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Tetrahedron Letters xxx (2014) xxx-xxx

Contents lists available at ScienceDirect

Tetrahedron Letters

journal homepage: www.elsevier.com/locate/tetlet



Total synthesis of catenioblin B

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ARTICLE INFO

ABSTRACT

Article history: Received 25 June 2014 Revised 6 September 2014 Accepted 8 September 2014 Available online xxxx

Herein we report the first total synthesis of catenioblin B (1) via a titanium(IV) mediated regioselective epoxide ring-opening, Grignard reaction, Luche's reduction and TEMPO/BAIB mediated intramolecular cyclization as the key steps.

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Keywords: Catenioblins Pyranone derivative Sharpless epoxidation Regioselective epoxide ring-opening Grignard reaction

In recent years polyol containing chiral δ -lactones have attracted much attention due to their importance as building blocks in natural product synthesis¹ and their various biological activity profiles. Recently, catenioblin B^2 (1) a polyketide derived pyranone was isolated from the insect pathogenic fungus Paecilomyces cateniobliquus. The fungal strain P. cateniobliquus. YMF1.01799 showed good inhibitory activity towards growth of cotton bollworms and nematodes and regulate growth of insects. Catenioblin B (1) is structurally related to prelactone C^3 but contains a *cis* vicinal diol at the C_3-C_4 positions (Fig. 1).

The retrosynthetic analysis of catenioblin (Scheme 1) depicts the regioselective epoxide ring-opening reaction⁴ of known epoxy alcohol **2**⁵ as the synthetic tool in accessing polyol system en route the crucial compound 11, while 11 in turn could be conceived from epoxy alcohol derivative 3 via a sequence of transformations followed by Grignard reaction, and oxidation. A key step in the synthesis of the crucial fragment 11 was a Luche's reduction, followed by the deprotection of the PMB-ether and pyranone ring cyclization under TEMPO/BIAB conditions and finally deprotection of the *cis*-diol would furnish the natural product **1**.

Accordingly, the known epoxy alcohol 2 (Scheme 2) was accessed from the readily available starting material, 1,3 propanediol by a reported procedure.⁵ Thus, the epoxy alcohol **2** when subjected to the regioselective ring-opening reaction with benzoic acid⁴ (Bz-OH/Ti(OⁱPr)₄/CH₂Cl₂/0 °C) afforded 1,2 diol **3** (85%). Diol **3** on selective silvl protection (TBSCl/*n*-Bu₂SnO/CH₂Cl₂/0 °C) gave

http://dx.doi.org/10.1016/j.tetlet.2014.09.034 0040-4039/© 2014 Published by Elsevier Ltd.



Prelactone C

Figure 1. Structures catenioblin B and prelactone C.

mono silvl ether compound 4(90%) which on benzovl group deprotection (K₂CO₃/MeOH/0 °C) resulted in diol 5 (85%). Masking of diol 5 as its acetonide (2,2 DMP/PPTS/CH₂Cl₂/0 °C) gave compound 6 (80%). Later, deprotection of TBS ether with TBAF furnished primary alcohol 7^{4b} (86%).

Next the primary alcohol 7 was subjected to Swern oxidation followed by Grignard reaction⁶ (1-propenyl magnesiumbromide/ THF/0 °C) to afford *syn/anti* diastereomeric mixture along with its E,Z-isomers of allyl alcohol 8 (82% over two steps). Further, the diasteromeric mixture (without separation) was subjected to Dess–Martin periodinane oxidation to provide the α , β -unsaturated ketone as a E/Z mixture 9 (92% in 3:2 ratio). The purified E-isomer 9 was reduced under Luche's reduction conditions⁷ (CeCl₃·7H₂O/ NaBH₄/MeOH/-78 °C) to afford chiral allylic alcohol 10 (90%). The de of allylic alcohol 10 was calculated as 93.4% by LCMS {column: XCB C18 column, 30% water (NH₄OAc 10 mm) in acetonitrile, flow rate: 1 mL/min, 254 nm, t_r (major) = 9.555 min, t_r (minor) = 8.843 min} in favor of S-alcohol (major).¹⁰ The absolute stereochemistry at C5 was tentatively assigned as 'S' based on literature

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Scheme 1. Retrosynthetic analysis of the catenioblin B (1).



