

Synthesis of γ -Oxo- α -amino Acids via Radical Acylation with **Carboxylic Acids**

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deoxygenative protocol to access γ -oxo- α -amino acid derivatives. This radical methodology employs photoredox catalysis, in combination with triphenylphosphine, to generate acyl radicals from readily available (hetero)aromatic and vinylic carboxylic acids. This approach allows for the straightforward synthesis of γ -oxo- α amino acids bearing a wide range of functional groups (e.g., Cl, CN, furan, thiophene, Bpin) in synthetically useful yields (~60% average yield). To further highlight the utility of the methodology, several deprotection and derivatization reactions were carried out.

 γ -Oxo- α -amino acids are highly versatile building blocks in organic synthesis, as well as key components in biologically active molecules. They can be used as precursors for homophenylalanine derivatives,¹ γ -hydroxy- α -amino acids,² γ valerolactones,³ or γ -valerolactames, for example. As it is often the case in synthetic chemistry, one of the main challenges associated with this interesting class of amino acids is their stereoselective synthesis. There are four main retrosynthetic pathways to achieve this goal: (a) via acylation reactions, starting from L- or D-aspartic acid, (b) via asymmetric or diastereoselective Mannich reactions, 5 (c) via asymmetric Stetter reactions,⁶ or (d) via transition metal-mediated crosscouplings with iodoalanine or β -metalated alanine derivatives (Figure 1A).⁷ While powerful, these methodologies often present limitations regarding the scope of coupling partners, or require the use of chiral catalysts.

Radical chemistry offers exciting and highly attractive approaches to access new chemical space in a rapid fashion.⁸ As such, it has been exploited for the synthesis and derivatization of amino acids and peptides.⁹ We recently contributed to this area with the development of a decarboxylative protocol for the diastereoselective synthesis of a wide range of unnatural amino acids (UAAs) using the Beckwith-Karady alkene I¹⁰ as radical acceptor.¹¹ Although this methodology granted access to γ -oxo- α -amino acids derivatives (II) when using α -keto acids as acylating reagents (Figure 1B), it afforded diminished yields with electron deficient or (hetero)aromatic systems. In addition, α -keto acids are not readily available, and their synthesis often requires the use of hazardous reagents, such SeO₂. Since II is a highly versatile species, we became interested in developing alternative methodologies for its synthesis using more readily available starting materials.¹²



Recently, the development of deoxygenative radical strategies to access acyl radicals has attracted increased attention.¹³ Seminal independent studies by Rovis and Doyle^{13e} and Zhu^{13d} described the use of photoredox catalysis to generate phosphine radical cations that swiftly react with carboxylates to generate acyl radicals and phosphine oxide after β -scission (vide infra).¹⁴ Encouraged by these reports, we envisioned that it might be possible to develop a diastereoselective synthesis of γ -oxo- α -amino acids using I and readily available carboxylic acids as acyl radical sources (Figure 1C).

Herein we present a highly efficient, light-mediated deoxygenative protocol to access products II from readily and commercially available (hetero)aromatic and vinylic carboxylic acids. In addition, the utility of this methodology is further highlighted by several derivatizations and deprotections of II.¹³

Initial optimization studies were carried out using benzoic acid as the acylating reagent.¹⁶ The targeted product (1) could be isolated in 95% yield and excellent diastereoselectivity (d.r. > 20:1) using 1.0 equiv of I, 1.5 equiv of benzoic acid, 1.8 equiv. of PPh₃, 2.0 equiv. of 2,4,6-collidine, and 1.0 mol% $[Ir(dFCF_3ppy)_2(dtbbpy)][PF_6]$ (Ir-F) in 1,4-dioxane (0.2) M) while irradiating with 32 W blue LEDs (λ_{max} = 440 nm) for 24 h at room temperature. Control experiments showed that the reaction needs both light and a photocatalyst to proceed, and that it does not proceed when using 4CzIPN,¹⁷ an

Received: December 15, 2020 Published: June 1, 2021







Figure 1. Synthetic strategies toward γ -oxo- α -amino acids.

organophotocatalyst possessing similar redox potentials to Ir–F.

With the optimized conditions in hand, the scope and limitations, as well as the scalability of the methodology, were explored (Scheme 1). The standard reaction with benzoic acid was scaled up to 5.0 mmol (1.4 g of II), affording 1 in 95% (1.9 g) and 73% (1.4 g) yield using 0.5 mol% and 0.25 mol% of Ir-F, respectively. This highlights the high catalytic efficiency of the methodology, affording TON up to 288. Regarding the scope, aromatic carboxylic acids were first tested (2-15). Both electron rich and poor *para*-substituents on the aromatic ring were tolerated (2-10), although the latter afforded diminished yields. However, this represents a significant improvement compared to our previous methodology employing α -keto acids as acylating reagents, e.g., compound 9 was isolated in 61% yield vs 31% yield using α keto acids.¹¹ Free nucleophilic motifs, such as hydroxy groups, were not tolerated (2); however, this limitation could be circumvented by the use of protecting groups (3 and 4). Challenging substrates bearing sensitive functional groups, such as nitriles (8) or aldehydes (10), afforded the desired products in moderate to poor yields, while compound 15, bearing a meta-boronic ester substituent, was obtained in 76% yield. Gratifyingly, ortho-substituents were well tolerated (12 and 13), and salicylic acid derived 13 was obtained in an excellent 92% yield. More complex aromatic carboxylic acids bearing multiple functional groups (14) afforded the targeted γ -oxo- α -amino acid derivative in excellent yield.

The use of heteroaromatic carboxylic acids was also investigated (16-22). While nicotinic acid afforded the

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desired product in moderate yields (16), no product was observed with picolinic or pyrazinoic acids (18–19). Surprisingly, when the reaction was carried out using 4-chloro-1,3-dimethylpyrazolo[3,4-b]pyridine-5-carboxylic acid, the main product was the dechlorinated species 17 (39%), while the expected product 17' was isolated in 18% yield. The use of 5-membered heterocycles (20–22), such as unprotected pyrroles (20), furans (21), and thiophenes (22) afforded the desired products in variable yields (21–71%). Overall, our new methodology presents a broad functional group tolerance, where compounds bearing several vectors for further functionalization, such as halides, boronic esters, or amines, can be readily obtained.

To further challenge the limits of our methodology, the use of aliphatic, cinnamic, and vinylic carboxylic acids as acylating reagents was evaluated. While hydrocinnamic acid failed to afford the desired product,¹⁶ cinnamic acid delivered a complex mixture, from where the targeted product could not be isolated.¹⁸ However, the use of cyclic, vinylic carboxylic acids afforded interesting γ -oxo- α -amino acid derivatives bearing 5- and 6-membered heterocycles, such dihydrofuranes (27), tetrahydropyrines (26), and tetrahydropyrans (24). To the best of our knowledge, this is the first time that vinylic carboxylic acids have been directly used as acyl radical precursors.

To highlight the utility of our methodology, a series of derivatization reactions were carried out. Acidic deprotection of II using concentrated HCl in 1,4-dioxane afforded γ -oxo- α -amino acid salts **28–30** in quantitative yields (Scheme 2A). To check the e.r. of the products after the deprotection step, compound **1** was deprotected and further derivatized to the corresponding N-Boc protected methyl ester. HPLC analysis of the latter revealed a 92:8 e.r. Our methodology can also be applied for the acylation of dehydroalanine derivative IA, affording the corresponding product **1A** in 55% yield (Scheme 2B), and for the synthesis of ¹³C-labeled amino acid derivatives (Scheme 2C).

Finally, a plausible reaction mechanism for this transformation is shown in Figure 2. First, the excited photocatalyst $(*Ir^{III}, *E_{1/2} = +1.21$ V vs SCE)¹⁹ undergoes reductive quenching by PPh₃ ($E_{1/2} = +0.98$ V vs SCE)²⁰ to generate triphenhylphosphine radical cation III and a IrII species. III reacts with the corresponding carboxylic acid to afford phosphoranyl radical IV, which readily undergoes β -scission to deliver OPPh₃ and the key acyl radical V. Subsequent radical addition of the latter to I affords α -amino radical VI, which after reduction by the reduced $Ir^{II} (E_{1/2} = -1.37 \text{ V vs SCE})^{19}$ and protonation delivers the desired product II. This mechanism is in accordance with previous proposals for acylation reactions using photoredox catalysis to access phosphoranyl radicals.^{13,14} Quantum yield determinations suggest that there is also a significant contribution from a radical-chain pathway ($\Phi = 13.5$).¹⁶ On the basis of further experiments, 2,4,6-collidine seems to play a crucial role in the chain process. However, at this point, the nature of the chain carrier remains elusive.¹⁶

In conclusion, we have developed a highly efficient, lightmediated, deoxygenative strategy for the synthesis of γ -oxo- α amino acid derivatives. This radical methodology exploits the addition of acyl radicals, generated from readily available carboxylic acids, to Beckwith–Karady alkene I, allowing for the straightforward synthesis of a wide range of γ -oxo- α -amino acid derivatives in excellent diastereoselectivities and synthetically

Scheme 1. Scope and Limitations of the Methodology^e



^a5.0 mmol scale, Ir-F 0.5 mol%. ^b5.0 mmol scale, Ir-F 0.25 mol%, 72 h. ^c48 h. ^dDMF (0.2 M). ^eReaction conditions: Acid (0.75 mmol, 1.5 equiv.), I (0.50 mmol, 1.0 equiv.), Ir-F (1.0 mol%), PPh₃ (0.9 mmol, 1.8 equiv.), 2,4,6-collidine (1.0 mmol, 2.0 equiv.), 1,4-dioxane (0.2 M), RT, 24 h; product d.r. > 20:1.

useful yields (\sim 60% average yield). Furthermore, the synthetic utility of this protocol was highlighted by a series of derivatization reactions, granting access to in good yields and diastereoselectivities.

EXPERIMENTAL SECTION

General Information. Commercial reagents and solvents were used as purchased. Unless otherwise noted, all reactions were carried out under an atmosphere of N₂ in flame-dried glassware. The solvents used were purified by distillation over standard drying agents and were stored over molecular sieves or transferred under N₂. Blue LEDs (Kessil PR-160, 32 W, $\lambda_{max} = 440$ nm or EvoluChem CREE XPE, 30 W, $\lambda_{max} = 450$ nm) were used for irradiation, in combination with an EvoluChem PhotoRedOx Box. The reaction temperature was kept at 27 °C thanks to the fans incorporated in the reactor.

