

Design, syntheses and antitumor activities evaluation of 1,5-diaryl substituted pyrazole secnidazole ester derivatives

Qing-Hu Teng, Gui-Xia Sun, Shu-Ying Luo, Kai Wang*, Fu-Pei Liang*

Guangxi Key Laboratory of Electrochemical and Magnetochemical Functional Materials, College of Chemistry and Bioengineering, Guilin University of Technology, Guilin, 541004, China.

*Corresponding author: e-mail: kaiwang2011@yahoo.com, fliangoffice@yahoo.com.

((Additional Supporting Information may be found in the online version of this article.))**(Please delete if not appropriate)**

Abstract. According to the drug hybridization principle, a series of novel 1,5-diaryl substituted pyrazole secnidazole ester derivatives (**6aa-6gc**) have been synthesized by the combinations of various 1,5-diarylpyrazole-3-carboxylic acids with secnidazole. The in vitro antitumor/ cytotoxicities activities against tumor and normal cell lines, including NCI-H460 (lung tumor cell), MCG-803 (gastric tumor cell), Skov-3 (ovarian tumor cell), BEL-7404 (liver tumor cell) and HL-7702 (normal liver cell), have been evaluated using MTT assay. All compounds showed promising inhibitory activities against four tumor cell lines. The IC₅₀ of **6bc** against the BEL-7404 cell was 2.03 μM, and those of **6fc** against the NCI-H460, MCG-803 and Skov-3 were 1.34, 0.14 and 0.87 μM, respectively. All these values were much lower than those of the cisplatin. Furthermore, **6fc** and **6bc** were also verified to be considerable safe for normal human liver cell, since the lower IC₅₀ values than cisplatin. Based on these results, the cell cycle analysis, apoptosis ratio detection, and mitochondrial membrane potential assay of **6fc** and **6bc** were further performed aiming to investigate their inhibition mechanism of BEL-7404 cells. It is revealed that they have effectively inhibited the cell growth by arresting the BEL-7404 cells at S phase and induced apoptosis through the mitochondria-mediated pathway.

Keywords: Drug hybridization principle; 1,5-Diaryl substituted pyrazole; Secnidazoles; Antitumor activities

which could behavior as the bifunctional conjugates with potent inhibitory activity towards cyclooxygenase (COX) and histone deacetylase (HDAC), could arrest the cell cycle progression of androgen dependent prostate tumor cell line (LNCaP) in the S- and G0/G1 phases.¹¹ Several hybridization of pyrazoles and substituted coumarins/morpholines groups have also found to be potent dual COX-2/5-LOX inhibitors and antitumor agents. Their lowest IC₅₀ have reached to 0.16 μM.¹² These results suggested that more extensive and prominent anti-tumor activities could be expected when such pyrazole parents were hybridized with other novel active groups.

As a next-generation of 5-nitroimidazole, secnidazole has been approved for a single-dose (2 g) treatment of bacterial vaginosis (BV), as well as commercially available for more than three decades in 31 countries.¹³ Moreover, secnidazole can also undergo a bioreduction of the nitro group to generate toxic reactive oxygen species (ROS), which leading to DNA helix damage, disruption of bacterial protein synthesis and replication, and ultimately, cell death.¹⁴ On this basis, special attention have been paid to its structural modifications. Several secnidazole derivatives, including esters and metal-complexes, have been reported as potential drug in tumor growth inhibition.¹⁵ It should be noted that, its derivatives that stem from the combination of pyrazoles with secnidazoles was rarely investigated yet. Up to now, only series of compounds synthesized form the coupling reaction of 4-(1-acetyl-3-phenyl-4,5-dihydro-1H-pyrazol-5-yl)benzoic acid with secnidazole has been developed. And it is gratifying that these compounds have been determined to bear excellent antitumor activities.¹⁶

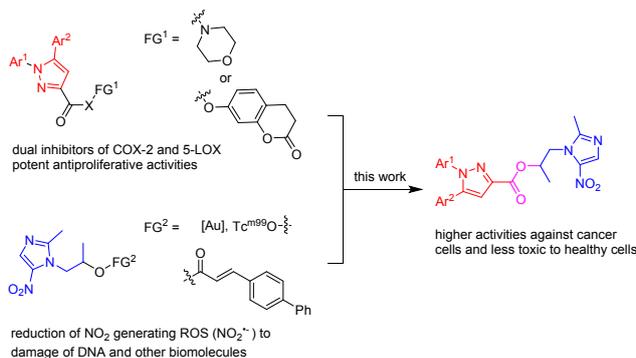
In recent years, our group have been interested in the development of new methodologies for the syntheses of new nitrogen heterocycles molecules, as well as the searching of novel antitumor targeted drug candidates.¹⁷ Based on these works and combined with excellent antitumor potentials of the pyrazole and secnidazole groups (Scheme 1), we have designed and