Scheme 2. Reagents and conditions: (a) BzOH, $Ti(O^{1}Pr)_{4}$, $CH_{2}Cl_{2}$, 0 °C to rt, 3 h, 85%; (b) TBSCl, *n*-Bu₂SnO, imidazole, $CH_{2}Cl_{2}$, 0 °C to rt, 3 h, 90%; (c) $K_{2}CO_{3}$, MeOH, 0 °C to rt, 2 h, 85%; (d) 2,2-DMP, PPTS, $CH_{2}Cl_{2}$, 0 °C to rt, 6 h, 80%; (e) TBAF, THF, 0 °C to rt, 2 h, 86%; (f) (i) (COCl)₂, DMSO, Et₃N, $CH_{2}Cl_{2}$, -78 °C, 1 h, (ii) 1-propenyl magnesium bromide, dry THF, 0 °C, 2 h (82% over two steps); (g) DMP, $CH_{2}Cl_{2}$, 0 °C to rt, 1 h, 92%; (h) CeCl₃, NaBH₄, MeOH, -78 °C, 4 h, 90%; (i) DDQ, 0 °C to rt, 2 h, 88%; (j) TEMPO, BAIB, $CH_{2}Cl_{2}$: H₂O (3:1) 0 °C to rt, 4 h, 88%; (k) Amberlyst, $CH_{3}CN$, 0 °C to rt, 5 h, 78%.



Figure 2. (a) NOESY spectrum (in CDCl₃) of 1; (b) energy minimized structure of 1.

evidence⁷ and confirmed by extensive NMR studies conducted subsequently. Next, deprotection of the PMB group in allylic alcohol **10** under DDQ conditions led to the crucial primary alcohol **11** (88%). And the assignment of stereochemistry at C5 stereogenic carbon as 'S' in compound **11** was based on the 1H–1H COSY and NOESY experiments (Fig. 2) that revealed the absence of NOE between C5H/C3H suggested that both protons are in *trans* orientation to each other and with opposite facial orientation. This was supported by NOE correlation between C3H/C1H, C3H/Me-B, C5H/Me-A, C5H/ C7H and C6H/C7H confirming its assigned stereochemistry.

Finally the primary alcohol **11** was subjected to TEMPO/BAIB conditions⁸ to afford the cyclized product **12** (88%) that upon

Please cite this article in press as: Srinivas, T.; et al. Tetrahedron Lett. (2014), http://dx.doi.org/10.1016/j.tetlet.2014.09.034

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Figure 3. (a) NOESY spectrum (in CDCl₃) of 11; (b) energy minimized structure of 11.

deprotection of acetonide group under acidic conditions with Amberlyst⁹ led to the final target **1** (78%). The spectral data of the target compound **1** was in fairly good agreement with that of the isolated natural product (see SI).¹⁰ However, the optical rotation of the synthetic **1** was found to be $[\alpha]_D^{25}$ –6.83 (*c* 1.5, MeOH); while that of Natural **1** was reported as: $[\alpha]_D^{25}$ –2.72, (*c* 0.16, MeOH).¹¹Additionally, the purity of synthetic **1** was determined by HPLC analysis {XCB C18 column; 4.6 × 150 mm; 40% water (NH₄OAc 10 mm) in acetonitrile; 1 mL/min; 254 nm, *t*_r(major) = 6.958, *t*_r(minor) = 3.019} and found to be 97.2%.¹⁰ Similarly, compound **1** (Fig. 3), on 1H–1H COSY and NOESY experiments, showed the correlation between C3H/C5H, C5H/C7H and C6H/C7H, confirming that the centre C5 assigned as 'S' was indeed correct and the energy diagrams are also incorporated in the Supporting information.

