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PPh₃ Mediated Reductive Annulation Reaction between Isatins and Electron Deficient Dienes to Construct Spirooxindole Compounds

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Abstract: A PPh₃ mediated reductive annulation reaction between isatins and 4,4-dicyano-2-methylenebut-3-enoates was developed. The reaction provided an alternative method for constructing five-membered and three-membered all-carbon spirooxindole compounds. Lithium chloride as a Lewis acid played a key role in the synthesis of spirocyclopentenyl oxindole compounds.

INTRODUCTION

Spirooxindole moiety is an important framework present in a variety of pharmaceutical molecules¹ and natural products². As representative compounds, compound **a** in Scheme 1 is a natural product that contains the spirooxindole framework, while compounds **b** and **c** are two pharmaceutical molecules which show distinct antitumor activities¹ since the strained spiro-cyclopropane at the C3 position of oxindoles plays an important role in their biological activities. So far, the spirooxindole framework has attracted great interest from chemists due to its biological importance and the challenge of synthesizing it³. Isatins have been widely used as raw materials in the synthesis of spirooxindole compounds⁴, however, to the best of our knowledge, isatins are rarely reported as a C1 synthon in the construction of all-carbon spirooxindole compounds⁵, since the deoxygenation step is needed.

In the early reports, α -dicarbonyl compounds could be reduced by phosphine⁶ with one of the carbonyls deoxygenated, and a carboxylic ester could be synthesized by a reductive coupling of α -keto ester with a carboxylic acid⁷. It is commonly believed that either a five-coordinate 1,3,2-dioxaphospholene intermediate or a dipolar ion (a in Scheme 2) is formed between the α -dicarbonyl compound and phosphine⁸. In our preliminary study, it was found that isatins could undergo a reductive coupling reaction at the C3 position with a carboxylic acid, and a benzoate at C3 position of isatins could be obtained, which also proved that a Kukhtin–Ramirez intermediate was formed between the N-methyl protected isatin and triphenylphosphine (a in Scheme 2)^{7c,8b}. In this process, the intermediate underwent a two-step process: first, the intermediate accepted a proton from the acid, then the carboxylic ion attacked the C-O bond of the intermediate, triphenylphosphine oxide departed and the ester was finally formed. Considering the feature of the above reaction, this intermediate of isatins might have potential in the synthesis of spirooxindole compounds with another synthon. With our ongoing interest in the exploration of heterocyclic scaffold construction through organocatalysis⁹, we used triphenylphosphine as a deoxygenation regent and developed a new reaction in which isatins could react directly with electron-deficient dienes and produce various spiro-oxindole compounds.

Scheme 1. Representative natural products and pharmaceutical molecules



Scheme 2. Different PPh₃ involved intermediates

a Kukhtin-Ramirez intermediate



RESULTS AND DISCUSSION

In our initial investigation, ethyl 4,4-dicyano-2-methylene-3-phenylbut-3-enoate $2a^{10,11}$, PPh₃ and an N-methyl protected isatin were used as substrates. When stoichiometric amount of PPh₃ was added, a yellow solid of tetravalent phosphinium zwitterion (b in Scheme 2) was formed immediately. An unexpected cyclopropanation compound was produced after heating for 24 h (Table 1, entry 1). The cyclopropanation product could also be obtained with the separated tetravalent phosphinium zwitterions and the N-methyl protected isatin under the same conditions. To improve the efficiency of this process, a series of N-protected isatins were tested. When the electron donating group protected isatins were used, the dimerization of isatins was observed, phosphorous ylides of isatins were also formed in the reaction system and the excessive PPh₃ was consumed by isatins. Finally, it was found that tosyl group was the best N-protection group for the reaction (Table 1, entries 1-5). The solvents were then screened and it was found that toluene performed the best. Polar protic solvents were not favorable for this process, and no product was observed when MeOH was used as the solvent (see Supporting Information, Table S1). Other polar solvents showed moderate yields in this process. Further tests showed that PPh₃ was the optimal trivalent phosphorus reagent, P(NMe₂)₃ (Table 1, entries 7, 8).

Table 1. Scope of N-protected isatins^{*a*}.



entry	\mathbb{R}^1	P reagent	temp (°C)	Time (h)	dr ^b	yield ^c (%)
1	Me	PPh ₃	60	24	>20:1	20
2	Boc	PPh ₃	60	24		n.r.
3	Ac	PPh ₃	60	20	>20:1	51
4	Bn	PPh ₃	60	42	>20:1	73
5	Ts	PPh ₃	60	24	6:1	83
6	Ts	P(NMe ₂) ₃	0	1	6:1	32
7	Ts	PBu ₃	60	6	6:1	43
8	Ts	Ph ₂ MeP	60	14	6:1	51

^{*a*}Standard procedure: substrate **1** (0.15 mmol) and substrate **2a** (0.1 mmol) were added into a reaction tube containing the solvent (0.6 mL), PR₃ solution (1.5 equiv) in the solvent (0.4 mL) was then added slowly into the tube with stirring at room temperature under N₂ atmosphere, and then the reaction tube was heated at 60 °C in an oil bath with the reaction monitored by TLC. ^{*b*}Determined by ¹H NMR analysis. ^{*c*}Isolated yield of the major product.

The generality of this reductive cyclopropanation was further explored under the optimized conditions. A variety of substituted dienes 2 (Table 2) were tested to react with N-tosyl protected isatin. Dienes 2 with the electron-donating and electron-withdrawing group substituted phenyls performed differently in this cyclopropanation reaction. Dienes 2 with

Table 2. The substrate scope of the cyclopropanation reaction^a



entry	R ²	R ³	time (h)	dr ^b	yield ^c (%)
1	Ph (2a)	Н (1а)	20	6:1	3a , 83
2	$2\text{-MeC}_6\text{H}_4(2\mathbf{b})$	Н	20	6:1	3b , 75
3	$3-MeC_6H_4(2c)$	Н	20	10:1	3c , 90
4	$4\text{-MeC}_6\text{H}_4(\textbf{2d})$	Н	20	6:1	3d , 83
5	$4\text{-OMeC}_{6}\text{H}_{4}\left(\mathbf{2e}\right)$	Н	32	8:1	3e , 80
6	$2\text{-FC}_{6}\text{H}_{4}\left(\mathbf{2f}\right)$	Н	20	>20:1	3f , 66
7	$2\text{-ClC}_6\text{H}_4(2\mathbf{g})$	Н	72	10:1	3g , 57
8	$3\text{-ClC}_6\text{H}_4(2\mathbf{h})$	Н	20	10:1	3h , 60
9	$4\text{-ClC}_6\text{H}_4(2\mathbf{i})$	Н	72	8:1	3i , 60
10	$4-BrC_{6}H_{4}(2j)$	Н	20	>20:1	3j , 52
11	$4-t-\mathrm{BuC}_{6}\mathrm{H}_{4}\left(\mathbf{2k}\right)$	Н	6	>20:1	3k , 92
12	3,4-diMe C ₆ H ₃ (2 I)	Н	20	>20:1	31 , 95
13	Ph (2a)	5-Cl (1b)	4	>20:1	3m , 96
14	Ph (2a)	5-Me (1c)	20	10:1	3n , 83