Introduction

As one type of the core frameworks in various natural products and medicines, nitrogen-containing heterocycles have played important roles in drug design and development.¹ The pyrazole is just one of such species. They have shown extensive bioactivities,² such as anti-tumor,³ antimicrobial,⁴ antifungal,⁵ hypoglycemic,⁶ anti-inflammatory,⁷ analgesic,⁸ against oxidative stress and diabetes,⁹ and various enzyme inhibiting activities,¹⁰ etc. In particular, promising potentials against the malignant tumor have been demonstrated in some of modified pyrazole derivatives when they were grafted with other active groups according to the drug hybridization principle. For instances, the methyl sulfone pyrazole derivatives,

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1002/jhet.4302

synthesized various 1,5-diaryl substituted pyrazole secnidazole ester derivatives. The antitumor activities of all obtained compounds have been evaluated using MTT assay. Several compounds have been found to bear much higher inhibiting activities against most of selected tumor cells than the cisplatin. Furthermore, they were also verified to be much safer for normal human liver cell than the cisplatin. In order to reveal their inhibition mechanism, the cell cycle analysis, apoptosis ratio detection, and mitochondrial membrane potential assay of selected compounds were also investigated.

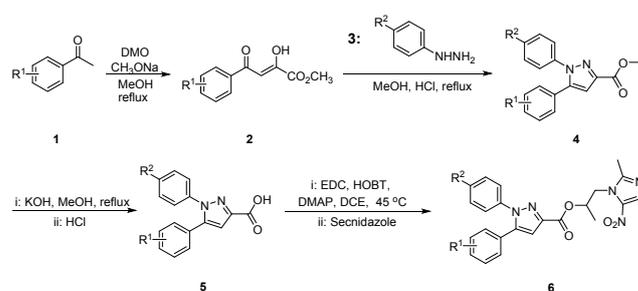


Scheme 1. Design philosophy of pyrazole secnidazole derivatives as antitumor agents.

Results and discussion

Chemistry

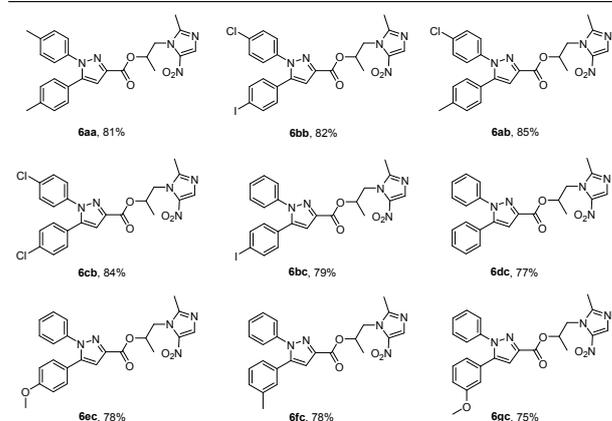
The syntheses of targeted compounds **6** are illustrated in Scheme 2. Their structural diversity stem from precursor compounds of 1,5-diaryl substituted 1*H*-pyrazole-3-carboxylic acid (**5**), in which various substituted benzene moieties have been introduced into the 1- and 5- positions of pyrazole moieties according to previously reported mature method.^{3,12b} Originally, methyl (*Z*)-2-hydroxy-4-oxo-4-phenylbut-2-enoate (**2**) were synthesized from substituted acetophenones (**1**) and dimethyl oxalate (DMO) through aldol type condensation reaction. The reaction of **2** with *para*-substituted phenylhydrazines (**3**) afforded methyl 1,5-diphenyl-1*H*-pyrazole-3-carboxylates (**4**) via an intermolecular cyclization mechanism. Various 1,5-diphenyl-1*H*-pyrazole-3-carboxylic acid (**5**) were subsequently synthesized through successively implemented hydrolysis and acidification processes of **4**. Finally, secnidazole active groups were introduced by simple esterification reactions, which yielded target compounds 1-(2-methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-1,5-diphenyl-1*H*-pyrazole-3-carboxylate (**6**) in moderate to good yields.



