

NJC

Accepted Manuscript



This article can be cited before page numbers have been issued, to do this please use: C. Yu, H. Chang and T. Chien, *New J. Chem.*, 2019, DOI: 10.1039/C9NJ01012B.



This is an Accepted Manuscript, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about Accepted Manuscripts in the [author guidelines](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the ethical guidelines, outlined in our [author and reviewer resource centre](#), still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this Accepted Manuscript or any consequences arising from the use of any information it contains.

ARTICLE

Total Synthesis of Pseudouridine *via* Heck-type C-Glycosylation[†]Cheng-Ping Yu,^a Hsin-Yun Chang,^a and Tun-Cheng Chien^{*,a,b}Received 00th January 20xx,
Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

The reaction of 2,4-dimethoxy-5-iodopyrimidine (**8**) and 3,5-di-*O*-*tert*-butyldimethyl protected ribofuranoid glycal **4** was carried out with Pd(OAc)₂ as the catalyst, PPh₃ as the ligand and Et₃N as the base in DMF at 70 °C followed by desilylation to afford exclusively the β-anomer of 5-(2,3-dideoxy-3-oxoribofuranosyl)-2,4-dimethoxypyrimidine (**11**) in a very good yield. Subsequent protecting group and functional group interconversions furnished pseudouridine (**Ψ**, **1**).

Introduction

Pseudouridine (**Ψ**, **1**), isolated from RNA mixture in 1957¹ and characterized in 1961,^{2,3} is the first C-nucleoside found in nature. It is the fifth nucleoside component in RNA and the most abundant natural C-nucleoside, which, therefore, received significant attention. Pseudouridine is the structural isomer of uridine with identical uracil base and ribosyl sugar moiety but differ only in the nucleosidic linkages (**Figure 1**). It is formed by post-translational isomerization from uridine in RNA catalyzed by pseudouridine synthase.⁴ Albeit decades of efforts have been devoted, the role of pseudouridine in RNA still remains to be further clarified.⁵ One of the major restrictions for the studies is due to the very limited commercial availability and expensive prices of pseudouridine. Thus, chemical synthesis of pseudouridine continues to offer an alternative supply for this biological important molecule. Nevertheless, the chemical synthesis of C-nucleosides (and also C-glycosides) is perceived to be challenging and the development of efficient syntheses for C-nucleosides is still an ongoing task.⁶⁻⁸

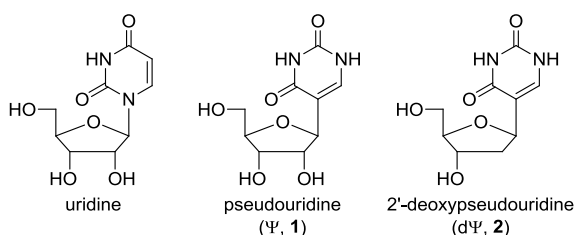
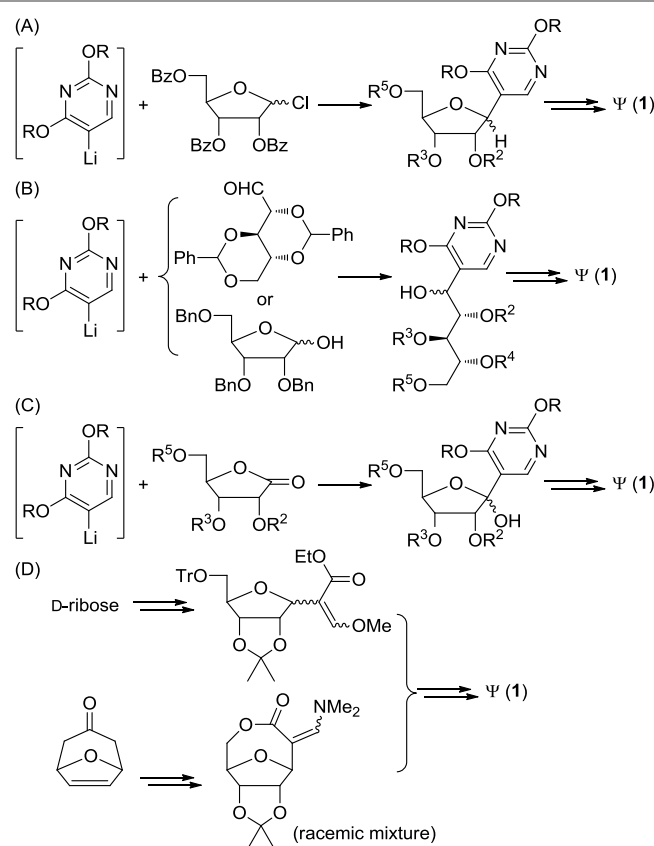


Figure 1. Uridine, pseudouridine (**Ψ**, **1**) and 2'-deoxypseudouridine (**dΨ**, **2**)



Scheme 1. Synthetic strategies toward pseudouridine (**1**)

A perusal of literature revealed that the previous synthesis of pseudouridine (**1**) was achieved mainly by four different strategies summarized in **Scheme 1**. (A) The first synthesis of pseudouridine was accomplished by Shapiro and Chambers in 1961. They exploited the substitution reaction between 5-lithiated 2,4-dimethoxypyrimidine and 2,3,5-tri-*O*-benzoylribofuranosyl chloride followed by acid-hydrolysis to afford pseudouridine.³ (B) Later on, the addition reaction of 5-lithiated 2,4-dialkoxypyrimidine to hydroxyl-protected ribose followed by global deprotection and acid-promoted dehydrative cyclization became an early approach for the

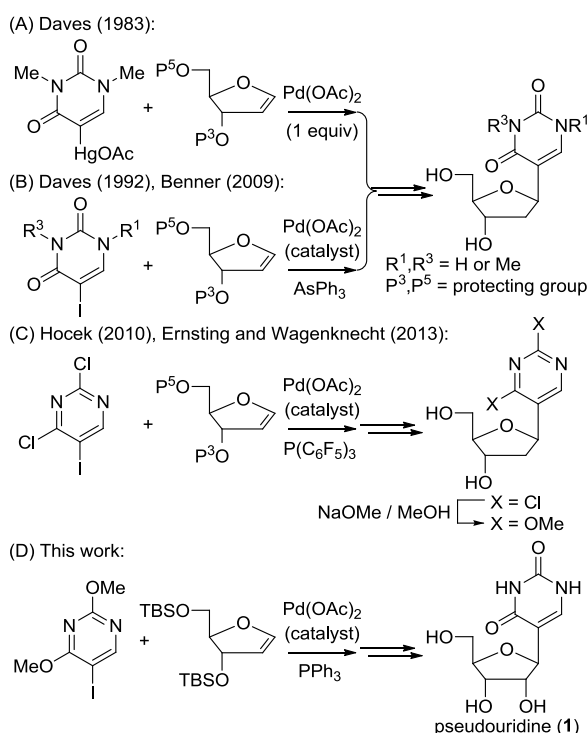
^a Department of Chemistry, National Taiwan Normal University, Taipei 11677, Taiwan. Email: tcchien@ntnu.edu.tw

^b Faculty of Pharmacy, National Yang-Ming University, Taipei 11221, Taiwan.

[†] This paper is dedicated to Professor Ji-Wang Chern on the occasion of his 65th birthday and retirement from National Taiwan University.

Electronic Supplementary Information (ESI) available: [copies of ¹H and ¹³C NMR spectra]. See DOI: 10.1039/x0xx00000x

synthesis of pseudouridine.⁹ However, both approaches (A) and (B) resulted in the formation of α/β -isomers in moderate yields accompanied with a minimal amount of pyranose-isomers. (C) The addition reaction of 5-lithiated 2,4-dialkoxy pyrimidine to hydroxyl-protected ribonolactone was found to be a superior method. The addition reaction formed an anomeric mixture of ribonolactol and subsequent reduction followed by deprotection rendered the synthesis of pseudouridine.¹⁰⁻¹³ It is noteworthy that the synthesis of C-nucleosides *via* the ribonolactone approach typically gives a mixture of α/β -stereoisomers. However, an improved protocol developed by Hanessian *et al.* allowed the synthesis of pseudouridine to be accomplished in an excellent β -stereoselectivity.¹¹⁻¹³ (D) Alternatively, pseudouridine can also be synthesized *via de novo* construction of the heterocyclic aglycon upon the ribofuranosyl precursors equipped with functionalized substituents at the anomeric position. This approach was demonstrated by Watanabe¹⁴ and Noyori,^{15,16} respectively.



