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Stimuli-responsive bile acid-based metallogels forming in aqueous

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- 1. Introduction

38 The formation of highly ordered nanostructures from small 39 molecules through non-covalent interactions, such as hydrogen bonding, π - π stacking, metal coordination, solvophobic forces, 40 and van der Waals interactions, has widely been utilized as a bot-41 tom-up approach to design and prepare nanostructured materials 42 43 with interesting properties [1,2]. Of these materials, supramolecular gels derived from low molecular weight gelators have attracted 44 ever increasing attention because of the wide range of applications 45 in which they may find use in our daily life. These applications 46 range from e.g., optoelectronics, light harvesting, and hybrid mate-47 48 rials to tissue engineering, regenerative medicine, and drug delivery, as exhaustively reviewed several times [3–7]. 49

50 The interest in the investigation of metal complexes as supramolecular metallogelators has been increasing only in the past 51 52 decade. The availability and the diversity of metal ligand coordina-53 tion that could readily induce or control the self-assembly process leading to the gel formation and thereby influence the properties of 54 the formed gel has raised the interest towards metallogels. Espe-55 cially the utilization of transition metal complexes as metallogela-56 57 tors has been found to lead to materials exhibiting interesting 58 optical, catalytic, and magnetic properties [8]. In general, the incor-59 poration of the metallic elements into the gel matrix can be

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ABSTRACT

The synthesis and gelation properties of a picolinic acid conjugated bile acid derivative in the presence of metal salts along with the stimuli-responsiveness of the systems are reported. The gels are formed in the presence of Cu²⁺ ions in the solvent systems composed of 30–50% of organic solvent (MeOH, acetonitrile, or acetone) in water. The gels respond to various stimuli: they can be formed upon sonication or shaking, and their gel–sol transformation can be triggered by a variety of chemical species. NMR, MS, and SEM techniques are exploited in order to gain a deeper insight on the self-assembled systems.

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achieved by either physical entrapment of noncoordinating metallic elements or by exploiting the metal-ligand interactions as the major driving force leading to the gel network formation. Metallogels have been shown to be responsive to external stimuli and exploit a range of spectroscopic, magnetic, catalytic, and redox properties that are inherent to the metals they contain.

Steroids are widespread natural products having a large and rigid steroidal nucleus combined with derivatizable functional groups leading to an adjustable polarity profile, which makes them attractive building blocks when designing novel low molecular weight gelators [9]. Bile acids are a group of steroidal compounds biosynthesized in the liver through several complementary pathways. Besides the rigid steroidal nucleus composed of four fused rings they have a short aliphatic side chain containing a carboxylic acid group in its end. In higher vertebrates the rings A and B of the steroidal nucleus are in cis-fusion, causing curvature to the steroidal nucleus, and thus making the molecules facially amphiphilic. The hydroxyl groups are located on the concave α -face, whereas the convex β -face possesses the three methyl groups [10]. Besides as low molecular weight gelators, bile acids and their derivatives have found use in pharmacology [11–13], asymmetric synthesis and chiral discrimination [14], or as receptors for molecular and ionic recognition [15,16], to name a few.

Herein, we report the synthesis and gelation properties of an aminoethyl amide derivative of deoxycholic acid conjugated *via* amide bond with picolinic acid (**4**, Scheme 1) in the presence of Cu²⁺ salts, along with the stimuli-responsiveness of the formed metallogels. Gelation properties of a series of aminoalkyl amides 87



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88 of different bile acids have been systematically studied by us lately 89 [17]. N-(2-aminoethyl)- 3α , 12α -dihydroxy- 5β -cholan-24-amide (**3**, 90 Scheme 1) employed as a starting compound in the current study 91 was not, however, shown to be capable of self-assembly leading to gelation in the conditions studied. Inspired by the free amino 92 93 group of the compound we were prompted to synthesize a new 94 conjugate with the desire of gaining better gelation capabilities. 95 Because picolinic acid (2, Scheme 1) is known to act as a chelating agent in the human body, it was chosen as the compound to be 96 conjugated with the deoxycholyl derivative. Indeed, compound **4** 97 98 was shown to form gels in the presence of Cu²⁺ ions in the solvent systems composed of 30–50% of methanol, acetonitrile, or acetone 99 in water. Moreover, the gels were shown to respond to various 100 stimuli: they could be formed upon sonication or shaking, and 101 102 their gel-sol transformation could be triggered by a variety of 103 chemical species.

