

Supporting Information

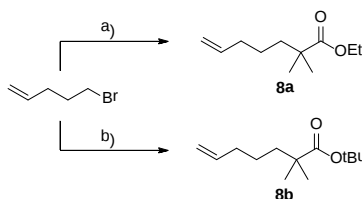
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Rational Design, Synthesis and Biological Evaluation of Modular Fluorogenic Substrates with High Affinity and Selectivity for PTP1B

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I. Synthetic procedures and compound characterization.

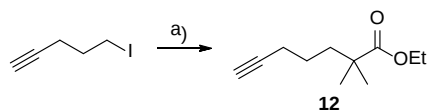


Scheme S1. Reagents and conditions: a) ethyl isobutyrate, LDA, THF, $-78\text{ }^{\circ}\text{C}$ to rt, overnight; b) *t*-butyl isobutyrate, LDA, THF, $-78\text{ }^{\circ}\text{C}$ to rt, overnight.

General procedure for the synthesis of compounds 8aⁱ and 8b: to a stirred solution of freshly prepared LDA (1.1 equiv.) in THF (0.05 M), the opportune isobutyrate (1.0 equiv., ethyl isobutyrate for **8a**; *t*-butyl isobutyrate for **8b**) was added at $-78\text{ }^{\circ}\text{C}$ and under argon atmosphere. After 2 hours 5-bromo-pent-1-ene (1.0 equiv) was added, the mixture was allowed to warm to room temperature and stirred overnight. The reaction was then quenched with HCl 2N and extracted with Et₂O. Organic layers were combined, washed with brine, dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was filtered over a short pad of silica gel by eluting with heptane and used as is for the next step.

Ethyl 2,2-dimethylhept-6-enoate (8a): the spectroscopic data recorded are in agreement with those reported in the reference. ¹H NMR (400 MHz, CDCl₃) δ 5.76 (ddt, $J = 16.9, 10.2, 6.6$ Hz, 1H), 4.98 (dd, $J = 17.1, 1.4$ Hz, 1H), 4.93 (d, $J = 10.2$ Hz, 1H), 4.10 (qd, $J = 7.2, 2.4$ Hz, 2H), 2.05 – 1.97 (m, 2H), 1.53 – 1.48 (m, 2H), 1.37 – 1.25 (m, 2H), 1.23 (t, $J = 7.1$ Hz, 4H), 1.14 (s, 1H) ppm; ¹³C NMR (101 MHz, CDCl₃) δ 178.0, 138.7, 114.6, 60.3, 42.2, 40.3, 34.2, 25.3, 24.4, 14.4 ppm.

***tert*-Butyl 2,2-dimethylhept-6-enoate (8b):** ¹H NMR (400 MHz, CDCl₃) δ 5.78 (ddt, $J = 17.0, 10.2, 6.7$ Hz, 1H), 4.98 (dd, $J = 17.1, 1.4$ Hz, 1H), 4.93 (d, $J = 10.2$ Hz, 1H), 2.02 (q, $J = 7.0$ Hz, 2H), 1.51 – 1.44 (m, 2H), 1.42 (s, 9H), 1.37 – 1.27 (m, 2H), 1.10 (s, 6H) ppm; ¹³C NMR (101 MHz, CDCl₃) δ 177.4, 138.8, 114.6, 79.7, 42.7, 40.3, 34.3, 28.1, 25.3, 24.4 ppm.



Scheme S2. Reagents and conditions: a) ethyl isobutyrate, LDA, THF, -78 °C to rt, overnight.

Synthesis of ethyl 2,2-dimethylhept-6-ynoate (12): to a stirred solution of freshly prepared LDA (1.1 equiv.) in THF (0.05 M), ethyl isobutyrate (1.1 equiv.) was added at -78 °C and under argon atmosphere. After 2 hours 5-iodopent-1-yne (1.0 equiv) was added, the mixture was allowed to warm to room temperature and stirred overnight. The reaction was then quenched with HCl 2N and extracted with Et₂O. Organic layers were combined, washed with brine, dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was filtered over a short pad o silica gel by eluting with heptane and used as is for the next step. ¹H NMR (400 MHz, CDCl₃) δ 4.12 (q, *J* = 7.1 Hz, 2H), 2.17 (td, *J* = 7.0, 2.6 Hz, 2H), 1.95 (t, *J* = 2.6 Hz, 1H), 1.66 – 1.59 (m, 2H), 1.52 – 1.42 (m, 2H), 1.25 (t, *J* = 7.2 Hz, 3H), 1.18 (s, 6H) ppm; ¹³C NMR (101 MHz, CDCl₃) δ 177.8, 84.3, 68.5, 60.4, 42.1, 39.9, 25.2, 24.2, 19.0, 14.3 ppm.

ABBREVIATIONS USED

TMSBr, bromotrimethylsilane; DCM, dicloromethane; SiO₂, silica gel; MOMCl, chloromethyl methyl ether; DIPEA, N,N-diisopropylethylamine; DMAP, 4-dimethylaminopyridine; DIBAL-H, diisobutylaluminium hydride; TEA, triethylamine; LDA, lithium diisopropylamide; LiTMP, lithium tetramethylpiperidine.

II. Stability Studies

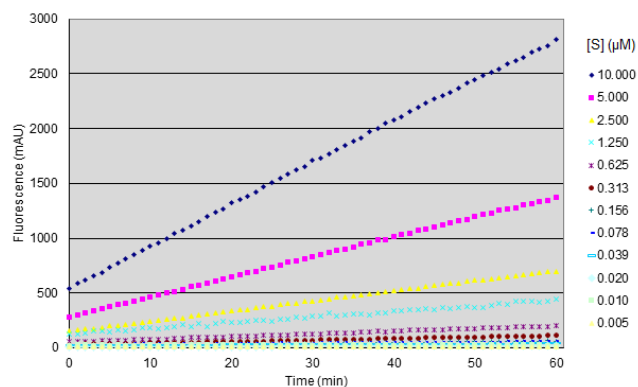


Figure S1. Spontaneous degradation of compound **4f**. Compound **4f** was serially diluted (10 μM , 5 μM , 2.5 μM , 1.25 μM , 0.625 μM , 0.313 μM , 0.156 μM , 0.078 μM , 0.039 μM , 0.020 μM , 0.010 μM and 0.005 μM) in cold reaction buffer (50 mM HEPES, pH 7.1, 50 mM KCl, 1mM EDTA, 3 mM DTT, 0.05% NP-40) for a total volume of 25 μl . Fluorescence production (mAU, y-axis) was monitored ($\lambda_{\text{ex}} = 342 \text{ nm}$, $\lambda_{\text{em}} = 441 \text{ nm}$) every minute for one hour. The assay was run in duplicate.

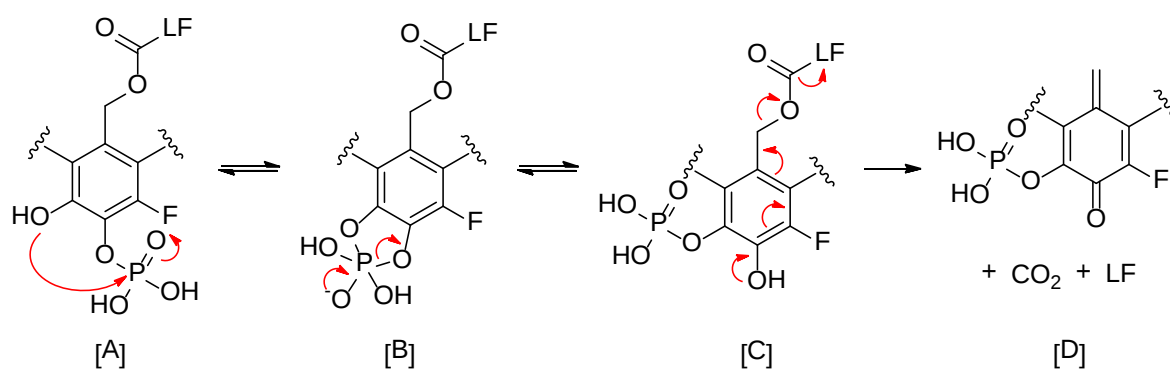


Figure S2. Proposed decomposition mechanism for compounds **4h** and **4k**. [A] When unmasked, the free phenol attacks the phosphate group to generate an unstable pentacyclic intermediate [B] which re-opens and triggers the 1,6-elimination [C] with subsequent decomposition of the scaffolds [D].

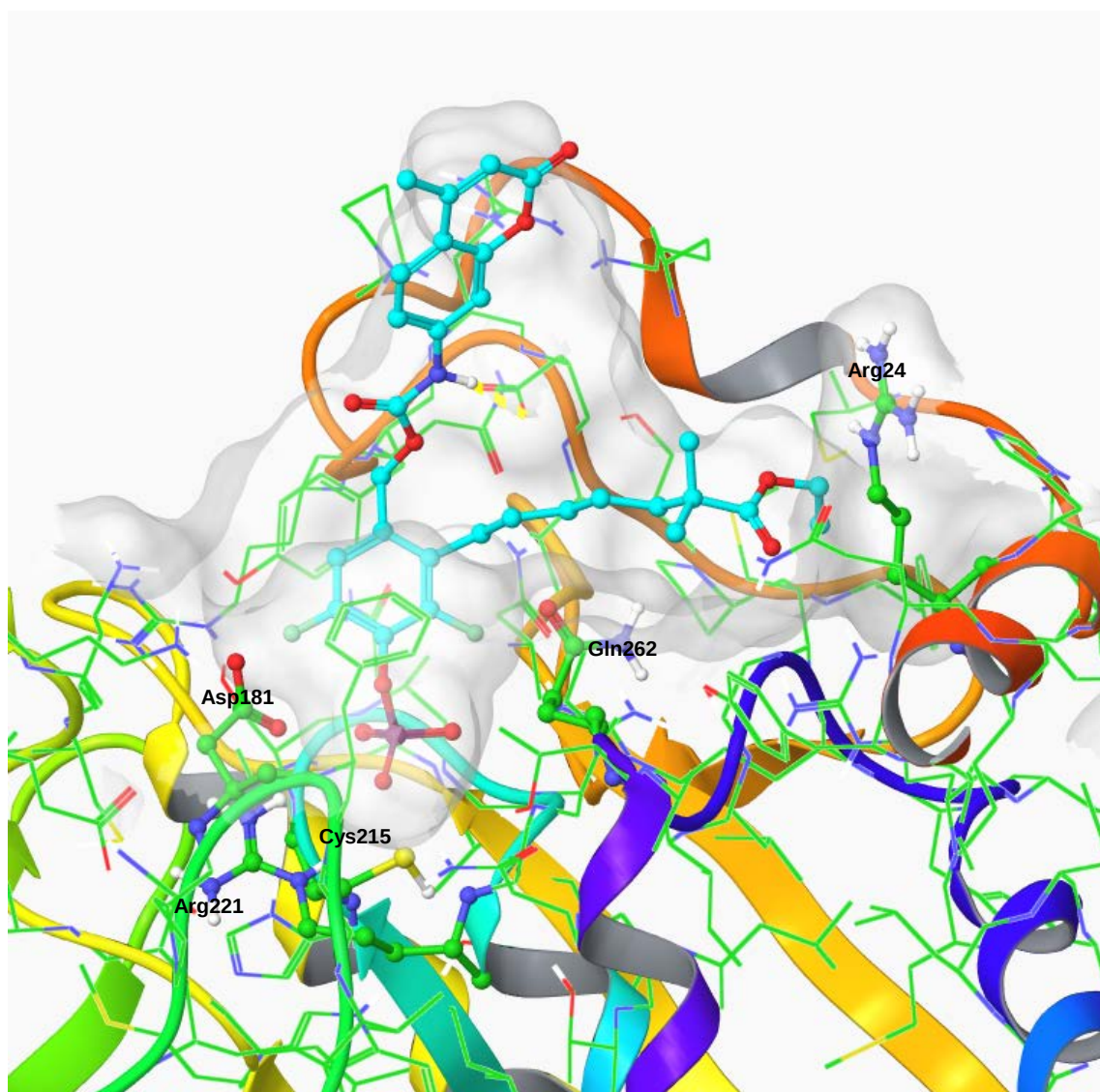
III. Computational Studies

Table S1 – Graphical summary of the BLAST output

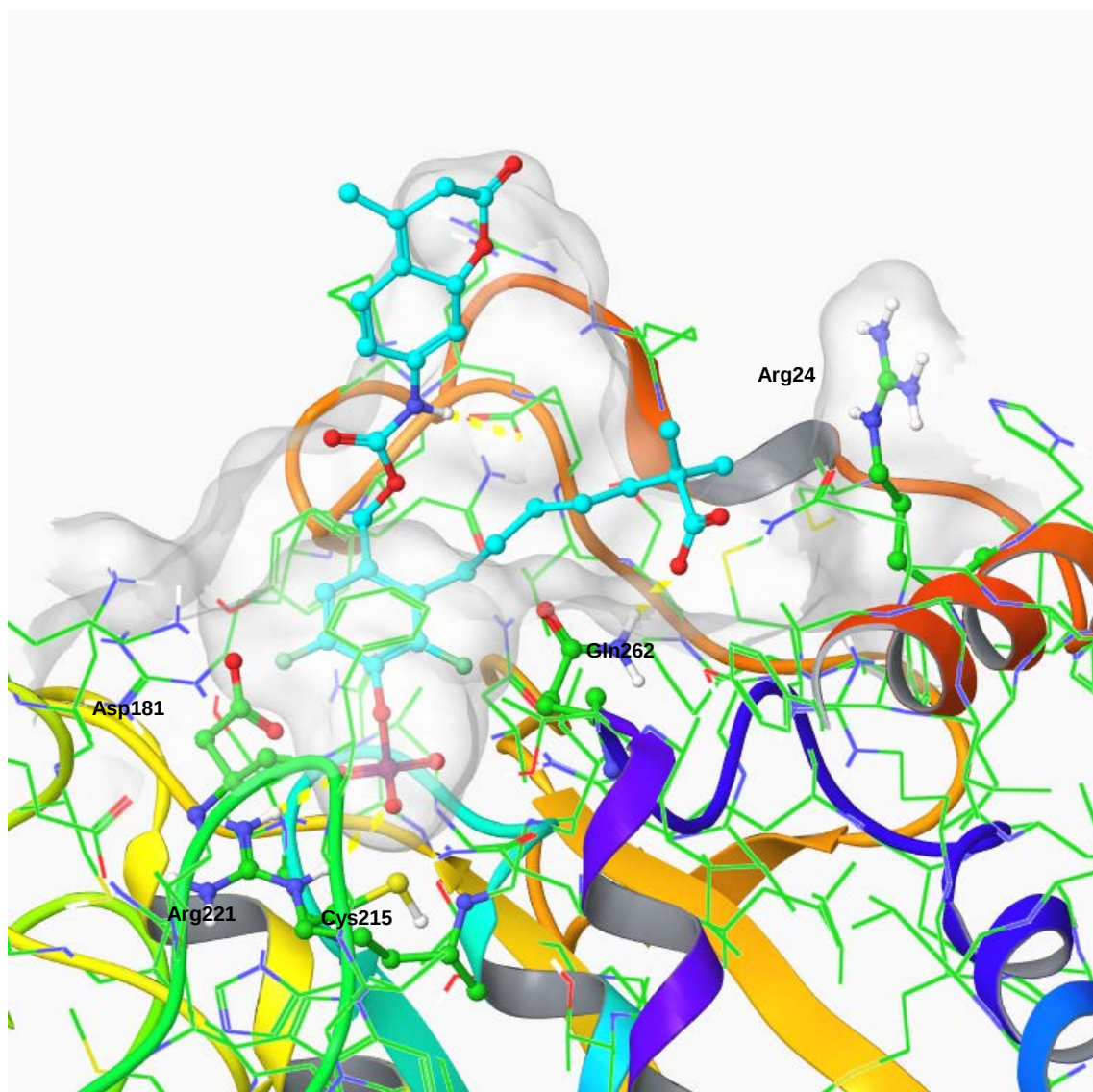
Protein	Pdb ID	Score ^a	Identities ^b	Positives ^c
TCPTP	1L8K	449.5	68 %	84 %
PTP- β	2I4E	197.2	39%	58%
SHP1	305X	186.2	38 %	59 %
LAR	1LAR	182.6	35 %	51 %
CD45	1YGR	115.2	25 %	45%
VHR*	ND	ND	ND	ND
PP2A*	ND	ND	ND	ND

a) numerical value describing the overall quality of an alignment: higher numbers correspond to a higher similarity; b) parameter indicating amino acid residues that are identical in the query (PTP1B) and in the hit when the two are optimally aligned; c) parameter indicating residues that are similar to each other.

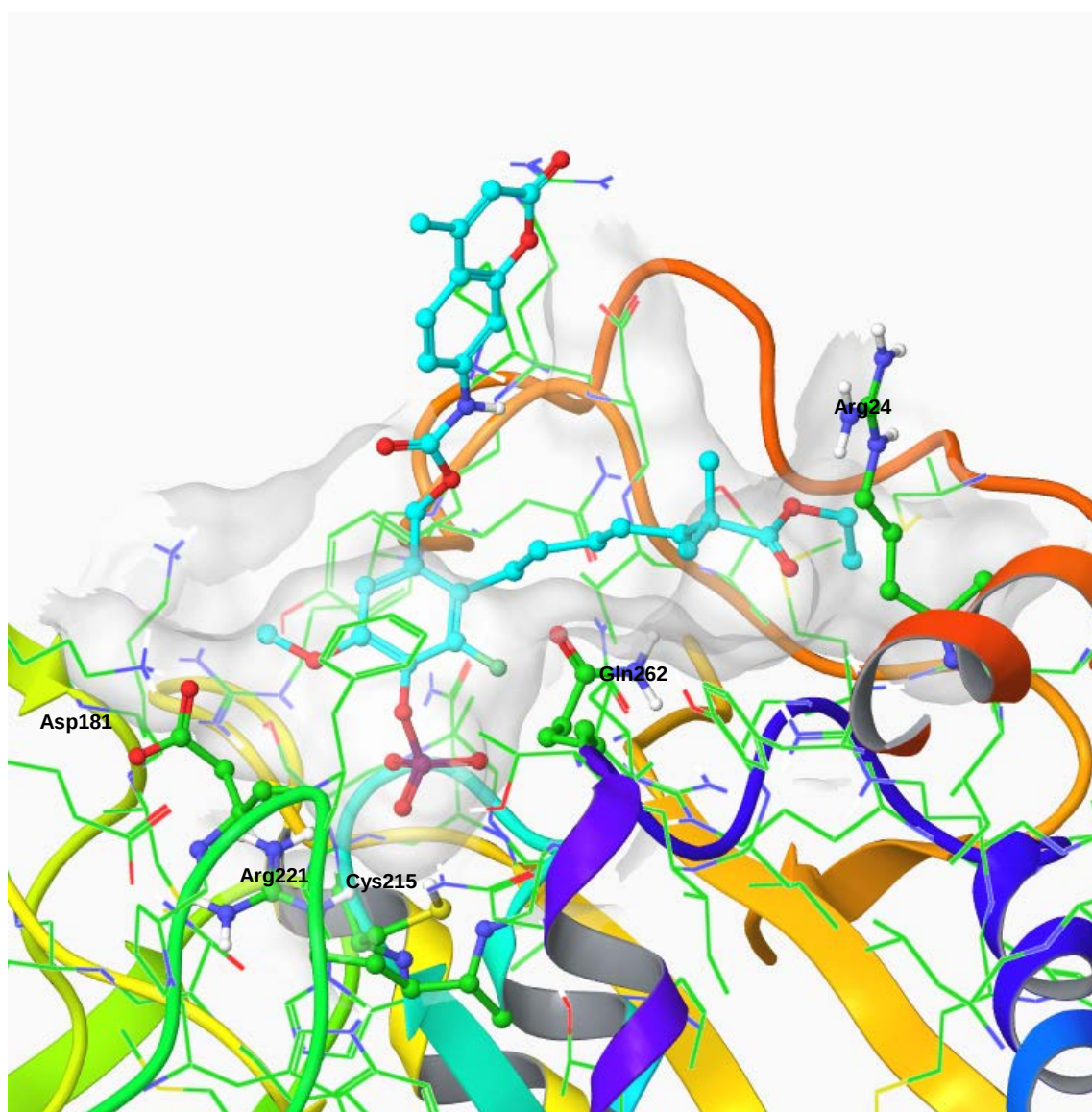
BLAST output highlights a very high degree of similarity between PPT1B and TCPTP. A minor degree of homology was found in PTP- β , SHP1, LAR and CD45 whereas PP2A and VHR scored below the threshold value. The obtained data was in complete agreement with the biological results on the potential secondary targets for the synthesized substrates.



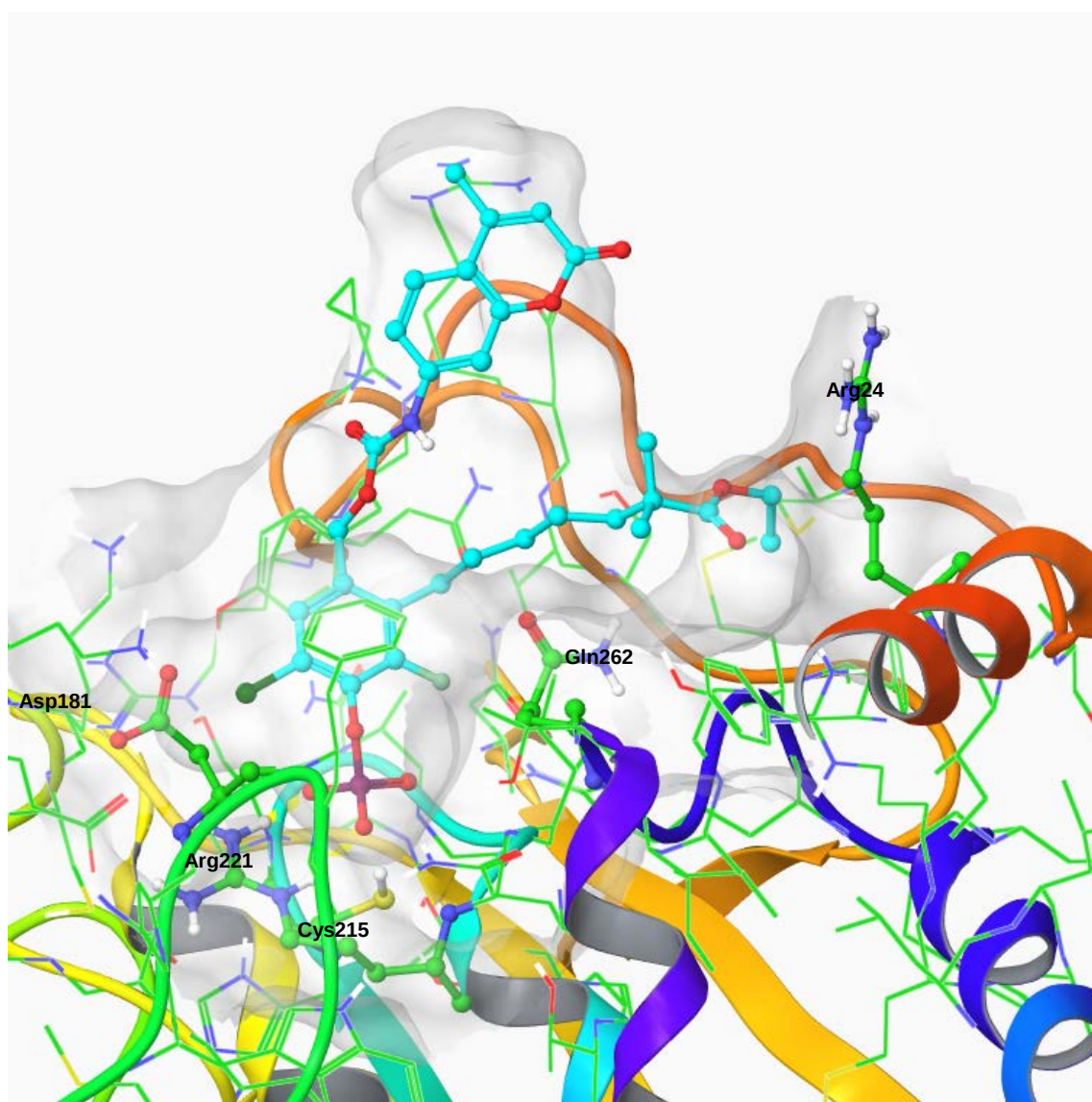
Compound **4d** docked into the active site of 2BGE



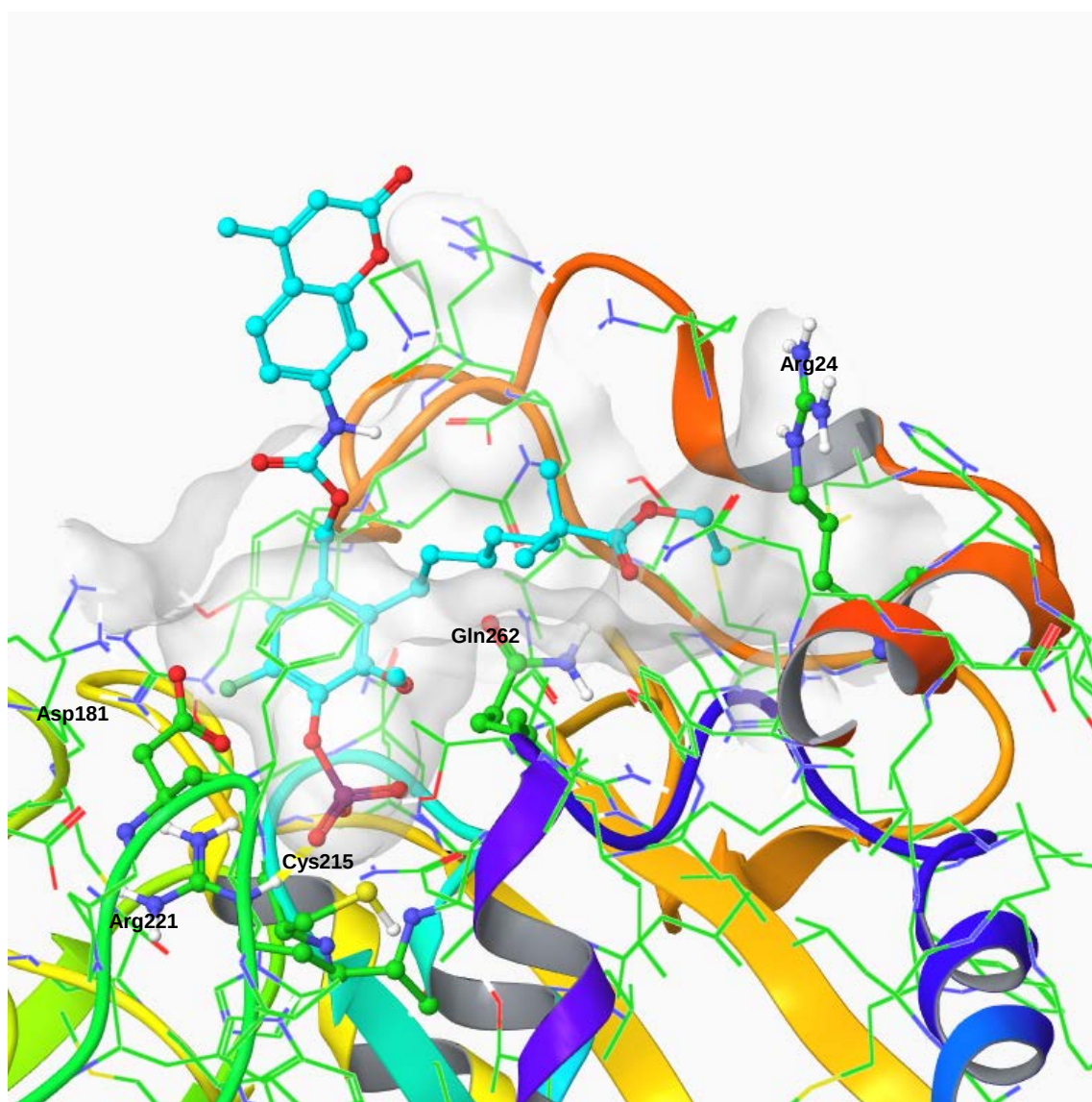
Compound **4e** docked into the active site of 2BGE



Compound **4g** docked into the active site of 2BGD

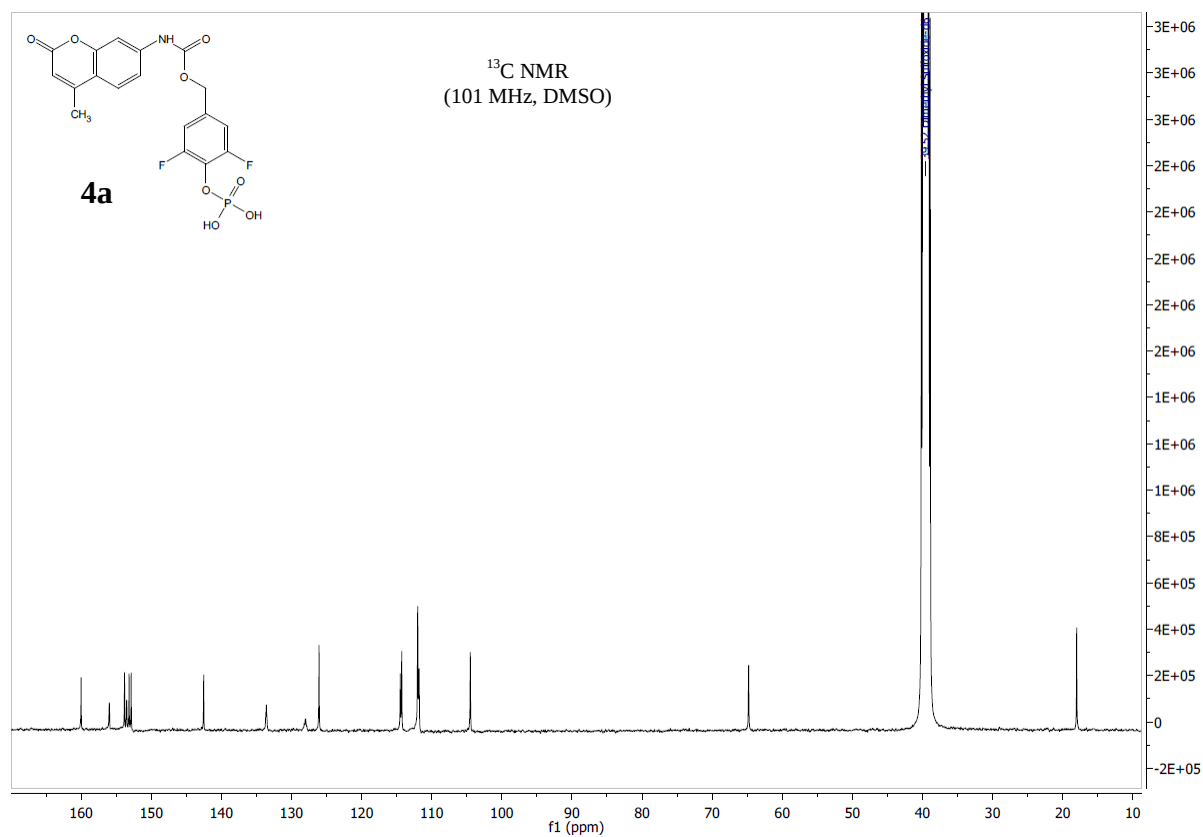
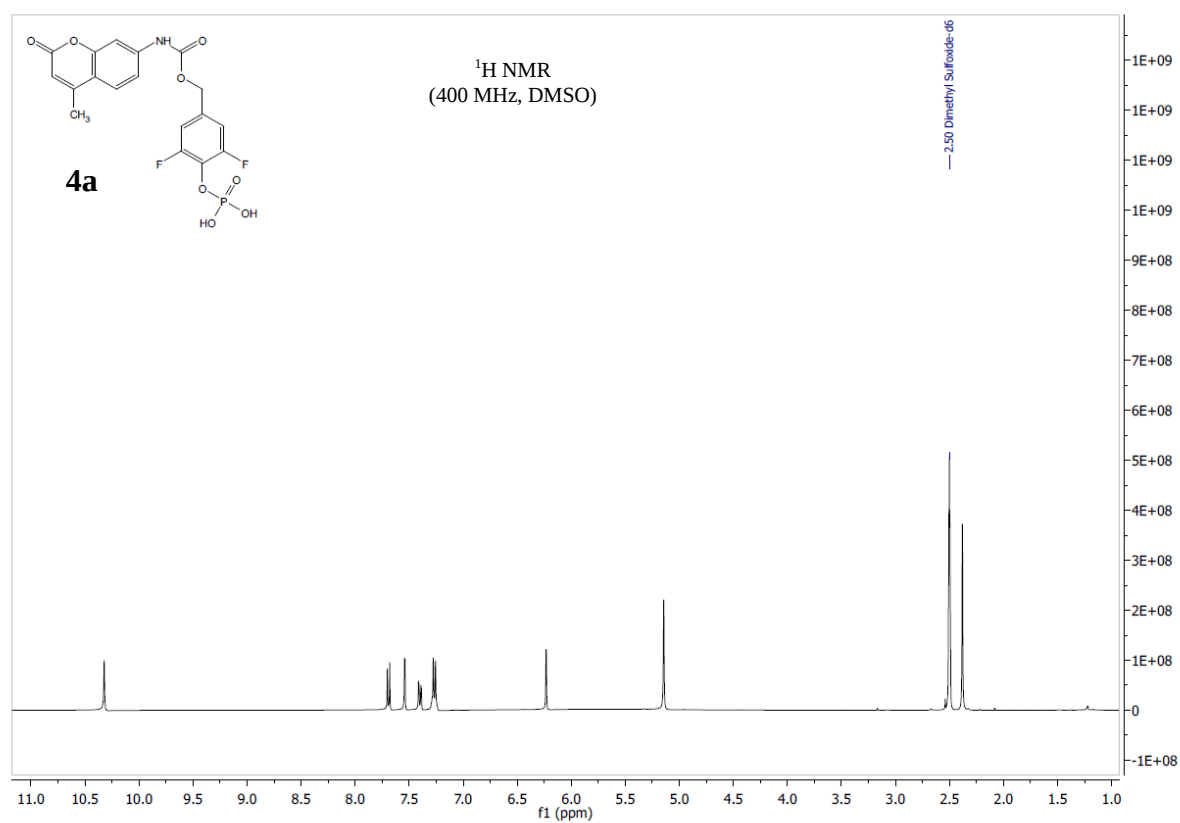


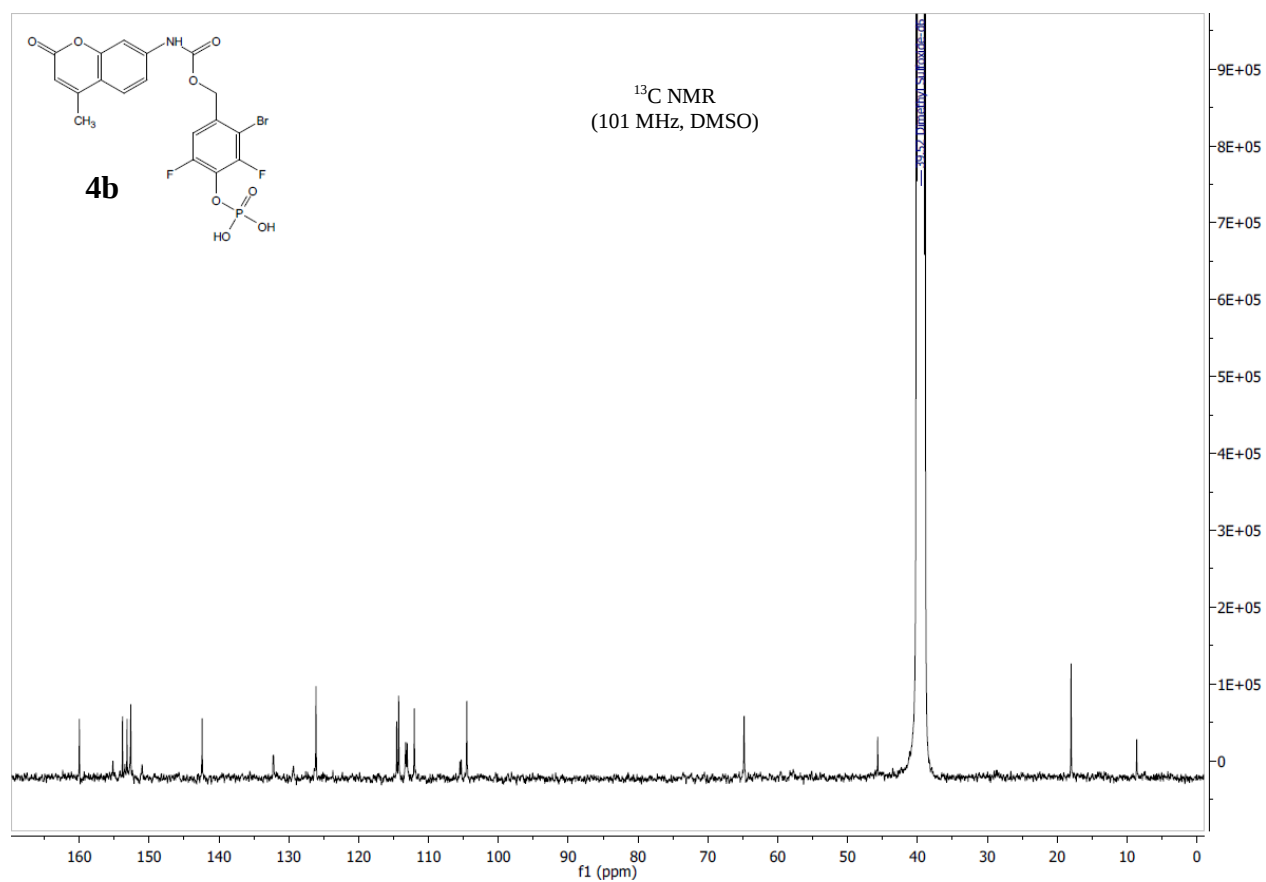
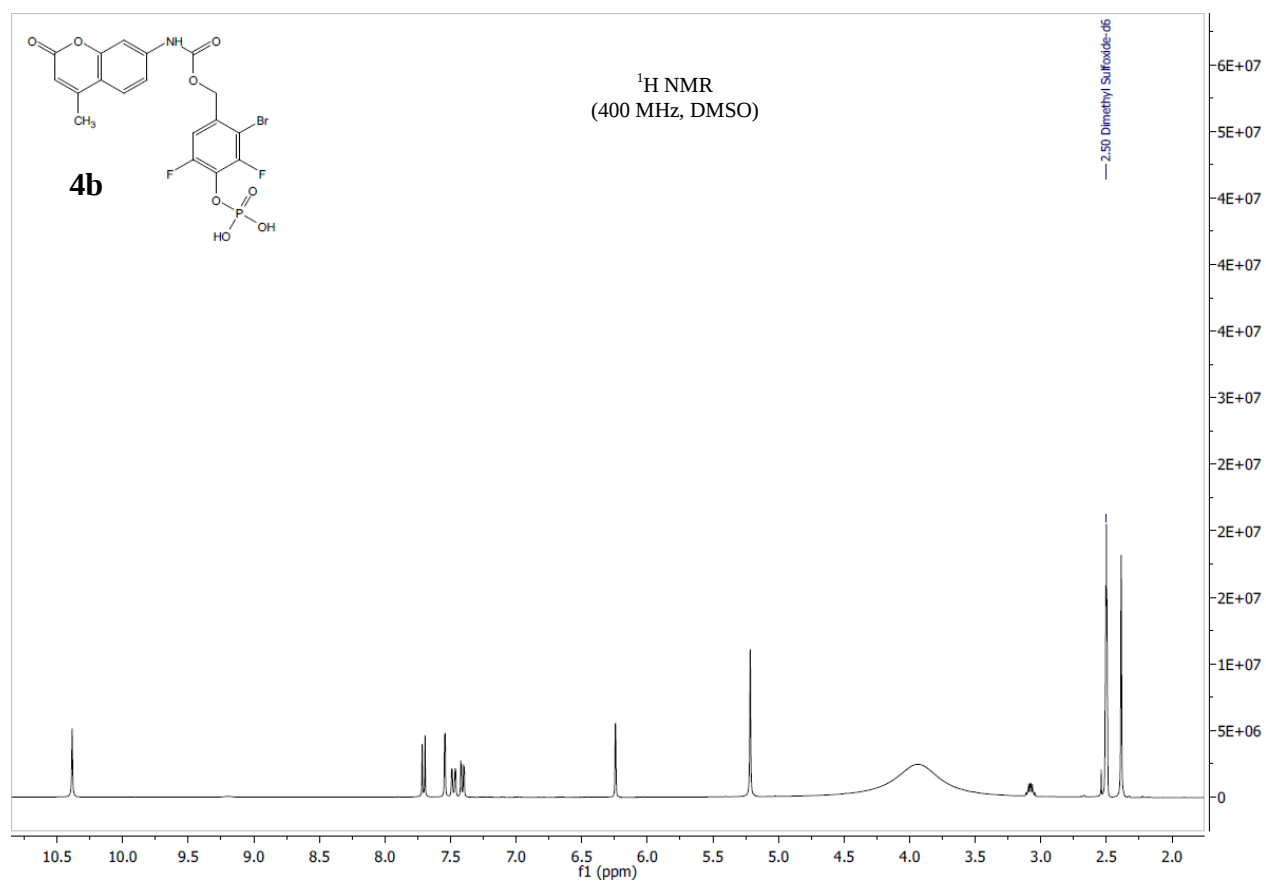
Compound **4i** docked into the active site of 2BGD

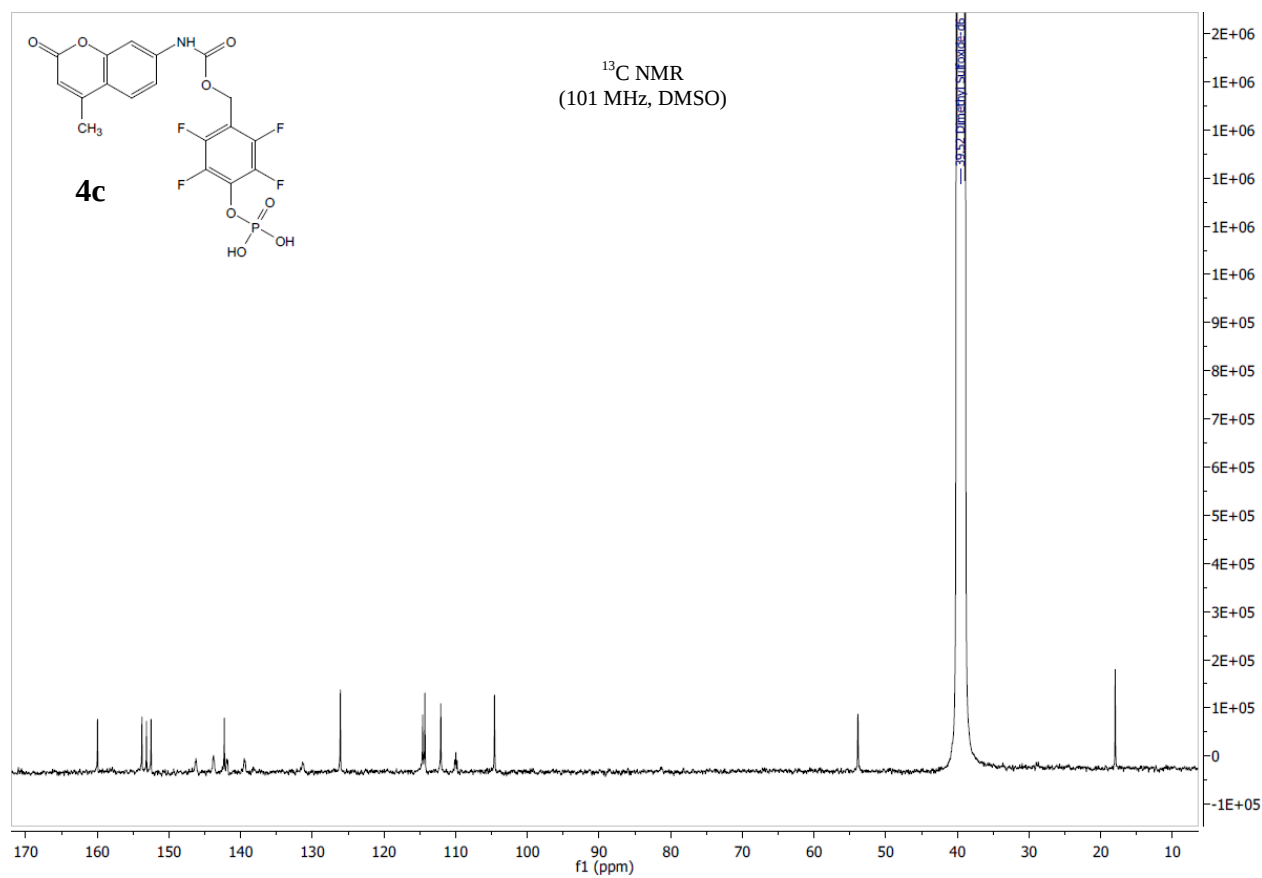
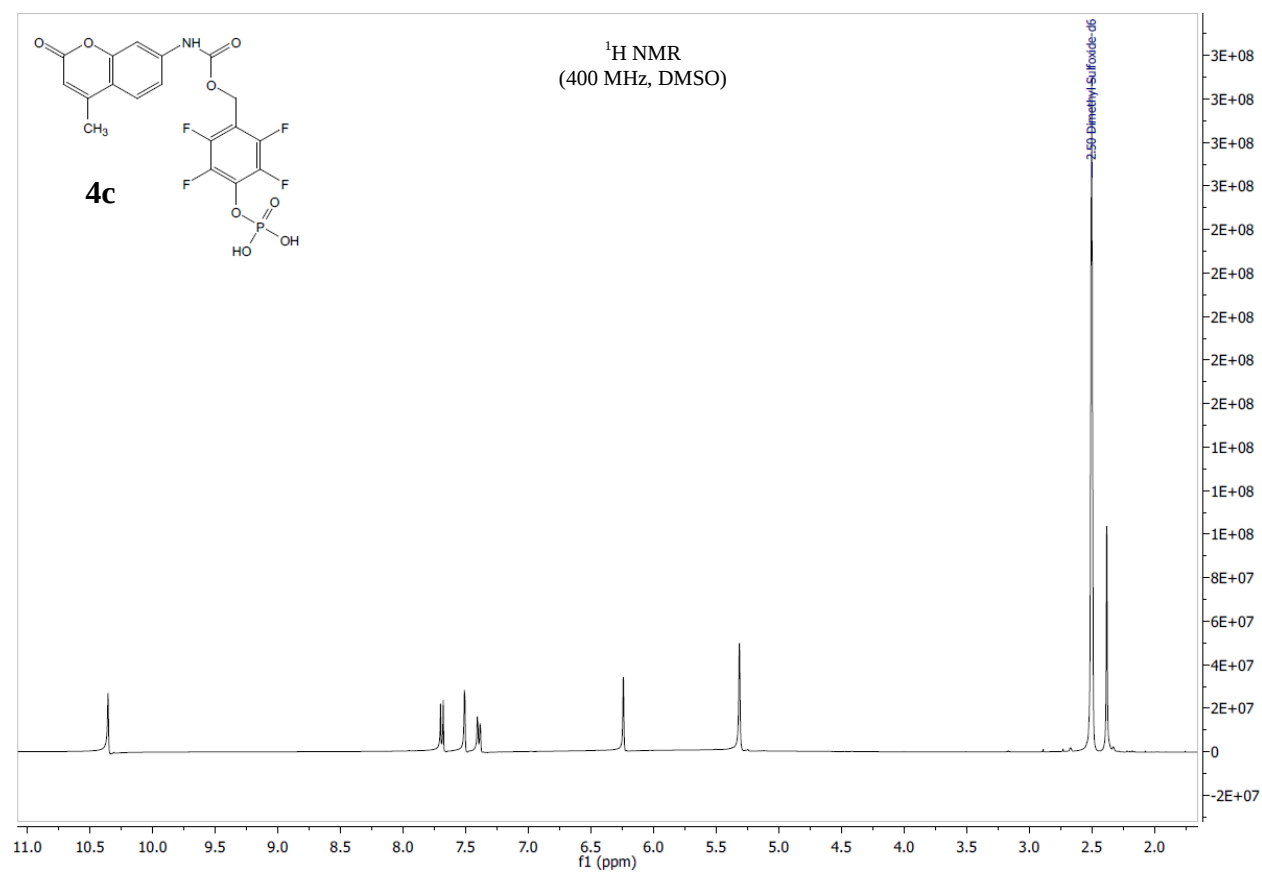


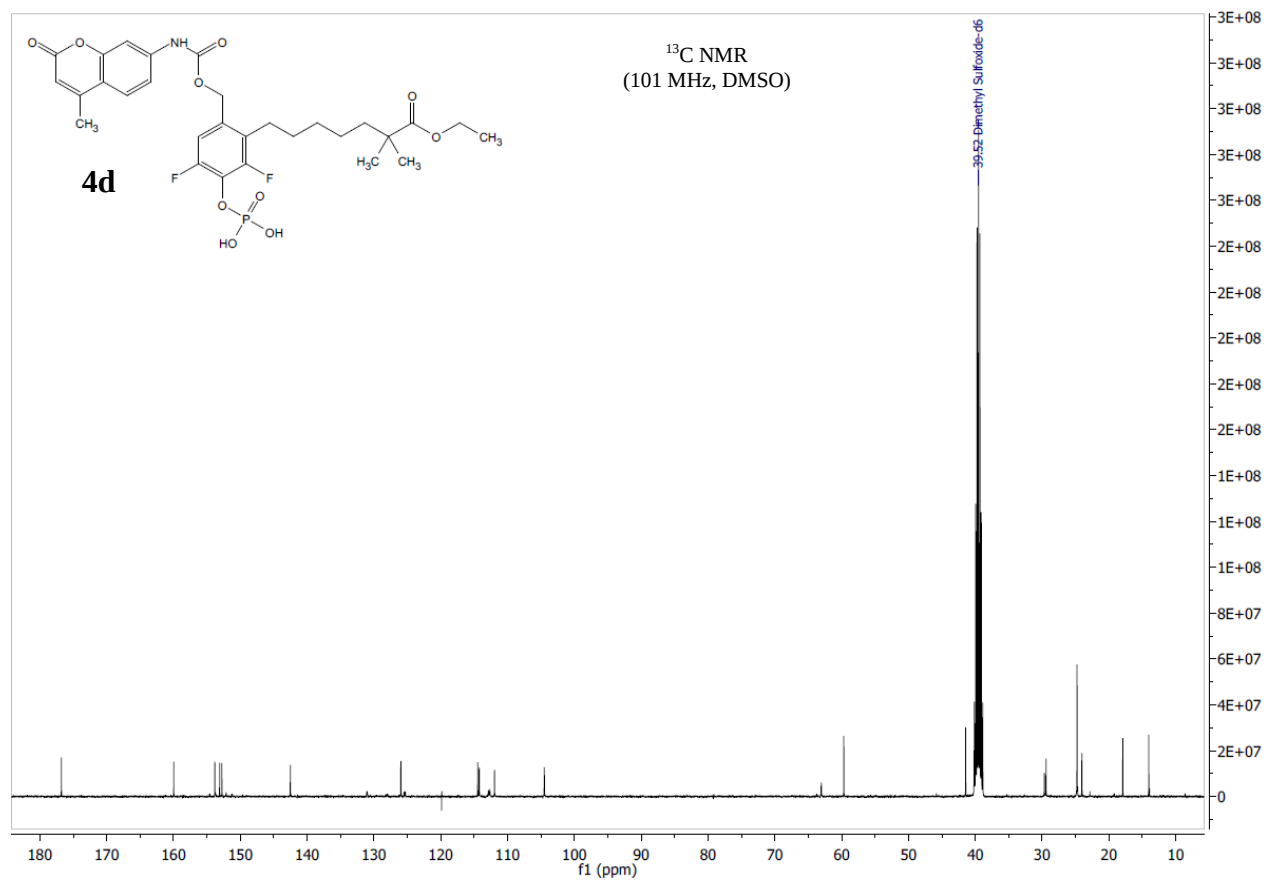
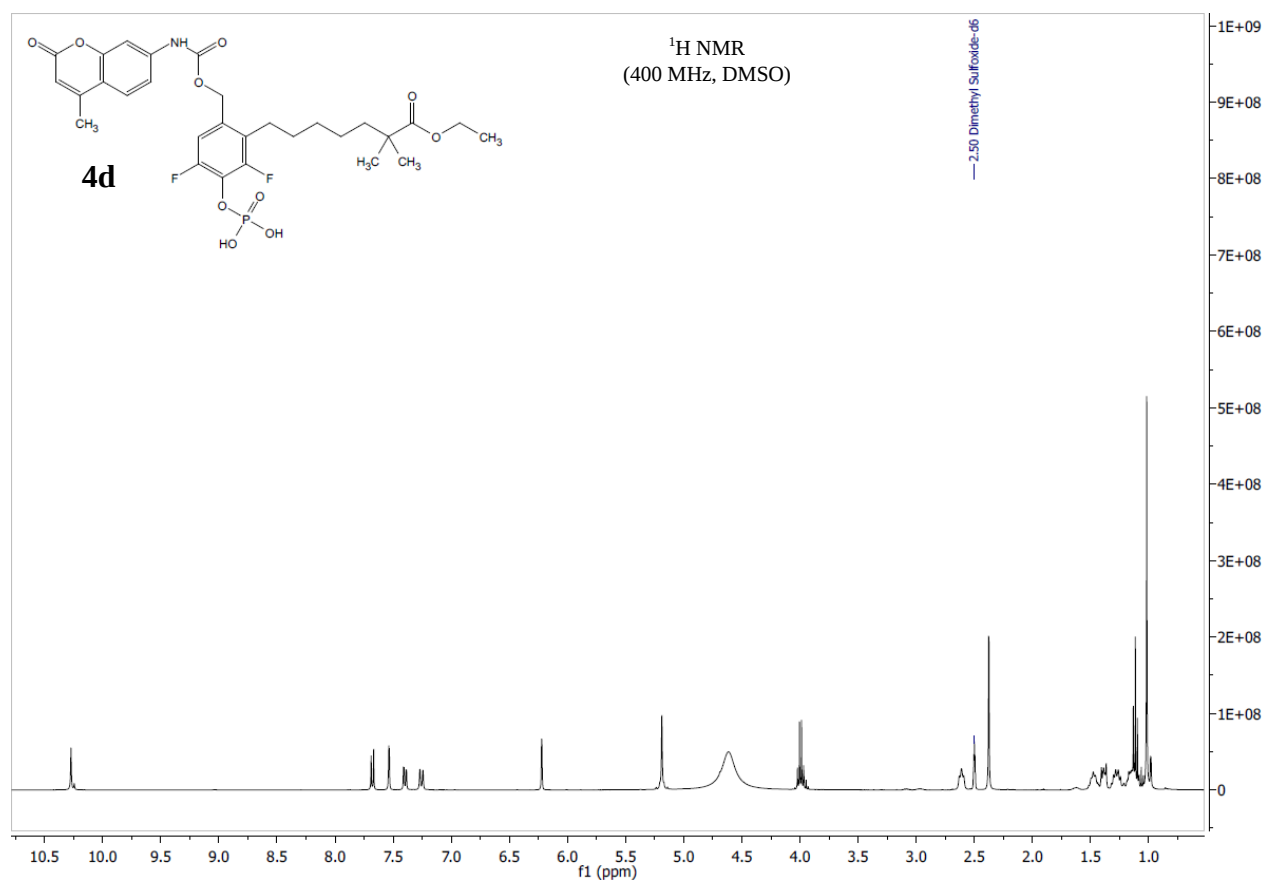
Compound **4j** docked into the active site of 2BGE

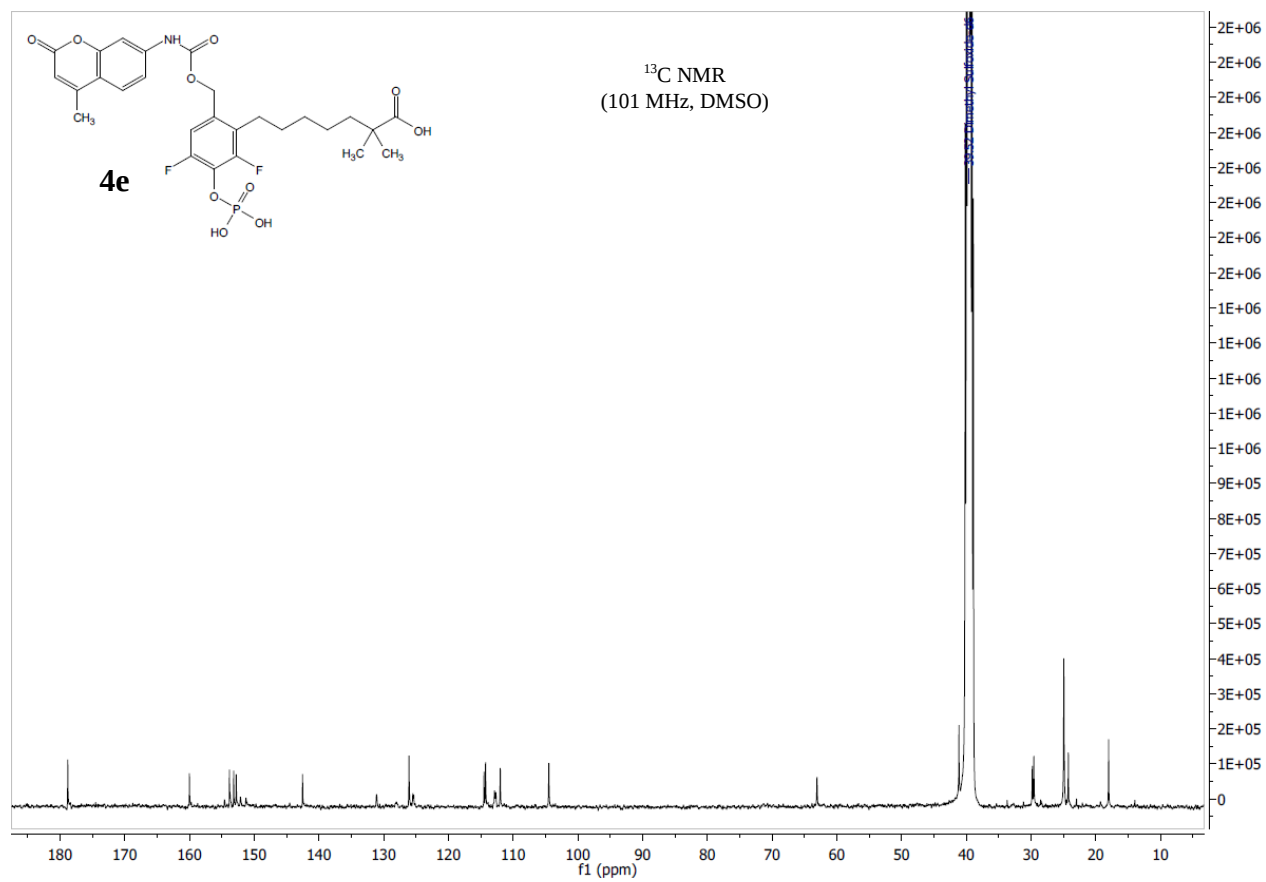
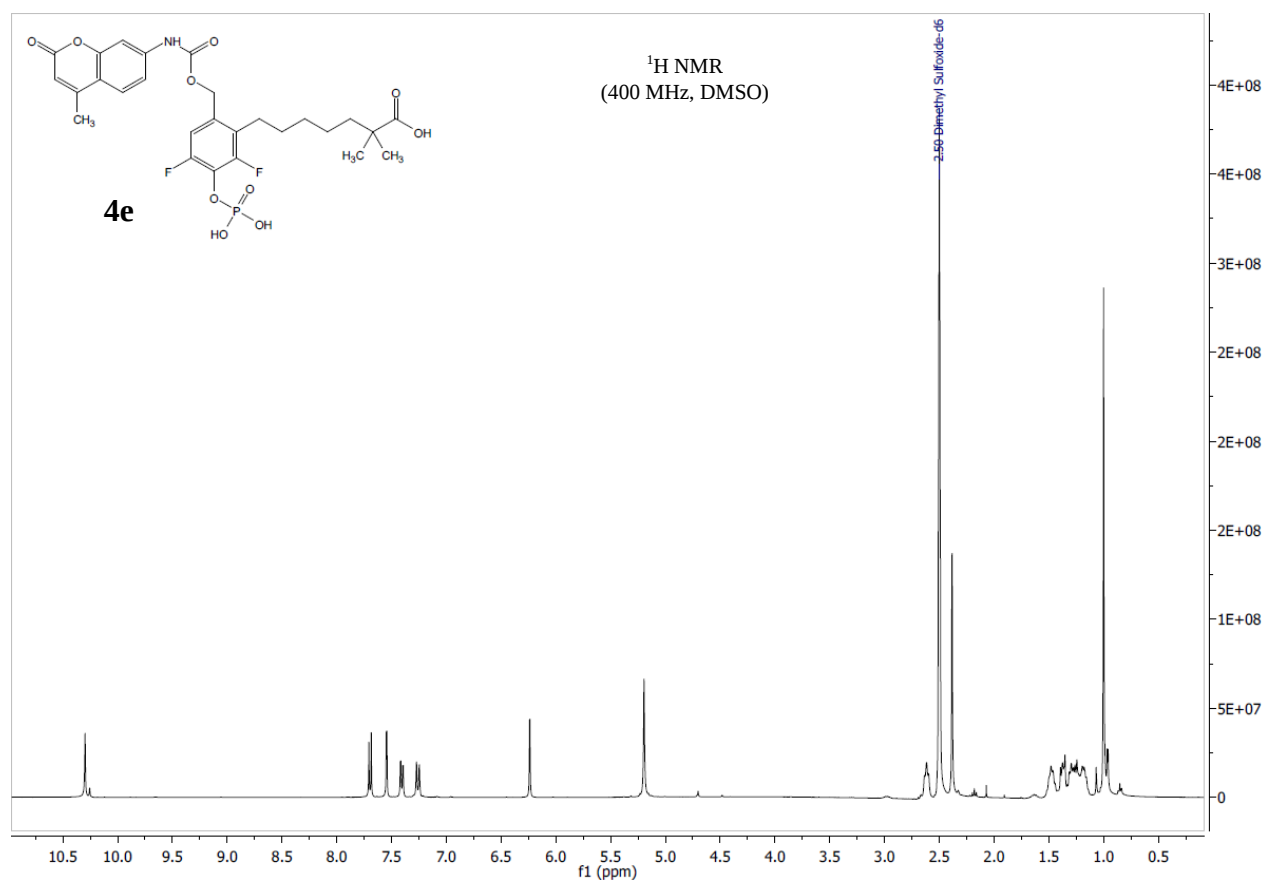
V. NMR spectra