^{*a*}Unless noted otherwise, the reaction is operated under the standard procedure A in the Experimental Section. ^{*b*}Determined by ¹H NMR analysis. ^{*c*}Isolated yield of the major product.

electron-donating group substituted phenyls afforded the products in high yields (Table 2, entries 11, 12), while those with electron- deficient aryl groups provided moderate yields (Table 2, entries 8-10). Due to the steric hindrance, dienes **2** with the ortho–substituted phenyls afforded the products in low yields (Table 2, entries 2, 6, 7), no cyclopropanation product was isolated with 1-napth-phenyl diene **2**; the furyl substituted diene **2** produced a mixture of cyclopropanation products with dr ratio 1:1, which could not be separated by silicagel column. Dienes **2** with *meta-* and *para-* substituted phenyls did not show any difference in the cyclopropanation reaction (Table 2, entries 8, 9). The substituent effect of isatins 1 was also investigated. Electron poor isatin produced the product in excellent yield (Table 2, entry 13), the electron rich group substituted isatin **1** also showed good activity in the process (Table 2, entry 14), we believed that isatins with electron-withdrawing groups could form the intermediates^{8b} more easily and therefore caused less side reactions of isatins.

Table 3. Screening of additives

		R^{1}	NC CN + PPh ₂ CO ₂ Et 2a	additive solvent	/ N N	
entry	\mathbb{R}^1	additives (equiv)	solvent	temp (°C)	time (h)	yield ^a (%)

2	Ts	LiCl (0.2)	toluene	70	20	55	
3	Ts	LiCl (2.0)	toluene	70	20	66	
4	Ts	LiBr (2.0)	toluene	70	20	65	
5	Ts	LiBF ₄ (2.0)	toluene	70	20	21	
6	Ts	LiOH (2.0)	toluene	70	20	N.R.	
7	Ts	LiCl (3.0)	toluene	80	20	68	
8	Ts	LiCl (3.0)	DCE	80	20	83	
9	Ts	LiCl (3.0)	toluene	100	20	57	
10	Ts	LiCl (3.0)	DCE	100	20	75	
^a Isolated yield of the major product							

"Isolated yield of the major product.

When we were optimizing the conditions for the cyclopropanation process, several Lewis acids were also tested to improve the yield. However, when anhydrous LiCl was used as an additive, the expected cyclopropanation did not occur, a cyclopentene annulation product was produced instead. Further research showed that only lithium salt was favorable for this reaction, other Lewis acids such as Scandium trifluoromethanesulfonate and Magnesium trifluoromethanesulfonate only gave cyclopropanation products. It's probably because the coordination between lithium and PPh₃ was weaker than those between other Lewis acids and PPh₃. With this interesting result obtained, we started to optimize the conditions of this [4+1] annulation reaction. It was found that the excessive amount of lithium salt was needed to improve the yield (Table 3), DCE was the optimal solvent, and 80 °C was the most favorable temperature. Only small amount of cyclopropanation product was produced in the initial stage, under the reaction conditions, the cyclopanation product rearranged to form cyclopentene annulation products (Table 4). The results showed that substrates **2** with electron deficient group substituted phenyls (Table 4, entry 6) resulted in moderate yields, while substrates **2** with electron rich group substituted phenyls (Table 4, entries 2, 3, 7, 8) resulted in high yields, and the electron deficient substrates **2** showed lower reactivities in cyclopentene annulation processes than in the cyclopropanation reactions.

All the cyclopropanation and cyclopentene annulation products were characterized by ¹H, ¹³C NMR and HRMS-ESI measurements. The structures of representative cyclopropanation and cyclopentene annulation products were determined by single-crystal X-ray diffraction analysis (CCDC number 1424174 for **3**j, 1424175 for **4**a). The major diastereomers of cycloprorantion compounds were made in analogy with representative 3**j**.

Table 4. Substrate scope of the [4+1] annulations^a



entry	R^2	R ³	yield ^{b} (%)	
1	Ph (2a)	Н (1а)	4a , 83	
2	$4\text{-Me-}C_6H_4\left(\mathbf{2d}\right)$	Н	4d , 77	
3	$4\text{-OMe-}C_6H_4(2\mathbf{e})$	Н	4e , 78	
4	$3\text{-Cl-C}_6\text{H}_4(2\mathbf{h})$	Н	4h , 60	
5	$3-Br-C_6H_4(2o)$	Н	40 , 65	
6	2-furyl (2p)	Н	4p , 58	
7	$4-t-Bu-C_6H_4(2\mathbf{k})$	Н	4k , 76	
8	$3,4-d1Me-C_6H_3(21)$		41 , 70	4
9	rn	5-CI (10)	4m , 80	4

The Journal of Organic Chemistry

10Ph5-Me (1c)4n, 81"Unless noted otherwise, the reaction was operated under the standard procedure B in the Experimental section." Isolated yield of the major product.

During our study, efforts were also made to understand the reaction mechanism. The N-protected isatin and triphenylphosphine can generate phosphorous ylide (as in Scheme 5, A), the ylide may attack substrate 2 via a tandem S_N^2 Michael addition to form product either 3 or 4. However, it was found that the phosphorous ylide of N-protected isatin did not react with substrate 2 under either standard procedure A (a in Scheme 3) or standard procedure B (b in Scheme 3), and the tetravalent phosphinium zwitterions (b in Scheme 2) did not react with benzaldehyde (c in Scheme 3), either. These experiments demonstrated that the reaction did not occur by a phosphorous ylide intermediate mechanism or a wittig reaction mechanism, further experiments showed that N-protected isatin reacted with 2-benzylidenemalononitrile to produce a cyclopropanation product under the standard procedure A (d in Scheme 3), which suggested that the reaction might proceed via a Michael addition process.

Scheme 3. Mechanism study



Further investigation proved that the isolated cyclopropanation product could be rearranged by PPh₃ in DCE, the rearrangement reached equilibrium by using PPh₃ after heating for 24 hours (a in Scheme 4). On the contrary, using both lithium chloride and PPh₃ resulted in a complete conversion, f completely disappeared even though the isolated yield of the product g was only 28% (b in Scheme 4). No rearrangement was observed by using lithium chloride alone (c in Scheme 4). The rearrangement was not observed at 60 °C in the cyclopropanation process, either, the reason might be that the low temperature was not favorable for the rearrangement. The rearrangement proceeded under the standard procedure B in a yield of only 28% (b in Scheme 4), most of the spirocyclopropane was decomposed in this process, which proved that the main reaction pathway of the cyclopentene product is not the rearrangement of cyclopropanation product.