104 2. Experimental

105 2.1. Materials

Pyridine-2-carboxylic acid (1) (99%) was purchased from Aldrich. Other reagents used in the synthetic steps as well as the solvents used in chromatography and gelation studies were of analytical grade. Triethylamine and ethyl chloroformate were distilled prior to use. The mixed anhydride method used in the preparation of the target molecule and slightly modified by us has been reported previously [18–20].

113 2.2. Synthesis of compound 4

114 Compound **4** was synthesized by conjugating the freshly pre-115 pared pyridine-2-carboxylic acid anhydride (**2**) with *N*-(2-amino-116 ethyl)- 3α , 12α -dihydroxy- 5β -cholan-24-amide (**3**) prepared 117 according to the previously reported synthetic protocol (Scheme 1) 118 [17].

Synthesis of compound **4** was performed in N₂-atmosphere. In a 119 round-bottomed three-necked 250 mL flask picolinic acid (pyri-120 121 dine-2-carboxylic acid; 5.3 mmol, 1 eq.) and dry dichloromethane 122 (40 mL) were cooled on an ice-water bath to +10 °C, after which tri-123 ethylamine (6.9 mmol, 1.3 eq.) was added to the solution from a 124 dropping funnel, followed by a dropwise addition of ethyl chlorofor-125 mate (6.9 mmol, 1.3 eq.) in dichloromethane (3 mL). The mixture 126 was stirred at rt for 40 min, after which N-(2-aminoethyl)- 3α , 12α -

dihydroxy-5_β-cholan-24-amide (5.3 mmol, 1 eq.) in dichlorometh-127 ane (30 mL) was added dropwise to the freshly prepared picolinic 128 acid anhydride. The stirring was continued for 20 h on an oil bath 129 (40 °C). The crude product obtained after the evaporation of the vol-130 atiles was dissolved in CHCl₃ (100 mL) and washed with water 131 $(2 \times 75 \text{ mL})$, 0.1 M HCl solution $(2 \times 75 \text{ mL})$, water (75 mL), and 132 finally with brine $(2 \times 75 \text{ mL})$. Then the yellowish organic layer 133 was dried (Na₂SO₄), filtered, and the volatiles evaporated under 134 reduced pressure. The crude product was purified by column chro-135 matography (silica gel, CH₂Cl₂:MeOH 90:10) to yield the compound 136 as a white solid. 137

2.3. Compound 4

Yield 84%. ¹H NMR (CDCl₃, 500 MHz, ppm): δ 8.56 (d, 1H, 139 32-CH), 8.38 (m, 1H, NH), 8.18 (d, 1H, 29-CH), 7.86 (m, 1H, 30-140 CH), 7.44 (m, 1H, 31-CH), 6.46 (m, 1H, NH), 3.94 (m, 1H, 12β-H), 141 3.61 (m, 3H, 3β-H + 26-CH₂), 3.49 (m, 2H, 25-CH₂), 2.24 (m, 1H, 142 23α/β-H), 2.09 (m, 1H, 23α/β-H), 1.890.96 (m, 26H), 0.95 (d, 3H, 143 21-CH₃), 0.89 (s, 3H, 19-CH₃), 0.63 (s, 3H, 18-CH₃). ¹³C NMR (CDCl₃, 144 126 MHz, ppm): δ 174.3 (C-24), 165.6 (C-27), 149.5 (C-28), 148.1 145 (C-32), 137.5 (C-30), 126.4 (C-31), 122.4 (C-29), 73.1 (C-12), 71.8 146 (C-3), 48.3 (C-14), 47.1 (C-17), 46.5 (C-13), 42.1 (C-5), 40.4 (C-147 25), 39.4 (C-26), 36.5 (C-4), 36.0 (C-8), 35.3 (C-1), 35.2 (C-20), 148 34.1 (C-10), 33.7 (C-9), 33.4 (C-23), 31.6 (C-22), 30.5 (C-2), 28.7 149 (C-11), 27.4 (C6/7), 27.1 (C6/7), 26.2 (C-16), 23.6 (C-15), 23.1 (C-150 19), 17.4 (C-21), 12.7 (C-18). ESI TOF MS: $[M+Na]^+ m/z = 562$, 151 $[M+K]^+$ m/z = 578. M.W. $(C_{32}H_{49}N_3O_4) = 539.75$. Elemental analy-152 sis: Found C, 70.73; H, 9.13; N, 7.67. Calc. for C₃₂H₄₉N₃O₄ 153 0.25H₂O: C, 70.62; H, 9.17; N, 7.72. 154