Scheme 2. Heck-type glycosylation of uracil/pyrimidine derivatives

Over the past three decades, palladium-catalyzed Heck-type reaction of ribofuranoid glycols with (het)aryl iodides has been developed as a promising approach for the synthesis of 2'-deoxy-C-nucleosides.⁸ The pioneering work was undertaken by Daves who first reported that the reaction of (1,3-dimethyluracil-5-yl)mercury(II) acetate with ribofuranoid glycols under stoichiometric amount of $\text{Pd}(\text{OAc})_2$ led to the formation of 1,3-dimethyl-2'-deoxypseudouridine (**Scheme 2 (A)**).¹⁷ Their continuous efforts led them to develop the reaction into a practical approach which utilizes palladium-catalyzed Heck cross-coupling reaction of aryl iodides with ribofuranoid glycols for the synthesis of 2'-deoxy-C-nucleosides

with remarkable β -stereoselectivity. In particular, Zhang and Daves demonstrated that 2'-deoxypseudouridine (**2**) can be prepared directly from 5-iodouracil and 3'-TBDPS-protected furanoid glycol under the catalysis of $\text{Pd}(\text{OAc})_2$ and AsPh_3 (**Scheme 2 (B)**).^{18,19} Since then, the Heck-type C-glycosylation has been extensively applied to the synthesis of a variety of (het)arene 2'-deoxy-C-nucleosides.^{20,21}

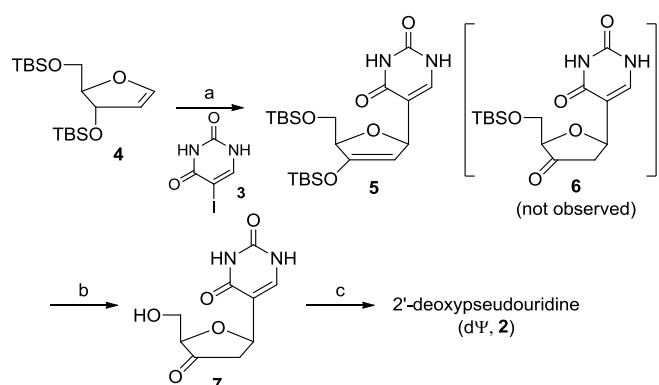
Nevertheless, it came to our attention that, besides 2'-deoxypseudouridine (**2**), several C5-(2'-deoxyribofuranosyl)pyrimidine derivatives have also been prepared by this method (**Scheme 2 (C)**).^{17-19,22} Despite they are structurally related to 2'-deoxypseudouridine (**2**), further elaboration of these pyrimidine 2'-deoxy-C-nucleosides into pseudouridine (**1**) has not been explored. Thus, we were prompted to investigate whether the palladium-catalyzed Heck-type glycosylation of 5-iodouracil derivatives followed by a sequential chemical manipulation would be a general and feasible strategy for the preparation of pseudouridine (**1**) and its derivatives (**Scheme 2 (D)**).

Results and discussion

In our attempts to study chemical elaboration of 2'-deoxypseudouridine derivatives into pseudouridine (**1**), the synthesis of 2'-deoxypseudouridine (**2**) based on the Heck reaction approach developed by Zhang and Daves was first conducted.¹⁹ It is worth to mention that the Heck-type glycosylation usually required the use of triphenylarsine or tris(pentafluorophenyl)phosphine as the ligand in the reaction in order to achieve high stereoselectivities and yields. Since we first aimed to avoid the use of toxic arsine or expensive phosphine ligands, the reaction of 5-iodouracil (**3**) with 3,5-di-*O*-*tert*-butyldimethyl protected ribofuranoid glycol (**4**),²³ prepared from thymidine by literature procedures, was subjected to optimization with various catalysts, ligands and bases. The survey of reaction conditions showed that the Heck glycosylation adduct **5** was obtained in the maximum yield as a single stereoisomer when the reaction was carried out in DMF at 70 °C with $\text{Pd}_2(\text{dba})_3$ as the catalyst, XantPhos as the ligand and *n*-Bu₃N as the base (**Table S1** in Supporting Information). The removal of TBS groups was then accomplished by TBAF with AcOH to form the 2'-deoxy-3'-oxo-C-ribonucleoside **7**.¹⁹ With the directing effect from 5'-hydroxy group, the 3'-keto group of **7** was stereoselectively reduced by $\text{NaB}(\text{OAc})_3\text{H}$ from the β -face to give 2'-deoxypseudouridine (**2**) as a single stereoisomer in an excellent yield (**Scheme 3**).^{19,24,25}

Literature survey revealed that transformations of silyl enol ethers from Heck glycosylation into ribofuranose C-nucleosides can be achieved mainly by three strategies, including the direct oxidation with osmium tetroxide²⁶ or dimethyldioxirane²⁷ of the silyl enol ethers to give the α -hydroxyketone, and hydroboration-oxidation of the silyl enol ethers to form the 2',3'-*trans*-diols.²⁸ However, our attempts to further elaborate 2'-deoxypseudouridine (**2**) or its silyl enol ether derivative **5** to pseudouridine (**1**) based on the above mentioned approaches were failed. We speculated that this failure could be attributed to the interference from the unprotected nitrogens/oxygens

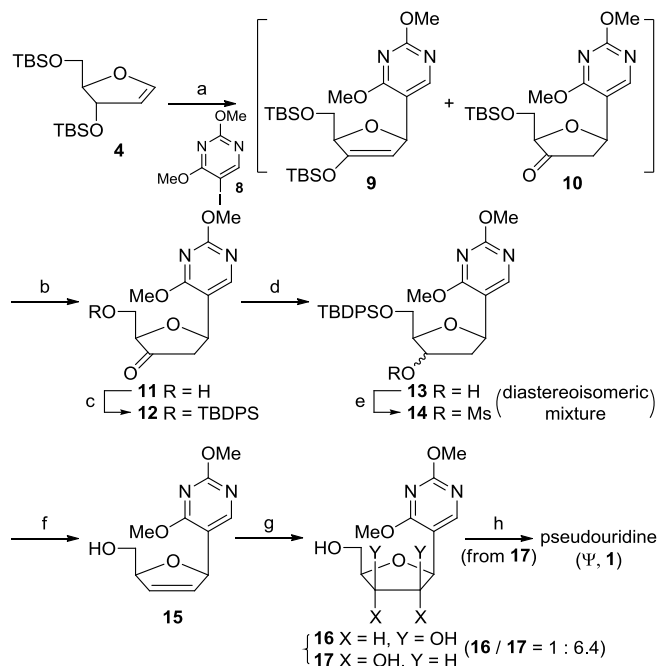
on the uracil ring. After several unsuccessful trials with *N*-protecting groups on uracil of 2'-deoxypseudouridine (**2**), we opted to protect the oxygens of uracil for subsequent transformation.



reagents and conditions:

(a) Pd₂dba₃ (0.1 equiv), XantPhos (0.2 equiv), 5-iodouracil (**3**, 2 equiv), glycal **4** (1 equiv), *n*-Bu₃N (1.5 equiv), DMF (0.05 M), 70 °C, 15 h, 83%; (b) AcOH (2 equiv), TBAF (1M in THF, containing ca. 5% water, 2 equiv), THF (0.2 M), 0 °C~rt, 8 h, 85%; (c) NaB(OAc)₃H (1.5 equiv), AcOH/CH₃CN (12 mL, v/v = 1 : 1, 0.066 M), 0 °C~rt, 3 h, 92%