Scheme 2. Synthesis route of compounds **6**.

The molecular structures and yields (%) of these targeted compounds were summarized in Table 1. All targeted compounds were first reported and structurally characterized by ¹H NMR spectroscopy, ¹³C NMR spectroscopy and ESI-MS. Taking **6aa** as the example, its ¹H NMR spectrum showed a doublet of Me proton of propane at N1-secnidazole at δ 1.44 (d, J = 6.1 Hz, 3H), three singlets of Me protons at *para*-5-phenyl, *para*-1-phenyl, and C2 of secnidazole appeared at δ 2.29 (s, 3H), 2.35 (s, 3H), 2.55 (s, 3H), respectively. The methylene (CH₂) protons of propane at N1-imidazole showed a multiplet at δ 4.54 (m, 1H), and a doublet at δ 4.69 (d, J = 14.1 Hz, 1H), respectively. There was a broad singlet of methylene (CH) proton of propane at N1-imidazole at δ 5.45 (br, 1H). The protons of pyrazole and imidazole ring displayed at δ 6.96 (s, 1H) and 8.02 (s, 1H) as singlets, respectively. The protons of 1, 5-diphenyl showed a multiplet at δ 7.11-7.19 (m, 6H), and a doublet at δ 7.26 (d, J = 7.8, 2H). Nearly similar patterns were noticed in ¹H and ¹³C NMR spectra of all the compounds (**6aa-6gc**). The HRMS (ESI) of compounds (**6aa-6gc**) showed characteristic [M+H]⁺ corresponding peaks equivalent to their molecular formulae. Relevant spectral data is furnished in supporting information.

Table 1. Molecular structures and the yields of compounds **6aa-6gc**.



Antiproliferation assays and MTT assays

The antiproliferative effects of all targeted compounds were evaluated against four human tumor cell lines and a normal cell line using MTT assay. These cell

lines include human lung tumor cell NCI-H460, human gastric tumor cell MCG-803, human ovarian tumor cell Skov-3, human liver tumor cell BEL-7404, and normal liver cell HL-7702. Cisplatin, a well-known marketed antitumor drug, was employed as the positive control in the MTT assay. As shown in Table 2, all compounds exhibited potent inhibitory activity against four tumor cells of NCI-H460, MCG-803, Skov-3, and BEL-7404. The IC_{50} values of all compounds against four tumor cells range from 0.14 to 14.08 μ M. Obviously, these values were all much lower than the corresponding IC_{50} values of the cisplatin. Among these, **6fc** showed the highest inhibiting effects on tumor cell lines of NCI-H460, MCG-803, and Skov-3, with the IC_{50} values of 1.34, 0.14, and 0.87 μ M, respectively. While for BEL-7404 tumor cell lines, the **6bc** possessed the most significant inhibiting activity, whose IC_{50} value was determined to be 2.03 μ M. It should be noted that the IC_{50} values of **6fc** and **6bc** against the human normal liver cell HL-7702 were 8.97 and 5.77 μ M, respectively, all of which were closed to that of the cisplatin (7.93 μ M). This meant that both of them displayed comparable cytotoxic activities as the cisplatin.

Table 2. IC_{50} (μ M) values of compounds **6aa-6gc** against four tumor cell lines and a normal cell line.^a

Compound	NCI-H460	MCG-803	Skov-3	BEL-7404	HL-7702
6aa	14.08	0.32	7.34	4.56	8.46
6bb	2.43	1.27	2.67	2.78	6.25
6ab	4.68	0.68	1.47	6.45	5.34
6cb	6.42	1.05	3.38	7.66	7.81
6bc	7.88	2.47	6.59	2.03	5.77
6dc	6.67	3.52	3.17	8.59	11.76
6ec	3.85	0.79	9.69	4.65	16.52
6fc	1.34	0.14	0.87	5.28	8.97
6gc	3.54	1.86	10.37	11.24	57.78
Cisplatin	48.52	4.19	84.21	24.87	7.93

^a IC_{50} : Concentration inhibits 50% of cell growth. Data are shown as the mean of three independent experiments run in triplicate.