2.4. NMR spectroscopy

¹H, ¹³C, ¹³C DEPT-135, and 2D PFG ¹H, ¹³C HMQC and HMBC 156 NMR spectra used for characterization of the prepared compound 157 was recorded with a Bruker Avance DRX 500 MHz spectrometer 158 equipped with a 5 mm diameter broad band inverse detection 159 probehead operating at 500.13 MHz in ¹H and 125.77 MHz in ¹³C 160 experiments, respectively. The ¹H NMR chemical shifts are refer-161 enced to the signal of residual CHCl₃ (7.26 ppm from internal 162 TMS). The ¹³C NMR chemical shifts are referenced to the centre 163 peak of the solvent CDCl₃ (77.0 ppm from internal TMS). A compos-164 ite pulse decoupling, Waltz-16, has been used to remove proton 165 couplings from ¹³C NMR spectra. Assignment of the ¹³C NMR 166



Scheme 1. Synthesis route leading to compound 4.

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167 chemical shifts has been made with the help of 2D NMR spectra 168 and literature [21].

the sample stub, and after drying in air for two days, coated with 190 191

2.5. Gelation studies 169

Gelation tests for compound 4 in the presence of metal salts 170 were performed as follows: 5 mg of compound 4 was weighed in 171 172 a test tube and 150-250 µL of MeOH/MeCN/acetone was added. 173 After that, depending on the solvent composition, 50-225 µL of 174 deionized water was added. Finally, depending on the desired 175 ligand:metal molar ratio (2:1, 1:1, or 1:2), 125 µL or 250 µL of aqueous metal salt solution was added, resulting in the total sol-176 177 vent volume of 500 µL. The obtained suspension or solution was then subjected to ultrasound for ca. 1 min and heated with a heat 178 gun until a clear mixture was obtained. The solution was allowed 179 to cool, after which observations with regard to gelation were 180 made. The test tube was inverted, and if the solid mass did not 181 182 move, it was defined as a gel. If the gel was formed already while sonicating, no heating was performed. 183

2.6. Microscopy 184

185 Scanning electron micrographs were taken with Bruker Quan-186 tax400 EDS microscope equipped with a digital camera. Samples of the xerogels were prepared either by placing a hot, clear solution 187 of the gelator and metal salt in an appropriate solvent mixture or a 188 scooped sample of the preformed gel on carbon tape placed over 189

Table 1

properties of varying $\mathbf{4}$ ·Cu²⁺ molar ratios in aqueous solvents Ω^2

gold in a JEOL Fine Coat Ion Sputter JFC-1100.

2.7. Mass spectrometry

The mass spectrometric experiments were performed with a 193 QSTAR Elite ESI-Q-TOF mass spectrometer equipped with an API 194 200 TurboIonSpray ESI source from AB Sciex (former MDS Sciex) 195 in Concord, Ontario (Canada). The gel samples were prepared by 196 utilizing ligand:Cu²⁺ molar ratio of 3:1 and 1:1 and 30% of MeOH 197 as the co-solvent. In order to investigate the effect of the counter 198 anion both CuSO₄ 5H₂O and Cu(NO₃)₂ 3H₂O were exploited as 199 the copper source. For the measurements, the gel-phase samples 200 were dissolved and diluted in MeCN to obtain 9.25 µM samples 201 for measurements. 202

3. Results and discussion

Compound **4** was synthesized with an overall yield of 84% by 204 modifying the well-known synthesis method reported in the liter-205 ature (Scheme 1). Conventional analysis methods were utilized in 206 order to prove the identity, and the full data are provided in the 207 experimental section. Compound 4 itself did not form gels in any 208 of the common solvents tested; in general, it was shown to be 209 rather poorly soluble in aromatic solvents, but then apparently 210 too soluble in more polar solvents. 211