TLC were conducted with precoated glass-backed plates (silica gel 60 F254) and visualized by exposure to UV light (254 nm) or stained with ceric ammonium molybdate (CAM), basic potassium permanganate (KMnO₄), Ninhydrin or *p*-anisaldehyde solutions,

and subsequent heating. Flash column chromatography was performed on silica gel (40–60 μ m), and the eluent used is reported in the respective experiments.

¹H NMR spectra were recorded with 400 or 600 MHz instruments, ¹³C{¹H} NMR spectra at 101 or 151 MHz. Chemical shifts are reported in ppm relative to the solvent signal, coupling constants *J* in Hz. Multiplicities were defined by standard abbreviations. Lowresolution mass spectra (LRMS) were recorded using a LC/MScombination (ESI). High-resolution mass spectra (HRMS) were obtained using ESI ionization (positive) on a Bruker micrOTOF.

All carboxylic acids were commercially available, while [Ir- $(dFCF_3ppy)_2(dtbbpy)$][PF₆] (**Ir**-**F**),²¹ I and IA were synthesized according to literature procedures.^{10c}

Synthesis and Characterization of Products. General Procedure A (GP-A). An 8 mL Biotage microwave vial was charged with the corresponding carboxylic acid (0.75 mmol, 1.5 equiv.), I (145 mg, 0.50 mmol, 1.0 equiv.), PPh₃ (235 mg, 0.9 mmol, 1.8 equiv.), Ir–F (5.5 mg, 5 μ mol, 1 mol%), and sealed with a septum cap. The vial was put under a vacuum for 1 min and refilled with N₂ (× 3). Afterward, 2,4,6-collidine (132 μ L, 1.0 mmol, 2.0 equiv.) and

Scheme 2. Deprotection and Derivatization Reactions



Figure 2. Plausible reaction mechanism.

degassed 1,4-dioxane (2.5 mL, 0.2 M) were added. The reaction mixture was then sparged with N₂ for 2–5 min and irradiated with blue LEDs (λ_{max} = 440 or 450 nm) in an EvoluChem PhotoRedOx Box for 24 h. Finally, the solvent was evaporated and the crude reaction mixture was purified by column chromatography over silica gel to afford the desired product.

General Procedure B (GP-B). An 8 mL Biotage microwave vial was charged with the corresponding carboxylic acid (0.75 mmol, 1.5 equiv.), I (145 mg, 0.50 mmol, 1.0 equiv.), PPh₃ (235 mg, 0.9 mmol, 1.8 equiv.), Ir–F (5.5 mg, 5 μ mol, 1 mol%), and sealed with a septum cap. The vial was put under a vacuum for 1 min and refilled with N₂ (× 3). Afterward, 2,4,6-collidine (132 μ L, 1.0 mmol, 2.0 equiv.) and degassed DMF (2.5 mL, 0.2 M) were added. The reaction mixture was then sparged with N₂ for 2–5 min and irradiated with blue LEDs (λ_{max} = 440 or 450 nm) in an EvoluChem PhotoRedOx Box for 24 h. Finally, the solvent was evaporated and the crude reaction mixture was purified by column chromatography over silica gel to afford the desired product.

Characterization Data. (25,45)-2-(tert-Butyl)-5-oxo-4-(2-oxo-2-phenylethyl)oxazolidine-3-carboxylate (1). In 0.5 mmol scale:

Synthesized following **GP-A** using benzoic acid (90 mg, 0.75 mmol 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide **1** as a yellow oil in 95% yield (190 mg, 0.48 mmol). The spectroscopic data are consistent with those previously reported.¹¹ ¹H NMR (400 MHz, CDCl₃) δ 7.94–7.88 (m, 2H), 7.62–7.55 (m, 1H), 7.49–7.42 (m, 2H), 7.32–7.27 (m, 3H), 7.24–7.20 (m, 2H), 5.61 (s, 1H), 5.24 (dd, *J* = 6.9, 5.0 Hz, 1H), 5.11 (d, *J* = 12.1 Hz, 1H), 5.00 (d, *J* = 12.1 Hz, 1H), 3.56 (dd, *J* = 16.4, 6.9 Hz, 1H), 3.38 (dd, *J* = 16.4, 5.0 Hz, 1H), 1.02 (s, 9H).

In 5.0 mmol scale: Synthesized following **GP-A** using benzoic acid (916 mg, 7.5 mmol 1.5 equiv.) and **Ir**-**F** (28 mg, 25 μ mol, 0.5 mol%). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide **1** as a yellow oil in 97% yield (1.9 g, 4.8 mmol).

In 5.0 mmol scale: Synthesized following **GP-A** using benzoic acid (916 mg, 7.5 mmol 1.5 equiv.) and **Ir**-F (14 mg, 12.5 μ mol, 0.25 mol %) for 72 h. The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide **1** as a yellow oil in 73% yield (1.4 g, 3.6 mmol).

Benzyl (2*R*,4*R*)-2-(tert-butyl)-5-oxo-4-(2-oxo-2-phenylethyl-2-¹³C)oxazolidine-3-carboxylate (1-¹³C). Synthesized following GP-A using benzoic acid- α -¹³C (92.3 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by column chromatography (cyclohexane/:EtOAc, 4:1) to provide 1-¹³C as a yellow oil in 85% yield (110 mg, 0.27 mmol). ¹H NMR (400 MHz, CDCl₃) δ 7.91 (ddd, *J* = 8.5, 3.8, 1.3 Hz, 2H), 7.58 (tt, *J* = 6.9, 1.4 Hz, 1H), 7.46 (tt, *J* = 7.1, 0.7 Hz, 2H), 7.31–7.27 (m, 3H), 7.25–7.19 (m, 2H), 5.61 (s, 1H), 5.24 (ddd, *J* = 6.9, 5.0, 3.6 Hz, 1H), 5.11 (d, *J* = 12.1 Hz, 1H), 5.00 (d, *J* = 12.0 Hz, 1H), 3.56 (ddd, *J* = 16.3, 6.9, 5.5 Hz, 1H), 3.38 (ddd, *J* = 16.3, 6.0, 5.0 Hz, 1H), 1.02 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 194.8, 172.2, 172.1, 155.7, 136.7, 136.2, 135.3, 133.6, 128.9, 128.7, 128.5, 128.4, 96.4, 68.4, 53.8, 42.3, 41.8, 37.6, 24.9. HRMS (FI) [*m*/z] calculated for C₂₂¹³CH₂₅NNaO₅ ([M + Na]⁺) 419.1664, found 419.1666.

Methyl 2-(*bis*(*tert-butoxycarbonyl*)*amino*)-4-*oxo*-4-*phenylbuta*-*noate* (**1***A*). Synthesized following **GP-B** using benzoic acid (91.5 mg, 0.75 mmol, 1.5 equiv.) and **IA** (190.1 mg, 0.50 mmol, 1.0 equiv.). The crude product was purified by column chromatography (cyclohexane/:EtOAc, 4:1) to provide **1A** as a yellow oil in 54% yield (110 mg, 0.27 mmol). ¹H NMR (400 MHz, CDCl₃) δ 8.00–7.95 (m, 2H), 7.57 (s, 1H), 7.48 (dd, *J* = 7.3, 0.9 Hz, 2H), 5.73 (dd, *J* = 6.8, 5.7 Hz, 1H), 4.04 (dd, *J* = 17.5, 6.8 Hz, 1H), 3.72 (s, 3H), 3.28 (dd, *J* = 17.5, 5.7 Hz, 1H), 1.51 (s, 18H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 196.7, 170.9, 152.1, 136.8, 133.3, 128.8, 128.3, 83.5, 54.8, 52.6, 40.0, 28.2. HRMS (FI) [*m*/*z*] calculated for C₂₁H₂₉NNaO₇ ([M + Na]⁺) 430.1840, found 430.1836. IR \tilde{v} [cm⁻¹] = 2980 (s), 2935 (s), 1797 (m), 1967 (m), 1478 (m), 1451 (m), 1367 (s), 1238 (s), 1094 (m), 1045 (s), 850 (s), 766 (s), 691 (s), 634 (s), 606 (m), 440 (m). *R*_f (cyclohexane/EtOAc, 4:1) = 0.62 [*p*-Anisaldehyde].

Benzyl (2S,4S)-2-(tert-butyl)-4-(2-(4-methoxyphenyl)-2-oxoethyl)-5-oxooxazolidine-3-carboxylate (3). Synthesized following GP-A using 4-methoxybenzoic acid (114 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide 3 as a yellow oil in 92% yield (195 mg, 0.46 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.92–7.85 (m, 2H), 7.38-7.35 (m, 1H), 7.30-7.28 (m, 2H), 7.24-7.21 (m, 2H), 6.96-6.87 (m, 2H), 5.61 (s, 1H), 5.23 (dd, J = 7.0, 5.0 Hz, 1H), 5.13-5.09 (m, 1H), 5.00 (d, J = 12.1 Hz, 1H), 3.87 (s, 3H), 3.50 (dd, J = 16.1, 7.0 Hz, 1H), 3.33 (dd, J = 16.1, 5.0 Hz, 1H), 1.01 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 193.3, 172.3, 163.9, 155.7, 135.3, 130.6, 129.5, 128.7, 128.7, 128.6, 128.5, 114.1, 96.4, 68.3, 55.7, 53.9, 41.7, 37.6, 24.9. HRMS (FI) [m/z] calculated for C₂₄H₂₇NNaO₆ $([M + Na]^+)$ 448.1731, found 448.1734. IR $\tilde{v} [cm^{-1}] = 2964$ (w), 2914 (w), 2874 (w), 2842 (w), 1792 (s), 1718 (s), 1677 (m), 1599 (s), 1575 (m), 1511 (m), 1481 (w), 1457 (m), 1393 (m), 1345 (s), 1291 (s), 1258 (s), 1238 (s), 1217 (s), 1200 (m), 1168 (s), 1118 (m), 1067 (m), 1040 (s), 1027 (s), 989 (s), 889 (m), 840 (m), 824 (m), 783 (m), 738 (m), 697 (s), 633 (m), 596 (m), 563 (m), 531

(m), 505 (m), 454 (m). R_f (cyclohexane/EtOAc, 5:1) = 0.31 [*p*-Anisaldehyde]. $[\alpha]_D^{2D}$ = +41.1 (ρ = 0.99, CH₂Cl₂).