Scheme 3. Synthesis of 2'-deoxypseudouridine (dΨ, **2**)



reagents and conditions:

(a) Pd(OAc)₂ (0.1 equiv), PPh₃ (0.2 equiv), 2,4-dimethoxy-5-iodopyrimidine (**8**, 2 equiv), glycal **4** (1 equiv), Et₃N (1.5 equiv), DMF (0.1 M), 70 °C, 15 h; (b) TBAF (1M in THF, containing ca. 5% water, 2.0 + 1.0 equiv), AcOH (2.0 equiv), THF (0.1 M), 0 °C~rt, overnight, 88% from **4**; (c) TBDPSCl (1.2 equiv), imidazole (2.2 equiv), DCM (0.1 M), rt, overnight, 97%; (d) NaBH₄ (1.5 equiv), MeOH (0.1 M), rt, 1 h, 96% (diastereoisomeric mixture); (e) MsCl (1.3 equiv), Et₃N (1.5 equiv), DCM (0.1 M), rt, 1 h (quantitative); (f) DBU (18 equiv), CH₃CN (0.1 M), 75 °C, 48 h, 49% from **13**; (g) OsO₄ (0.05 equiv), NMO (3.0 equiv), acetone (0.1 M), -20 °C~rt, overnight, 76% (**16** : **17** = 1 : 6.4); (h) NaI (4 equiv), AcOH (0.04 M), reflux, 45 min, 90%

Scheme 4. Synthesis of pseudouridine (Ψ, **1**)

Thus, 2,4-dimethoxy-5-iodopyrimidine (**8**),²⁹ an oxygen-protected equivalent of uracil, was subjected to the Heck glycosylation with ribofuranoid glycal **4**. In a continuous screening of palladium catalyst and phosphine ligand combinations, the silyl enol ether adduct **9** from Heck glycosylation was obtained as a single diastereomer in good yields (76%) when the reaction was carried out in DMF at 70 °C with Pd₂(dba)₃ as the catalyst and PPh₃ as the ligand (entries 3 and 5 in **Table S2** in Supporting Information). In contrast, when the reaction was carried out with Pd(OAc)₂ and PPh₃, only a minimum yield of the silyl enol ether **9** was obtained accompanied with the formation of the corresponding desilylated adduct **10** in 83% yield (**Table S2**). Our investigation showed that the use of Pd(OAc)₂ as the catalyst resulted in partial desilylation of the immediate Heck adduct **9** to form a product mixtures of **9** and **10**.^{21,30} Regardless, the overall yield for the C-C bond formation is excellent and the condition is more suitable for multi-gram synthesis. Although **9** and **10** can be readily separated by column chromatography for characterization purpose, in our later work, the mixture of **9** and **10** was treated with TBAF and AcOH for global desilylation to give 5-(2'-deoxy-3'-oxoribofuranosyl)-2,4-dimethoxypyrimidine (**11**) in an excellent yield.

As a result, instead of elaborating the silyl enol ether **9**, we decided to derive the 3'-keto adduct **11** into the targeted molecule. After protecting the 5'-hydroxy group of **11** with TBDPS, the 3'-keto group was reduced with NaBH₄ to form an epimeric mixture of **13**. Subsequently, mesylation of the mixture **13** followed by elimination and concomitant removal of TBDPS group with DBU in CH₃CN furnished the 2',3'-dideoxydihydro-C-nucleoside **15**. *cis*-Dihydroxylation of **15** with OsO₄ gave a mixture of lyxofuranosyl and ribofuranosyl adducts (**16** and **17**, respectively) in the ratio of 1 to 6.4.³¹ Fortunately, the desired product **17** is predominant and can be readily separated.³² Finally, *O*-demethylation of **17** was accomplished by treatment with NaI in acetic acid at reflux temperature to afford pseudouridine (**1**).¹¹ The spectroscopic data of our synthesized pseudouridine (**1**) is in accordance with data previously reported in the literature (**Tables S3~S6** in Supporting Information).^{11,15,33-35}

Conclusion

In summary, we herein reported a facile and practical synthesis for pseudouridine (**1**), which provides a complementary access to this important biological molecule. Further application of this approach to manipulate the Heck-type glycosylation adducts into ribonucleosides would be amenable to the synthesis of versatile C-nucleosides.

Experimental section

General chemical procedures

The chemical shift values are reported in δ values (parts per million, ppm) relative to the standard chemical shift for the hydrogen residue peak and carbon-13 peak in the deuterated

solvent, CDCl₃, or DMSO-*d*₆.³⁶ The coupling constant (*J*) values are expressed in hertz (Hz). The numbers of protons directly attached to the individual carbons were determined by ¹³C NMR DEPT experiments. Thin-layer chromatography (TLC) was performed on silica gel plates. Compounds on TLC were visualized by illumination under UV light (254 nm), and dipped into 10% *conc.* H₂SO₄ in EtOH, *p*-anisaldehyde stain (with sulfuric acid in EtOH), or 10% phosphomolybdic acid in EtOH followed by charring on a hot plate. Solvent systems are expressed with respect to the volumetric ratio of the less polar component to the more polar component (v/v). Silica gel (230-400 mesh) was used for flash column chromatography and this technique has been described by W. C. Still *et al.*³⁷ Evaporations were carried out under reduced pressure (water aspirator or vacuum pump) with the bath temperature below 50 °C unless specified otherwise. Materials obtained from commercial suppliers were used without further purification.

3',5'-Di-*O*-(*tert*-butyldimethylsilyl)-2',3'-didehydro-2'-deoxypseudouridine (5)

To a solution of XantPhos (0.1389 g, 0.24 mmol, 0.2 equiv) in DMF (6.0 mL) was added tris(dibenzylideneacetone)dipalladium (Pd₂dba₃, 0.1099 g, 0.12 mmol, 0.1 equiv) at room temperature. The Pd₂dba₃ solution was stirred under argon atmosphere at room temperature for 30 min. In a separate flask, to the solution of 5-iodouracil (**3**, 0.5712 g, 2.40 mmol, 2.0 equiv) and furanoid glycal **4**²³ (0.4164 g, 1.20 mmol) in DMF (18.0 mL) was added tri-*n*-butylamine (0.43 mL, 1.8.0 mmol, 1.5 equiv) under argon atmosphere at room temperature. The 5-iodouracil solution was then added to the Pd₂dba₃ solution and the resulting solution was stirred under argon atmosphere at 70 °C for 15 h. After cooling to room temperature, the solvent was evaporated under reduced pressure. The residue was dissolved in CH₂Cl₂ and the solution was washed with H₂O and saturated aqueous NaCl solution. The organic layer was dried over anhydrous MgSO₄ and then concentrated under reduced pressure. The residue was purified by flash column chromatography (Hex / EtOAc = 5 : 5, *R_f* = 0.35) to give **5** (colorless oil, 0.4539 g, 1.00 mmol, 83%). ¹H NMR (400 MHz, CDCl₃) δ 9.29 (br s, 2H), 7.68 (s, 1H), 5.68 (t, *J* = 1.8 Hz, 1H), 4.90 (s, 1H), 4.51 (s, 1H), 3.85 and 3.73 (ABX system, *J* = 11.3, 2.8, 1.8 Hz, 2H), 0.92 (s, 9H), 0.87 (s, 9H), 0.21 (s, 3H), 0.18 (s, 3H), 0.05 (s, 3H), 0.03 (s, 3H); ¹³C NMR (400 MHz, CDCl₃) δ 163.2, 152.1, 150.1, 138.3 (CH), 116.6, 100.5 (CH), 83.8 (CH), 77.2 (CH), 63.2 (CH₂), 25.9 (CH₃), 25.5 (CH₃), 18.5, 18.0, -4.8 (CH₃), -5.2 (CH₃), -5.3 (CH₃), -5.4 (CH₃); MS (ESI-) *m/z* (%) 453 ([*M* - H], 100); HRMS (ESI+, TOF) calcd for C₂₁H₃₉N₂O₅Si₂ [*M* + H]: 455.2398. Found 455.2400.