The above results confirmed the validity of our design, the hybridization of pyrazole and secnidazole could result in compounds with excellent antitumor activities. In order to further probe antitumor mechanism of targeted compounds, compounds **6fc** and **6bc** were selected to determine the cell cycle analysis, apoptosis ratio detection, and mitochondrial membrane potential assay using BEL-7404 cell line.

Apoptosis study by flow cytometry and confocal microscopy

In order to verify whether the compounds induced cell death was caused by apoptosis or necrosis, the interactions of BEL-7404 cells with **6fc** or **6bc** was

investigated by an annexin V-FITC/propidium iodide assay using flow cytometry. Considering the fact that in apoptosis process the phosphatidylserine (PS) exposure usually occurs before the loss of plasma membrane integrity, the presence of annexin V+/PI-cells was regarded as an indicator of apoptosis. As shown in Fig. 1, the presence of both **6fc** and **6bc** have led to the obvious increasing of the population of annexin V+/PI-cells (26.1% for **6fc** and 23.96% for **6bc**) in comparison with that of control (9.56%). These results suggested that apoptotic death of BEL-7404 cells was induced by two tested compounds.

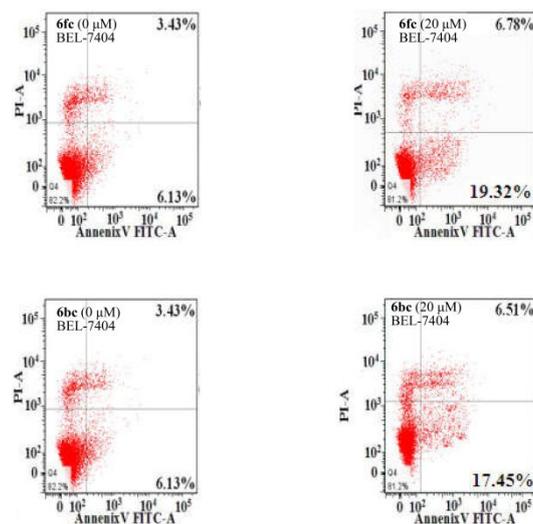


Fig. 1 Annexin-V/propidium iodide assay of the BEL-7404 cells treated with **6fc** and **6bc** (20.0 μ M).

Cell cycle arrest

Meanwhile, the flow cytometry analysis was also employed to determine the apoptosis ration of BEL-7404 cell line induced by **6fc** and **6bc**, thereby the relationships between inhibiting activities of above compounds and the cell cycle arrest of corresponding BEL-7404 cells could be illustrated. In this assay, the BEL-7404 cells were treated with **6fc** and **6bc** at their IC_{50} concentrations, following which the cell cycle phase was assayed by evaluating the DNA content of cells stained with propidium iodide. As shown in Fig. 2, with the extending of treating time, compound **6fc** has caused increased accumulation of BEL-7404 cells in the S phases but a decrease in the accumulation in the G1 phase. The percentage of S phase for control and compound **6fc** at 24 and 48 h were 31.39, 37.40 and 55.26%, respectively. As for the cells treated with **6bc**, the percentage of G1 phase among them also decreased along with the treating time extending. And the percentage of S phase also underwent a substantially rising. It has increased from 26.21% for control to the 33.57% at 24 h, and finally reached to the 45.85% at 48 h. Therefore, the **6fc** and **6bc** could arrest cell cycle of BEL-7404 cells at S phase. And based on these results it could be concluded that DNA

may be one of the possible intracellular targets for **6fc** or **6bc**, because most DNA replication occurs within this S stage.

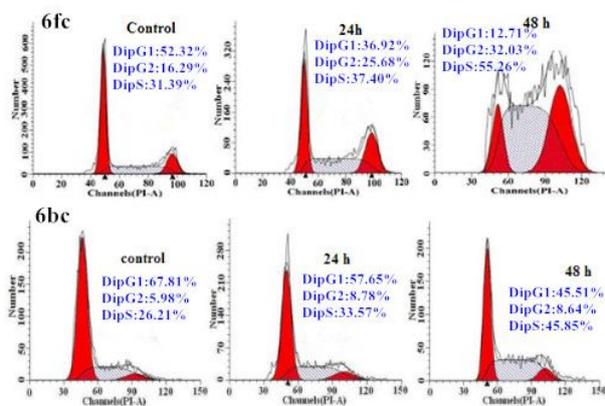
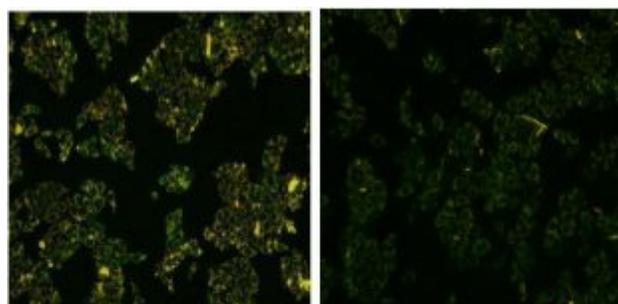