Benzyl (2S,4S)-4-(2-(4-((tert-butoxycarbonyl)amino)phenyl)-2oxoethyl)-2-(tert-butyl)-5-oxooxazolidine-3-carboxylate (4). Synthesized following GP-A using 4-((tert-butoxycarbonyl)amino)benzoic acid (178 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide 4 as a fine yellowish powder in 80% yield (203 mg, 0.40 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.92-7.85 (m, 2H), 7.38-7.35 (m, 1H), 7.30-7.28 (m, 2H), 7.26-7.20 (m, 2H), 6.96-6.87 (m, 2H), 5.61 (s, 1H), 5.23 (dd, J = 7.0, 5.0 Hz, 1H), 5.12-5.10 (m, 1H), 5.00 (d, J = 12.1 Hz, 1H), 3.87 (s, 3H), 3.50 (dd, J = 16.1, 7.0 Hz, 1H), 3.33 (dd, J = 16.1, 5.0 Hz, 1H), 1.01 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 193.4, 172.2, 155.7, 152.2, 143.4, 135.3, 131.0, 129.8, 128.7, 128.6, 128.5, 117.7, 96.4, 81.5, 68.3, 53.9, 41.7, 37.6, 28.4, 24.9. HRMS (FI) [m/z] calculated for C₂₈H₃₄N₂NaO₇ $([M + Na]^+)$ 533.2258, found 533.2265. IR \tilde{v} $[cm^{-1}] = 3338$ (w), 2972 (w), 2930 (w), 2875 (w), 2852 (w), 1792 (m), 1718 (m), 1679 (m), 1588 (m), 1525 (m), 1502 (m), 1481 (w), 1454 (w), 1393 (m), 1366 (m), 1344 (m), 1312 (m), 1290 (m), 1229 (s), 1149 (s), 1122 (m), 1045 (m), 990 (m), 900 (w), 838 (m), 822 (m), 769 (m), 747 (m), 696 (m), 626 (m), 593 (w), 576 (w), 540 (m), 510 (m), 458 (m). R_f (cyclohexane/EtOAc, 4:1) = 0.29 [p-Anisaldehyde]. $[\alpha]_D^{20}$ = +35.6 ($\rho = 1.02$, CH₂Cl₂).

Benzyl (2S,4S)-2-(tert-butyl)-5-oxo-4-(2-oxo-2-(p-tolyl)ethyl)oxazolidine-3-carboxylate (5). Synthesized following GP-A using 4-methylbenzoic acid (103 mg, 0.76 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/ EtOAc, 5:1) to provide 5 as a yellow oil in 80% yield (165 mg, 0.40 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.84-7.77 (m, 2H), 7.30-7.27 (m, 3H), 7.26-7.24 (m, 2H), 7.23-7.20 (m, 2H), 5.61 (s, 1H), 5.23 (dd, J = 6.9, 5.0 Hz, 1H), 5.11 (d, J = 12.1 Hz, 1H), 4.99 (d, J = 12.1 Hz, 1H), 3.53 (dd, J = 16.2, 6.9 Hz, 1H), 3.35 (dd, J = 16.2, 5.1 Hz, 1H), 2.42 (s, 3H), 1.01 (s, 9H). ¹³C{¹H} NMR (151 MHz, ${\rm CDCl}_3)\,\delta$ 194.4, 172.2, 155.7, 144.5, 135.3, 134.0, 129.6, 128.7, 128.6, 128.5, 128.5, 96.4, 68.3, 53.9, 41.9, 37.6, 24.9, 21.8. HRMS (FI) [m/ z] calculated for $C_{24}H_{27}NNaO_5$ ([M + Na]⁺) 432.1781, found 432.1789. IR \tilde{v} [cm⁻¹] = 3063 (w), 3034 (w), 2966 (w), 2875 (w), 1792 (s), 1719 (s), 1682 (s), 1606 (m), 1573 (m), 1480 (w), 1453 (m), 1393 (s), 1344 (s), 1289 (s), 1235 (s), 1174 (s), 1120 (s), 1068 (m), 1041 (s), 1018 (s), 984 (s), 890 (m), 840 (m), 807 (m), 745 (s), 697 (s), 635 (w), 592 (m), 560 (m), 531 (m), 506 (m), 455 (m). R_f (cyclohexane/EtOAc, 4:1) = 0.45 [p-Anisaldehyde]. $[\alpha]_D^{20} = +43.6$ $(\rho = 1.03, CH_2Cl_2).$

Benzyl (2S,4S)-4-(2-(4-bromophenyl)-2-oxoethyl)-2-(tert-butyl)-5-oxooxazolidine-3-carboxylate (6). Synthesized following GP-A using 4-bromobenzoic acid (151 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 10:1) to provide 6 as a yellow oil in 74% yield (175 mg, 0.37 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.73 (d, J = 8.6 Hz, 2H), 7.57 (d, J = 8.6 Hz, 2H), 7.31 (d, J = 1.5 Hz, 3H), 7.24-7.17 (m, 2H), 5.61 (s, 1H), 5.18 (dd, J = 6.9, 5.1 Hz, 1H), 5.10 (d, J = 12.0 Hz, 1H), 5.01 (d, J = 12.0 Hz, 1H), 3.49 (dd, J = 16.2, 6.9 Hz, 1H), 3.33 (dd, J = 16.2, 5.1 Hz, 1H), 1.01 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 193.9, 172.0, 155.6, 135.1, 135.1, 132.2, 129.8, 128.8, 128.8, 128.8, 128.6, 110.0, 96.5, 68.5, 53.8, 41.9, 37.6, 24.9. IR \tilde{v} [cm⁻¹] = 2963 (w), 2874 (w), 1791 (s), 1719 (m), 1688 (m), 1584 (s), 1581 (m), 1457 (m), 1393 (m), 1343 (m), 1286 (m), 1235 (m), 1173 (m), 1120 (m), 1068 (m), 1042 (m), 989 (w), 823 (m), 733 (m), 697 (s), 509 (s), 453 (s). HRMS (ESI) [m/z] calculated for $C_{23}H_{24}BrNNaO_5$ ([M + Na]⁺) 496.0729, found 496.0730. R_f (cyclohexane/EtOAc, 4:1) = 0.62 [p-Anisaldehyde]. $\left[\alpha\right]_{D}^{20} = +40.5$ $(\rho = 0.93, CH_2Cl_2).$

Benzyl (25,45)-2-(tert-butyl)-4-(2-(4-fluorophenyl)-2-oxoethyl)-5oxooxazolidine-3-carboxylate (7). Synthesized following GP-A using 4-fluorobenzoic acid (105 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/ EtOAc, 4:1) to provide 7 as a yellowish oil in 61% yield (127 mg, 0.31 mmol). ¹H NMR (400 MHz, CDCl₃) δ 7.96–7.86 (m, 2H), 7.35– 7.24 (m, 3H), 7.28–7.19 (m, 2H), 7.16–7.05 (m, 2H), 5.62 (s, 1H), pubs.acs.org/joc

5.21 (dd, J = 7.0, 5.0 Hz, 1H), 5.11 (d, J = 12.1 Hz, 1H), 5.03 (d, J = 12.1 Hz, 1H), 3.51 (dd, J = 16.3, 7.0 Hz, 1H), 3.34 (dd, J = 16.2, 5.0 Hz, 1H), 1.01 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 193.3, 172.1, 166.1 (d, ¹ $J_{C-F} = 255.5$ Hz), 155.7, 135.2, 132.8 (d, ⁴ $J_{C-F} = 3.1$ Hz), 131.0 (d, ³ $J_{C-F} = 9.3$ Hz), 128.8, 128.7, 128.5, 116.1 (d, ² $J_{C-F} = 22.0$ Hz), 96.4, 68.5, 53.8, 41.9, 37.6, 24.9. ¹⁹F{¹H} NMR (376 MHz, CDCl₃) δ –104.4. HRMS (FI) [m/z] calculated for C₂₃H₂₄FNNaO₅ ([M + Na]⁺) 436.1531, found 436.1535. IR \tilde{v} [cm⁻¹] = 3067 (w), 3035 (w), 2969 (w), 2913 (w), 2875 (w), 1791 (s), 1719 (s), 1687 (s), 1596 (s), 1507 (w), 1481 (w), 1456 (w), 1392 (m), 1345 (s), 1289 (s), 1230 (s), 1199 (s), 1178 (s), 1156 (s), 1122 (m), 1069 (m), 1042 (s), 1018 (m), 993 (m), 911 (w), 891 (w), 842 (m), 822 (m), 786 (w), 732 (m), 697 (m), 637 (w), 593 (m), 561 (w), 531 (W), 510 (w), 456 (w), 419 (w). R_f (cyclohexane/EtOAc, 4:1) = 0.43 [KMnO₄]. [α]²⁰₂ = +29.5 (ρ = 1.02, CH₂Cl₂).

Benzyl (2S,4S)-2-(tert-butyl)-4-(2-(4-cyanophenyl)-2-oxoethyl)-5oxooxazolidine-3-carboxylate (8). Synthesized following GP-A using 4-cyanobenzoic acid (111 mg, 0.75 mmol, 1.5 equiv.) and irradiating for 48 h. The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide 8 as an offwhite solid in 35% yield (73 mg, 0.17 mmol). ¹H NMR (600 MHz, $CDCl_3$) δ 7.93 (d, J = 8.0 Hz, 2H), 7.72 (d, J = 8.1 Hz, 2H), 7.35– 7.29 (m, 3H), 7.23 (dd, J = 7.2, 2.0 Hz, 2H), 5.62 (s, 1H), 5.19–5.14 (m, 1H), 5.10 (d, J = 11.9 Hz, 1H), 5.04 (d, J = 12.0 Hz, 1H), 3.52 (dd, J = 16.4, 7.0 Hz, 1H), 3.37 (dd, J = 16.3, 5.0 Hz, 1H), 1.01 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 193.8, 171.8, 155.6, 139.3, 135.0, 132.7, 128.9, 128.8, 128.7, 128.6, 117.9, 116.9, 96.6, 68.6, 53.8, 42.2, 37.6, 24.9. HRMS (FI) [m/z] calculated for C₂₄H₂₄N₂NaO₅ $([M + Na]^{+})$ 443.1577, found 443.1580. IR $\tilde{v} [cm^{-1}] = 2964$ (w), 2875 (w), 2230 (w), 1790 (s), 1719 (s), 1695 (s), 1607 (w), 1567 (w), 1498 (w), 1481 (m), 1455 (m), 1394 (s), 1336 (s), 1289 (s), 1269 (s), 1233 (s), 1198 (s), 1174 (s), 1119 (s), 1069 (m), 1041 (s), 1016 (s), 996 (s), 890 (m), 844 (m), 821 (m), 782 (m), 749 (s), 697 (s), 636 (m), 567 (m), 545 (m), 532 (m), 505 (m), 455 (m). R_f (cyclohexane/EtOAc, 4:1) = 0.30 [KMnO₄]. $[\alpha]_{D}^{20}$ = +38.7 (ρ = 0.99, CH_2Cl_2).