In summary, we have accomplished the first total synthesis of catenioblin B (1) in 12 steps featuring a Sharpless epoxidation, Ti(IV) promoted nucleophilic regioselective epoxide ring-opening and intramolecular cyclization as the key reaction steps.

Acknowledgments

One of the authors (T.S.) is thankful to the IICT-RMIT Research Programme (CLP-0092) for the financial support in the form of research fellowship. The authors are thankful to the respective Program Directors Dr. M. Lakshmi Kantam (IICT) and Prof. Suresh Bhargava (RMIT) for the generous support and encouragement.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.tetlet.2014.09. 034.

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- Spectral data of selected compounds: (E)-1-((45,55)-5-(2-(4-Methoxybenzyloxy)ethyl)-2,2-dimethyl-1,3-dioxolan-4-yl)but-2-en-1-one (9). [α]⁵⁵/₂ 14.23 (c 1.2, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.24 (d, J = 8.6 Hz, 2H), 6.93-7.05 (m, 1H), δ 6.87 (d, J = 8.6 Hz, 2H), 6.5 (dd, J = 1.7, 15.6 Hz, 1H), 4.52 (d, J = 8.6 Hz, 2H), 6.55 (d, J = 1.7, 15.6 Hz, 1H), 4.52 4.62 (m, 2H), 4.42 (s, 2H), 3.80 (s, 3H), 3.55 (q, J = 5.2, 7.3 Hz, 2H), 1.91 (dd, J = 1.7, 6.9 Hz, 3H), 1.66-1.77 (m, 2H), 1.61 (s, 3H), 1.39 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 197.2, 159.1, 144.5, 130.4, 127.5, 113.6, 113.7, 109.9, 81.7, 74.9, 72.6, 66.6, 55.2, 30.9, 27.1, 25.0,18.5; MS-ESI: m/z C₁₉H₂₆O₅ [M+NH₄]*: 352.

(S,E)-1-((4R,5S)-5-(2-(4-Methoxybenzyloxy)ethyl)-2,2-dimethyl-1,3-dioxolan-4yl)but-2-en-1-ol (**10**). $[x]_{D}^{25}$ -8.3 (c 0.5, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.26 (d, J = 8.6 Hz, 2H), 6.88 (d, J = 8.6 Hz, 2H), 5.71-5.84 (m, 1H), 5.45 (dddd, J = 1.5, 7.1, 15.48 Hz, 1H), 4.4 (s, 2H), 4.30 (q, J = 6.0, 13.5 Hz, 1H), 4.09 (br s, 1H), 3.96 (t, J = 6.0 Hz, 1H), 3.81 (s, 3H), 3.55-3.61 (m, 2H), 2.35 (br s, 1H), 1.85-1.90 (m, 2H), δ 1.71 (d, J = 6.4 Hz, 3H), 1.49 (s, 3H), 1.37 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 159.1, 129.8, 129.2, 113.7, 107.9, 80.2, 74.2, 72.7, 70.7, 67.1, 55.2, 30.2, 25.3, 27.7, 17.9; MS-ESI: m/z Cl₃Hz₂O₅ [M+NH₄]*; 335. (*S*,*E*)-1-((4*R*,*S*))-5-(2-Hydroxyethyl)-2,2-dimethyl-1,3-dioxolan-4-yl)but-2-en-1-

of (**11**). $[α]_{25}^{25} - 12.9$ (*c* 0.5, CHCl₃); ¹H NMR (300 MHz, CDCl₃): 5.77–5.90 (m, 1H), 5.46 (ddd, *J* = 1.5, 6.7, 15.1 Hz, 1H), 4.29–4.36 (m, 1H), 4.11 (t, *J* = 6.7 Hz, 1H), 4.01 (t, *J* = 6.0 Hz, 1H), 3.80–3.85 (m, 2H), 2.37 (br s, 1H), 1.86–1.99 (m, 2H), δ 1.73 (d, *J* = 6.0 Hz, 3H), 1.52 (s, 3H), 1.38 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 130.3, 129.5, 108.4, 80.4, 76.1, 70.7, 61.1, 32.1, 27.8, 25.4, 17.9; MS-ESI: *m/z* C₁₁H₂₀O₄ [M+Na]⁺: 239.

