Trans-Esterification of Aromatic Esters As a Model for the Depolymerization of Poly (ethylene) Terephthalate, J. Phys. Chem. A. 116 (2012) 12389–12398. https://doi.org/10.1021/jp304212y.
- [29] F.F. Wong, P.-W. Chang, H.-C. Lin, B.-J. You, J.-J. Huang, S.-K. Lin, An efficient and convenient transformation of α-haloketones to α-hydroxyketones using cesium formate, J. Organomet. Chem. 694 (2009) 3452–3455, https://doi.org/10.1016/j. jorganchem.2009.06.031.
- [30] W. Lin, L. Yang, S.C. Chai, Y. Lu, T. Chen, Development of CINPA1 analogs as novel and potent inverse agonists of constitutive androstane receptor, Eur. J. Med. Chem. 108 (2016) 505–528, https://doi.org/10.1016/j.ejmech.2015.12.018.
- [31] (a) J.-J. Huang, S.-H. Lu, Y.H. Chung, F.F. Wong, Vilsmeier-Haack reagent-promoted formyloxylation of α-chloro-N-arylacetamides by formamide, RSC Adv. 5 (2015) 35934–35939, https://doi.org/10.1039/C5RA05779E;
  (b) S.-H. Lu, P.-L. Liu, F.F. Wong, Vilsmeier reagent-mediated synthesis of 6-[[formyloxy)methyl]-pyrazolopyrimidines via a one-pot multiple tandem reaction, RSC Adv. 5 (2015) 47098–47107. https://doi.org/10.1039/C5RA07707A.
- [32] Y.J. Pu, R.K. Vaid, S.K. Boini, R.W. Towsley, C.W. Doecke, D. Mitchell, A Practical Method for Functionalized Peptide or Amide Bond Formation in Aqueous–Ethanol Media with EDC as Activator, Org. Process Res. Dev. 13 (2009) 310–314, https:// doi.org/10.1021/op800240d.
- [33] P. Mampuys, E. Ruijter, R.V.A. Orru, B.U.W. Maes, Synthesis of Secondary Amides from Thiocarbamates, Org. Lett. 20 (2018) 4235–4239, https://doi.org/10.1021/ acs.orglett.8b01654.
- [34] (a)R. Sustmann, Synthesis of Acid Halides, Anhydrides and Related Compounds, in: B. M. Trost, I. Fleming (Eds.), Comprehensive Organic Synthesis, Pergamon Press, Oxford, 1991, Vol. 6, pp. 301–318.
  (b)N. Teno, K. Wanaka, Y. Okada, Y. Tsuda, U. Okamoto, A. Hyikata, A. Okunomiya, T. Naito, S. Okamoto, S., Development of Active Center-Directed Inhibitors against Plasmin., Chem. Pharm. Bull. (Tokyo). 39 (1991) 2340–2346. https://doi.org/10.1248/cpb.39.2340.
- [35] S. Ohta, A. Shimabayashi, M. Aono, M. Okamoto, A General Convenient One-Pot Procedure for the Conversion of Carboxylic Acids into their t-Butyl Esters which is also Applicable to Aliphatic Carboxylic Acids, Synthesis (Stuttg). 1982 (1982) 833–834, https://doi.org/10.1055/s-1982-29961.
- [36] E. Turos, B. Bhattacharya, Antibiotic Compositions and Methods of use US 2017/ 9670179 B11.
- [37] E. DeVita, P. Schüler, S. Lovell, J. Lohbeck, S. Kullmann, E. Rabinovich, A. Sananes, B. Heßling, V. Hamon, N. Papo, J. Hess, E.W. Tate, N. Gunkel, A.K. Miller, Depsipeptides Featuring a Neutral P1 Are Potent Inhibitors of Kallikrein-Related Peptidase 6 with On-Target Cellular Activity, J. Med. Chem. 61 (2018) 8859–8874, https://doi.org/10.1021/acs.jmedchem.8b01106.
- $[38] E. Frérot, J. Coste, A. Pantaloni, M.-N. Dufour, P. Jouin, PyBOP® and PyBroP: Two reagents for the difficult coupling of the <math display="inline">\alpha$ ,  $\alpha$ -dialkyl amino acid, Aib, Tetrahedron. 47 (1991) 259–270, https://doi.org/10.1016/S0040-4020(01)80922-4.
- [39] J. Diago-Meseguer, A.L. Palomo-Coll, J.R. Fernández-Lizarbe, A. Zugaza-Bilbao, A New Reagent for Activating Carboxyl Groups; Preparation and Reactions of N, N -Bis[2-oxo-3-ox-azolidinyl]phosphorodiamidic Chloride, Synthesis (Stuttg). 1980 (1980) 547–551, https://doi.org/10.1055/s-1980-29116.
- [40] N.D. Yadav, R.S. Bhide, R.O. Bora, P. Gunaga, M. Panda, E.S. Priestley, J. Richter, Substituted Nitrogen containing Compounds, WO 2018/222795 A1.