Scheme 4. Rearrangement investigation



The Journal of Organic Chemistry

Based on this study and the early related reports^{5, 8}, a reasonable mechanism for the cyclopropanation and [4+1] type annulation process was proposed. First, isatins and triphenylphosphine form the unstable intermediate **A** in the solvent. In the cyclopropanation process, **A** reacts with substrate **2** by a Michael addition process to produce the carbanion **C**, **C** then undergoes the replacement reaction via an intramolecular S_N^2 process, where the C-O bond is broken and triphenylphosphine oxide departs, and the cyclopropanation product is formed. In the [4+1] type annulation process, the carbanion **C** formed after the Michael addition is stabilized by Li⁺ to afford the new carbanion **D** (*E*-isomer is favorable), an intramolecular S_N^2 process occurs in **D** with the C3 position on the indole ring of **A** attacked by Li⁺-activated diene, then triphenylphosphine oxide departs and the cyclopentene annulation product is formed. In the Li⁺ promoted rearrangement process, PPh₃ attacks the electron deficient olefinic bond of cyclopropanation product to form carbanion **E**, then another intramolecular S_N^2 process occurs in **E** with the C-C bond of cyclopropane attacked and the ring is opened, PPh₃ is then eliminated and the cyclopentene is obtained. According to our study, the cyclopentene product is mainly produced via carbanion **D**, not carbanion **E**.

Scheme 5. Proposed mechanism



CONCLUSION

In conclusion, we have developed an alternative method to construct spirocyclopropyl oxindole compounds and spirocyclopentenyl oxindole compounds. PPh_3 acted as a reductant and activated electronic deficient olefinic bonds in the rearrangement process. Lithium chloride as a Lewis acid activated 4,4-dicyano-2-methylenebut-3-enoates in the [4+1] annulation process. In addition, a plausible reaction mechanism was proposed, and the key intermediate of this reaction was proved to be the Kukhtin–Ramirez intermediate.

EXPERIMENTAL SECTION

All reagents were purchased from commercial vendors and used as received unless otherwise noted. 1,2-dichloroethane was dried over P_2O_5 for 12 hours prior to use. Toluene was dried over sodium. ¹H, ¹³C, and ³¹P NMR spectra were recorded on a Bruker JNM-ECS 400M. ¹H NMR chemical shifts are given in ppm with respect to the solvent residual peak (CDCl₃, δ = 7.26 ppm), ¹³CNMR shifts are given in ppm with respect to CDCl₃ (δ = 77.2 ppm), and ³¹P shifts are given in ppm with respect to an external sample of 85% H₃PO₄ (δ = 0.0 ppm). HRMS spectra were acquired in the ESI mode (positive ion) with the mass analyzer of Obitrap used. Coupling constants are reported as *J*-values in Hz. Column chromatography was performed using 200-300 mesh silica gel as the stationary phase. 4,4-dicyano-2-methylenebut-3-enoate **2** was prepared according to a procedure in the literature^{11a}. N-tosyl protected isatin **1** was prepared according to the procedure below: isatin (1.47 g) was dissolved in DMF (10 ml) in a 50 ml round-bottom flask at 0 °C, and sodium hydride (0.48 g) was added slowly into the flask, after stirring for 1 h, tosyl chloride (2.28 g) was added slowly into this flask, the mixture was then stirred for 3 h at room temperature, then the flask was heated at 70 °C for 8 h, removed from the heat and cooled down to room temperature, water was then added into the mixture and the product was precipitated, filtrated under reduced pressure and washed with EtOH and Et₂O. N-tosyl protected isatin was obtained in 90% yield.

Standard procedure A: N-tosyl protected isatin 1 (0.15 mmol) and 4,4-dicyano-2-methylenebut-3-enoate 2 (0.1 mmol) were added into toluene (1 mL) in a reaction tube, PPh₃ (0.15 mmol) was then added, and the reaction was heated at 60 °C under N₂ atmosphere and monitored by TLC until the starting material disappeared. The solvent was removed under reduced pressure, and the residue was isolated by silica gel column chromatography (petroleum ether/ethyl acetate 5:1) to give product **3**.

Standard procedure B: N-tosyl protected isatin (0.15 mmol) and 4,4-dicyano-2-methylenebut-3-enoate (0.1 mmol) were added into DCE (1 mL) in a flask, PPh₃ (0.15 mmol) and LiCl (0.3 mmol) were then added, and the reactions was heated at 80 °C under N₂ atmosphere for 20 h. The solvent was removed and the residue was isolated by silica gel column chromatography (petroleum ether/ethyl acetate 4:1) to give product **4**.

Ethyl 2-(2,2-*dicyano-1-phenylvinyl*)-2'-*oxo-1*'-*tosylspiro*[*cyclopropane-1,3*'-*indoline*]-2-*carboxylate* (**3***a*) white solid, 44.5mg, 83% yield; mp 88–90 °C; IR (KBr) cm⁻¹: 2983, 2230, 1734, 1597, 1462, 1380, 1254, 1238, 1190, 1177, 1086, 961, 910, 734, 665, 575, 542, ¹H NMR (400 MHz, CDCl₃, TMS) : δ 8.11 (d, 2H, J = 8.3 Hz), 8.08 (d, 1H, J = 8.8 Hz), 7.46–7.42 (m, 2H), 7.54–7.49 (m, 5H), 7.39 (d, 1H, J = 7.0 Hz), 7.37 (d, 1H, J = 6.5 Hz), 7.14 (t, 1H, J = 7.6 Hz), 4.39–4.18 (m, 2H, CH₂), 2.76 (d, 1H, J = 5.6 Hz), 2.49 (s, 3H), 1.71(d, 1H, J = 5.5 Hz), 1.19 (t, 3H, J = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.3, 169.8, 164.1, 146.5, 140.7, 134.8, 134.1, 132.1, 129.8, 129.7, 129.0, 128.8, 128.2, 124.6, 123.9, 122.8, 113.8, 112.3, 111.8, 90.6, 64.0, 47.0, 44.2, 27.5, 21.8, 13.9; HRMS (ESI) Calcd for C₃₀H₂₃N₃O₅S₁Na₁[M+Na]⁺: 560.1251; found : 560.1245.

Ethyl 2-(2, 2-*dicyano-1-(o-tolyl)vinyl)-2'-oxo-1'-tosylspiro[cyclopropane-1,3'-indoline]-2-carboxylate* (**3b**) white solid, 41.3mg, 75% yield; mp 158–160 °C; IR (KBr) cm⁻¹: 2925, 2853, 2232, 1733, 1598, 1462, 1381, 1253, 1238, 1190, 1177, 1086, 910, 733, 665, 575, 542 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.14 (d, 1H, J = 8.2 Hz), 8.08 (d, 1H, J = 8.1 Hz), 7.67 (d, 1H, J = 7.3 Hz), 7.45 (d, 2H, J = 8.2Hz), 7.39 (t, 3H, J = 8.0 Hz), 7.25 (t, 2H, J = 8.0 Hz), 7.13 (t, 1H, J = 7.6 Hz), 4.35–4.15 (m, 2H, CH₂), 2.71 (d, 1H, J = 5.6 Hz), 2.19 (s, 3H), 2.48 (s, 3H, CH₃), 1.80 (d, 1H, J = 5.6 Hz), 1.19 (t, 3H, J = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 171.4, 170.2, 163.7, 146.5, 140.6, 134.6, 134.1, 131.2, 131.1, 130.1, 129.9, 129.8, 128.9, 128.1, 124.48, 124.46, 122.8, 122.7, 113.8, 111.7, 111.5, 94.6, 63.9, 47.6, 44.2, 25.6, 21.8, 20.0, 13.8; HRMS (ESI) calcd for C₃₁H₂₆N₃O₅S₁ [M+H]⁺: 552.1588; found: 552.1583.