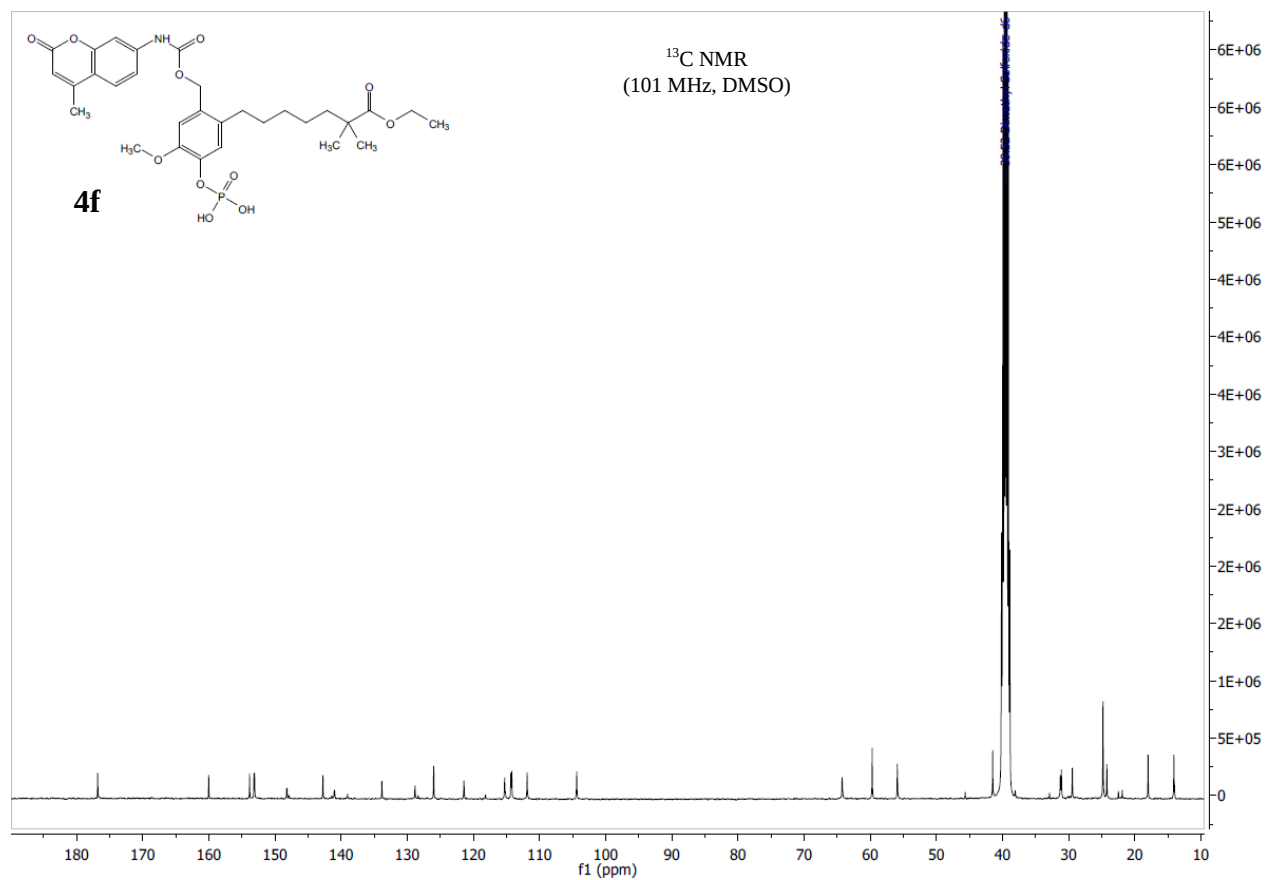
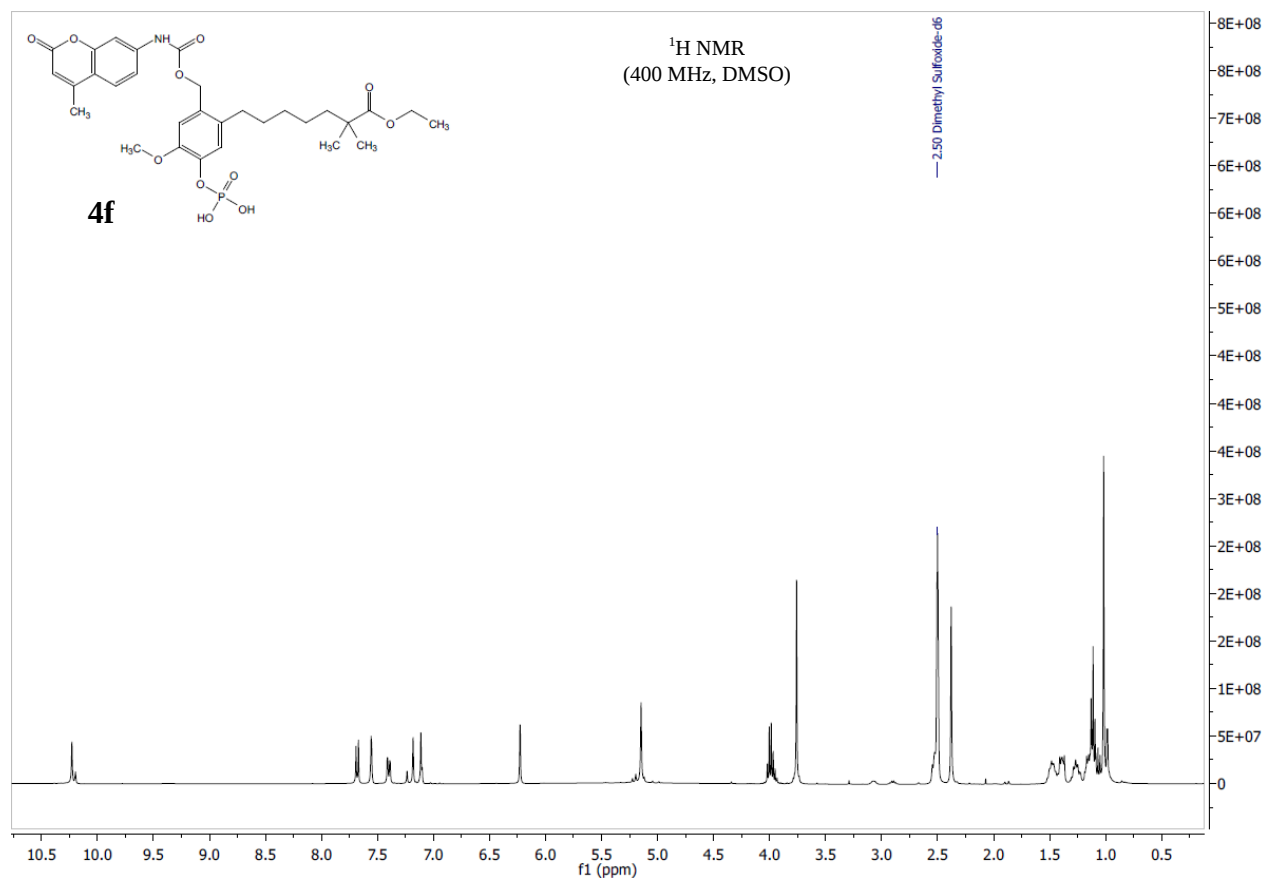


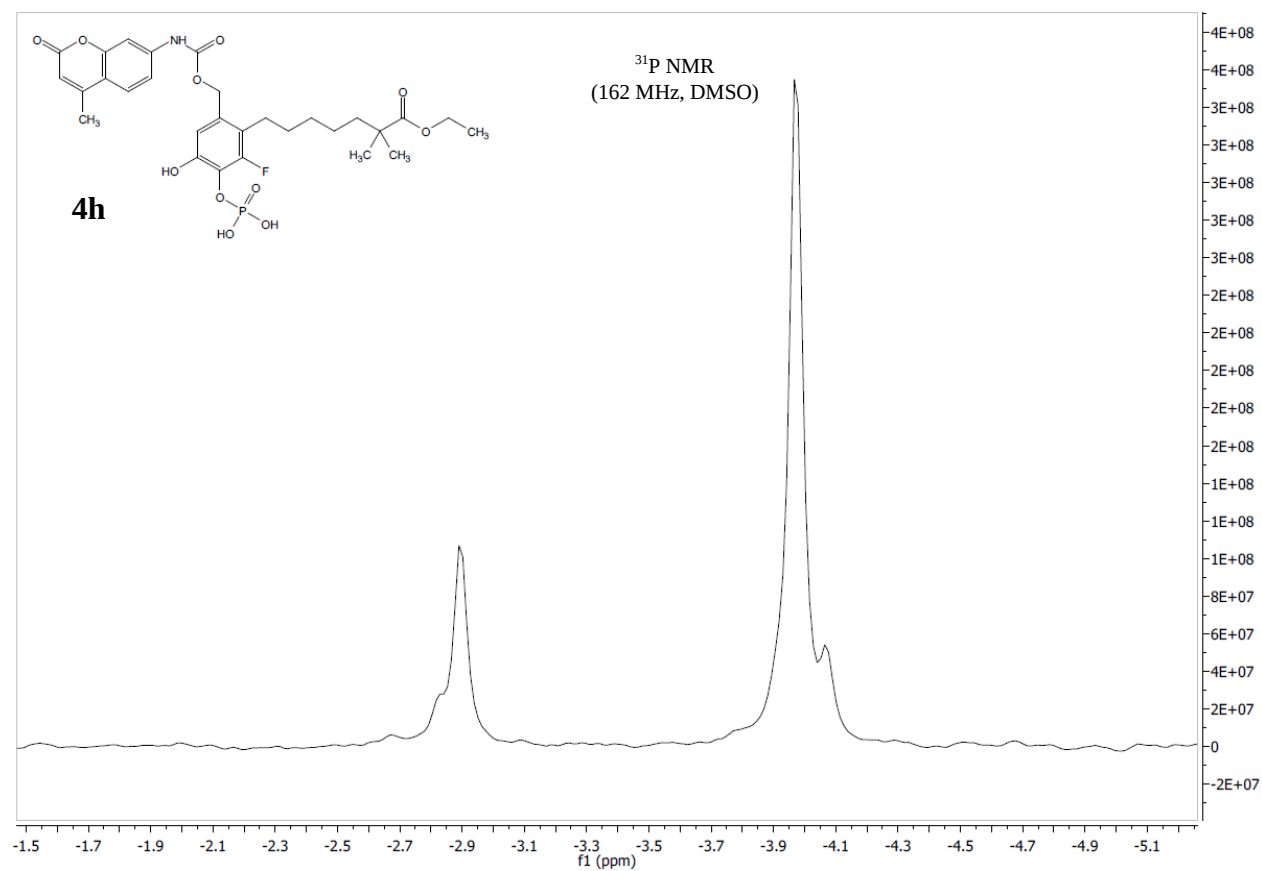
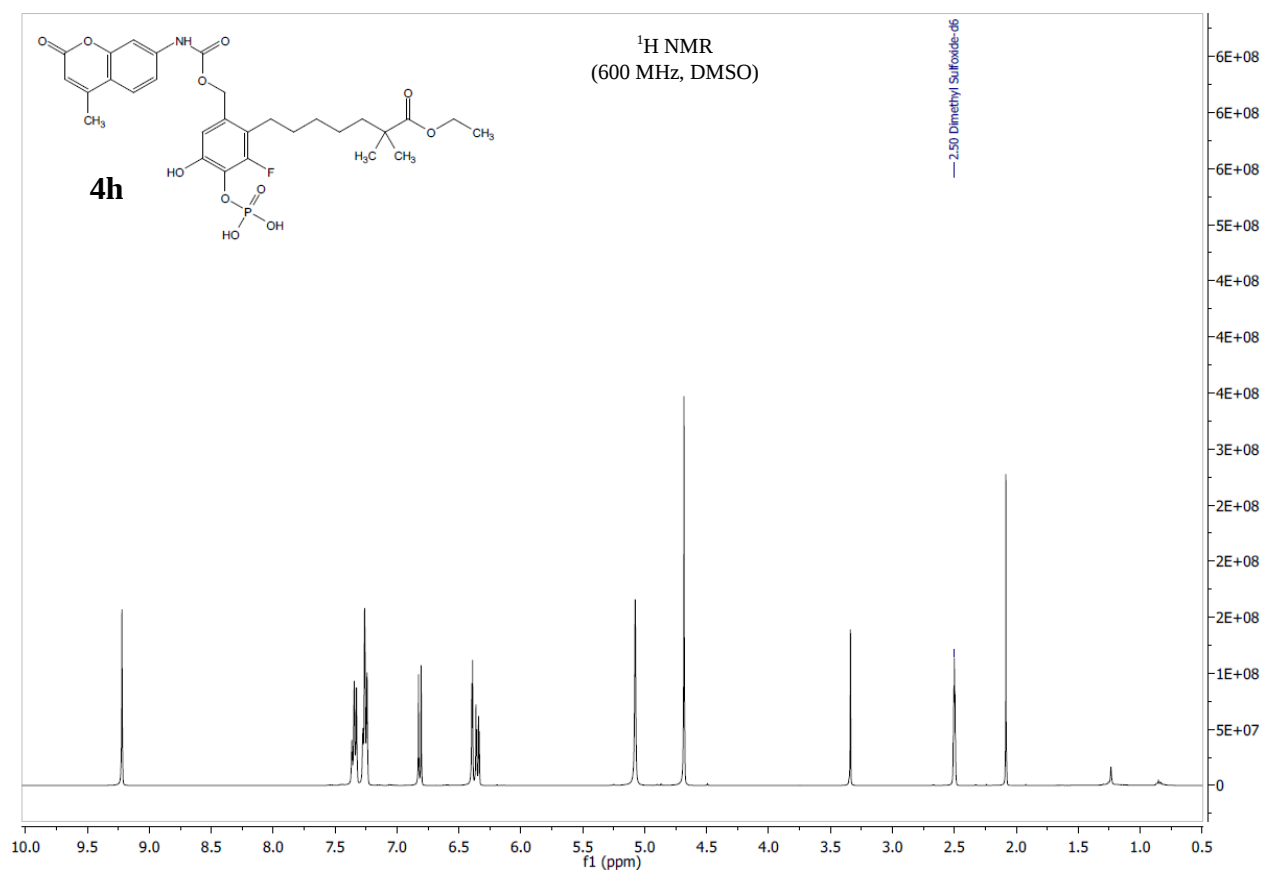


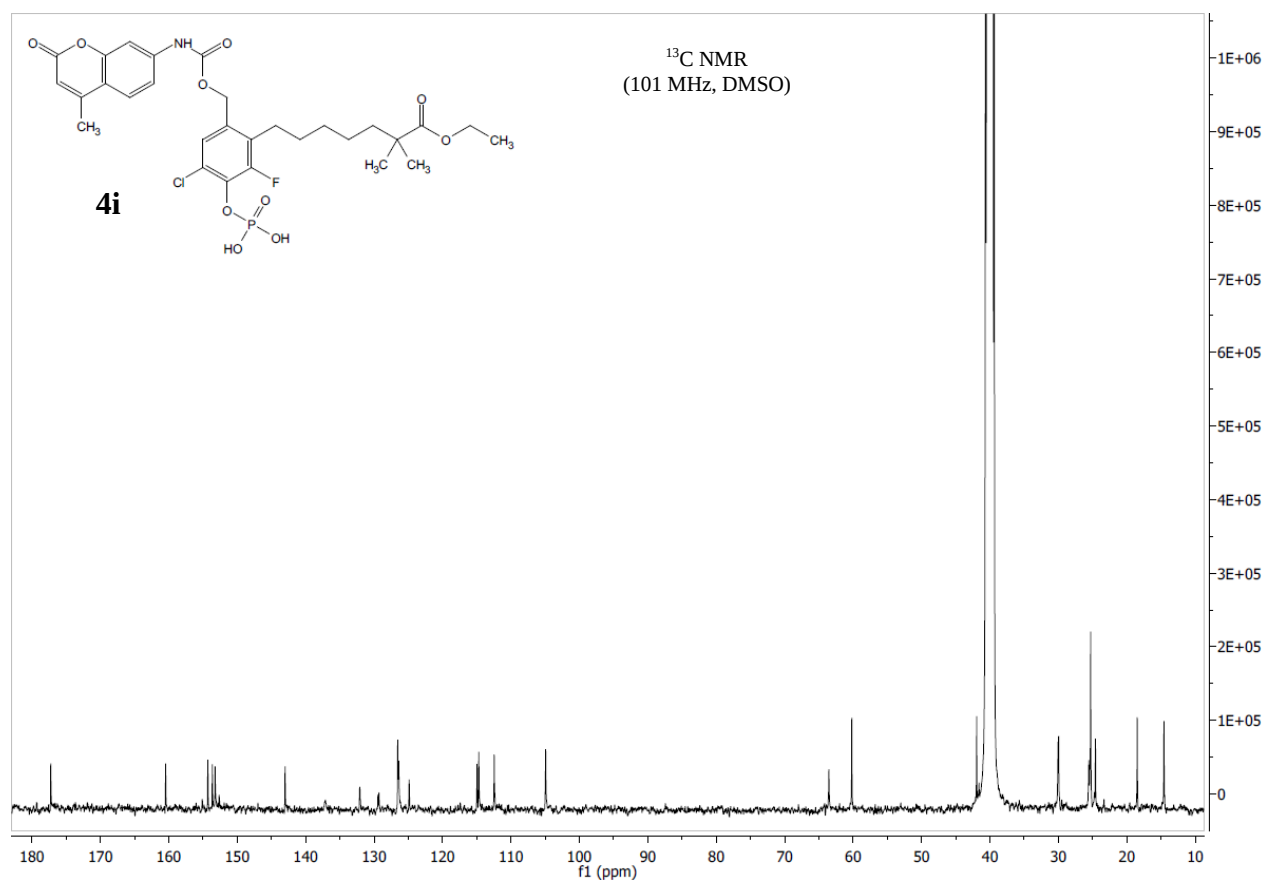
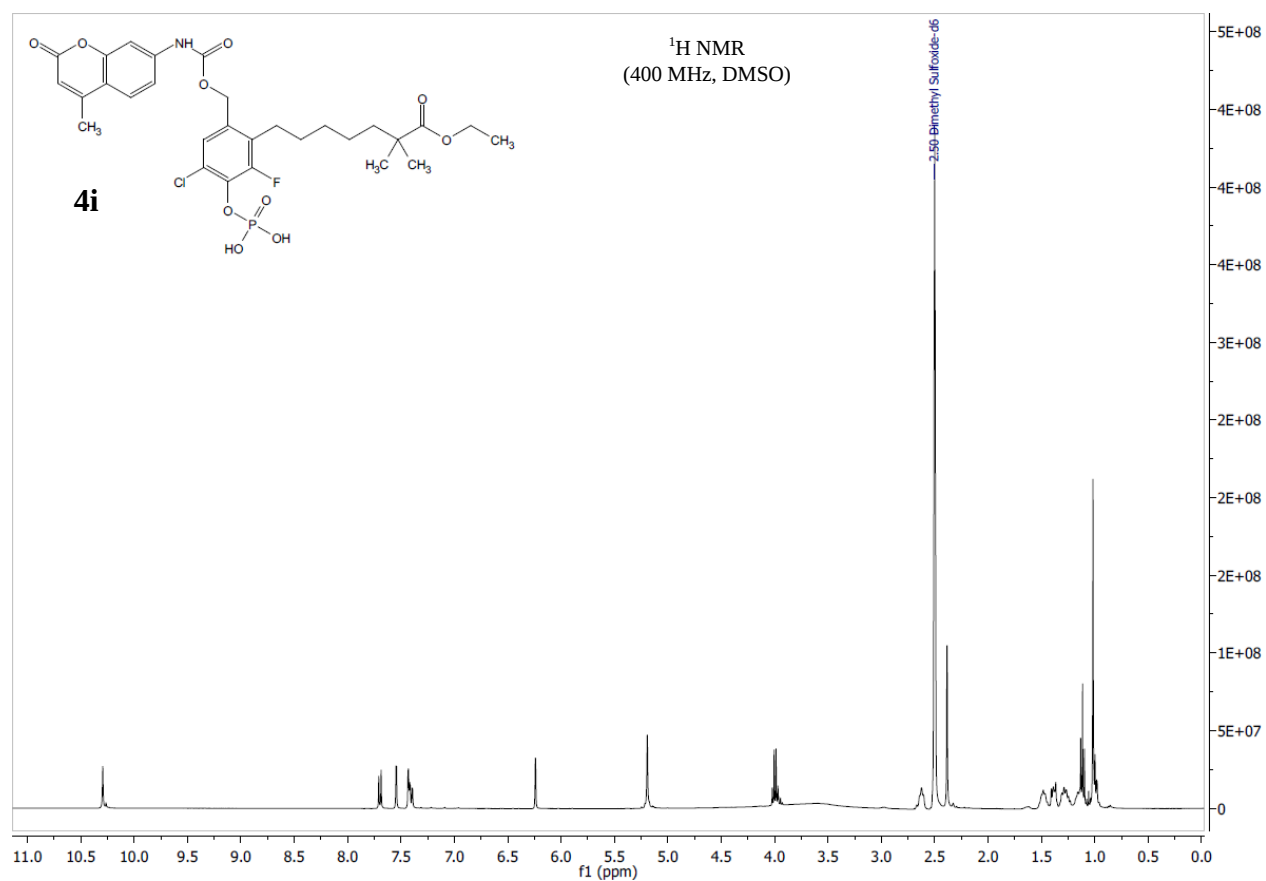


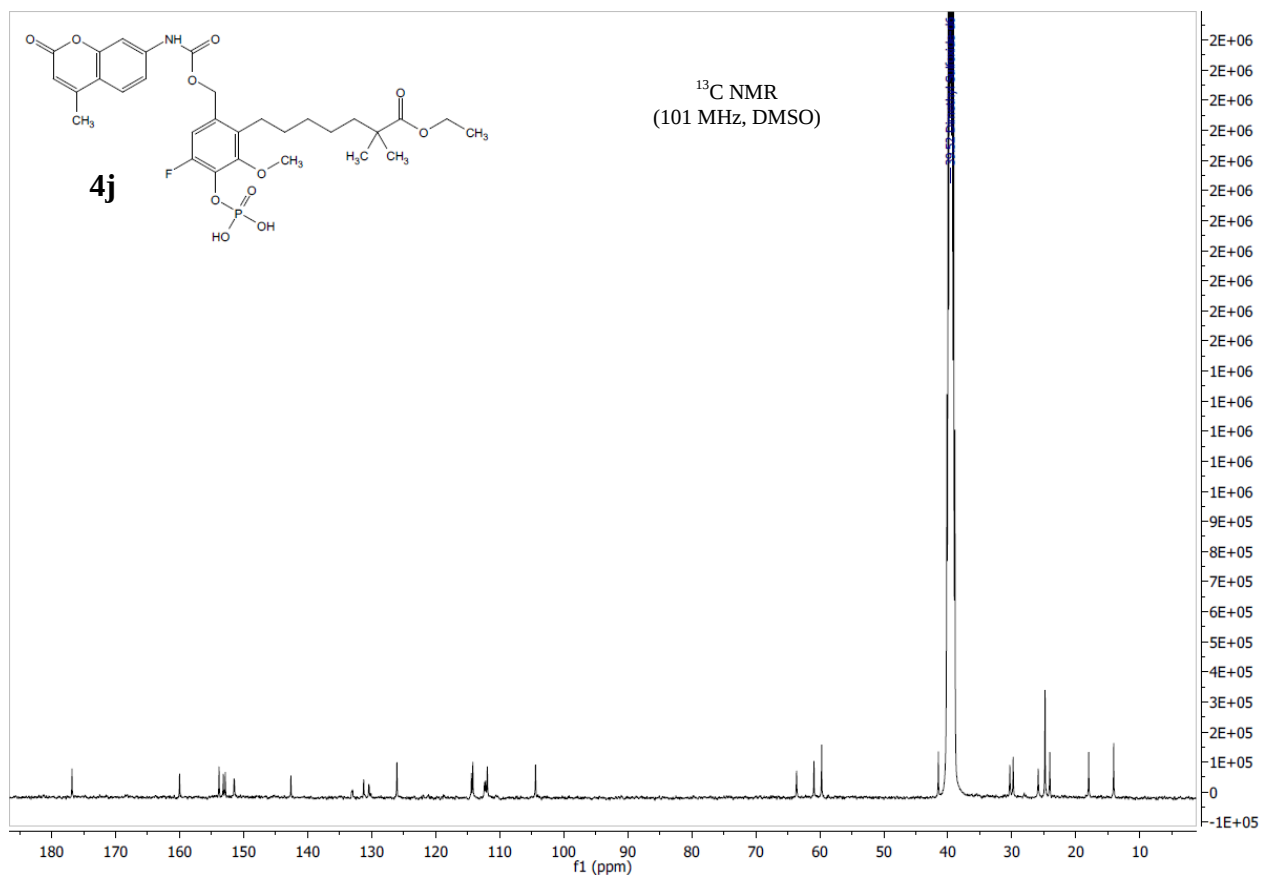
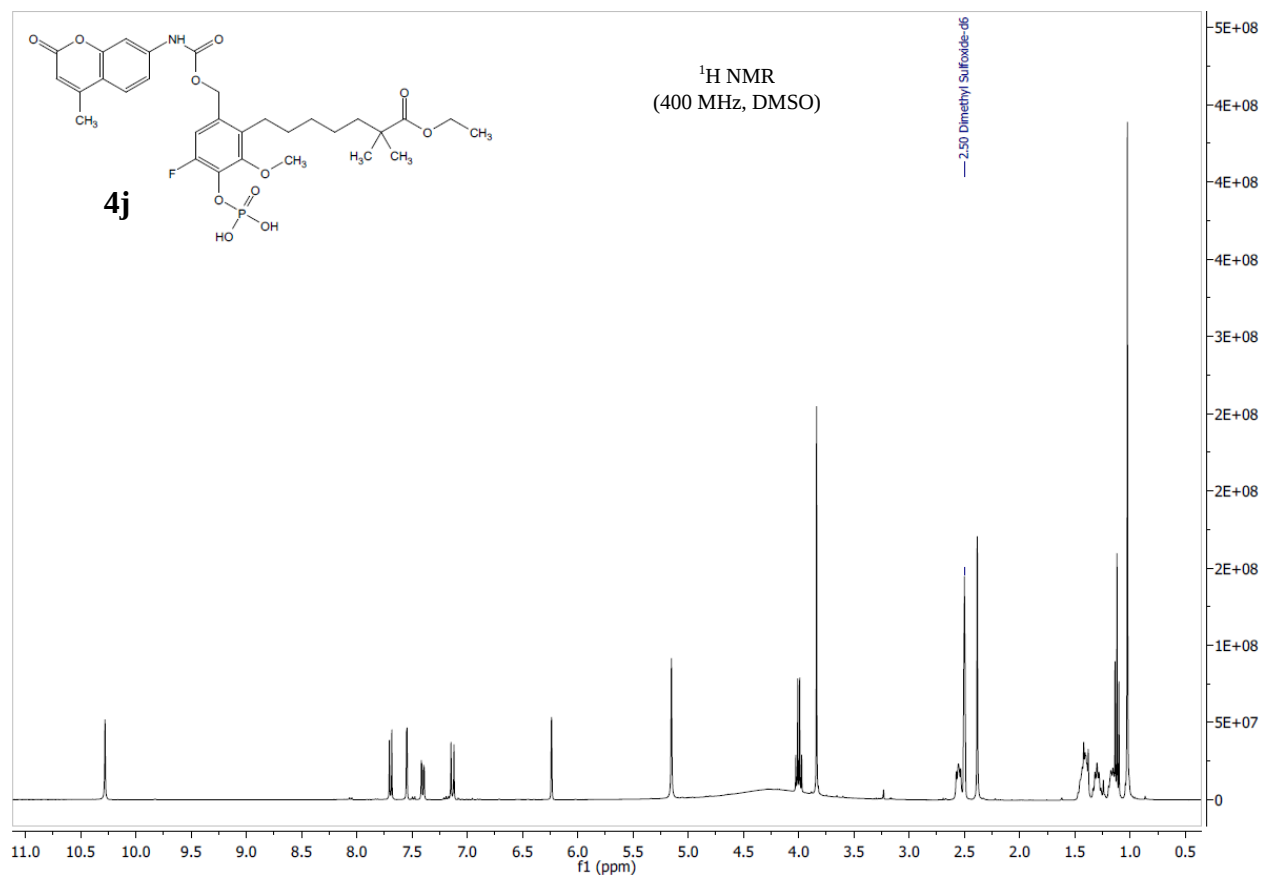


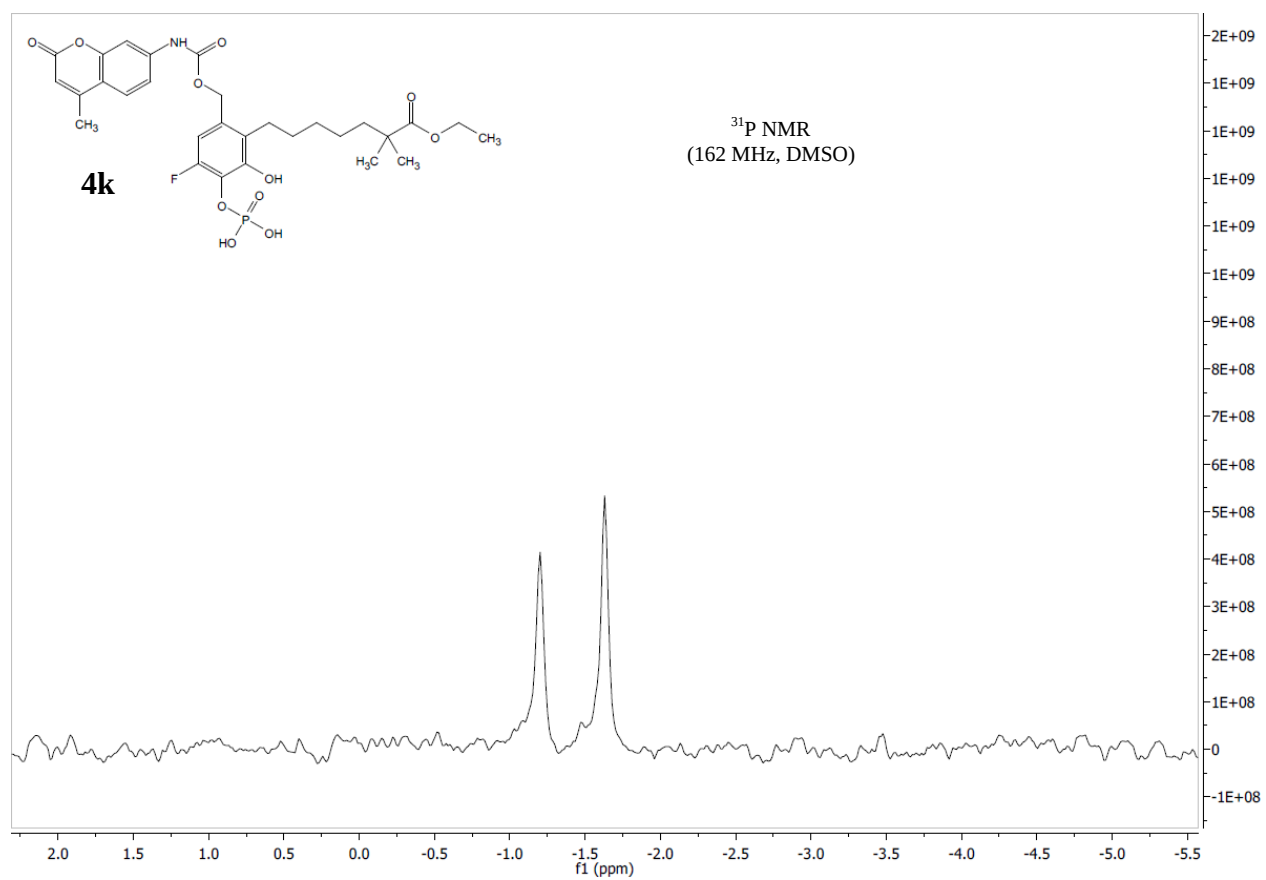
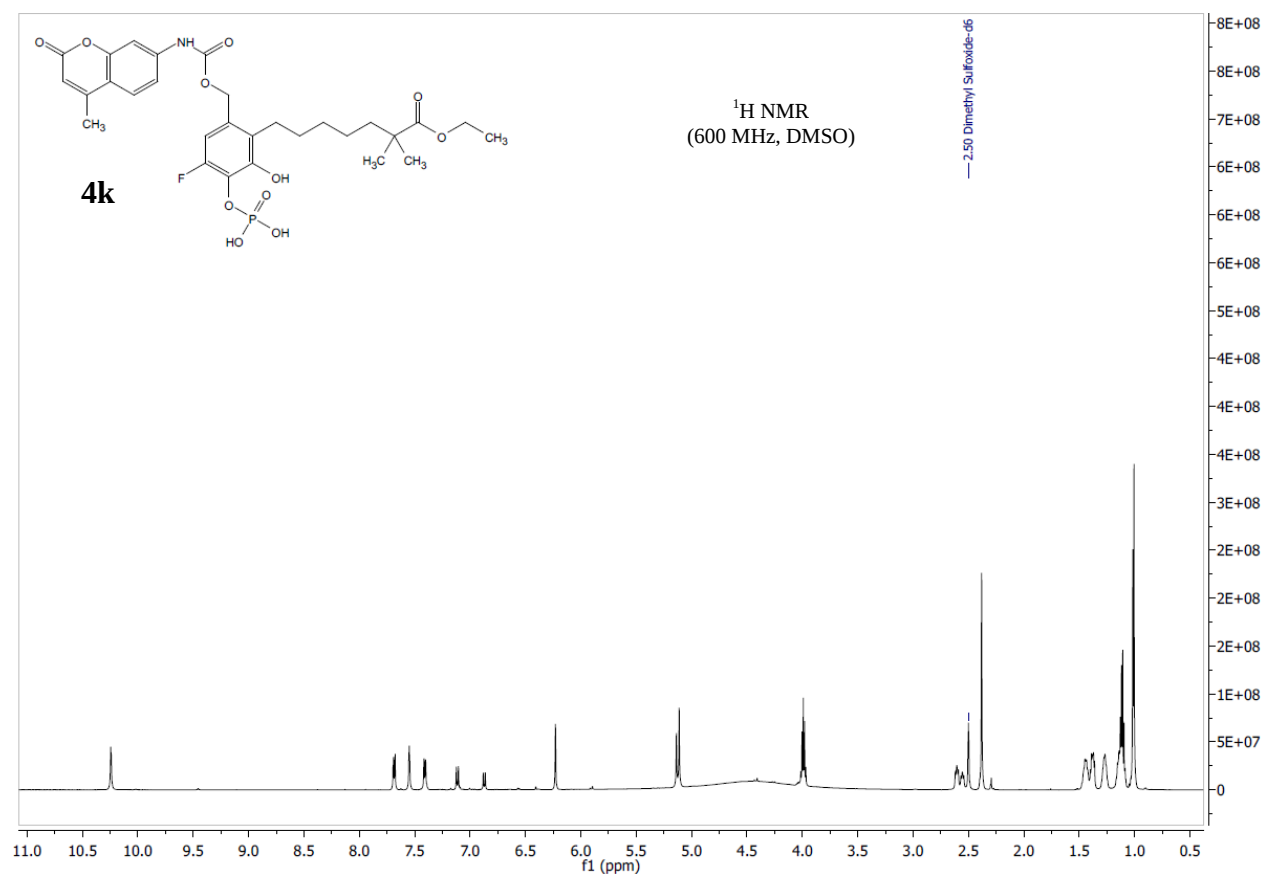


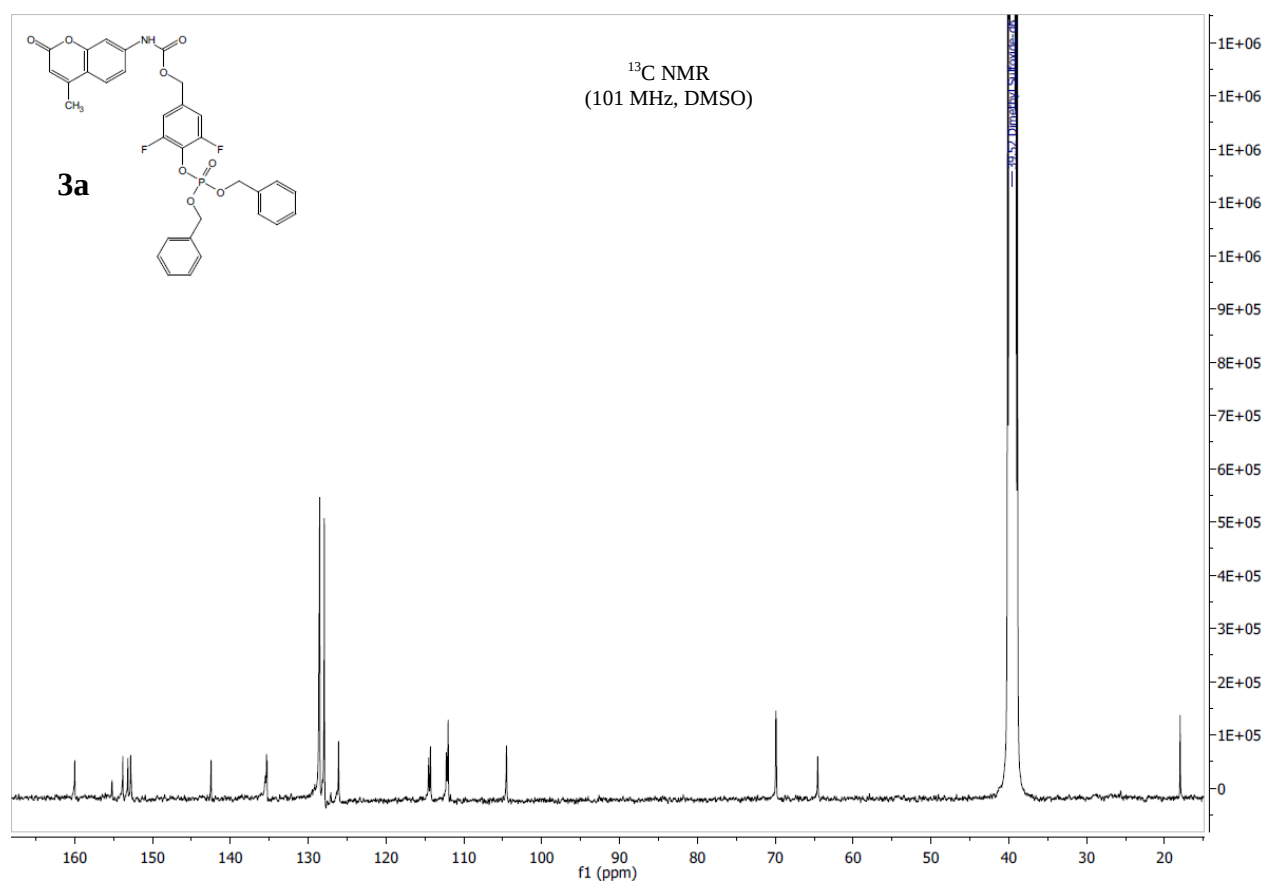
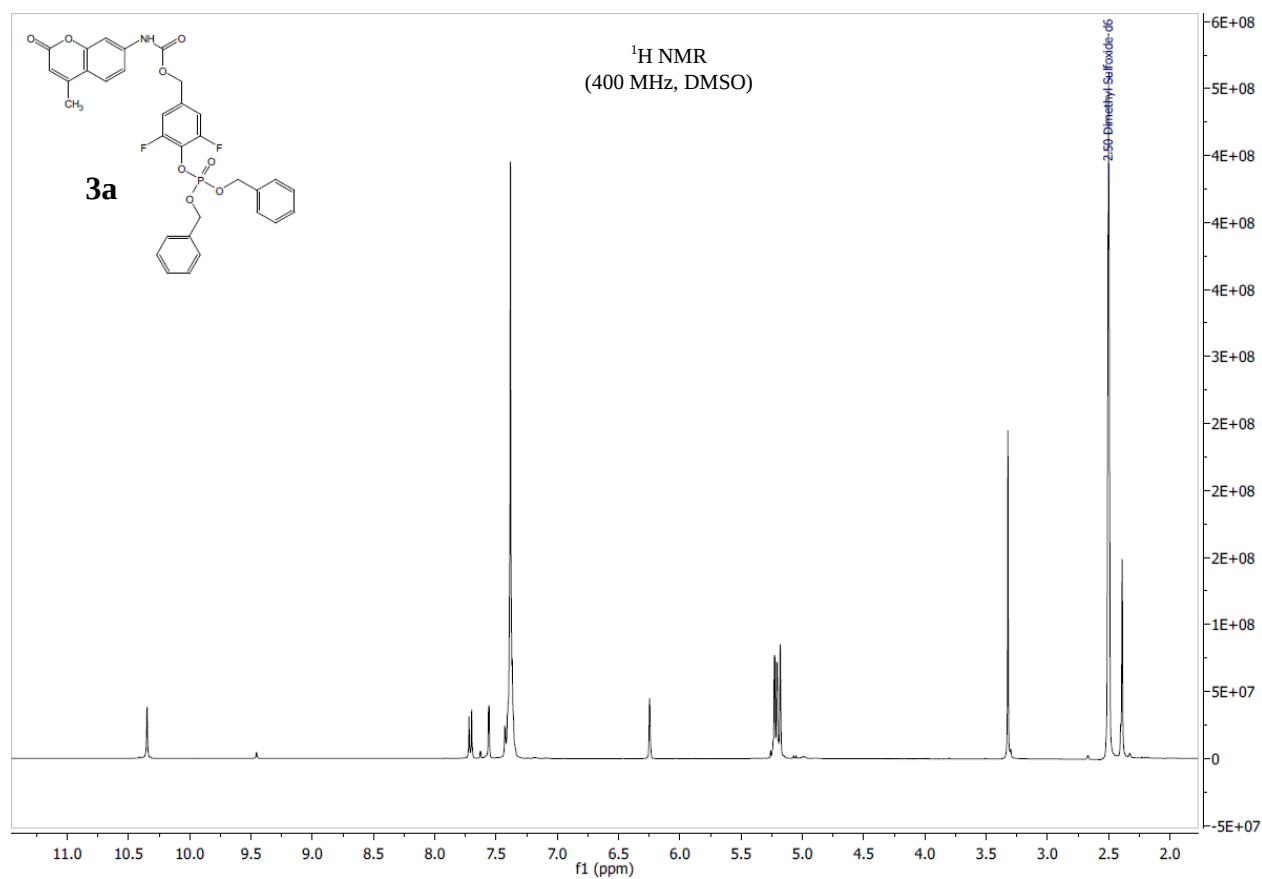


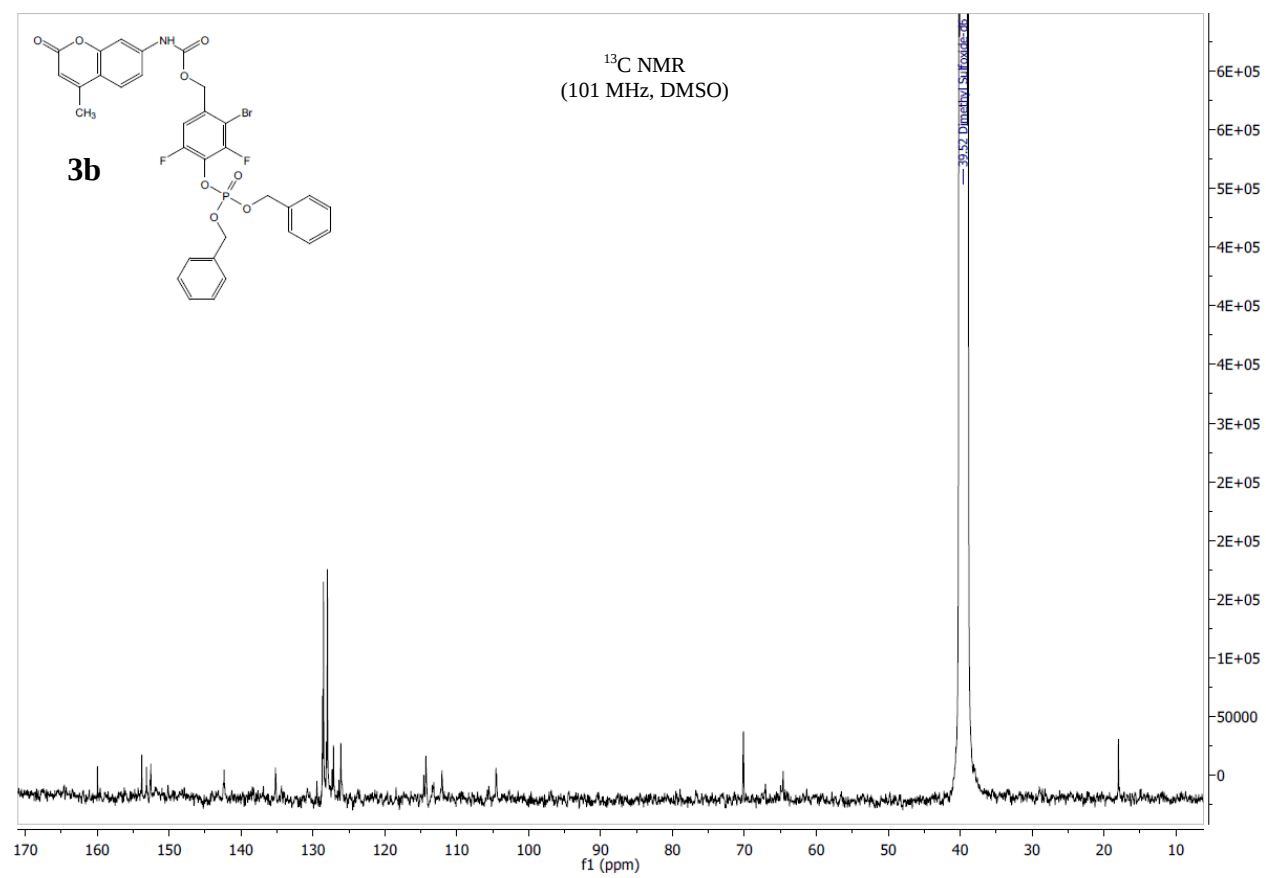
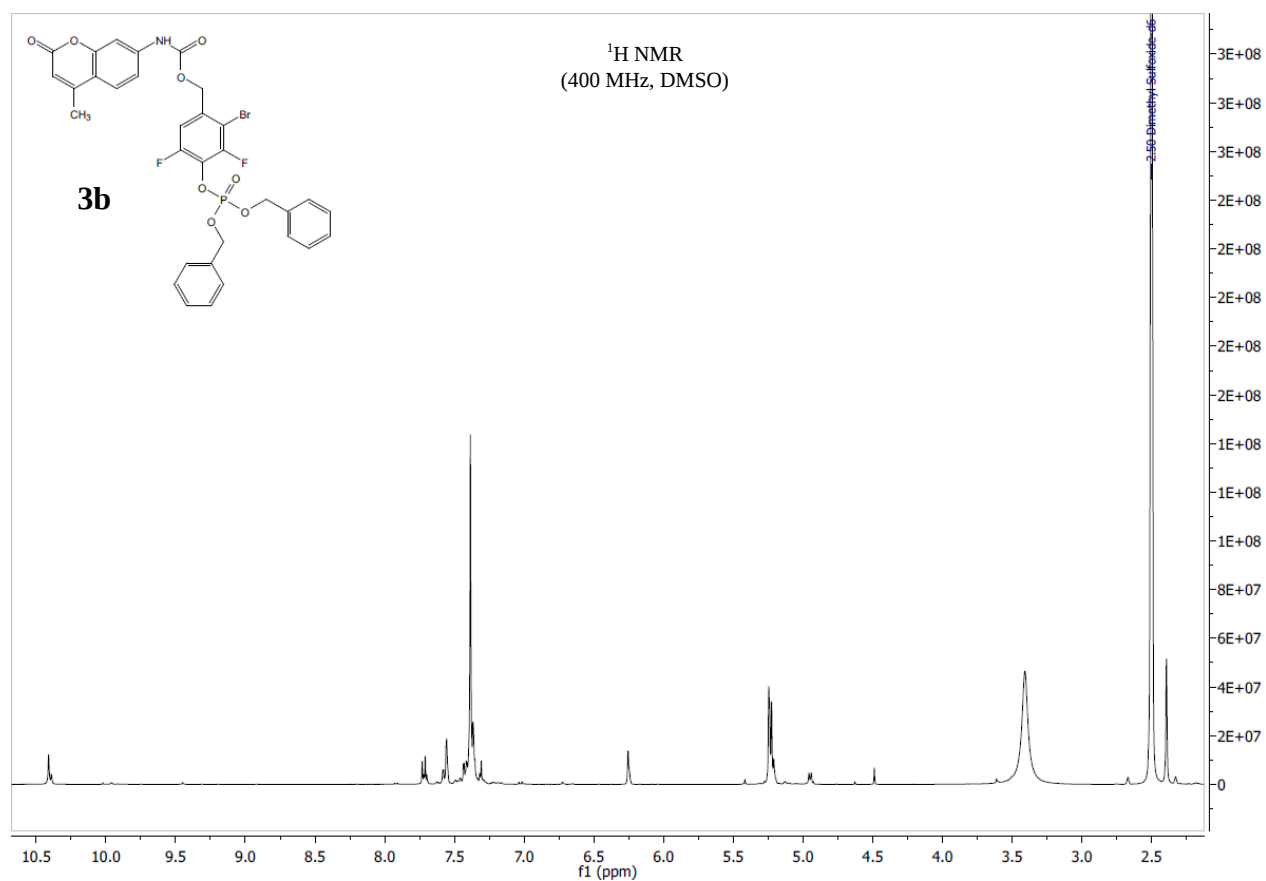


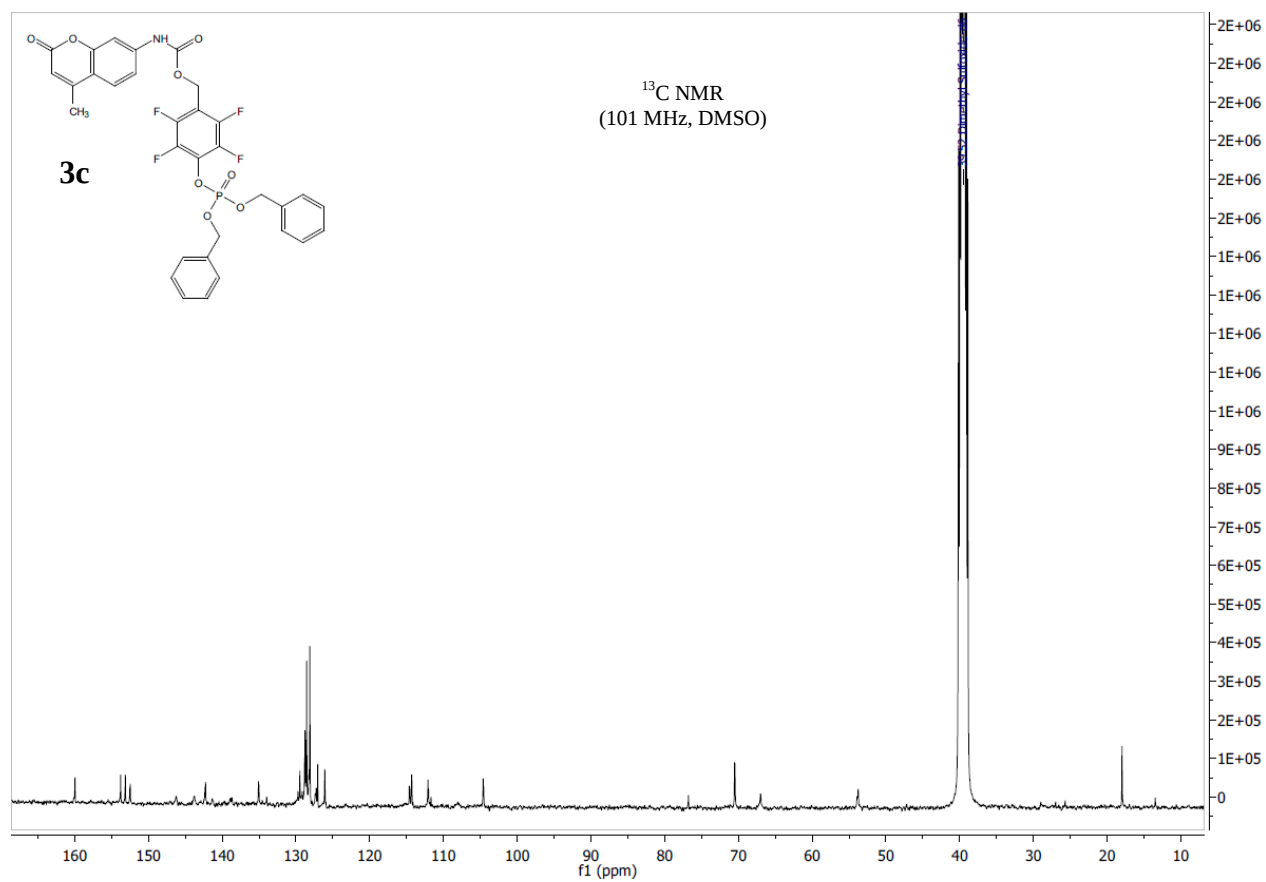
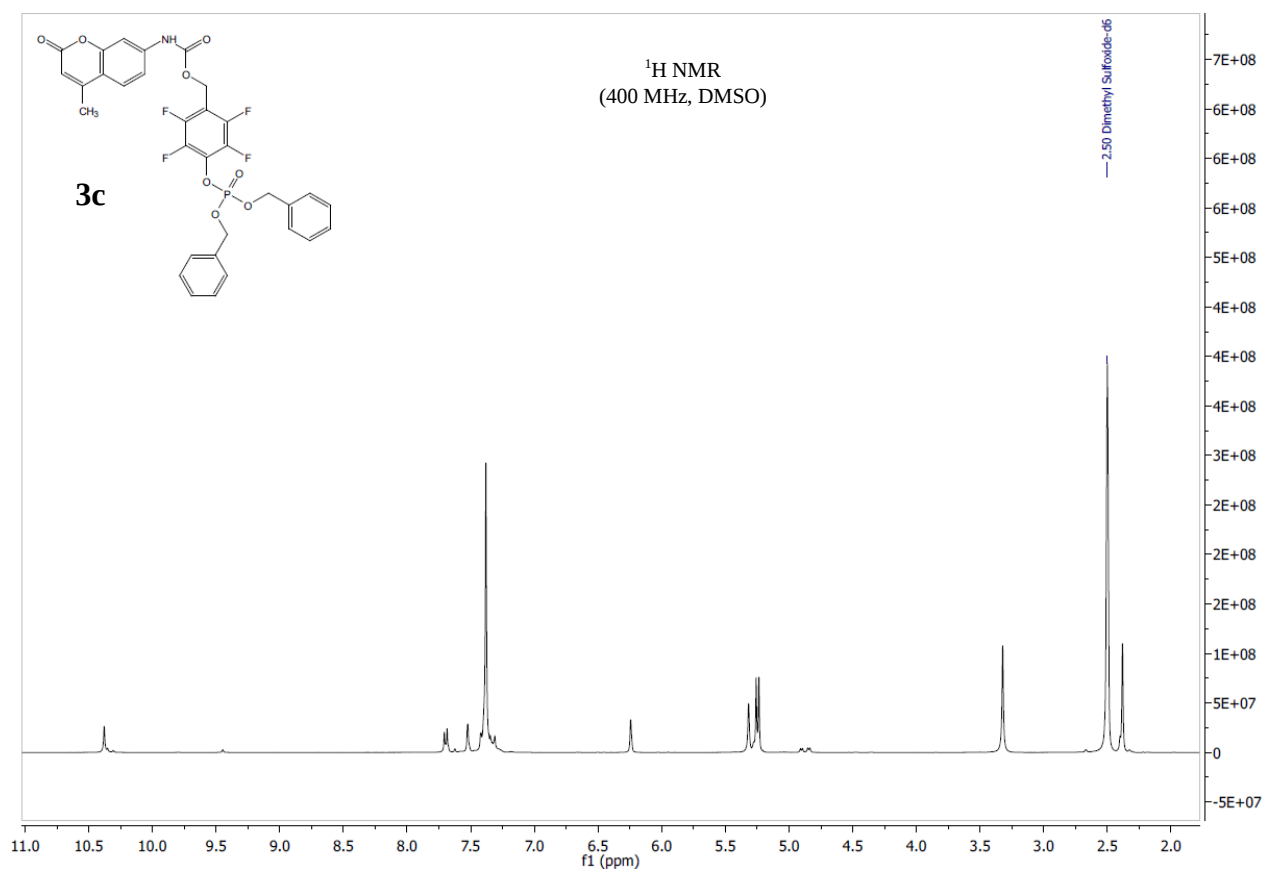


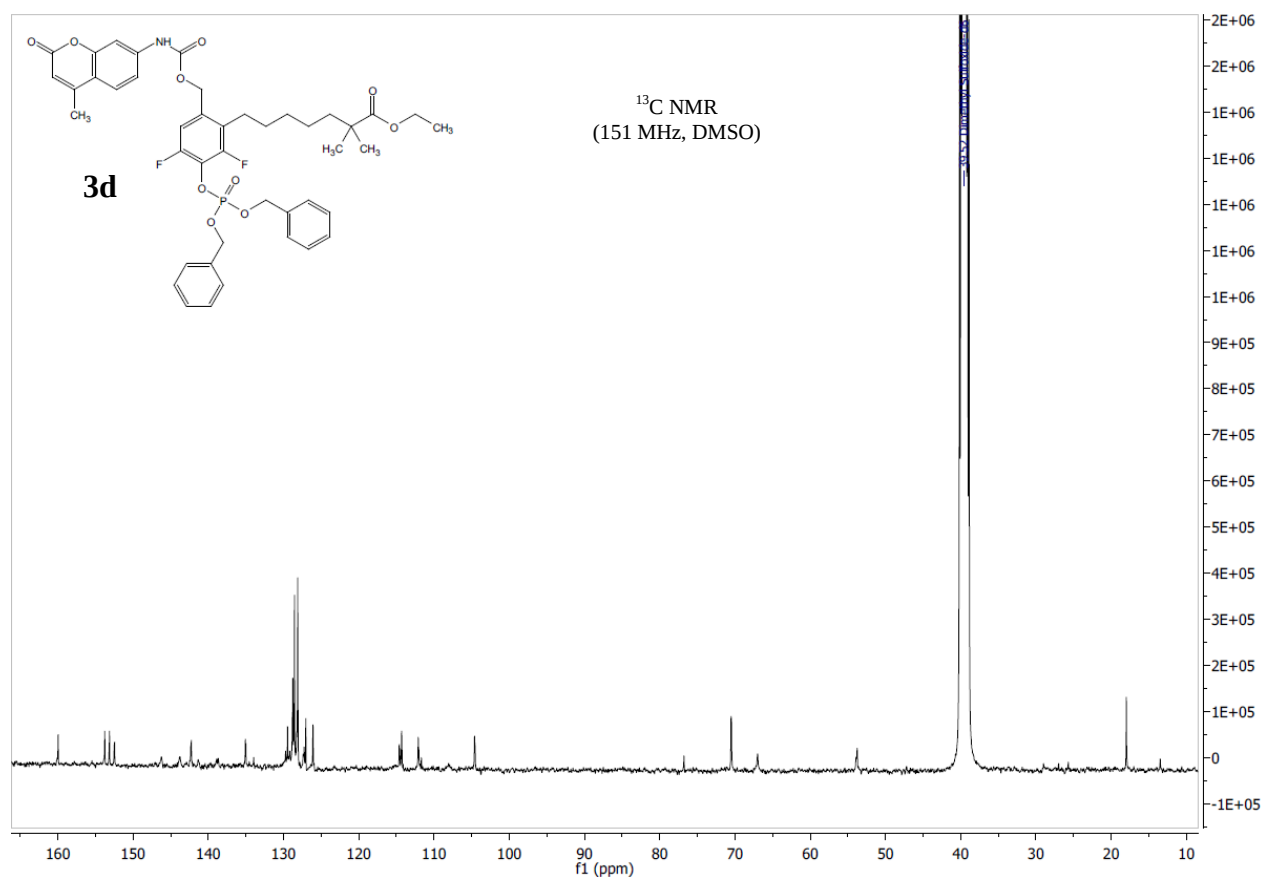
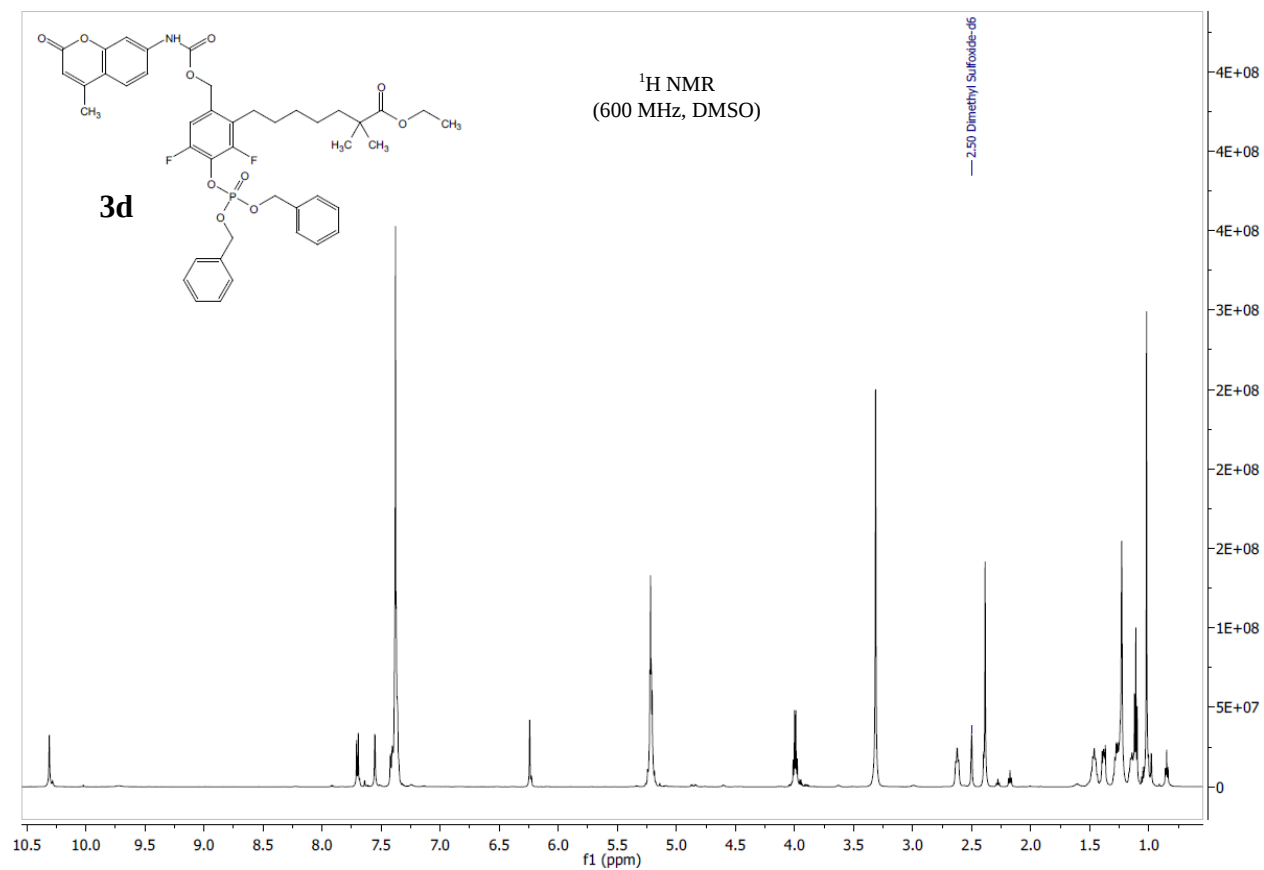