- [41] R.G. Jones, C. Ainsworth, 1,2,4-Triazole-3-alanine, J. Am. Chem. Soc. 77 (1955) 1538–1540, https://doi.org/10.1021/ja01611a040.
- [42] A. Madin, A.P. Owens, Triazolo-pyridazine Derivatives as Ligands for Gaba Receptors, WO 1999/037645 A1.
- [43] E. Gunic, S. Chow, F. Rong, K. Ramasamy, A. Raney, D. Yunzhi Li, J. Huang, R. K. Hamatake, Z. Hong, J.-L. Girardet, 6-Hydrazinopurine 2'-methyl ribonucleosides and their 5'-monophosphate prodrugs as potent hepatitis C virus inhibitors, Bioorg. Med. Chem. Lett. 17 (2007) 2456–2458, https://doi.org/10.1016/j. bmcl.2007.02.029.
- [44] N. Li, L.-J. Wang, B. Jiang, S.-J. Guo, X.-Q. Li, X.-C. Chen, J. Luo, C. Li, Y. Wang, D.-Y. Shi, Design, synthesis and biological evaluation of novel pyrimidinedione derivatives as DPP-4 inhibitors, Bioorg. Med. Chem. Lett. 28 (2018) 2131–2135.
- [45] L. Wagner, C. Klemann, M. Stephan, S. von Hörsten, Unravelling the immunological roles of dipeptidyl peptidase 4 (DPP4) activity and/or structure homologue (DASH) proteins, Clin. Exp. Immunol. 184 (2016) 265–283, https:// doi.org/10.1111/cei.12757.
- [46] L.L. Brockunier, J. He, L.F. Colwell, B. Habulihaz, H. He, B. Leiting, K.A. Lyons, F. Marsilio, R.A. Patel, Y. Teffera, J.K. Wu, N.A. Thornberry, A.E. Weber, E. R. Parmee, Substituted piperazines as novel dipeptidyl peptidase IV inhibitors, Bioorganic Med. Chem. Lett. 14 (2004) 4763–4766, https://doi.org/10.1016/j. bmcl.2004.06.065.
- [47] G.R. Lankas, B. Leiting, R.S. Roy, G.J. Eiermann, M.G. Beconi, T. Biftu, C.-C. Chan, S. Edmondson, W.P. Feeney, H. He, D.E. Ippolito, D. Kim, K.A. Lyons, H.O. Ok, R.

A. Patel, A.N. Petrov, K.A. Pryor, X. Qian, L. Reigle, A. Woods, J.K. Wu, D. Zaller, X. Zhang, L. Zhu, A.E. Weber, N.A. Thornberry, Dipeptidyl Peptidase IV Inhibition for the Treatment of Type 2 Diabetes: Potential Importance of Selectivity Over Dipeptidyl Peptidases 8 and 9, Diabetes. 54 (2005) 2988–2994, https://doi.org/10.2337/diabetes.54.10.2988.

- [48] J.L. Velázquez-Libera, F. Durán-Verdugo, A. Valdés-Jiménez, G. Núñez-Vivanco, J. Caballero, LigRMSD: a web server for automatic structure matching and RMSD calculations among identical and similar compounds in protein-ligand docking, Bioinformatics. 36 (2020) 2912–2914, https://doi.org/10.1093/bioinformatics/ btaa018.
- [49] C. Rummey, G. Metz, Homology models of dipeptidyl peptidases 8 and 9 with a focus on loop predictions near the active site, Proteins Struct. Funct. Bioinforma. 66 (2006) 160–171, https://doi.org/10.1002/prot.21138.
- [50] G.F. Pauli, S.-N. Chen, C. Simmler, D.C. Lankin, T. Gödecke, B.U. Jaki, J.B. Friesen, J.B. McAlpine, J.G. Napolitano, Importance of Purity Evaluation and the Potential of Quantitative <sup>1</sup>H NMR as a Purity Assay, J. Med. Chem. 57 (2014) 9220–9231, https://doi.org/10.1021/jm500734a.
- [51] A. Aljuhani, M.R. Aouad, N. Rezki, O.A. Aljaldy, S.A. Al-Sodies, M. Messali, I. Ali, Novel pyridinium based ionic liquids with amide tethers: Microwave assisted synthesis, molecular docking and anticancer studies, J. Mol. Liq. 285 (2019) 790–802, https://doi.org/10.1016/j.molliq.2019.04.143.
- [52] Y. Ran, H. Pei, C. Xie, L. Ma, Y. Wu, K. Lei, M. Shao, M. Tang, M. Xiang, A. Peng, Y. Wei, L. Chen, Mol. Divers. 19 (2015) 333–346.