2',3'-Dideoxy-3'-oxopseudouridine¹⁹ (**7**)

To a solution of **5** (0.6223 g, 1.37 mmol) in THF (7.0 mL) at 0 °C were added AcOH (0.16 mL, 2.74 mmol, 2.0 equiv) and tetra-*n*-butylammonium fluoride (TBAF, 1 M solution in THF, containing *ca.* 5% H₂O, 2.74 mL, 2.74 mmol, 2.0 equiv). The reaction mixture was stirred for 8 h while the reaction temperature was allowed to rise to room temperature. The solvent was evaporated under reduced pressure and the

residue was purified by flash column chromatography (CH₂Cl₂ / MeOH = 9 : 1, *R_f* = 0.25) to give **7** (white solid, 0.2634 g, 1.16 mmol, 85%).¹⁹ m.p. 206-208 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.15 (br s, 2H), 7.61 (s, 1H), 5.00 (dd, *J* = 10.0, 6.4 Hz, 1H), 4.91 (br s, 1H), 3.93 (t, *J* = 3.0 Hz, 1H), 3.65-3.58 (m, 2H), 2.68 and 2.38 (ABX system, *J* = 18.0, 10.0, 6.4 Hz, 2H); ¹³C NMR (400 MHz, DMSO-*d*₆) δ 214.2, 163.5, 151.1, 139.3 (CH), 111.8, 82.1 (CH), 70.6 (CH), 60.6 (CH₂), 42.7 (CH₂); MS (ESI-) *m/z* (%) 225 ([*M* - H], 100); HRMS (ESI-, TOF) calcd for C₉H₉N₂O₅ [*M* - H]: 225.0511. Found 225.0510.

2'-Deoxypseudouridine^{19,24,25,38} (**dψ**, **2**)

To a solution of **7** (0.1797 g, 0.79 mmol) in AcOH (6.0 mL) and acetonitrile (6.0 mL) was added sodium triacetoxymethylborohydride (NaB(OAc)₃H, 0.2522 g, 1.20 mmol, 1.5 equiv) at 0 °C. The reaction mixture was stirred for 3 h while the reaction temperature was allowed to rise to room temperature. The solvent was evaporated under reduced pressure. The residue was dissolved in EtOAc and the organic layer was washed with H₂O and saturated aqueous NaCl solution. The organic layer was dried over anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography (CH₂Cl₂ / MeOH = 8 : 2, *R_f* = 0.18) to give **2** (white solid, 0.1659 g, 0.73 mmol, 92%). m.p. 220-222 °C;^{24,38} ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.95 (br s, 2H), 7.37 (s, 1H), 4.99 (d, *J* = 3.6 Hz, 1H, OH), 4.80-4.75 (m, 2H, including 1 x OH), 4.13-4.09 (m, 1H), 3.67 (td, *J* = 4.6, 2.4 Hz, 1H), 3.41-3.38 (m, 2H), 2.00 (ddd, *J* = 12.6, 5.8, 1.8 Hz, 1H), 1.73 (ddd, *J* = 12.5, 9.7, 5.7 Hz, 1H); ¹³C NMR (400 MHz, DMSO-*d*₆) δ 163.5, 151.2, 137.9 (CH), 113.2, 87.1 (CH), 73.2 (CH), 72.1 (CH), 62.2 (CH₂), 40.9 (CH₂); MS (ESI-) *m/z* (%) 227 ([*M* - H], 100); HRMS (ESI-, TOF) calcd for C₉H₁₁N₂O₅ [*M* - H]: 227.0668. Found 227.0669.

2,4-Dimethoxy-5-(3,5-di-*O*-*tert*-butyldimethylsilyl-2-deoxy-2,3-didehydro-β-D-ribofuranosyl)pyrimidine (**9**)

To a solution of PPh₃ (0.0276 g, 0.11 mmol, 0.2 equiv) in DMF (1.0 mL) was added Pd₂dba₃ (0.0483 g, 0.053 mmol, 0.1 equiv) at room temperature. The Pd₂dba₃ solution was stirred under argon atmosphere at room temperature for 30 min. In a separate flask, to the solution of 2,4-dimethoxy-5-iodopyrimidine²⁹ (**8**, 0.2655 g, 1.00 mmol, 2.0 equiv) and furanoid glycal **4**²³ (0.1725 g, 0.50 mmol) in DMF (4.0 mL) was added triethylamine (0.11 mL, 0.79 mmol, 1.5 equiv) under argon atmosphere at room temperature. The 2,4-dimethoxy-5-iodopyrimidine solution was then added to the Pd₂dba₃ solution and the resulting solution was stirred under argon atmosphere at 70 °C for 15 h. After cooling to room temperature, the solvent was evaporated under reduced pressure. The residue was dissolved in EtOAc and the solution was washed with H₂O and saturated aqueous NaCl solution. The organic layer was dried over anhydrous MgSO₄ and then concentrated under reduced pressure. The residue was purified by flash column chromatography (Hex / EtOAc = 20 : 1) to give **9** (oil, 0.1866 g, 0.39 mmol, 77%, *R_f* = 0.60 (Hex / EtOAc = 2 : 1)). ¹H NMR (400 MHz, CDCl₃) δ 8.38 (s, 1H), 5.87 (dd, *J* = 3.4, 1.4 Hz, 1H), 4.77 (dd, *J* = 1.8 Hz, 1H), 4.54 (ddd, *J* = 5.6, 3.8, 2.0 Hz, 1H), 3.98 (s, 3H), 3.96 (s, 3H), 3.82 and 3.70 (ABX

system, $J = 11.2, 4.0, 2.4$ Hz, 2H), 0.93 (s, 9H), 0.83 (s, 9H), 0.22 (s, 3H), 0.20 (s, 3H), -0.0095 (s, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 168.2, 164.5, 157.3 (CH), 151.0, 116.3, 99.3 (CH), 83.9 (CH), 76.8 (CH), 63.8 (CH_2), 54.4 (CH_3), 53.6 (CH_3), 25.8 (CH_3), 25.3 (CH_3), 18.3, 17.8, -5.1 (CH_3), -5.5 (CH_3), -5.6 (CH_3); MS (ESI+) m/z (%) 483 ([M + H], 100); HRMS (ESI+, TOF) calcd for $\text{C}_{23}\text{H}_{43}\text{N}_2\text{O}_5\text{Si}_2$ [M + H]: 483.2711. Found: 483.2707.

2,4-Dimethoxy-5-(5-*O*-*tert*-butyldimethylsilyl-2,3-dideoxy-3-oxo- β -D-ribofuranosyl)pyrimidine (10)

To a solution of **9** (0.9800 g, 2.03 mmol) in THF (28 mL) was added AcOH (0.13 mL, 2.16 mmol, 1.0 equiv) and TBAF (1 M solution in THF, containing *ca.* 5% H_2O , 4.2 mL, 4.20 mmol, 2.0 equiv) at 0 °C. The solution was stirred at 0 °C for 20 min till no starting material was detected by TLC. The solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography (Hex / EtOAc = 6 : 1) to give **10** (oil, 0.6451 g, 1.75 mmol, 86%, $R_f = 0.35$ (Hex / EtOAc = 2 : 1)). ^1H NMR (400 MHz, CDCl_3) δ 8.48 (s, 1H), 5.23 (dd, $J = 11.0, 5.8$ Hz, 1H), 3.99-3.96 (m, 1H), 3.98 (s, 3H), 3.97 (s, 3H), 3.94-3.93 (m, 1H), 3.90 (dd, $J = 11.4, 3.4$ Hz, 1H), 2.86 and 2.29 (ABX system, $J = 17.7, 10.8, 6.0$ Hz, 2H), 0.82 (s, 9H), 0.040 (s, 3H), 0.021 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 213.4, 168.0, 164.9, 155.7 (CH), 114.5, 82.4 (CH), 70.8 (CH), 62.5 (CH_2), 54.8 (CH_3), 53.9 (CH_3), 44.3 (CH_2), 25.7 (CH_3), 18.2, -5.5 (CH_3), -5.7 (CH_3); MS (ESI+) m/z (%) 369 ([M + H], 100); HRMS (ESI+, TOF) calcd for $\text{C}_{17}\text{H}_{29}\text{N}_2\text{O}_5\text{Si}$ [M + H]: 369.1846. Found: 369.1844.