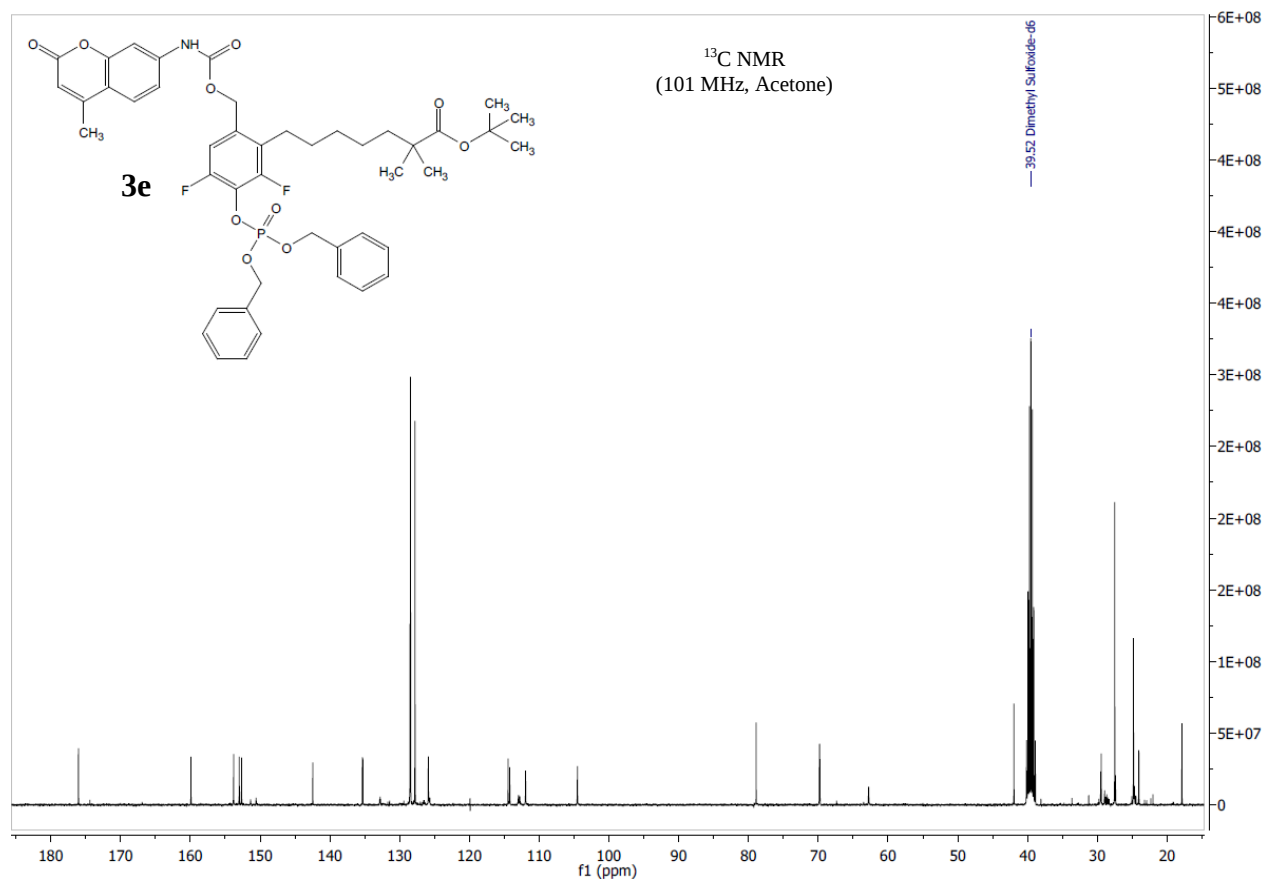
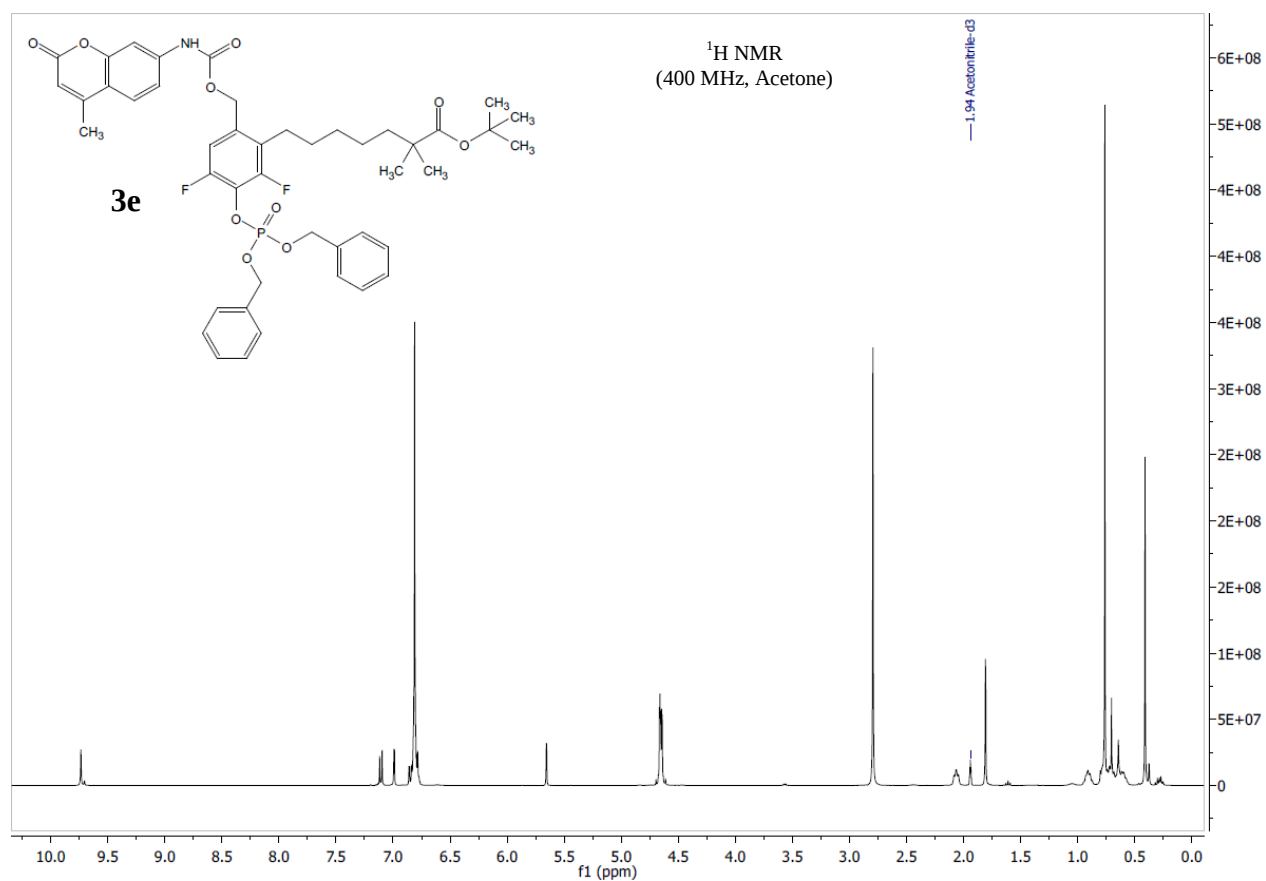


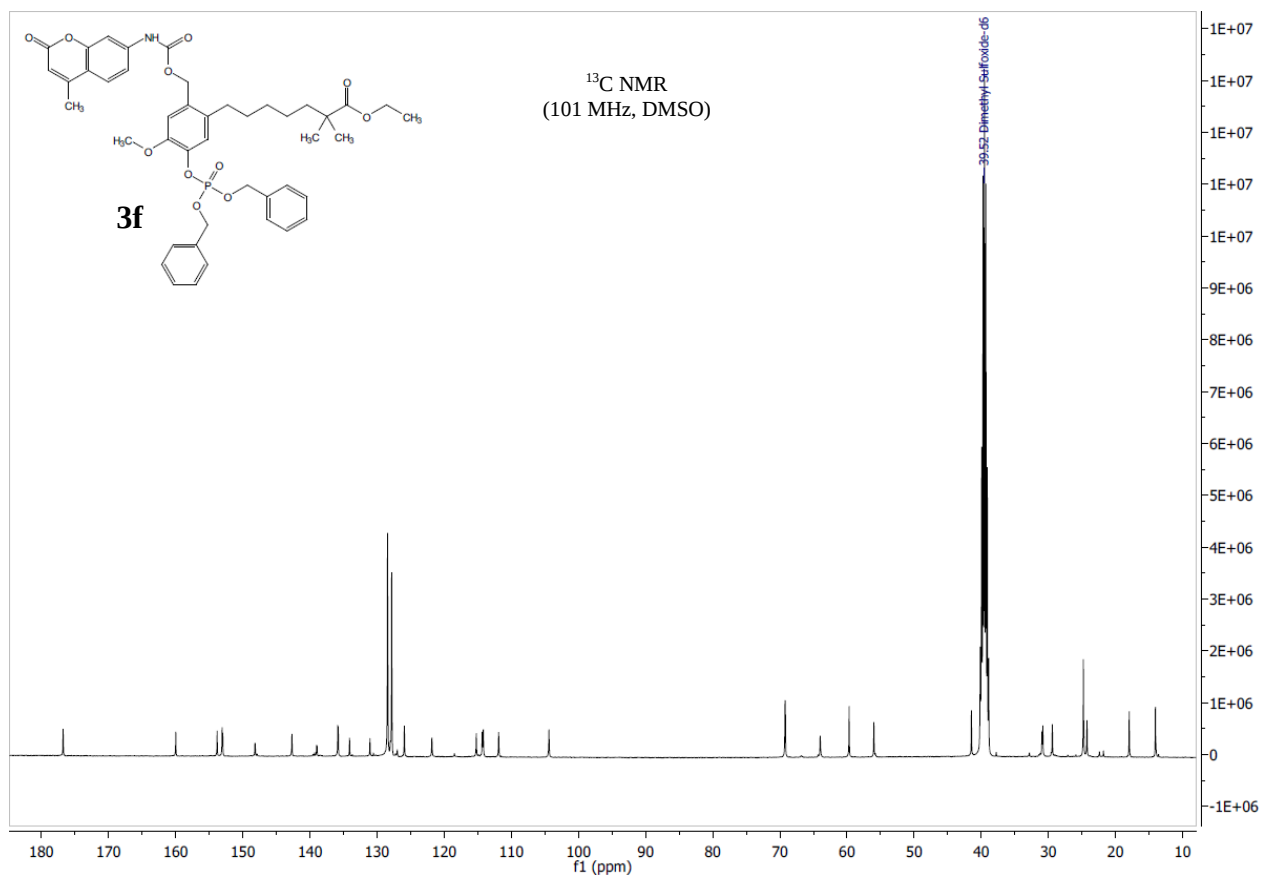
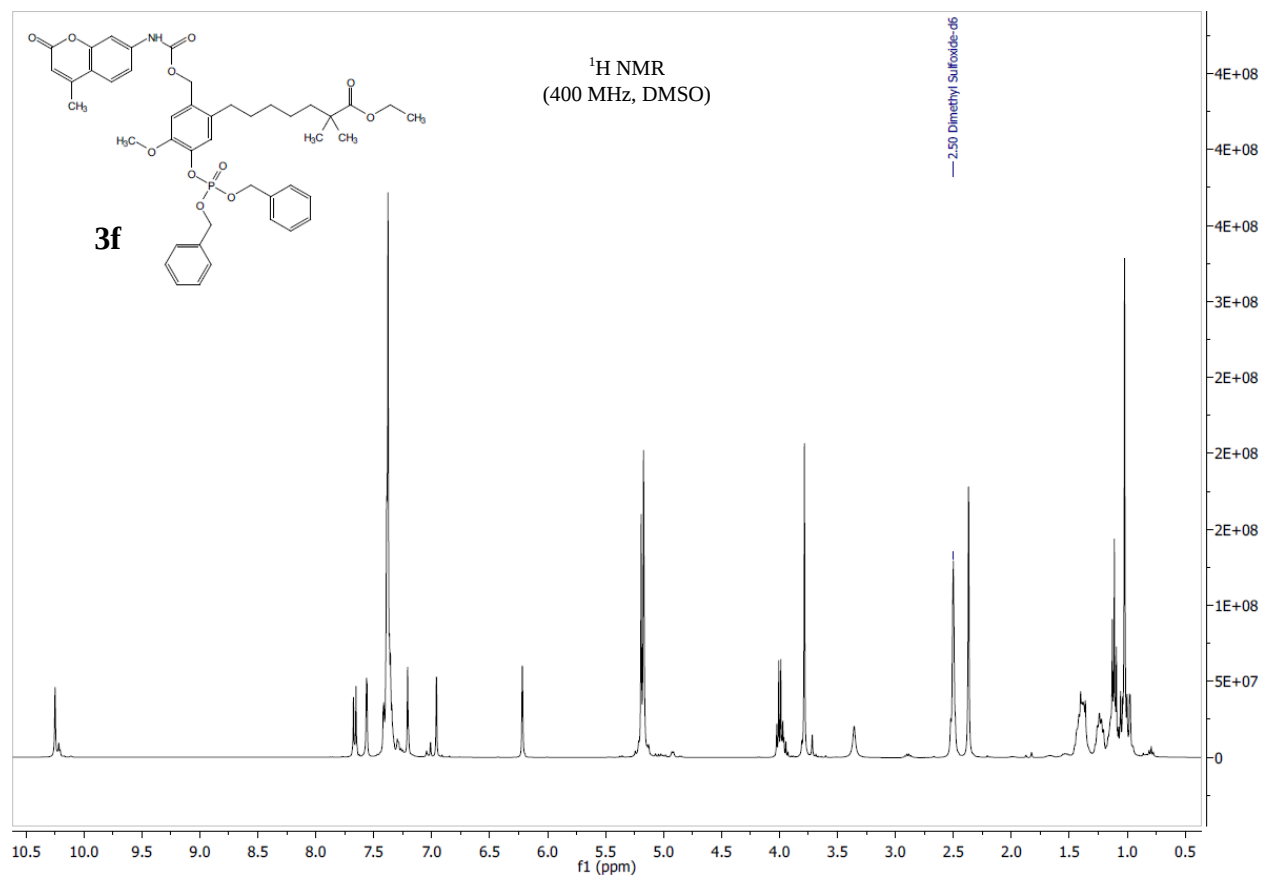


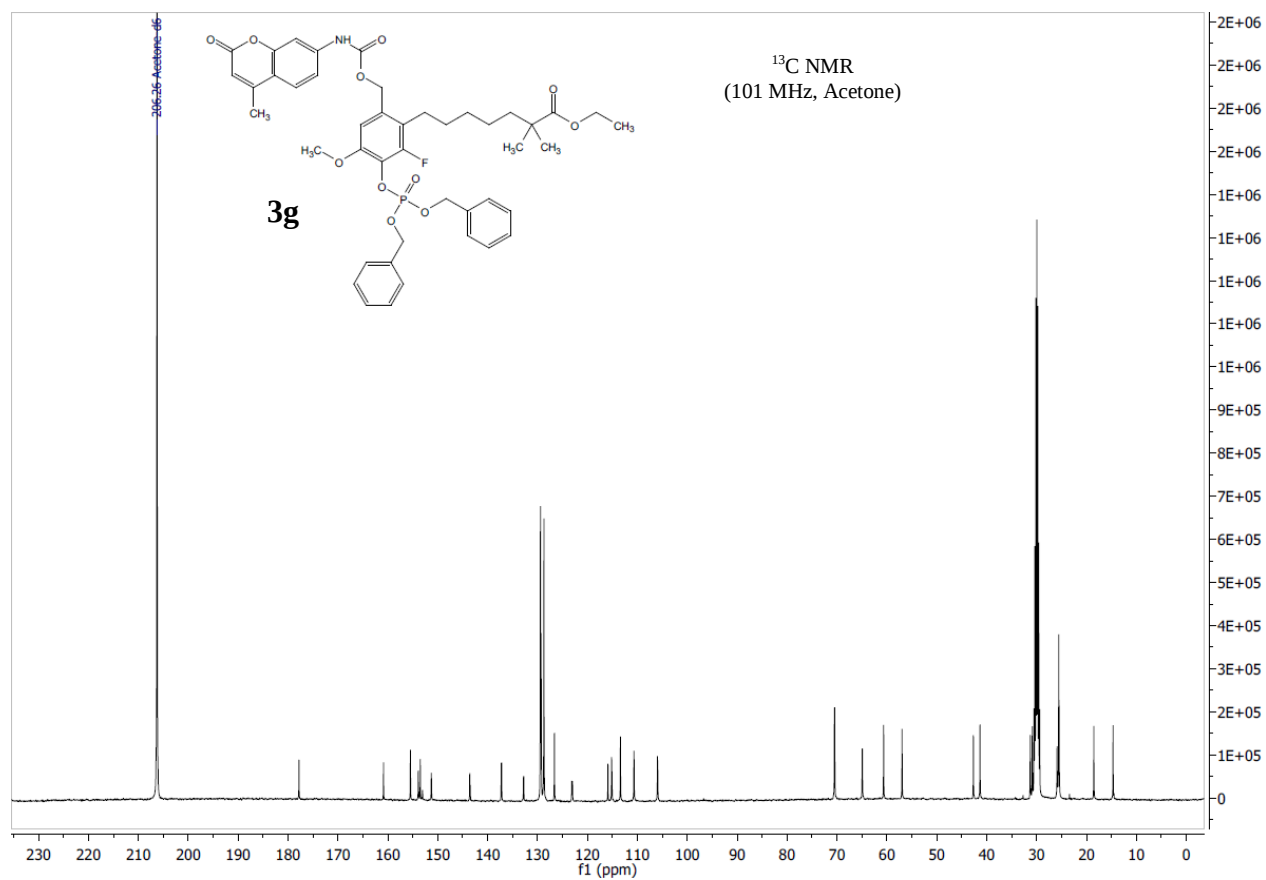
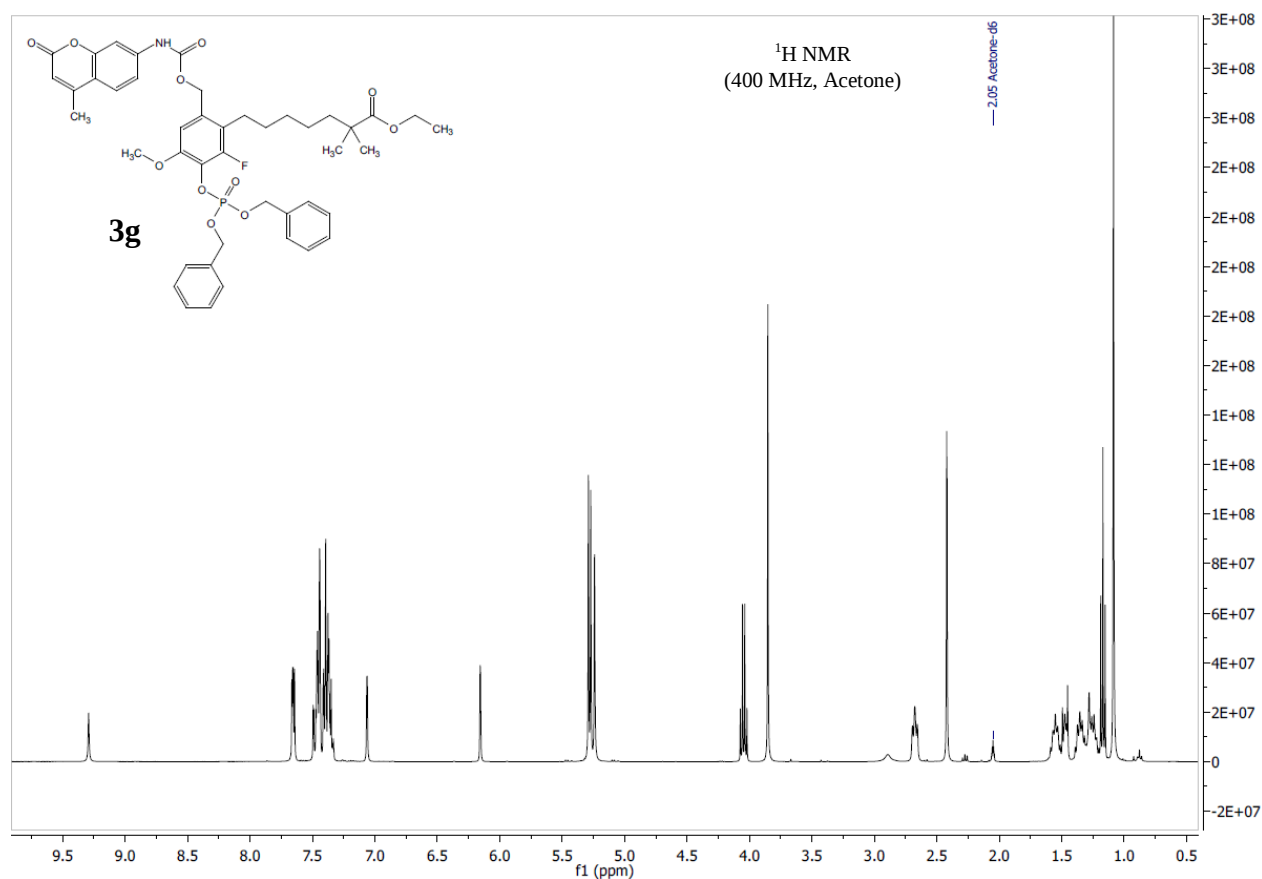


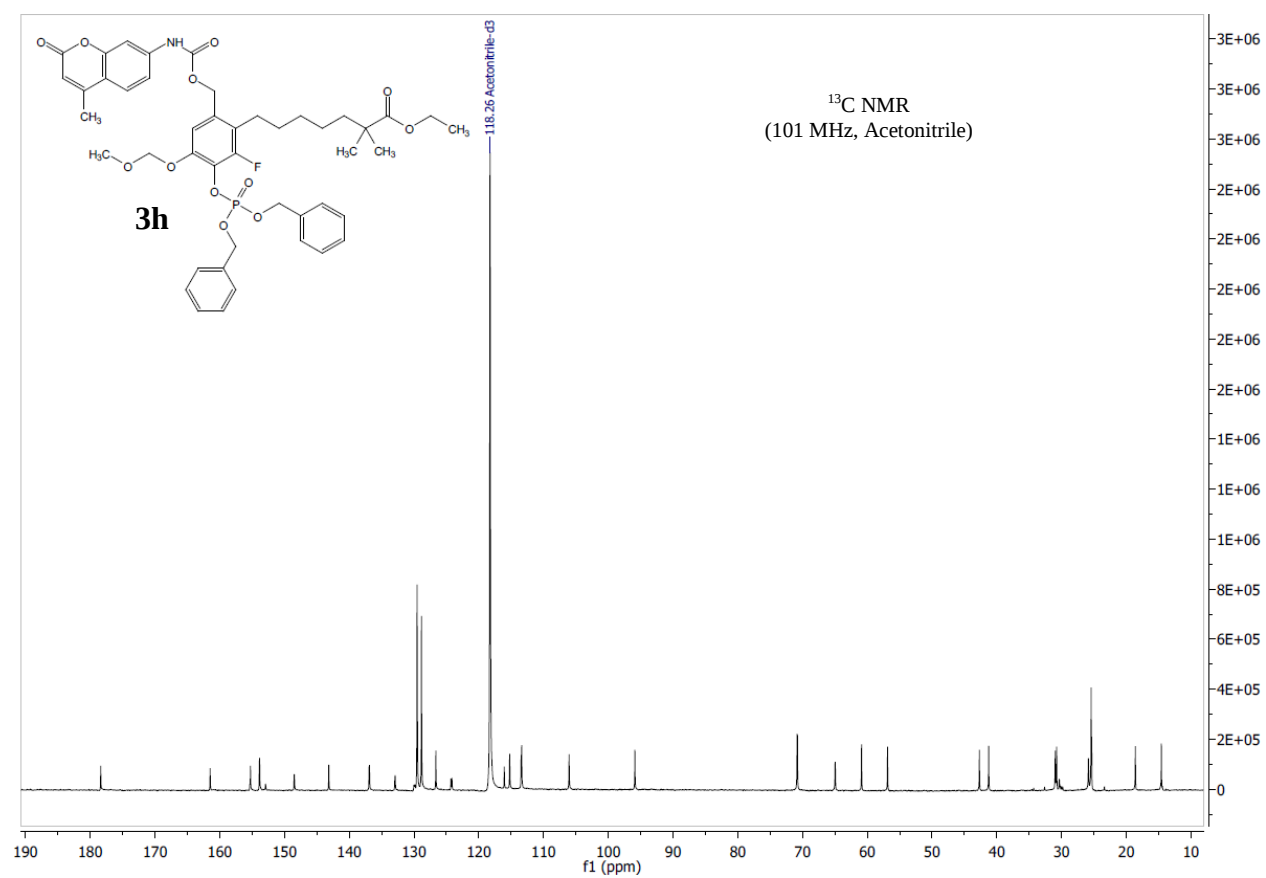
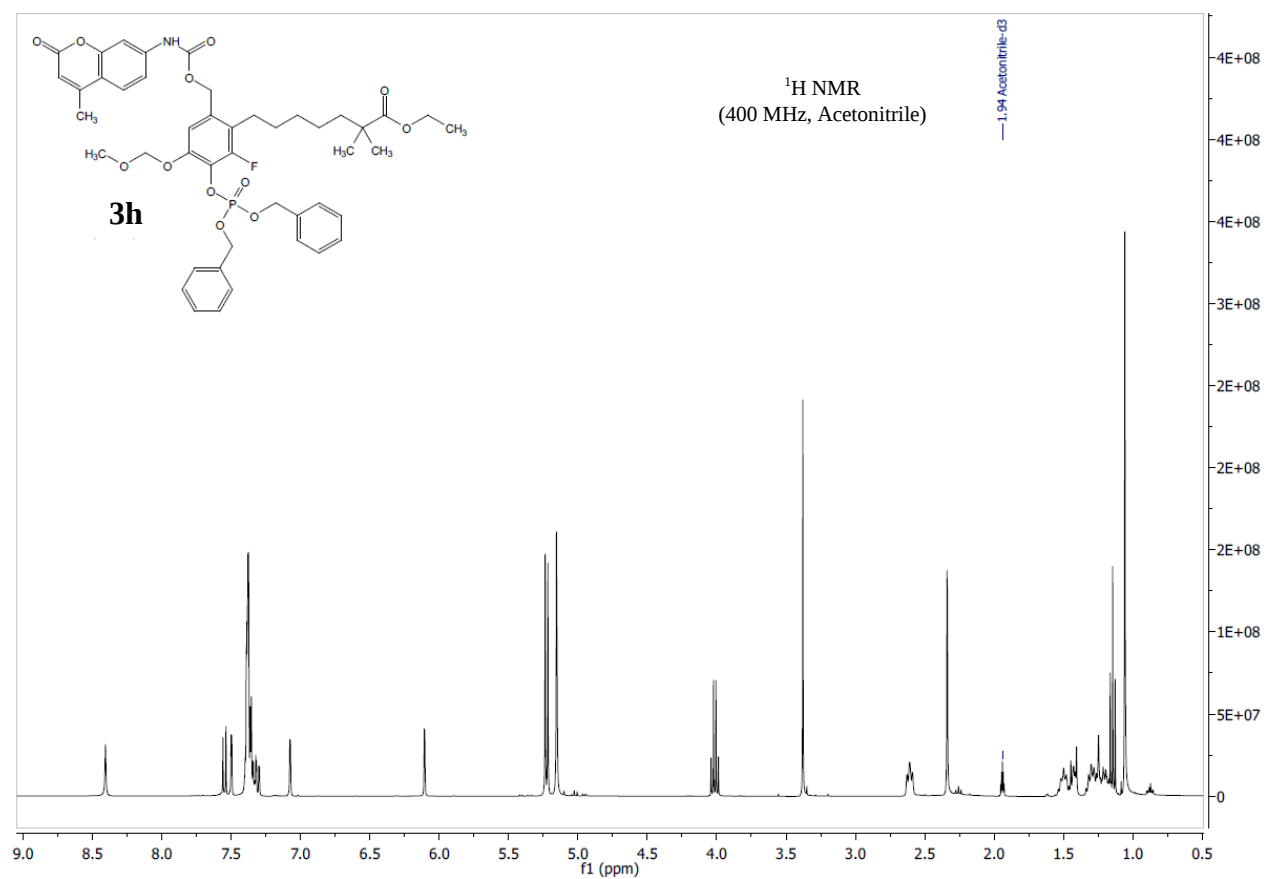


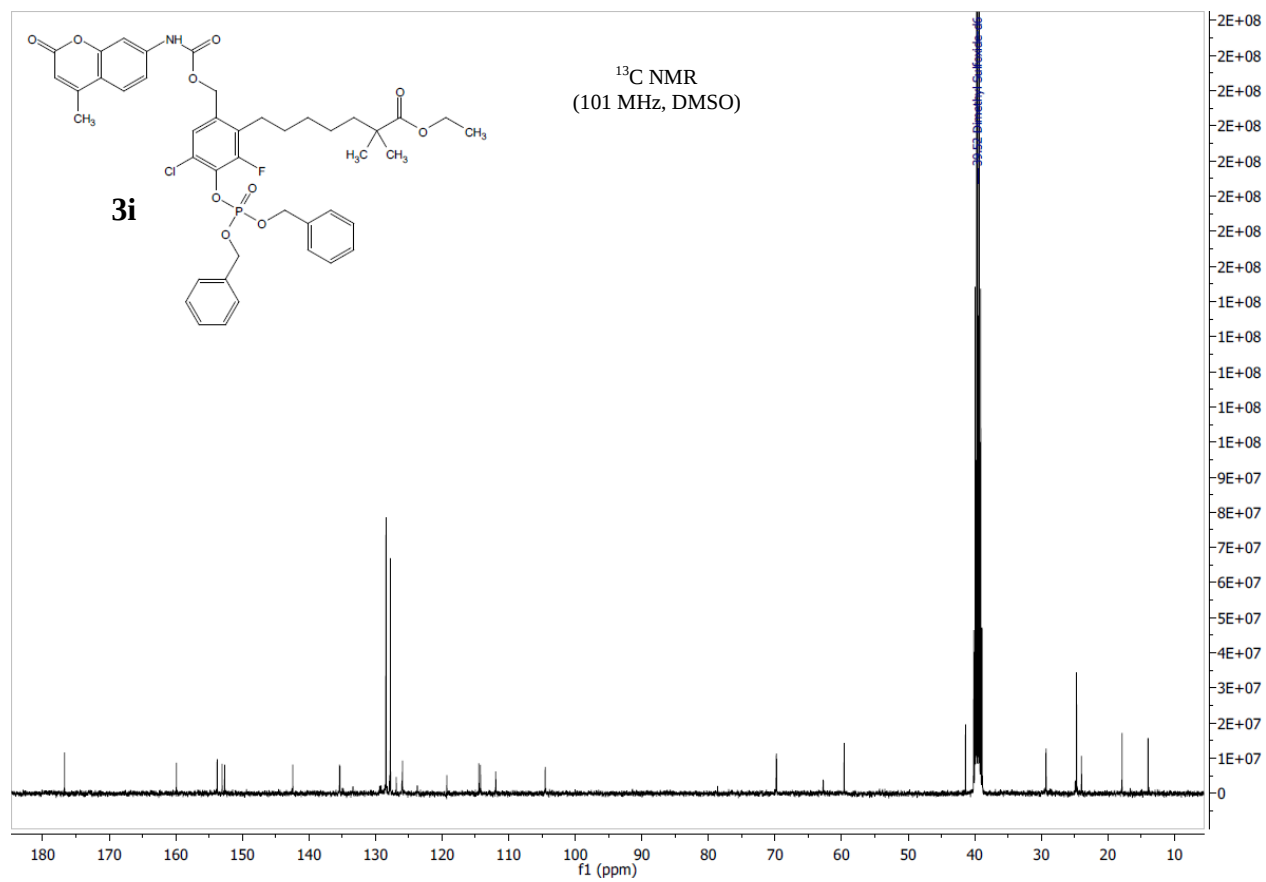
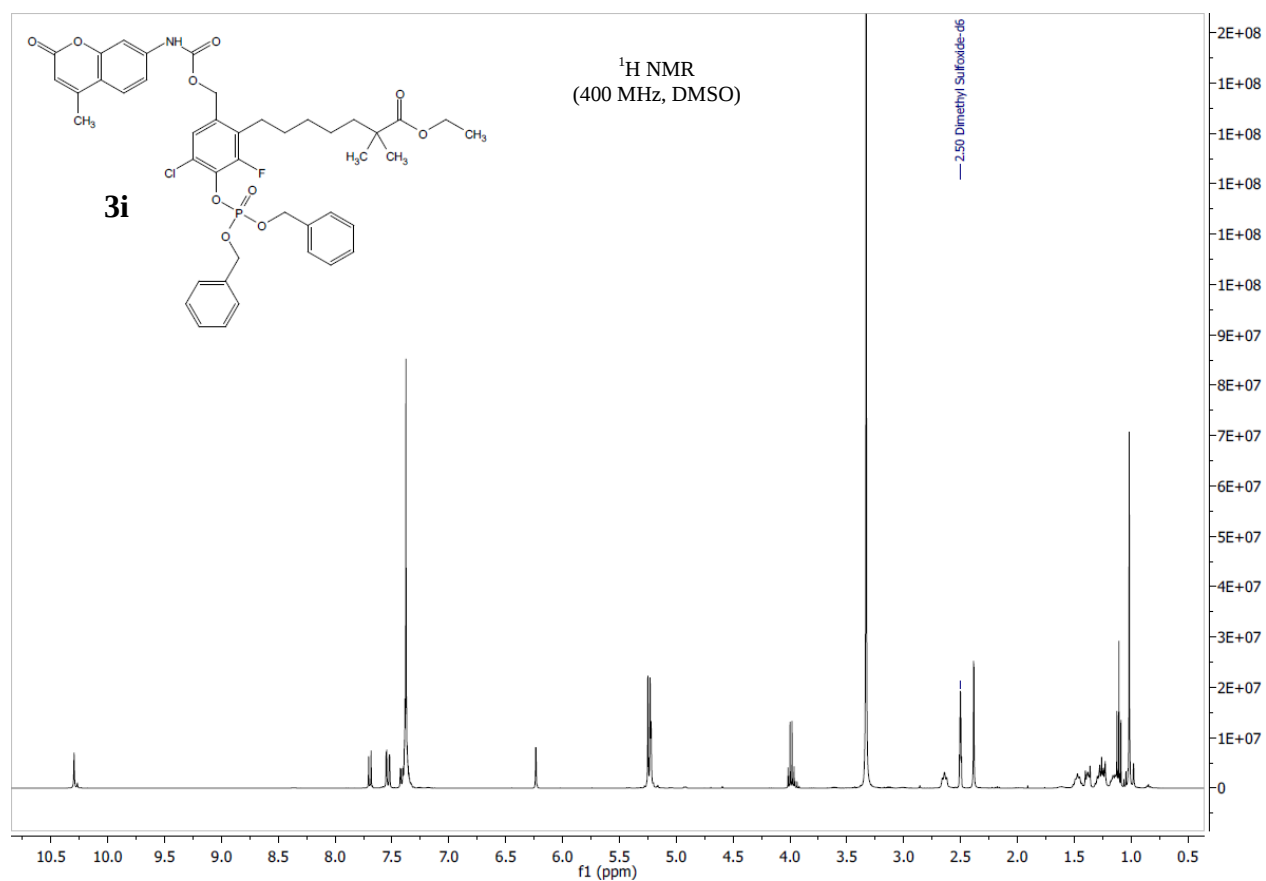


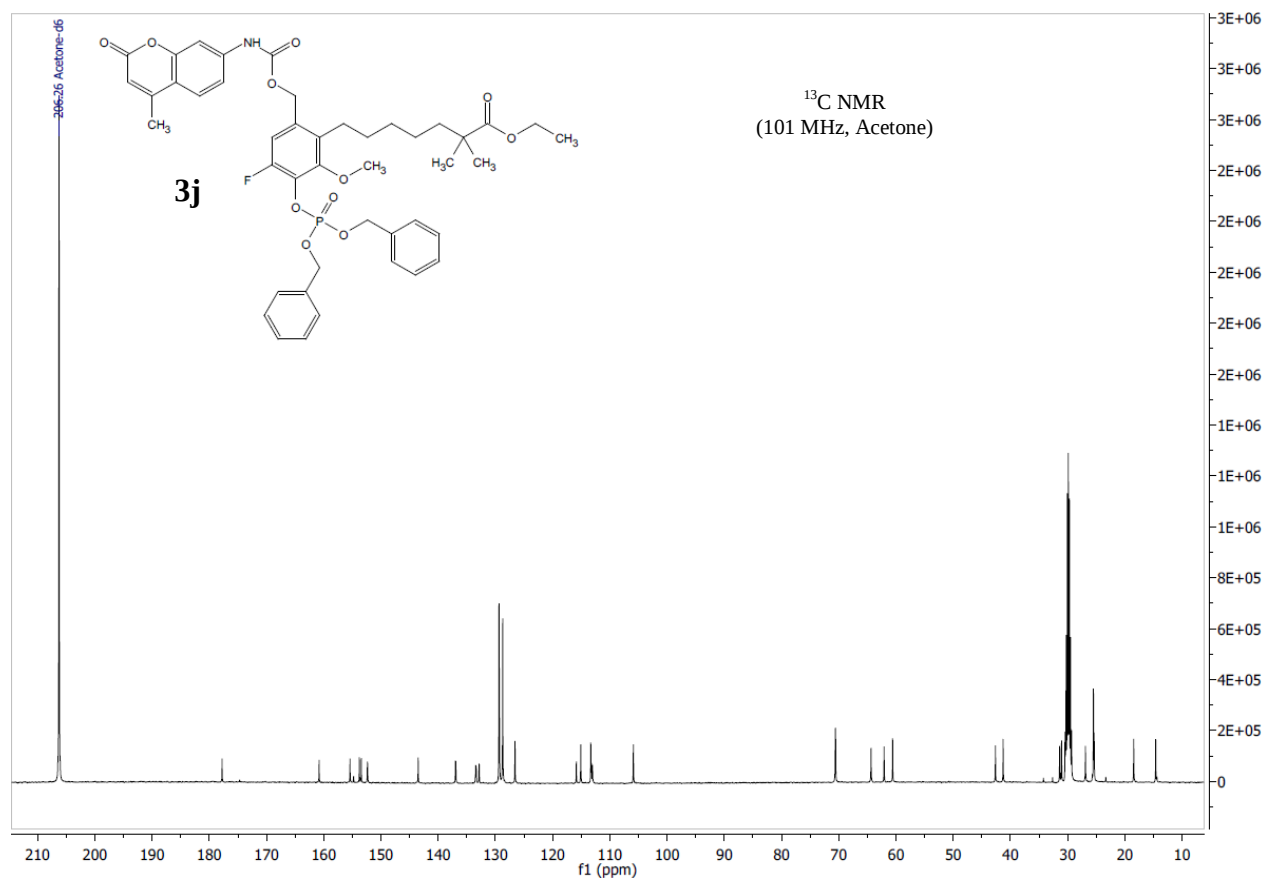
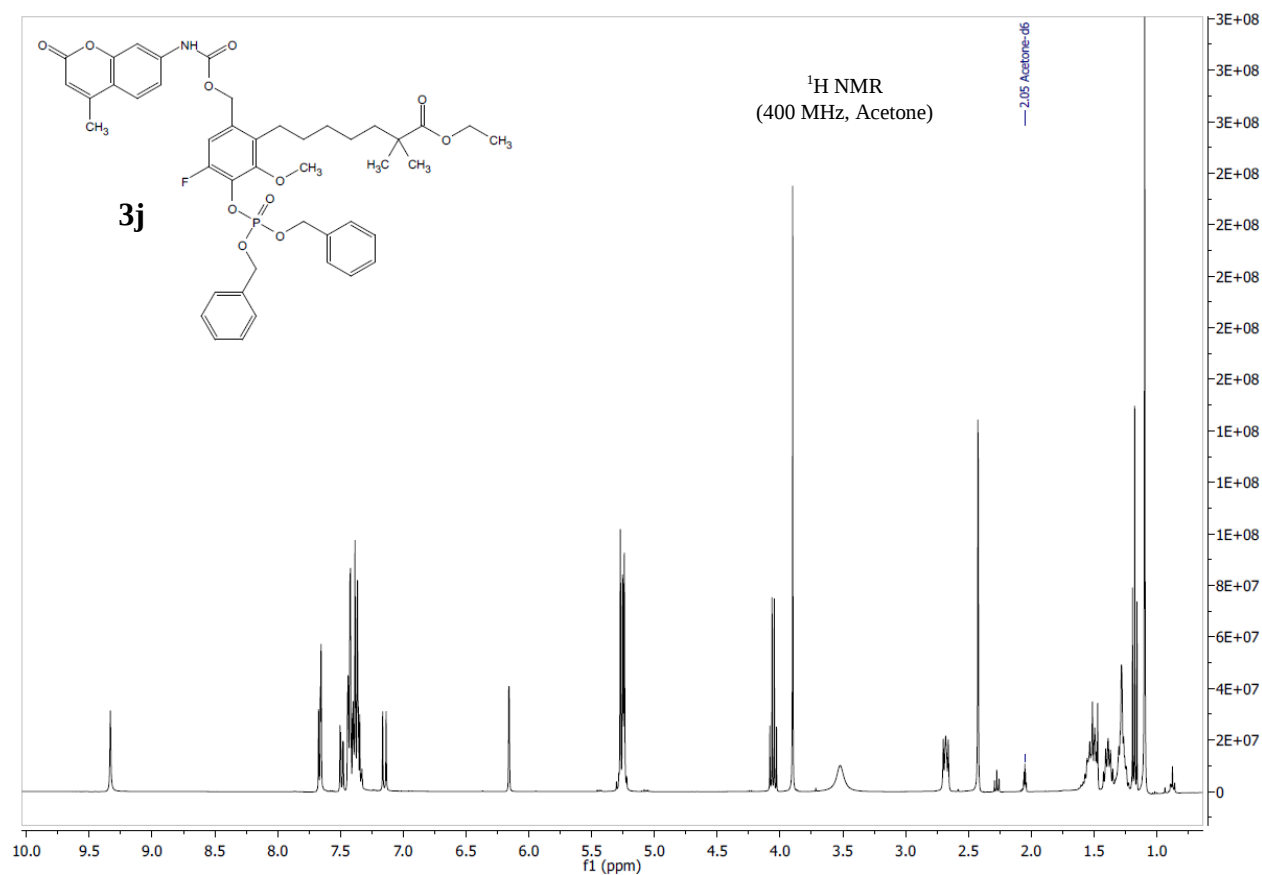


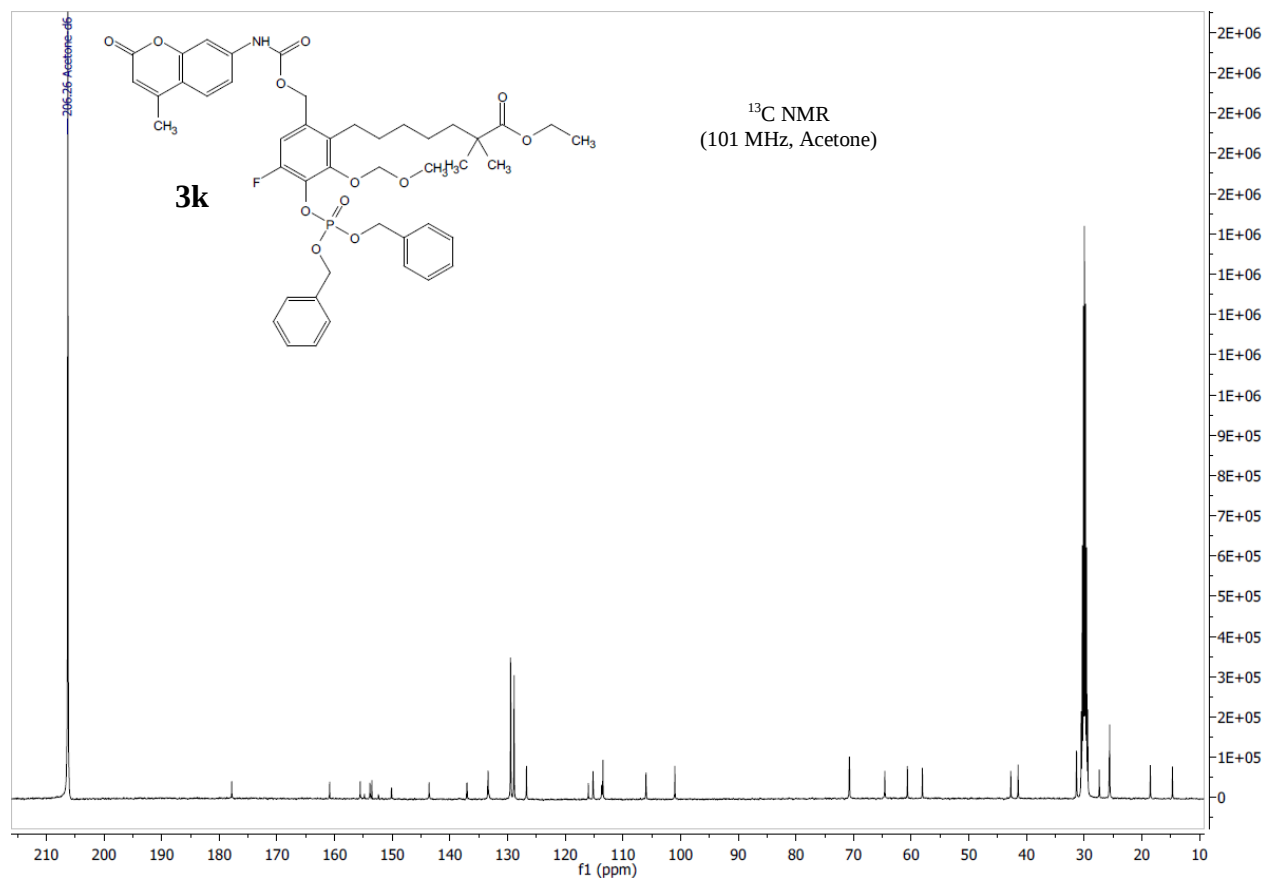
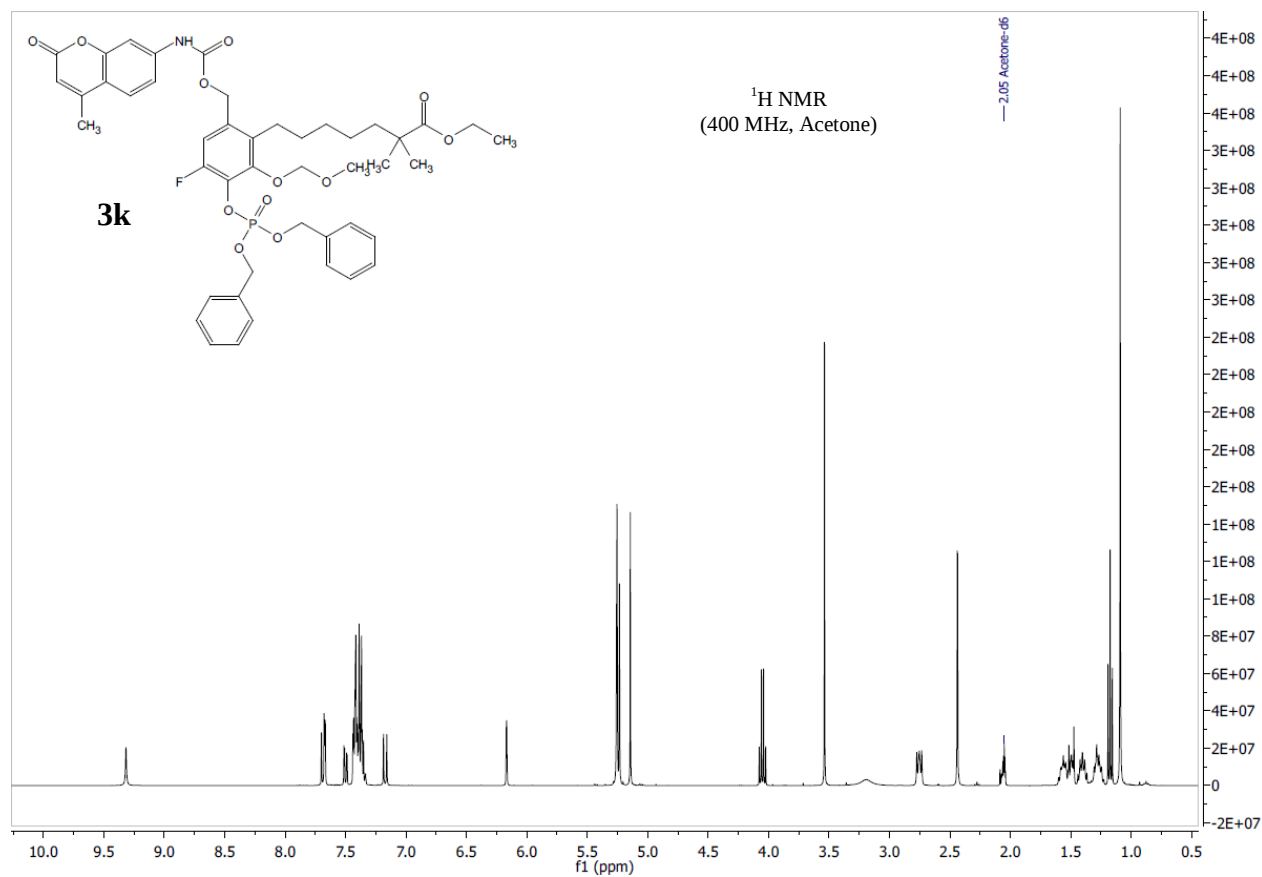


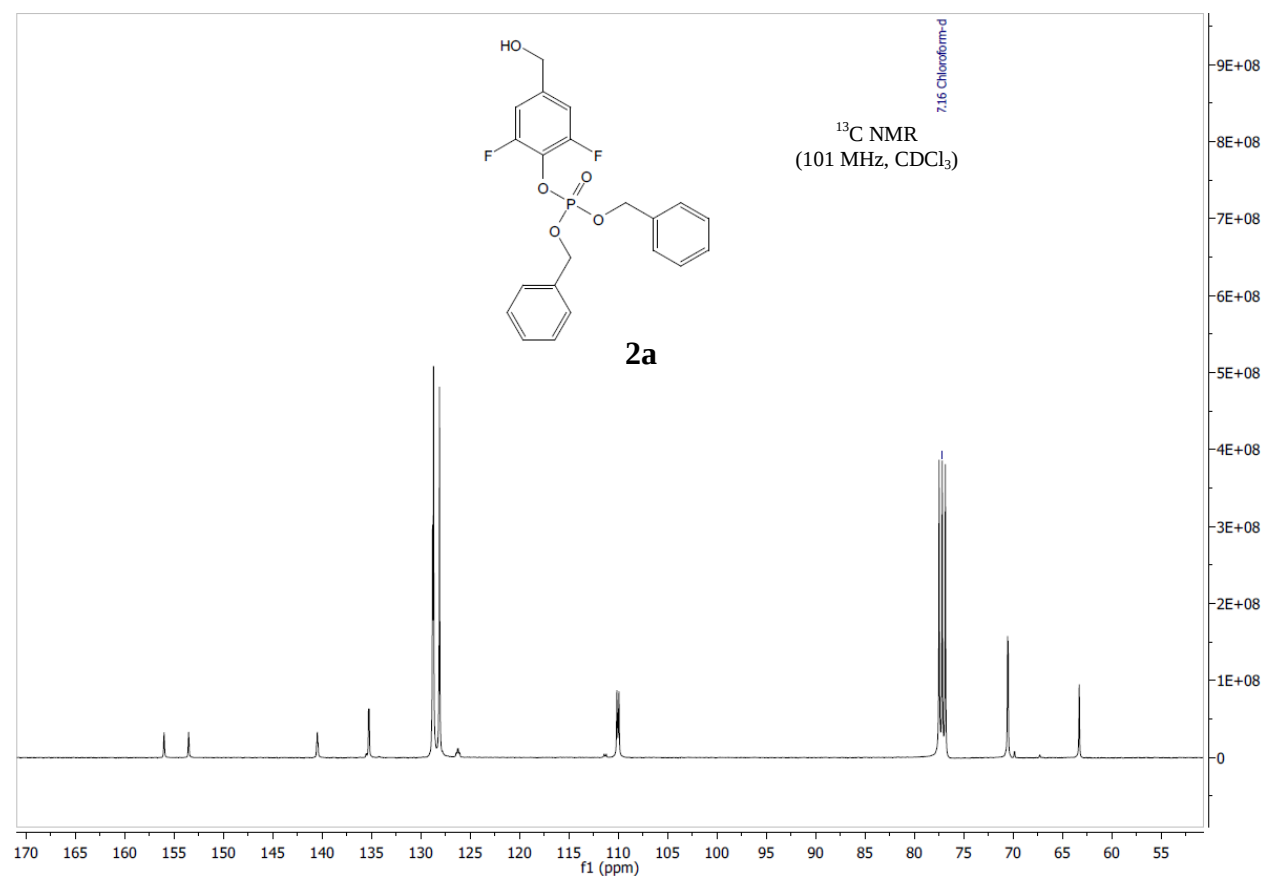
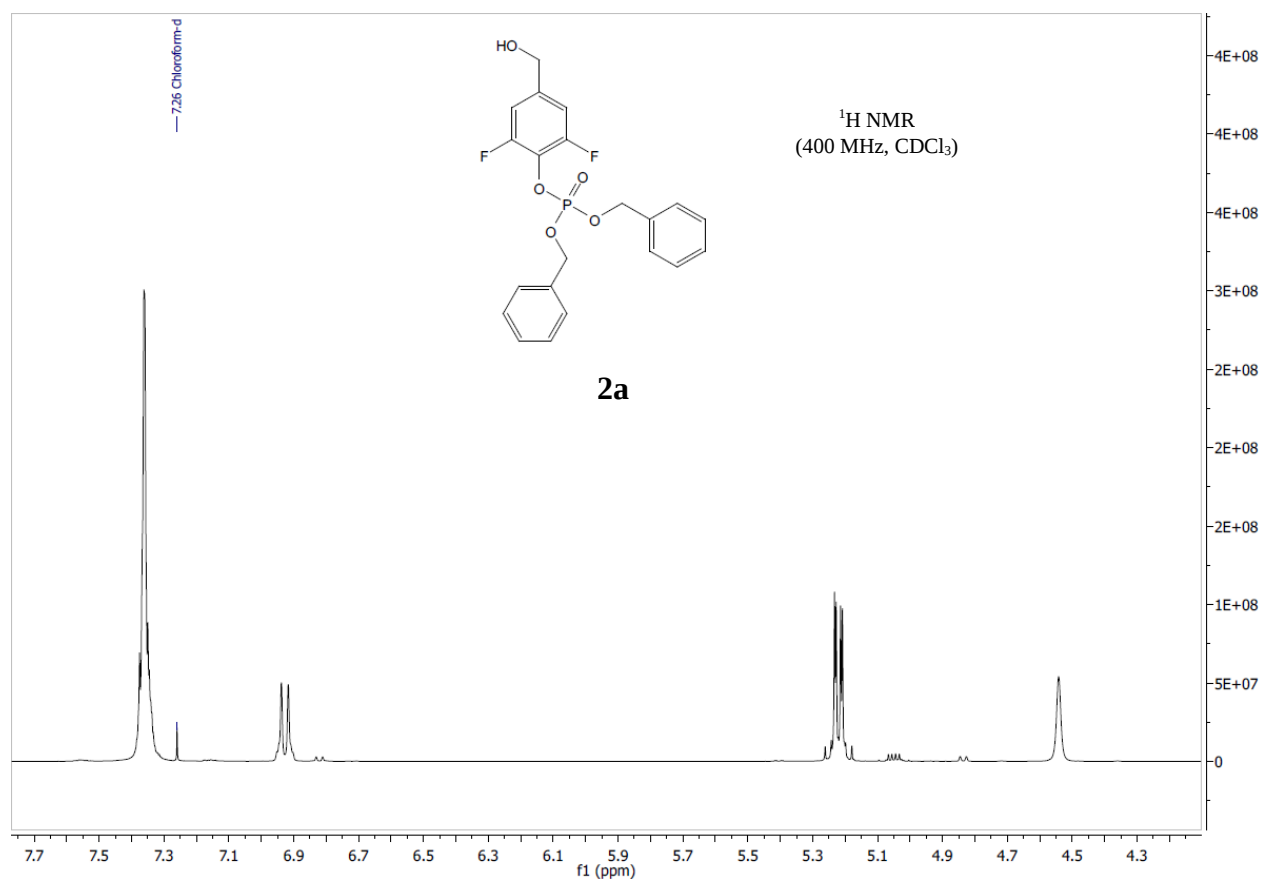


