

Towards the Synthesis of the Skeleton of Salvianolic Acid D

Clémence Queffélec, Fabrice Bailly, Philippe Cotelte*

Laboratoire de Chimie Organique et Macromoléculaire, UMR CNRS 8009, Université de Lille 1, 59655 Villeneuve d'Ascq, France
Fax +33(320)336309; E-mail: Philippe.Cotelte@univ-lille1.fr

Received 23 September 2005; revised 10 October 2005

Abstract: A successful synthesis of the acid part of salvianolic acid D is described (eight steps from isovanillin, 22% overall yield). The benzaldehyde key intermediate was obtained in six steps in 52% overall yield and was converted into the trimethylated precursor molecule using the Knoevenagel procedure. Finally, the acid part of salvianolic acid D was obtained by the exhaustive deprotection of the methyl groups with boron tribromide.

Key words: natural polyphenols, antivirals, boron tribromide

Salvia is an important genus widely cultivated and used in traditional medicines. It is a rich source of polyphenols and the water-soluble extracts of *Salvia* are of great importance due to their bioactivities (antioxidant, antiplatelet, antitumor, and antiviral activities).¹ Some of them exhibit potent effects against HIV-1 enzymes.^{2–4} Due to our convergent interests in the total synthesis of natural polyphenols^{5–6} and the discovery of new HIV-1 integrase inhibitors,^{7–8} we decided to systematically synthesize acid moieties of salvianolic acids and related compounds (Figure 1).

This work deals with the elaboration of a key intermediate in the synthesis of a series of dimers of caffeic acid and with the total synthesis of the acid part of salvianolic acid D (1).

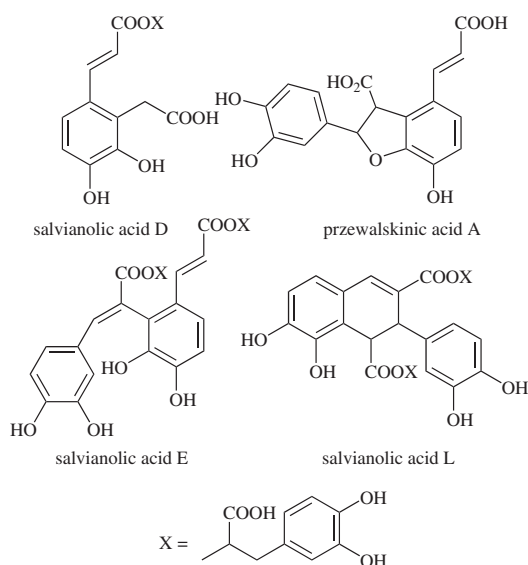
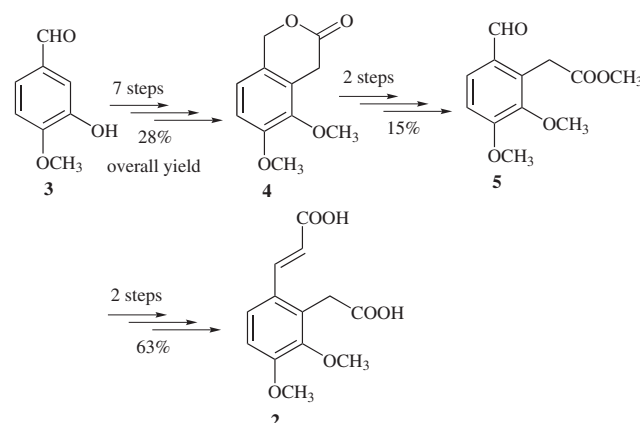


Figure 1 Structures of salvianolic acids and related compounds

SYNTHESIS 2006, No. 5, pp 0768–0770
Advanced online publication: 07.02.2006
DOI: 10.1055/s-2006-926332; Art ID: Z17705SS
© Georg Thieme Verlag Stuttgart · New York

Salvianolic acid D was isolated from *Salvia miltiorrhiza* and the synthesis of the dimethylated acid part of salvianolic acid D (2) was described.⁹ The prerequisite for the elaboration of the target polyphenol needs the building of the intermediate 5, which can be considered a milestone in the synthesis of various natural dimers of caffeic acid. Compound 5 can be obtained either from isovanillin or 2,3-dimethoxybenzaldehyde.