2,4-Dimethoxy-5-(2,3-dideoxy-3-oxo- β -D-ribofuranosyl)pyrimidine (11)

To a solution of PPh_3 (0.5816 g, 2.22 mmol, 0.2 equiv) in DMF (22 mL) was added $\text{Pd}(\text{OAc})_2$ (0.2502 g, 1.11 mmol, 0.1 equiv) at room temperature. The $\text{Pd}(\text{OAc})_2$ solution was stirred under argon atmosphere at room temperature for 30 min. In a separate flask, to the solution of 2,4-dimethoxy-5-iodopyrimidine²⁹ (**8**, 5.8817 g, 22.11 mmol, 2.0 equiv) and furanoid glycal **4**²³ (3.8105 g, 11.06 mmol) in DMF (88 mL) was added triethylamine (2.4 mL, 17.21 mmol, 1.5 equiv) under argon atmosphere at room temperature. The 2,4-dimethoxy-5-iodopyrimidine solution was then added to $\text{Pd}(\text{OAc})_2$ solution and the resulting mixture was stirred under argon atmosphere at 70 °C for 15 h. After cooling to room temperature, the solvent was evaporated under reduced pressure and the resulting residue (containing the product mixture of **9** and **10**) was used for the subsequent reaction without further purification.

To the solution of previous residue in THF (110 mL) were added AcOH (1.4 mL, 23.31 mmol, 2.0 equiv) and TBAF (1 M solution in THF, containing *ca.* 5% H_2O , 22 mL, 22.0 mmol, 2.0 equiv) at 0 °C. The solution was stirred overnight while the reaction temperature was allowed to rise to room temperature. Additional TBAF (1 M in THF, 11 mL, 11.00 mmol, 1.0 equiv) was added at 0 °C and the solution was stirred for additional 2 h while the reaction temperature was allowed to rise to room temperature. The solvent was evaporated under reduced pressure. The residue was dissolved in EtOAc and the solution was washed with H_2O and saturated aqueous NaCl

solution. The organic layer was dried over anhydrous MgSO_4 and then concentrated under reduced pressure. The residue was purified by flash column chromatography (Hex / EtOAc = 1 : 2, $R_f = 0.20$) to give **11** (oil, 2.4834 g, 9.77 mmol, 88%). ^1H NMR (400 MHz, CDCl_3) δ 8.34 (s, 1H), 5.20 (dd, $J = 10.6, 6.2$ Hz, 1H), 3.97 (s, 3H), 3.95 (s, 3H), 3.99-3.87 (m, 3H), 3.12 (bs, 1 H, OH), 2.81 and 2.54 (ABX system, $J = 18.1, 10.8, 6.1$ Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 213.5, 168.3, 165.1, 156.4 (CH), 113.4, 82.0 (CH), 71.9 (CH), 61.3 (CH_2), 54.9 (CH_3), 54.1 (CH_3), 43.0 (CH_2); MS (ESI+) m/z (%) 255 ([M + H], 100); HRMS (ESI+, TOF) calcd for $\text{C}_{11}\text{H}_{15}\text{N}_2\text{O}_5$ [M + H]: 255.0981. Found: 255.0981.

2,4-Dimethoxy-5-(5-*O*-*tert*-butyldiphenylsilyl-2,3-dideoxy-3-oxo- β -D-ribofuranosyl)pyrimidine (12)

To a mixture of **11** (2.5213 g, 9.92 mmol) and imidazole (1.4924 g, 21.92 mmol, 2.2 equiv) in CH_2Cl_2 (99 mL) at 0 °C was added dropwise *tert*-butylchlorodiphenylsilane (TBDPSCI, 3.4 mL, 12.37 mmol, 1.2 equiv). The reaction mixture was stirred overnight while the reaction temperature was allowed to rise to room temperature. The solvent was evaporated under reduced pressure. The residue was partitioned between CH_2Cl_2 and H_2O . The organic layer was washed with aqueous 1 N HCl solution, saturated aqueous Na_2CO_3 solution, saturated aqueous NaCl solution and dried over anhydrous MgSO_4 . The solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography (Hex / EtOAc = 4 : 1) to give **12** (oil, 4.7250 g, 9.59 mmol, 97%, $R_f = 0.45$ (Hex / EtOAc = 2 : 1)). ^1H NMR (400 MHz, CDCl_3) δ 8.54 (s, 1H), 7.71 (d, $J = 6.0$ Hz, 2H), 7.66 (d, $J = 6.4$ Hz, 2H), 7.44-7.36 (m, 6H), 5.27 (dd, $J = 11.0, 5.8$ Hz, 1H), 4.04-3.99 (m, 3H), 4.02 (s, 3H), 3.98 (s, 3H), 2.95 and 2.44 (ABX system, $J = 17.4, 11.0, 5.8$ Hz, 2H), 0.99 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 213.2, 168.0, 165.0, 155.6 (CH), 135.5 (CH), 135.4 (CH), 132.8, 132.6, 129.7 (CH), 127.7 (CH), 127.6 (CH), 114.2, 82.4 (CH), 71.0 (CH), 62.9 (CH_2), 54.8 (CH_3), 53.9 (CH_3), 44.3 (CH_2), 26.6 (CH_3), 19.1; MS (ESI+) m/z (%) 493 ([M + H], 100); HRMS (ESI+, TOF) calcd for $\text{C}_{27}\text{H}_{33}\text{N}_2\text{O}_5\text{Si}$ [M + H]: 493.2159. Found: 493.2159.

Mixture of 2,4-dimethoxy-5-(2-deoxy- β -D-ribofuranosyl)pyrimidine and 3'-epimer (13)

To a solution of **12** (3.7421 g, 7.60 mmol) in MeOH (76 mL) at 0 °C was added NaBH_4 (0.4389 g, 11.60 mmol, 1.5 equiv). The reaction mixture was stirred for 1 h till no starting material was detected by TLC while the reaction temperature was allowed to rise to room temperature. The reaction was quenched with saturated aqueous NH_4Cl solution at 0 °C and the mixture was stirred for 10 min. The solvents were evaporated under reduced pressure and the residue was partitioned between EtOAc and saturated aqueous NH_4Cl solution. The organic layer was washed with saturated aqueous NaCl solution and dried over anhydrous MgSO_4 . The solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography (Hex / EtOAc = 2 : 1, $R_f = 0.13$ and 0.20) to give the product mixture of **13** (oil, 3.6205 g, 7.32 mmol, 96%). Diastereoisomeric mixture: ^1H NMR (400 MHz, CDCl_3) δ 8.41 (s, 1H), 8.29 (s, 0.18H), 7.73-7.65 (m, 4H+0.72H, Ph), 7.47-7.37 (s, 6H+1.08H, Ph), 5.21 (dd, $J = 10.0, 5.6$ Hz, 0.18H), 4.97 (t, $J = 7.4$