Ethyl 2-(2, 2-dicyano-1-(m-tolyl)vinyl)-2'-oxo-1'-tosylspiro[*cyclopropane-1,3'-indoline*]*-2-carboxylate* (*3c*) white solid, 52.4mg, 95% yield; mp 158–160 °C; IR (KBr) cm⁻¹: 2983, 2961, 2257, 2230, 1734, 1598, 1463, 1381, 1239, 1190, 1177, 1086, 962, 910, 734, 665, 575, 542; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.11 (d, 2H, *J* = 8.2 Hz), 8.07 (d, 1H, *J* = 8.2Hz), 7.46–7.42 (m, 2H), 7.40–7.33 (m, 4H), 7.28 (s, 2H), 7.14 (t, *J* = 7.6 Hz), 4.40–4.18 (m, 2H), 2.77 (d, 1H, *J* = 5.7 Hz), 2.49 (s, 3H), 2.44 (s, 3H), 1.73 (d, 1H, *J* = 5.7 Hz), 1.18 (t, 3H, *J* = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.3, 169.9, 164.2, 146.5, 140.6, 139.0, 134.8, 134.1, 133.0, 129.8, 128.9, 128.8, 128.5, 125.4, 124.5, 122.92, 122.87, 113.8, 112.4, 111.9, 90.4, 63.9, 47.0, 44.0, 27.5, 21.8, 21.4, 13.9; HRMS (ESI) calcd for C₃₁H₂₅N₃O₅S₁Na₁ [M+Na]⁺: 574.1407; found: 574.1403.

Ethyl 2-(2, 2-*dicyano-1-(p-tolyl)vinyl)-2'-oxo-1'-tosylspiro[cyclopropane-1,3'-indoline]-2-carboxylate (3d) white solid, 45.7mg, 83% yield, mp 162–164 °C, IR (KBr) cm⁻¹: 2984, 2950, 2256, 2229, 1734, 1606, 1574, 1553, 1462, 1380, 1255, 1237, 1190, 1177, 1086, 961, 909, 815, 753, 732, 665, 574, 542; ¹H NMR (400 MHz, CDCl₃, TMS): \delta 8.11–8.06 (m, 3H), 7.44 (d, 2H, <i>J* = 8.0 Hz), 7.36–7.40 (m, 4H), 7.33 (d, 2H, *J* = 7.6 Hz), 7.14 (t, 1H, *J* = 7.5 Hz), 4.36–4.18 (m, 2H), 2.77 (d, 1H), 2.49 (s, 3H), 2.43 (s, 3H), 1.73 (d, 1H, *J* = 5.4 Hz), 1.17 (t, 3H, *J* = 7.0 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.2, 169.7, 164.3, 146.5, 143.3, 140.6, 134.0, 132.0, 129.8, 129.77, 129.7, 128.8, 128.3, 124.5, 122.91, 122.86, 113.8, 112.7, 112.0, 89.4, 63.9, 47.0, 44.0, 27.6, 21.8, 21.7, 13.9; HRMS (ESI) calcd for C₃₁H₂₅N₃O₅S₁Na₁[M+Na]⁺ : 574.1407; found: 574.1401.

Ethyl 2-(2,2-*dicyano-1*-(4-*methoxyphenyl*)*vinyl*)-2'-*oxo-1*'-*tosylspiro*[*cyclopropane-1,3*'-*indoline*]-2-*carboxylate* (**3***e*) white solid ,45.3mg, 80% yield, mp 68–70 °C, IR (KBr) cm⁻¹: 2982, 2934, 2256, 2227, 1733, 1603, 1510, 1494, 1462, 1381, 1262, 1238, 1189, 1177, 1153, 1086, 1027, 961, 909, 837, 733, 665, 575, 542; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.09 (d, 2H, *J* = 8.4Hz), 8.06 (d, 1H, *J* = 8.4 Hz), 7.50 (d, 2H, *J* = 8.8 Hz), 7.44–7.35 (m, 4H), 7.13 (t, 1H, *J* = 7.6 Hz), 6.99 (d, 2H, *J* = 8.9 Hz), 4.37–4.16 (m, 2H), 3.87 (s, 3H), 2.78 (d, 1H, *J* = 5.6 Hz), 2.48 (s, 3H), 1.74 (d, 1H, *J* = 5.6 Hz), 1.17 (t, 3H, *J* = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.2, 168.7, 164.4, 162.9, 146.4, 140.6, 134.1, 131.2, 130.6, 129.7, 128.8, 127.0, 124.4, 123.0, 122.8, 114.4, 113.8, 113.0, 112.3, 87.7, 63.8, 55.5, 46.9, 43.8, 27.8, 21.7, 13.9; HRMS (ESI) calcd for C₃₁H₂₅N₃O₆S₁Na₁ [M+Na]⁺: 590.1356; found : 590.1348.

Ethyl 2-(2,2-dicyano-1-(2-fluorophenyl)vinyl)-2'-oxo-1'-tosylspiro[cyclopropane-1,3'-indoline]-2-carboxylate (3f) white solid, 36.8mg, 66% yield, mp 163–165 °C, IR (KBr) cm⁻¹: 2984, 2926, 2257, 2232, 1736, 1610, 1462, 1381, 1254, 1238, 1190, 1177, 1154, 1087, 962, 909, 754, 733, 665, 576, 542; ¹HNMR (400 MHz, CDCl₃, TMS): δ 7.68 (t, 1H, *J* = 6.4 Hz), 7.58–7.52 (m, 1H), 7.45 (d, 2H, *J* = 8.2 Hz), 7.42–7.38 (m, 2H), 7.34 (t, 1H, *J* = 7.6 Hz), 7.19–7.12 (m, 2H), 4.33–4.20 (m, 2H), 2.76 (d, 1H, *J* = 5.4 Hz), 2.49 (s, 3H), 1.70 (d, 1H, *J* = 5.4 Hz), 1.20 (t, 3H, *J* = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.6, 166.6, 163.1, 158.8 (d, 1*J*C-F = 250.4 Hz), 146.6, 140.6, 134.2, 134.1 (d, 3*J*C-F = 8.7 Hz), 131.7, 129.8, 128.9, 124.9 (d, 4*J*C-F = 3.4 Hz), 124.6, 123.2, 122.9, 122.6, 122.5, 116.2 (d, 2*J*C-F = 21.0 Hz), 113.8, 111.7, 111.3, 93.4, 63.7, 47.0, 44.2 26.4, 21.8, 13.9; HRMS (ESI) calcd for C₃₀H₂₂F₁N₃O₅S₁Na₁ [M+Na]⁺ : 578.1156; found: 578.1149