	Solvent composition	Ligand:Cu ²⁺ molar r	Ligand:Cu ²⁺ molar ratio		
		2:1	1:1	1:2	
CuSO4-5H2O	MeOH 50%	Gs	S	S	
	MeOH 40%	Gs	Gs	Gs	
	MeOH 30%	Gs	Gs	Gs	
	MeCN 50%	S	S	S	
	MeCN 40%	S	S	S	
	MeCN 30%	G	G	G ^t	
	Acetone 50%	$Gs \rightarrow C$	$PG \rightarrow C$	$PG \rightarrow$	
	Acetone 40%	Gs	Gs	$Gs \rightarrow$	
	Acetone 30%	Gs	Gs	Gs	
Cu(NO ₃) ₂ .3H ₂ O	MeOH 50%	S	S	S	
	MeOH 40%	S	$S \rightarrow PG$	$S \rightarrow P$	
	MeOH 30%	PG	Gs	Gs	
	MeCN 50%	S	S	S	
	MeCN 40%	S	S	S	
	MeCN 30%	S	S	S	
	Acetone 50%	S	S	S	
	Acetone 40%	S	S	S	
	Acetone 30%	S	G	Gs	

G = gel formed after heating and cooling; Gs = gel formed after sonication; G^t = transparent gel formed after heating and cooling; PG = partial gel; PG = partial weak gel; S = clear blue solution; C = crystallizes.



Fig. 1. Sonication-induced gel formation of compound **4** with $CuSO_4$ (1:1) in aqueous MeOH (30%).

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212 Compound 4 bearing a pyridine group, a classic metal-coordina-213 tion site, along with the aminoethyl amide linker between the 214 deoxycholic acid and picolinic acid was anticipated to be able to interact with metal cations. Intrigued by the possibility of obtain-215 ing metallogels Cu²⁺, Ag⁺, and Zn²⁺ cations were tested. Cu²⁺ was 216 shown to give the most promising results in the preliminary tests, 217 and was thus chosen to be the metal cation concentrated on. Three 218 different Cu²⁺ salts, namely CuSO₄·5H₂O, Cu(NO₃)₂·3H₂O, and 219 CuCl₂·2H₂O were employed. The gels were shown to be formed 220 in water with 50-30% of methanol, acetonitrile, or acetone as the 221

co-solvent according to the Table 1. Since hydrogels are better222compatible with bodily tissues than organogels, the water content223of the solvent mixture was increased to as high a level as possible.224If it exceeded 70% precipitation instead of gel formation occurred.225

The counter anion of the copper salt was shown to play a role in the gel formation. The different properties of the copper salts, for example in terms of complex formation in aqueous solutions, may have affected on the behavior of the systems. In the studied conditions, $CuSO_4 \cdot 5H_2O$ was shown to promote gel formation more often than $Cu(NO_3)_2 \cdot 3H_2O$. Additionally, in the case of $Cu(NO_3)_2$. 231



Fig. 2. Appearance of the samples with ligand:Cu²⁺ molar ratio of 1:1 in aqueous solvents (30% of organic solvent).



Fig. 3. Illustration showing the gel-sol transformation of Cu²⁺-gels of compound 4 by the addition of external chemical stimuli.

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Fig. 4. Illustration of the reversible gel-sol transformation induced by the reduction of Cu²⁺ to Cu⁺ by ascorbic acid and subsequent oxidation back to Cu²⁺ by HNO₃.



Fig. 5. SEM images of the xerogels of compound 4 formed in the presence of Cu^{2+} . Samples prepared by scooping a portion of the preformed gel on a sample stub. (A) Ligand: Cu^{2+} 2:1 gel in MeOH, (B) Ligand: Cu^{2+} 2:1 gel in MeOH, (C) Ligand: Cu^{2+} 2:1 gel in acetone, (D) Ligand: Cu^{2+} 1:1 gel in MeOH, (E) Ligand:Cu

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·3H₂O, in most of the cases ligand:Cu²⁺ molar ratio of 1:1 or 1:2 232 233 was required for the gel formation, and the gels were only formed 234 in rather high water concentrations. However, the gels formed 235 with $Cu(NO_3)_2 \cdot 3H_2O$ were shown to be rather stable, whereas the gels formed with CuSO₄·5H₂O were shown to crystallize/precipi-236 tate moderately quickly. CuCl₂·2H₂O, however, was only able to 237 238 give clear blue solutions; no gels were detected. Interestingly, very recently Džolić and Cametti with their co-workers reported the 239 metallogel formation of pyridyloxalamide in alcohols in the pres-240 ence of only $CuCl_2$ of several other Cu^{2+} salts tested [22]. 241