Benzyl (25, 45)-2-(tert-butyl)-5-0x0-4-(2-0x0-2-(4-(trifluoromethyl)phenyl)ethyl)oxazolidine-3-carboxylate (9). Synthesized following GP-A using 4-(trifluoromethyl)benzoic acid (142 mg, 0.75 mmol, 1.5 equiv.) and irradiating for 48 h. The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide 9 as a yellow oil in 61% yield (141 mg, 0.30 mmol). The spectroscopic data are consistent with those previously reported.¹¹ ¹H NMR (400 MHz, CDCl₃) δ 7.96 (d, *J* = 8.1 Hz, 2H), 7.69 (d, *J* = 8.2 Hz, 2H), 7.29 (dtt, *J* = 8.3, 5.5, 2.7 Hz, 4H), 7.24–7.19 (m, 2H), 5.63 (s, 1H), 5.18 (dd, *J* = 6.9, 5.1 Hz, 1H), 5.10 (d, *J* = 12.0 Hz, 1H), 3.39 (dd, *J* = 16.2, 5.2 Hz, 1H), 1.02 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) 194.1, 171.9, 155.6, 139.1, 135.1, 134.9 (q, ²*J*_{C-F} = 32.8 Hz), 128.8, 128.7, 128.6, 126.0 (q, ³*J*_{C-F} = 3.7 Hz), 123.6 (q, ¹*J*_{C-F} = 272.6 Hz), 96.5, 68.6, 53.9, 42.2, 37.6, 24.9. ¹⁹F{¹H} NMR (376 MHz, CDCl₃) δ -63.2.

Benzyl (25,45)-2-(tert-butyl)-4-(2-(4-formylphenyl)-2-oxoethyl)-5-oxooxazolidine-3-carboxylate (10). Synthesized following GP-A using 4-formylbenzoic acid (113 mg, 0.75 mmol 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide 10 as a yellow oil in 10% yield (21.2 mg, 0.05 mmol). ¹H NMR (600 MHz, CDCl₃) δ 10.10 (s, 1H), 8.01 (t, *J* = 8.1 Hz, 2H), 7.94 (d, *J* = 8.4 Hz, 2H), 7.30 (dd, *J* = 5.1, 2.1 Hz, 3H), 7.25–7.20 (m, 2H), 5.63 (s, 1H), 5.21 (dd, *J* = 7.0, 5.0 Hz, 1H), 5.11 (d, *J* = 12.0 Hz, 1H), 5.03 (d, *J* = 12.1 Hz, 1H), 3.57 (dd, *J* = 16.3, 7.0 Hz, 1H), 3.41 (dd, *J* = 16.3, 5.0 Hz, 1H), 1.02 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 194.4, 191.5, 171.9, 155.6, 140.5, 139.4, 135.1, 130.0, 128.8, 128.8, 128.6, 96.5, 68.6, 53.8, 42.4, 37.6, 27.1, 24.9. HRMS (ESI) [*m*/*z*] calculated for C₂₄H₂₅NNaO₆ ([M + Na]⁺) 446.1574, found 446.1573. *R_f* (cyclohexane/EtOAc, 4:1) = 0.16 [*p*-Anisaldehyde].

(25,45)-Benzyl 4-(2-(benzo[d][1,3]dioxol-5-yl)-2-oxoethyl)-2-(tert-butyl)-5-oxooxazolidine-3-carboxylate (11). Synthesized following GP-A using piperonylic acid (124.5 mg, 0.75 mmol, 1.5

equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide 11 as a white solid in 95% yield (208 mg, 0.47 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.48 (dd, J = 8.2, 1.8 Hz, 1H), 7.37 (d, J = 1.8 Hz, 1H), 7.32-7.28 (m, 3H), 7.23 (dd, J = 6.6, 2.9 Hz, 2H), 6.81 (d, J =8.1 Hz, 1H), 6.04 (q, J = 1.4 Hz, 2H), 5.61 (s, 1H), 5.21 (dd, J = 6.9, 5.0 Hz, 1H), 5.11 (d, J = 12.1 Hz, 1H), 5.02 (d, J = 12.1 Hz, 1H), 3.46 (dd, J = 16.1, 7.0 Hz, 1H), 3.29 (dd, J = 16.1, 5.1 Hz, 1H), 1.01 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 192.8, 172.2, 155.7, 152.2, 148.5, 135.3, 131.3, 128.7, 128.6, 128.5, 124.6, 108.1, 108.0, 102.1, 96.3, 68.3, 53.9, 41.7, 37.5, 24.9. HRMS (ESI) [m/z] calculated for $C_{24}H_{25}NNaO_7$ ([M + Na]⁺) 462.1520, found 462.1523. IR \tilde{v} $[cm^{-1}] = 2963$ (w), 1791 (s), 1718 (m), 1676 (m), 1604 (m), 1485 (s), 1443 (s), 1393 (s), 1345 (m), 1235 (w), 1176 (m), 1110 (s), 1034 (s), 930 (m), 808 (w). 733 (s), 697 (s). R_f (cyclohexane/ EtOAc, 4:1) = 0.32 [p-Anisaldehyde]. $[\alpha]_D^{20}$ = +41.5 (ρ = 0.95, CH₂Cl₂).

Benzyl (2S,4S)-2-(tert-butyl)-4-(2-(2-chlorophenyl)-2-oxoethyl)-5-oxooxazolidine-3-carboxylate (12). Synthesized following GP-A using 2-chlorobenzoic acid (117.4 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 10:1) to provide 12 as a yellow oil in 95% yield (203 mg, 0.47 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.47 (d, J = 7.7 Hz, 1H), 7.41-7.39 (m, 2H), 7.33 (s, 5H), 7.32-7.28 (m, 1H), 5.61 (s, 1H), 5.22 (t, J = 6.1 Hz, 1H), 5.20-5.12 (m, 2H), 3.51-3.48 (m, 2H), 0.97 (s, 9H). ${}^{13}C{}^{1}H$ NMR (151 MHz, CDCl₃) δ 197.4, 172.1, 155.6, 138.5, 135.3, 132.4, 131.3, 130.7, 130.0, 128.8, 128.7, 128.6, 127.2, 96.4, 68.5, 53.7, 45.8, 37.6, 24.9. HRMS (ESI) [m/z]calculated for $C_{23}H_{24}CINNaO_5$ ([M + Na]⁺) 452.1215, found 452.1235. IR \tilde{v} [cm⁻¹] = 2968 (w), 1791 (s), 1716 (s), 1589 (s), 1392 (m), 1345 (m), 1284 (m), 1176 (m), 1120 (m), 1076 (m), 1040 (m), 976 (m), 757 (m), 697 (m), 697 (s), 633 (m). R_f (cyclohexane/EtOAc, 4:1) = 0.45 [p-Anisaldehyde]. $\left[\alpha\right]_{D}^{20}$ = +47.2 $(\rho = 1.03, CH_2Cl_2).$

Benzyl (2S,4S)-4-(2-(2-acetoxyphenyl)-2-oxoethyl)-2-(tert-butyl)-5-oxooxazolidine-3-carboxylate (13). Synthesized following GP-A using 2-acetoxybenzoic acid (135 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 10:1) to provide 13 as a yellow oil in 92% yield (208 mg, 0.45 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.67 (dd, J = 7.8, 1.7 Hz, 1H), 7.56–7.52 (m, 1H), 7.33–7.27 (m, 5H), 7.15 (dd, J = 8.1, 1.1 Hz, 1H), 5.60 (s, 1H), 5.19 (dd, J = 7.0, 4.8 Hz, 1H), 5.15 (d, J = 12.1 Hz, 1H), 5.06 (d, J = 12.1 Hz, 1H), 3.47 (dd, J = 16.7, 7.0 Hz, 1H), 3.31 (dd, J = 16.7, 4.8 Hz, 1H), 2.31 (s, 3H), 0.98 (s, 9H). ${}^{13}C{}^{1}H$ NMR (151 MHz, CDCl₃) δ 194.6, 172.1, 169.4, 155.6, 149.2, 135.4, 133.6, 130.5, 129.9, 128.8, 128.7, 128.5, 126.2, 124.1, 96.3, 68.4, 53.5, 44.9, 37.6, 24.9, 21.2. HRMS (ESI) [m/z] calculated for C₂₅H₂₇NNaO₇ ([M + Na]⁺) 476.1681, found 476.1680. IR $\tilde{v} [cm^{-1}] = 2965$ (w), 2875 (m), 1791 (m), 1718 (m), 1603 (s), 1346 (m), 1286 (m), 1177 (w), 1120 (m), 1041 (m), 909 (s), 7534 (m), 697 (s), 503 (s). R_f (cyclohexane/EtOAc, 4:1) = 0.37 [p-Anisaldehyde]. $[\alpha]_{D}^{20} = +41.5 \ (\rho = 1.01, \text{ CH}_2\text{Cl}_2).$

Benzyl (2S,4S)-2-(tert-butyl)-4-(2-(3-(5-(2-fluorophenyl)-1,2,4-oxadiazol-3-yl)phenyl)-2-oxoethyl)-5-oxooxazolidine-3-carboxylate (14). Synthesized following GP-A using 3-[5-(2-fluorophenyl)-1,2,4oxadiazol-3-yl] benzoic acid (213 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide 14 as a yellow oil in 87% yield (243.7 mg, 0.44 mmol). ¹H NMR (400 MHz, CDCl₃) δ 8.69 (t, J = 1.7 Hz, 1H), 8.40 (dt, J = 7.8, 1.4 Hz, 1H), 8.24 (ddd, J = 7.8, 7.0, 1.8 Hz, 1H), 8.09–8.03 (m, 1H), 7.62 (t, J = 7.8 Hz, 2H), 7.26 (t, J = 1.3 Hz, 4H), 5.64 (s, 1H), 5.27 (dd, J = 6.8, 5.1 Hz, 1H), 5.16-4.99 (m, 2H), 3.61 (dd, J = 16.4, 6.8 Hz, 1H), 3.49 (dd, J = 16.4, 5.1 Hz, 1H), 1.04 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 194.3, 173.3 $(d, {}^{3}J_{C-F} = 4.4 \text{ Hz}), 172.1, 168.1, 162.3 (d, {}^{1}J_{C-F} = 260.8 \text{ Hz}), 155.7,$ 137.1, 135.2, 135.0 (d, ${}^{3}J_{C-F} = 8.6$ Hz), 132.3, 131.1, 130.8, 129.6, 128.7, 128.7, 128.5, 127.8 (d, ${}^{3}J_{C-F}$ = 5.0 Hz), 127.4, 124.9 (d, ${}^{3}J_{C-F}$ = 3.8 Hz), 117.5 (d, ${}^{2}J_{C-F} = 20.9$ Hz), 112.8(d, ${}^{3}J_{C-F} = 11.3$ Hz), 96.5, 68.5, 53.8, 42.0, 37.6, 24.9. $^{19}F{^1H}$ NMR (376 MHz, CDCl₃) δ -108.1. HRMS (ESI) [m/z] calculated for $C_{31}H_{28}FN_3NaO_6$ ([M +

Na]⁺) 580.1868, found 580.1854 IR \tilde{v} [cm⁻¹] = 2962 (w), 2358 (w), 1791 (s), 1718 (m), 1621 (s), 1459 (m), 1393 (m), 1344 (m), 1289 (m), 1176 (m), 1121 (m), 1041 (m), 753 (s), 695 (m). R_f (cyclohexane/EtOAc, 4:1) = 0.25 [*p*-Anisaldehyde]. [α]_D²⁰ = +36.9 (ρ = 0.95, CH₂Cl₂).