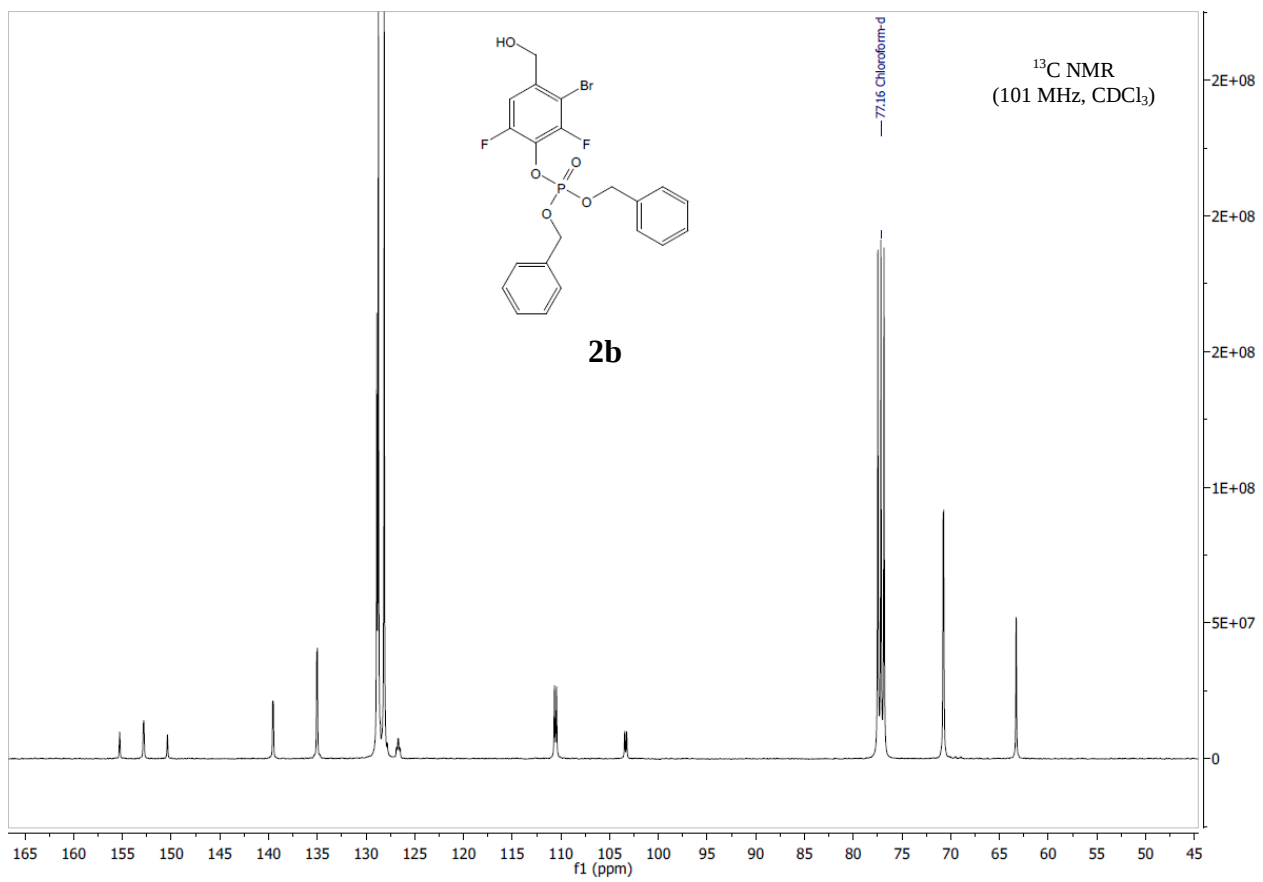
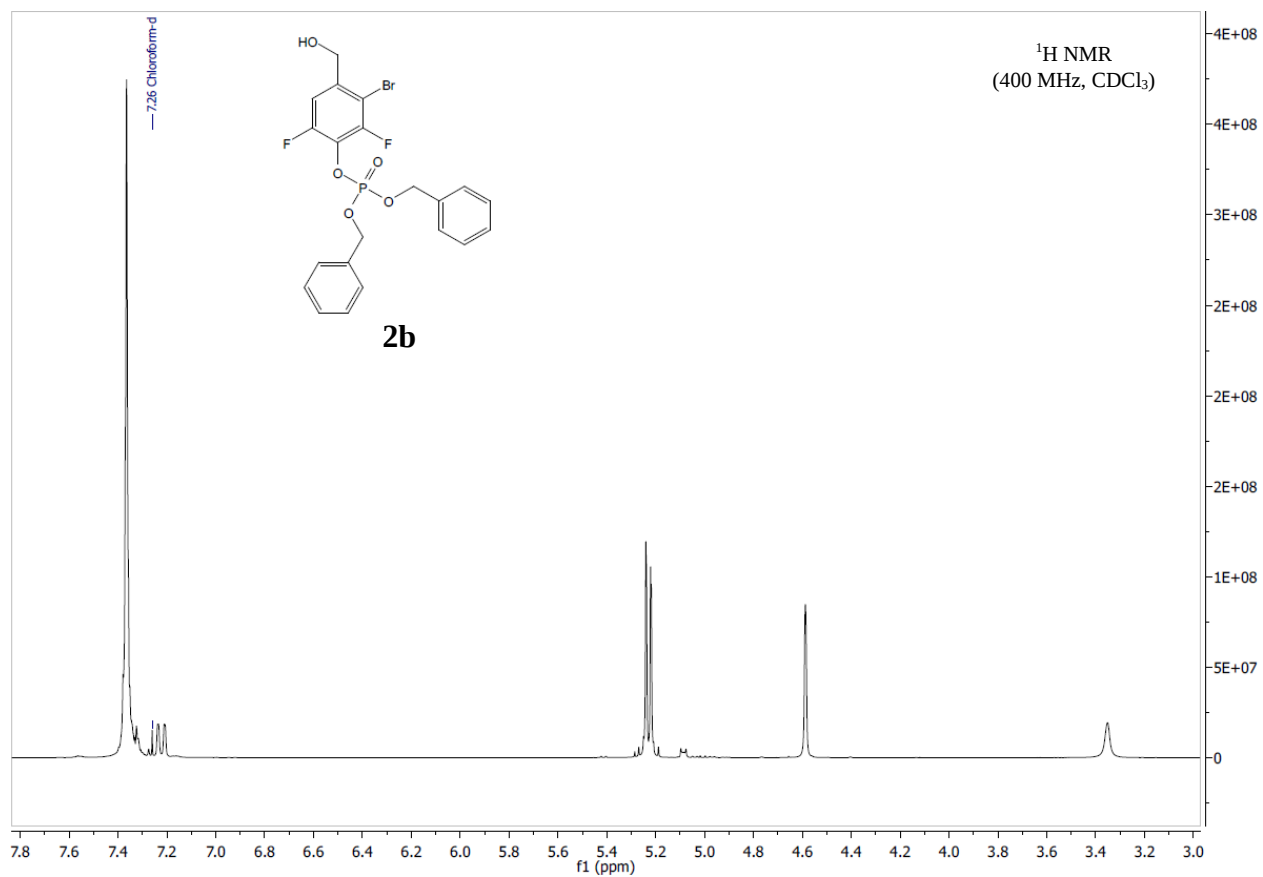


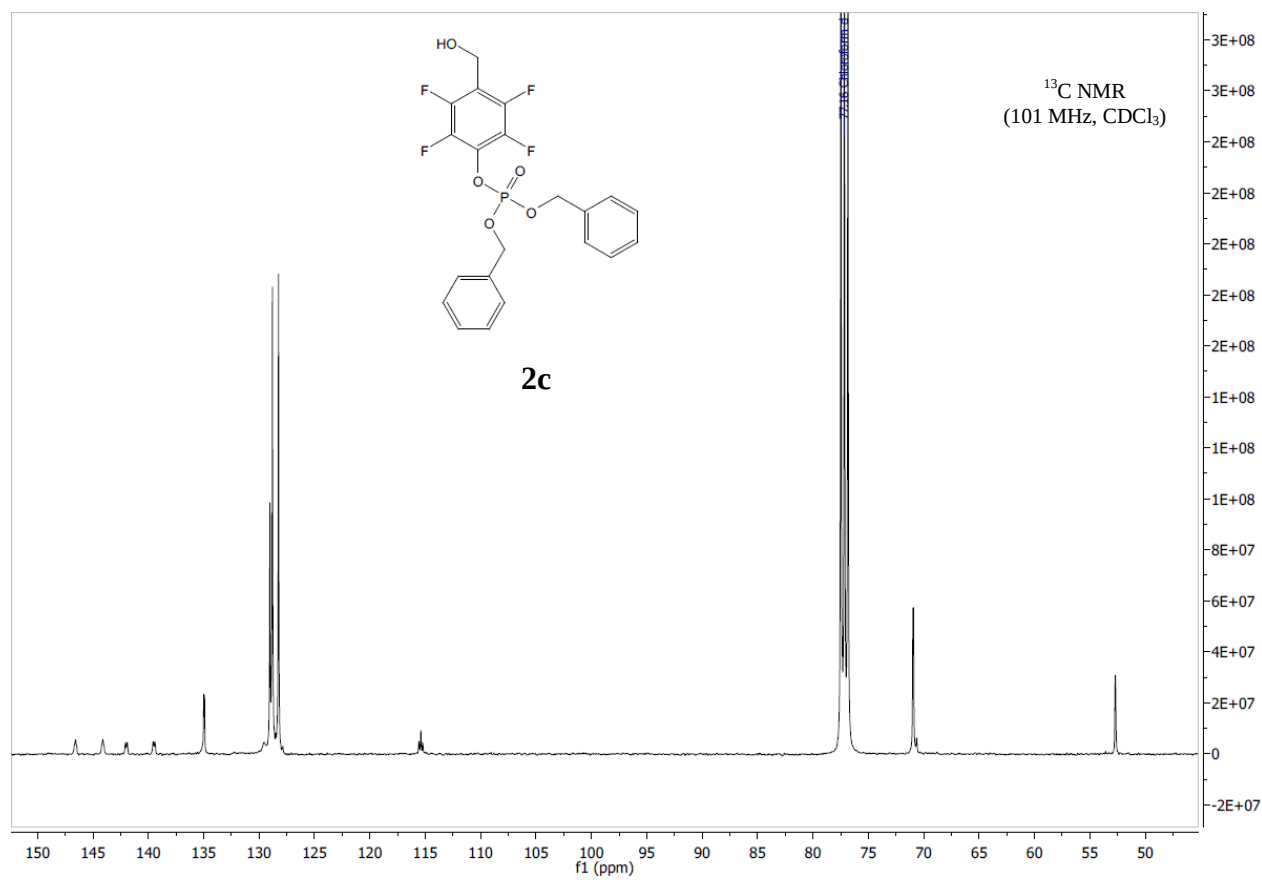
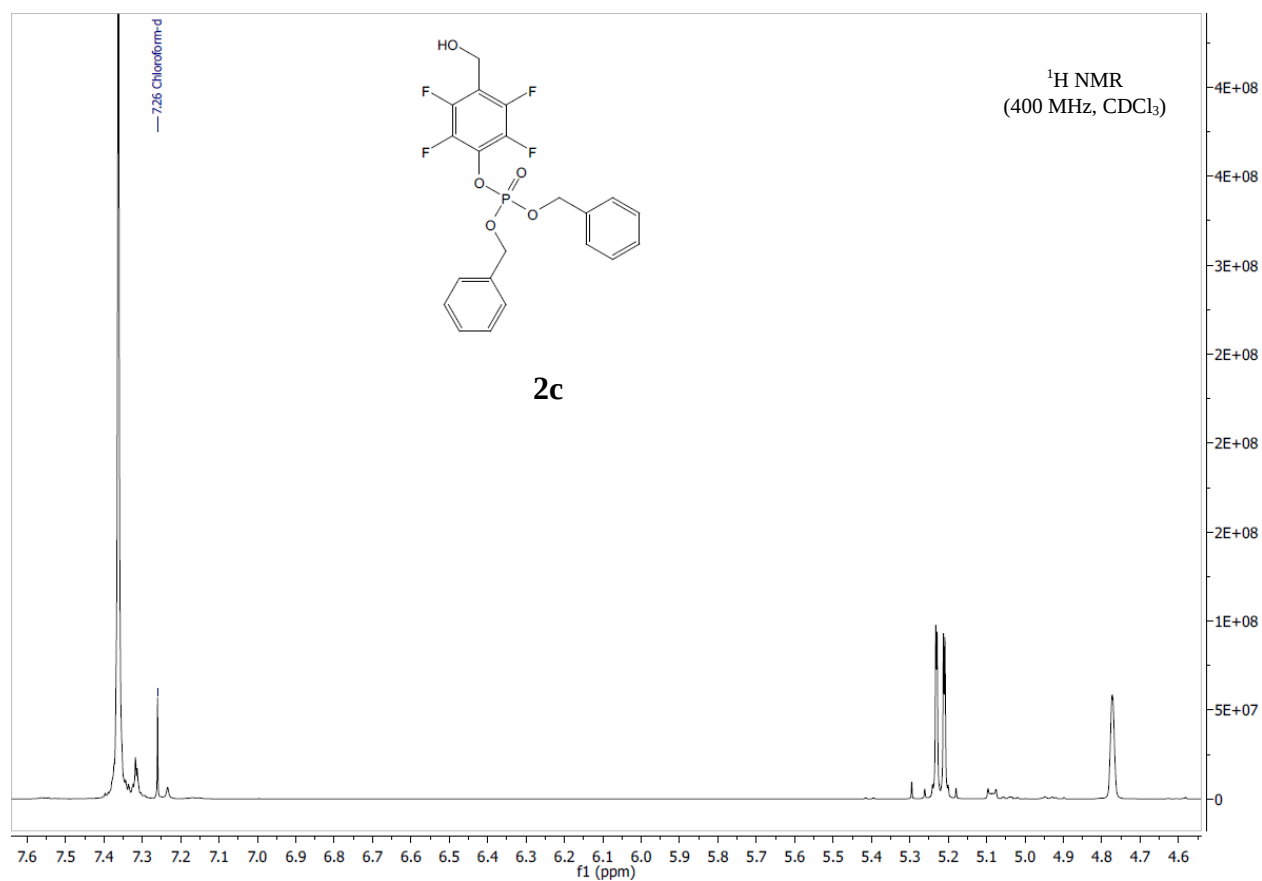


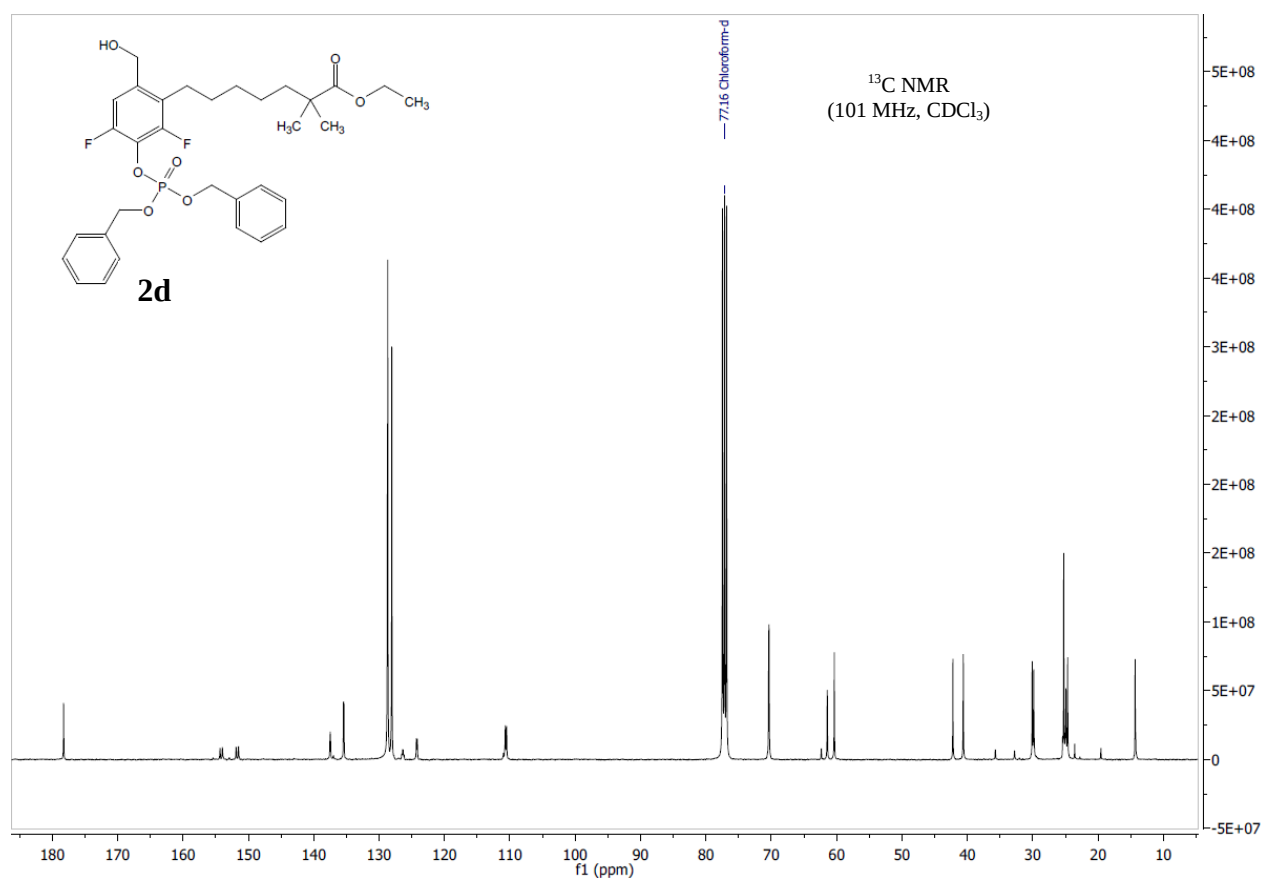
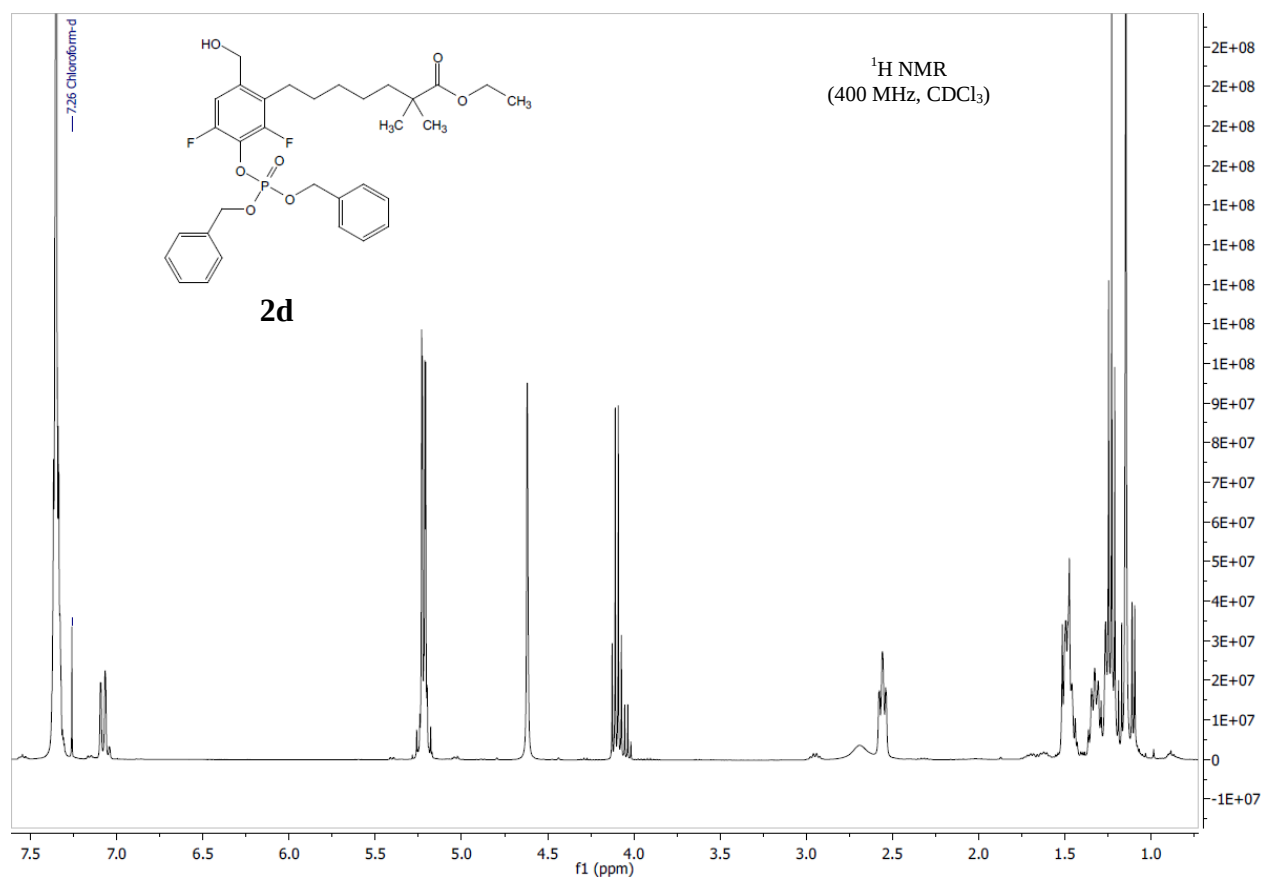


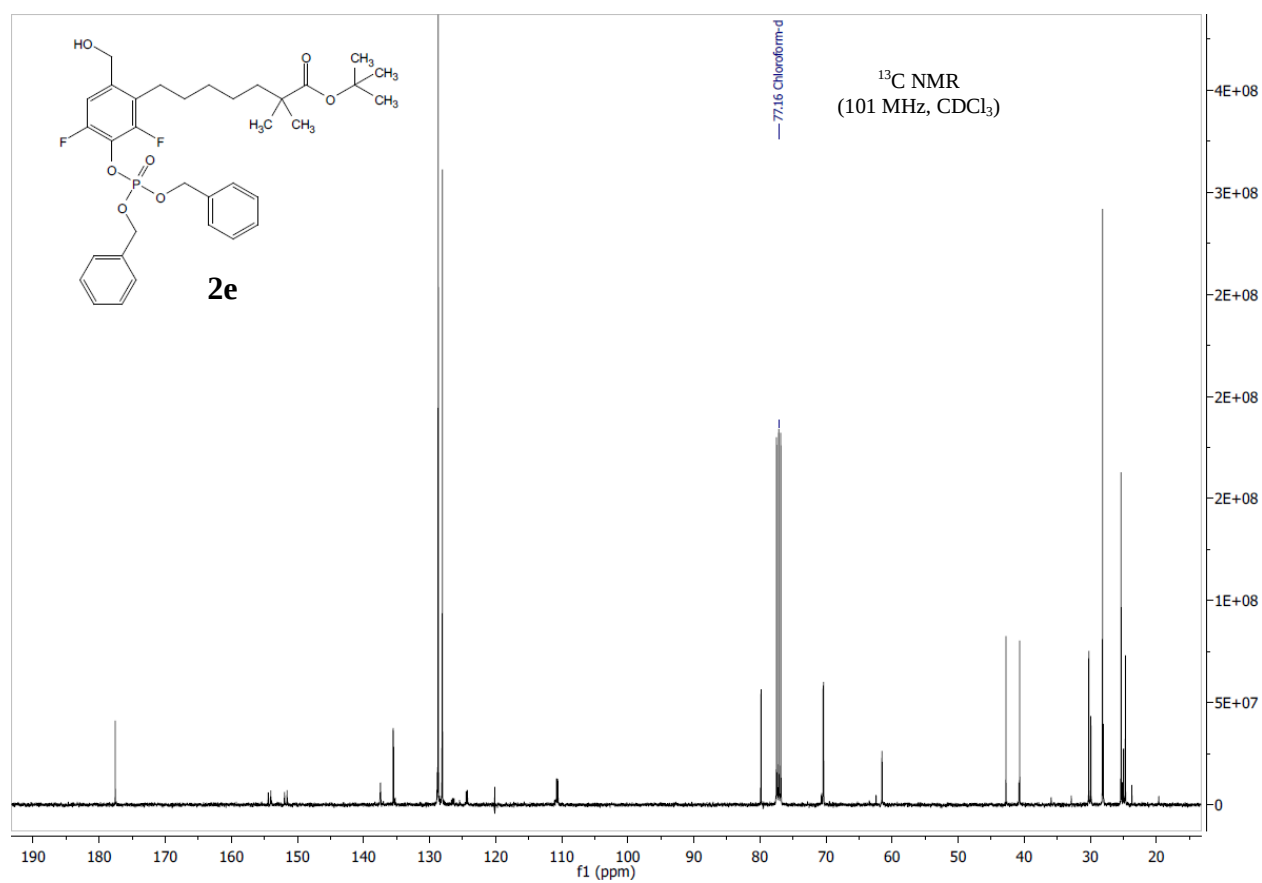
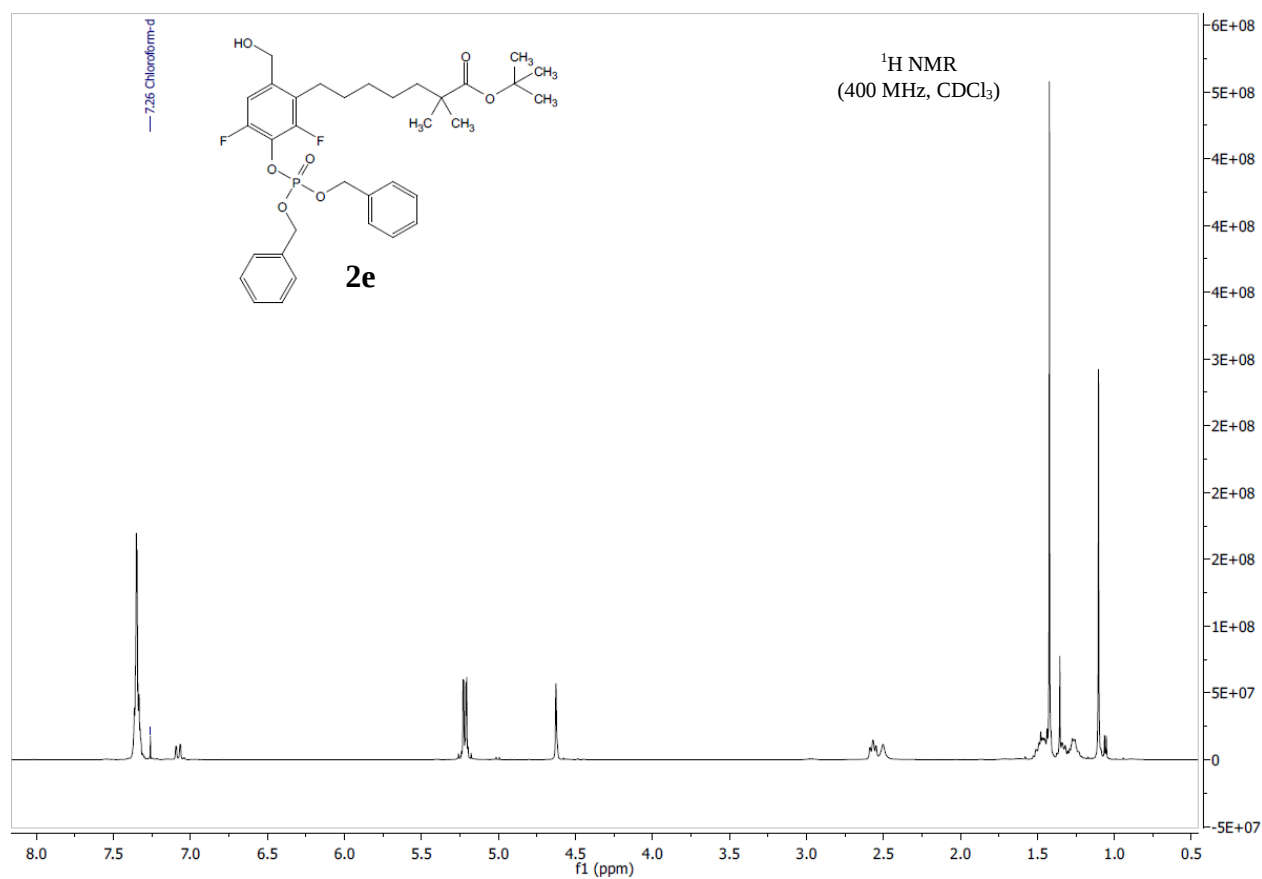


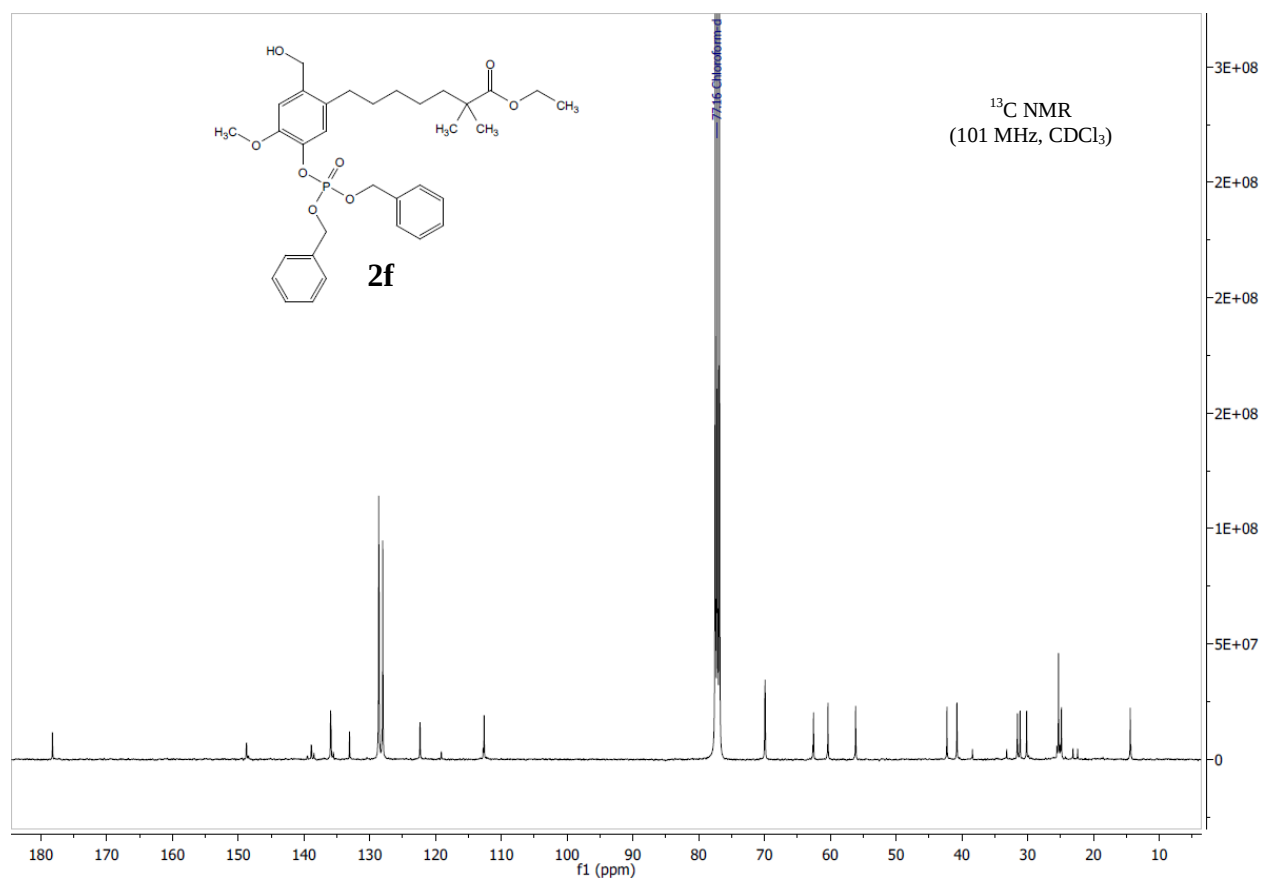
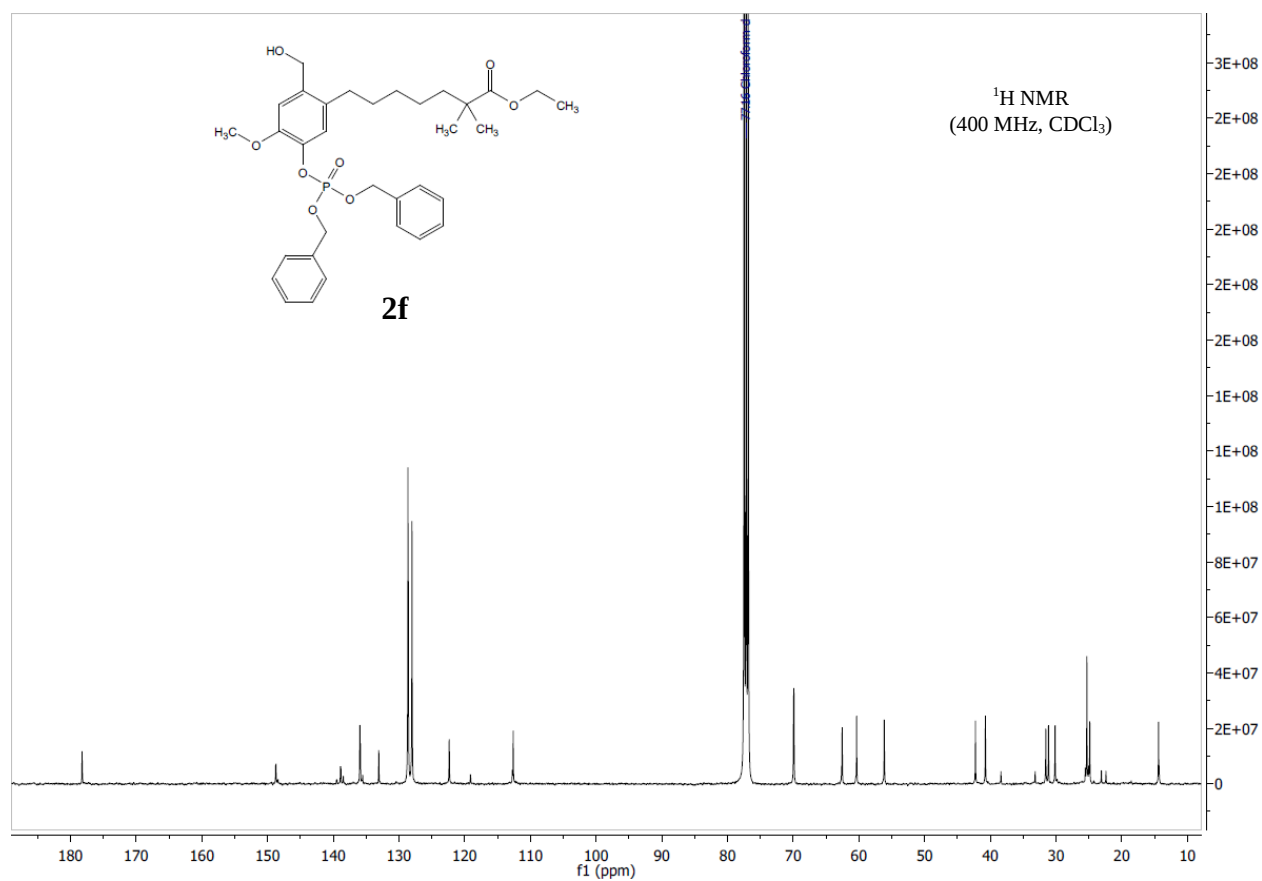


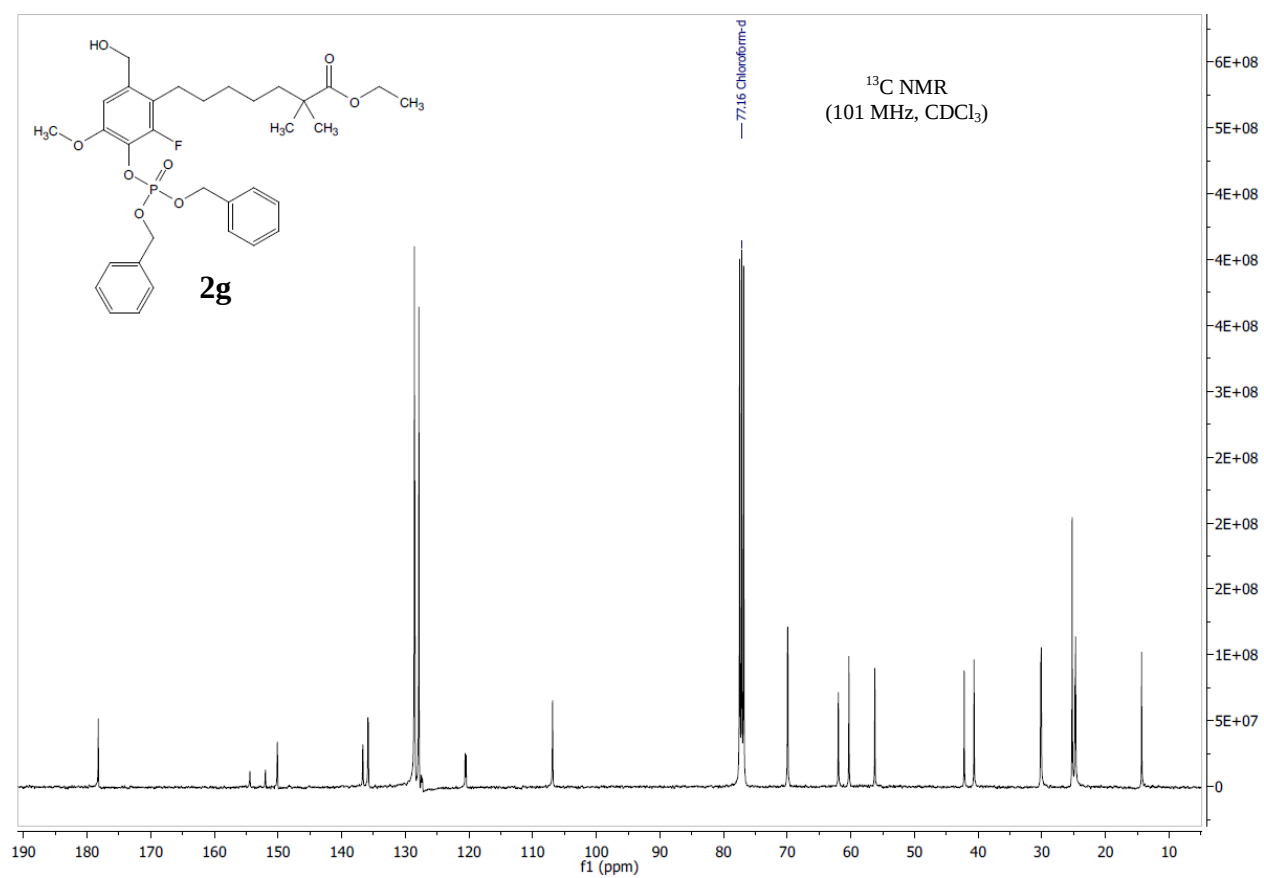
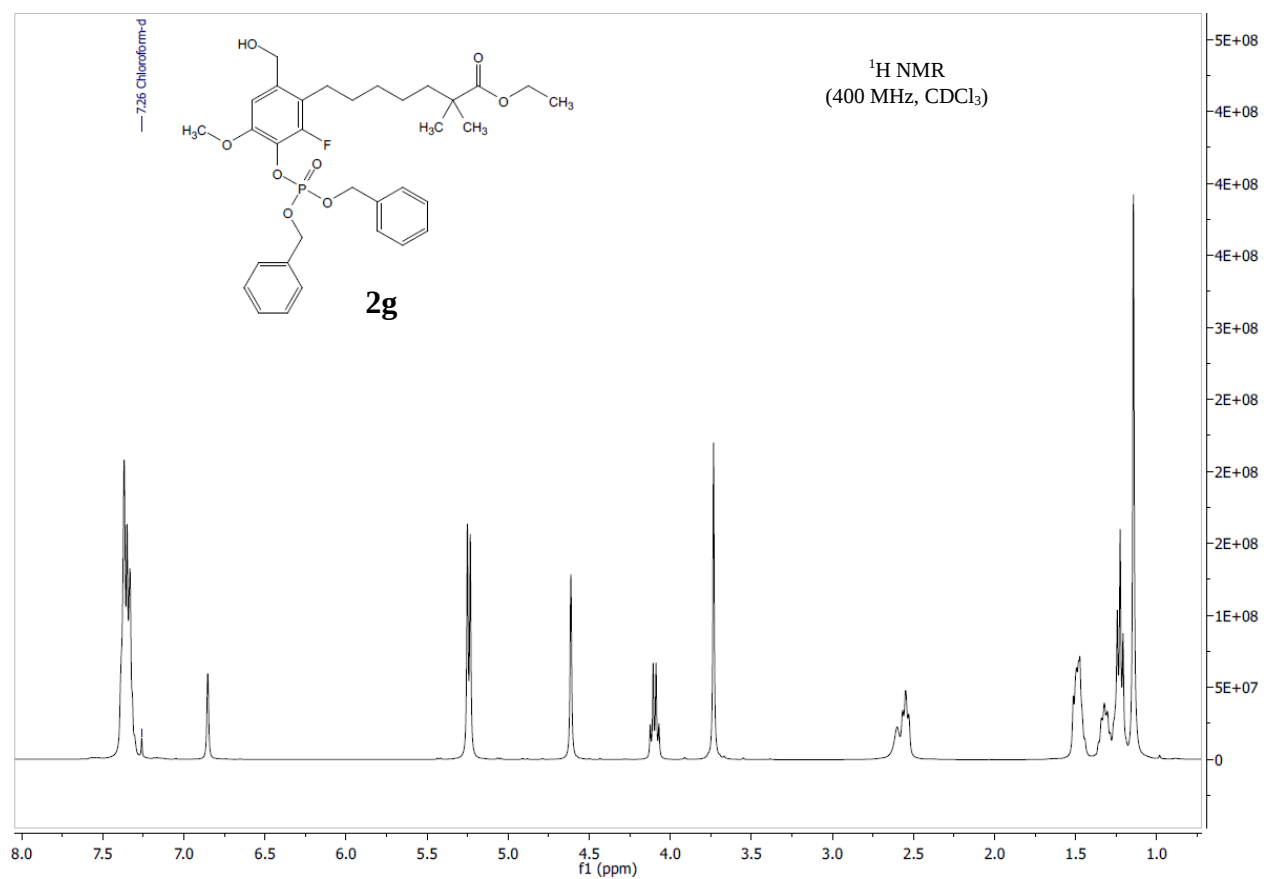


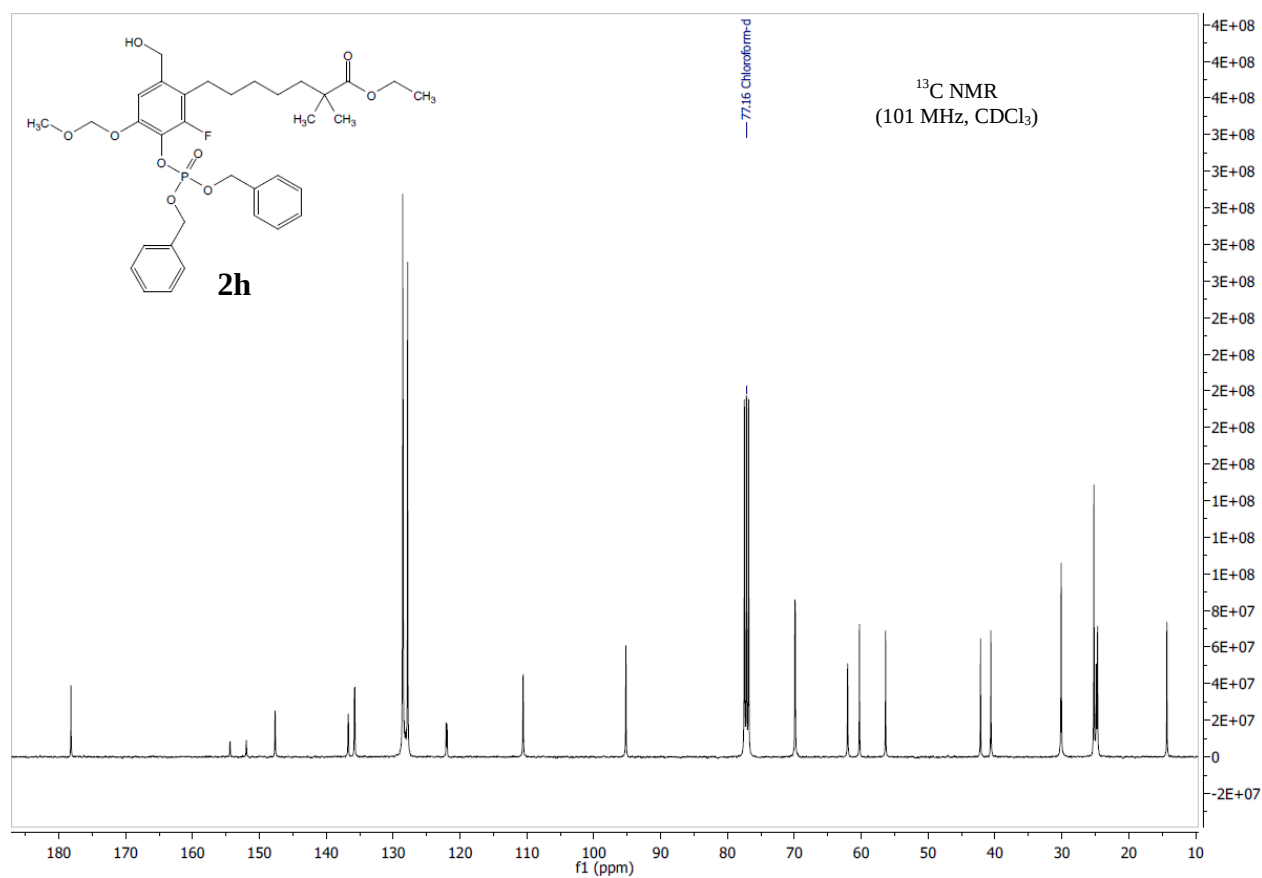
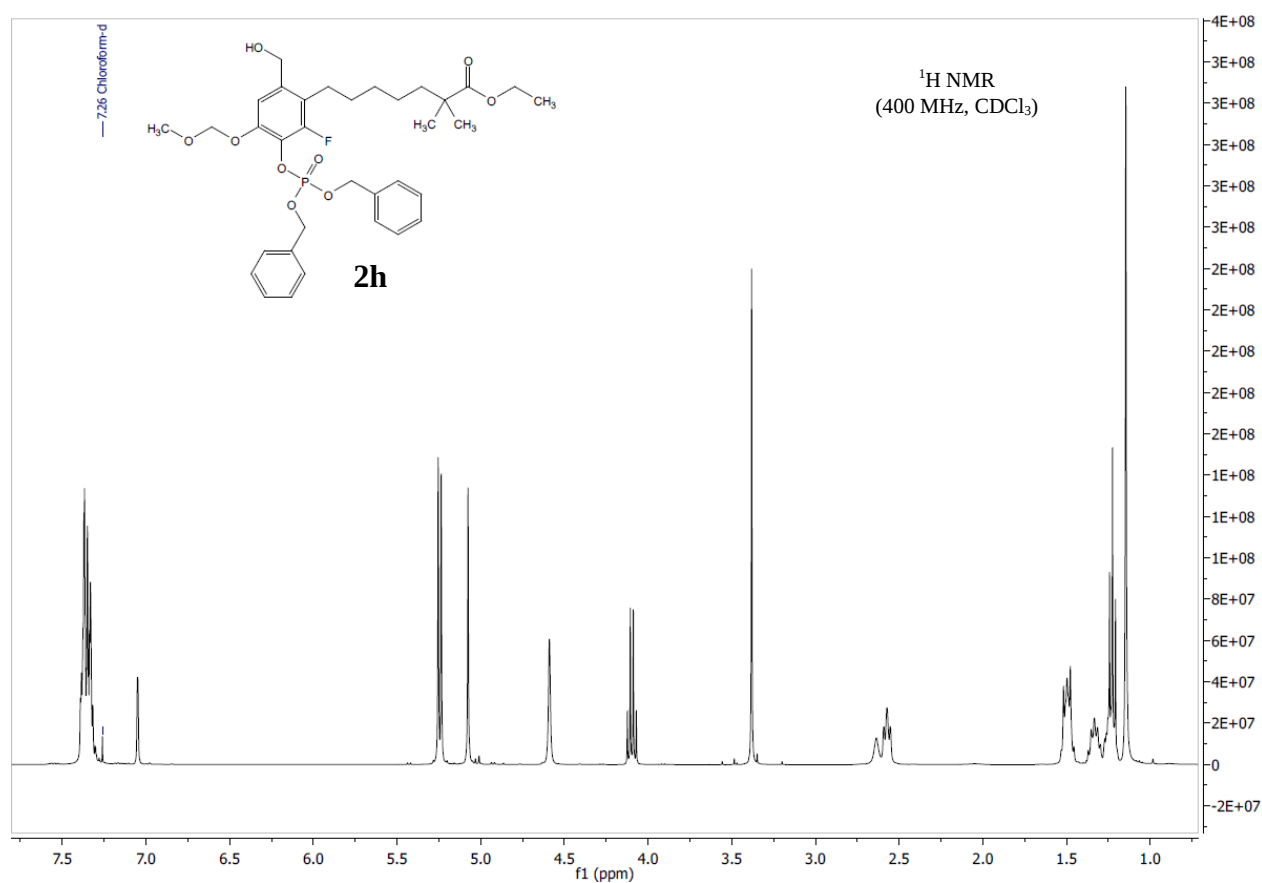


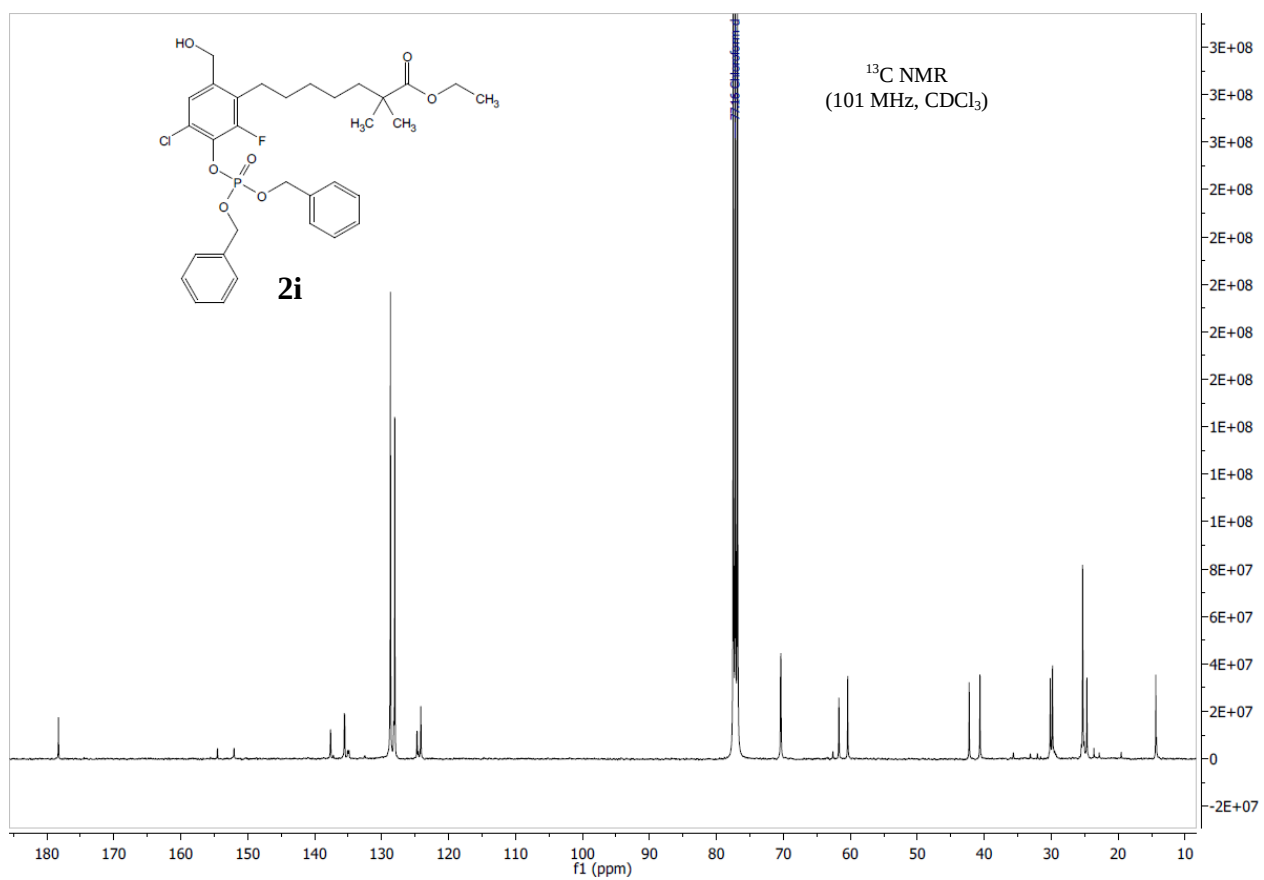
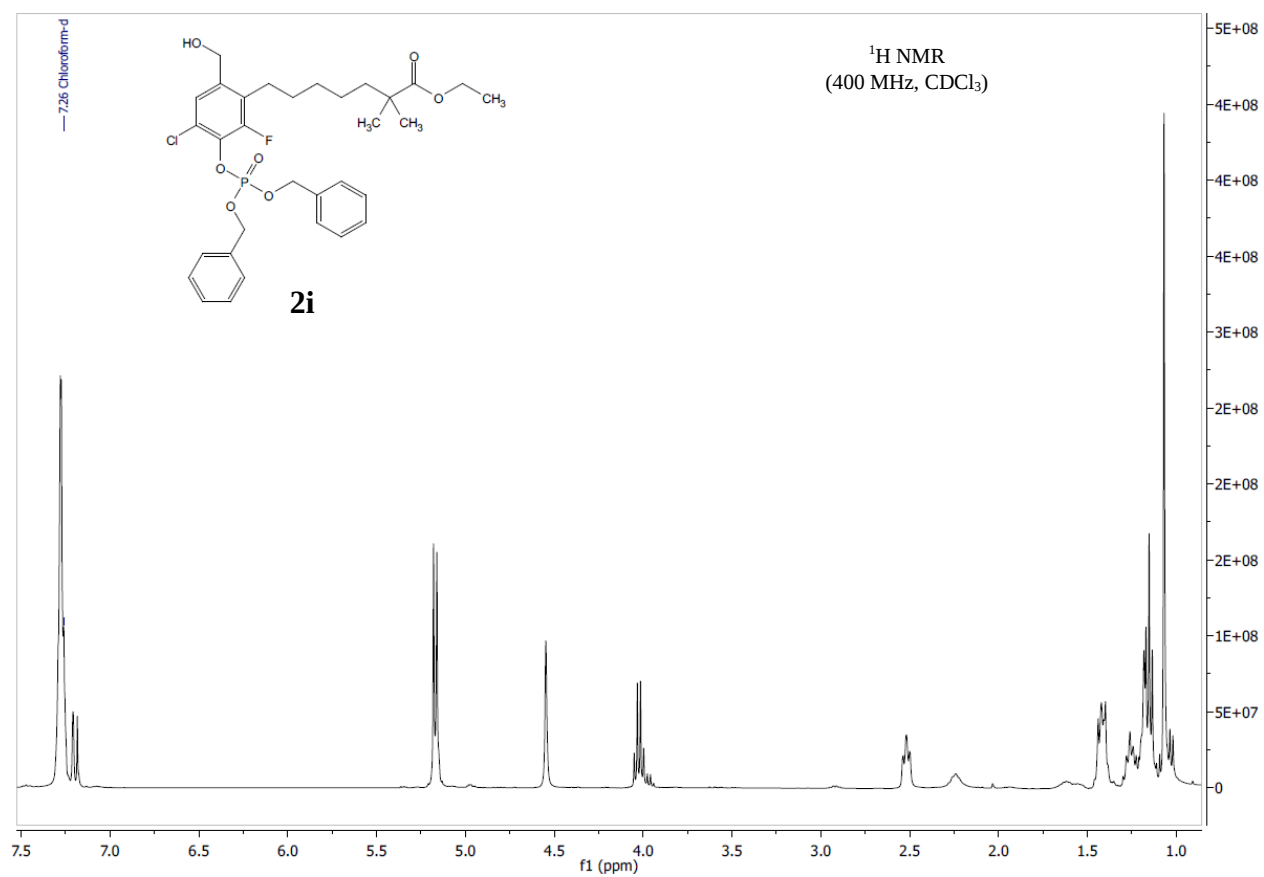


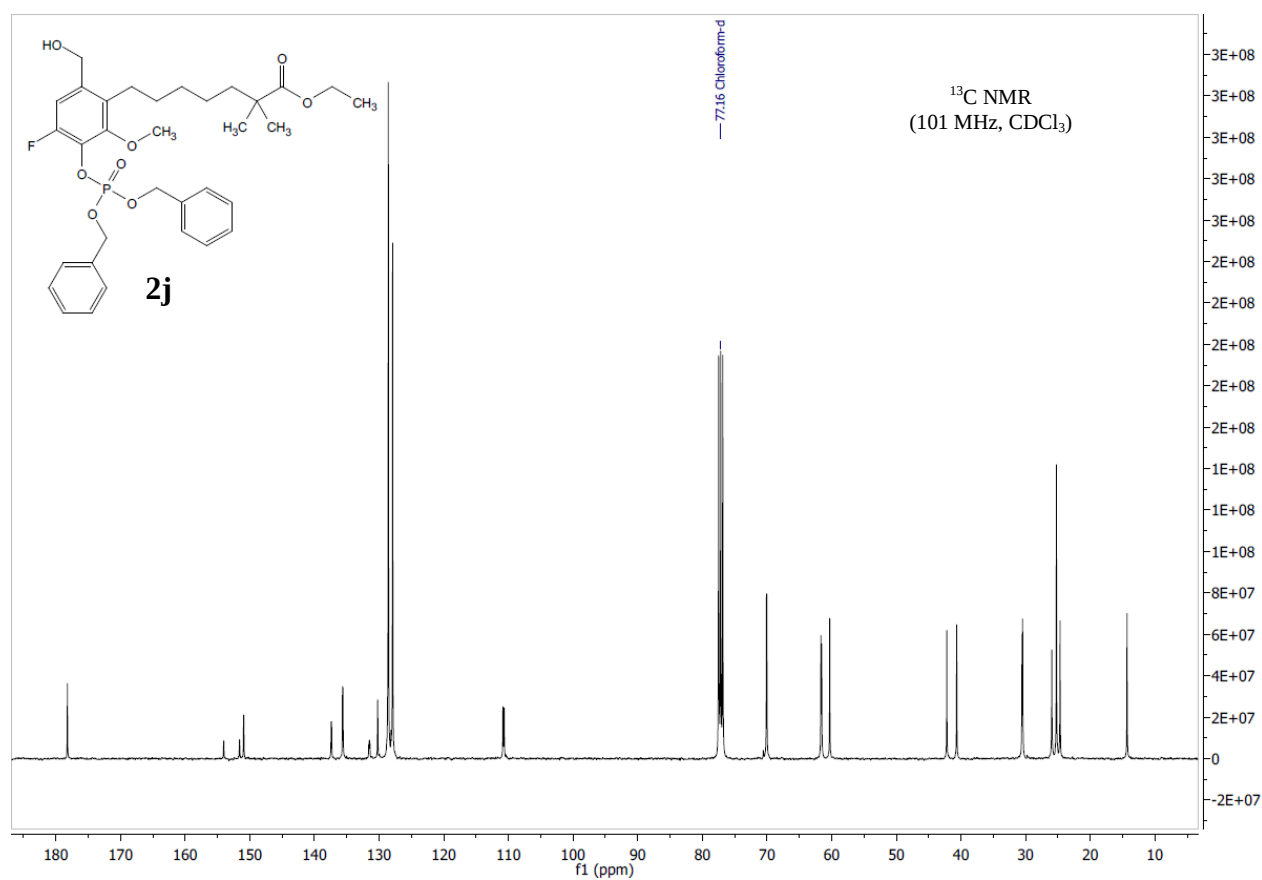
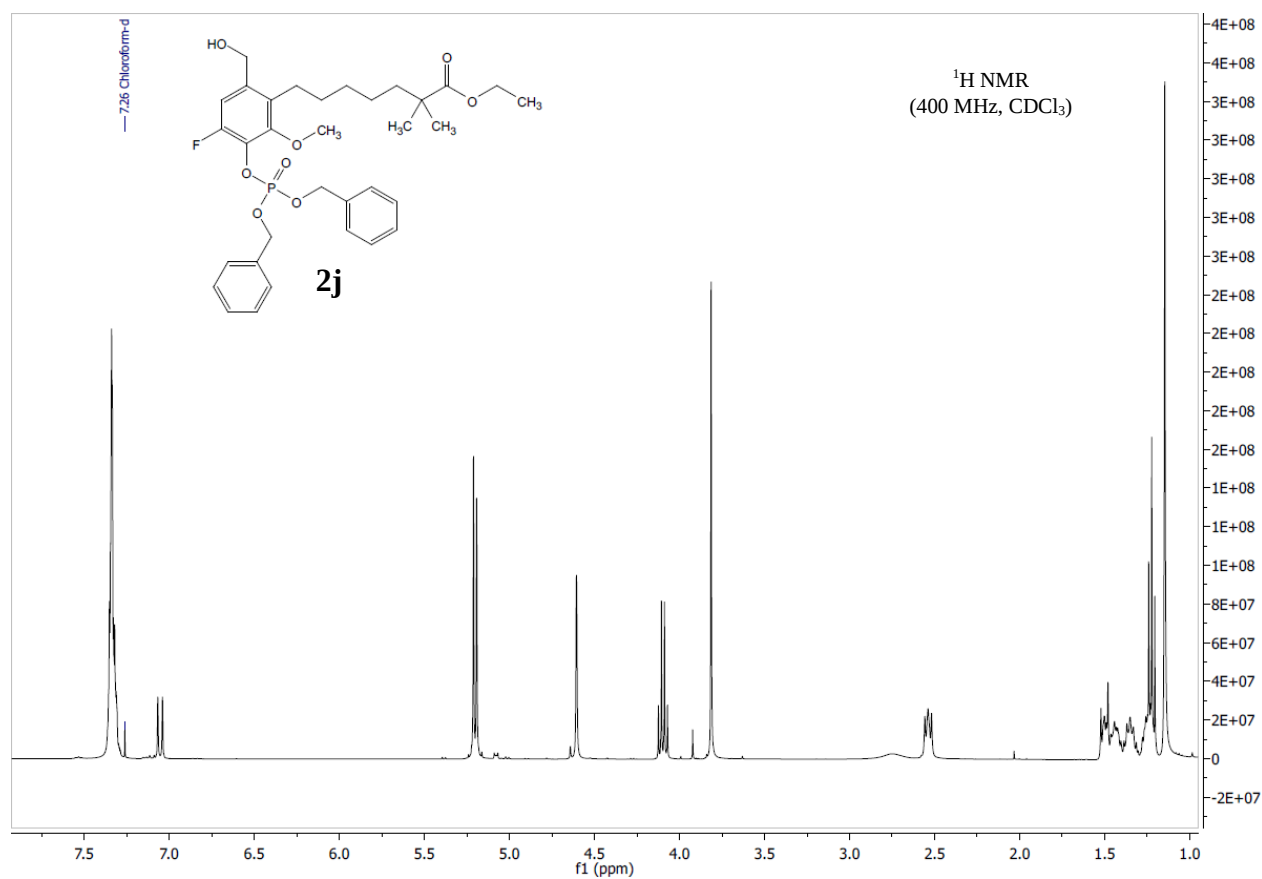