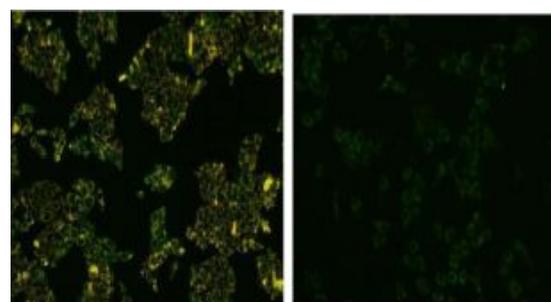
Fig. 2. Cell cycle analysis by flow cytometry for BEL-7404 cells treated with **6fc** and **6bc**.

Detection of mitochondrial membrane potential

It is known that mitochondria play significant roles in the process of cell apoptosis. They could act as a point of integration for apoptotic signals, which stem from both the extrinsic and intrinsic apoptotic pathways. Particularly, the loss of mitochondrial membrane potential (MMP, $\Delta\Psi_m$) is involved in apoptotic cell death due to the cytotoxicity of the antitumor drugs. Because of this, it is regarded as an important indicator of mitochondrial dysfunction. To investigate these roles of mitochondria in BEL-7404 apoptosis induced by **6fc** or **6bc**, the change of the mitochondrial membrane potential was evaluated. From Fig. 3 it could be observed that the BEL-7404 cells were obviously changed into green by treating with compounds **6fc** and **6bc** in the concentration of 20 μM , which indicated that the MMP was declined comparing with the control (Fig. 3.). That meant the mitochondrial-mediated pathways were involved in the apoptosis of BEL-7404 cells induced by **6fc** and **6bc**.



(a) control (b) **6fc** (20 μM)



(c) control (d) **6bc** (20 μM)

Fig. 3 Loss of $\Delta\Psi_m$ induced by **6fc** and **6bc**.

Conclusion

In summary, nine novel hybrid compounds containing both diaryl substituted pyrazole and secnidazole groups have been designed and synthesized. They have been determined to showed potent in vitro inhibiting activities against selected four human tumor cell lines, which were all superior to those of the cisplatin. Among them, compound **6fc** displayed the most significant inhibiting activities against human tumor cell lines of NCI-H460, MCG-803 and Skov-3. While for BEL-7404 cell, compound **6bc** did the best. Further studies revealed the possible antitumor mechanism of **6fc** and **6bc**. Both of them could arrest the BEL-7404 cell cycle at S phase, and induced apoptosis of BEL-7404 cells through mitochondrial-mediated pathway.

Experimental Section

Synthesis and characterization of 1-(2-methyl-5-nitro-1H-imidazol-1-yl)propan-2-yl 1,5-diphenyl-1H-pyrazole-3-carboxylate (**6**)

Conventional procedure:

Synthesis of compounds **2**

The reaction mixture of **1** (8 mmol), dimethyl oxalate (16 mmol), and dry CH_3OH (20 mL) was slowly added into the mixture of $\text{CH}_3\text{ONa}/\text{CH}_3\text{OH}$ (16 mmol/5 mL). The reaction mixture was stirred at reflux for 5 h, and monitored periodically by TLC. Upon completion, the reaction mixture was cooled to room temperature and poured into water (50 mL). Then, the mixture was stirred and added HCl (1 M) until $\text{pH} = 3-4$. After neutralization, the mixture was extracted with ethyl acetate (3×100 mL). The combined organic layers were washed with water and brine, dried over NaSO_4 and filtered. The solvent was removed under vacuum. The residue was purified by flash column chromatography to afford **2**.