Gelation was shown to take place by solely sonication (denoted 242 Gs in Table 1, Fig. 1) in several cases. If sonication was not suffi-243 cient enough in order to promote gelation, the traditional heating 244 and cooling-cycle was performed. Interestingly, in some cases the 245 246 gelation of the clear blue solution obtained after the heating and 247 cooling/settling period of one day could be induced by vigorous shaking. Similar phenomenon of mechanically triggered gel forma-248 tion has been recently reported by us [23] and others [24]. In most 249 of the cases rather translucent light blue gels were obtained 250 (Fig. 2).

The gel systems obtained were shown to act as responsive soft materials responding to a variety of chemical stimuli (Figs. 3 and 4). Three drops of pyridine, triethylamine, or aqueous ammonia or small amount of solid disodium salt of EDTA (ethylenediamine tetra-acetic acid) were added onto the surface of a preformed gel. The upper layer of the gel was immediately shown to transform into a solution, and after a gentle agitation, the gel-sol transformation was completed. In the case of pyridine an intensively blue-colored clear solution was obtained (Fig. 3).

The gels also showed reversible redox-responsiveness (Fig. 4): 261 the Cu²⁺ ions could be reduced to Cu⁺ by adding ascorbic acid 262 (1 eq. in 20 µL of water) onto the preformed gel. The upper layer 263 264 of the gel was immediately shown to transform into solution. In 265 order to speed up the process the sample was heated until a clear 266 yellow solution was obtained. When cooled down, the clear yellow solution first became opaque and finally after some shaking a light 267 green solution with some green precipitate was obtained. In order 268 to oxidize Cu^+ again to Cu^{2+} 20 µL of 0.5 M nitric acid was added to 269 the solution, which was then heated until a clear light blue solution 270 was obtained. This solution was finally sonicated in order to revive 271 the gel. 272

In order to gain some visual insight into the microscopic morphology of the dried self-assembled materials the scanning electron microscopy (SEM) was utilized. The xerogel samples for the SEM were prepared by following two different protocols: in the first one, the gel was formed by the heating-cooling cycle and allowed to stabilize, after which the preformed gel was scooped onto a sample stub covered with the carbon tape (Fig. 5). In the second experiment, the clear hot solution containing the ligand, copper salt, and the aqueous solvent was pipetted onto the sample stub and allowed to dry (Fig. 6). As expected, and observed by us previously [23], as well, differences in the sample appearance between these two sample preparation protocols were observed. The sample appearance, however, was shown to be independent of the ligand:Cu²⁺ molar ratio, or the solvent system used.

From the gelation experiments it was clear that Cu²⁺ was essential for gel formation. The role of the cation in the process was further investigated by utilizing NMR spectroscopy. First, the ¹H NMR spectra were measured in absence of the cation for samples of 0.17%, 0.5%, and 1% in concentration (4 in MeOD: D_2O mixture). The spectra appeared well-resolved. Increasing of the concentration did not have any particular effect on the values of the chemical shifts. From the spectra it was obvious that there was no gel formed. When the cation was added to the sample solutions, the overall resolution and S/N in all spectra dramatically decreased. The most probable explanation is, naturally, the paramagnetic nature of Cu²⁺.

The spectra were measured for ligand:Cu²⁺ 1:1 samples of 299 0.17%, 0.5% (gel), and 1% (gel) in concentration (**4**:Cu²⁺ in 300 MeOD:D₂O mixture). Increasing of the concentration resulted in 301



Fig. 6. SEM images of the xerogels of compound 4 formed in the presence of Cu²⁺. Samples prepared from a hot clear solution. (A) Ligand:Cu²⁺ 2:1 in MeOH, (B) Ligand:Cu²⁺ 1:1 in MeOH, (C and D) Ligand:Cu²⁺ 1:1 in acetone. Scale bars: (Å) 2 μm, (B) 20 μm, (C) 20 μm, (D) 2 μm.