Benzyl (25,45)-2-(tert-butyl)-5-oxo-4-(2-oxo-2-(3-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl)ethyl)oxazolidine-3-carboxylate (15). Synthesized following GP-A using 4-carboxylphenylboronic acid pinacol ester (186 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/ EtOAc, 4:1) to provide 15 as a white solid in 76% yield (198 mg, 0.38 mmol). ¹H NMR (600 MHz, CDCl₃) δ 8.31 (s, 1H), 8.03-7.98 (m, 2H), 7.47 (d, J = 7.7 Hz, 1H), 7.29-7.27 (m, 3H), 7.23-7.18 (m, 2H), 5.61 (s, 1H), 5.24 (dd, J = 6.7, 5.2 Hz, 1H), 5.11 (d, J = 12.1 Hz, 1H), 4.96 (d, J = 12.1 Hz, 1H), 3.55 (dd, J = 16.3, 6.8 Hz, 1H), 3.44 (dd, J = 16.3, 5.2 Hz, 1H), 1.36 (s, 12H), 1.03 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 195.1, 172.2, 155.7, 139.9, 136.0, 135.3, 134.6, 131.0, 128.7, 128.6, 128.5, 128.3, 96.4, 84.4, 68.3, 53.9, 41.9, 37.6, 25.1, 25.0. HRMS (ESI) [m/z] calculated for C₂₉H₃₆BNNaO₇ $([M + Na]^+)$ 544.2480, found 544.2477. IR $\tilde{v} [cm^{-1}] = 2974$ (w), 2931 (w), 2358 (w), 1793 (s), 1720 (s), 1687 (m), 1482 (m), 1390 (m), 1348 (m), 1287 (m), 1140 (m), 1041 (m), 697 (s). R_f (cyclohexane/EtOAc, 4:1) = 0.5 [p-Anisaldehyde]. $[\alpha]_{D}^{20} = +37.1$ $(\rho = 1.14, CH_2Cl_2).$

Benzyl (2S,4S)-2-(tert-butyl)-5-oxo-4-(2-oxo-2-(pyridin-3-yl)ethyl)oxazolidine-3-carboxylate (16). Synthesized following GP-A using nicotinic acid (92.25 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/ EtOAc, 4:1) to provide 16 as a yellow oil in 45% yield (110 mg, 0.28 mmol). ¹H NMR (400 MHz, CDCl₃) δ 9.11-9.08 (m, 1H), 8.80 (dd, *J* = 5.0, 1.7 Hz, 1H), 8.19 (dt, *J* = 8.0, 1.9 Hz, 1H), 7.44 (ddd, *J* = 8.0, 4.9, 0.8 Hz, 1H), 7.32-7.28 (m, 3H), 7.25-7.22 (m, 2H), 5.62 (s, 1H), 5.21 (dd, J = 6.9, 5.0 Hz, 1H), 5.07 (q, J = 12.0 Hz, 2H), 3.54 (dd, J = 16.4, 6.9 Hz, 1H), 3.39 (dd, J = 16.4, 5.0 Hz, 1H), 1.01 (s, 1)9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 193.6, 171.9, 155.6, 153.3, 149.1, 136.2, 135.0, 132.0, 128.8, 128.8, 128.6, 124.1, 96.5, 68.6, 53.6, 42.1, 37.6, 24.9. HRMS (ESI) [m/z] calculated for C₂₂H₂₅N₂O₅ ([M + Na]⁺) 397.1757, found 397.1758. IR $\tilde{v} [cm^{-1}] = 3035 (\tilde{w}), 1790$ (s), 1719 (m), 1480 (m), 1392 (m), 1343 (m), 1285 (m), 1176 (m), 1120 (s), 1040 (m), 1020 (m), 779 (m), 698 (m). R_f (cyclohexane/ EtOAc, 1:1) = 0.4 [p-Anisaldehyde]. $[\alpha]_{D}^{20} = +42.3$ ($\rho = 1.04$, CH_2Cl_2).

Benzyl (2S,4S)-2-(tert-butyl)-4-(2-(1,3-dimethyl-1H-pyrazolo[3,4b]pyridin-5-yl)-2-oxoethyl)-5-oxooxazolidine-3-carboxylate (17) and Benzyl (2S,4S)-2-(tert-butyl)-4-(2-(4-chloro-1,3-dimethyl-1Hpyrazolo[3,4-b]pyridin-5-yl)-2-oxoethyl)-5-oxooxazolidine-3-carboxylate (17'). Synthesized following GP-B using 4-Chloro-1,3dimethylpyrazolo[3,4-b]pyridine-5-carboxylic acid (169 mg, 0.75 mmol 1.5 equiv.), and irradiating for 48 h. The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 2:1) to provide 17 a yellow foam in 39% (98 mg, 0.20 mmol) and 17' as a yellow solid in 18% yield (45 mg, 0.10 mmol). Spectroscopic data for 17: ¹H NMR (600 MHz, CDCl₃) δ 9.05 (d, J = 2.0 Hz, 1H), 8.49 (d, J = 2.0 Hz, 1H), 7.23 (q, J = 2.9 Hz, 3H), 7.19 (dd, J = 6.8, 3.1 Hz, 2H), 5.64 (s, 1H), 5.26 (dd, J = 6.8, 5.2 Hz, 1H), 5.11–5.01 (m, 2H), 4.11 (s, 3H), 3.59 (dd, J = 16.0, 6.8 Hz, 1H), 3.43 (dd, J = 16.0, 5.2 Hz, 1H), 2.59 (s, 3H), 1.04 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 193.2, 172.0, 155.7, 152.2, 149.5, 143.2, 135.0, 130.8, 128.7, 128.6, 128.5, 125.1, 114.8, 96.5, 68.5, 54.0, 42.0, 37.6, 33.9, 24.9, 12.6. HRMS (ESI) [m/z] calculated for $C_{25}H_{28}N_4NaO_5$ ([M + Na]+) 487.1957, found 487.1952. IR $\tilde{v} [cm^{-1}] = 2961$ (s), 2926 (s), 1790 (w), 1719 (w), 1678 (w), 1599 (w), 1564 (m), 1520 (m), 1478 (m), 1392 (w), 1281 (w), 1177 (w), 1121 (m), 1041 (w), 983 (w), 748 (m), 697 (w), 578 (m), 503 (m), 456 (m), 428 (m). R_f (cyclohexane/ EtOAc, 1:1) = 0.33 [p-Anisaldehyde]. $[\alpha]_{D}^{20} = +40.1$ ($\rho = 0.92$, CH_2Cl_2).

Spectroscopic data for 17': ¹H NMR (400 MHz, CDCl₃) δ 8.67 (s, 1H), 7.40–7.29 (m, 1H), 7.27 (tdd, *J* = 4.6, 3.4, 2.0 Hz, 4H), 5.63 (s, 1H), 5.23 (dd, *J* = 6.9, 5.4 Hz, 1H), 5.14 (d, *J* = 1.0 Hz, 2H), 4.07 (s, 3H), 3.60 (dd, *J* = 16.3, 6.9 Hz, 1H), 3.53 (dd, *J* = 16.3, 5.5 Hz, 1H),

2.73 (s, 3H), 1.00 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 195.0, 171.9, 155.6, 152.3, 151.0, 142.9, 138.0, 135.1, 128.7, 128.7, 128.6, 126.1, 113.2, 96.5, 68.6, 54.2, 45.9, 37.6, 34.0, 24.9, 15.1. HRMS (ESI) [*m*/*z*] calculated for C₂₅H₂₇ClN₄NaO₅ ([M + Na]⁺) 521.1562, found 521.1560. IR \tilde{v} [cm⁻¹] = 2962 (s), 2874 (s), 1791 (m), 1717 (w), 1582 (s), 1547 (m), 1515 (w), 1456 (w), 1391 (m), 1332 (w), 1285 (w), 1234 (w), 1175 (w), 1117 (m), 1038 (w), 729 (w), 697 (w), 582 (m), 503 (m). *R*_f (cyclohexane/EtOAc, 1:1) = 0.43 [*p*-Anisaldehyde]. [α]_D²⁰ = +38.1 (ρ = 0.96, CH₂Cl₂).