Ethyl 2-(1-(2-chlorophenyl)-2,2-dicyanovinyl)-2'-oxo-1'-tosylspiro[cyclopropane-1,3'-indoline]-2-carboxylate (**3***g*) white solid, 32.5mg, 57% yield, mp 164–166 °C, IR (KBr) cm⁻¹: 2984, 2926, 2256, 2234, 1736, 1597, 1463, 1380, 1311, 1238, 1190, 1178, 1087, 962, 910, 812, 733, 665, 575, 541; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.13 (d, 2H, *J* = 8.3 Hz), 8.08 (d, 1H, *J* = 8.3 Hz), 7.78 (t, 1H, *J* = 3.0 Hz), 7.49–7.43 (m, 5H), 7.40 (d, 1H, *J* = 8.0 Hz), 7.33 (d, 1H, *J* = 7.6 Hz), 7.14 (t, 1H, *J* = 7.6 Hz), 4.33–4.14 (m, 2H), 2.78 (d, 1H, *J* = 5.9 Hz), 2.48 (s, 3H), 1.70 (d, 1H, *J* = 5.9 Hz) 1.19 (t, 3H, *J* = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.5, 168.8,163.4, 146.6, 140.5, 134.0, 133.4, 133.2, 132.7, 131.3, 130.3, 129.9, 129.8, 128.9, 127.4, 124.6, 123.1, 122.8, 113.8, 111.6, 111.2, 95.2, 63.7, 47.3, 44.7, 25.8, 21.8,13.9; HRMS (ESI) calcd for C₃₀H₂₂Cl₁N₃O₅S₁Na₁ [M+Na]⁺: 594.0861; found: 594.0855.

Ethyl 2-(*1*-(*3*-chlorophenyl)-2,2-dicyanovinyl)-2'-oxo-1'-tosylspiro[cyclopropane-1,3'-indoline]-2-carboxylate (**3h**) white solid, 34.2mg, 60% yield, mp 76–78 °C, IR (KBr) cm⁻¹: 2982, 2927, 2232, 1732, 1603, 1565, 1462, 1385, 1308, 1249, 1179, 1086, 960, 906, 861, 806, 754, 695, 572; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.12–8.07 (m, 3H), 7.53–7.51 (m, 1H), 7.49–7.44 (m, 3H), 7.42–7.38 (m, 4H), 7.16 (t, 1H, *J* = 7.6 Hz), 4.40–4.19 (m, 2H), 2.77 (d, 1H, *J* = 5.8 Hz), 2.49 (s, 3H), 1.72 (d, 1H, *J* = 5.8 Hz), 1.20 (t, 3H, *J* = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.3, 168.4, 163.8, 146.6, 140.6, 136.3, 136.1, 134.0, 132.0, 130.4, 130.0, 129.8, 128.8, 127.7, 126.5, 124.6, 123.1, 122.6, 113.9, 111.8, 111.4, 91.8, 64.2, 46.7, 44.4, 27.3, 21.8, 13.9; HRMS(ESI) calcd for C₃₀H₂₂Cl₁N₃O₅S₁Na₁ [M+Na]⁺ : 594.0861; found: 594.0851.

Ethyl 2-(1-(4-chlorophenyl)-2,2-dicyanovinyl)-2'-oxo-1'-tosylspiro[cyclopropane-1,3'-indoline]-2-carboxylate (**3i**) white solid, 34.2mg, 60% yield, mp 82–84 °C, IR (KBr) cm⁻¹: 2935, 2232, 1737, 1598, 1461, 1381, 1243, 1089, 960, 909, 821, 745, 664, 573; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.10–8.06 (m, 3H), 7.49 (d, 2H, *J* = 8.5 Hz), 7.44–7.41 (m, 5H), 7.39–7.36 (m, 1H), 7.15 (t, 1H, *J* = 7.6 Hz), 4.39–4.17 (m, 2H), 2.77 (d, 1H, *J* = 5.7 Hz), 2.49 (s, 3H), 1.69 (d, 1H, *J* = 5.7 Hz), 1.19 (t, 3H, *J* = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.3, 168.6, 163.9, 146.6, 140.6, 138.6, 134.0, 133.0, 130.0, 129.8, 129.6, 129.4, 128.8, 124.6, 123.0, 122.6, 113.9, 112.1, 111.6, 90.9, 64.1, 46.7, 44.3, 27.4, 21.8, 13.9; HRMS (ESI) calcd for C₃₀H₂₂Cl₁N₃O₅S₁Na₁ [M+Na]⁺: 594.0861; found: 594.0855.

Ethyl 2-(1-(4-bromophenyl)-2,2-dicyanovinyl)-2'-oxo-1'-tosylspiro[cyclopropane-1,3'-indoline]-2-carboxylate (3j) white solid, 31.9mg, 52% yield, mp 81–84 °C, IR (KBr) cm⁻¹: 2983, 2928, 2256, 2230, 1735, 1596, 1585, 1462, 1381, 1237, 1177, 1086, 1010, 961, 813, 752, 665, 574, 542; ¹H NMR (400 MHz, CDCl₃, TMS): 8.06–8.10 (m, 3H), 7.65 (d, 2H, *J* = 8.32 Hz), 7.44–7.40(m, 3H), 7.39–7.34 (m, 3H), 7.14 (t, 1H, *J* = 7.56 Hz), 4.39–4.18 (m), 2.75 (d, *J* = 5.6Hz), 2.48 (s, 3H), 1.69 (d, 1H, *J* = 5.6 Hz), 1.19 (t, 3H, *J* = 7.12 Hz); ¹³C7 NMR (100 MHz, CDCl₃, TMS): δ 170.3, 168.7, 164.0, 146.6, 140.7, 134.1, 133.5, 132.4, 131.4, 130.0, 129.7, 128.8, 127.0, 124.6, 123.0,

122.7, 113.9, 112.1, 111.6, 91.0, 64.1, 46.7, 44.4, 27.4, 21.8, 13.9; HRMS (ESI) calcd for $C_{30}H_{22}Br_1N_3O_5S_1Na_1$ [M+Na]⁺: 638.0356; found: 638.0350.

Ethyl 2-(*1*-(*4*-(*tert-butyl*)*phenyl*)-2,2-*dicyanovinyl*)-2'-*oxo-1'*-*tosylspiro*[*cyclopropane-1,3'*-*indoline*]-2-*carboxylate* (**3***k*) white solid, 54.5mg, 92% yield, mp 178–180 °C, IR (KBr) cm⁻¹: 2966, 2906, 2870, 2256, 2229, 1734, 1605, 1462, 1382, 1255, 1238, 1190, 1177, 1086, 910, 734, 664, 573, 542; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.10 (d, 2H, *J* = 8.4 Hz), 8.07 (d, 1H, *J* = 8.4 Hz), 7.51 (d, 2H, *J* = 8.5 Hz), 7.46–7.41 (m, 4H), 7.40–7.37 (m, 2H), 7.14 (t, 1H, *J* = 7.3 Hz), 4.37–4.16 (m, 2H), 2.78 (d, 1H, *J* = 5.7 Hz), 2.49 (s, 3H), 1.75 (d, 1H, *J* = 5.7 Hz), 1.34 (s, 9H), 1.18 (t, 3H, *J* = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.3, 169.5, 169.3, 164.3, 156.3, 146.5, 140.7, 134.2, 131.9, 129.8, 129.7, 128.9, 128.3, 126.0, 124.5, 123.0, 123.9, 122.9, 113.8, 112.2, 89.2, 63.9, 46.9, 44.0, 35.2, 31.0, 27.8, 21.8,13.9; HRMS (ESI) calcd for C₃₄H₃₁N₃O₅S₁Na₁ [M+Na]⁺ : 616.1877; found: 616.1873.