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Fig. 7. Samples with 1% of 4 in MeOD with varied ligand: Cu^{2+} molar ratios: 10:1 (undermost) \rightarrow 10:2 (5:1) \rightarrow 10:3 \rightarrow 10:4 (5:2) \rightarrow 10:5 (2:1) uppermost.

302 gel formation, which was evident from the ever broadening signals of ¹H NMR spectra due to restriction of molecular tumbling as a 303 result of gel formation. The ligand:Cu²⁺ molar ratio was further 304 increased to 1:2, and the measurements repeated for the same 305 set of concentrations. The appearances of the spectra were close 306 to identical, when compared with the ligand:Cu²⁺ molar ratio of 307 1:1. When the temperature was increased from 30 °C to 60 °C in 308 5 °C temperature steps for the 1% sample of ligand:Cu²⁺ in a molar 309 ratio of 1:2, the breakdown of the gel network could be deduced 310 311 from the ever improving resolution of the ¹H NMR spectra. The 312 pyridine protons became visible and were observed to move 313 towards the upper field region as the gel was melting (*i.e.*, the temperature was increased). This indicates pyridine playing an impor-314 tant role in the process leading to gel formation, most probably by 315 316 chelating Cu²⁺ ions. When the gel-forming network breaks down the Cu²⁺ cation does not interact with pyridine as intensively, 317 and consequently withdrawal of the pyridine electrons by the 318 positively charged cation diminishes. This results in decreased 319 320 deshielding, which can be seen as movement of the chemical shift to the upper field. 321

Next, the molar ratio of copper to ligand was increased from 322 323 1:10 to 1:2. Due to the simultaneous effects of unfavorable NMR properties of copper and restricted molecular tumbling due to gel 324 formation, not much but the decrease in resolution as a conse-325 326 quence of the increasing amount of copper could be observed. The measurements were further repeated in pure MeOD, a solvent 327 which did not induce gel formation. Paramagnetic properties of 328 Cu²⁺ affected on the appearance of the spectrum, but when the 329 amount of Cu²⁺ was increased, the pyridine protons broadened 330 and moved to downfield direction (Fig. 7). This further confirms 331 the role of pyridine chelating copper as the key process in the gel 332 333 formation. A tendency of the 12 β proton on the steroidal skeleton to move to downfield direction was observed as well. 334

335 In addition to the role we were also interested in the coordina-336 tion degree of copper in the current gel-forming system. Encouraged by the work of Bunzen and coworkers [25], the complex 337 responsible for gelation was tried to be determined by using mass 338 spectrometry. The samples were prepared by using ligand:Cu²⁺ 339 340 molar ratios of 3:1, 2:1, and 1:1 and 30% of MeOH as the co-sol-341 vent. In order to investigate the effect of the counter anion both 342 sulfate and nitrate were exploited as the copper source. For the measurements, the gel samples were further dissolved and diluted in MeCN to obtain 9.25 µM samples for measurements.

Unfortunately, for compound 4 the results were not straightforward. For each sample, a collection of different ligand:metal adducts were observed in the spectrum. While copper undoubtedly is essential to gel formation, based on the mass spectrometric experiments a certain ligand:copper complex inducing the gel formation is not likely - rather several different adducts synergetically end in gelation.

4. Conclusions

In this work we report the synthesis and structural analysis of an aminoethyl amide derivative of deoxycholic acid conjugated via amide bond with picolinic acid. Furthermore, the compound is shown to be able to form gels in the presence of Cu^{2+} ions in the solvent systems composed of 30–50% of methanol. acetonitrile. or acetone in water. The formed gels are shown to respond to various stimuli: they can be formed upon sonication or shaking, and their gel-sol transformation can be triggered by a variety of chemical species. The gels also show reversible redox-responsiveness: the Cu²⁺ ions can be reduced to Cu⁺ followed by gel-sol transition. Oxidation of the Cu^+ ions again to Cu^{2+} revives the gel. The essential role of the pyridine moiety chelating the Cu²⁺ ions in the process leading to gel formation was proved by NMR experiments. Mass spectrometry and SEM techniques were used in further characterization the systems.

The important physiological roles of bile acids and picolinic acid combined with the formation of stimuli-responsive gels in aqueous media responding to several stimuli may in the future lead to fascinating biomedical applications, especially due to the neuroprotective, immunological, and anti-proliferative activities of picolinic acid.

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