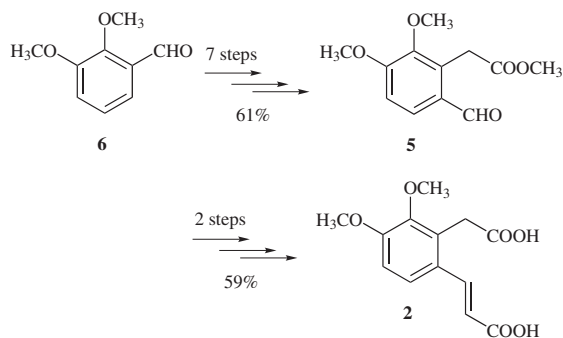
Ai and Li reported the synthesis of 5 starting from isovanillin 3 (Scheme 1). Using the same procedure, we were able to obtain compound 4 in 28% overall yield but the cleavage of the lactone previously described in 20% yield, was unsuccessful in our hands.



Scheme 1 Ai and Li procedure⁹

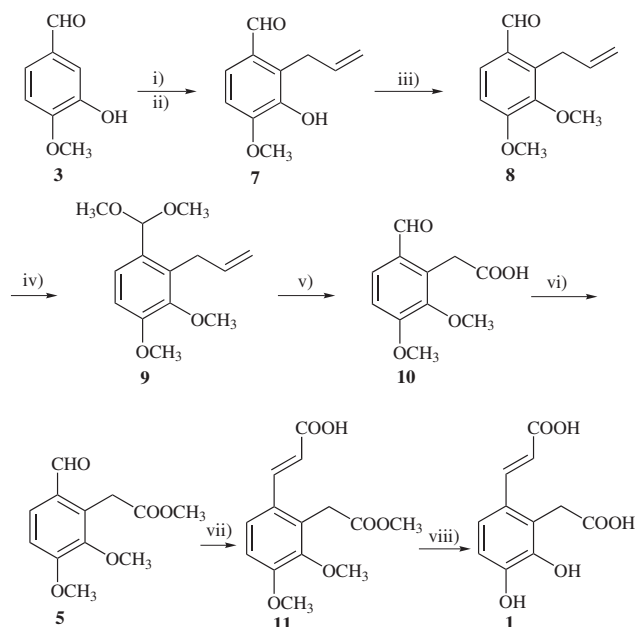
During the course of this work, 5 was also obtained in 61% yield by another method¹⁰ using 2,3-dimethoxybenzaldehyde 6 as starting material (Scheme 2). Since the sixth step of this procedure requires purification by column chromatography of two regioisomers and we had the need for a multigram-scale synthesis of 5 in mind, we decided to explore an alternative strategy (Scheme 3).

We started, as in the Ai and Li procedure, and in a short communication dealing with the total synthesis of heptamethyl lithospermate,¹¹ by the O-allylation of isovanillin followed by the Claisen rearrangement of O-allylisovanillin in dimethylacetamide. After methylation of the phenol groups, the protection of the formyl group was required, which was perfectly achieved by its transformation into the acetal derivative 9 with trimethyl orthoformate. A Lemieux–Rudloff oxidation led to the acid 10. The dimethylacetal was hydrolyzed during the work-up. Finally 10 was converted into its methyl ester to give 5.



Scheme 2 Detterbeck and Hesse procedure¹⁰

Two steps were necessary to obtain the acid part of salvianolic acid D from **5**. First, a Knoevenagel condensation was required and the α,β -unsaturated acid side-chain was received in good yield. Secondly, treatment with boron tribromide as deprotecting reagent afforded the desired product **1** in moderate yield.



Scheme 3 Reagents and conditions: i) Allyl bromide, K_2CO_3 , acetone, reflux, 3 h, 99%; ii) Dimethylacetamide, reflux, 10 h, 90%; iii) MeI, K_2CO_3 , DMF, r.t., 10 h, 91%; iv) $HC(OMe)_3$, MeOH, NH_4Cl , reflux, 2 h, 86%; v) $KMnO_4$, $NaIO_4$, K_2CO_3 , t -BuOH– H_2O , r.t., 4 h, 77%; vi) $SOCl_2$, MeOH, 0 °C, 1.5 h, 96%; vii) Malonic acid, py, piperidine, 60 °C, 10 h, 94%; viii) BBr_3 , CH_2Cl_2 , r.t., 10 h, then H_2O , 48 h, 45%.

Herein, we have described an efficient synthesis of the benzaldehyde **5** in a slightly lower overall yield (52%) than in a previous work (61%) but in a more convenient manner for a multigram scale. This key intermediate **5** is expected to allow for the total synthesis of przewalskinic acid A and the acid parts of salvianolic acid L and salvianolic acid E (Figure 1). As a first example, **5** was converted into the skeleton of salvianolic acid D (**1**) in a satisfactory 22% overall yield.