Ethyl 2-(2,2-*dicyano-1-(3,4-dimethylphenyl)vinyl)-2'-oxo-1'-tosylspiro[cyclopropane-1,3'-indoline]-2-carboxylate* (**3***l*) white solid, 53.6mg, 95% yield, mp 84–86 °C, IR (KBr) cm⁻¹: 2982, 2924, 2256, 2229, 1734, 1605, 1462, 1380, 1256, 1238, 1190, 1177, 1086, 962, 910, 814, 753, 733, 665, 574, 542; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.11 (d, J = 8.2 Hz), 8.06 (d, 1H, J = 8.2 Hz), 7.45 (d, 2H), 7.35–7.42 (m, 2H), 7.25 (d, 3H, J = 4.1 Hz), 7.14 (t, 1H, J = 7.6 Hz), 4.17–4.38 (m, 2H), 2.78 (d, 1H, J = 5.7 Hz), 2.49 (s, 3H), 2.34 (s, 3H), 2.33 (s, 3H), 1.74 (d, 1H, J = 5.7 Hz), 1.17 (t, 3H, J = 7.12 Hz,); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.2, 169.7, 164.2, 146.4, 142.1, 140.5,137.5, 134.0, 132.3, 130.2, 129.7, 129.0, 128.7, 125.9, 124.5, 122.9, 122.8, 113.7, 112.7, 112.1, 89.1, 63.9, 47.0, 43.8, 27.6, 21.8, 20.0, 19.9, 13.9; HRMS (ESI) calcd for C₃₂H₂₇N₃O₅S₁Na₁ [M+Na]⁺: 588.1564; found: 588.1559.

Ethyl 5'-chloro-2-(2,2-dicyano-1-phenylvinyl)-2'-oxo-1'-tosylspiro[cyclopropane-1,3'-indoline]-2-carboxylate (*3m*) white solid, 54.8mg, 96% yield, mp 176–178 °C, IR (KBr) cm⁻¹: 2983, 2927, 2256, 2231, 1734, 1597, 1582, 1460, 1383, 1254, 1238, 1190, 1178, 1088, 910, 819, 733, 663, 581, 543; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.08 (d, 2H, *J* = 8.1 Hz), 8.02 (d, 1H, *J* = 8.7 Hz), 7.56–7.49 (m, 3H), 7.47–7.45 (m, 4H), 7.41–7.37 (m, 2H), 4.44–4.21 (m, 2H), 2.77 (d, 1H, *J* = 5.7 Hz), 2.50 (s, 3H), 1.74 (d, 1H, *J* = 5.7 Hz), 1.23 (t, 3H, *J* = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 169.8, 169.6, 164.0, 146.8, 139.1, 134.5, 133.7, 132.3, 130.3, 129.8, 129.1, 128.9, 128.2, 124.6, 123.5, 114.9, 112.2, 111.8, 90.5, 64.3, 47.2, 43.7, 27.8, 21.8, 13.9; HRMS (ESI) calcd for C₃₀H₂₂Cl₁N₃O₅S₁Na₁ [M+Na]⁺ : 594.0861; found: 594.0853

Ethyl 2-(2,2-*dicyano-1-phenylvinyl)-5'-methyl-2'-oxo-1'-tosylspiro[cyclopropane-1,3'-indoline]-2-carboxylate* (**3***n*) white solid, 45.1mg, 82% yield, mp 160–162 °C, IR (KBr) cm⁻¹: 2961, 2927, 2257, 1754, 1719, 1596, 1485, 1383, 1262, 1237, 1191, 1178, 1021, 911, 815, 734, 664, 569, 545; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.09 (d, 1H, J = 8.2 Hz), 7.94 (d, 1H, J = 8.4 Hz), 7.55–7.50 (m, 5H), 7.44 (d, 2H, J = 8.2 Hz), 7.20 (d, 1H, J = 8.4 Hz), 7.16 (s,1H), 4.40–4.18 (m, 2H), 2.74 (d, 1H, J = 5.6 Hz), 2.48 (s, 3H), 2.32 (s, 3H), 1.69 (d, 2H, J = 5.6 Hz), 1.20 (t, 3H, J = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 170.4, 170.0, 164.2, 146.4, 138.2, 134.7, 134.3, 134.1, 132.1, 130.3, 129.8, 128.7, 128.2, 123.4, 122.8, 113.5, 112.4, 111.8, 90.4, 64.0, 46.9, 44.1, 27.5, 21.8, 21.2, 13.9; HRMS (ESI) calcd for C₃₁H₂₅N₃O₅S₁Na₁[M+Na]⁺: 574.1407; found: 574.1404

Ethyl 2,2-dicyano-2'-oxo-3-phenyl-1'-tosylspiro[cyclopent[3]ene-1,3'-indoline]-4-carboxylate (4a) red solid, 44.5mg, 83% yield, mp 202–204 °C, IR (KBr) cm⁻¹: 2979, 2959, 2869, 2249, 2230, 1958, 1755, 1730, 1605, 1596, 1465, 1374, 1326, 1239, 1226, 1179, 1157, 1102, 1088, 1018, 962, 767, 717, 666, 572, 545; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.04 (d, 1H, J = 8.3 Hz), 8.00 (d, 2H, J = 8.4 Hz), 7.56–7.52 (m, 1H), 7.47–7.43 (m, 5H), 7.35–7.31 (m, 3H), 4.07 (q, 2H, J = 7.1Hz), 3.30 (dd, 2H, J = 64.3 Hz, 17.6 Hz), 2.40 (s, 3H), 1.03 (t, 3H, J = 7.1Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 171.8, 162.2, 146.5, 142.7, 139.3, 135.2, 134.1, 131.7, 130.1, 130.0, 128.5, 128.1, 125.8, 124.7, 123.4, 114.4, 111.1, 110.1, 61.6, 58.1, 54.0, 42.3, 21.7, 13.6; HRMS (ESI) calcd for C₃₀H₂₃N₃O₅S₁Na₁[M+Na]⁺ : 560.1251; found: 560.1245.