Hz, 1H), 4.60-4.55 (m, 1H), 4.51 (dt, $J = 5.6, 2.7$ Hz, 0.18H), 4.11 (dd, $J = 11.0, 6.2$ Hz, 1H), 4.05 (dd, $J = 10.8, 4.0$ Hz, 1H), 4.00 (dd, $J = 3.9, 2.8$ Hz, 0.18H), 3.98 (s, 3H, OMe), 3.97 (s, 3H, OMe), 3.97 (s, 0.54H, OMe), 3.95 (s, 0.54H, OMe), 3.94-3.90 (m, 1H), 3.85 and 3.69 (ABX system, $J = 10.7, 6.4, 4.2$ Hz, 0.36H), 3.08 (d, $J = 4.4$ Hz, 1H), 2.65 (ddd, $J = 13.8, 7.8, 6.2$ Hz, 1H), 2.30 (ddd, $J = 13.2, 5.8, 2.2$ Hz, 0.18H), 1.93 (ddd, $J = 13.4, 6.6, 3.2$ Hz, 2H+0.36H), 1.07 (s, 9H, *t*-butyl), 1.05 (s, 1.62H, *t*-butyl); ^{13}C NMR (100 MHz, CDCl_3) δ 167.9, 167.7, 164.5, 164.4, 155.7 (CH), 155.2 (CH), 135.4 (CH), 133.01, 132.96, 132.6, 132.5, 129.7 (CH), 129.6 (CH), 127.7 (CH), 127.6 (CH), 115.8, 115.3, 86.9 (CH), 81.7 (CH), 73.6 (CH), 73.3 (CH), 73.1 (CH), 72.6 (CH), 64.4 (CH₂), 62.8 (CH₂), 54.5 (CH₃), 53.7 (CH₃), 53.6 (CH₃), 41.6 (CH₂), 41.2 (CH₂), 26.6 (CH₃), 19.0, 18.9; MS (ESI+) m/z (%) 495 ([M + H], 100); HRMS (ESI+, TOF) calcd for $\text{C}_{27}\text{H}_{35}\text{N}_2\text{O}_5\text{Si}$ [M + H]: 495.2315. Found: 495.2316.

2,4-Dimethoxy-5-(2,3-didehydro-2,3-dideoxy- β -D-ribofuranosyl)pyrimidine (15)

To a solution of **13** (1.8260 g, 3.69 mmol) and Et_3N (0.78 mL, 5.59 mmol, 1.5 equiv) in CH_2Cl_2 (37 mL) at 0 °C was added methanesulfonyl chloride (MsCl , 0.38 mL, 4.91 mmol, 1.3 equiv). The reaction mixture was stirred at room temperature for 1 h till no starting material was detected by TLC. The solvent was evaporated under reduced pressure and the residue was partitioned between EtOAc and H_2O . The organic layer was washed with saturated aqueous NaCl solution and dried over anhydrous MgSO_4 . The solvent was evaporated under reduced pressure and the resulting residue ($R_f = 0.075$, Hex / $\text{EtOAc} = 4 : 1$) was used for the subsequent reaction without further purification.

To the solution of previous residue in CH_3CN (37 mL, 0.1 M) at 0 °C was added 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU, 10.5 mL, 68.97 mmol, 18.0 equiv). The reaction mixture was stirred at 75 °C for 48 h. After cooling to room temperature, the solvent was evaporated under reduced pressure and the residue was partitioned between EtOAc and H_2O . The organic layer was washed with saturated aqueous NaCl solution and dried over anhydrous MgSO_4 . The solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography (Hex / $\text{EtOAc} = 1 : 2 \sim 1 : 4$) to give **15** (oil, 0.4336 g, 1.82 mmol, 49%, $R_f = 0.35$ (Hex / $\text{EtOAc} = 1 : 4$)). ^1H NMR (400 MHz, CDCl_3) δ 8.20 (s, 1H), 5.97 (dt, $J = 6.3, 1.9$ Hz, 1H), 5.91 (ddd, $J = 6.1, 2.3, 1.5$ Hz, 1H), 5.87-5.85 (m, 1H), 4.94-4.95 (m, 1H), 3.98 (s, 3H), 3.94 (s, 3H), 3.73 and 3.63 (ABX system, $J = 11.7, 4.6, 3.2$ Hz, 2H), 2.53 (br s, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 168.5, 164.9, 156.7 (CH), 129.8 (CH), 128.1 (CH), 114.8, 87.2 (CH), 80.8 (CH), 64.7 (CH₂), 54.8 (CH₃), 54.0 (CH₃); MS (ESI+) m/z (%) 239 ([M + H], 100); HRMS (ESI+, TOF) calcd for $\text{C}_{11}\text{H}_{15}\text{N}_2\text{O}_4$ [M + H]: 239.1032. Found: 239.1031.

2,4-Dimethoxy-5-(β -D-ribofuranosyl)pyrimidine³² (17)

To a mixture of **15** (0.0709 g, 0.30 mmol) and 4-methylmorpholine *N*-oxide (50 wt% in H_2O , 0.22 mL, 0.94 mmol, 3.0 equiv) in acetone (3.1 mL) at -20 °C was added dropwise OsO_4 (4.0 wt% in H_2O , 0.09 mL, 0.014 mmol, 0.050 equiv). The reaction mixture was stirred overnight while the

reaction temperature was allowed to rise gradually to room temperature. The solvent was evaporated under reduced pressure and the residue was partitioned between CHCl_3 and saturated aqueous $\text{Na}_2\text{S}_2\text{O}_3$ solution. The organic layer was washed with saturated aqueous NaCl solution and dried over anhydrous MgSO_4 . The solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography (CH_2Cl_2 / $\text{MeOH} = 20 : 1$) to give two diastereoisomeric products **17** and **16** (oil, 0.0613 g, 0.23 mmol, 76%, **17** / **16** = 6.4 : 1, $R_f = 0.18$ (more polar, the desired product, **17**) and 0.28 (less polar, the byproduct, **16**) (CH_2Cl_2 / $\text{MeOH} = 15 : 1$)).

2,4-Dimethoxy-5-(β -D-ribofuranosyl)pyrimidine³² (17) (more polar product)

^1H NMR (500 MHz, D_2O) δ 8.24 (s, 1H), 4.89 (d, $J = 5.0$ Hz, 1H), 4.25 (t, $J = 5.3$ Hz, 1H), 4.14 (t, $J = 5.8$ Hz, 1H), 4.04 (dt, $J = 5.8, 3.5$ Hz, 1H), 3.99 (s, 3H), 3.95 (s, 3H), 3.91 and 3.79 (ABX system, $J = 12.4, 5.5, 3.3$ Hz, 2H); ^{13}C NMR (125 MHz, D_2O) δ 168.9, 164.7, 155.7 (CH), 112.9, 83.0 (CH), 78.8 (CH), 74.5 (CH), 70.6 (CH), 61.4 (CH₂), 55.0 (CH₃), 54.3 (CH₃); MS (ESI+) m/z (%) 273 ([M + H], 100); HRMS (ESI+, TOF) calcd for $\text{C}_{11}\text{H}_{17}\text{N}_2\text{O}_6$ [M + H]: 273.1087. Found: 273.1088.

2,4-Dimethoxy-5-(β -D-lyxofuranosyl)pyrimidine (16) (less polar byproduct)

^1H NMR (400 MHz, D_2O) δ 8.25 (s, 1H), 5.02 (d, $J = 4.0$ Hz, 1H), 4.63 (t, $J = 6.0$ Hz, 1H), 4.44 (t, $J = 4.6$ Hz, 1H), 4.20 (dt, $J = 6.2, 4.0$ Hz, 1H), 3.96 (s, 3H), 3.95 (s, 3H), 3.89 and 3.81 (ABX system, $J = 12.2, 5.8, 3.4$ Hz, 2H); ^{13}C NMR (100 MHz, D_2O) δ 168.2, 164.4, 155.5 (CH), 111.5, 79.4, 75.6 (CH), 72.0 (CH), 71.1 (CH), 60.4 (CH₂), 54.9 (CH₃), 54.3 (CH₃); MS (ESI+) m/z (%) 273 ([M + H], 100); HRMS (ESI+, TOF) calcd for $\text{C}_{11}\text{H}_{17}\text{N}_2\text{O}_6$ [M + H]: 273.1087. Found: 273.1087.