SiO_2 , 200–400 mesh (Merck) was used for column chromatography. Melting points were obtained on a Reichert Thermopan melting point apparatus, equipped with a microscope. NMR spectra were obtained with a AC 200 Bruker spectrometer in the appropriate solvent with TMS as internal reference. J values are given in Hz. Mass spectra were recorded on a Thermo-Finnigan PolarisQ mass spectrometer (70 eV, Electronic Impact). Elemental analyses were performed by CNRS laboratories (Vernaison) and were within 0.4% of the theoretical values.

3-Allyloxy-4-methoxybenzaldehyde

To isovanillin (**3**) (5.00 g, 32.9 mmol) in anhyd acetone (20 mL), allyl bromide (5.00 g, 3.6 mL, 41.3 mmol) and K_2CO_3 (5.00 g, 36.2 mmol) were added. Then the mixture was refluxed for 3 h. The removal of K_2CO_3 by filtration and the evaporation of acetone gave the desired compound (6.32 g, 99%) as a yellow oil.

1H NMR (200 MHz, $CDCl_3$): δ = 3.80 (s, 3 H, OMe), 4.55 (dt, 2 H, 4J = 1.45 Hz, 3J = 5.4 Hz, CH_2), 5.17 (dq, 1 H, 2J = 4J = 1.45 Hz, $^3J_{cis}$ = 10.2 Hz), 5.29 (dq, 1 H, 2J = 4J = 1.45 Hz, $^3J_{trans}$ = 17.9 Hz), 5.99 (m, 1 H, $^3J_{cis}$ = 10.2 Hz, $^3J_{trans}$ = 17.9 Hz, 3J = 5.4 Hz, =CH), 6.83 (d, 1 H, 3J = 8.3 Hz), 7.25 (d, 1 H, 4J = 1.9 Hz), 7.31 (dd, 1 H, 3J = 8.3 Hz, 4J = 1.9 Hz), 9.61 (s, 1 H, CHO).

2-Allyl-3-hydroxy-4-methoxybenzaldehyde (**7**)

A solution of 3-allyloxy-4-methoxybenzaldehyde (6.316 g, 32.90 mmol) in dimethylacetamide (9.2 mL) was stirred at 180 °C for 10 h. The cold solution was poured into aq 2 N NaOH soln (4 × 30 mL) and washed with Et_2O (2 × 20 mL). The aq layer was acidified with conc HCl soln and was extracted with $EtOAc$. Evaporation gave **7** as a brown oil (5.68 g, 90%).

1H NMR (200 MHz, $CDCl_3$): δ = 3.80 (d, 2 H, 3J = 5.9 Hz, CH_2), 3.95 (s, 3 H, OMe), 5.00 (m, 2 H), 5.81 (br s, 1 H), 6.05 (m, 1 H, $^3J_{cis}$ = 10.5 Hz, $^3J_{trans}$ = 16.5 Hz, 3J = 6.0 Hz, =CH), 6.83 (d, 1 H, 3J = 8.3 Hz), 7.41 (d, 1 H, 3J = 8.3 Hz), 10.05 (s, 1 H, CHO).

2-Allyl-3,4-dimethoxybenzaldehyde (**8**)

To **7** (6.00 g, 31.2 mmol) in DMF (34 mL), K_2CO_3 (8.65 g, 62.7 mmol) and MeI (6.61 g, 2.9 mL, 46.6 mmol) were added. The mixture was stirred at r.t. for 10 h, then taken up in H_2O (20 mL) and extracted with Et_2O (3 × 20 mL). Washing the organic phase with aq 2 N NaOH soln and evaporation gave **8** (5.85 g, 91%) as a brown oil.

1H NMR (200 MHz, $CDCl_3$): δ = 3.80 (s, 3 H, OMe), 3.82 (dt, 2 H, 3J = 3.5 Hz, CH_2), 3.95 (s, 3 H, OMe), 5.00 (m, 2 H), 6.05 (m, 1 H, $^3J_{cis}$ = 10.5 Hz, $^3J_{trans}$ = 16.5 Hz, 3J = 6.0 Hz, =CH), 6.90 (d, 1 H, 3J = 8.6 Hz), 7.61 (d, 1 H, 3J = 8.6 Hz), 10.05 (s, 1 H, CHO).

^{13}C NMR (50 MHz, $CDCl_3$): δ = 28.4 (t), 55.9 (q), 60.5 (q), 109.7 (d), 115.2 (d), 127.7 (s), 129 (d), 135.7 (s), 137 (d), 147 (s), 157.2 (s), 190.5 (d).