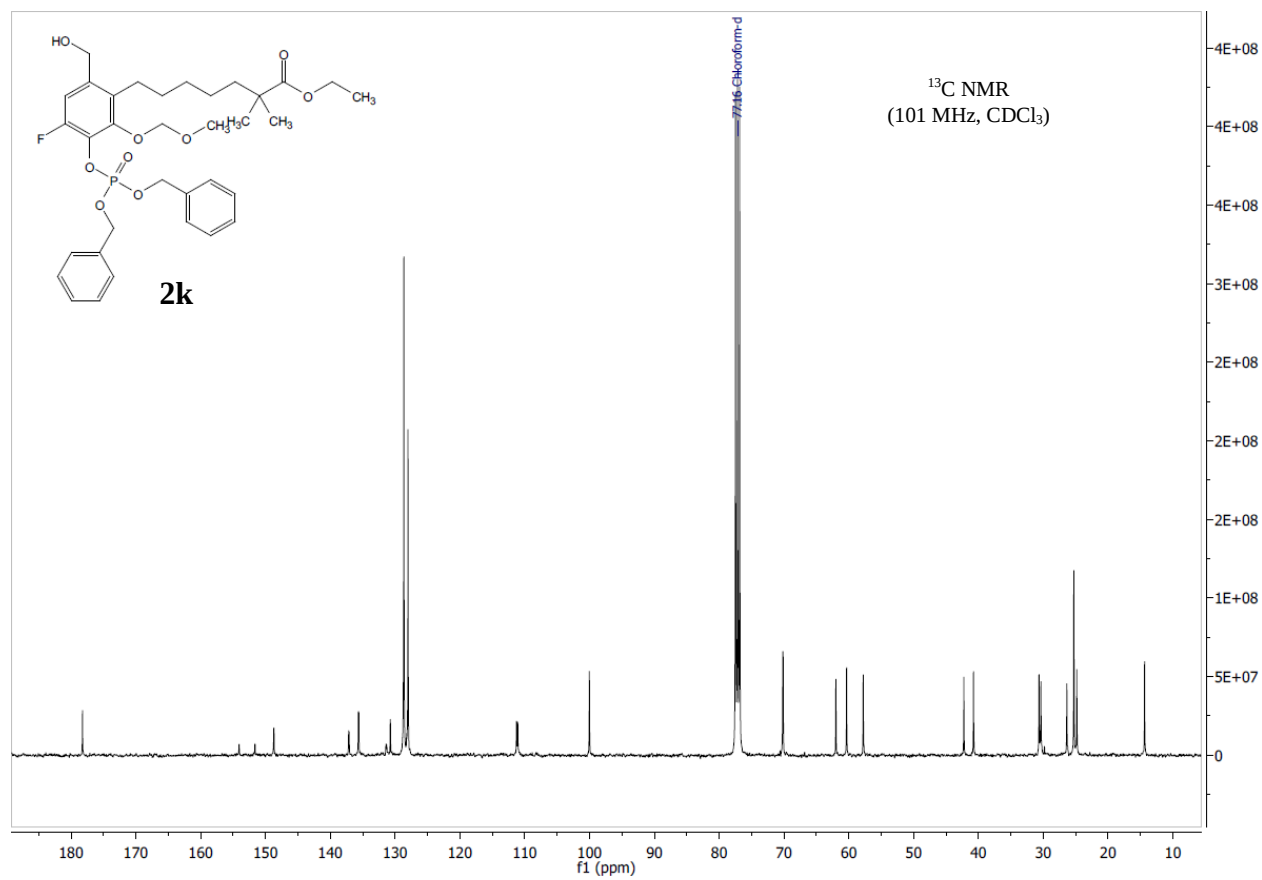
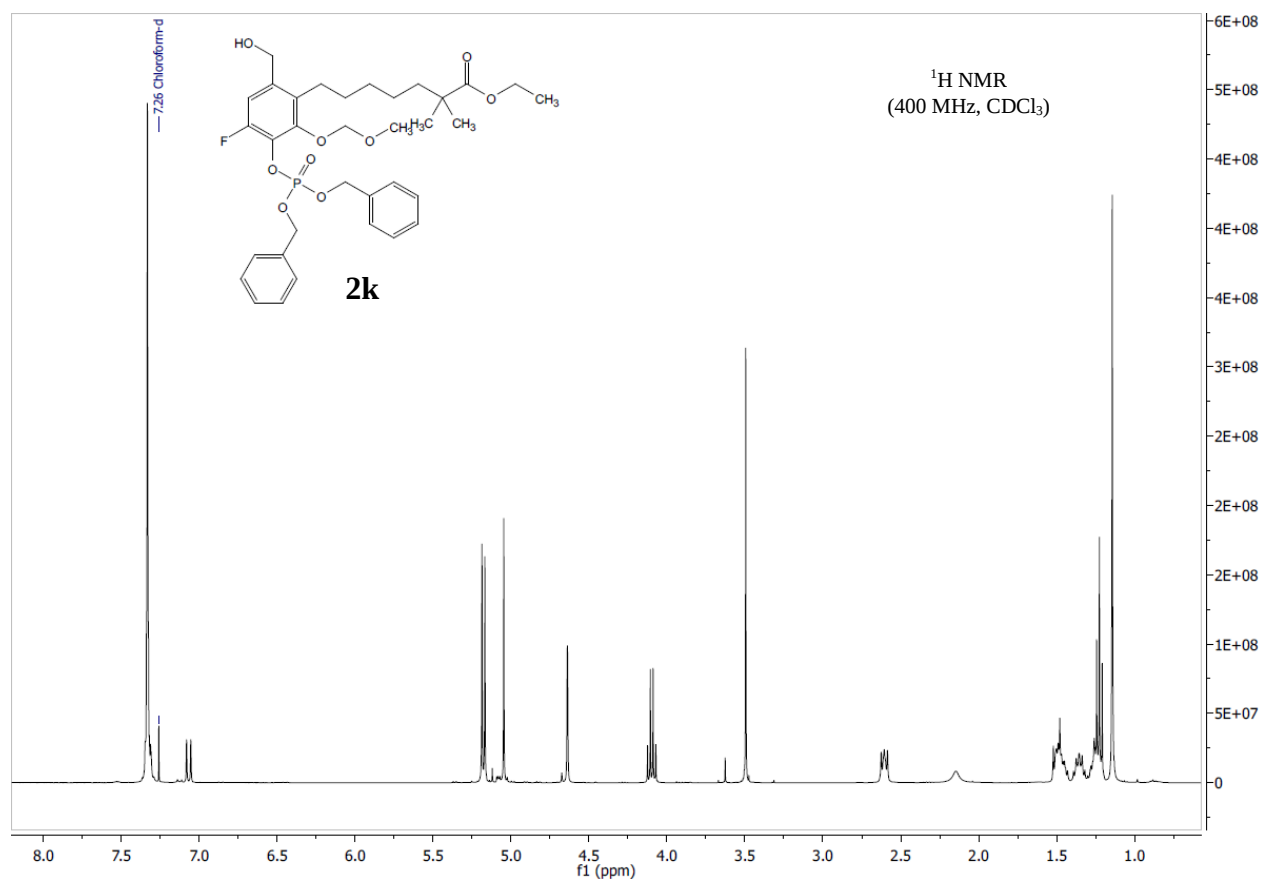


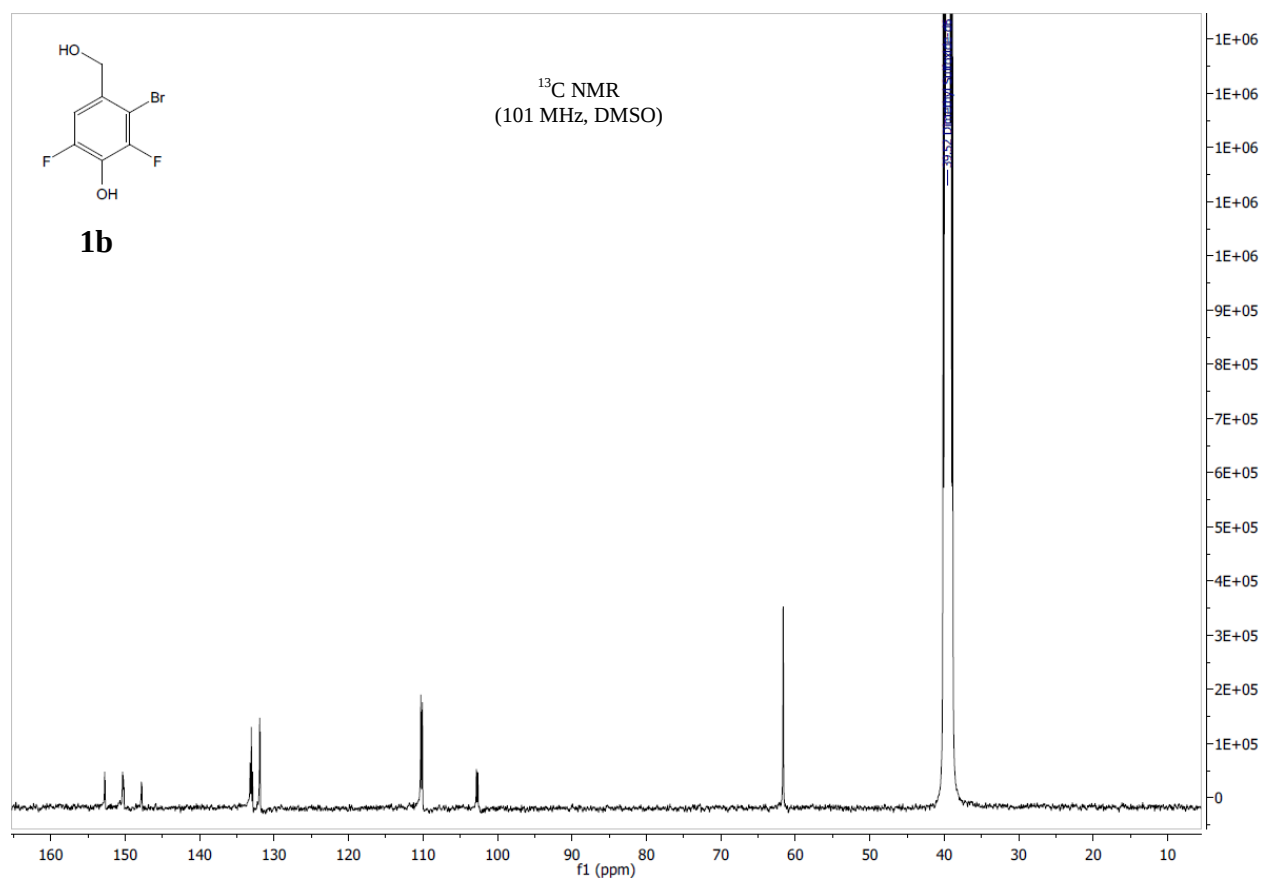
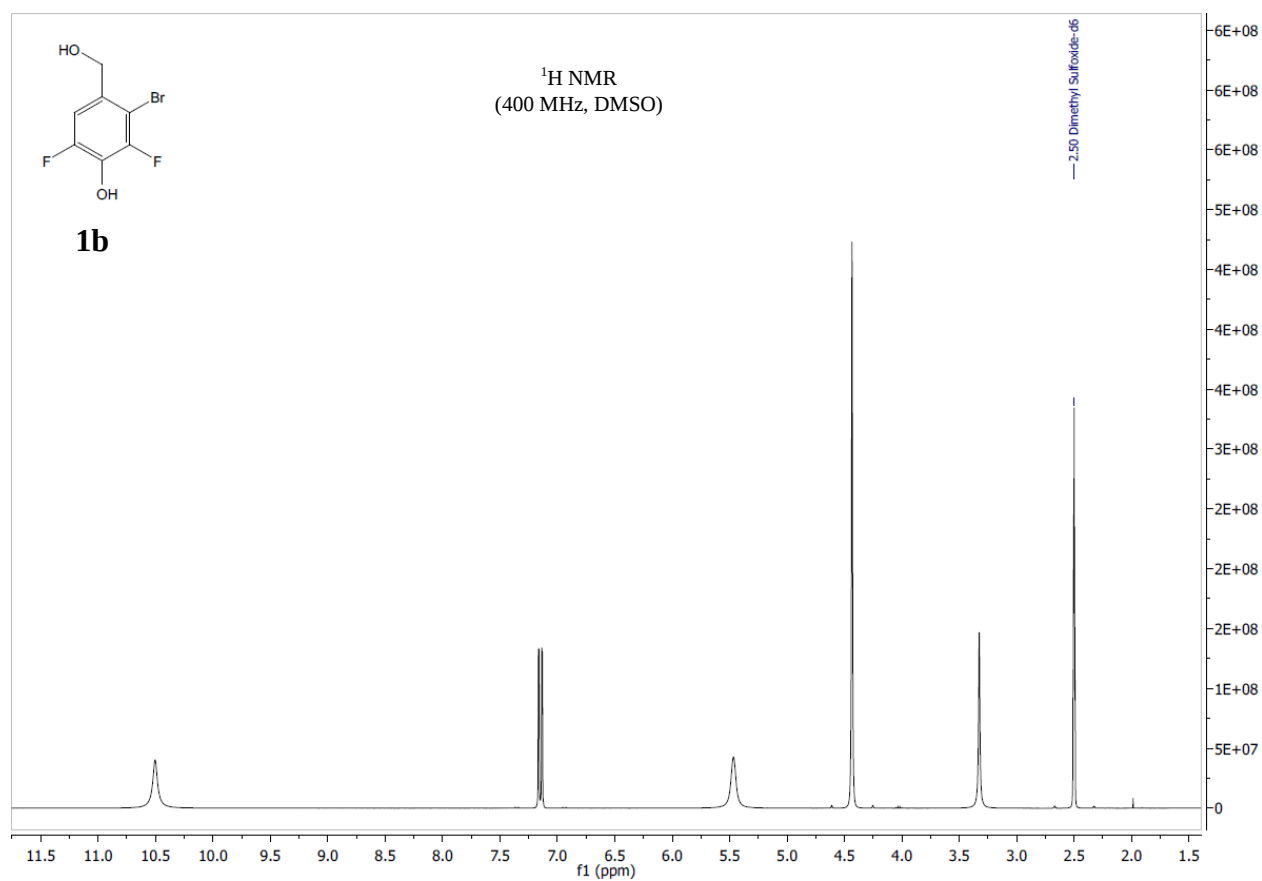


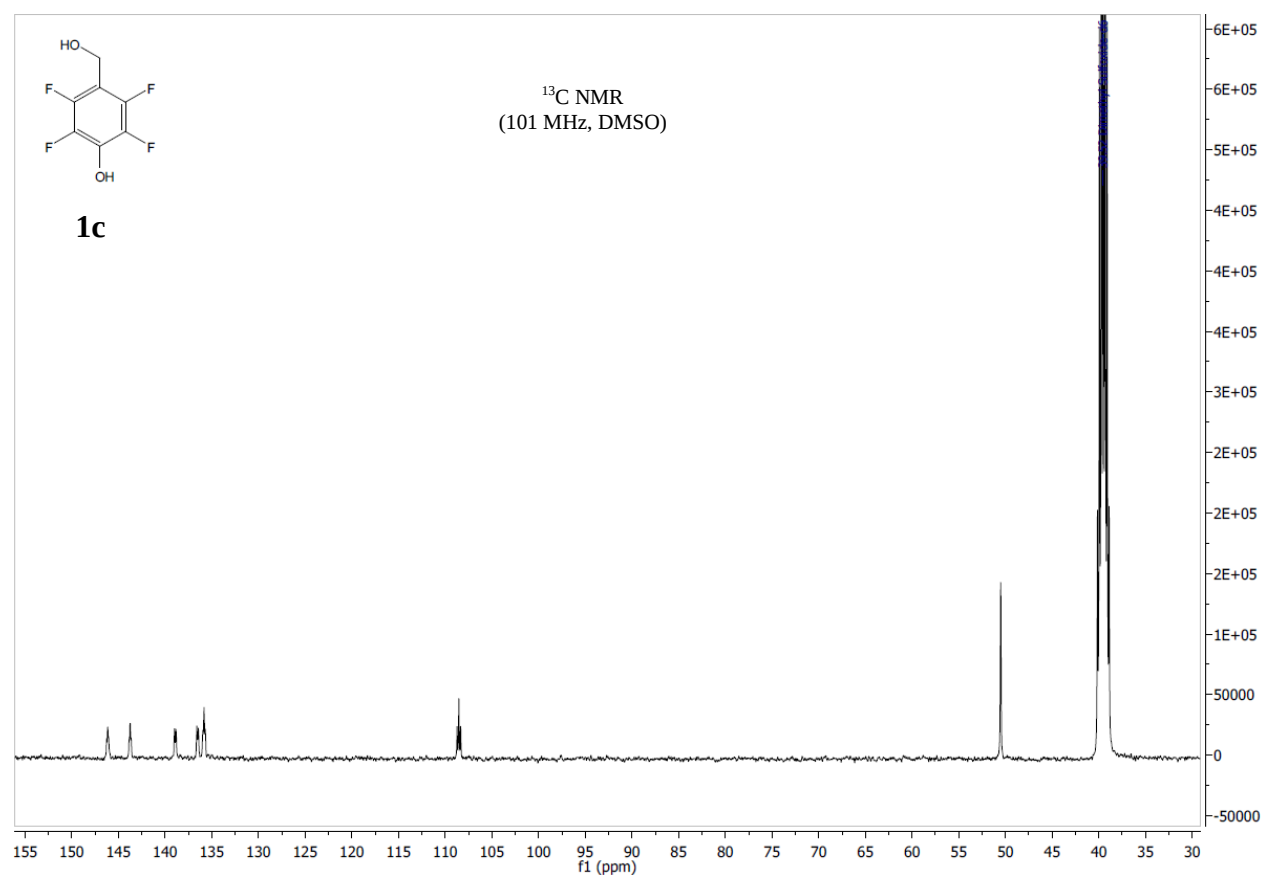
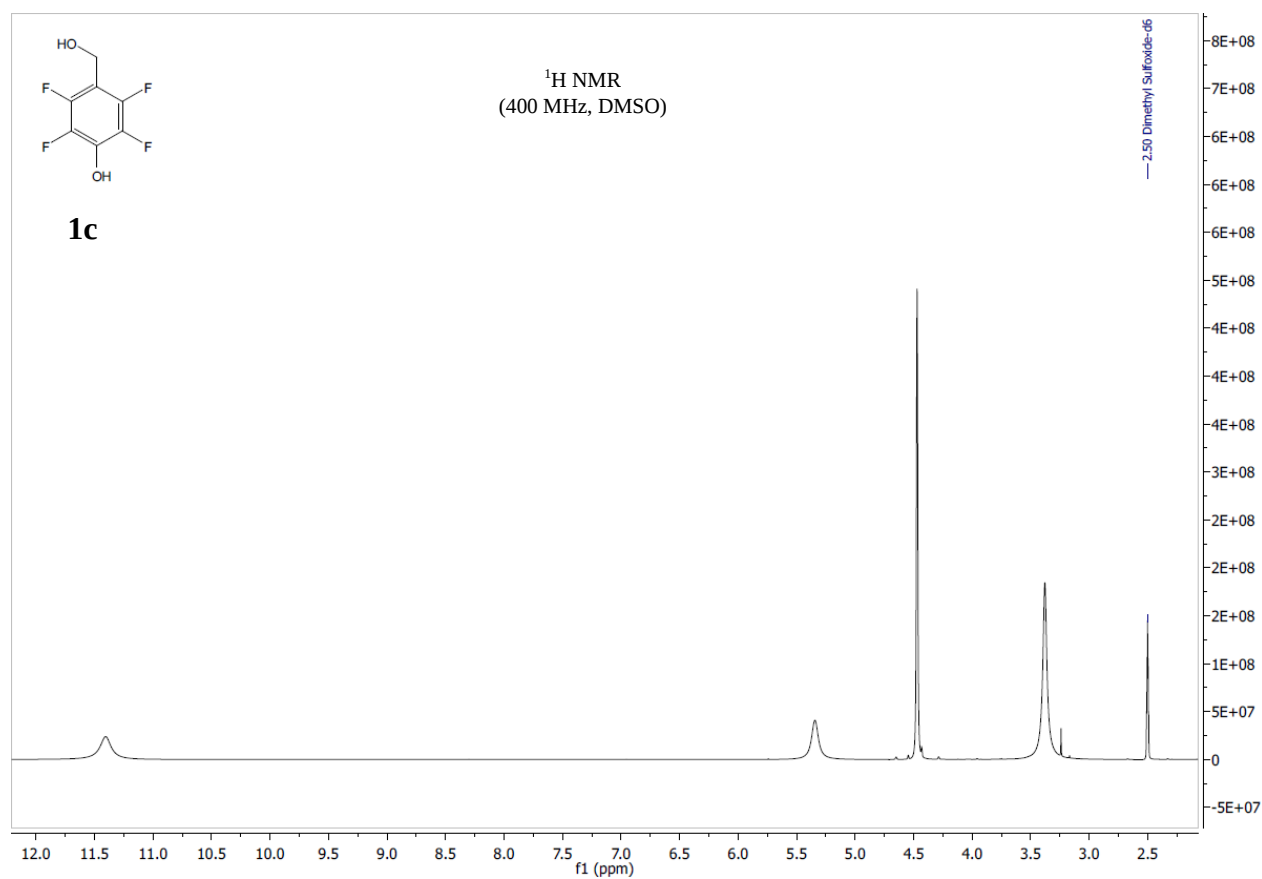


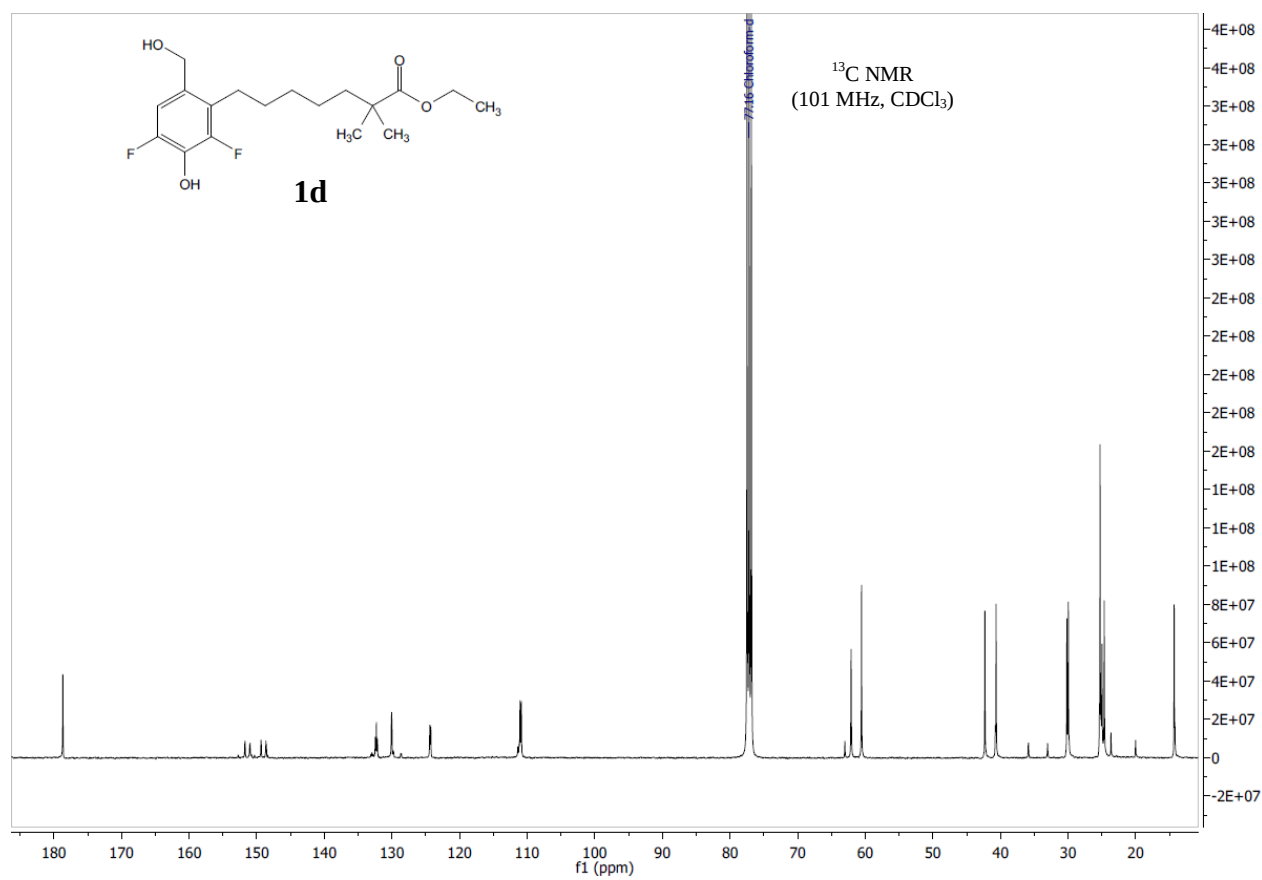
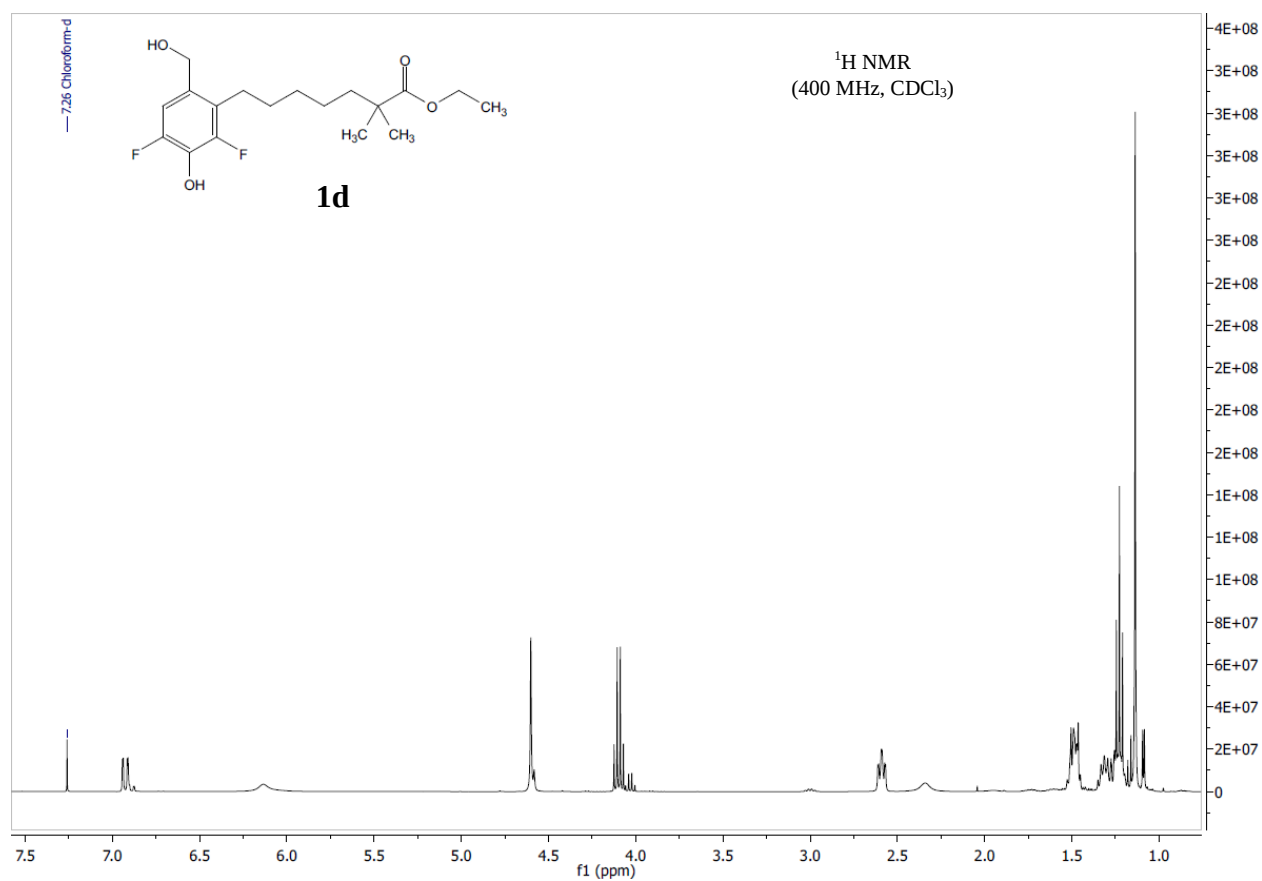


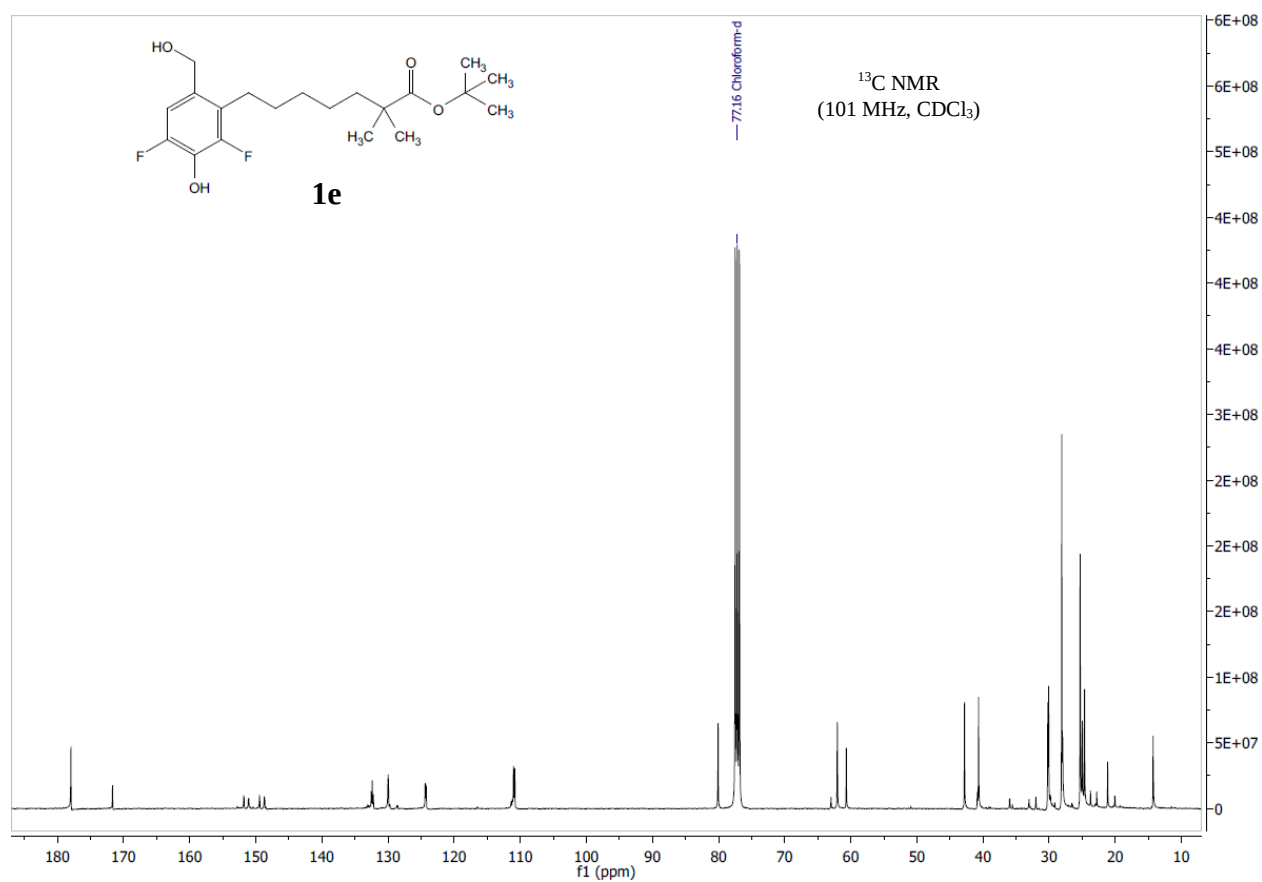
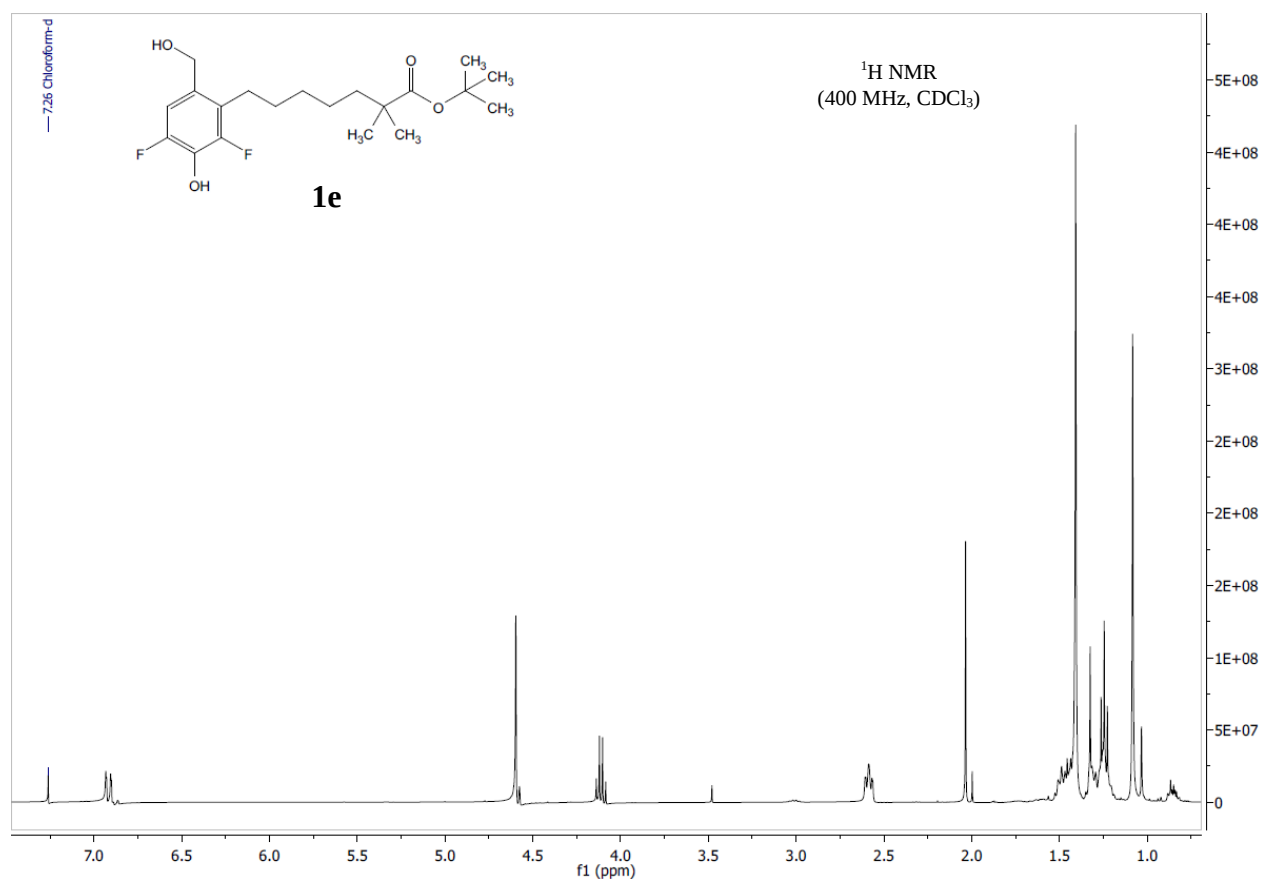


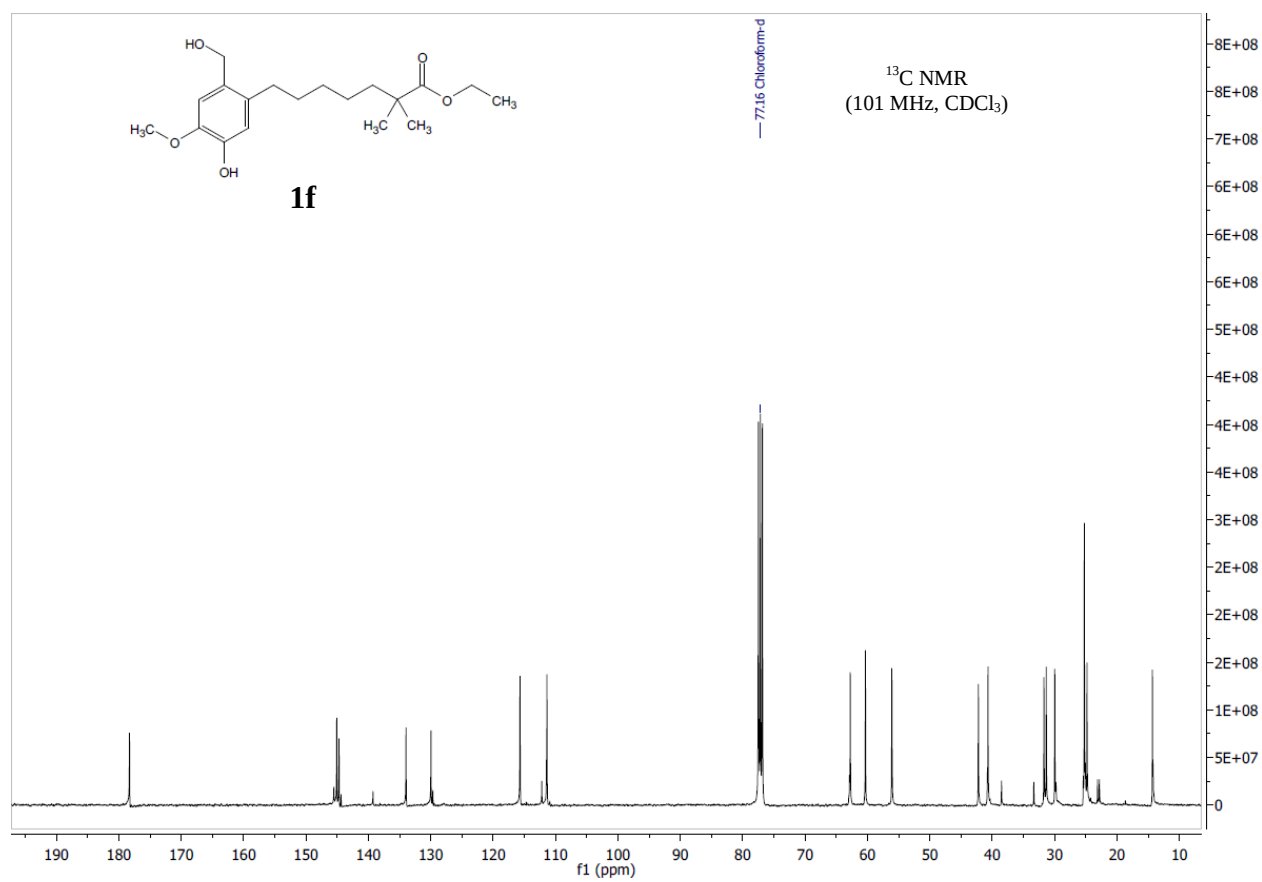
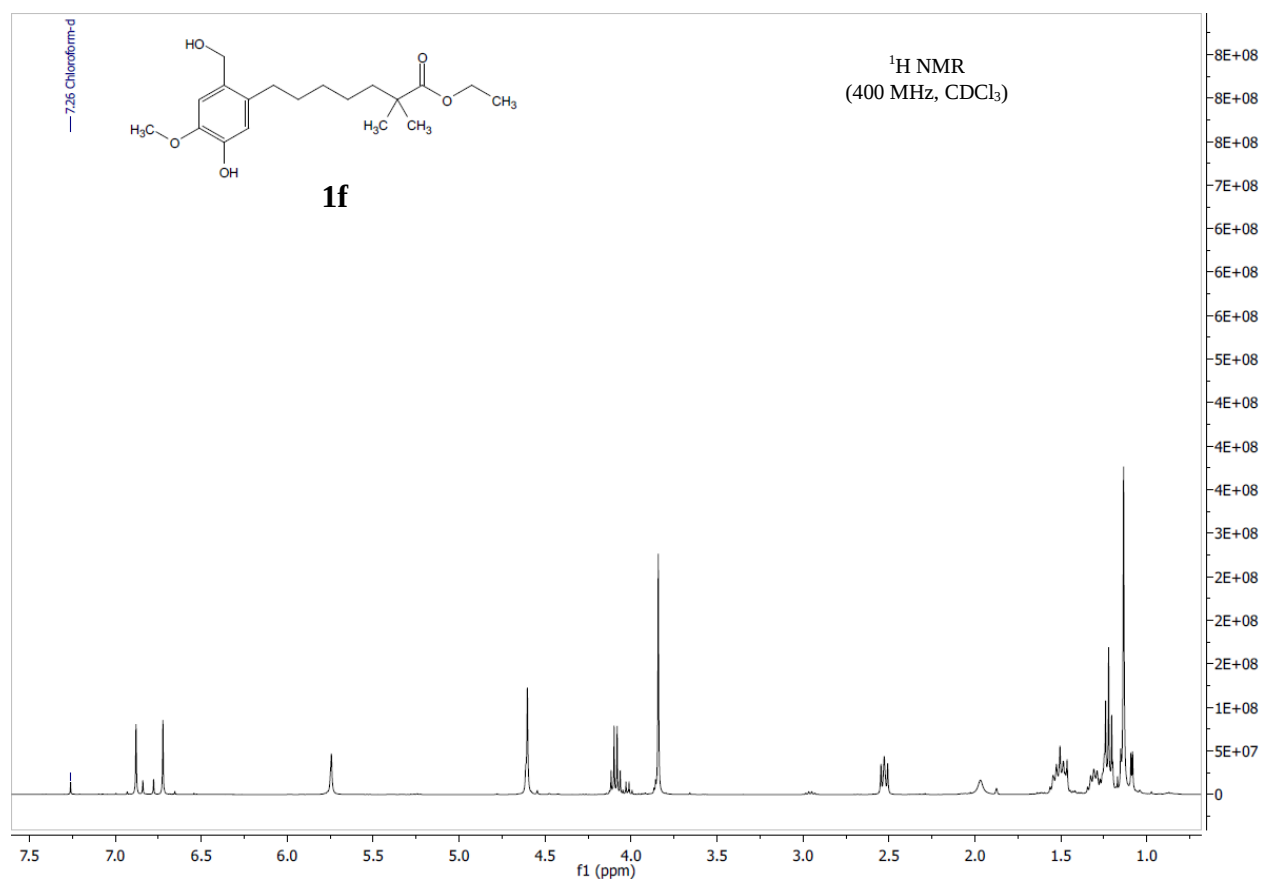


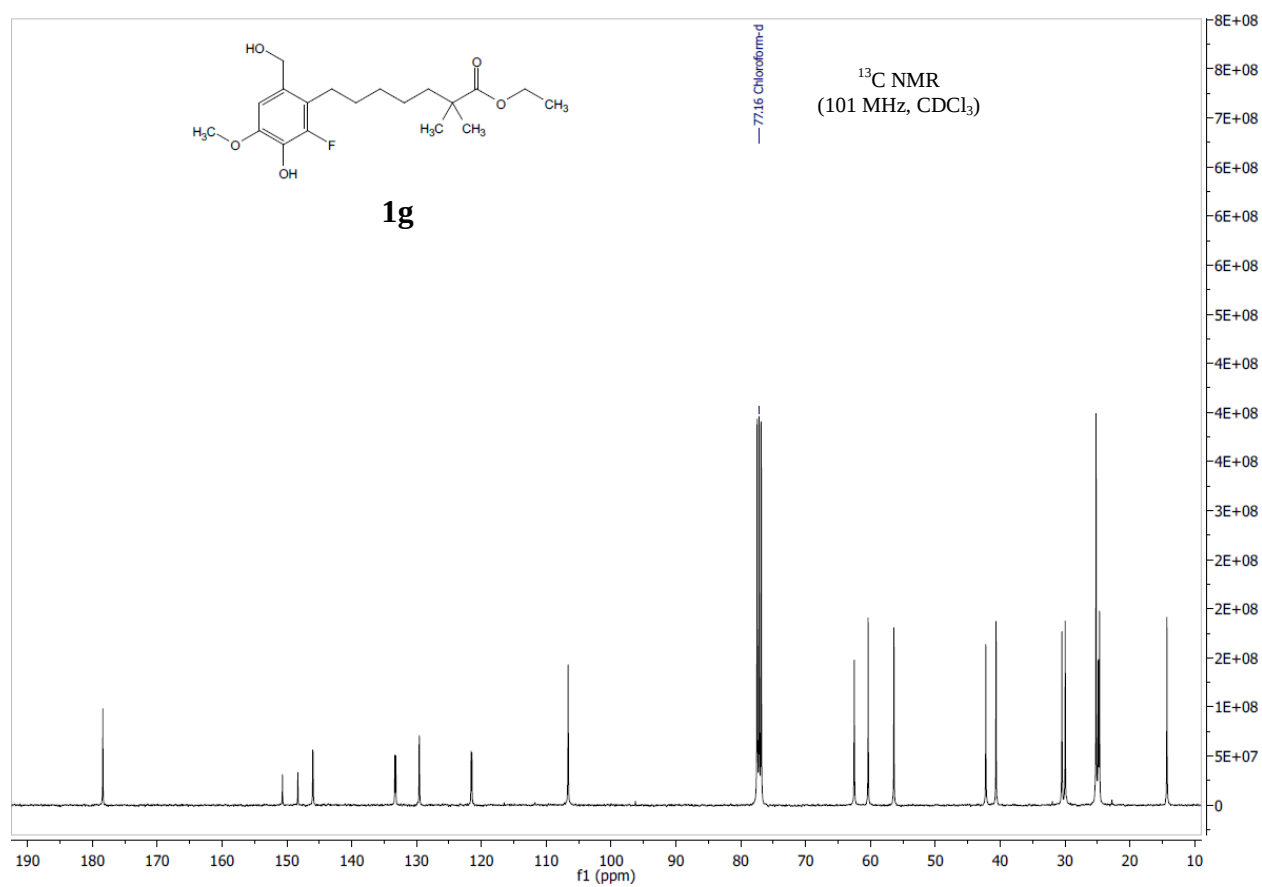
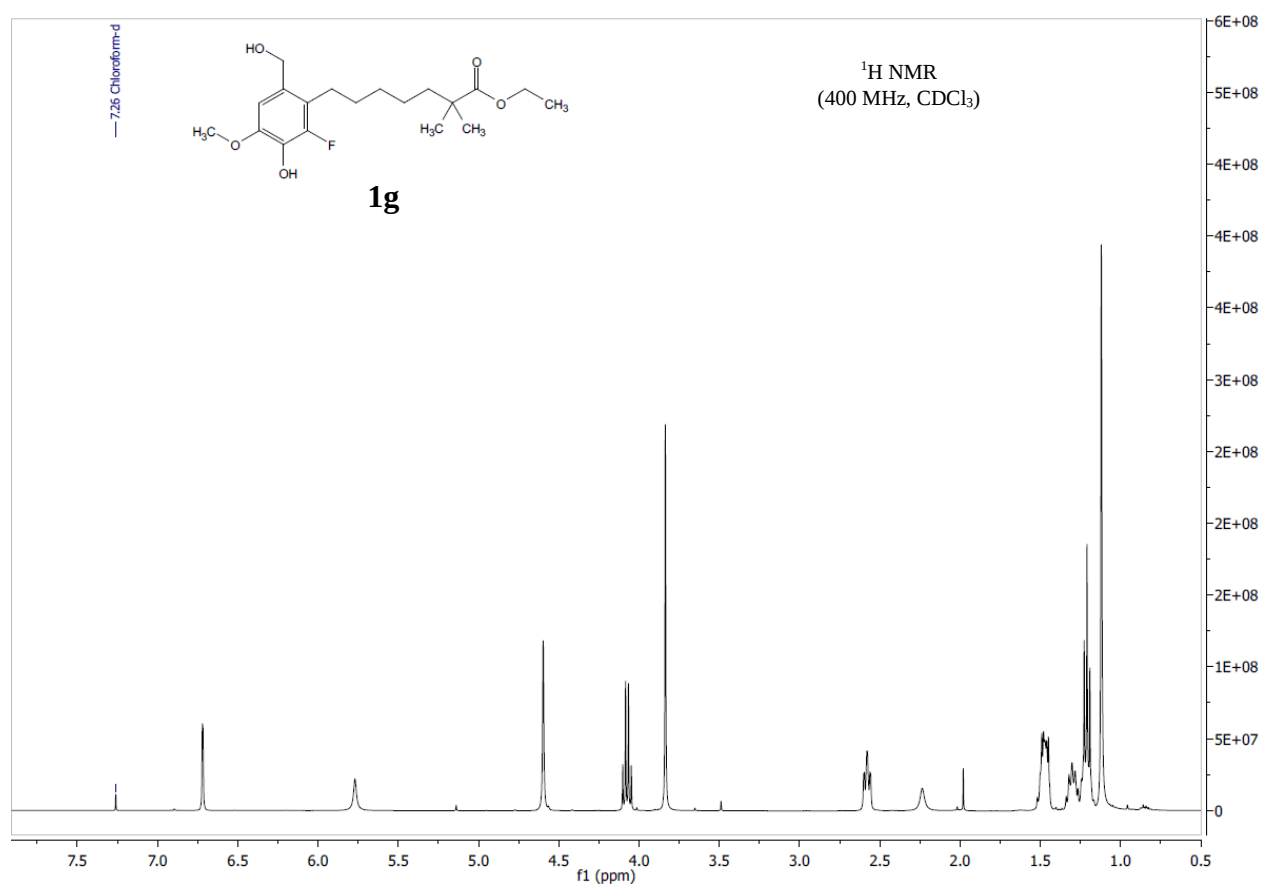


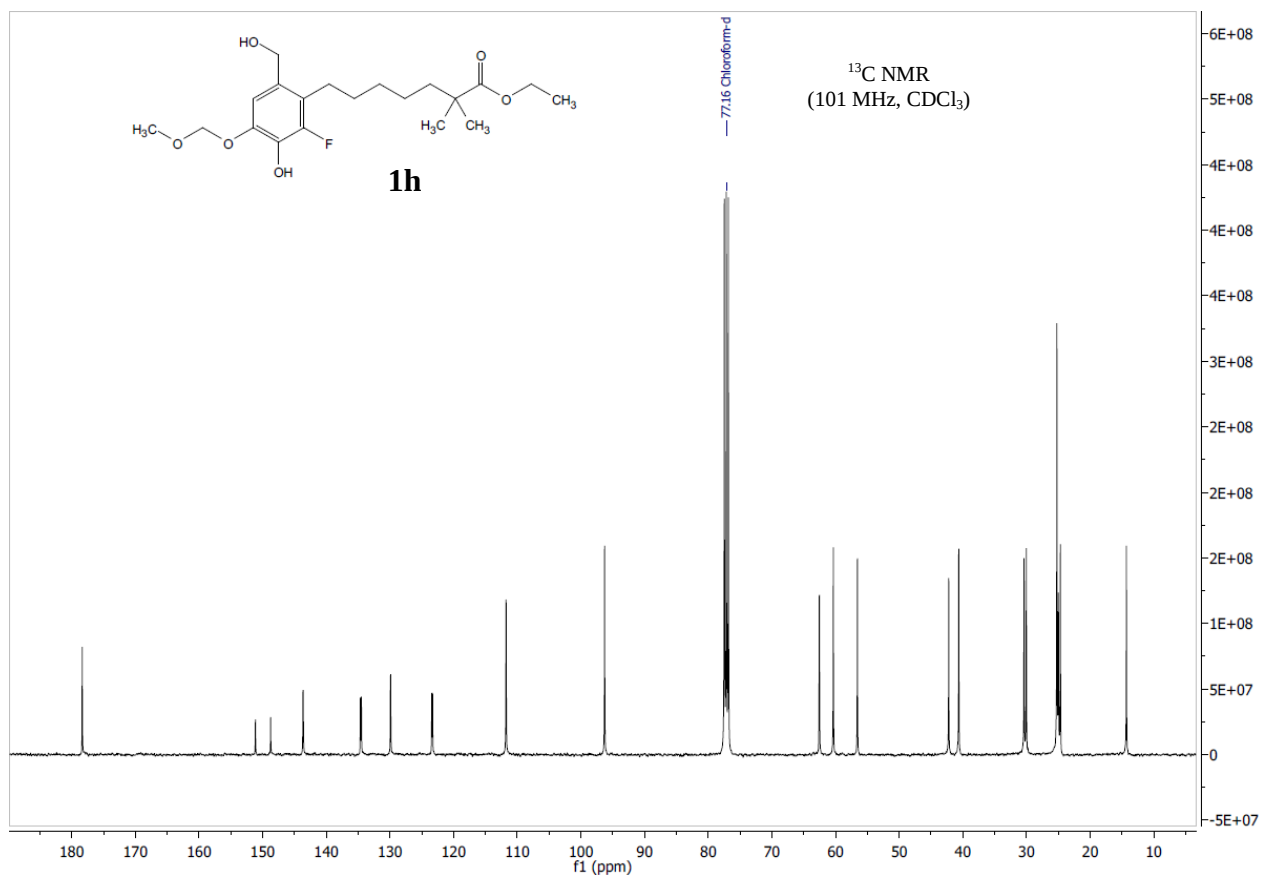
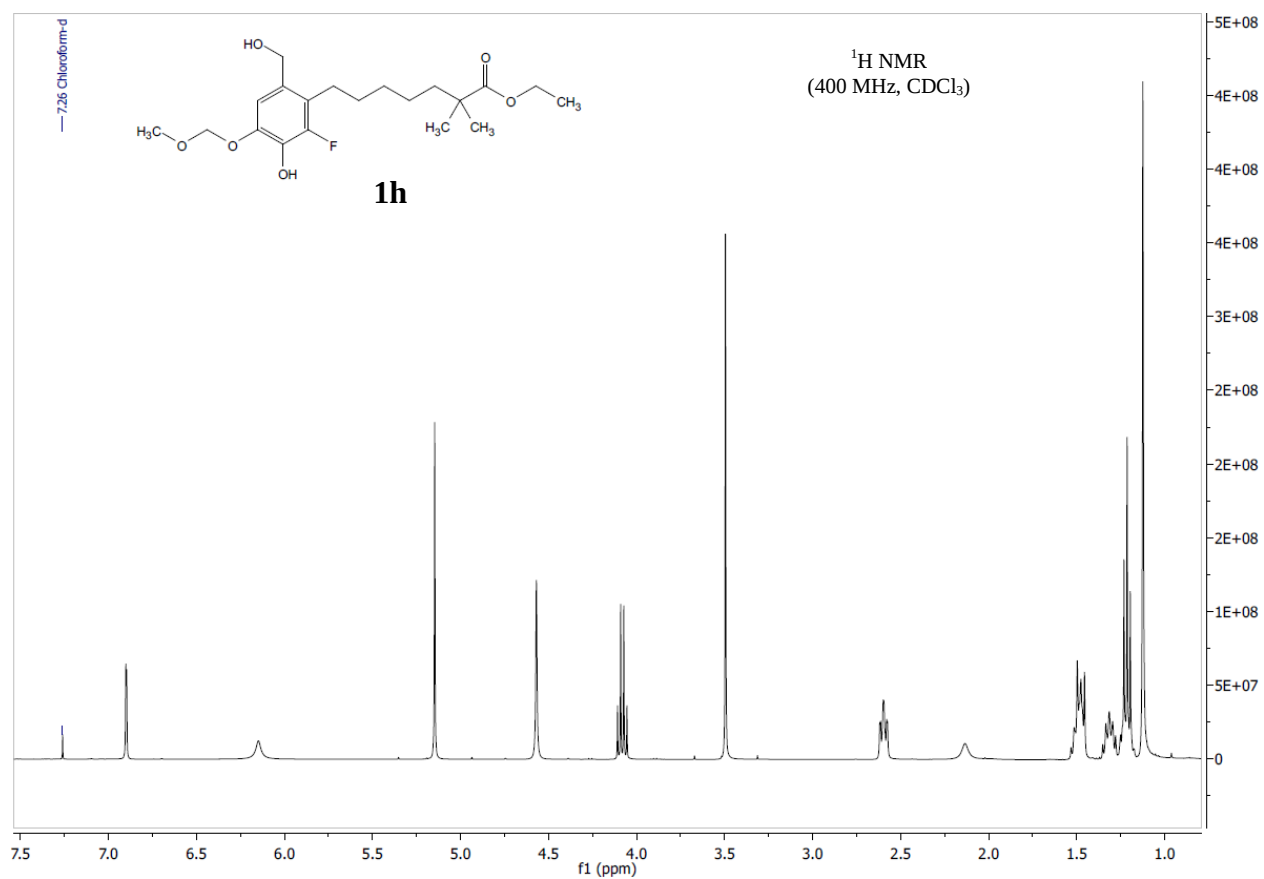


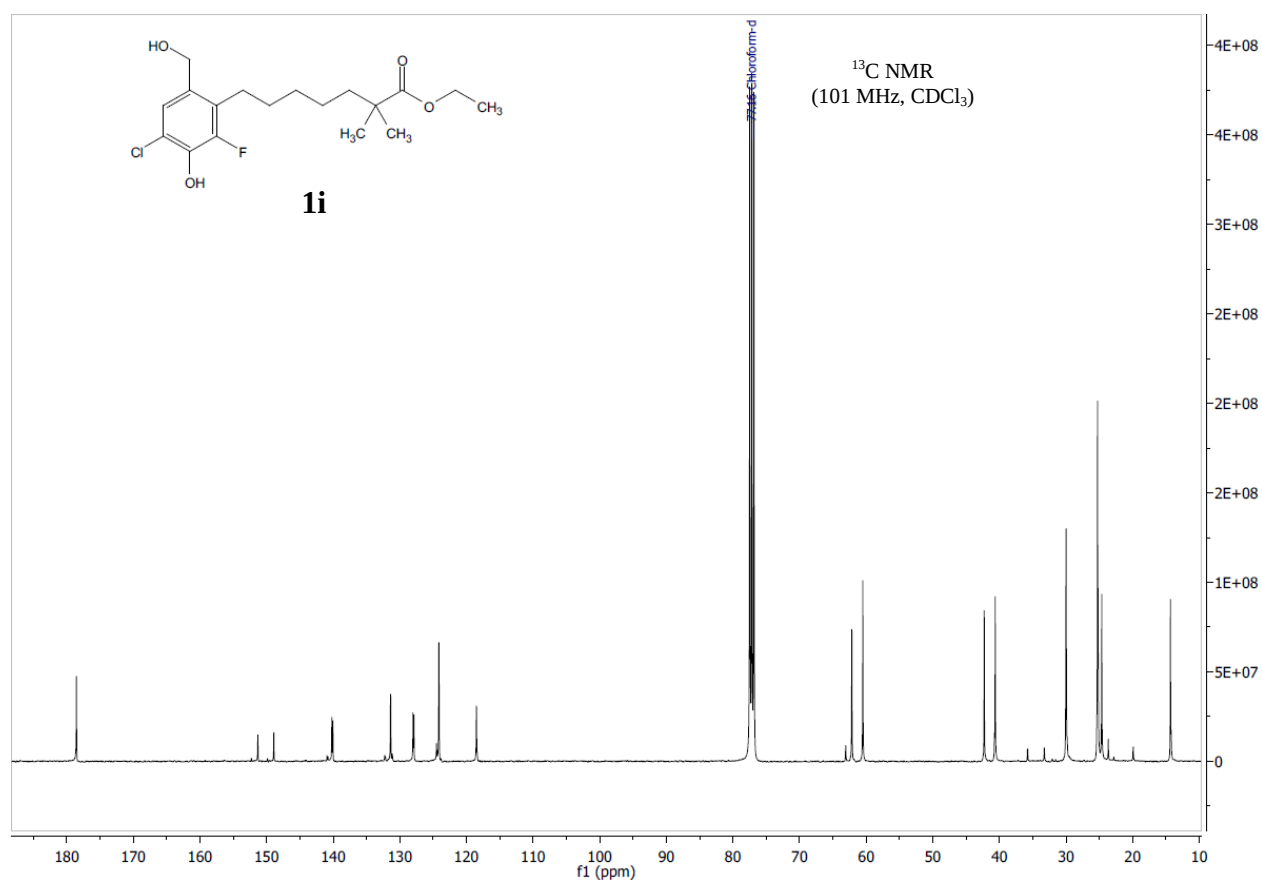
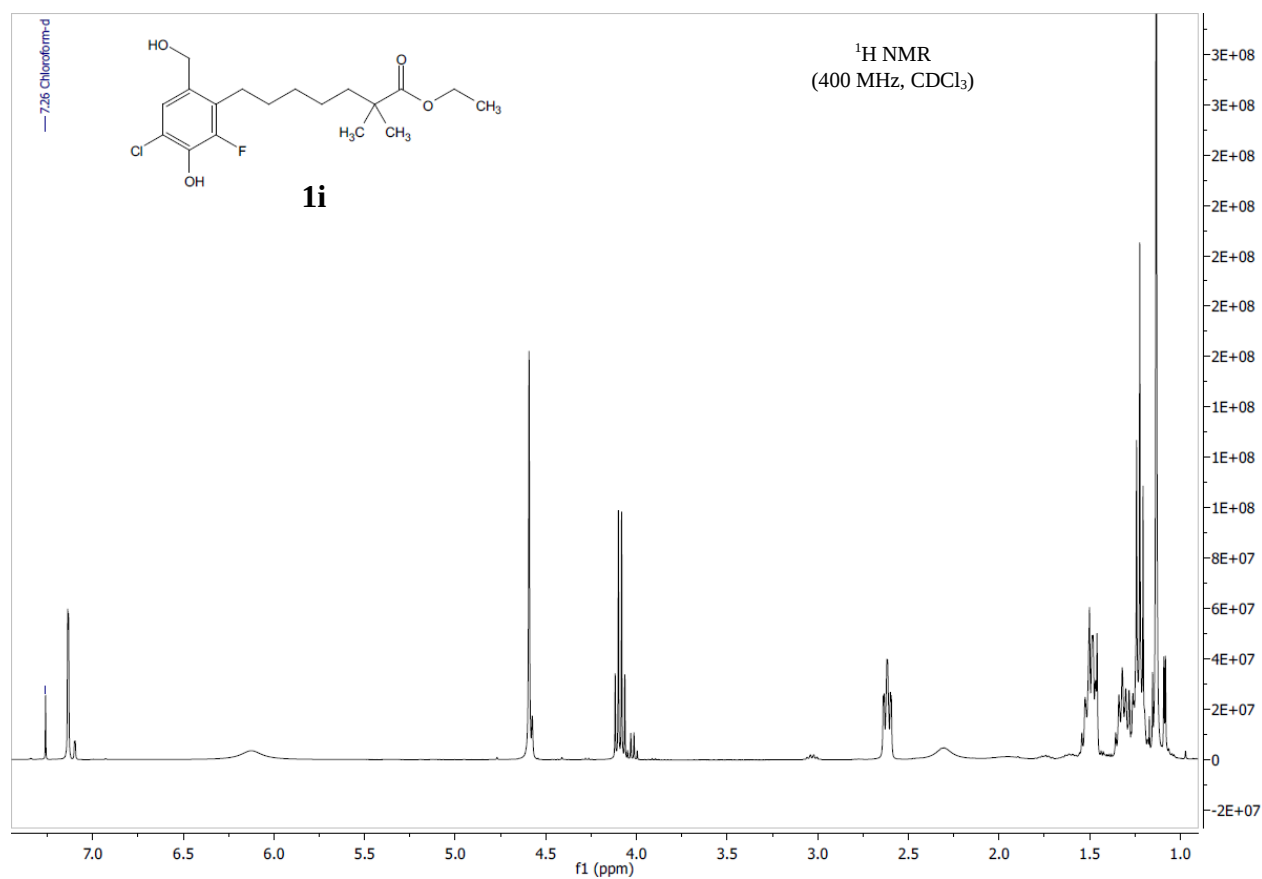