Pseudouridine (Ψ , 1)

To a mixture of **17** (0.0361 g, 0.13 mmol) in AcOH (3.3 mL) at room temperature was added NaI (0.0815 g, 0.54 mmol, 4.0 equiv). The reaction mixture was stirred at reflux temperature under argon atmosphere for 45 min till no starting material was detected by TLC. After cooling to room temperature, the solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography (CHCl_3 / $\text{MeOH} = 4 : 1$) to give pseudouridine (Ψ , **1**, white solid, 0.0268 g, 0.12 mmol, 90%, ($R_f = 0.08$, CH_2Cl_2 / $\text{MeOH} = 4 : 1$)). m.p. 219-222 °C; ^1H NMR (500 MHz, $\text{DMSO}-d_6$)^{15,33} δ 7.52 (s, 1H), 4.46 (d, $J = 4.5$ Hz, 1H), 3.92 (t, $J = 4.8$ Hz, 1H), 3.86 (t, $J = 5.3$ Hz, 1H), 3.69 (dt, $J = 5.5, 3.2$ Hz, 1H), 3.59 and 3.44 (ABX system, $J = 11.9, 3.7, 3.1$ Hz, 2H); ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$)^{15,34} δ 163.7, 151.3, 140.1 (CH), 111.0, 83.2 (CH), 78.9 (CH), 73.9 (CH), 70.4 (CH), 61.2 (CH₂); ^1H NMR (500 MHz, D_2O)^{11,33,35} δ 7.73 (d, $J = 0.6$ Hz, 1H), 4.74 (d, $J = 5.6$ Hz, 1H), 4.35 (t, $J = 5.5$ Hz, 1H), 4.20 (t, $J = 5.4$ Hz, 1H), 4.08 (dt, $J = 5.3, 3.3$ Hz, 1H), 3.90 (dd, $J = 12.5, 3.2$ Hz, 1H), 3.78 (dd, $J = 12.5, 4.9$ Hz, 1H); ^{13}C NMR (125 MHz, D_2O)¹¹ δ 165.4, 152.9, 141.6 (CH), 110.5, 83.4 (CH), 79.1 (CH), 73.4 (CH), 70.9 (CH), 61.5 (CH₂); MS (ESI-) m/z (%) 243 ([M - H], 100); HRMS (ESI-, TOF) calcd for $\text{C}_9\text{H}_{11}\text{N}_2\text{O}_6$ [M - H]: 243.0617. Found: 243.0618.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by National Taiwan Normal University and Research Grants MOST 107-2113-M-003-002- and MOST 106-2113-M-003-006- from Ministry of Science and Technology, Taiwan.