MS (EI, 70 eV): m/z (%) = 206 (50) [M^+], 191 (100), 190 (15), 175 (32), 115 (16), 91 (27).

Anal. Calc for $C_{12}H_{14}O_3$: C, 69.88; H, 6.84; O, 23.27. Found: C, 69.44; H, 6.96; O, 23.39.

2-Allyl-1-(dimethoxymethyl)-3,4-dimethoxybenzene (**9**)

A mixture of **8** (5.85 g, 28.4 mmol), trimethyl orthoformate (6.05 g, 6.2 mL, 56.8 mmol), and a cat. amount of NH_4Cl (ca 0.1 g) in MeOH (54 mL) was refluxed for 2 h. After cooling to r.t., the mixture was poured into ice-cooled aq 2 N NaOH soln (10 mL) and extracted quickly with Et_2O (3 × 20 mL). Evaporation gave **9** (6.15 g, 86%) as a brown oil.

1H NMR (200 MHz, $CDCl_3$): δ = 3.26 (s, 6 H, OMe), 3.5 (dt, 2 H, CH_2), 3.76 (s, 3 H, OMe), 3.83 (s, 3 H, OMe), 4.95 (m, 2 H), 5.35 (s, 1 H), 5.95 (m, 1 H, =CH), 6.77 (d, 1 H, 3J = 8.6 Hz), 7.25 (d, 1 H, 3J = 8.6 Hz).

^{13}C NMR (50 MHz, CDCl_3): δ = 23.4 (t), 47 (q, 2OCH_3), 49.3 (q), 54.4 (q), 95.2 (d), 103.4 (d), 108.6 (d), 116.1 (d), 122.8 (s), 125.7 (s), 131 (d), 141 (s), 146.5 (s).

MS (EI, 70 eV): m/z (%) = 252 (6) $[\text{M}^+]$, 221 (33) $[\text{M}^+ - \text{OCH}_3]$, 206 (36), 191 (82), 189 (100), 188 (32), 175 (36), 174 (77), 149 (38).

Anal. Calc for $\text{C}_{14}\text{H}_{20}\text{O}_4$: C, 66.65; H, 7.99; O, 25.37. Found: C, 66.45; H, 7.57; O, 25.20.

(6-Formyl-2,3-dimethoxyphenyl)acetic Acid (**10**)

To **9** (6.15 g, 24.4 mmol) in *t*-BuOH (40 mL) was added a solution of K_2CO_3 (10.1 g, 73.1 mmol) in H_2O (79 mL). To the resulting suspension were added NaIO_4 (21.6 g, 101 mmol) and KMnO_4 (3.15 g, 19.5 mmol). The mixture started to warm and was stirred for 4 h at r.t. The mixture was washed with EtOAc (10 mL) and this organic layer discarded. The aq layer was acidified with concd HCl soln and again exhaustively extracted with EtOAc. Evaporation yielded **10** (4.2 g, 77%) as a brown oil.

^1H NMR (200 MHz, CDCl_3): δ = 3.75 (s, 3 H, OMe), 3.88 (s, 3 H, OMe), 4.12 (s, 2 H, CH_2), 6.85 (d, 1 H, 3J = 8.3 Hz), 7.5 (d, 1 H, 3J = 8.3 Hz), 9.8 (s, 1 H, CHO).

^{13}C NMR (50 MHz, CDCl_3): δ = 31.3 (t), 56.1 (q), 61.1 (q), 110.7 (d), 128.3 (s), 129.3 (s), 132.9 (d), 148.3 (s), 157.6 (s), 177 (s), 192.2 (d).

MS (EI, 70 eV): m/z (%) = 194 (38) $[\text{M}^+ - \text{CH}_2\text{O}]$, 180 (20), 179 (100), 91 (15).

Anal. Calc for $\text{C}_{11}\text{H}_{12}\text{O}_5$: C, 58.93; H, 5.39; O, 35.68. Found: C, 58.67; H, 5.25; O, 35.90.

Methyl (6-Formyl-2,3-dimethoxyphenyl)acetate (**5**)

To an ice-cooled solution of **10** (4.20 g, 18.8 mmol) in MeOH (31 mL), SOCl_2 (3.72 g, 2.3 mL, 31.3 mmol) was slowly added dropwise. After 1.5 h stirring, evaporation yielded **5** (4.28 g, 96%) as a brown oil.