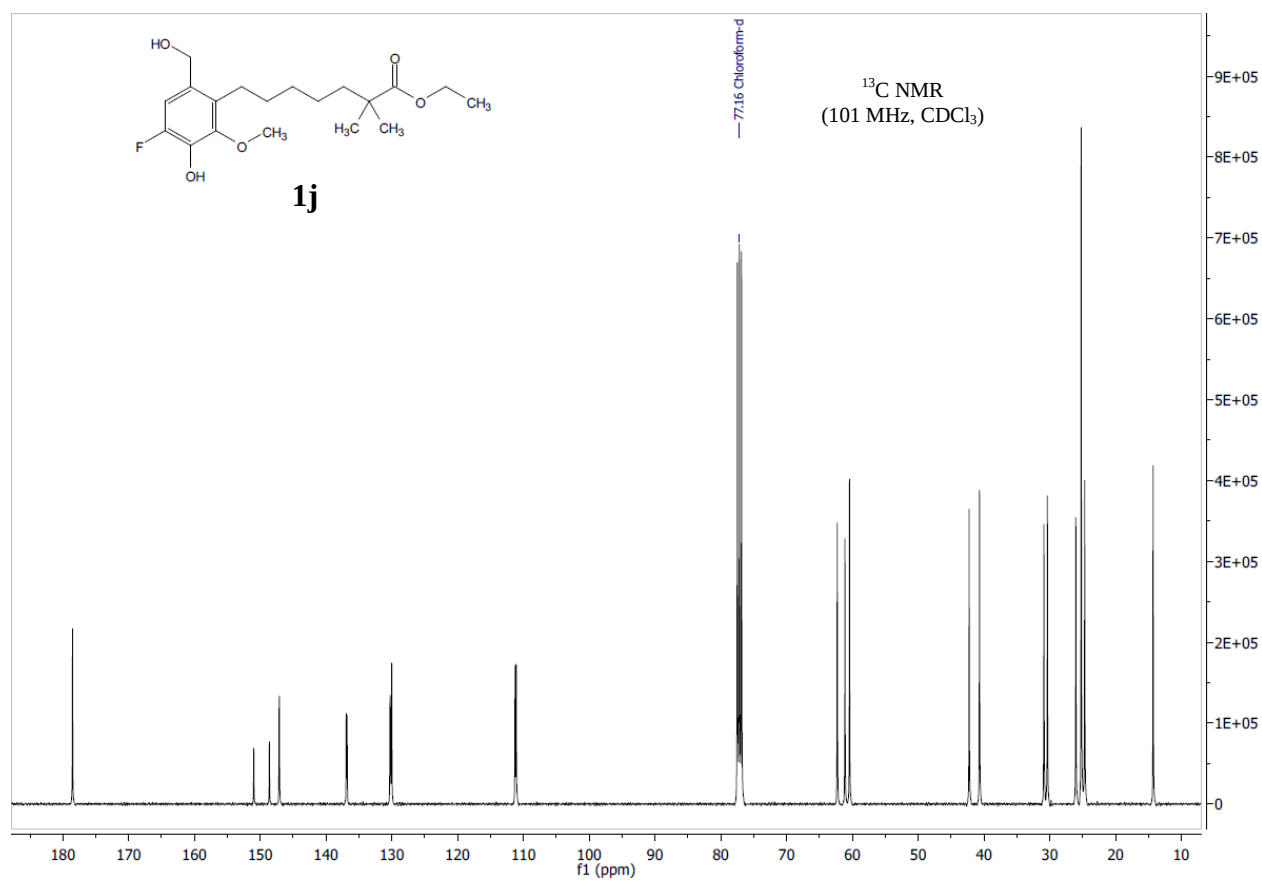
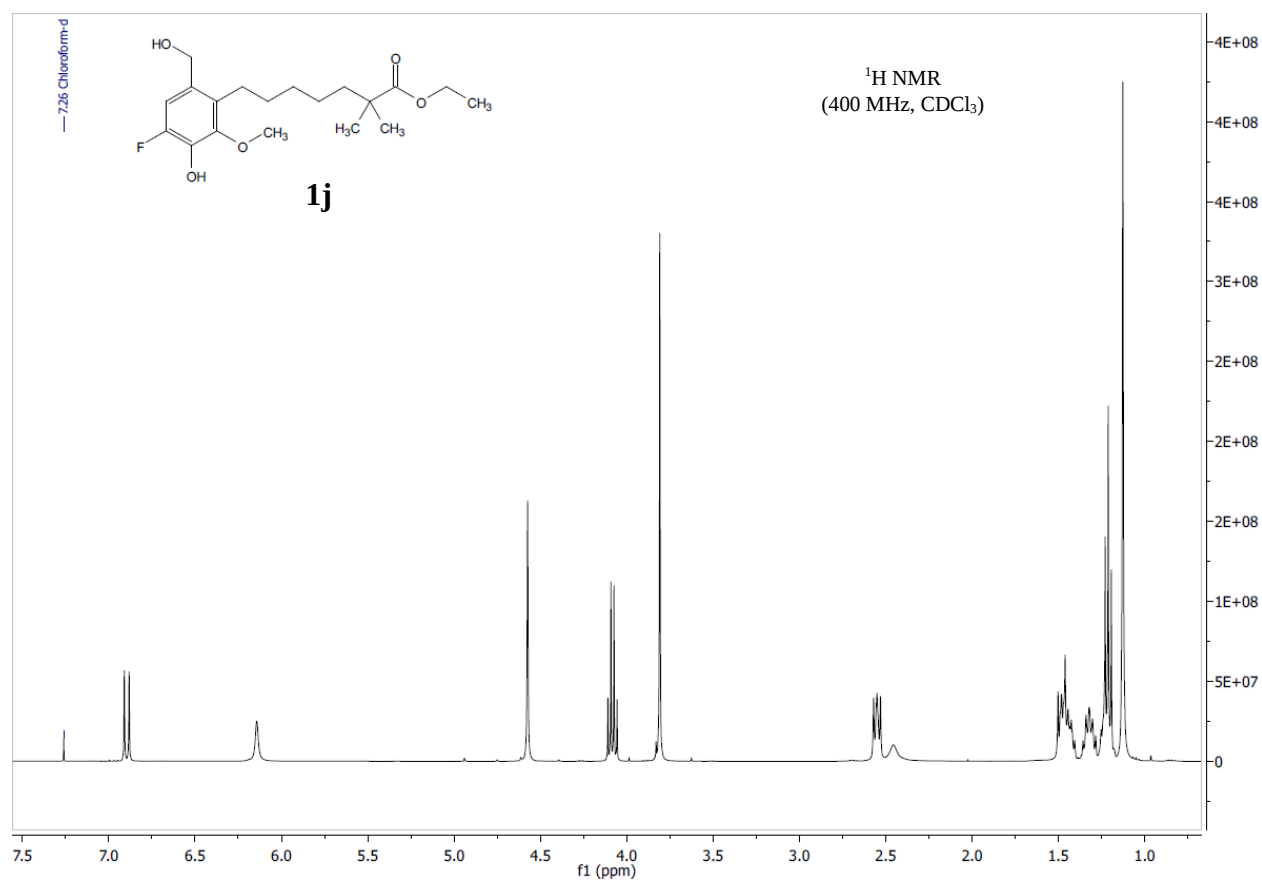


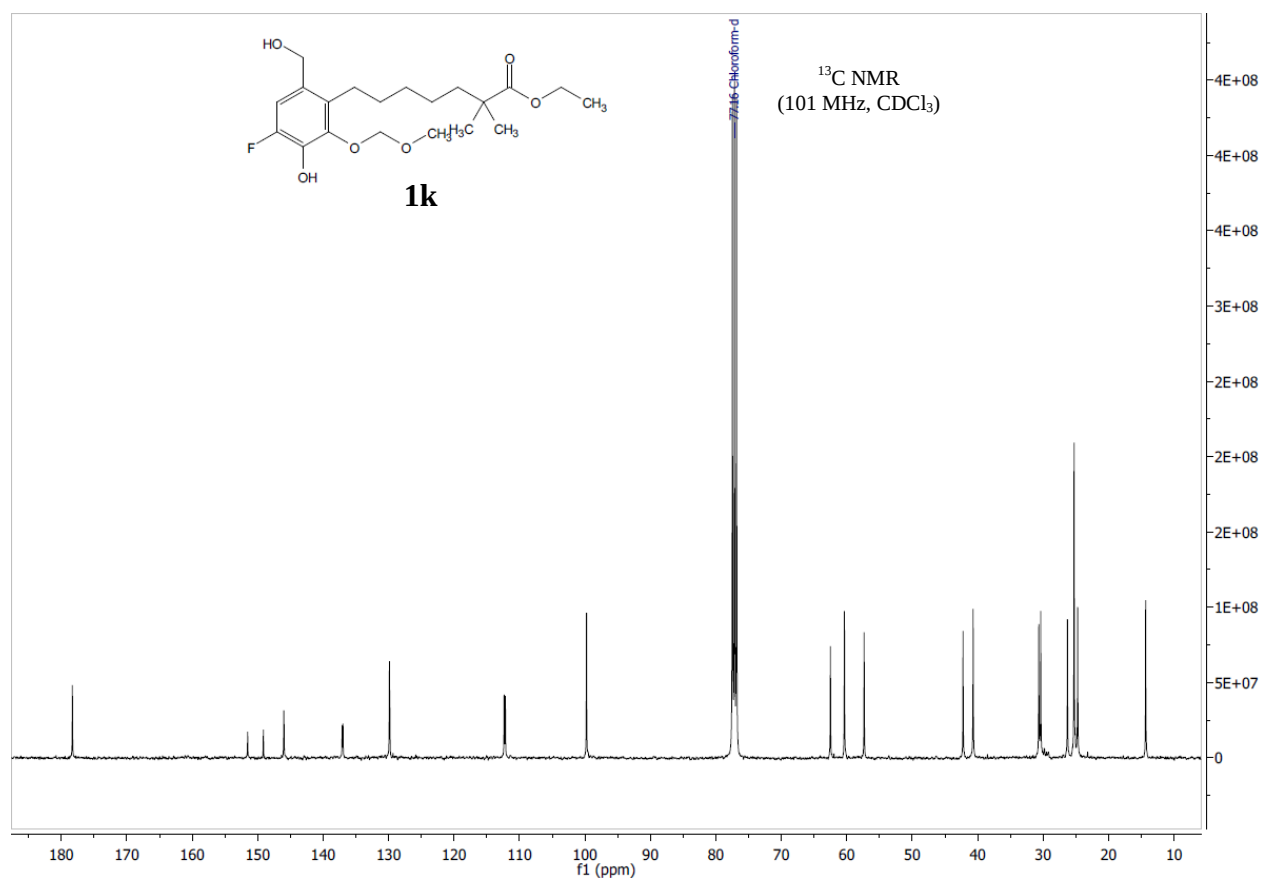
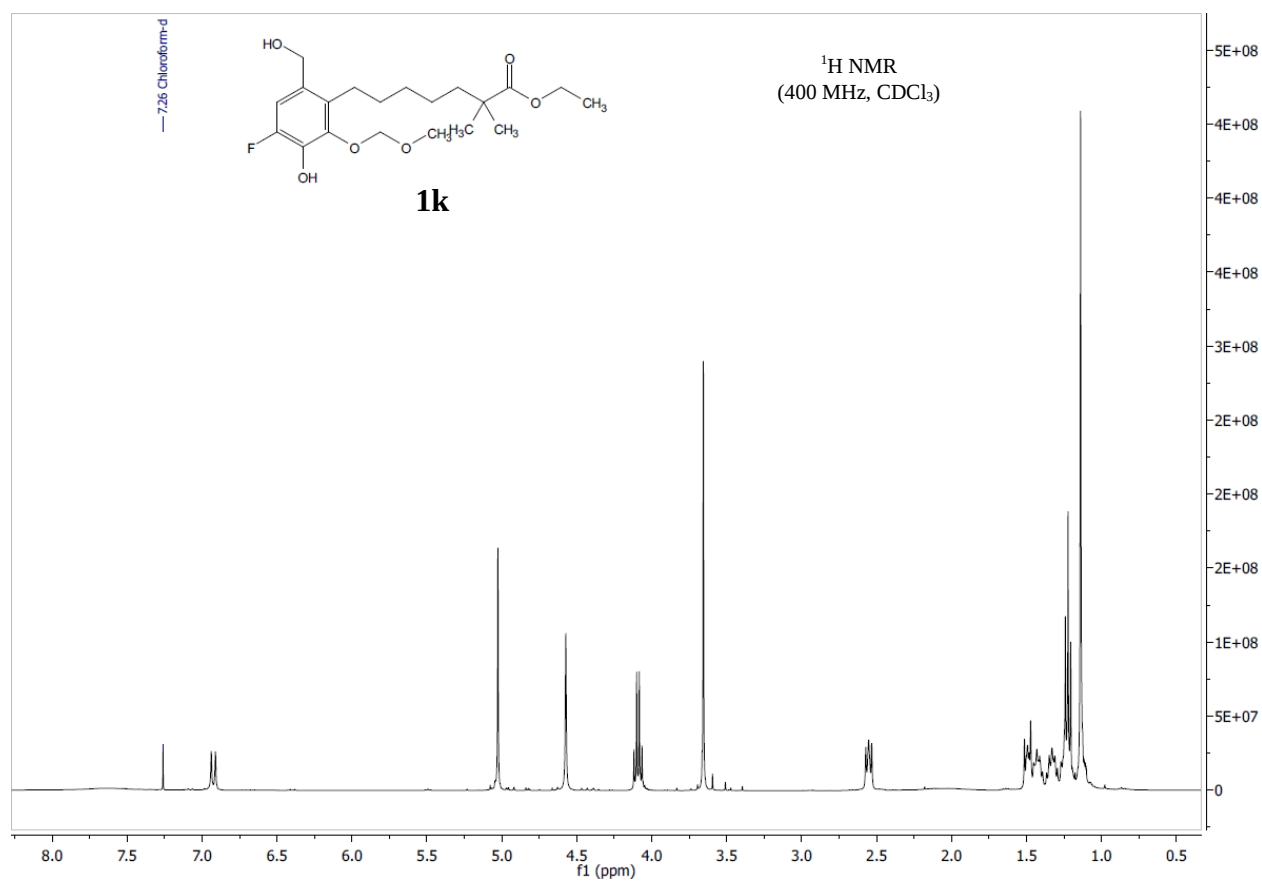


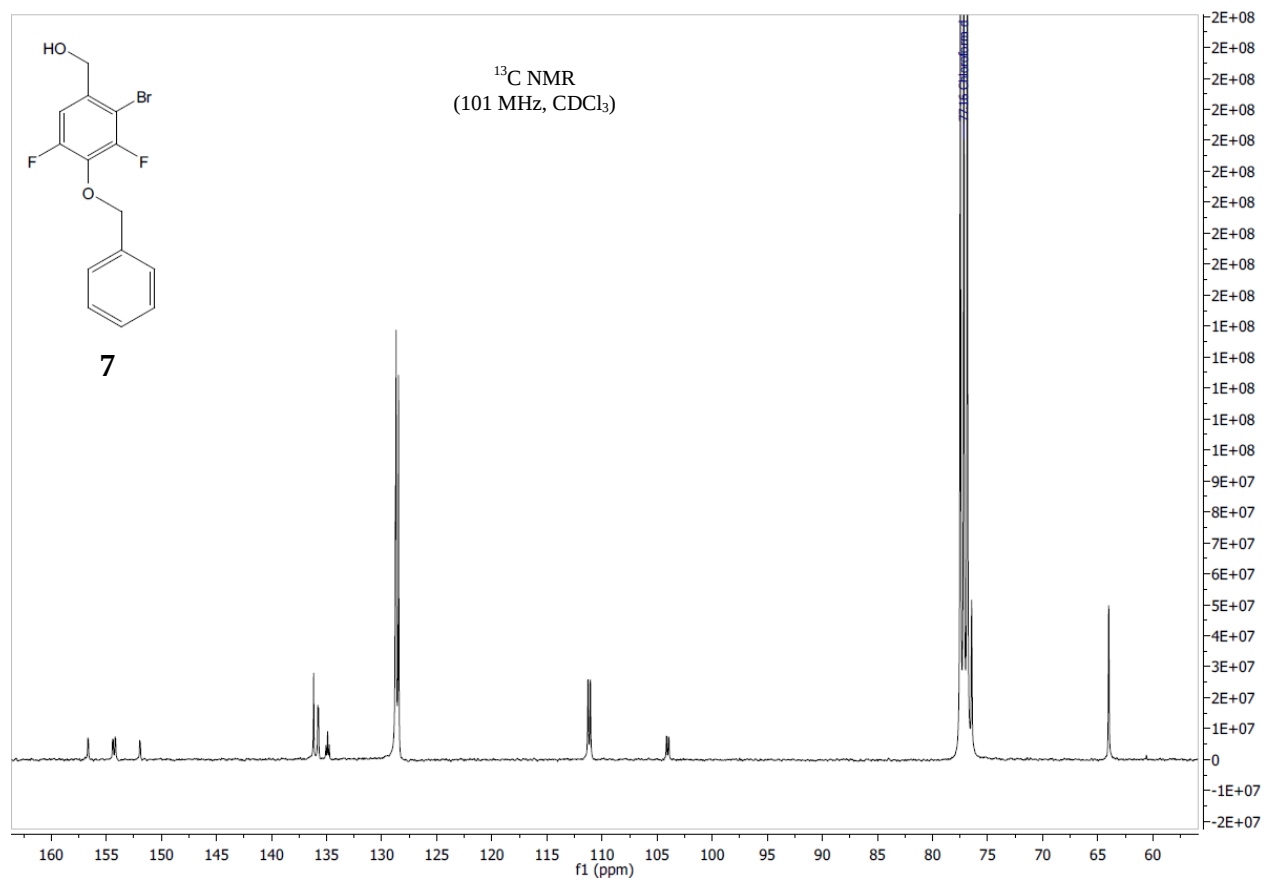
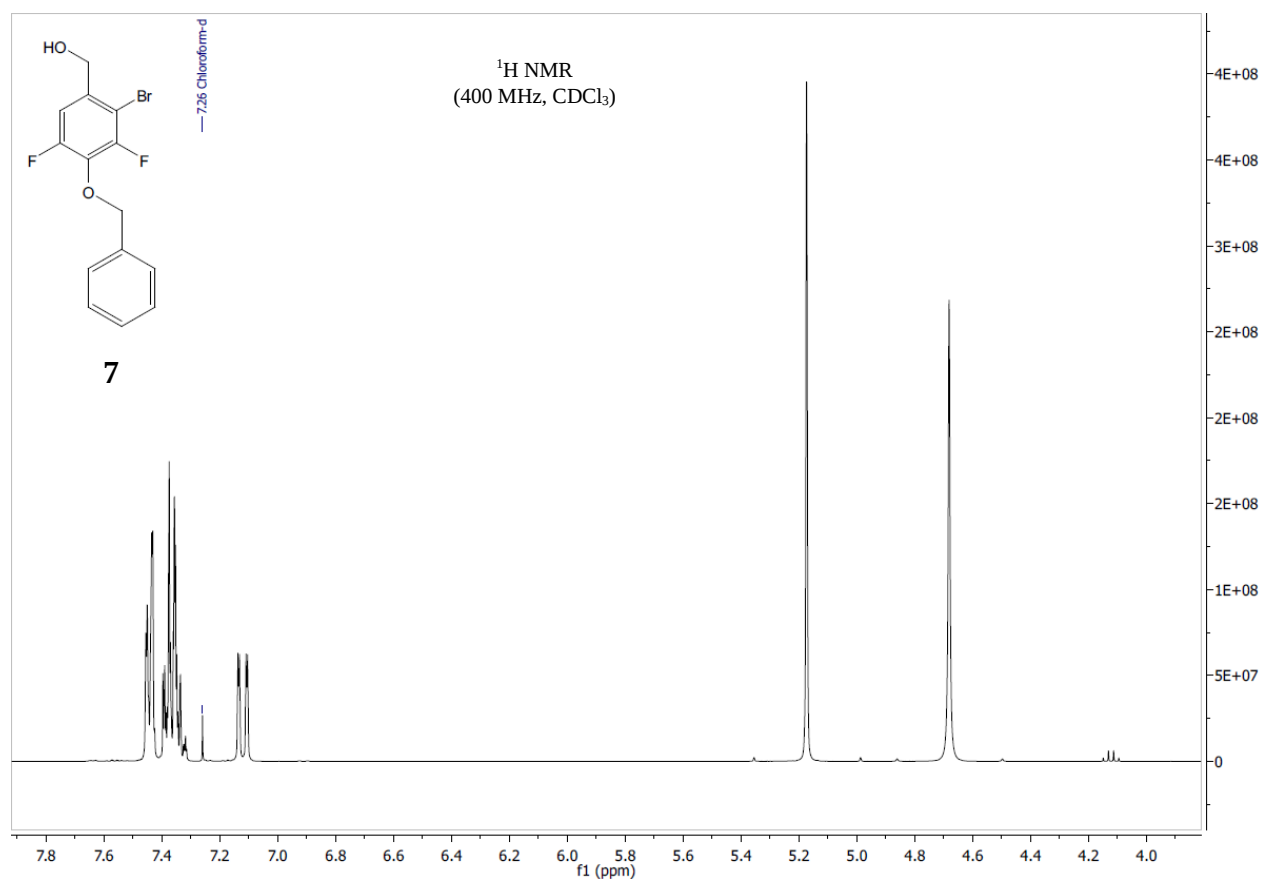


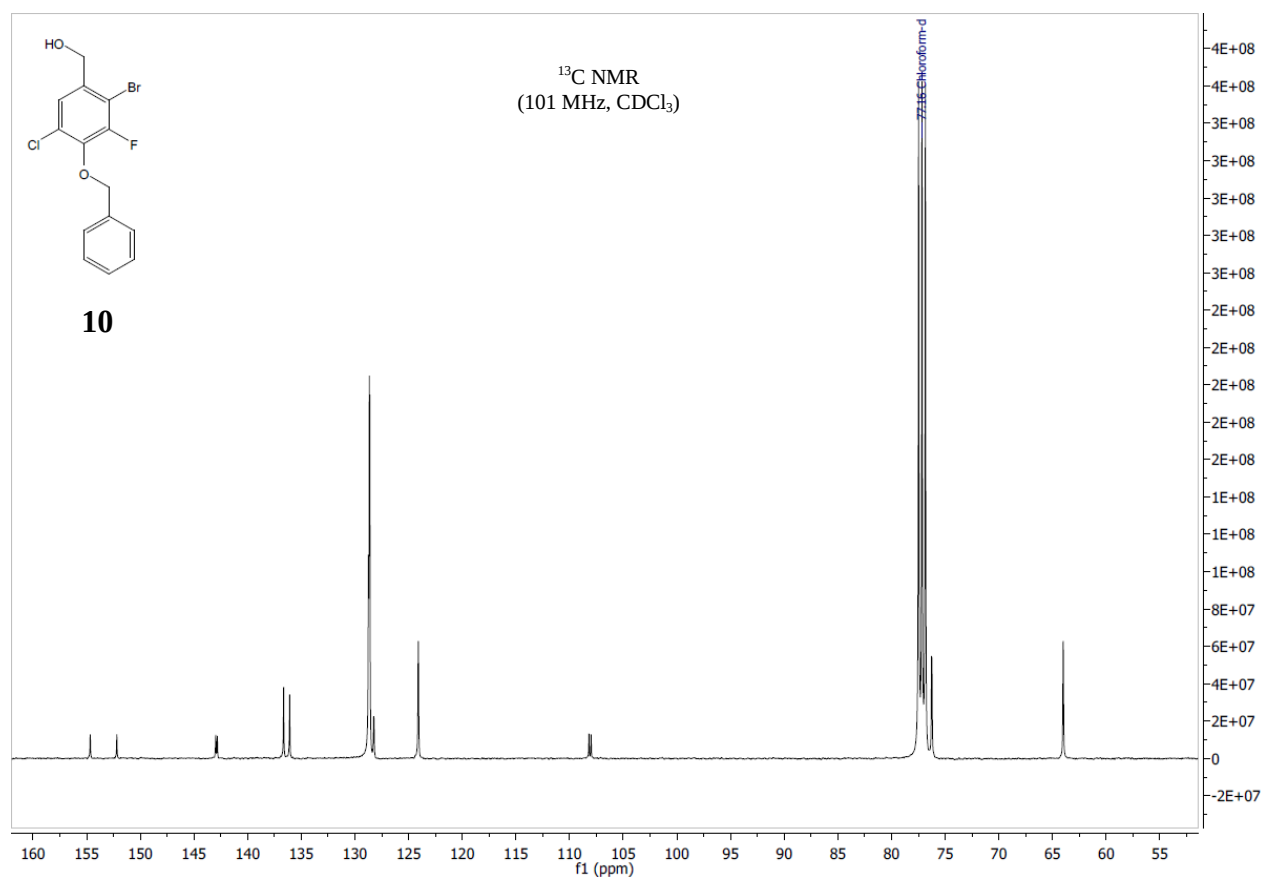
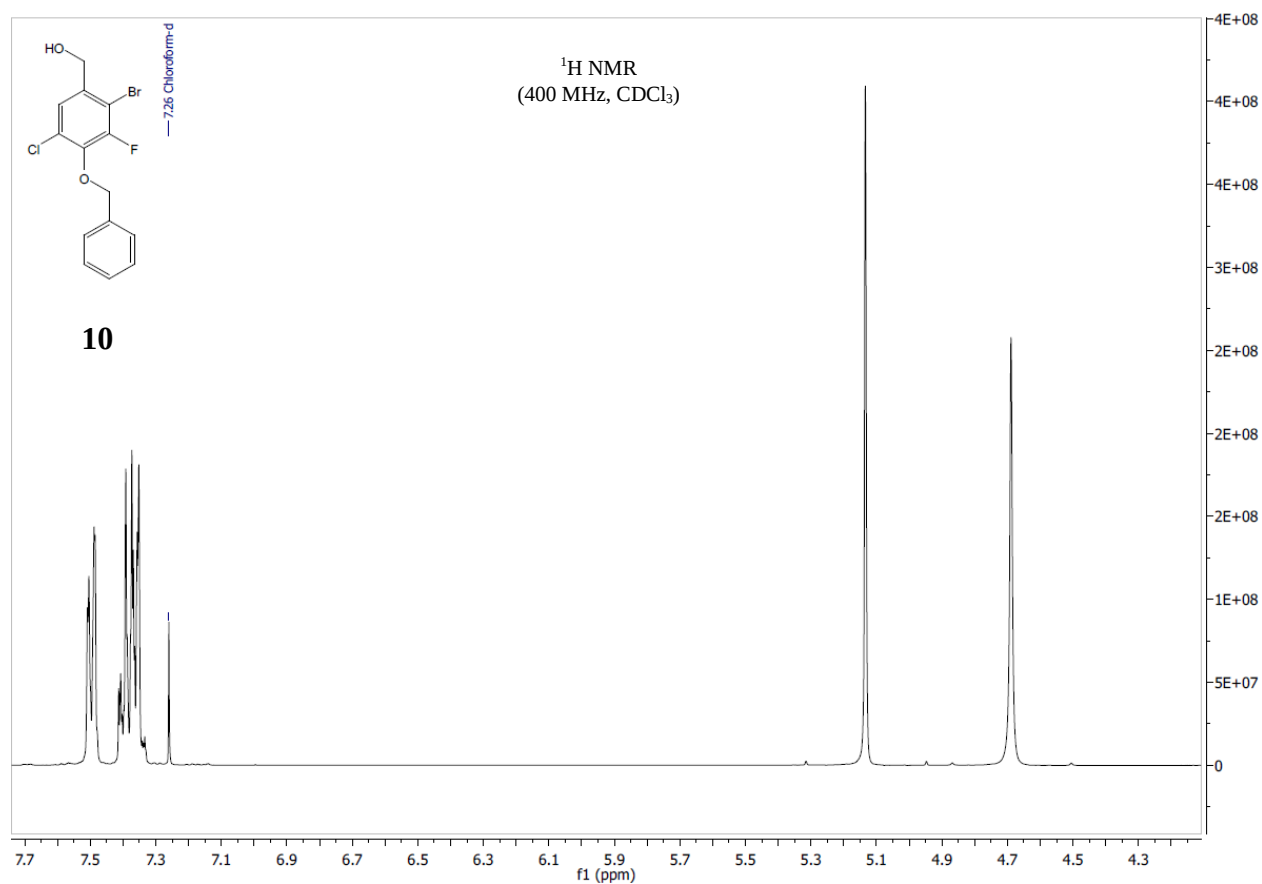


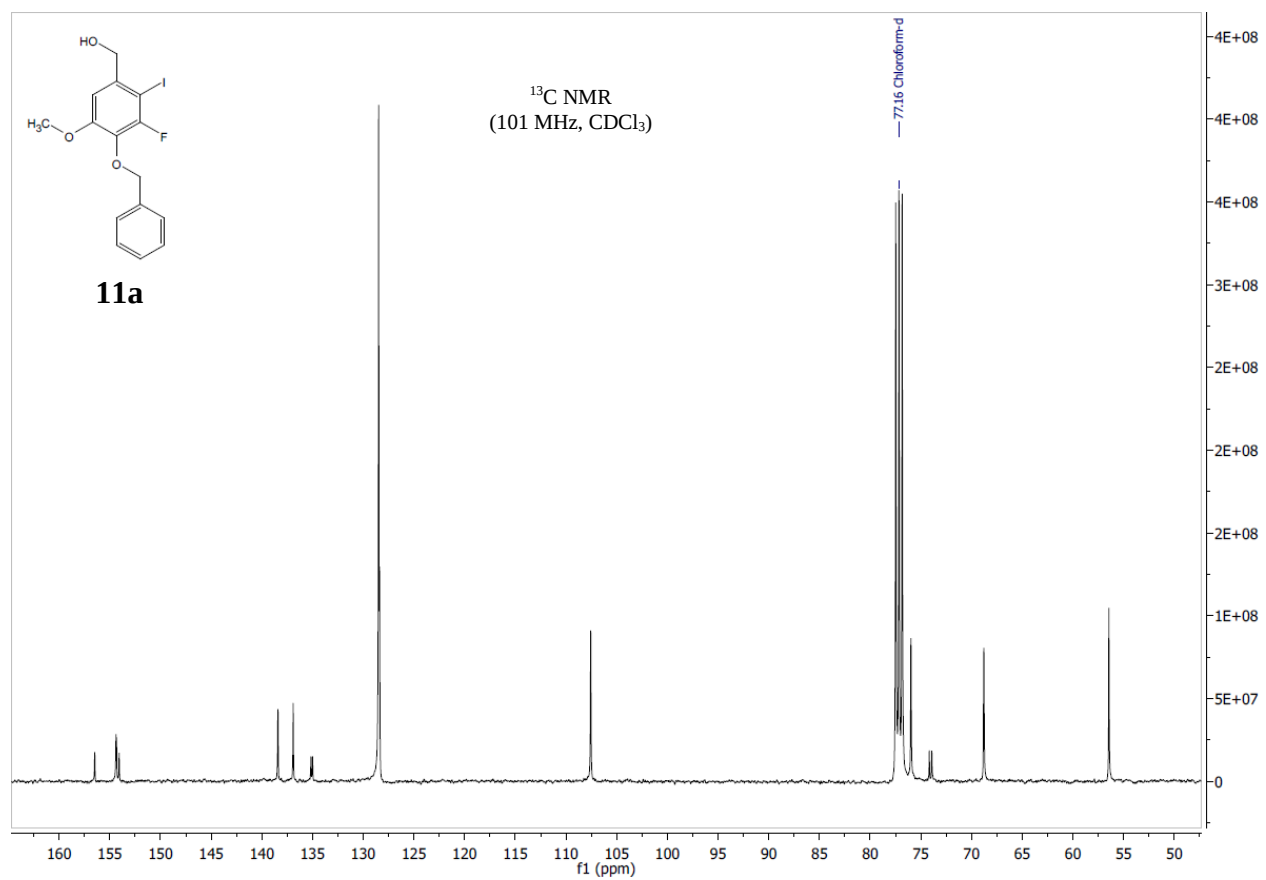
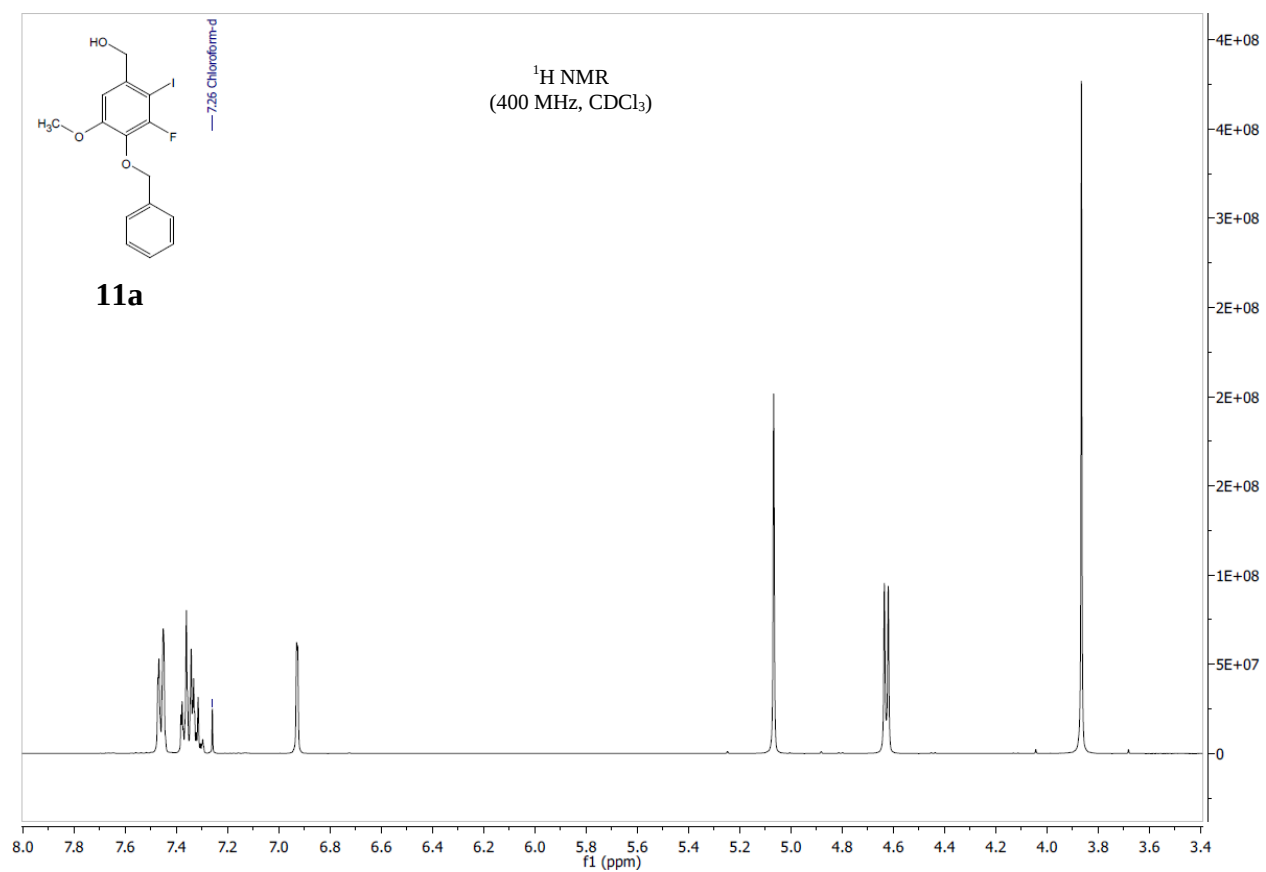


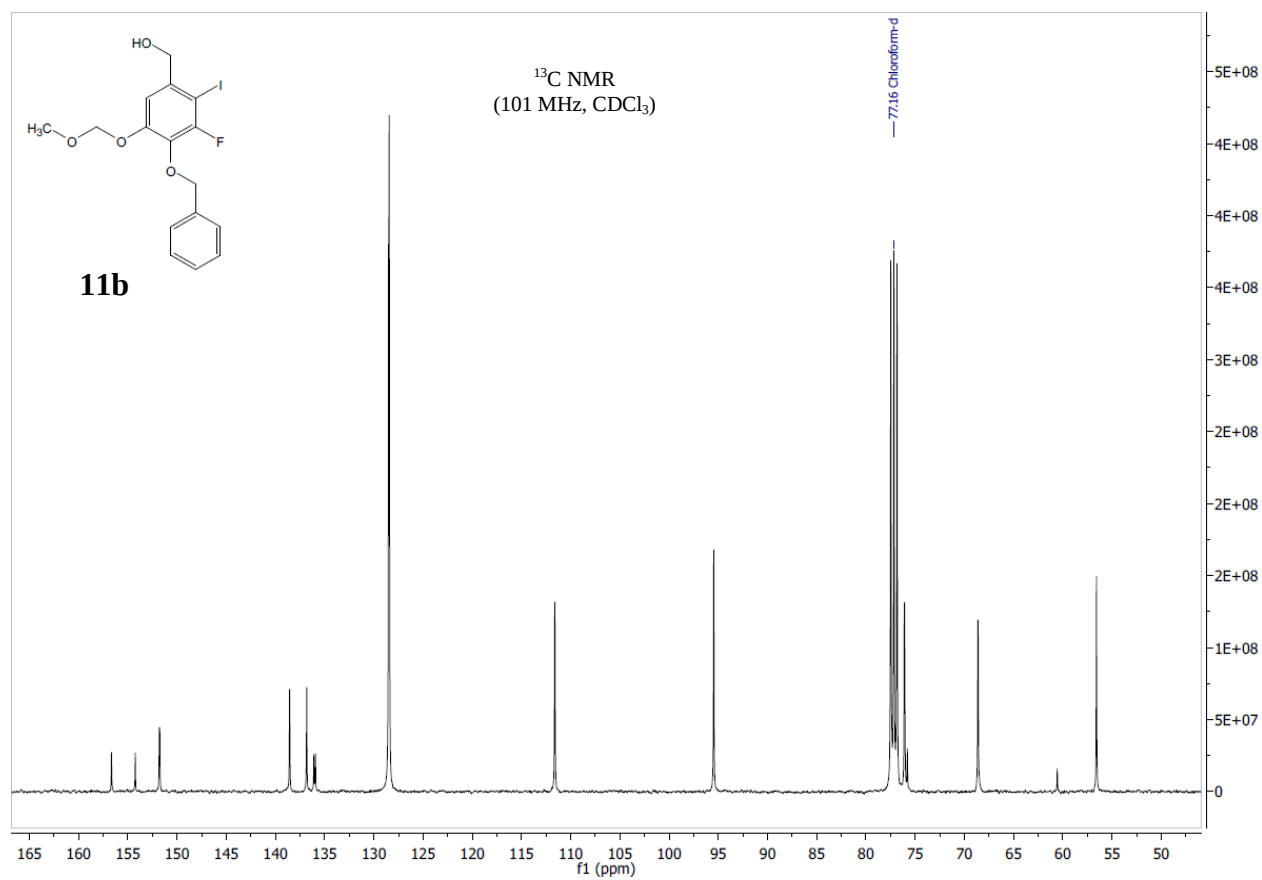
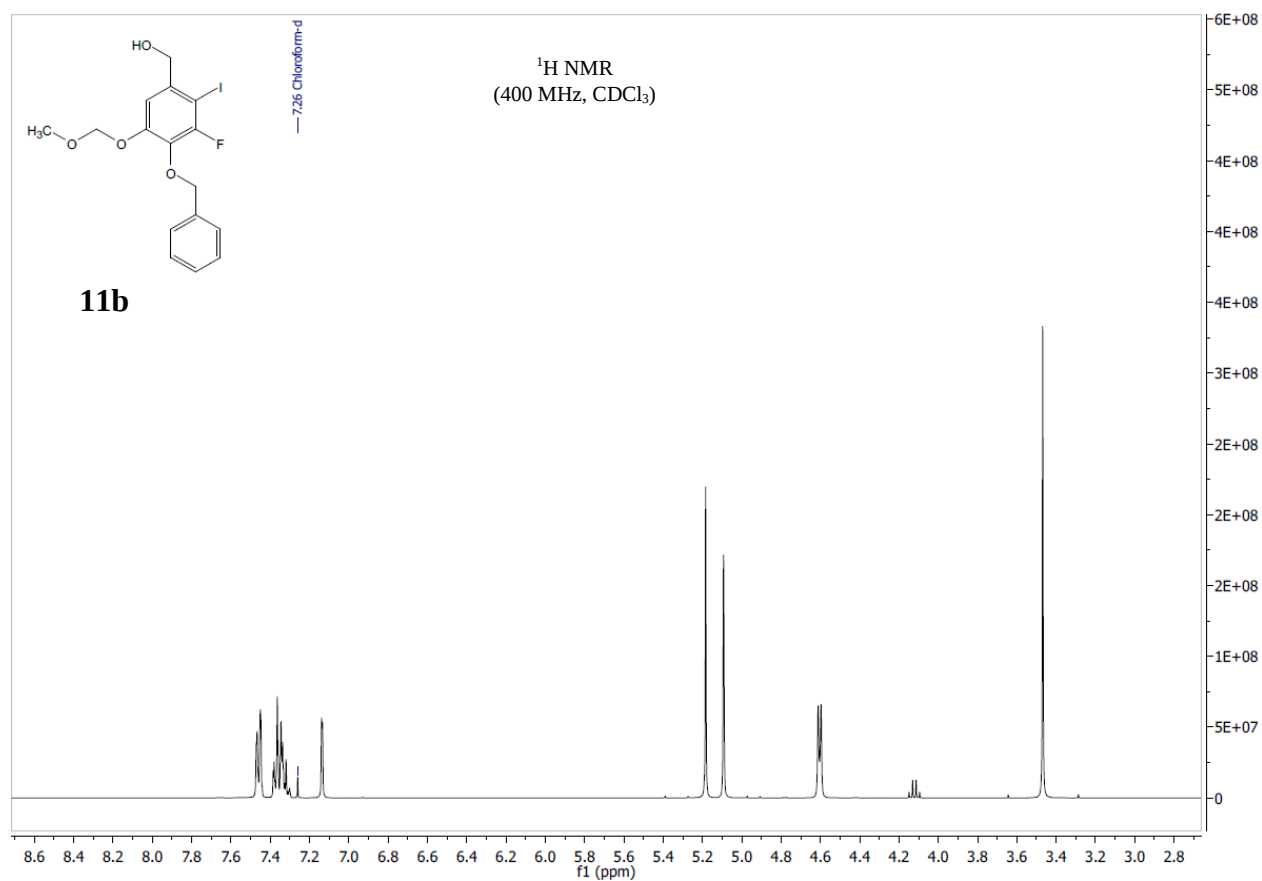


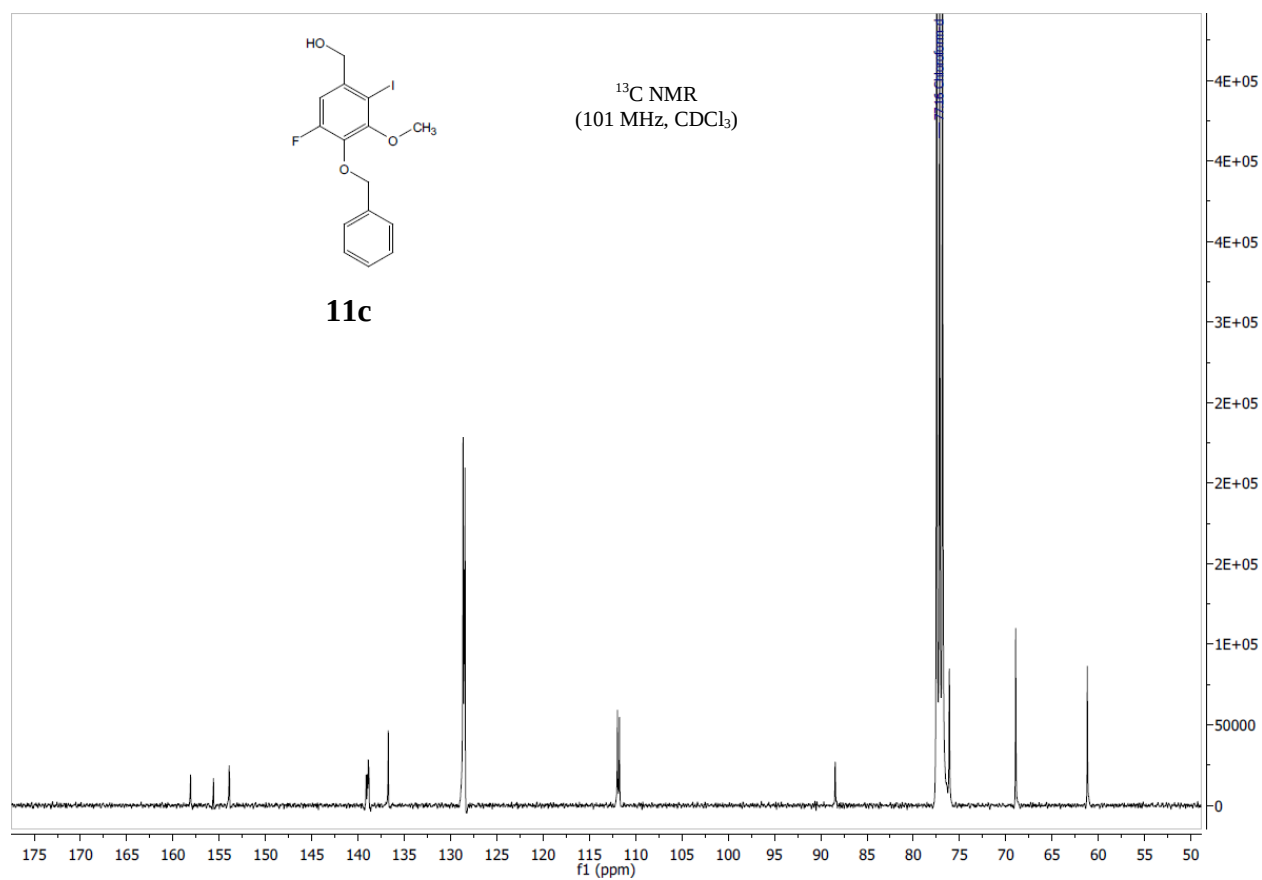
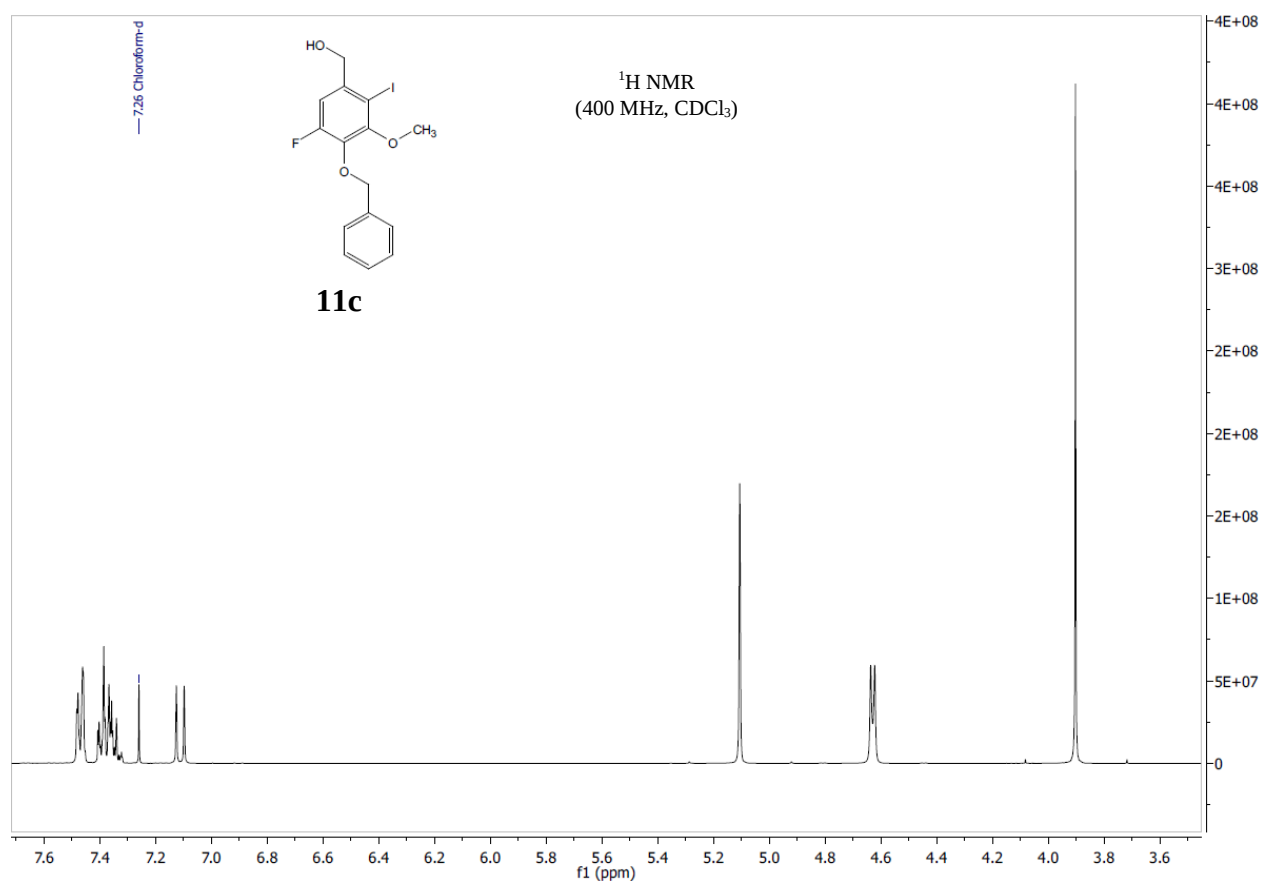


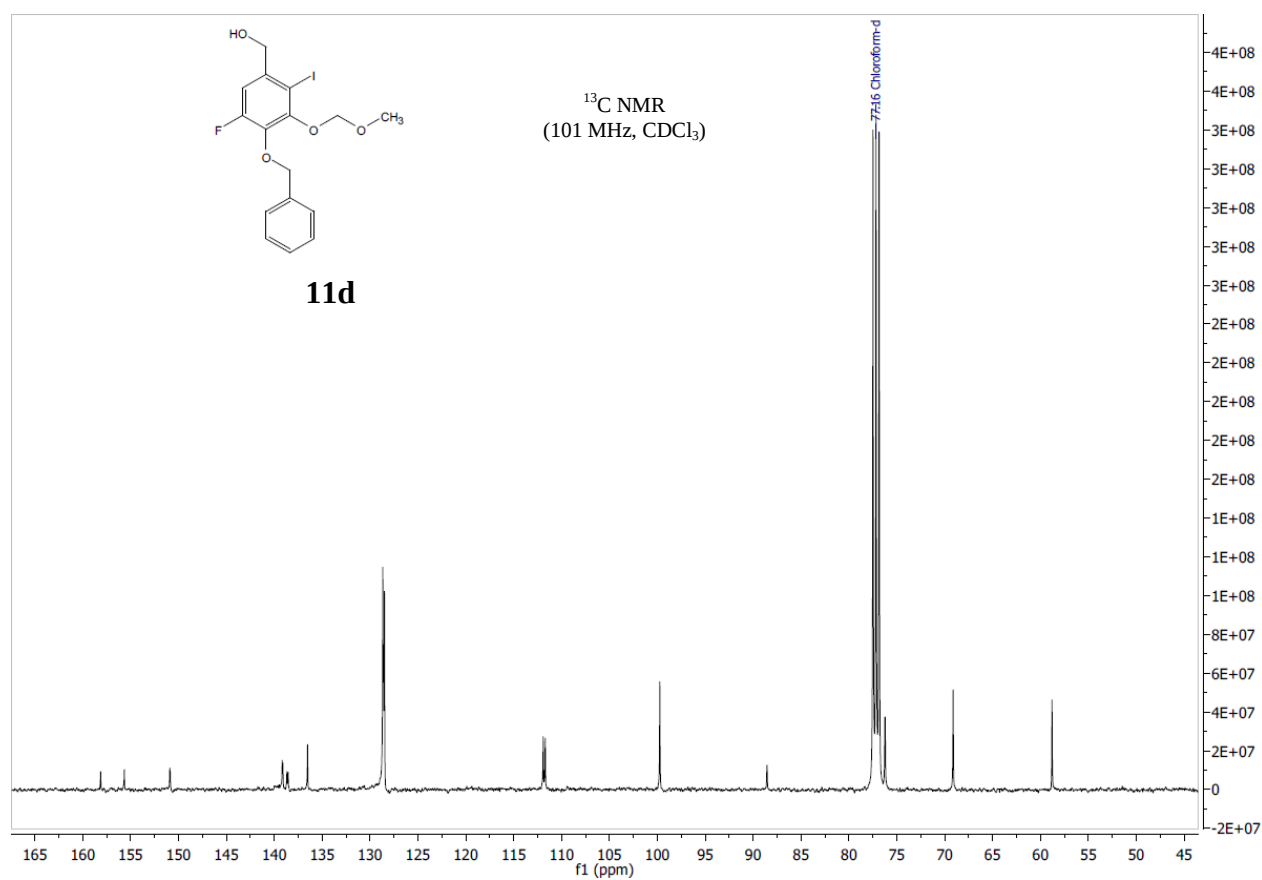
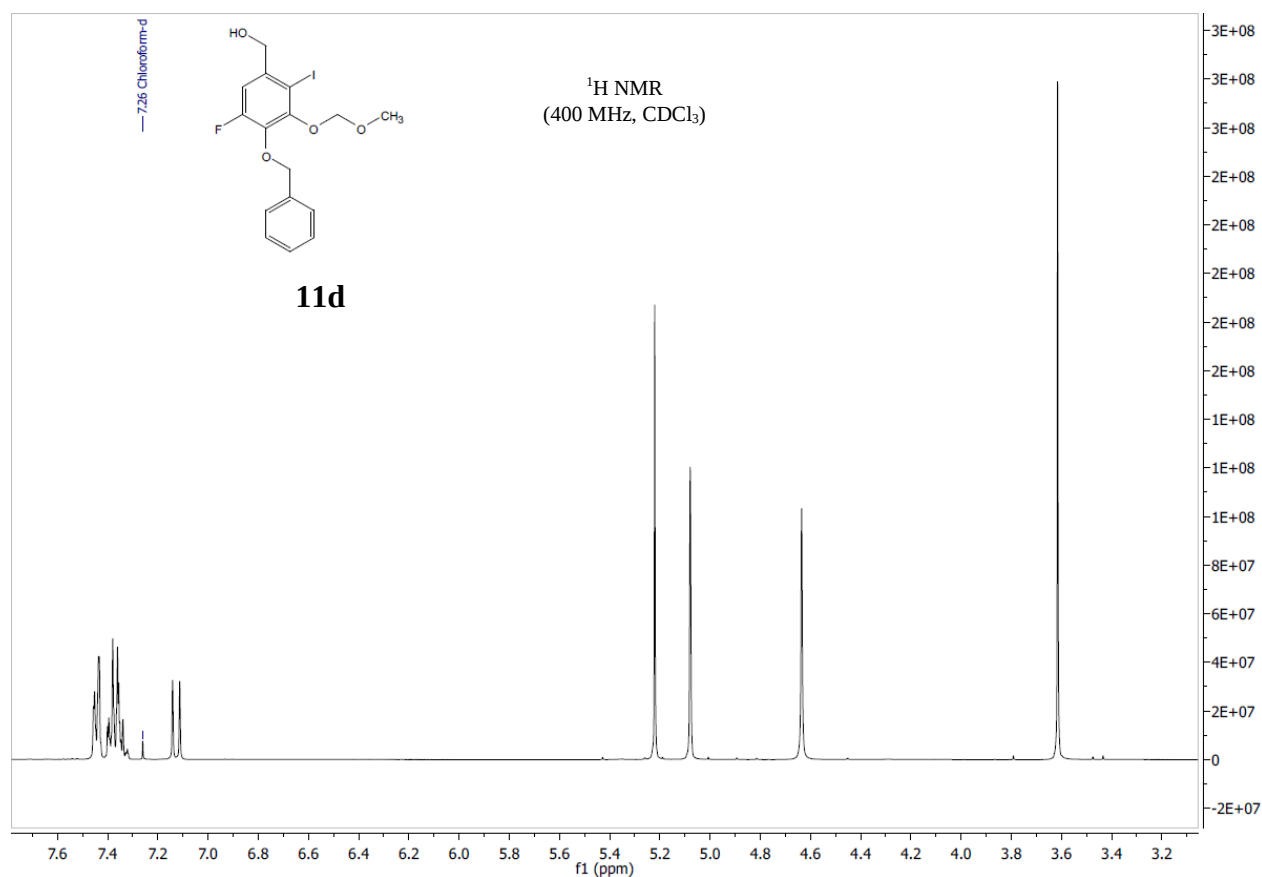