Benzyl (2S,4S)-2-(tert-butyl)-5-oxo-4-(2-oxo-2-(1H-pyrrol-2-yl)ethyl)oxazolidine-3-carboxylate (20). Synthesized following GP-A using 1H-pyrrole-2-carboxylic acid (84 mg, 0.75 mmol, 1.5 equiv.), and irradiating for 48 h. The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 5:1) to provide 20 as a dark yellow oil in 20% yield (39 mg, 0.10 mmol). ¹H NMR (600 MHz, CDCl₃) δ 9.42 (br, 1H), 7.40–7.27 (m, 3H), 7.24 (dq, J = 6.5, 4.2 Hz, 2H), 7.00 (td, J = 2.7, 1.2 Hz, 1H), 6.87 (ddd, J = 3.8, 2.4, 1.3 Hz, 1H), 6.26 (dt, J = 3.9, 2.5 Hz, 1H), 5.61 (s, 1H), 5.13 (t, J = 6.3 Hz, 1H), 5.09 (d, J = 12.1 Hz, 1H), 5.02 (d, J = 12.1 Hz, 1H), 3.33 (dd, J = 15.3, 6.6 Hz, 1H), 3.20 (dd, J = 15.3, 5.9 Hz, 1H), 1.01 (s, 10.1 Hz)9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 184.7, 172.0, 155.7, 135.3, 131.5, 128.7, 128.6, 128.5, 125.5, 116.7, 111.0, 96.5, 68.4, 54.4, 41.3, 37.5, 25.0. HRMS (ESI) [m/z] calculated for C₂₁H₂₄N₂NaO₅ ([M + Na]⁺) 407.1577, found 407.1587. IR v [cm⁻¹] = 3290(w), 2968 (w), 2875 (w), 2254 (w), 1790 (m), 1715 (s), 1640 (m), 1547 (w), 1481 (w), 1397 (s), 1345 (m), 1334 (m), 1292 (s), 1231 (m), 1179 (m), 1109 (s), 1067 (m), 1041 (s), 1017 (m), 979 (m), 909 (m), 727 (s), 697 (s), 648 (w), 602 (w), 582 (w), 512 (w), 455 (w). R_t (cyclohexane/EtOAc, 4:1) = 0.19 [p-Anisaldehyde]. $[\alpha]_D^{20} = +32.9$ $(\rho = 1.00, CH_2Cl_2).$

Benzyl (2S,4S)-2-(tert-butyl)-4-(2-(furan-2-yl)-2-oxoethyl)-5-oxooxazolidine-3-carboxylate (21). Synthesized following GP-A using furan-2-carboxylic acid (84 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 1:1) to provide 21 as a yellow oil in 58% yield (90 mg, 0.23 mmol). ¹H NMR (400 MHz, CDCl₃) δ 7.55 (dd, J = 1.7, 0.8 Hz, 1H), 7.32 (dd, J = 5.0, 2.0 Hz, 3H), 7.28–7.24 (m, 2H), 7.19 (dd, J = 3.6, 0.8 Hz, 1H), 6.53 (dd, J = 3.6, 1.7 Hz, 1H), 5.60 (s, 1H), 5.17-5.10 (m, 2H), 5.00 (d, J = 12.1 Hz, 1H), 3.43 (dd, J = 12.1 Hz, 1H), 3.415.7, 6.5 Hz, 1H), 3.24 (dd, J = 15.7, 6.0 Hz, 1H), 1.01 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 184.0, 171.9, 155.6, 152.5, 146.6, 135.3, 128.8, 128.7, 128.5, 117.4, 112.7, 96.5, 68.4, 53.8, 41.8, 37.5, 24.9. HRMS (ESI) [m/z] calculated for C₂₁H₂₃NNaO₆ ([M + Na]⁺) 408.1412, found 408.1418. IR $\tilde{v} [cm^{-1}] = 2967$ (w), 1790 (s), 1719 (s), 1676 (s), 1569 (s), 1467 (s), 1392 (s), 1345 (s), 1296 (s), 1178 (s), 1121 (s), 1017 (m), 978 (m), 756 (w), 697 (s), 594 (s), 507 (s). R_f (cyclohexane/EtOAc, 4:1) = 0.3 [p-Anisaldehyde]. $[\alpha]_D^{20}$ = +41.4 ($\rho = 0.95$, CH₂Cl₂).

Benzyl (2S,4S)-2-(tert-butyl)-5-oxo-4-(2-oxo-2-(thiophen-2-yl)ethyl)oxazolidine-3-carboxylate (22). Synthesized following GP-A using thiophene-2-carboxylic acid (96 mg, 0.75 mmol 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide 22 as a yellow oil in 71% yield (134 mg, 0.35 mmol). $^1\mathrm{H}$ NMR (600 MHz, CDCl₃) δ 7.71– 7.64 (m, 2H), 7.33–7.27 (m, 3H), 7.24 (dd, J = 6.7, 2.9 Hz, 2H), 7.12 (dd, J = 4.9, 3.8 Hz, 1H), 5.61 (s, 1H), 5.18–5.09 (m, 2H), 5.00 (d, J = 12.1 Hz, 1H), 3.47 (dd, J = 15.7, 6.6 Hz, 1H), 3.30 (dd, J = 15.6, 5.6 Hz, 1H), 1.01 (s, 9H). $^{13}C{^{1}H}$ NMR (151 MHz, CDCl₃) δ 187.6, 171.9, 155.7, 143.7, 135.3, 134.5, 132.3, 128.8, 128.7, 128.5, 128.4, 96.5, 68.4, 54.1, 42.6, 37.6, 24.9. HRMS (ESI) [m/z] calculated for $C_{21}H_{23}NNaO_5S$ ([M + Na]⁺) 424.1190, found 424.1189. IR \tilde{v} [cm⁻¹] = 2962 (s), 2874 (s), 1791 (m), 1717 (w), 1582 (s), 1547 (m), 1515 (w), 1456 (w), 1391 (m), 1332 (w), 1285 (w), 1234 (w), 1175 (w), 1117 (m), 1038 (w), 729 (w), 697 (w), 582 (m), 503 (m). R_f (cyclohexane/EtOAc, 4:1) = 0.20 [p-Anisaldehyde]. $[\alpha]_D^{20} = +26.6 (\rho)$ $= 0.93, CH_2Cl_2$).

Benzyl (25,45)-2-(tert-butyl)-4-(2-(cyclohex-1-en-1-yl)-2-oxoethyl)-5-oxooxazolidine-3-carboxylate (23). Synthesized following GP-A using 1-cyclohexene-1-carboxylic acid (85.8 μ L, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column pubs.acs.org/joc

chromatography (cyclohexane/EtOAc, 20:1–7:1) to provide **23** as a yellow oil in 35% yield (70 mg, 0.18 mmol). ¹H NMR 600 MHz, CDCl₃) δ 7.37–7.33 (m, 3H), 7.32 (d, *J* = 2.0 Hz, 3H), 6.84–6.81 (m, 1H), 5.58 (s, 1H), 5.16–5.09 (m, 3H), 3.23 (dd, *J* = 16.1, 7.3 Hz, 1H), 3.04 (dd, *J* = 16.1, 4.7 Hz, 1H), 2.25–2.20 (m, 4H), 1.65–1.56 (m, 6H), 0.98 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 128.6, 128.3, 96.1, 68.2, 53.7, 40.3, 37.4, 24.8, 21.5. HRMS (FI) [*m*/*z*] calculated for C₂₃H₂₉NNaO₅ ([M + Na]⁺) 422.1941, found 422.1938. IR \tilde{v} [cm⁻¹] = 2958 (w), 2935 (w), 2872 (w), 1972 (s), 1719 (s), 1668 (s), 1497 (m), 1452 (m), 1391 (m). 1345 (m). 1289 (m). 1173 (m), 1235 (m), 1121 (m). 1040 (s), 985 (m), 736 (w), 697 (s). *R*_f (cyclohexane/EtOAc, 4:1) = 0.50 [*p*-Anisaldehyde]. [α]_D²⁰ = +37.7 (ρ = 1.03, CH₂Cl₃).

Benzyl (2S,4S)-2-(tert-butyl)-4-(2-(3,4-dihydro-2H-pyran-5-yl)-2oxoethyl)-5-oxooxazolidine-3-carboxylate (24). Synthesized following GP-A using 3,4-dihydro-2H-pyran-5-carboxylic acid (96 mg, 0.75 mmol, 1.5 equiv.). The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide 24 as a yellow oil in 48% yield (39 mg, 0.24 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.53 (s, 1H), 7.37–7.31 (m, 5H), 5.58 (s, 1H), 5.16 (dd, J = 7.3, 4.8 Hz, 1H), 5.14 (dd, J = 12.2, 9.7 Hz, 2H), 4.05 (t, J = 5.3 Hz, 2H), 3.08 (dd, J = 15.7, 7.4 Hz, 1H), 2.94 (dd, J = 15.7, 4.8 Hz, 1H), 2.34-2.18 (m, 2H), 1.87–1.79 (m, 2H), 0.98 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 192.8, 172.3, 157.1, 155.8, 135.5, 128.8, 128.6, 128.4, 116.9, 96.2, 68.3, 67.3, 53.9, 40.0, 37.5, 24.9, 21.1, 18.5. HRMS (FI) [m/z] calculated for C₂₂H₂₇NNaO₆ ($[M + Na]^+$) 424.1731, found 424.1739. IR \tilde{v} [cm⁻¹] = 3065 (w), 3034 (w), 2961 (w), 2879 (w), 1791 (s), 1717 (s), 1656 (m), 1617 (s), 1481 (w), 1465 (w), 1450 (w), 1393 (m), 1345 (m), 1329 (s), 1289 (s), 1267 (m), 1230 (s), 1171 (s), 1122 (m), 1086 (m), 1041 (s), 1005 (s), 985 (s), 963 (m), 930 (m), 908 (m), 852 (m), 779 (m), 736 (m), 697 (s), 676 (m), 637 (w), 581 (w), 557 (w), 531 (w), 508 (m), 449 (m). R_f (cyclohexane/ EtOAc, 4:1) = 0.26 [p-Anisaldehyde]. $[\alpha]_{D}^{20}$ = +39.2 (ρ = 1.04, CH₂Cl₂).