^1H NMR (200 MHz, CDCl_3): δ = 3.67 (s, 3 H, CO_2Me), 3.8 (s, 3 H, OMe), 3.95 (s, 3 H, OMe), 4.18 (s, 2 H, CH_2), 7.00 (d, 1 H, 3J = 8.6 Hz), 7.58 (d, 1 H, 3J = 8.6 Hz), 9.90 (s, 1 H, CHO).

3-[3,4-Dimethoxy-2-(2-methoxy-2-oxoethyl)phenyl]acrylic Acid (**11**)

To a solution of **5** (1.10 g, 4.62 mmol) in pyridine (136 mL) were added malonic acid (979 mg, 9.41 mmol) and two drops of piperidine. The mixture was refluxed for 10 h and then poured into an excess of concd HCl soln. Extraction with EtOAc and evaporation gave **11** (1.216 g, 94%) as a brown solid.

^1H NMR (200 MHz, CDCl_3): δ = 3.7 (s, 3 H), 3.82 (s, 3 H), 3.88 (s, 2 H), 3.91 (s, 3 H), 6.29 (d, 1 H, 3J = 15.6 Hz), 6.9 (d, 1 H, 3J = 8.9 Hz), 7.42 (d, 1 H, 3J = 8.9 Hz), 7.92 (d, 1 H, 3J = 15.6 Hz).

Acid Part of Salvianolic Acid D (**1**)

To a solution of **11** (4.88 g, 17.4 mmol) in CH_2Cl_2 (20 mL), a 1 M BBr_3 soln in CH_2Cl_2 (69.5 mL) was added dropwise at 20 °C. After 24 h of stirring, the reaction was carefully quenched with H_2O (40 mL). A brown solid precipitated rapidly and filtration of this gave **1** (1.87 g, 45%) as a brown solid.

^1H NMR (200 MHz, CD_3COCD_3): δ = 4.09 (s, 2 H), 6.29 (d, 1 H, 3J = 16.13 Hz), 6.96 (d, 1 H, 3J = 8.51 Hz), 7.38 (d, 1 H, 3J = 8.51 Hz), 7.58 (d, 1 H, 3J = 16.0 Hz).

^{13}C NMR (50 MHz, CDCl_3): δ = 32.9 (t), 116.4 (d), 117.9 (d), 121.5 (s), 124.7 (d), 125.7 (s), 140.9 (d), 141.4 (s), 142.9 (s), 167.6 (s), 173.9 (s).

MS (EI, 70 eV): m/z (%) = 220 (46) $[\text{M} - \text{H}_2\text{O}]^+$, 176 (100) $[\text{M} - \text{H}_2\text{O} - \text{CO}_2]^+$, 148 (45), 147 (50), 146 (20), 119 (31), 91 (46).

Anal. Calc for $\text{C}_{11}\text{H}_{10}\text{O}_6$: C, 55.47; H, 4.23; O, 40.30. Found: C, 55.87; H, 4.61; O, 40.57.

Acknowledgment

This work was financially supported by grants from Centre National de la Recherche Scientifique (CNRS) and Agence Nationale de la Recherche contre le Sida (ANRS).

References

- (1) Lu, Y.; Foo, L. Y. *Phytochemistry* **2002**, *59*, 117.
- (2) Abd-Elazem, I. S.; Chen, H. S.; Bates, R. B.; Huang, R. C. *Antiviral Res.* **2002**, *55*, 91.
- (3) Han, M. K.; Lee, P. WO Patent 9966942, **1999**; *Chem. Abstr.* **2000**, *132*, 231935.
- (4) Han, M. K.; Lee, P. US Patent 6043276, **2000**; *Chem. Abstr.* **2000**, *132*, 69303.
- (5) Dalla, V.; Cotellet, P. *Tetrahedron Lett.* **1998**, *39*, 8285.
- (6) Dalla, V.; Cotellet, P. *Tetrahedron* **1999**, *55*, 6923.
- (7) Dupont, R.; Jeanson, L.; Mouscadet, J. F.; Cotellet, P. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 3175.
- (8) Maurin, C.; Bailly, F.; Buisine, E.; Vezin, H.; Mbemba, G.; Mouscadet, J. F.; Cotellet, P. *J. Med. Chem.* **2004**, *47*, 5583.
- (9) Ai, C. B.; Li, L. N. *Planta Med.* **1992**, *58*, 197.
- (10) Detterbeck, R.; Hesse, M. *Helv. Chim. Acta* **2003**, *86*, 343.
- (11) Nicolaou, K. C.; Seitz, S. P.; Pavia, M. R.; Petasis, N. A. *J. Org. Chem.* **1979**, *44*, 4013.