References

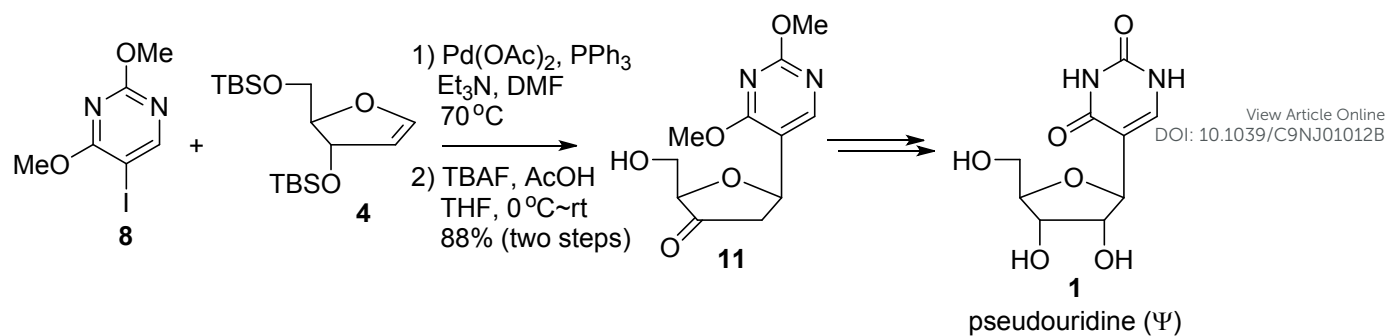
- (a) Cohn, W. E. *Biochim. Biophys. Acta*, 1959, **32**, 569; (b) Davis, F. F.; Allen, F. W. *J. Biol. Chem.*, 1957, **227**, 907; (c) Cohn, W. E. *Fed. Proc.*, 1957, **16**, 166.
- (a) Chambers, R. W.; Kurkov, V.; Shapiro, R. *Biochemistry*, 1963, **2**, 1192; (b) Cohn, W. E. *J. Biol. Chem.*, 1960, **235**, 1488.
- Shapiro, R.; Chambers, R. W. *J. Am. Chem. Soc.*, 1961, **83**, 3920.
- Hamma, T.; Ferré-D'Amaré, A. R. *Chem. Biol.*, 2006, **13**, 1125.
- (a) Rintala-Dempsey, A. C.; Kothe, U. *RNA Biol.*, 2017, **14**, 1185; (b) Li, X.; Ma, S.; Yi, C. *Curr. Opin. Chem. Biol.*, 2016, **33**, 108; (c) Spenkuch, F.; Motorin, Y.; Helm, M. *RNA Biol.*, 2014, **11**, 1540; (d) Durairaj, A.; Limbach, P. A. *Anal. Chim. Acta*, 2008, **623**, 117; (e) Charette, M.; Gray, M. W. *IUBMB Life*, 2000, **49**, 341.
- Kitamura, K.; Ando, Y.; Matsumoto, T.; Suzuki, K. *Chem. Rev.*, 2018, **118**, 1495.
- (a) Temburnikar, K.; Seley-Radtke, K. L. *Beilstein J. Org. Chem.*, 2018, **14**, 772; (b) De Clercq, E. J. *Med. Chem.*, 2016, **59**, 2301; (c) Štambaský, J.; Hocek, M.; Kočovský, P. *Chem. Rev.*, 2009, **109**, 6729; (d) Wu, Q.; Simons, C. *Synthesis*, 2004, 1533; (e) Shaban, M. A. E.; Nasr, A. Z. *Adv. Heterocycl. Chem.*, 1997, **68**, 223; (f) Shaban, M. A. E. *Adv. Heterocycl. Chem.*, 1997, **70**, 163.
- Wellington, K. W.; Benner, S. A. *Nucleosides Nucleotides Nucleic Acids*, 2006, **25**, 1309.
- (a) Brown, D. M.; Ogden, R. C. *J. Chem. Soc., Perkin Trans. 1*, 1981, 723; (b) Habermehl, G.; Christ, B. G. *Justus Liebigs Ann. Chem.*, 1978, 427; (c) Lerch, U.; Burdon, M. G.; Moffatt, J. G. *J. Org. Chem.*, 1971, **36**, 1507; (d) Brown, D. M.; Burdon, M. G.; Slatcliff, R. P. *J. Chem. Soc. C*, 1968, 1051; (e) Brown, D. M.; Burdon, M. G.; Slatcliff, R. P. *Chem. Commun.*, 1965, 77.
- (a) van Rijssel, E. R.; van Delft, P.; van Marle, D. V.; Bijvoets, S. M.; Lodder, G.; Overkleeft, H. S.; van der Marel, G. A.; Filippov, D. V.; Codée, J. D. C. *J. Org. Chem.*, 2015, **80**, 4553; (b) Hanessian, S.; Marcotte, S.; Machaalani, R.; Huang, G. B. *Org. Lett.*, 2003, **5**, 4277; (c) Asbun, W.; Binkley, S. B. *J. Org. Chem.*, 1968, **33**, 140; (d) Grohar, P. J.; Chow, C. S. *Tetrahedron Lett.*, 1999, **40**, 2049.
- Desaulniers, J.-P.; Chang, Y.-C.; Aduri, R.; Abeyirigunawardena, S. C.; SantaLucia, J. J.; Chow, C. S. *Org. Biomol. Chem.*, 2008, **6**, 3892.
- Chang, Y.-C.; Herath, J.; Wang, T. H. H.; Chow, C. S. *Bioorg. Med. Chem.*, 2008, **16**, 2676.
- Hanessian, S.; Machaalani, R. *Tetrahedron Lett.*, 2003, **44**, 8321.
- Chu, C. K.; Wempen, I.; Watanabe, K. A.; Fox, J. J. *J. Org. Chem.*, 1976, **41**, 2793.
- Sato, T.; Hayakawa, Y.; Noyori, R. *Bull. Chem. Soc. Jpn.*, 1984, **57**, 2515.
- Noyori, R.; Sato, T.; Hayakawa, Y. *J. Am. Chem. Soc.*, 1978, **100**, 2561.
- Cheng, J. C. Y.; Hacksell, U.; Daves, G. D., Jr. *J. Org. Chem.*, 1986, **51**, 3093.
- Kim, H.-J.; Leal, N. A.; Benner, S. A. *Bioorg. Med. Chem.*, 2009, **17**, 3728. DOI: 10.1039/C9NJ01012B
- Zhang, H. C.; Daves, G. D., Jr. *J. Org. Chem.*, 1992, **57**, 4690.
- Recent examples for Heck-type glycosylation: (a) T. Kaewsomboon, S. Nishizawa, T. Kanamori, H. Yuasa, A. Ohkubo, *J. Org. Chem.*, 2018, **83**, 1320; (b) T. J. Walter, C. Richert, *Nucleic Acids Res.*, 2018, **46**, 8069; (c) D. U. Ukale, T. Lönnberg, *Angew. Chem. Int. Ed.*, 2018, **57**, 16171; (d) R. Wang, C. Jin, X. Zhu, L. Zhou, W. Xuan, Y. Liu, Q. Liu, W. Tan, J. Am. Chem. Soc., 2017, **139**, 9104; (f) M. Merkel, L. Dehm, N. P. Ernsting, H.-A. Wagenknecht, *Angew. Chem. Int. Ed.*, 2017, **56**, 384; (g) M. Xia, W. Hu, S. Sun, J.-T. Yu, J. Cheng, *Org. Biomol. Chem.*, 2017, **15**, 4064; (h) T. Goldau, K. Murayama, C. Brieke, S. Steinwand, P. Mondal, M. Biswas, I. Burghardt, J. Wachtveitl, H. Asanuma, A. Heckel, *Chem. Eur. J.*, 2015, **21**, 2845; (i) N. Gaß, H.-A. Wagenknecht, *Eur. J. Org. Chem.*, 2015, 6661; (j) D. Hou, M. M. Greenberg, *J. Org. Chem.*, 2014, **79**, 1877; (k) K. Temburnikar, K. Brace, K. L. Seley-Radtke, *J. Org. Chem.*, 2013, **78**, 7305; (l) T. Ehrenschröder, W. Schmucker, C. Wellner, T. Augenstein, P. Carl, J. Harmer, F. Breher, H.-A. Wagenknecht, *Chem. Eur. J.*, 2013, **19**, 12547; (m) M. Minuth, C. Richert, *Angew. Chem., Int. Ed.*, 2013, **52**, 10874; (n) T. Kubelka, L. Slavětinská, V. Eigner, M. Hocek, *Org. Biomol. Chem.*, 2013, **11**, 4702; (o) S. H. Lee, S. Wang, E. T. Kool, *Chem. Commun.*, 2012, **48**, 8069; (p) J.-Y. Heo, G. T. Hwang, *Bull. Korean Chem. Soc.*, 2010, **31**, 3794; (q) S. Matsuda, A. M. Leconte, F. E. Romesberg, *J. Am. Chem. Soc.*, 2007, **129**, 5551; (r) N. Joubert, R. Pohl, B. Klepetarova, M. Hocek, *J. Org. Chem.*, 2007, **72**, 6797; (s) H.-P. Hsieh, L. W. McLaughlin, *J. Org. Chem.*, 1995, **60**, 5356.
- H. Chapuis, T. Kubelka, N. Joubert, R. Pohl, M. Hocek, *Eur. J. Org. Chem.*, 2012, **2012**, 1759.
- (a) Weinberger, M.; Berndt, F.; Mahrwald, R.; Ernsting, N. P.; Wagenknecht, H.-A. *J. Org. Chem.*, 2013, **78**, 2589; (b) Kubelka, T.; Slavětinská, L.; Hocek, M. *Synthesis*, 2012, **44**, 953; (c) Kubelka, T.; Slavětinská, L.; Klepetářová, B.; Hocek, M. *Eur. J. Org. Chem.*, 2010, 2666.
- (a) Cameron, M. A.; Cush, S. B.; Hammer, R. P. *J. Org. Chem.*, 1997, **62**, 9065; (b) Walker, J. A.; Chen, J. J.; Wise, D. S.; Townsend, L. B. *J. Org. Chem.*, 1996, **61**, 2219.
- Reese, C. B.; Wu, Q. *Org. Biomol. Chem.*, 2003, **1**, 3160.
- Ramzaeva, N.; Rosemeyer, H.; Leonard, P.; Mühlegger, K.; Bergmann, F.; von der Eltz, H.; Seela, F. *Helv. Chim. Acta*, 2000, **83**, 1108.
- Alonso, D.; Caballero, E.; Medarde, M.; Tomé, F. *Tetrahedron Lett.*, 2007, **48**, 907.
- Paquette, L. A.; Zhao, M. *J. Am. Chem. Soc.*, 1998, **120**, 5203.
- Kim, H.-J.; Leal, N. A.; Hoshika, S.; Benner, S. A. *J. Org. Chem.*, 2014, **79**, 3194.
- Prystaš, M. Š., F. *Collect. Czech. Chem. Commun.*, 1964, **29**, 121.
- T. Mabit, A. Siard, F. Legros, S. Guilleme, A. Martel, J. Lebreton, F. Carreaux, G. Dujardin, S. Collet, *Chem. Eur. J.*, 2018, **24**, 14069.
- Gudmundsson, K. S.; Drach, J. C.; Townsend, L. B. *J. Org. Chem.*, 1998, **63**, 984.
- Hanessian, S.; Marcotte, S.; Machaalani, R.; Huang, G. B.; Pierron, J.; Loiseleur, O. *Tetrahedron*, 2006, **62**, 5201.
- Deslauriers, R.; Smith, I. C. P. *Can. J. Chem.*, 1973, **51**, 833.
- (a) Wenkert, E.; Hagaman, E. W.; Gutowski, G. E. *Biochem. Biophys. Res. Commun.*, 1973, **51**, 318; (b) Chenon, M.-T.; Pugmire, R. J.; Grant, D. M.; Panzica, R. P.; Townsend, L. B. *J. Heterocycl. Chem.*, 1973, **10**, 427.
- (a) Sasaki, K.; Kusakabe, Y.; Esumi, S. *J. Antibiot.*, 1972, **25**, 151; (b) Hruska, F. E.; Grey, A. A.; Smith, I. C. P. *J. Am. Chem. Soc.*, 1970, **92**, 4088; (c) Luyten, I.; Pankiewicz, K. W.; Watanabe, K. A.; Chattopadhyaya, J. *J. Org. Chem.*, 1998, **63**, 1033.

ARTICLE

Journal Name

- 36 Gottlieb, H. E.; Kotlyar, V.; Nudelman, A. *J. Org. Chem.*, 1997, **62**, 7512.
- 37 Still, W. C.; Kahn, M.; Mitra, A. *J. Org. Chem.*, 1978, **43**, 2923.
- 38 (a) Pankiewicz, K.; Matsuda, A.; Watanabe, K. A. *J. Org. Chem.*, 1982, **47**, 485; (b) Matsuda, A.; Chu, C. K.; Reichman, U.; Pankiewicz, K.; Watanabe, K. A.; Fox, J. J. *J. Org. Chem.*, 1981, **46**, 3603; (c) Chu, C. K.; Reichman, U.; Watanabe, K. A.; Fox, J. J. *J. Heterocycl. Chem.*, 1977, **14**, 1119; (d) Bridges, S. D.; Brown, D. M.; Ogden, R. C. *J. Chem. Soc., Chem. Commun.*, 1977, 460.

View Article Online
DOI: 10.1039/C9NJ01012B



Pseudouridine (**1**) was synthesized by functional group interconversions of the Heck adduct **11** from 2,4-dimethoxy-5-iodopyrimidine (**8**) and ribofuranoid glycal **4**.