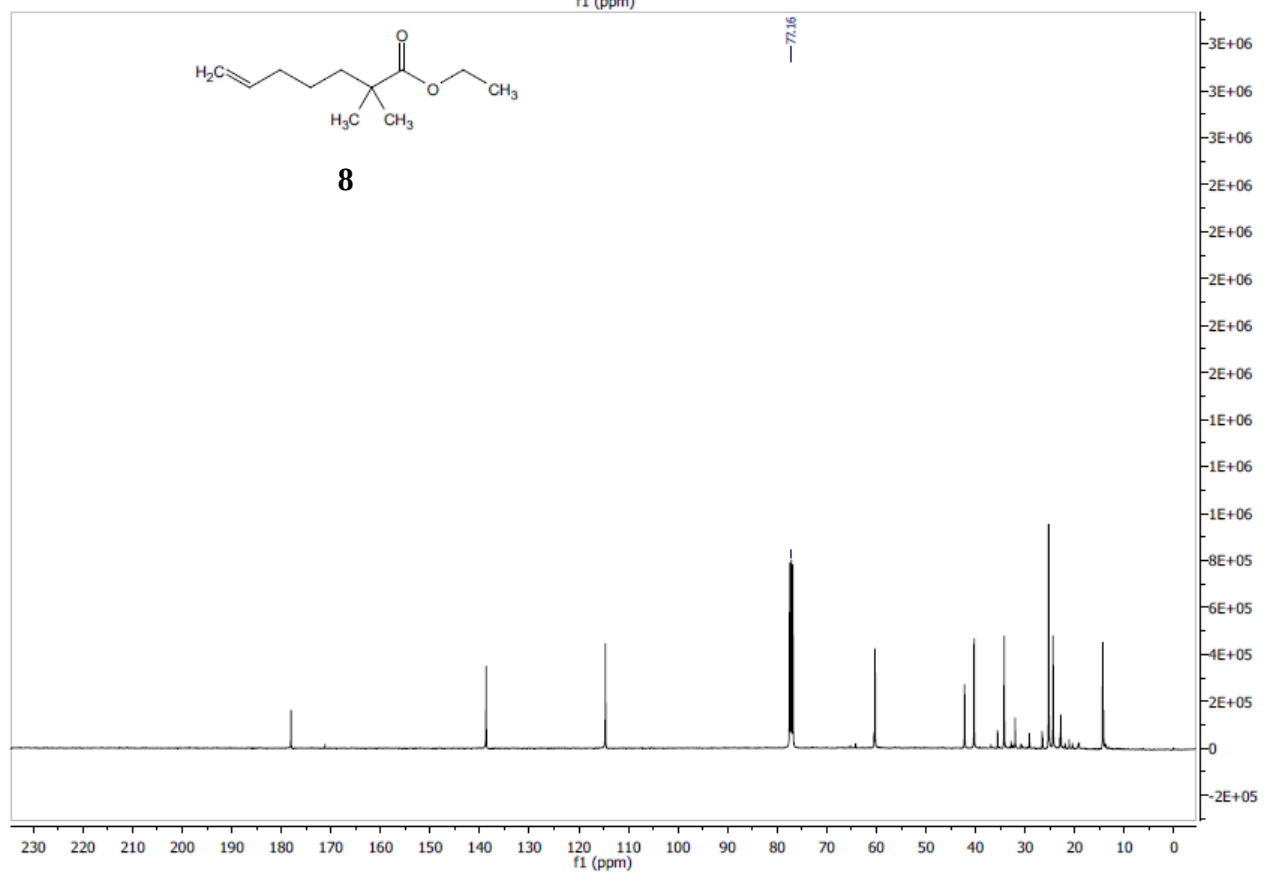
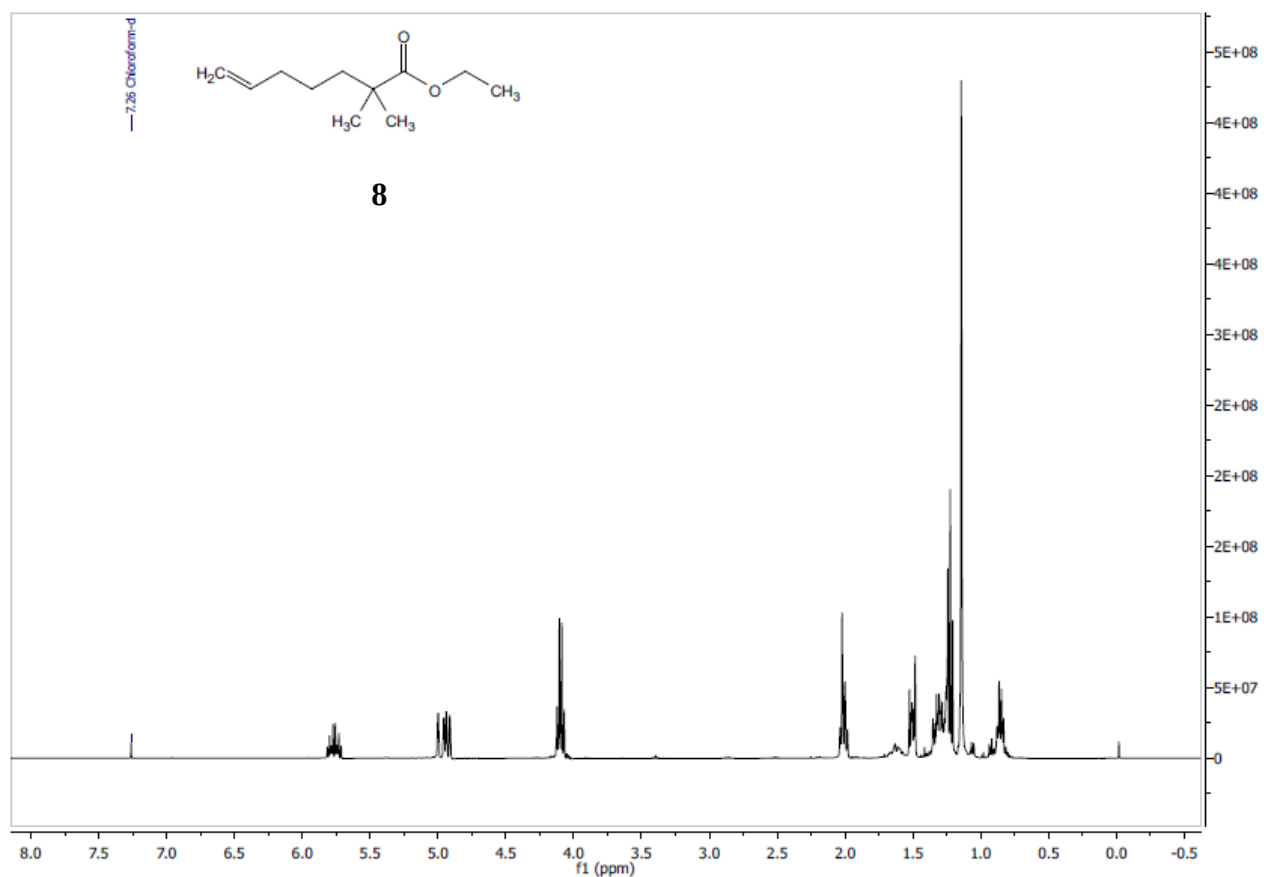


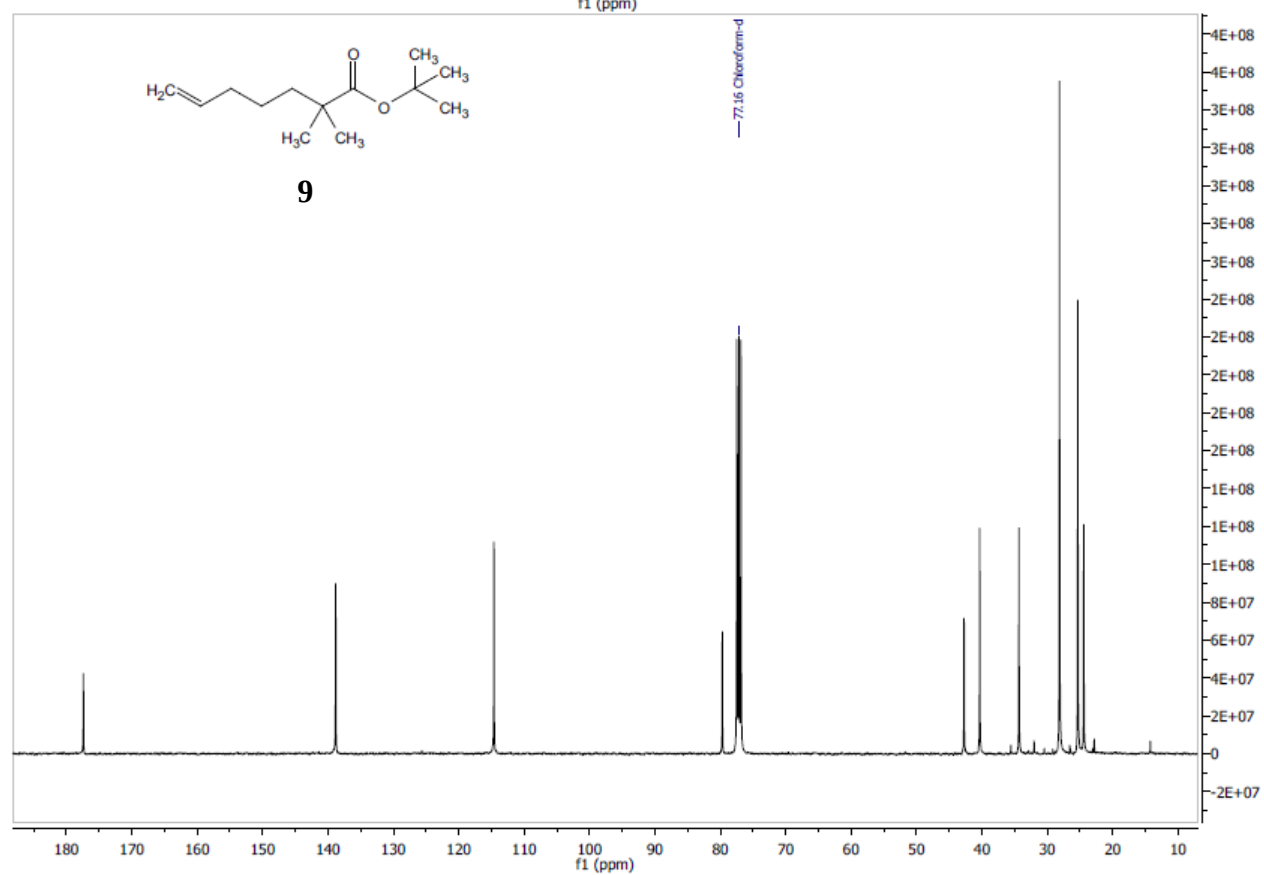
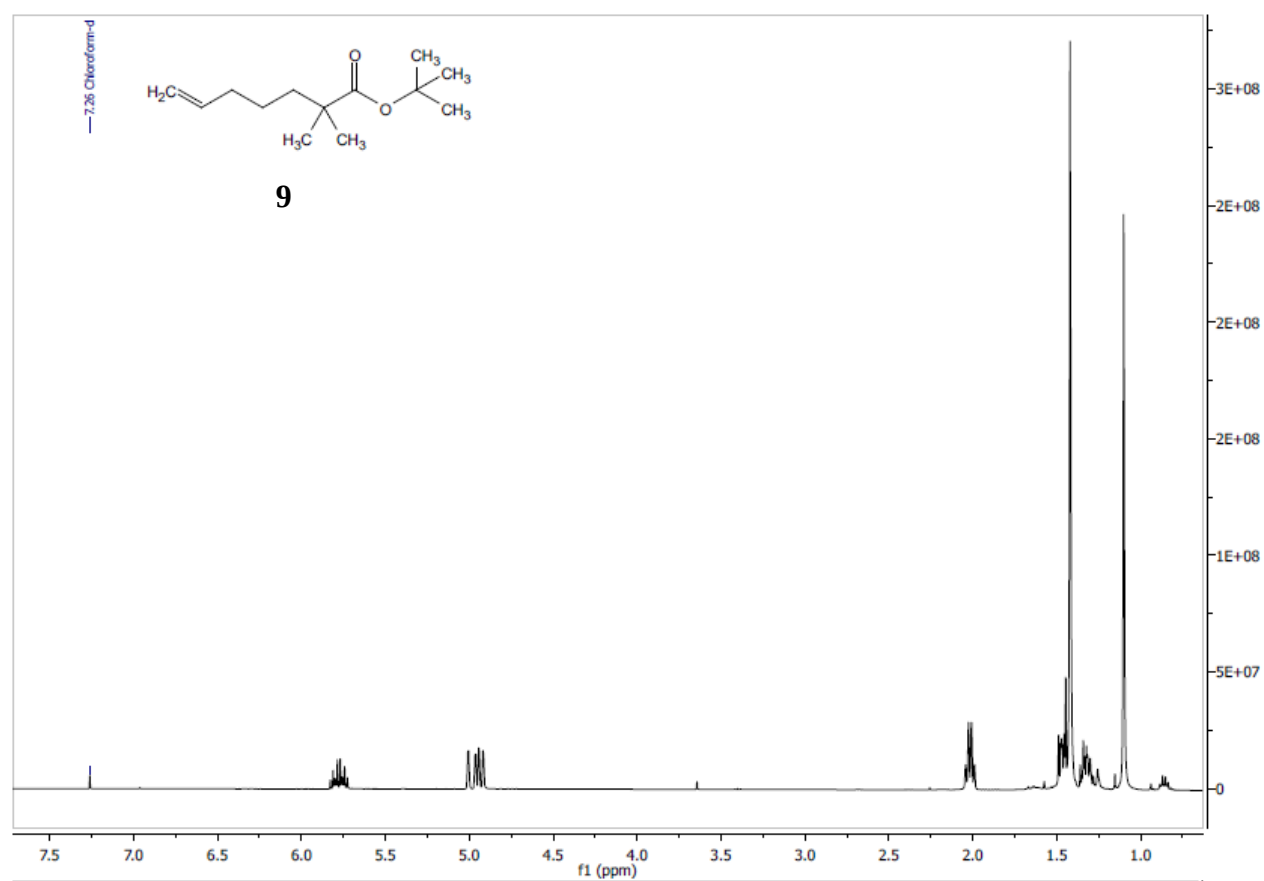


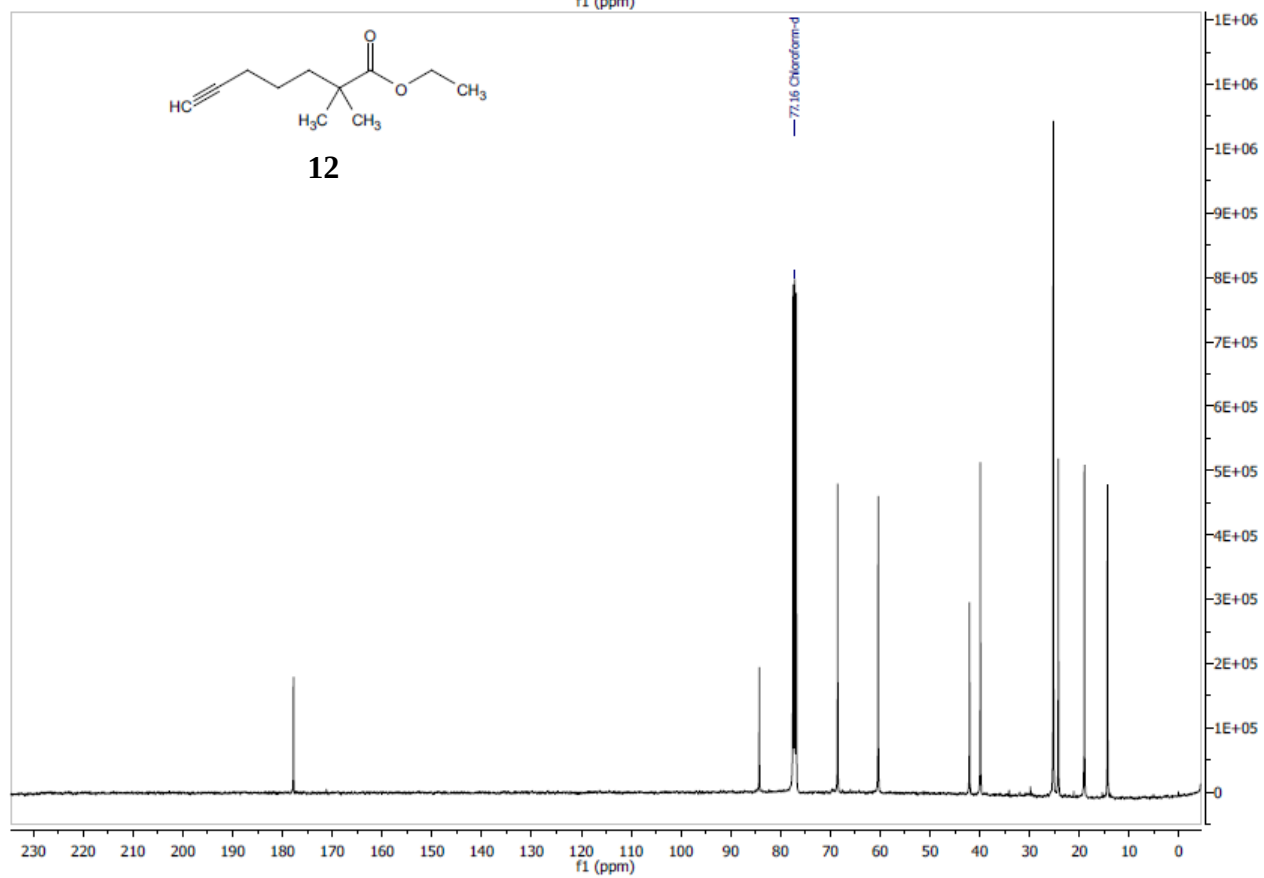
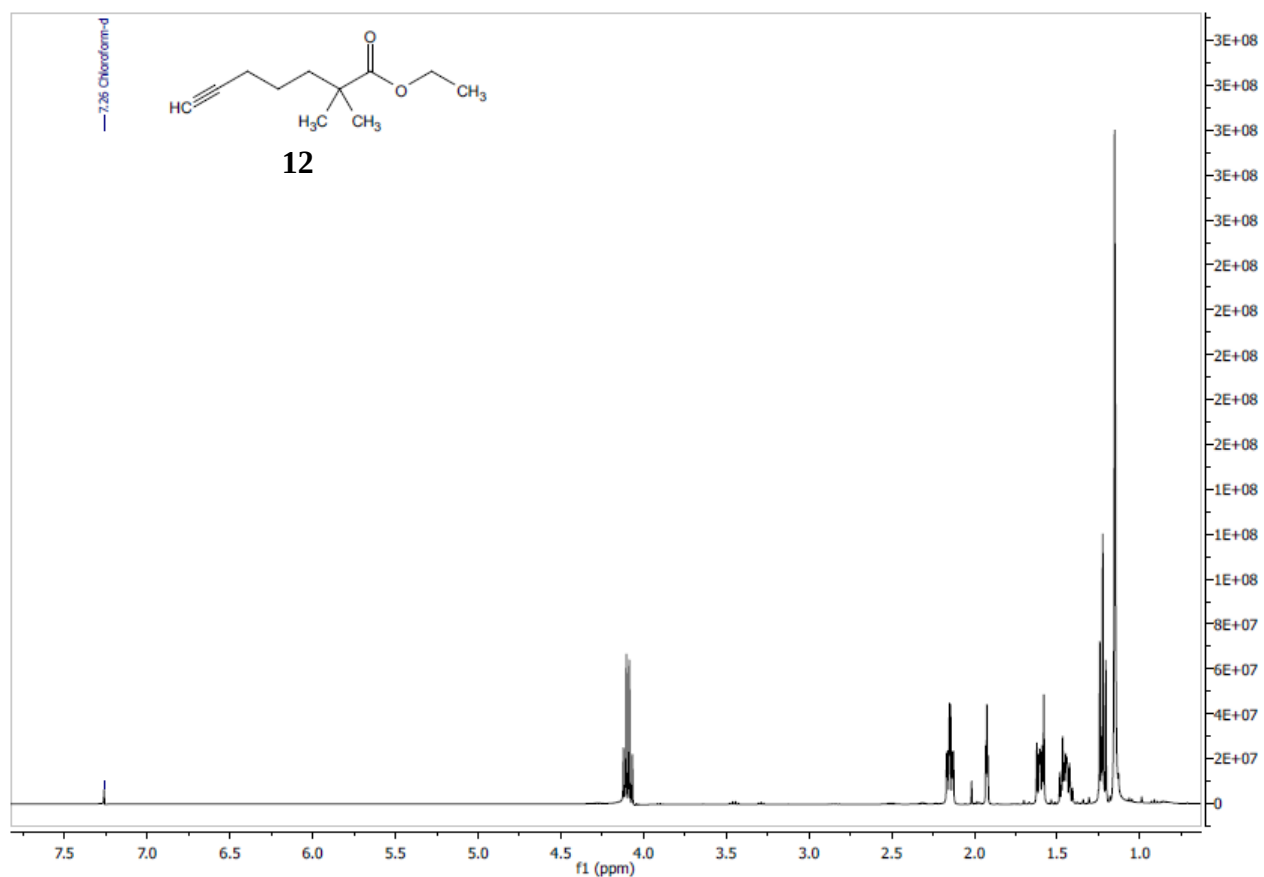


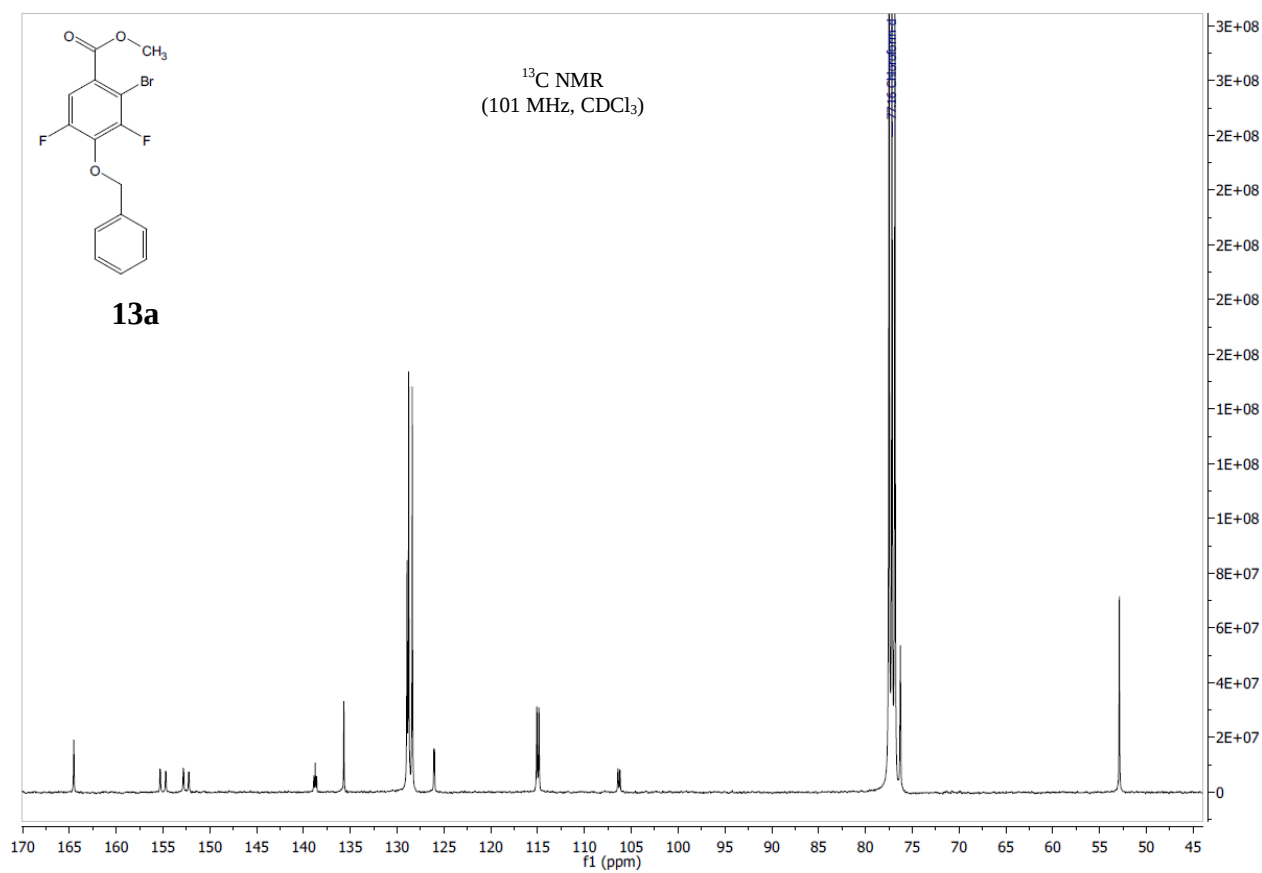
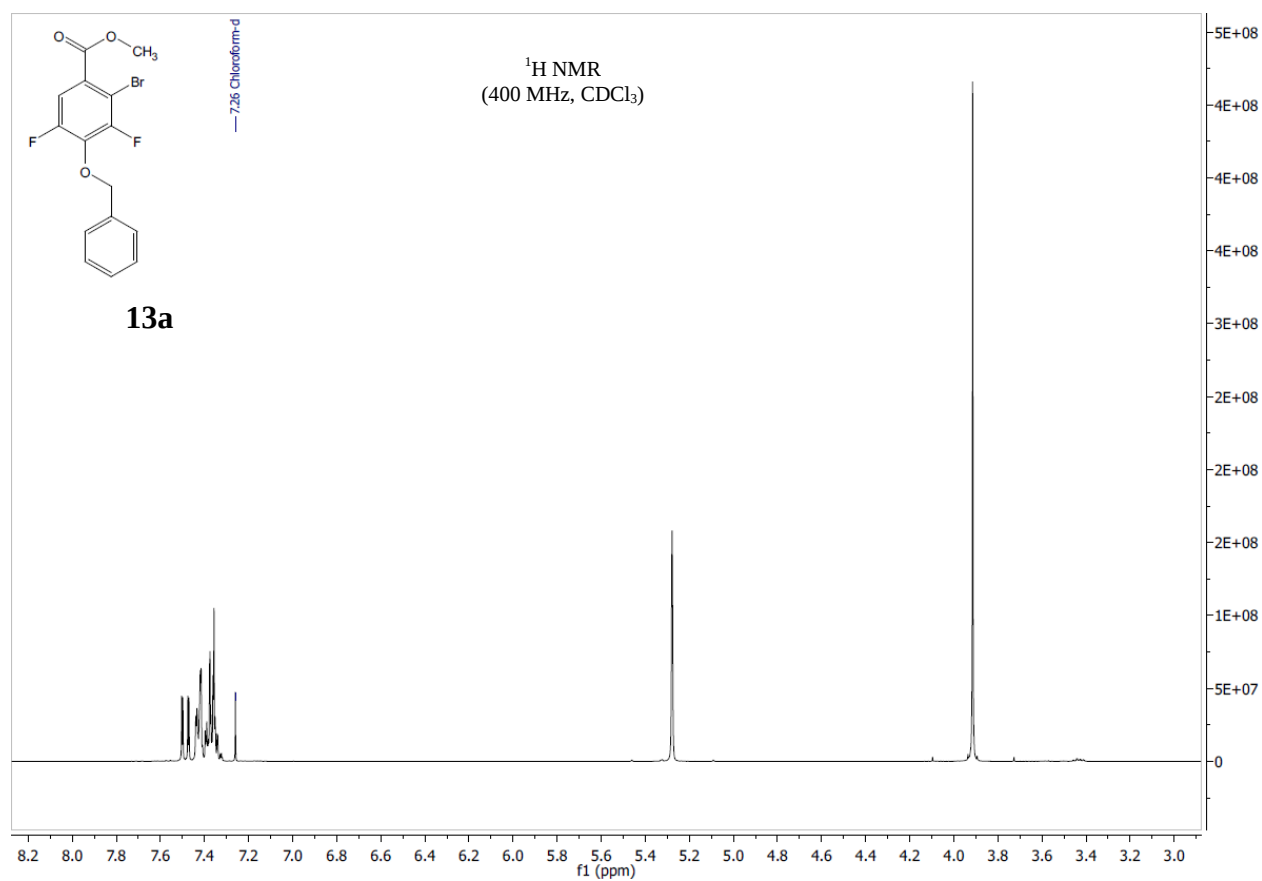


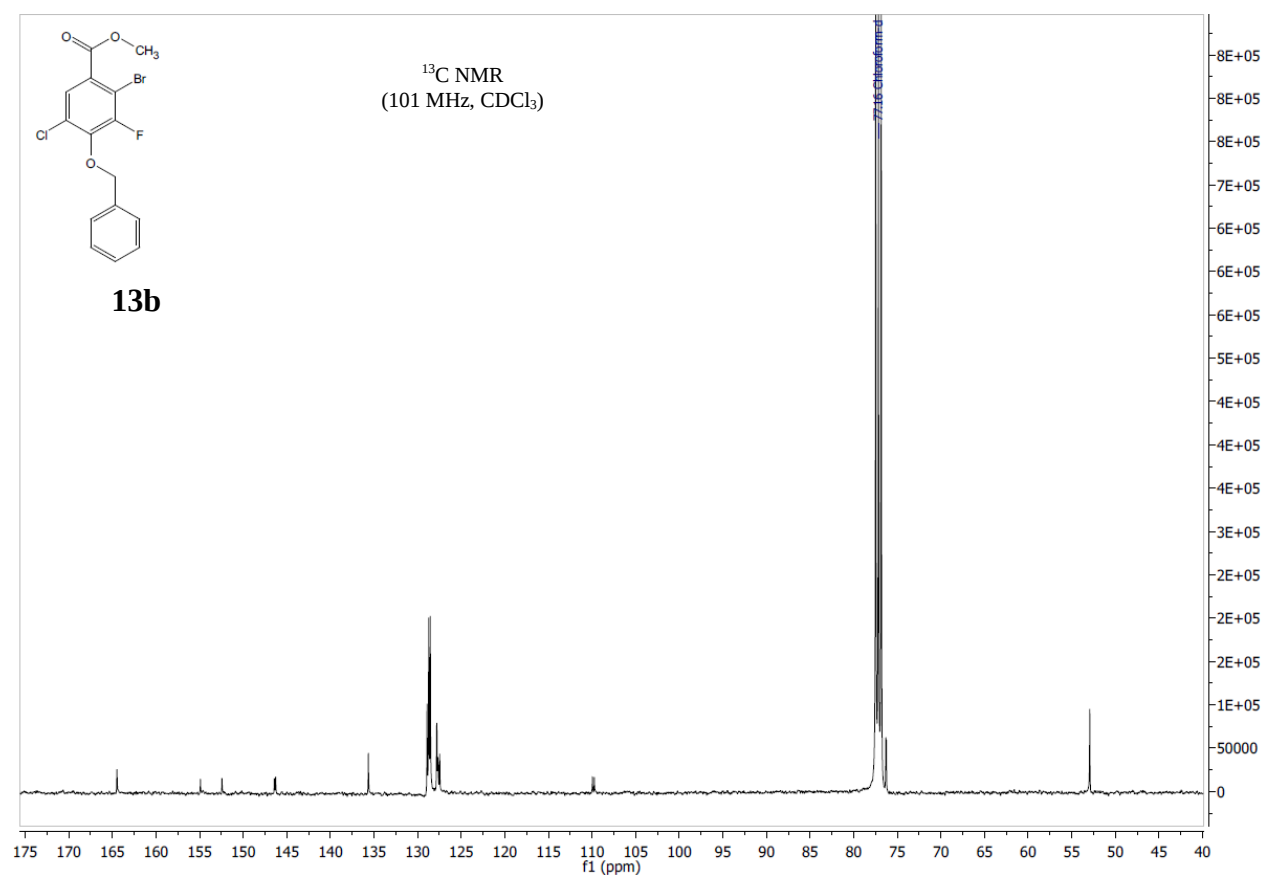
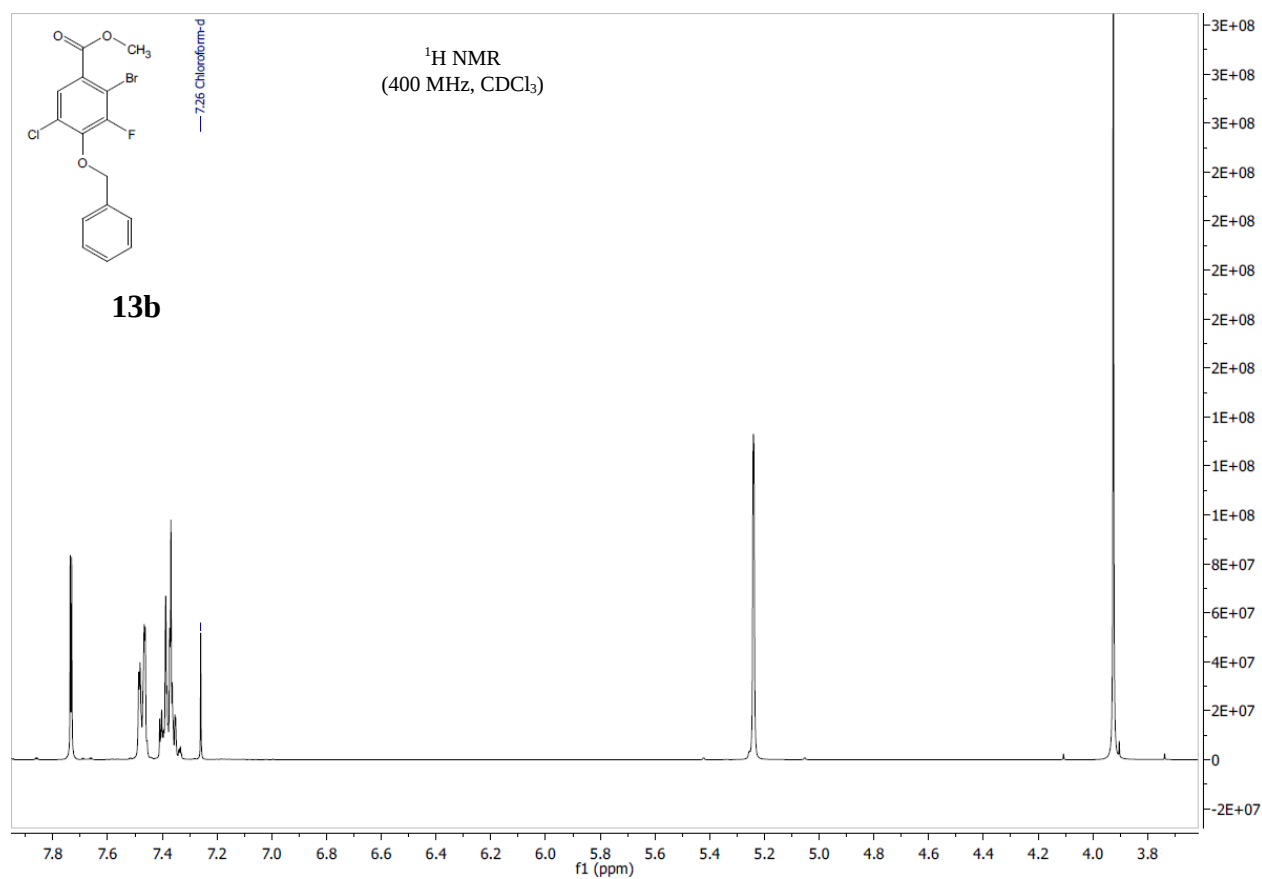


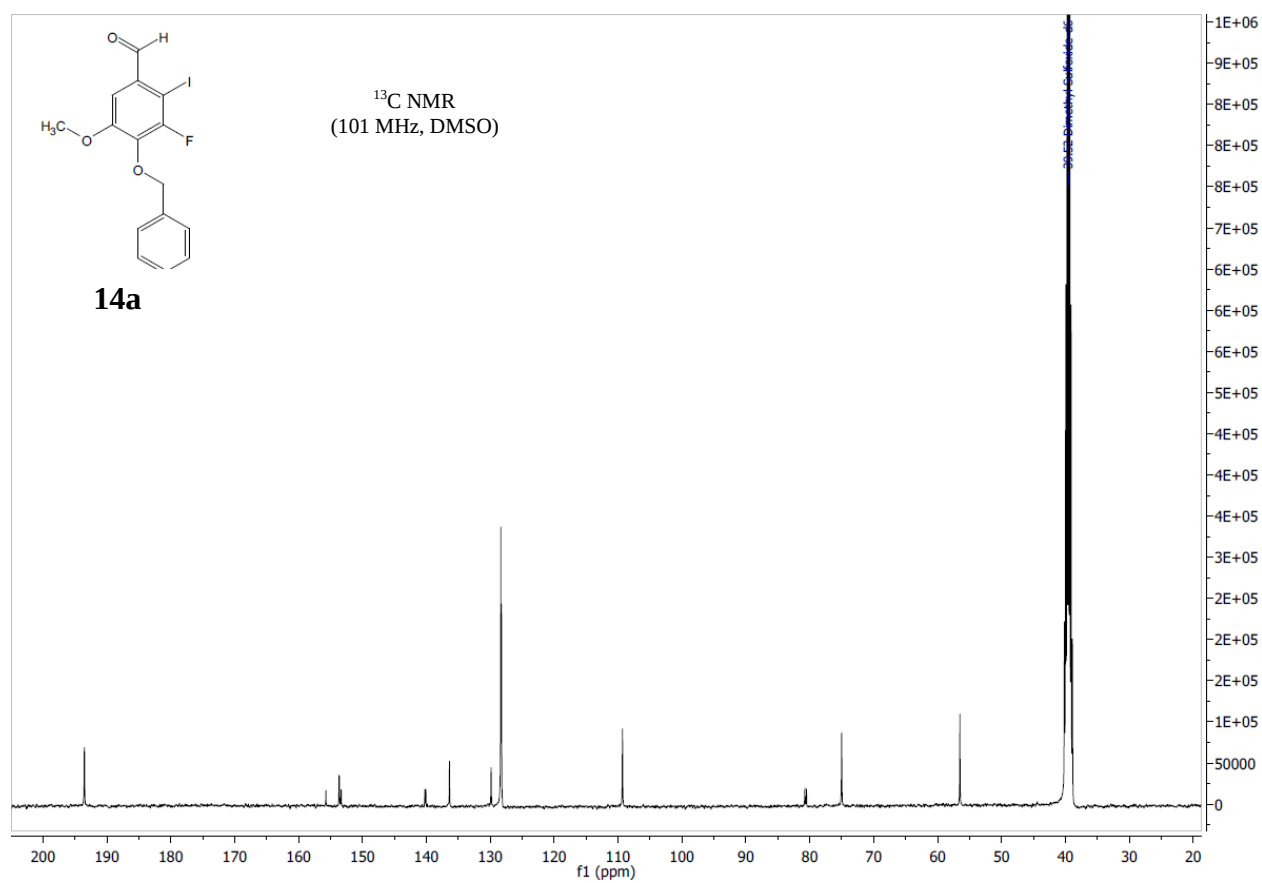
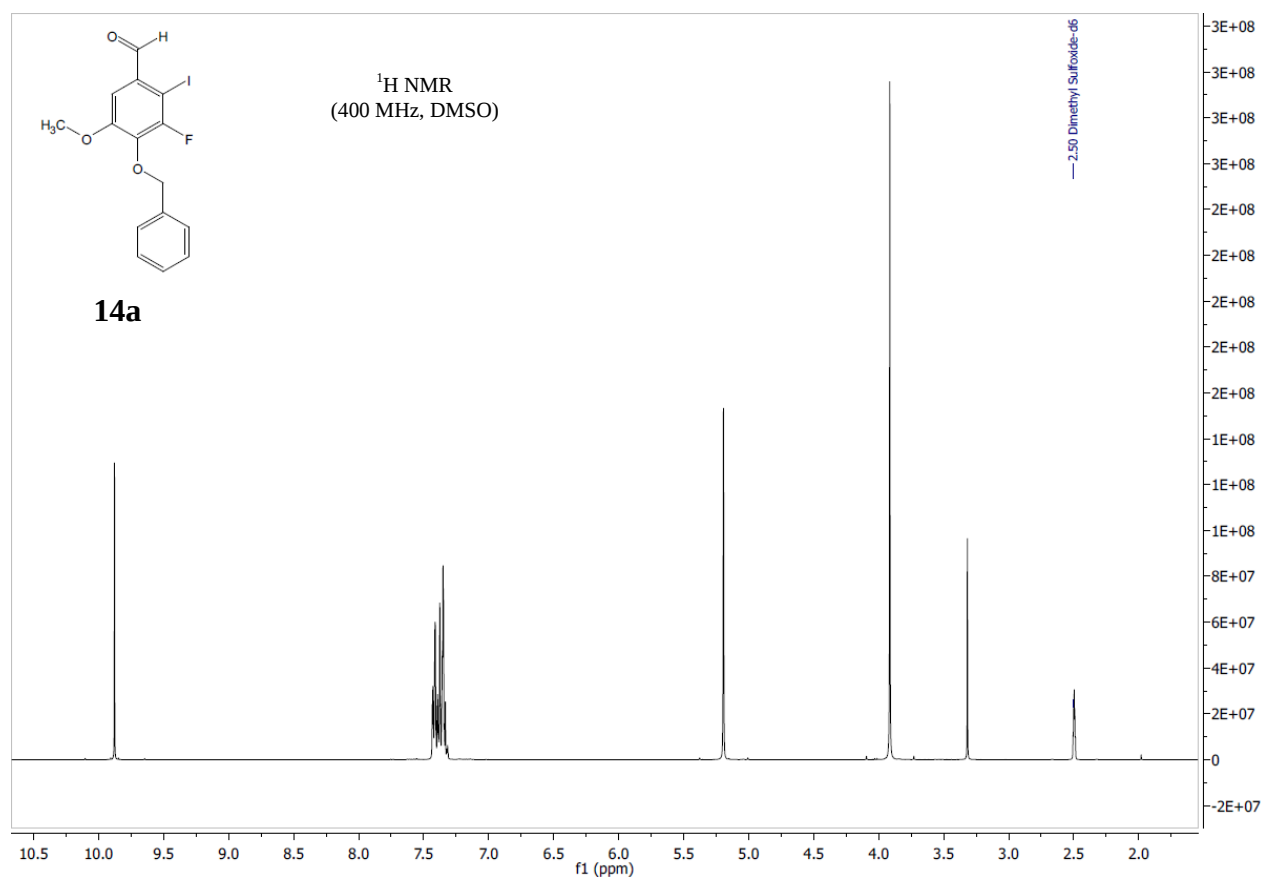


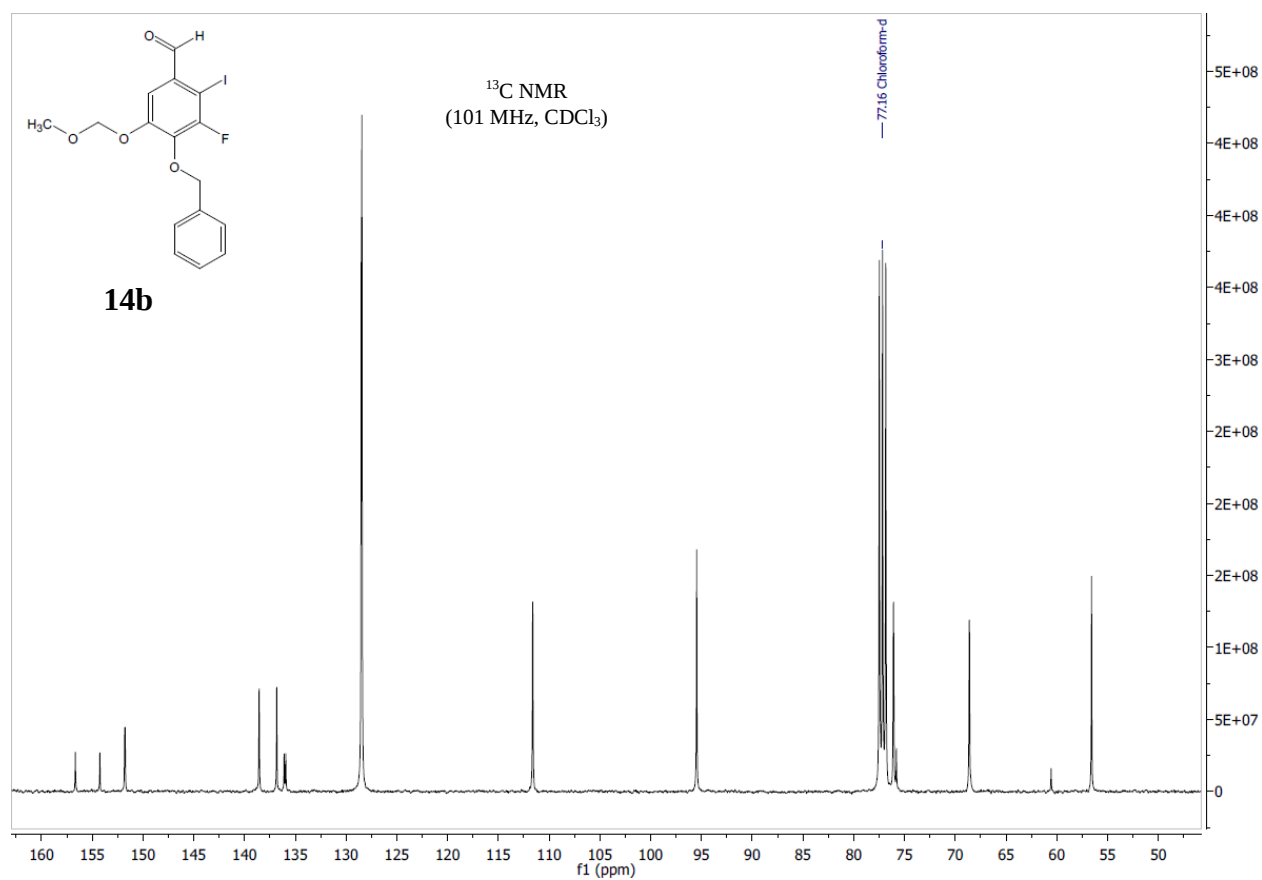
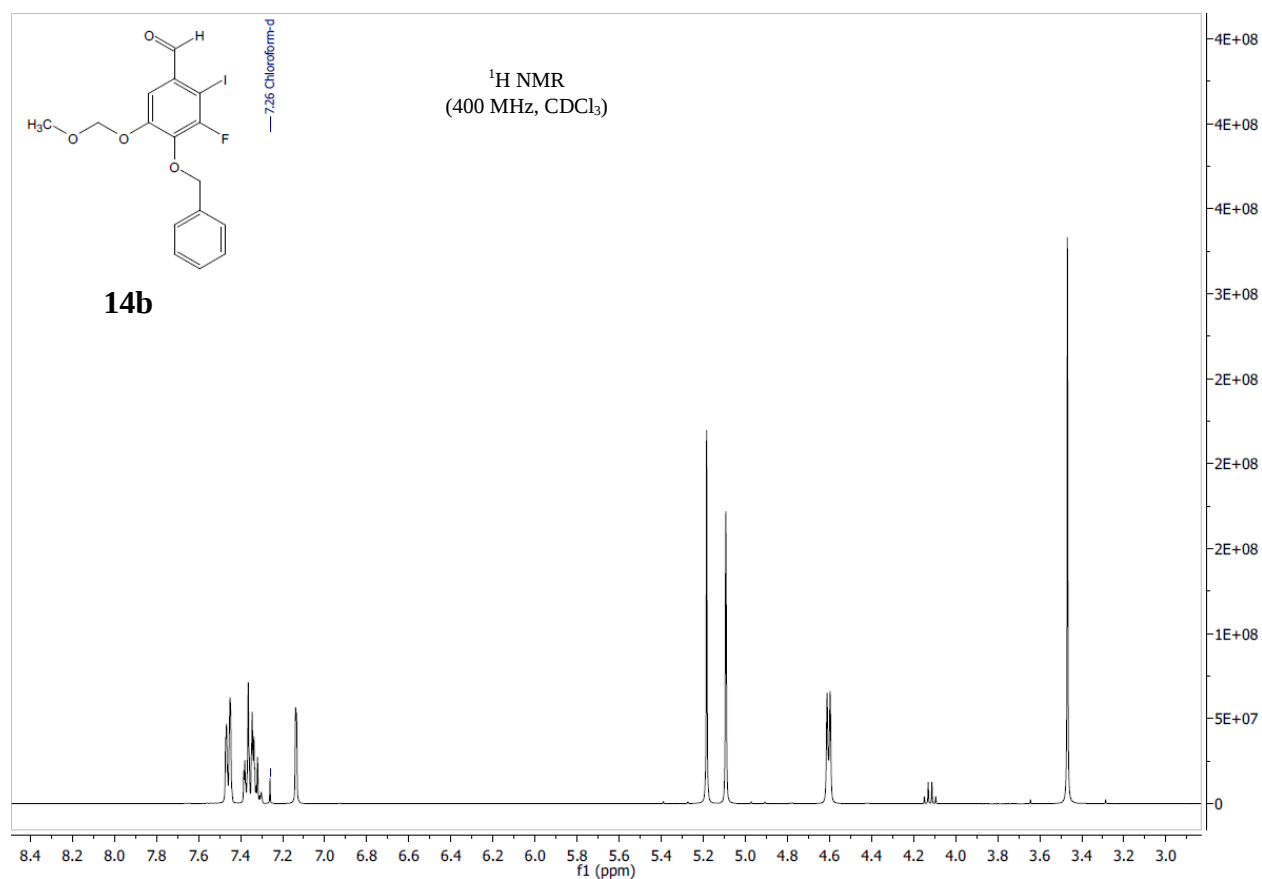


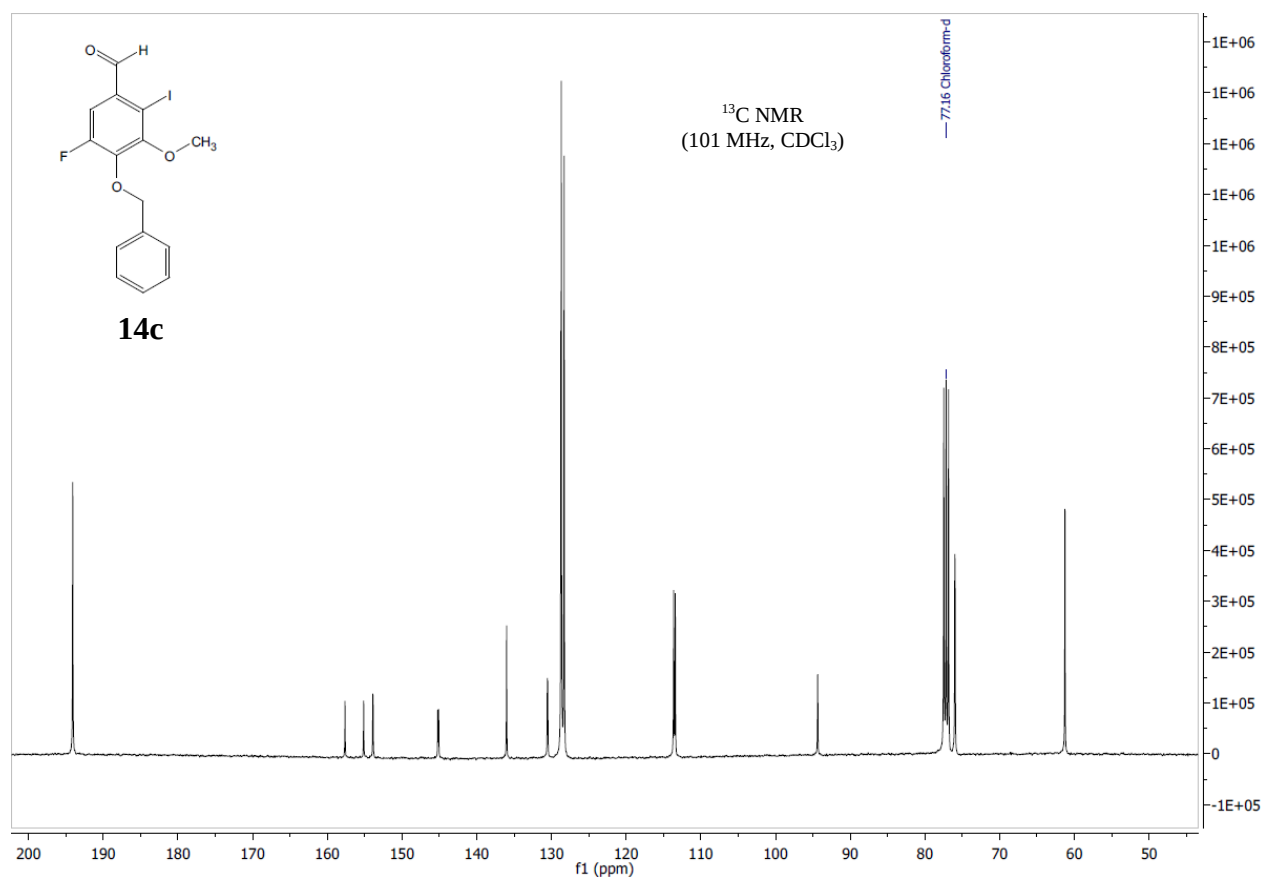
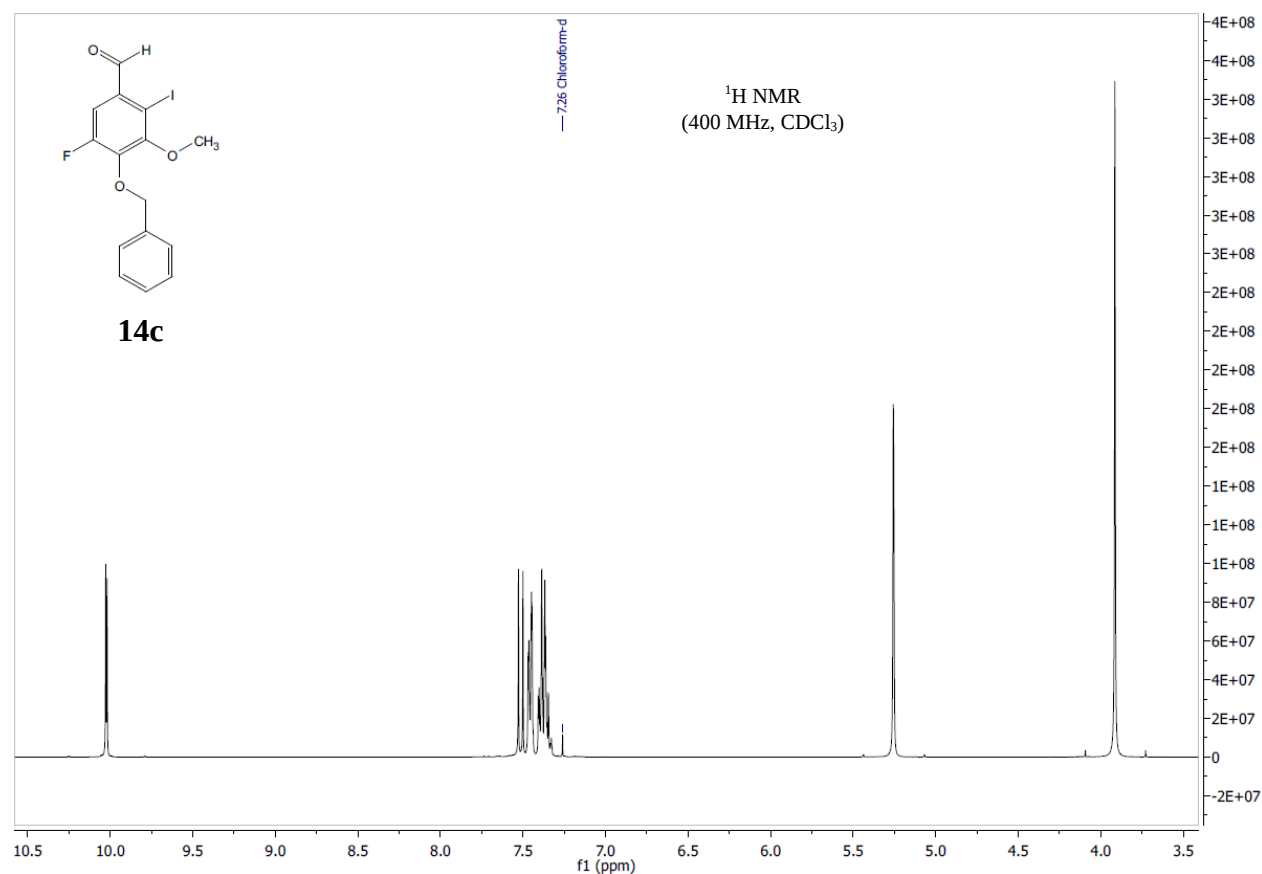


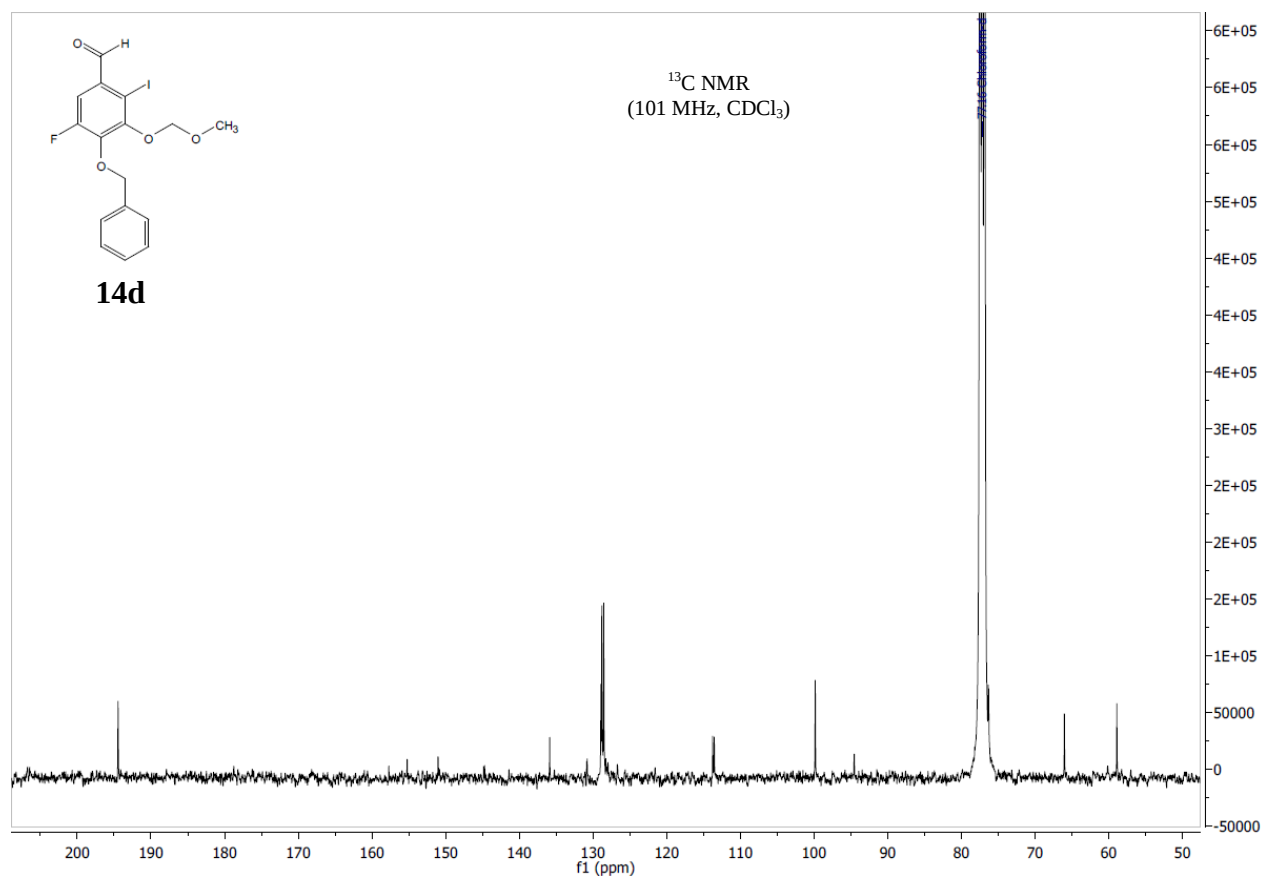
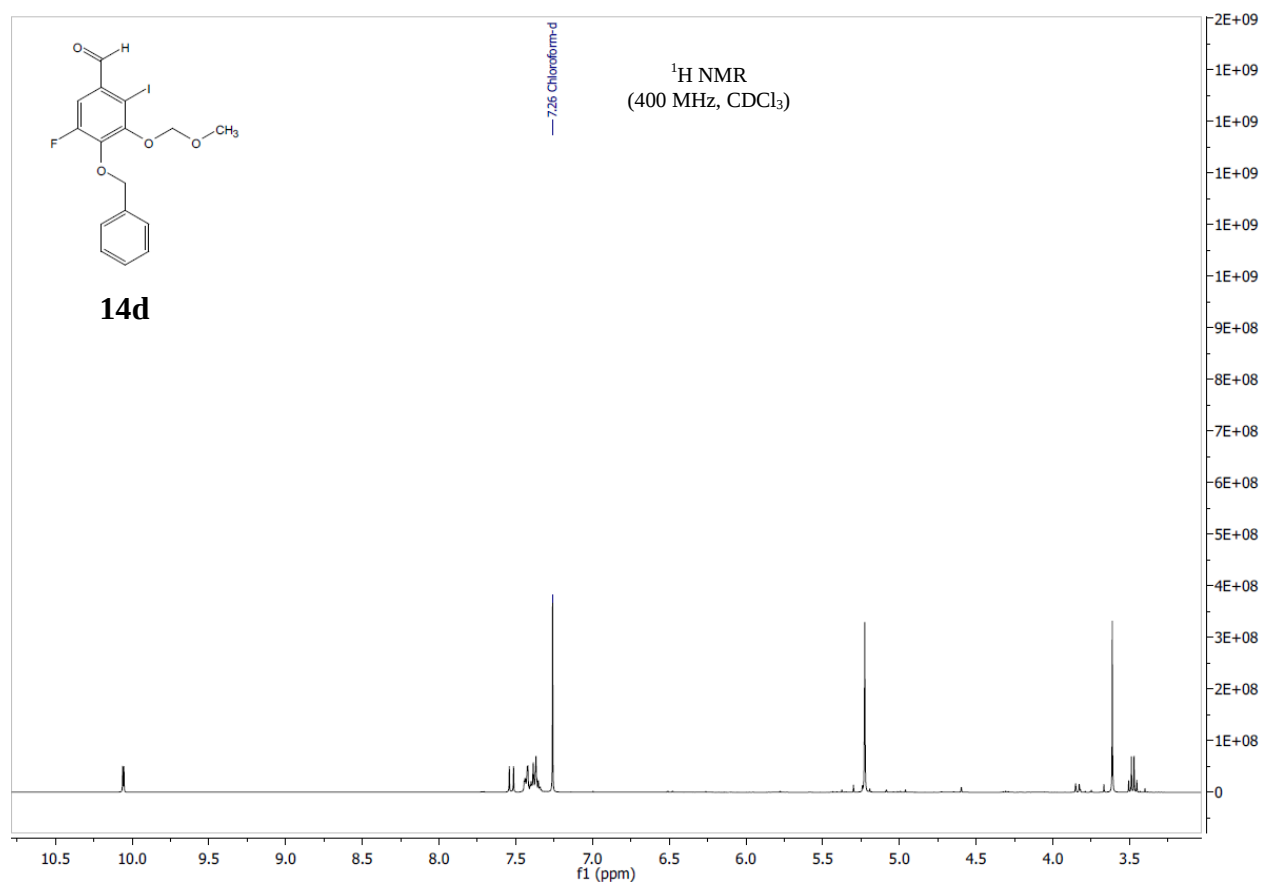