Benzyl (2S,4S)-4-(2-(1-(tert-butoxycarbonyl)-1,2,5,6-tetrahydropyridin-3-yl)-2-oxoethyl)-2-(tert-butyl)-5-oxooxazolidine-3-carboxylate (26). Synthesized following GP-A using 1-(tert-butoxycarbonyl)-1,2,5,6-tetrahydropyridine-3-carboxylic acid (171 mg, 0.75 mmol, 1.5 equiv. The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 4:1) to provide 26 as an offwhite solid in 68% yield (170 mg, 0.34 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.35–7.29 (m, 5H), 6.89 (s, 1H), 5.57 (s, 1H), 5.18–5.08 (m, 3H), 5.09–5.06 (m, 1H), 4.09 (s, 1H), 3.45 (s, 2H), 3.23 (dd, J = 16.0, 7.2 Hz, 1H), 3.07-3.00 (m, 1H), 2.35-2.27 (m, 2H), 1.46 (s, 9H), 0.97 (s, 9H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 193.6, 172.1, 155.6, 154.9, 138.8, 137.0, 135.3, 128.8, 128.7, 128.5, 96.3, 80.2, 77.4, 77.2, 76.9, 68.4, 53.7, 42.3, 40.3, 38.6, 37.5, 28.5, 25.9, 24.8. HRMS (FI) [m/z] calculated for $C_{27}H_{36}N_2NaO_7$ ($[M + Na]^+$) 523.2415, found 523.2410. IR \tilde{v} [cm⁻¹] = 2972 (w), 2935 (w), 2874 (w), 1792 (m), 1720 (m), 1693 (s), 1673 (s), 1621 (w), 1479 (m), 1455 (m), 1391 (m), 1365 (m), 1342 (m), 1285 (s), 1234 (s), 1159 (s), 1111 (s), 1068 (m), 1040 (s), 985 (m), 890 (w), 865 (m), 826 (w), 766 (m), 743 (m), 697 (m), 637 (w), 593 (w), 582 (w), 508 (m), 455 (m). R_f (cyclohexane/EtOAc, 4:1) = 0.20 [KMnO₄]. $[\alpha]_D^{20}$ = +37.1 (ρ $= 1.00, CH_2Cl_2).$

Benzyl 2-(tert-butyl)-4-(2-(4,5-dihydrofuran-3-yl)-2-oxoethyl)-5oxooxazolidine-3-carboxylate (27). Synthesized following GP-A using 4,5-dihydro-furan-3-carboxylic acid (86 mg, 0.75 mmol 1.5 equiv.), and irradiating for 48 h. The crude product was purified by flash column chromatography (cyclohexane/EtOAc, 2:1) to provide 27 as a yellow oil in 31% yield (60 mg, 0.15 mmol). ¹H NMR (400 MHz, CDCl₃) δ 7.40–7.31 (m, 5H), 7.26–7.25 (m, 1H), 5.59 (s, 1H), 5.15 (s, 2H), 5.08 (dd, *J* = 7.1, 5.3 Hz, 1H), 4.56–4.46 (m, 2H), 3.08 (dd, *J* = 15.2, 7.1 Hz, 1H), 2.95 (dd, *J* = 15.2, 5.3 Hz, 1H), 2.87– 2.70 (m, 2H), 0.98 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 189.6, 172.1, 158.5, 155.8, 135.5, 128.8, 128.7, 128.5, 120.4, 96.4, 73.8, 68.4, 54.2, 42.4, 37.5, 27.4, 24.9. HRMS (ESI) [*m*/*z*] calculated for C₂₁H₂₅NNaO₆ ([M + Na]⁺) 410.1585, found 410.1574. IR \tilde{v} [cm⁻¹] = 2968 (s), 1790 (w), 1716 (w), 1648 (m), 1604 (w), 1392 (m), 1334 (m), 1291 (m), 1178 (m), 1129 (w), 1042 (m), 910 (m),

728 (w), 697 (w), 456 (m). R_f (cyclohexane/EtOAc, 1:1) = 0.33 [*p*-Anisaldehyde]. $[\alpha]_D^{20}$ = +35.2 (ρ = 1.05, CH₂Cl₂).

Derivatization Reactions. General Procedure C (**GP-C**). In a 4 mL vial, the corresponding oxazolidinone was dissolved in a mixture of 1,4-dioxane (1.0 mL) and conc. HCl (2.0 mL), and stirred at 80 °C in an oil bath for 2 h, monitoring by TLC. Afterwards, the reaction was concentrated *in vacuo*. To the resulting solid, cyclohexane (3 × 2.0 mL) was added and evaporated *in vacuo* to azeotrope any water residues, affording the desired α -amino acid salts.

(S)-2-Amino-4-(2-hydroxyphenyl)-4-oxobutanoic acid hydrochloride salt (28). Synthesized following GP-C using 12 (62.4 mg, 0.14 mmol) to afford the desired α -amino acid salt 28 as an off-yellow solid in 97% yield (35.5 mg, 0.136 mmol). ¹H NMR (400 MHz, D₂O) δ 7.98 (dd, *J* = 8.1, 1.6 Hz, 1H), 7.70 (ddd, *J* = 8.4, 7.3, 1.6 Hz, 1H), 7.18–7.07 (m, 2H), 4.53 (t, *J* = 5.3 Hz, 1H), 3.98 (d, *J* = 5.3 Hz, 2H). ¹³C{¹H} NMR (101 MHz, D₂O) δ 204.5, 173.9, 162.1, 139.4, 132.5, 122.0, 120.8, 119.6, 50.9, 40.2. HRMS (ESI) [*m*/*z*] calculated for C₁₀H₁₁ClNO₄ ([M]⁻) 244.0383, found 244.0382. [α]_D²⁰ = +26.7 (ρ = 0.30, MeOH).

(S)-2-Amino-4-(2-chlorophenyl)-4-oxobutanoic acid hydrochloride salt (29). Synthesized following GP-C using 13 (51.3 mg, 0.11 mmol) to afford the desired α -amino acid salt 29 as an off-brown solid in quantitative yield (27.0 mg, 0.11 mmol). ¹H NMR (400 MHz, D₂O) δ 7.80 (dt, *J* = 7.7, 1.1 Hz, 1H), 7.69–7.61 (m, 2H), 7.55 (dt, *J* = 7.7, 4.3 Hz, 1H), 4.50 (t, *J* = 5.3 Hz, 1H), 3.90 (d, *J* = 5.3 Hz, 2H). The characterization data matches the reported literature.²²

(*S*)-2-Amino-4-oxo-4-(pyridin-3-yl)butanoic acid hydrochloride salt (**30**). Synthesized following **GP-C** using 7 (78.0 mg, 0.19 mmol) to afford the desired α -amino acid salt **30** as an off-brown solid in 95% yield (45.0 mg, 0.18 mmol). ¹H NMR (400 MHz, D₂O) δ 8.15–8.09 (m, 2H), 7.37–7.29 (m, 2H), 4.45 (dd, *J* = 5.8, 4.7 Hz, 1H), 3.93– 3.83 (m, 2H). The characterization data matches the reported literature.^{4b}

Procedure for the Synthesis of Methyl 2-((tert-Butoxycarbonyl)amino)-4-oxo-4-phenylbutanoate for e.r. Measurement. In a 4 mL vial, 1 was dissolved in a mixture of 1,4-dioxane (1.0 mL) and conc. HCl (2.0 mL) and stirred at 80 °C in an oil bath for 2 h, monitoring by TLC. Afterward, the reaction was concentrated in vacuo. To resulting solid, cyclohexane $(3 \times 2.0 \text{ mL})$ was added and evaporated in vacuo to azeotrope any water residues, affording the desired salt as an off-white solid. This was dissolved in 2,2-DMP (0.1 mL) and conc. HCl (0.3 mL) was added. The resulting suspension was stirred at room temperature for 18 h. The reaction was then concentrated in vacuo. The resulting oil was dissolved in MeOH (2.0 mL) and NaHCO₃ (94.9 mg) and Boc₂O (97.5 mg) were added. The reaction was stirred at 50 °C for 18 h. After cooling to room temperature, the reaction was quenched with sat. aq. NH₄Cl (2.0 mL) and the organic phase was extracted with EtOAc (3 \times 2.0 mL). The combined organic layers were dried over Na2SO4 and concentrated in vacuo. The crude product was purified by flash column chromatography (cyclohexane/EtOAc 4:1) to give the final product in 62% yield and 92:8 e.r. The spectroscopic data are consistent with those previously reported.²³ ¹H NMR (600 MHz, CDCl₃) δ 7.94 (dd, J = 8.4, 1.3 Hz, 2H), 7.59 (t, J = 7.4 Hz, 1H), 7.47 (dd, J = 8.3, 7.4 Hz, 2H), 5.61 (d, J = 8.8 Hz, 1H), 4.75-4.64 (m, J)1H), 3.74 (s, 3H), 3.72 (dd, J = 18.0, 4.3 Hz, 1H), 3.53 (dd, J = 18.0, 4.2 Hz, 1H), 1.44 (s, 9H). The enantiomeric excess was determined by HPLC (CHIRAL PAK IA, heptane/ethanol 98:2 (v/v), λ = 220 nm, flow rate: 0.8 mL/min); $t_{major} = 41.866 \text{ min}$, $t_{minor} = 45.837 \text{ min}$.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.joc.0c02951.

Emission spectra of LEDs; optimization and mechanistic studies; 1H , $^{13}C\{^1H\}$, and $^{19}F\{^1H\}$ NMR spectra; and HPLC chromatogram (PDF)

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Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This work was supported by the Fonds der Chemischen Industrie (Liebig fellowship to A.G.S. and Ph.D. scholarship to F.J.A.T.), by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) 443074366, and the Bergische Universität Wuppertal (BUW). We thank Dr. Lisa Candish and Jun. Prof. Matthew N. Hopkinson for fruitful discussions and proof reading of the manuscript. Umicore A.G. is acknowledged for its generous donation of materials. Prof. Stefan Kirsch (BUW) is greatly acknowledged for his continuous support.

REFERENCES

(1) Lin, W.; He, Z.; Zhang, H.; Zhang, X.; Mi, A.; Jiang, Y. Amino Acid Anhydride Hydrochlorides as Acylating Agents in Friedel-Crafts Reaction: A Practical Synthesis of l-Homophenylalanine. *Synthesis* **2001**, 2001, 1007.

(2) Berkeš, D.; Kolarovič, A.; Považanec, F. Stereoselective sodium borohydride reduction, catalyzed by manganese(II) chloride, of γ -oxo- α -amino acids. A practical approach to syn- γ -hydroxy- α -amino acids. *Tetrahedron Lett.* **2000**, *41*, 5257.

(3) (a) Ďuriš, A.; Wiesenganger, T.; Moravčíková, D.; Baran, P.; Kožíšek, J.; Daïch, A.; Berkeš, D. Expedient and Practical Synthesis of CERT-Dependent Ceramide Trafficking Inhibitor HPA-12 and Its Analogues. Org. Lett. 2011, 13, 1642. (b) Ďuriš, A.; Berkeš, D.; Jakubec, P. Stereodivergent synthesis of cyclic γ-aminobutyric acid -GABA analogues. Tetrahedron Lett. 2019, 60, 480.

(4) (a) Dardir, A. H.; Hazari, N.; Miller, S. J.; Shugrue, C. R. Palladium-Catalyzed Suzuki-Miyaura Reactions of Aspartic Acid Derived Phenyl Esters. Org. Lett. 2019, 21, 5762. (b) Golubev, A. S.; Sewald, N.; Burger, K. Synthesis of γ -oxo α -amino acids from L-aspartic acid. Tetrahedron 1996, 52, 14757.