Ethyl 2,2-*dicyano*-2'-*oxo*-3-(*p*-*tolyl*)-1'-*tosylspiro*[*cyclopent*[3]*ene*-1,3'-*indoline*]-4-*carboxylate* (4d) red solid 42.4mg, 77% yield, mp 64–66 °C, IR (KBr) cm⁻¹: 2983, 2925, 2257, 1754, 1717, 1603, 1475, 1383, 1237, 1191, 1178, 1150, 1088, 1018, 959, 911, 815, 733, 661, 569, 544; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.04 (d, 1H, *J* = 8.2 Hz), 7.99 (d, 2H, *J* = 8.4 Hz), 7.69–7.67 (m, 1H), 7.53 (td,1H, *J* = 8.3 Hz, 1.0 Hz), 7.36–7.30(m, 5H), 7.24 (d, 2H, 9.0 Hz), 4.09 (q, 2H, *J* = 7.1 Hz), 3.28 (dd, 2H, *J* = 17. 6 Hz, 68.7 Hz), 2.40 (s, 3H), 2.38 (s, 3H), 1.08 (t, 3H, *J* = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 171.8, 162.2, 146.4, 143.0, 140.4, 139.2, 134.5, 134.1, 131.7, 130.0, 129.2, 128.2, 128.1, 127.2, 125.7, 124.7, 123.5, 114.4, 111.1, 110.2, 61.5, 58.0, 53.9, 42.3, 21.7, 21.4, 13.7; HRMS (ESI) calcd for C₃₁H₂₅N₃O₅S₁Na₁[M+Na]⁺ : 574.1407; found: 574.1400.

Ethyl 2,2-*dicyano-3-(4-methoxyphenyl)-2'-oxo-1'-tosylspiro[cyclopent[3]ene-1,3'-indoline]-4-carboxylate* (*4e*) yellow solid, 44.2mg, 78% yield, mp 44–46 °C, IR (KBr) cm⁻¹ :2982, 2961, 2257, 2227, 1754, 1728, 1605, 1511, 1463, 1383, 1295, 1253, 1237, 1178, 1151, 1088, 1024, 960, 910, 841, 813, 733, 661, 569, 544; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.04 (d, 1H, *J* = 8.3Hz), 7.99 (d, 2H, *J* = 8.4 Hz), 7.67 (dd, 1H, *J* = 0.9 Hz, 7.7 Hz), 7.53 (td, 1H, *J* = 1.2 Hz, 8.2 Hz), 7.43–7.41 (m, 2H), 7.33–7.29 (m, 3H), 6.95 (d, 2H, *J* = 8.8 Hz), 4.10 (q, 2H, *J* = 7.1 Hz), 3.83 (s, 3H), 3.37 (dd, 2H, *J* = 68.7 Hz, 17.6 Hz), 2.40 (s, 3H), 1.11(t, 3H, *J* = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 171.8, 162.4, 161.1, 146.5, 142.9, 139.3, 134.2, 133.9, 131.7, 130.1, 130.0, 128.1, 125.7, 124.7, 123.6, 122.3, 114.4, 114.0, 61.6, 58.0, 55.3, 42.4, 21.8, 18.4, 13.8; HRMS (ESI) calcd for C₃₁H₂₅N₃O₆S₁Na₁[M+Na]⁺: 590.1356; found: 590.1348.

Ethyl 3-(3-chlorophenyl)-2,2-dicyano-2'-oxo-1'-tosylspiro[cyclopent[3]ene-1,3'-indoline]-4-carboxylate (*4h*) red solid, 34.2mg, 60% yield, mp 60–62 °C, IR (KBr) cm⁻¹: 2926, 2256, 2234, 1736, 1597, 1463, 1380, 1238, 1178, 1087, 962, 910, 812, 733, 665, 575, 541; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.04 (d, 1H, J = 8.3Hz), 8.00 (d, 2H, J = 8.4 Hz), 7.69 (d, 1H, J = 7.69 Hz), 7.55 (td, 1H, J = 7.3 Hz, 1.0Hz), 7.46–7.44 (m, 2H), 7.42–7.38 (m, 1H), 7.36–7.32 (m, 4H), 4.09 (q, 2H, J = 7.12 Hz), 3.38 (dd, 2H, J = 56.5 Hz, 17.7 Hz), 2.40 (s, 3H), 1.07 (t, 3H, J = 7.16 Hz); ¹³C NMR (100MHz, CDCl₃, TMS): δ 171.8, 161.8, 146.6, 141.0, 139.3, 136.6, 134.0, 131.89, 131.86, 130.3, 130.1, 129.9, 128.7, 128.1, 126.4, 125.8, 122.9, 114.4, 110.9, 109.9, 61.8, 58.2, 53.7, 42.2, 21.8, 13.6; HRMS (ESI) calcd for C₃₀H₂₂Cl₁N₃O₅S₁Na₁ [M+Na]⁺ : 594.0861; found: 594.0853.

Ethyl 3-(3-bromophenyl)-2,2-dicyano-2'-oxo-1'-tosylspiro[cyclopent[3]ene-1,3'-indoline]-4-carboxylate (*4o*) red solid, 39.9mg, 65% yield, mp 62–66 °C, IR (KBr) cm⁻¹: 2983, 2926, 2258, 1763, 1720, 1599, 1464, 1384, 1237, 1191, 1179, 1151, 1088, 959, 910, 732, 661, 569, 544; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.04 (d, 1H, *J* = 8.3 Hz), 8.00 (d, 2H, *J* = 8.4 Hz), 7.69 (d, 1H, *J* = 7.6 Hz), 7.61–7.59 (m, 2H), 7.55 (td, 1H, *J* = 8.4 Hz, 11.3 Hz), 7.41–7.39 (m, 1H), 7.36–7.31 (m,4H), 4.08 (q, 2H, *J* = 7.1 Hz), 3.37 (dd, 2H, *J* = 17.5 Hz, 70.2 Hz), 2.41 (s, 3H), 1.33 (s, 9H), 1.03 (t, 3H, *J* = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 171.8, 161.8, 146.6, 140.9, 139.4, 136.3, 134.0, 133.3, 132.1, 131.9, 131.5, 130.1, 128.2, 126.8, 125.8, 124.8, 122.9, 114.4, 110.9, 109.9, 61.8, 58.2, 53.7, 42.2, 21.8, 13.6; HRMS (ESI) calcd for C₃₀H₂₂Br₁N₃O₅S₁Na₁ [M+Na]⁺ : 638.0356; found: 638.0348

Ethyl 2,2-dicyano-3-(furan-2-yl)-2'-oxo-1'-tosylspiro[cyclopent[3]ene-1,3'-indoline]-4-carboxylate (4p) red liquid, 30.5mg, 58% yield, IR (KBr) cm⁻¹: 2959, 2926, 2256, 2228, 1752, 1723, 1607, 1598, 1460, 1382, 1236, 1187, 1157, 1087, 910, 734, 661, 568, 543; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.06 (d, 1H, *J* = 8.2 Hz), 7.99 (d, 2H, *J* = 8.4 Hz), 7.75 (d, 1H, *J* = 3.6 Hz), 7.66 (dd, 1H, *J* = 0.7 Hz, 7.7 Hz), ⁸

7.51–7.58 (m, 2H), 7.29–7.34 (m, 3H), 4.24–4.32 (m, 2H), 3.39 (dd, J = 17.9 Hz, 32.7 Hz), 2.41 (s, 3H), 1.31 (t, 3H, J = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 171.8, 162.2, 149.5, 146.4, 145.1, 144.0, 143.0, 139.5, 134.0, 131.8, 131.1, 130.0, 128.4, 128.3, 125.6, 125.0, 123.2, 119.3, 114.3, 112.9, 113.4, 110.7, 61.8, 57.6, 50.5, 42.8, 21.8, 14.1; HRMS (ESI) calcd for C₂₈H₂₁N₃O₆S₁Na₁ [M+Na]⁺:550.1043; found: 550.1037