Let: m/2 C1112004 [in: Hu] : 2.5. (363, 45, 763) -2,2-Dimethyl-4-([E]-prop-1-enyl)dihydro-3aH-[1,3]dioxolo[4,5c]pyran-6(4H)-one (12). $[\alpha]_D^{25} - 12.43$ (c 1.8, CHCl₃); ¹H NMR (300 MHz, CDCl₃): 5.87-5.93 (m, 1H), 5.72 (ddd, J = 1.6, 7.7, 15.5 Hz, 1H), 4.70-4.73 (m, 1H), 4.48 (d, J = 7.7 Hz, 1H), 4.38 (dd, J = 1.8, 7.6 Hz, 1H), 2.88 (dd, J = 2.3, 15.8 Hz, 1H), 2.52 (dd, J = 3.6, 15.8 Hz, 1H), 1.78 (dd, J = 1.5, 6.4 Hz, 3H), 1.47 (s, 3H), 1.34 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 171.7, 132.1, 124.2, 109.4, 78.9, 74.7, 71.4, 34.5, 25.9, 24.1, 17.8; MS-ESI: m/z C₁₁H₁₆O₄ [M+Na]*: 230.

 $\begin{array}{l} \label{eq:constraint} (13,5,65)-4,5-dihydroxy-6-((E)-prop-1-enyl)tetrahydro-2H-pyran-2-one \\ \label{eq:constraint} (45,55,65)-4,5-dihydroxy-6-((E)-prop-1-enyl)tetrahydro-2H-pyran-2-one \\ \label{eq:constraint} (12,2,2)-2,5,65) \\ \label{eq:constraint} (23,5,65)-4,5,67) \\ \label{eq:constraint} (14,25)-4,5,67) \\ \label{eq:constraint} (15,67)-1,5,1,14,2,11),4,25-4,5,9 \\ \label{eq:constraint} (15,67)-1,5,1,14,2,11),4,25-4,5,9 \\ \label{eq:constraint} (14,25)-4,5,9) \\ \label{eq:constraint} (15,67)-1,5,1,14,2,11),4,25-4,5,9 \\ \label{eq:constraint} (15,67)-1,5,1,14,2,11),4,25-4,5,9 \\ \label{eq:constraint} (15,67)-1,5,1,14,2,11),4,25-4,5,9 \\ \label{eq:constraint} (15,67)-1,5,1,14,2,11),4,25-4,25 \\ \label{eq:constraint} (15,67)-1,25,2,15,14,2,14),4,25-4,25 \\ \label{eq:constraint} (15,67)-1,25,2,15,14,2,14),2,5,14,2,14 \\ \label{eq:constraint} (15,67)-1,25,2,15,14,2,14),2,5,14,2,14 \\ \label{eq:constraint} (15,67)-1,25,2,15,14,2,14 \\ \label{eq:constraint} (15,67)-1,25,2,15,24,24 \\ \label{eq:constraint} (15,67)-1,25,24,24 \\ \labe$

11. In order to rationalize the differences in optical rotation values between the synthetic **1** and Natural product **1**, it was decided to record the same at different concentrations for the synthetic **1**. While, no specific rotation was detected at *c* 0.16, only at *c* 0.7 it was detected as $[\alpha]_D^{25}$ –6.95 (*c* 0.7, MeOH). Since the reported value is very low, the difference in optical rotation values between the two might be explained due to error in concentration and temperature.