(5) (a) Yang, C.-F.; Shen, C.; Wang, J.-Y.; Tian, S.-K. A Highly Diastereoselective Decarboxylative Mannich Reaction of β -Keto Acids with Optically Active N-Sulfinyl α -Imino Esters. Org. Lett. **2012**, 14, 3092. (b) Zhang, Y.; Li, J.-K.; Zhang, F.-G.; Ma, J.-A. Catalytic Asymmetric Access to Noncanonical Chiral α -Amino Acids from Cyclic Iminoglyoxylates and Enamides. J. Org. Chem. **2020**, 85, 5580. (c) Perera, S.; Sinha, D.; Rana, N. K.; Trieu-Do, V.; Zhao, J. C.-G.

List-Barbas-Mannich Reaction Catalyzed by Modularly Designed Organocatalysts. J. Org. Chem. 2013, 78, 10947.

(6) Jousseaume, T.; Wurz, N. E.; Glorius, F. Highly Enantioselective Synthesis of α -Amino Acid Derivatives by an NHC-Catalyzed Intermolecular Stetter Reaction. *Angew. Chem., Int. Ed.* **2011**, *50*, 1410.

(7) (a) Lu, X.; Yi, J.; Zhang, Z.-Q.; Dai, J.-J.; Liu, J.-H.; Xiao, B.; Fu, Y.; Liu, L. Expedient Synthesis of Chiral α -Amino Acids through Nickel-Catalyzed Reductive Cross-Coupling. *Chem. - Eur. J.* **2014**, *20*, 15339. (b) Jackson, R. F. W.; Wood, A.; Wythes, M. J. An Approach to the Synthesis of Enantiomerically Pure Hydroxylated α -Amino Acids Using Zinc Homoenolate Chemistry. *Synlett* **1990**, *1990*, 735. (c) Jackson, R. F. W.; James, K.; Wythes, M. J.; Wood, A. A new direct method for the synthesis of enantiomerically pure protected α -amino acids. *J. Chem. Soc., Chem. Commun.* **1989**, 644.

(8) (a) Yan, M.; Lo, J. C.; Edwards, J. T.; Baran, P. S. Radicals: Reactive Intermediates with Translational Potential. *J. Am. Chem. Soc.* **2016**, 138, 12692. (b) Zard, S. Z. Radicals in Action: A Festival of Radical Transformations. *Org. Lett.* **2017**, *19*, 1257.

(9) (a) Easton, C. J. Free-Radical Reactions in the Synthesis of α -Amino Acids and Derivatives. Chem. Rev. 1997, 97, 53. (b) Hansen, S. G.; Skrydstrup, T. Modification of Amino Acids, Peptides, and Carbohydrates through Radical Chemistry. In Radicals in Synthesis II; Gansäuer, A., Ed.; Springer: Berlin, Heidelberg, 2006; p 135. (c) Deska, J. Radical-Mediated Synthesis of α -Amino Acids and Peptides. In Amino Acids, Peptides and Proteins in Organic Chemistry 2011, 115. (d) Brandhofer, T.; García Mancheño, O. Site-Selective C-H Bond Activation/Functionalization of Alpha-Amino Acids and Peptide-Like Derivatives. Eur. J. Org. Chem. 2018, 2018, 6050. (e) Liu, J.-Q.; Shatskiy, A.; Matsuura, B. S.; Kärkäs, M. D. Recent Advances in Photoredox Catalysis Enabled Functionalization of α -Amino Acids and Peptides: Concepts, Strategies and Mechanisms. Synthesis 2019, 51, 2759. (f) Bottecchia, C.; Noël, T. Photocatalytic Modification of Amino Acids, Peptides, and Proteins. Chem. - Eur. J. 2019, 25, 26. (g) Larionov, V. A.; Stoletova, N. V.; Maleev, V. I. Advances in Asymmetric Amino Acid Synthesis Enabled by Radical Chemistry. Adv. Synth. Catal. 2020, 362, 4325. (h) Aguilar Troyano, F. J.; Merkens, K.; Anwar, K.; Gomez-Suarez, A. Radical-Based Synthesis and Modification of Amino Acids. Angew. Chem., Int. Ed. 2021, 60, 1098. (i) King, T. A.; Mandrup Kandemir, J.; Walsh, S. J.; Spring, D. R. Photocatalytic methods for amino acid modification. Chem. Soc. Rev. 2021, 50, 39.

(10) (a) Beckwith, A. L. J.; Chai, C. L. L. Diastereoselective radical addition to derivatives of dehydroalanine and of dehydrolactic acid. J. Chem. Soc., Chem. Commun. 1990, 1087. (b) Axon, J. R.; Beckwith, A. L. J. Diastereoselective radical addition to methyleneoxazolidinones: an enantioselective route to α -amino acids. J. Chem. Soc., Chem. Commun. 1995, 549. (c) Aycock, R. A.; Vogt, D. B.; Jui, N. T. A practical and scalable system for heteroaryl amino acid synthesis. Chem. Sci. 2017, 8, 7998. (d) Aycock, R. A.; Pratt, C. J.; Jui, N. T. Aminoalkyl Radicals as Powerful Intermediates for the Synthesis of Unnatural Amino Acids and Peptides. ACS Catal. 2018, 8, 9115. (e) Trowbridge, A.; Reich, D.; Gaunt, M. J. Multicomponent synthesis of tertiary alkylamines by photocatalytic olefin-hydroaminoalkylation. Nature 2018, 561, 522. (f) Kanegusuku, A. L. G.; Castanheiro, T.; Ayer, S. K.; Roizen, J. L. Sulfamyl Radicals Direct Photoredox-Mediated Giese Reactions at Unactivated C(3)-H Bonds. Org. Lett. 2019, 21, 6089. (g) Gaunt, M. J.; Reich, D.; Trowbridge, A. Rapid syntheses of (-)-FR901483 and (+)-TAN1251C enabled by complexity-generating photocatalytic olefin hydroaminoalkylation. Angew. Chem., Int. Ed. 2020, 59, 2256. (h) Zhang, O.; Schubert, J. W. Derivatization of Amino Acids and Peptides via Photoredox-Mediated Conjugate Addition. J. Org. Chem. 2020, 85, 6225. (i) Ji, P.; Zhang, Y.; Dong, Y.; Huang, H.; Wei, Y.; Wang, W. Synthesis of Enantioenriched alpha-Deuterated alpha-Amino Acids Enabled by an Organophotocatalytic Radical Approach. Org. Lett. 2020, 22, 1557.

(11) Merkens, K.; Aguilar Troyano, F. J.; Djossou, J.; Gómez-Suárez, A. Synthesis of Unnatural α -Amino Acid Derivatives via Light-

Mediated Radical Decarboxylative Processes. Adv. Synth. Catal. 2020, 362, 2354.

(12) Wang, X.; Chen, Y.; Song, H.; Liu, Y.; Wang, Q. Synthesis of Unnatural alpha-Amino Acids via Photoinduced Decatungstate-Catalyzed Giese Reactions of Aldehydes. *Org. Lett.* **2021**, *23*, 2199–2204.

(13) (a) Zheng, L.; Xia, P. J.; Zhao, Q. L.; Qian, Y. E.; Jiang, W. N.; Xiang, H. Y.; Yang, H. Photocatalytic Hydroacylation of Alkenes by Directly Using Acyl Oximes. J. Org. Chem. 2020, 85, 11989. (b) Zhang, M.; Yuan, X. A.; Zhu, C.; Xie, J. Deoxygenative Deuteration of Carboxylic Acids with D2O. Angew. Chem., Int. Ed. 2019, 58, 312. (c) Martinez Alvarado, J. I.; Ertel, A. B.; Stegner, A.; Stache, E. E.; Doyle, A. G. Direct Use of Carboxylic Acids in the Photocatalytic Hydroacylation of Styrenes To Generate Dialkyl Ketones. Org. Lett. 2019, 21, 9940. (d) Zhang, M.; Xie, J.; Zhu, C. A general deoxygenation approach for synthesis of ketones from aromatic carboxylic acids and alkenes. Nat. Commun. 2018, 9, 3517. (e) Stache, E. E.; Ertel, A. B.; Rovis, T.; Doyle, A. G. Generation of Phosphoranyl Radicals via Photoredox Catalysis Enables Voltage-Independent Activation of Strong C-O Bonds. ACS Catal. 2018, 8, 11134.

(14) Rossi-Ashton, J. A.; Clarke, A. K.; Unsworth, W. P.; Taylor, R. J. K. Phosphoranyl Radical Fragmentation Reactions Driven by Photoredox Catalysis. *ACS Catal.* **2020**, *10*, 7250.

(15) Kay, M.; Francisco José, A. T.; Khadijah, A.; Adrián, G. S. Synthesis of γ -Oxo- α -amino Acids via Radical Acylation with Carboxylic Acids. *ChemRxiv*, Jun 11, **2020**. DOI: 10.26434/ chemrxiv.13194920.v2.

(16) See the Supporting Information for further details.

(17) Shang, T.-Y.; Lu, L.-H.; Cao, Z.; Liu, Y.; He, W.-M.; Yu, B. Recent advances of 1,2,3,5-tetrakis(carbazol-9-yl)-4,6-dicyanobenzene (4CzIPN) in photocatalytic transformations. *Chem. Commun.* **2019**, *55*, 5408.

(18) Potential side reactions arising from interactions of the excited photocatalyst with cinnamic acid might be the cause of the observed complex mixture.

(19) Lowry, M. S.; Goldsmith, J. I.; Slinker, J. D.; Rohl, R.; Pascal, R. A.; Malliaras, G. G.; Bernhard, S. Single-Layer Electroluminescent Devices and Photoinduced Hydrogen Production from an Ionic Iridium(III) Complex. *Chem. Mater.* **2005**, *17*, 5712.

(20) Pandey, G.; Pooranchand, D.; Bhalerao, U. T. Photoinduced single electron transfer activation of organophosphines: Nucleophilic trapping of phosphine radical cation. *Tetrahedron* **1991**, *47*, 1745.

(21) Monos, T. M.; Sun, A. C.; McAtee, R. C.; Devery, J. J.; Stephenson, C. R. J. Microwave-Assisted Synthesis of Heteroleptic Ir(III)+ Polypyridyl Complexes. J. Org. Chem. 2016, 81, 6988.

(22) Kumar, S.; Gawandi, V. B.; Capito, N.; Phillips, R. S. Substituent Effects on the Reaction of β -Benzoylalanines with Pseudomonas fluorescens Kynureninase. *Biochemistry* **2010**, *49*, 7913.

(23) Nakamura, Y.; Matsubara, R.; Kiyohara, H.; Kobayashi, S. Catalytic, Asymmetric Mannich-Type Reactions of α -Imino Esters Bearing Readily Removable Substituents on Nitrogen. *Org. Lett.* **2003**, *5*, 2481.