Ethyl 3-(4-(*tert-butyl*)*phenyl*)-2,2-*dicyano*-2'-*oxo*-1'-*tosylspiro*[*cyclopent*[3]*ene*-1,3'-*indoline*]-4-*carboxylate* (**4***k*) red solid, 54.5mg, 92% yield, mp 62–64 °C, IR (KBr) cm⁻¹: 2965, 2869, 2257, 1754, 1718, 1603, 1464, 1384, 1237, 1179, 1150, 1088, 1017, 958, 910, 733, 661, 568, 543; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.04 (d, 1H, *J* = 8.3 Hz), 8.00 (d, 2H, *J* = 8.4 Hz), 7.68 (d, 1H, *J* = 7.16 Hz), 7.53 (td, 1H, *J* = 8.0 Hz, 1.0 Hz), 7.42 (dd, 4H, *J* = 8.4 Hz, 22.6 Hz), 7.35–7.30 (m, 3H), 4.07 (q, 2H, *J* = 7.1 Hz), 3.37 (dd, 2H, *J* = 17.5 Hz, 70.2 Hz), 2.40 (s, 3H), 1.33 (s, 9H), 1.03 (t, 3H, *J* = 7.1 Hz, CH₃); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 171.8, 162.3, 153.5, 146.4, 143.0, 139.3, 134.5, 134.1, 131.7, 130.0, 128.1, 127.1, 125.7, 125.4, 124.7, 123.6, 114.4, 111.2, 110.3, 61.5, 58.0, 53.9, 42.3, 31.1, 21.8, 13.5; HRMS (ESI) calcd for C₃₄H₃₁N₃O₅S₁Na₁[M+Na]⁺ : 616.1877; found : 616.1873

Ethyl 2,2-*dicyano-3-(3,4-dimethylphenyl)-2'-oxo-1'-tosylspiro[cyclopent[3]ene-1,3'-indoline]-4-carboxylate* (**4**) red solid, 53.6mg, 95% yield, mp 61–63 °C, IR (KBr) cm⁻¹: 2981, 2924, 2257, 1756, 1723, 1603, 1464, 1384, 1327, 1238, 1191, 1178, 1151, 1088, 1039, 959, 911, 813, 729, 681, 661, 570, 544; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.04 (d, 1H, J = 8.3Hz), 8.00 (d, 2H, J = 8.3 Hz), 7.68 (d, 1H, J = 7.3Hz), 7.55–7.51 (m, 1H), 7.34–7.30 (m, 3H), 7.21–7.19 (m, 3H), 4.09 (q, 2H, J = 7.1 Hz), 3.36 (dd, 2H, J = 71.0 Hz, 17.5 Hz), 2.40 (s, 3H), 2.28 (s, 6H), 1.09 (t, 3H, J = 7.1 Hz), ¹³C NMR (100 MHz, CDCl₃, TMS): δ 171.8, 162.3, 146.4, 143.1, 139.21, 139.16, 136.8, 136.1, 134.1, 131.7, 130.0, 129.7, 129.3, 128.1, 127.6, 125.72, 125.70, 124.7, 123.6, 114.4, 111.2, 110.3, 61.5, 58.1, 53.9, 42.3, 21.7, 19.8, 13.7; HRMS (ESI) calcd for C₃₂H₂₇N₃O₅S₁Na₁ [M+Na]⁺ : 588.1564; found: 588.1561;

Ethyl 5'-chloro-2,2-dicyano-2'-oxo-3-phenyl-1'-tosylspiro[cyclopent[3]ene-1,3'-indoline]-4-carboxylate (**4m**) red solid, 49.1mg, 86% yield, mp 58–60 °C, IR (KBr) cm⁻¹: 2983, 2929, 2366, 2258, 1889, 1757, 1717, 1656, 1596, 1463, 1444, 1385, 1341, 1283, 1236, 1179, 1191, 1155, 1109, 1088, 1021, 958, 911, 814, 734, 700, 663, 581, 553, 544; ¹H NMR (400 MHz, CDCl₃, TMS): δ 8.00 (d, 1H, J = 8.9 Hz, 1H), 7.97 (d, 2H, J = 8.4 Hz), 7.66 (d, 1H, J = 2.08 Hz), 7.53–7.50 (m, 1H), 7.47–7.44(m, 5H), 7.34 (d, 2H, J = 8.2 Hz), 4.07 (q, 2H, J = 7.1 Hz), 3.38 (dd, 2H, J = 71.1 Hz, 17.1 Hz), 2.40 (s, 3H), 1.04 (t, 3H, J = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 171.2, 161.9, 146.8, 142.6, 137.7, 134.9, 133.7, 131.8, 131.4, 130.2, 130.1, 130.0, 128.6, 128.3, 128.1, 125.1, 125.0, 115.0, 110.8, 109.8, 61.6, 57.8, 53.8, 42.1, 21.8, 13.6; HRMS (ESI) calcd for C₃₀H₂₂Cl₁N₃O₅S₁Na₁[M+Na]⁺: 594.0861; found: 594.0871

Ethyl 2,2-*dicyano-5'-methyl-2'-oxo-3-phenyl-1'-tosylspiro[cyclopent[3]ene-1,3'-indoline]-4-carboxylate* (*4n*) red solid, 44.6mg, 81% yield, mp 54–56 °C, IR (KBr) cm⁻¹: 3061, 2982, 2961, 2927, 2871, 2257, 2231, 1754, 1719, 1656, 1596, 1485, 1444, 1383, 1340, 1298, 1262, 1237, 1191, 1178, 1155, 1113, 1089, 1021, 958, 911, 847, 815, 734, 699, 664, 569, 545; ¹H NMR (400 MHz, CDCl₃, TMS): δ 7.98 (d, 1H, *J* = 8.4 Hz), 7.90 (d, 2H, *J* = 8.4 Hz), 7.48 (s, 1H), 7.47–7.42 (m, 5H), 7.33–7.31 (m, 3H), 4.07 (q, 2H, *J* = 7.1 Hz), 3.36 (dd, 2H, *J* = 59.9 Hz, 17.6 Hz), 2.41 (s, 3H), 2.39 (s, 3H), 1.04 (t, 3H, *J* = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 172.0, 162.2, 146.4, 142.9, 136.9, 135.8, 135.1, 134.2, 132.3, 130.4, 130.1, 130.0, 128.6, 128.4, 128.1, 125.3, 123.3, 114.2, 111.2, 110.2, 61.6, 58.2, 54.0, 42.4, 21.8, 21.2, 13.6; HRMS (ESI) calcd for C₃₁H₂₅N₃O₅S₁Na₁ [M+Na]⁺:574.1407; found: 574.1398

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website with control experiments, NMR spectra, X-ray data (CIF).

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Notes

The authors declare no competing financial interest.

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