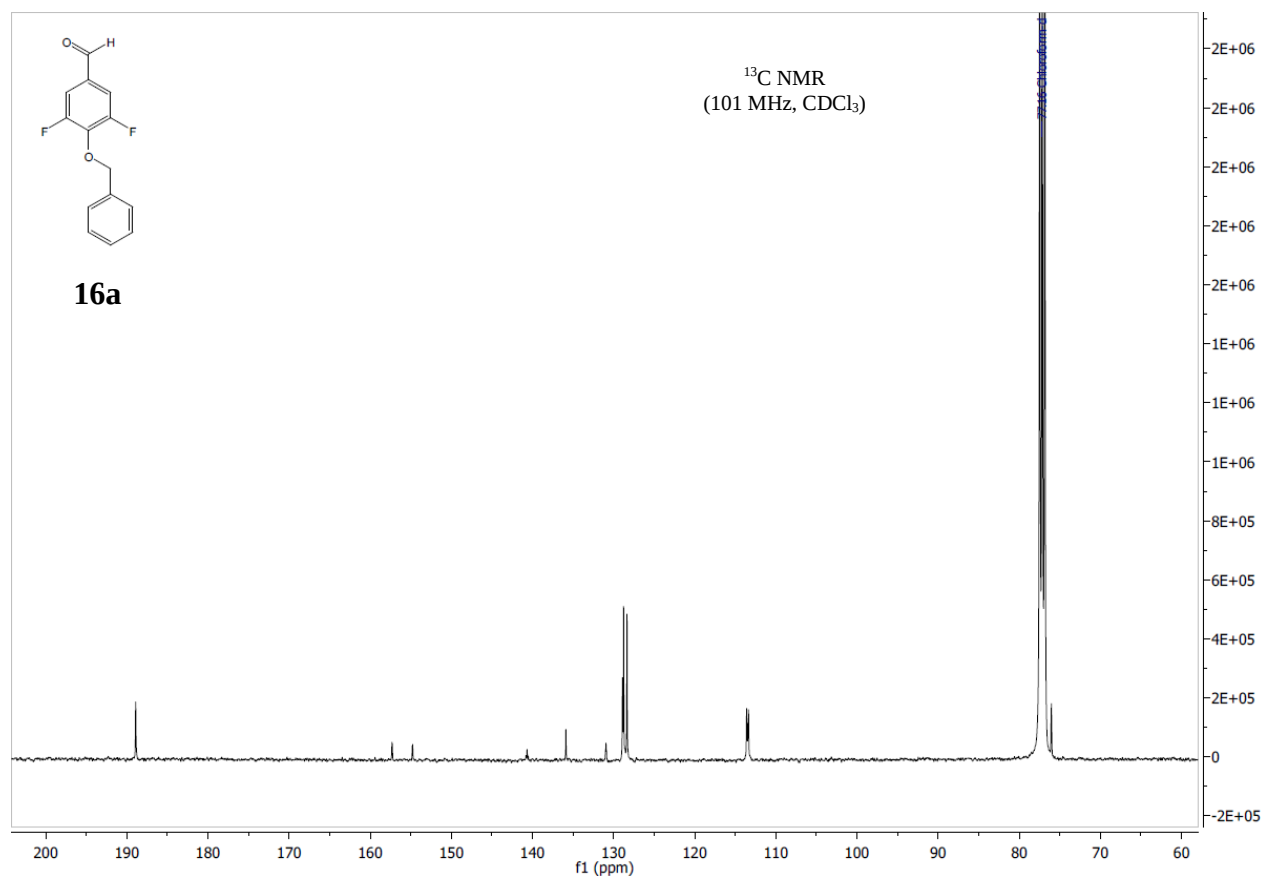
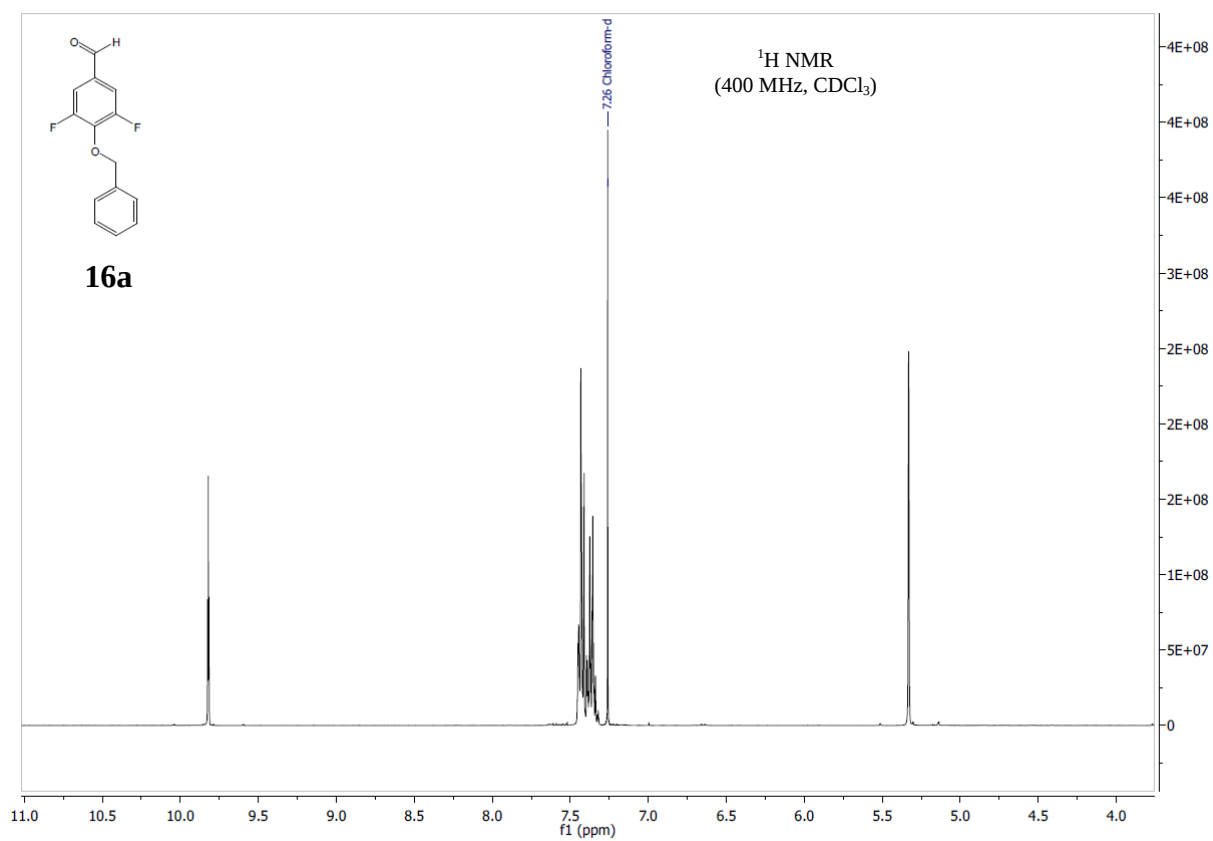


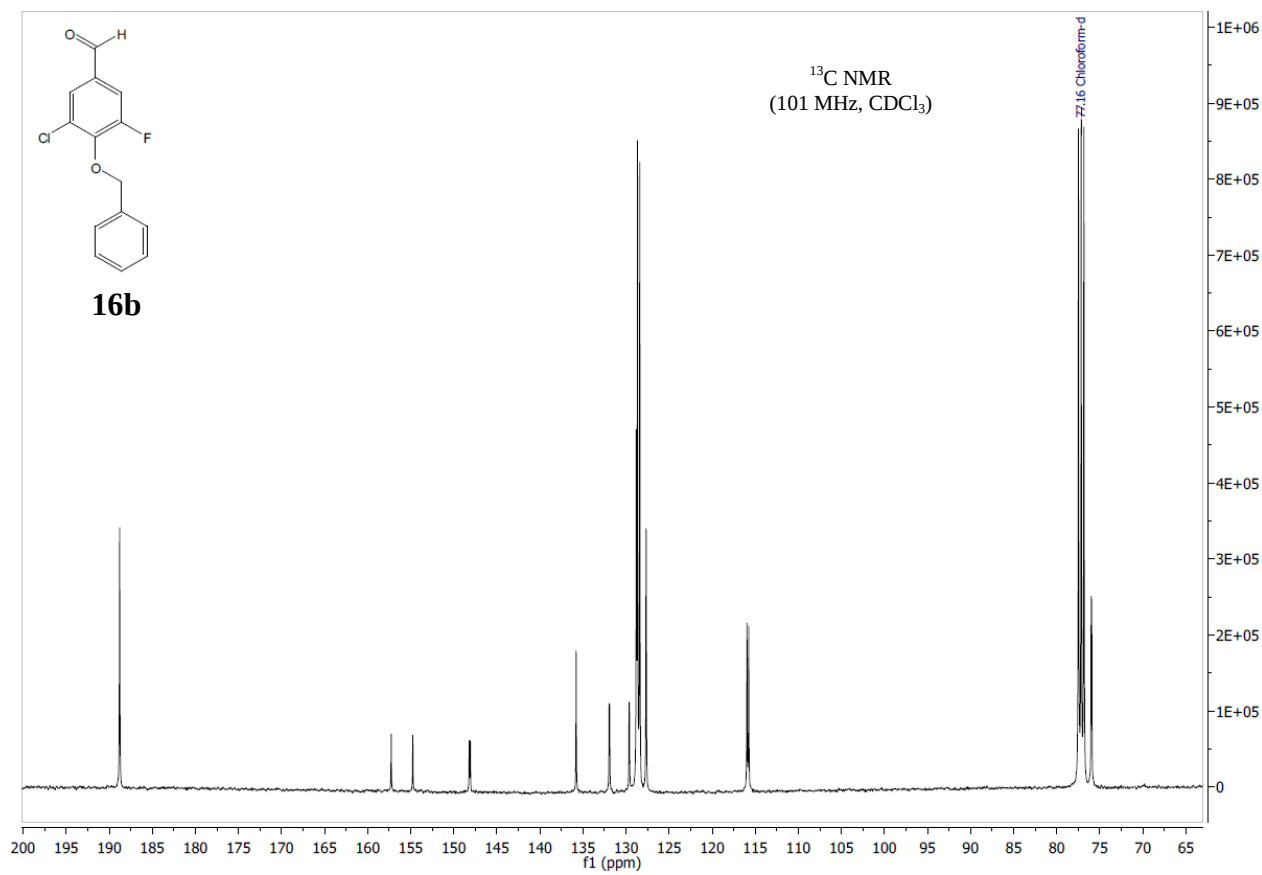
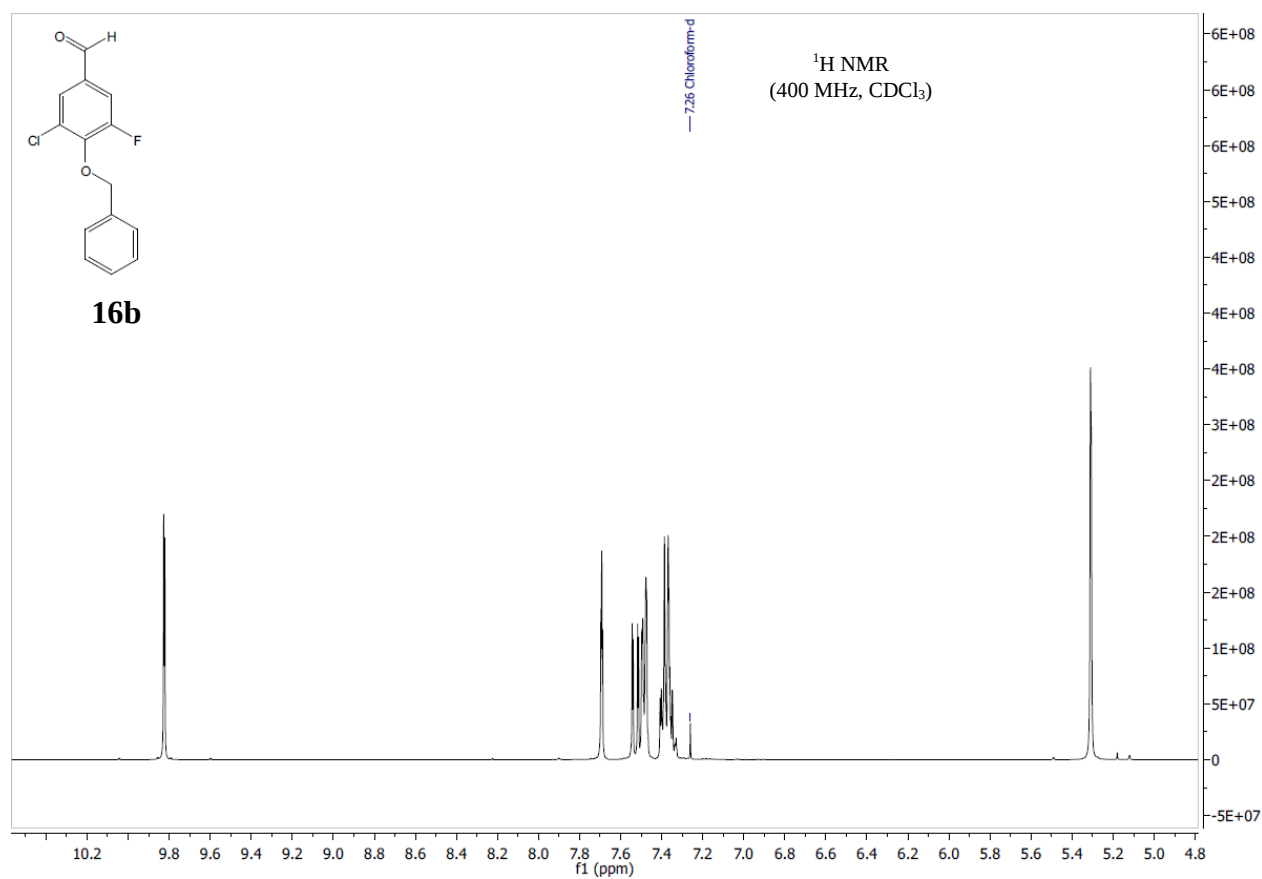


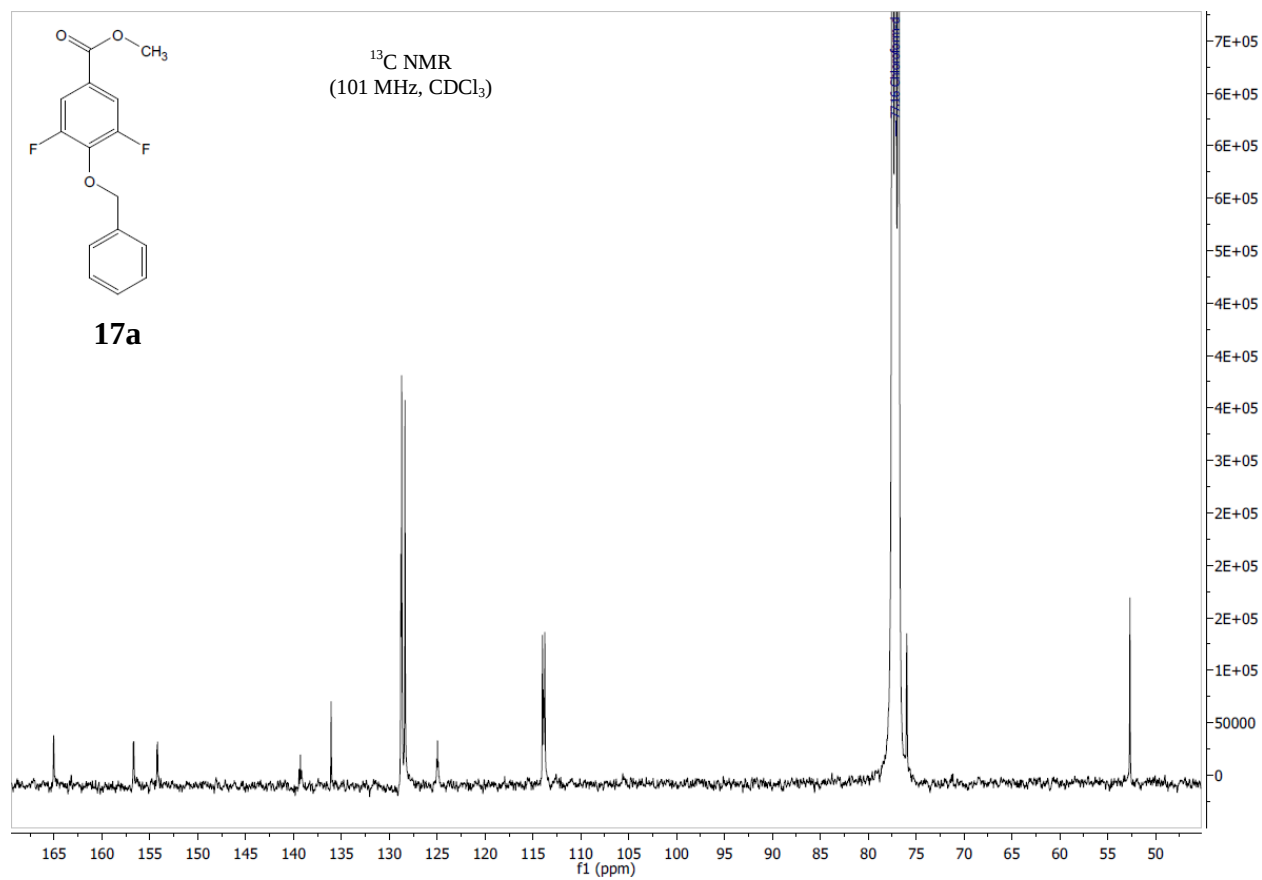
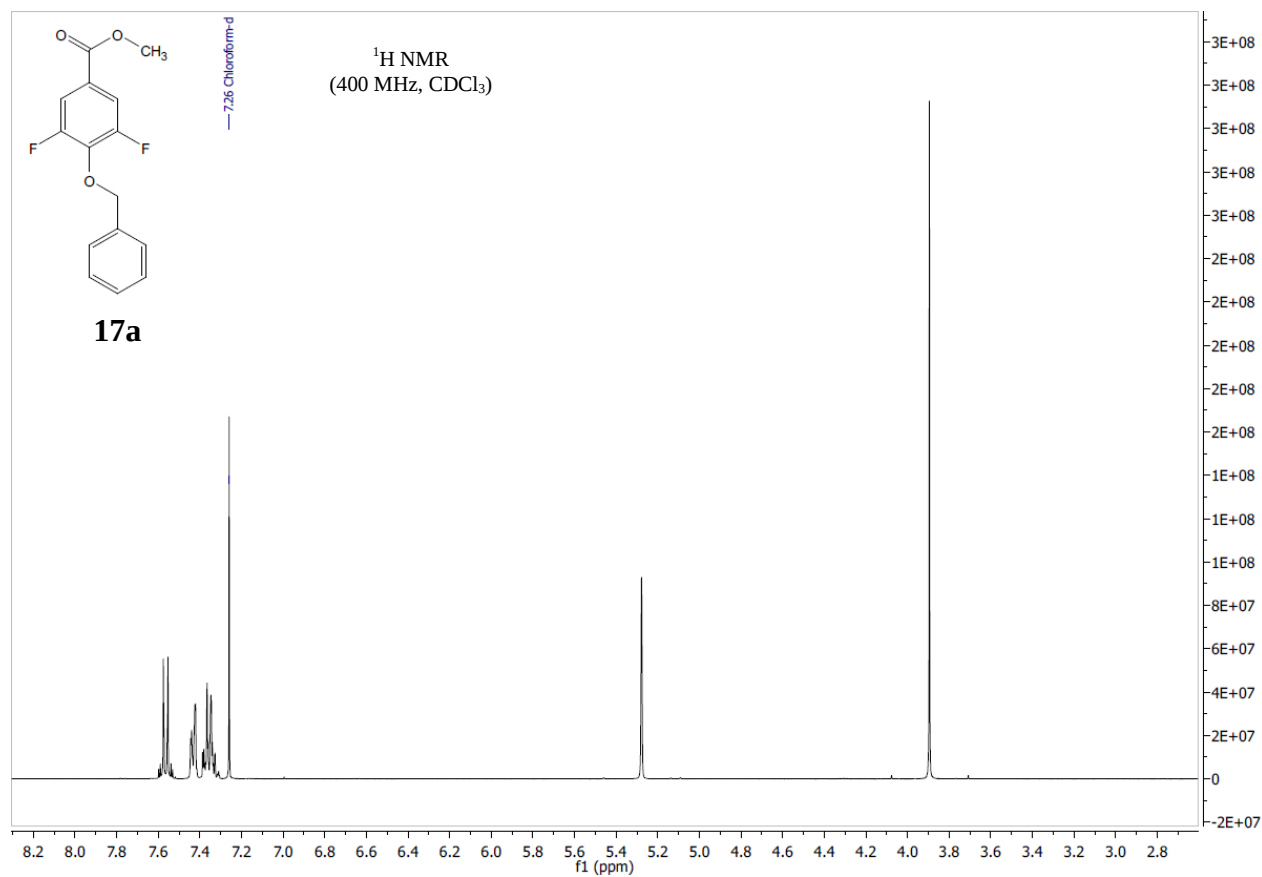


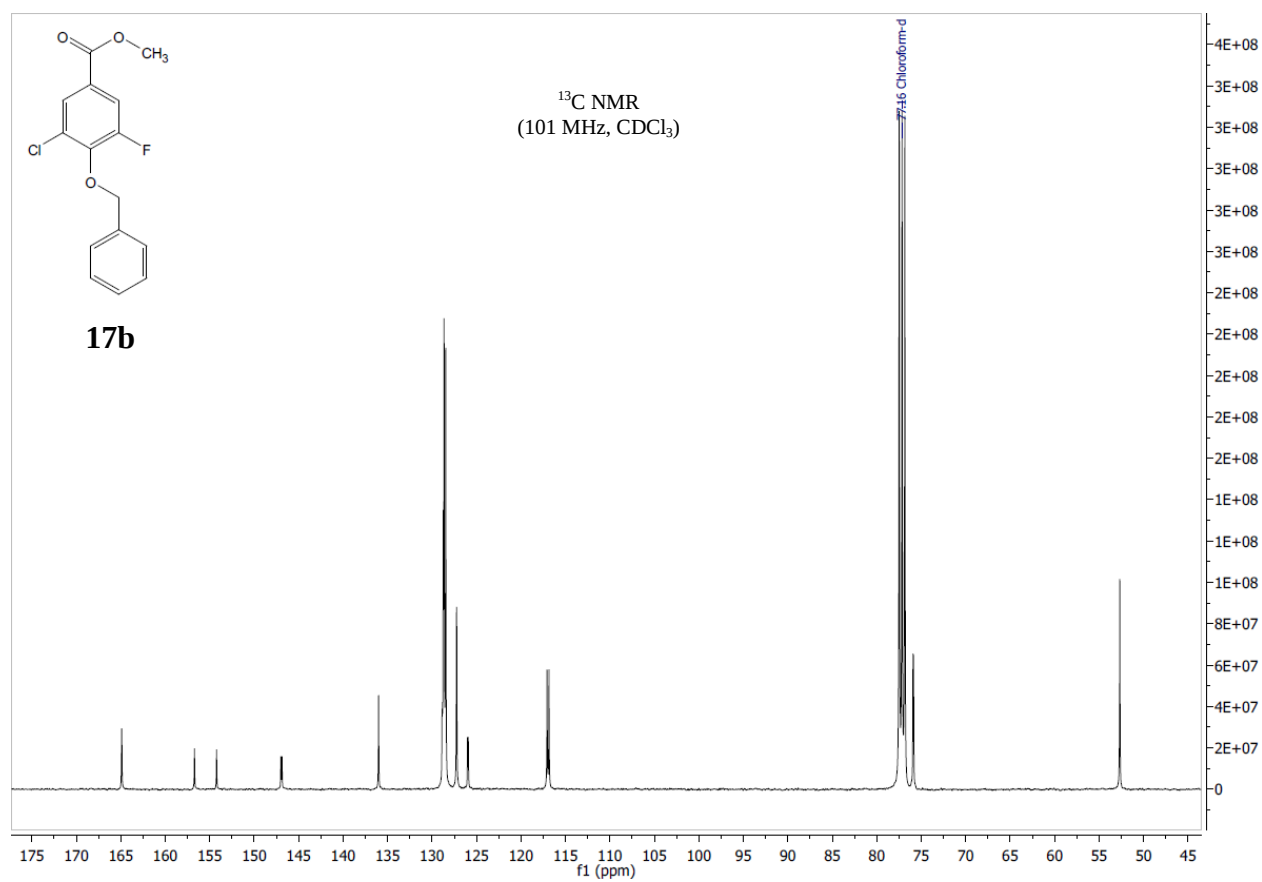
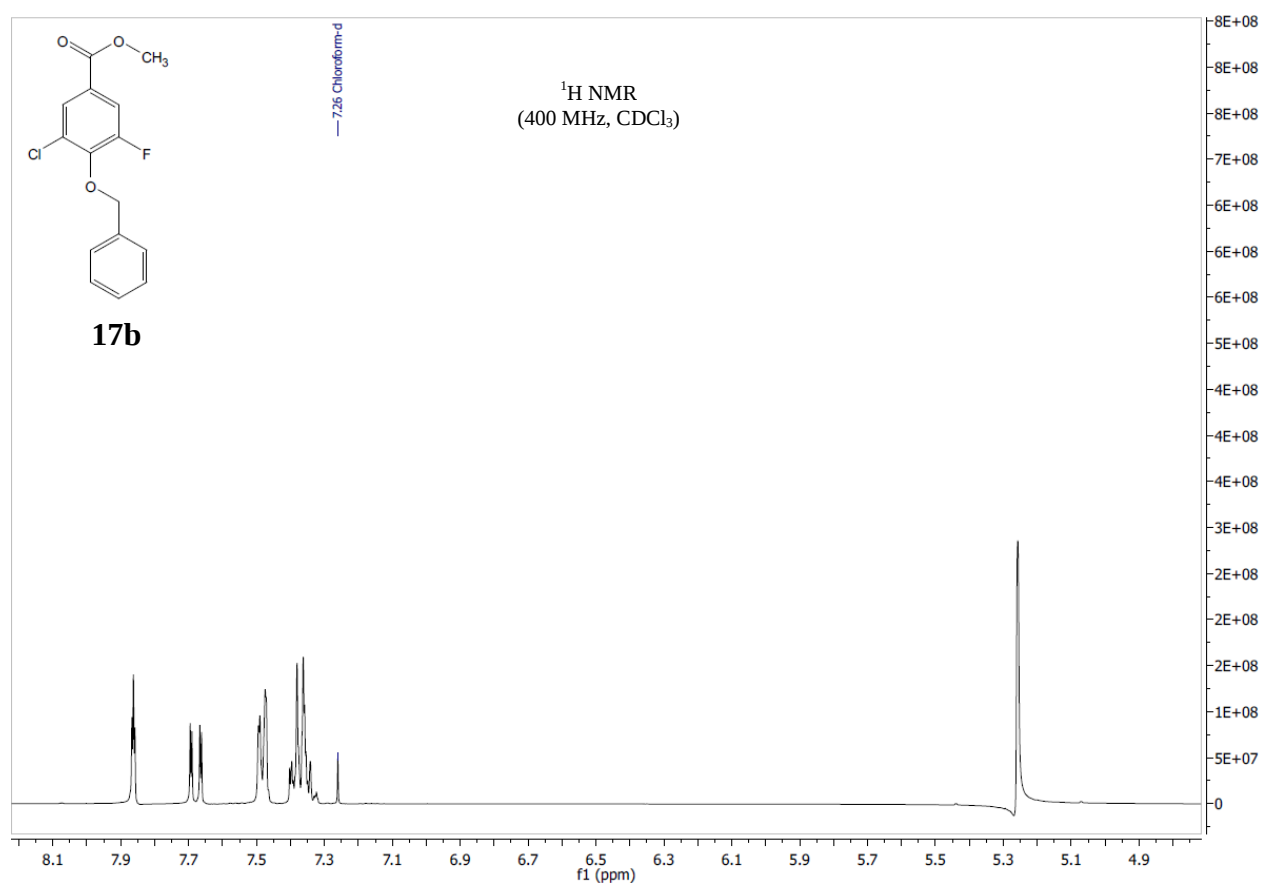


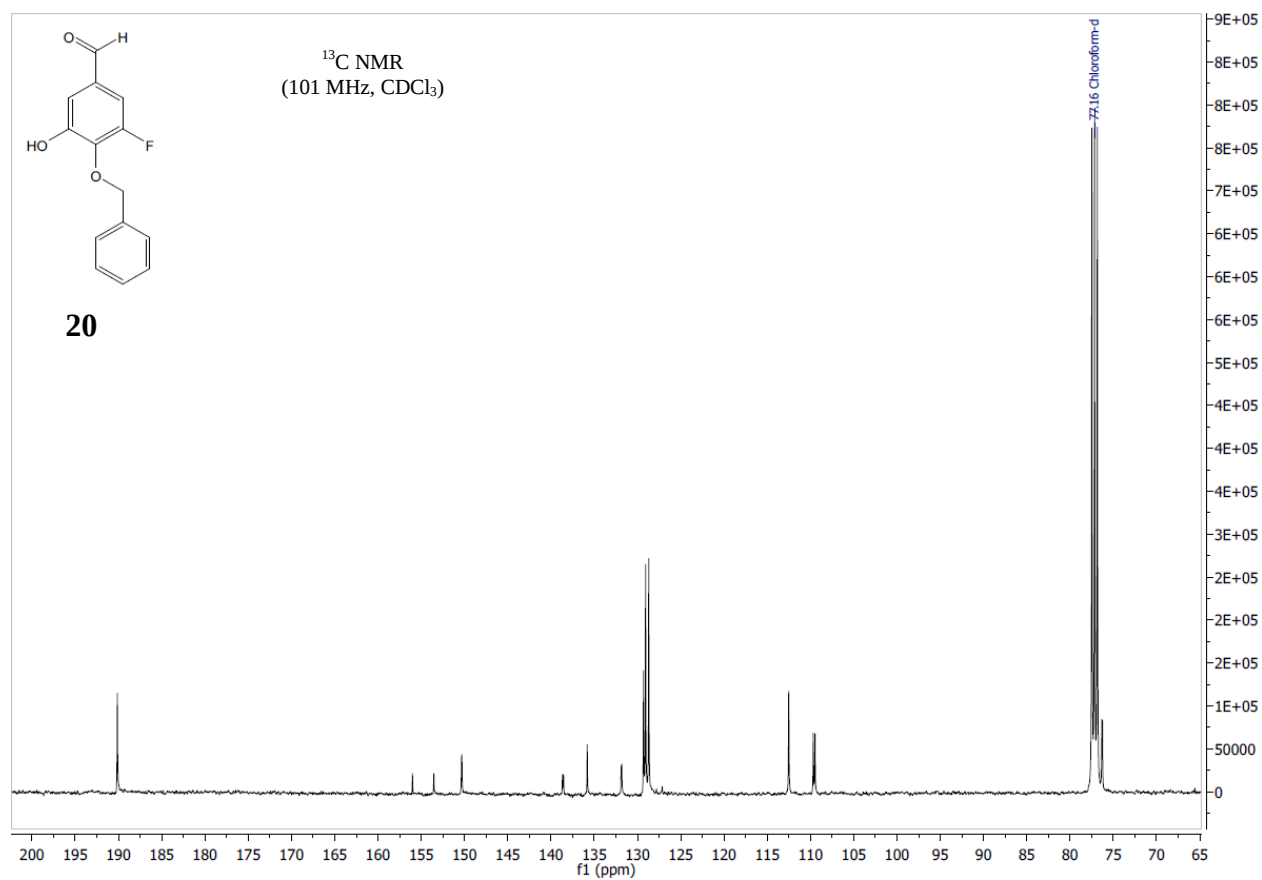
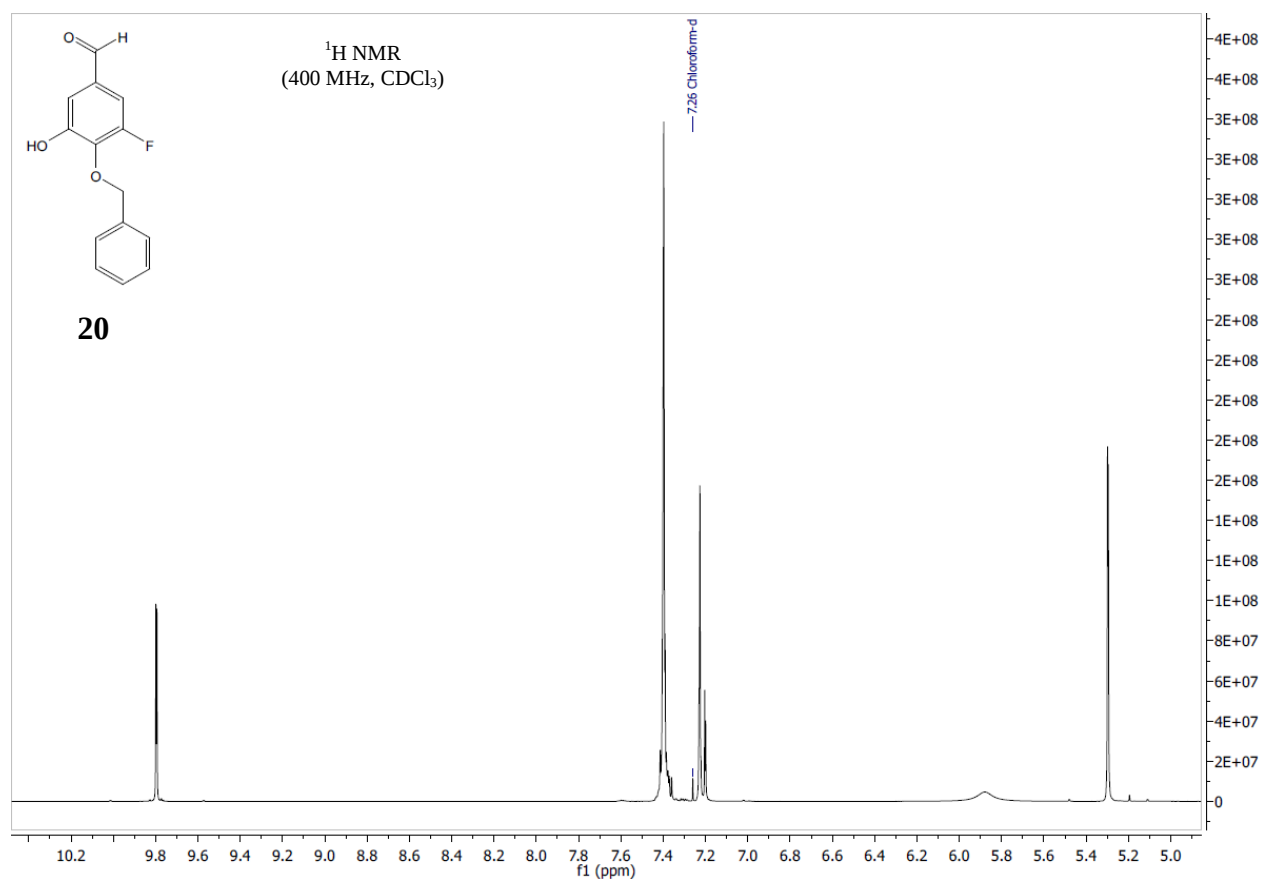


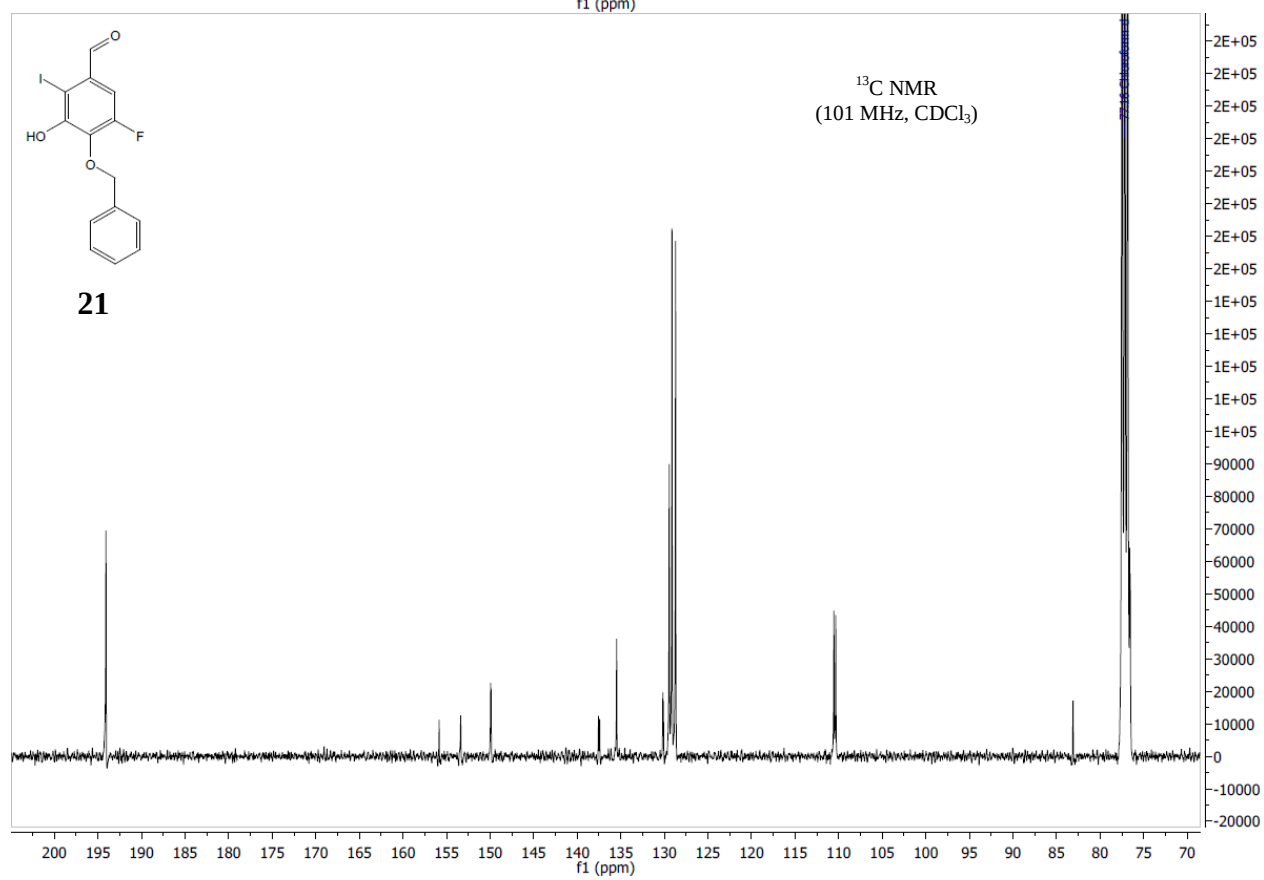
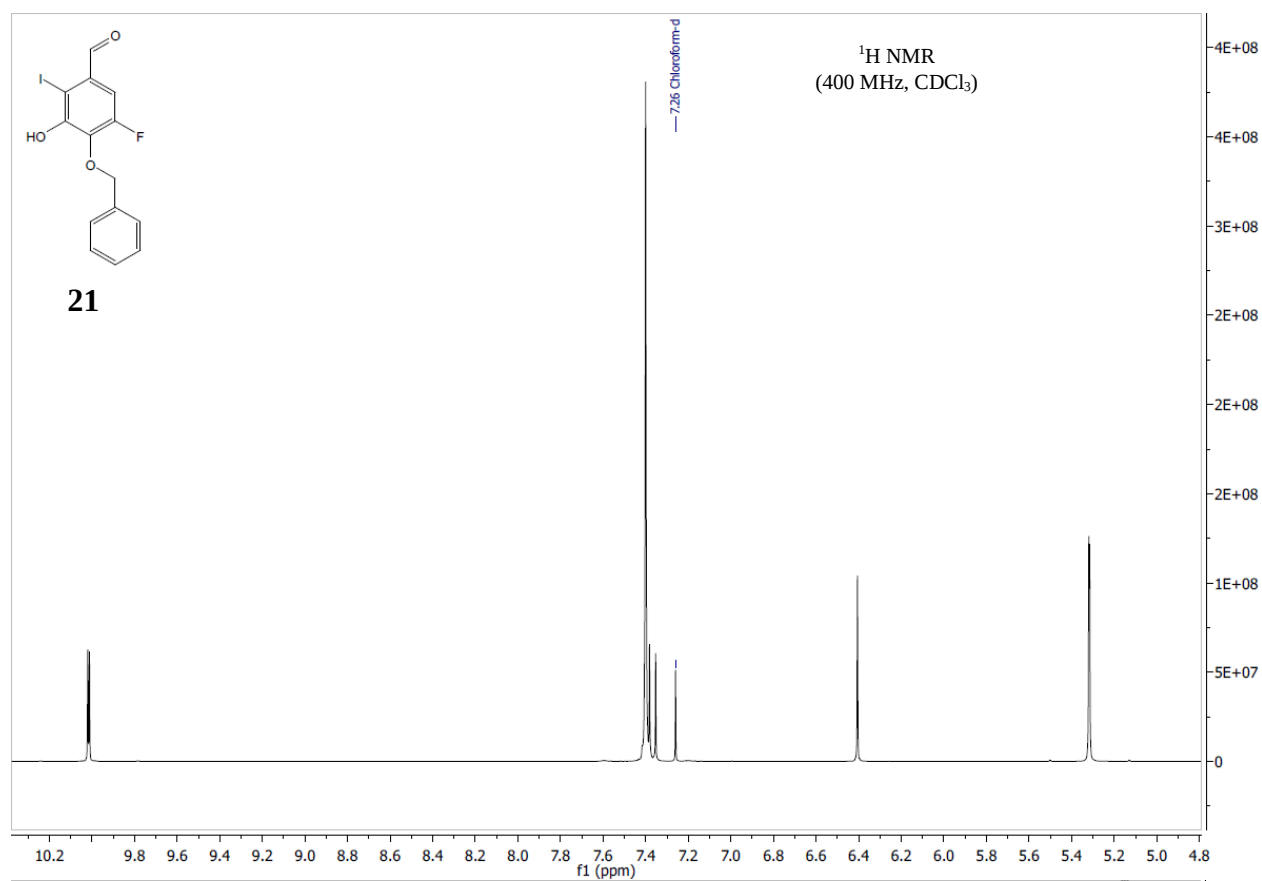


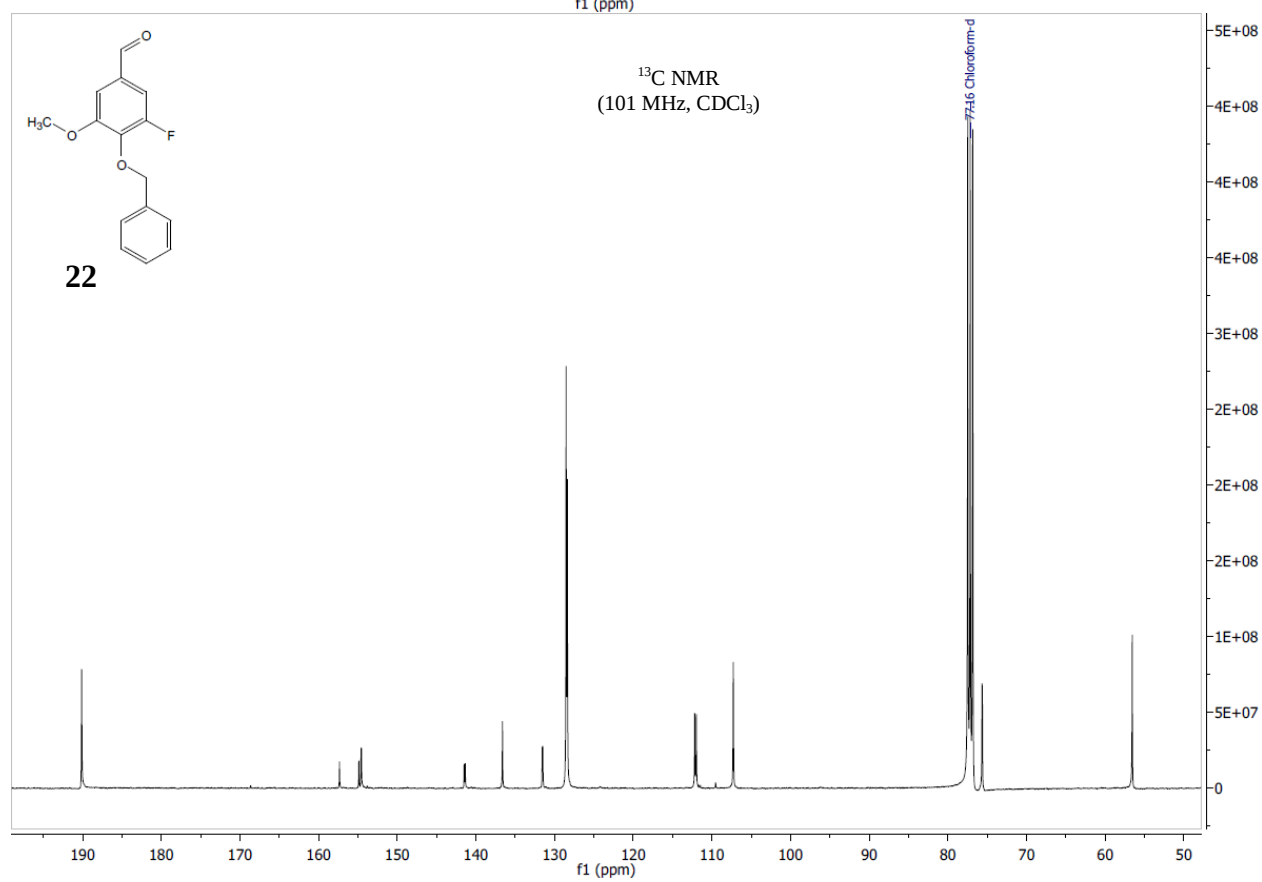
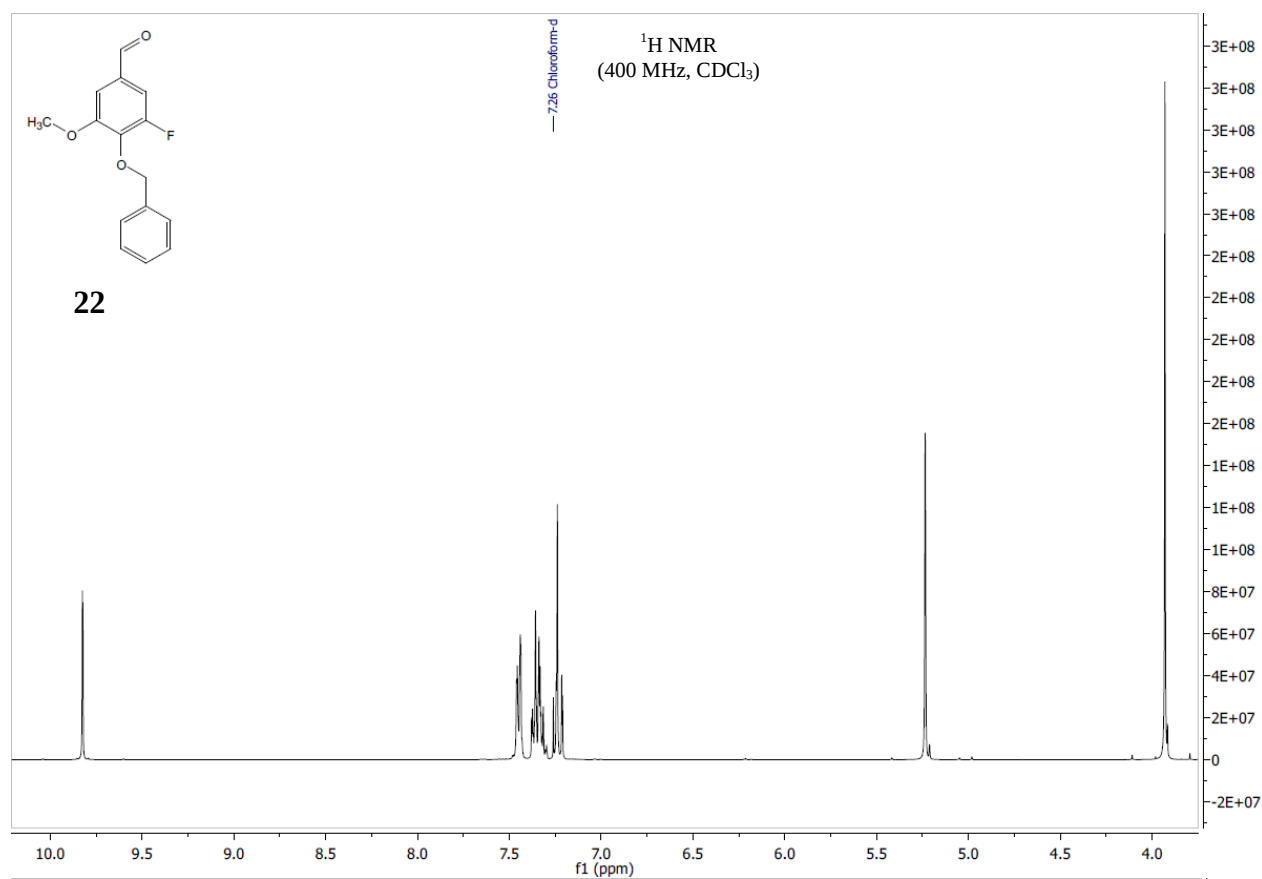


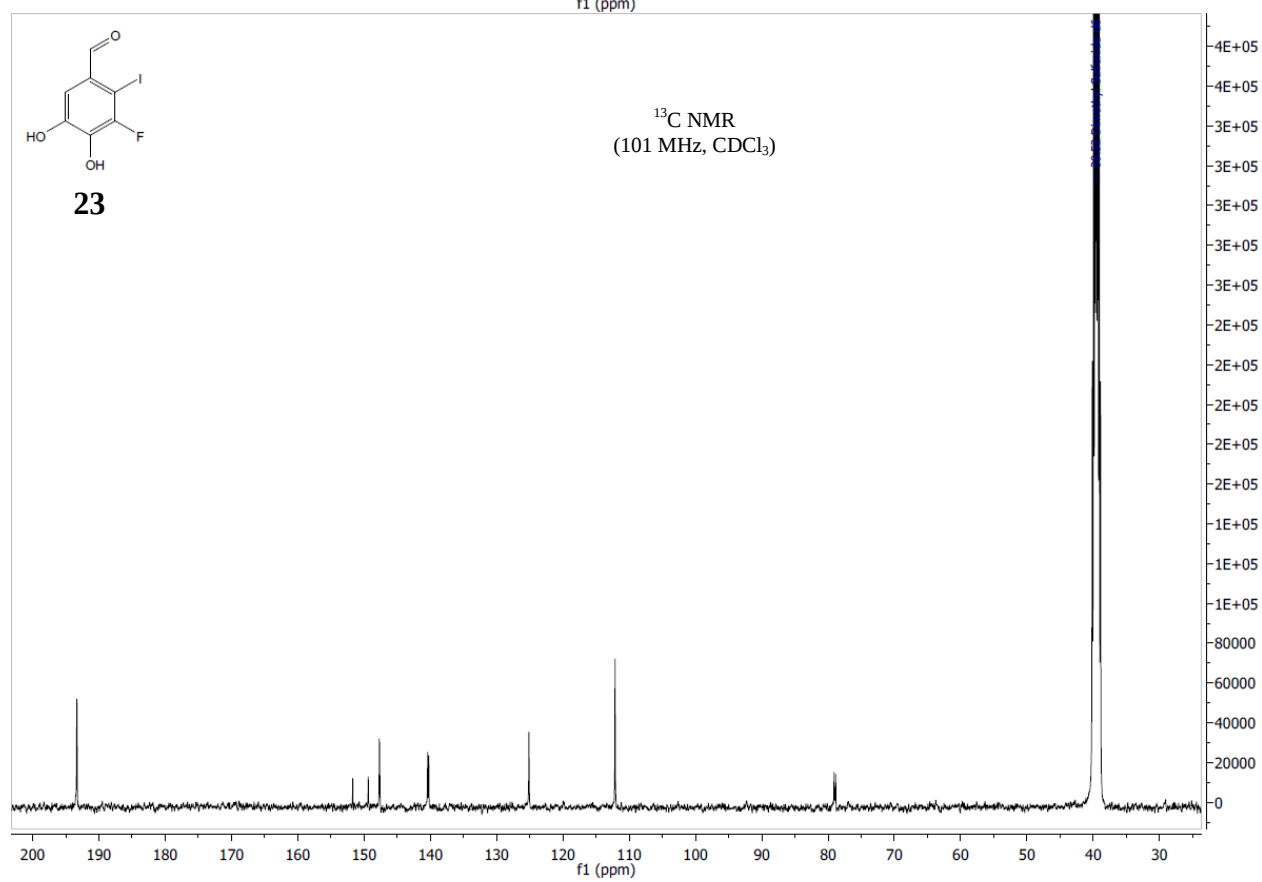
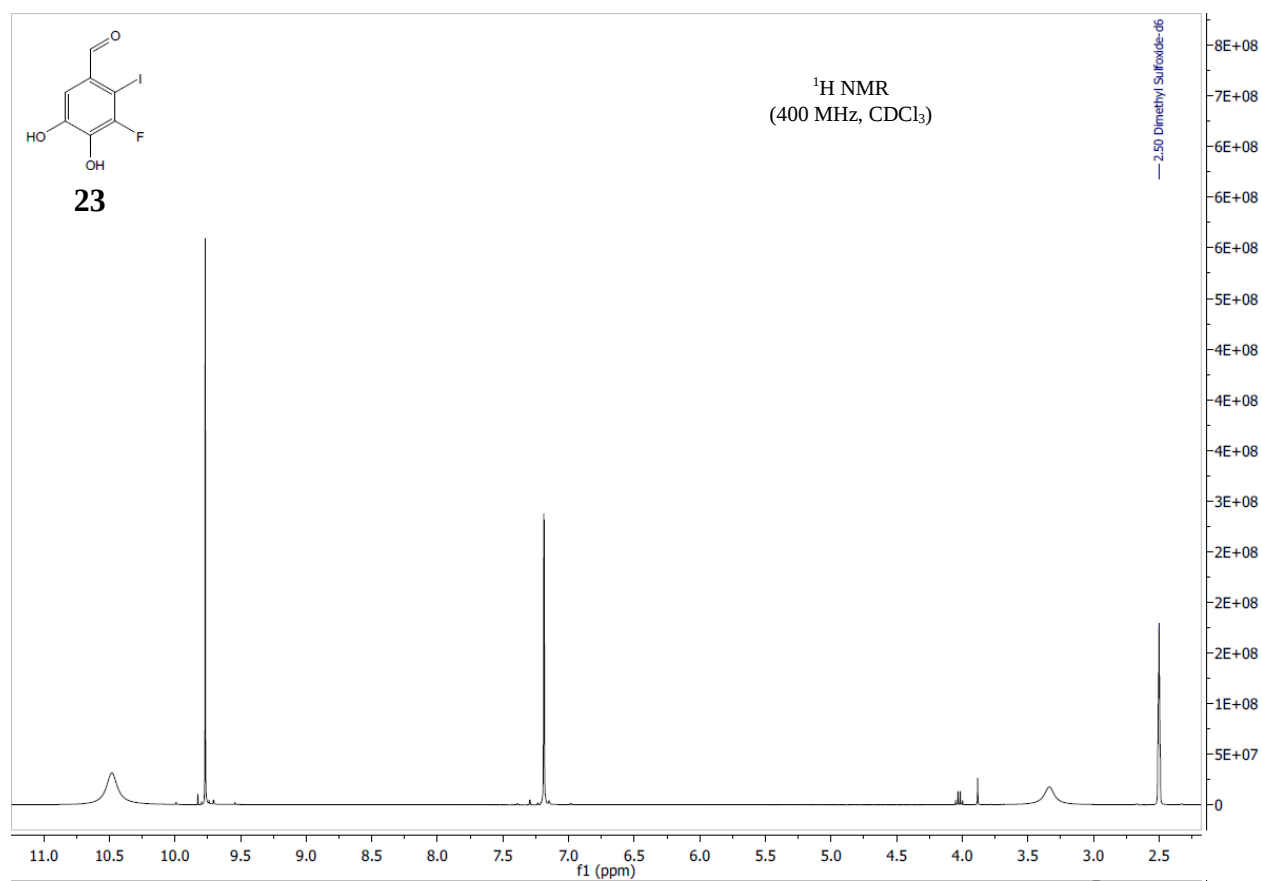


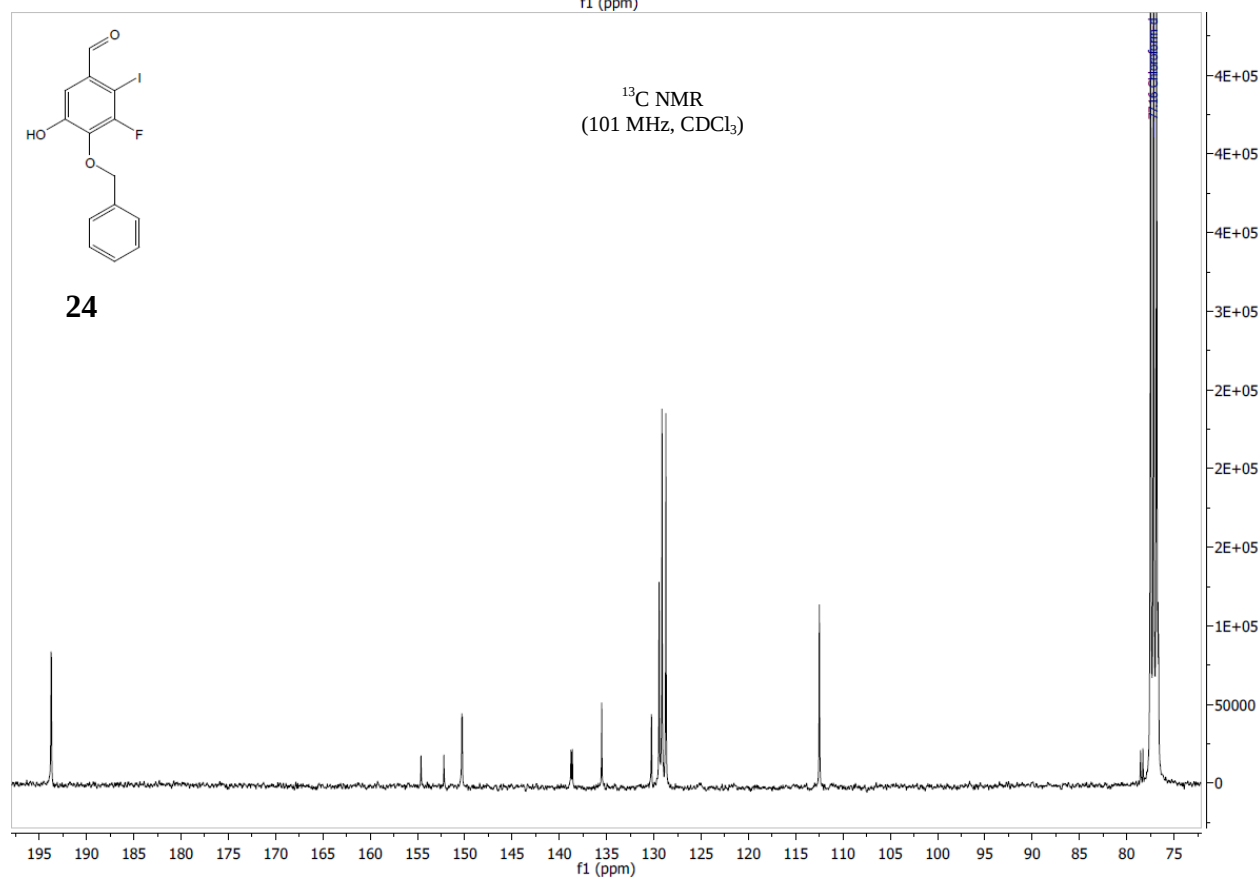
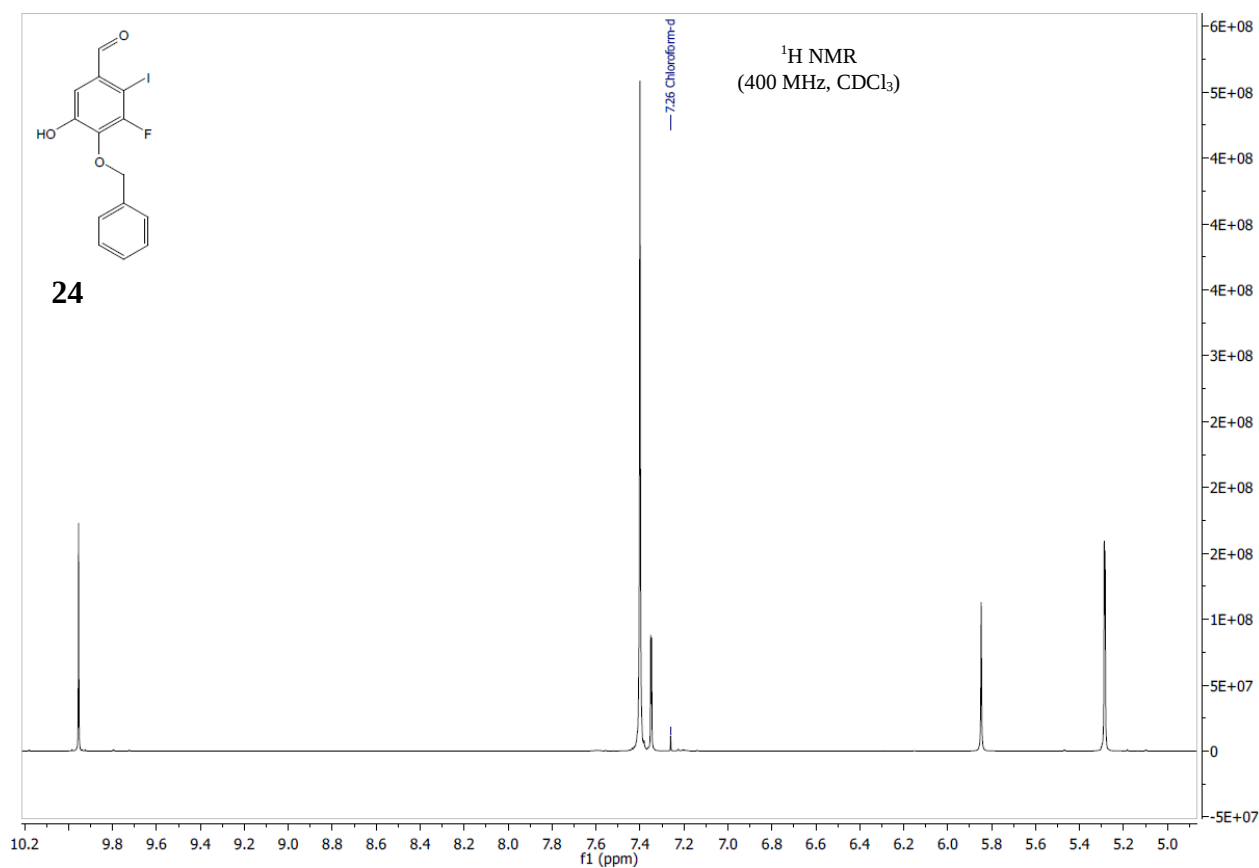












VI. References

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- ⁱ. H. M. Walborsky, M. Topolski, C. Hamdouchi, J. Pankowski, *J. Org. Chem.* **1992** 57, 6188–6191.