Synthesis of compounds **4**

The diluted HCl (1 mL/1 M) was added into the reaction mixture of **2** (1 mmol), phenylhydrazines **3** (2 mmol), and CH_3OH (12 mL). The reaction mixture was reflux for 6 h, and monitored periodically by TLC. Upon completion, the reaction mixture was cooled to room temperature and extracted with ethyl acetate (3×20 mL). The combined

organic layers were washed with water and brine, dried over NaSO₄ and filtered. The solvent was removed under vacuum. The residue was purified by flash column chromatography to afford **4**.

Synthesis of compounds 5

H₂O (1 mL) was slowly added into the reaction mixture of **4** (0.5 mmol), KOH (1.75 mmol) and CH₃OH (10 mL). The mixture was reflux for 2 h, and monitored periodically by TLC. Upon completion, the reaction mixture was cooled to room temperature and poured into H₂O (15 mL). Then, HCl (1 M) was added into the reaction mixture until pH = 3-4, followed by extracted with ethyl acetate (3 × 20 mL). The combined organic layers were washed with water and brine, dried over NaSO₄ and filtered. The solvent was removed under vacuum. The residue was purified by flash column chromatography to afford **5**.

Synthesis of compounds 6

The reaction mixture of **5** (0.4 mmol), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDC, 0.48 mmol), 1-Hydroxybenzotriazole hydrate (HOBT, 0.48 mmol), 4-dimethylaminopyridine (DMAP, 0.2 mmol), and DCE (10 mL) was stirred at 45 °C for 0.5 h. Then, sycnidazole (0.52 mmol) was added into the reaction mixture and heated to reflux overnight. Upon completion, the reaction mixture was cooled to room temperature and extracted with DCM (3 × 20 mL). The combined organic layers were washed with water and brine, dried over NaSO₄ and filtered. The solvent was removed under vacuum. The residue was purified by flash column chromatography or recrystallized in EtOH to afford **6**.

Characterization of the compounds

1-(2-Methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-1,5-dip-tolyl-1*H*-pyrazole-3-carboxylate (**6aa**) : yellow solid, mp: 171-172 °C; ¹H NMR (500 MHz, DMSO-*d*⁶): δ = 1.44 (d, *J* = 6.1 Hz, 3H), 2.29 (s, 3H), 2.35 (s, 3H), 2.55 (s, 3H), 4.54 (m, 1H), 4.69 (d, *J* = 14.1 Hz, 1H), 5.45 (br, 1H), 6.96 (s, 1H), 7.11-7.19 (m, 6H), 7.26 (d, *J* = 7.8, 2H), 8.02 (s, 1H) ppm; ¹³C NMR (125 MHz, DMSO-*d*⁶): δ = 14.58, 17.75, 21.12, 21.23, 50.01, 70.11, 109.66, 125.78, 126.45, 128.93, 129.70, 130.14, 133.67, 137.26, 138.76, 138.94, 142.91, 144.90, 152.37, 161.13 ppm. *m/z* (MS): calcd for C₂₅H₂₆N₅O₄ [M+H]⁺ 460.20, found 460.25.

1-(2-Methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-1-(4-chlorophenyl)-5-(4-iodophenyl)-1*H*-pyrazole-1-(2-Methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-1-(4-chlorophenyl)-5-(*p*-tolyl)-1*H*-pyrazole-3-carboxylate (**6ab**) : yellow solid, mp: 169-171 °C; ¹H NMR (500 MHz, DMSO-*d*⁶): δ = 1.44 (d, *J* = 6.4 Hz, 3H), 2.30 (s, 3H), 2.55 (s, 3H), 4.55 (dd, *J* = 15.0, 9.7 Hz, 1H), 4.69 (dd, *J* = 15.0, 2.7 Hz, 1H), 5.45 (ddd, *J* = 9.4, 6.4, 2.8 Hz, 1H), 6.99 (s, 1H), 7.14 (d, *J* = 8.2, 2H), 7.20 (d, *J* = 8.1, 2H), 7.34 (m, 2H), 7.54 (m, 2H), 8.02 (s, 1H) ppm; ¹³C NMR (125 MHz, DMSO-*d*⁶): δ = 14.58, 17.74, 19.91, 21.26, 48.48, 70.24, 80.47, 109.99, 126.15, 127.70, 129.05, 129.78, 129.82, 133.67, 139.20, 141.38, 143.39, 145.16, 148.84, 152.36, 156.64, 161.01 ppm. *m/z* (MS): calcd for C₂₄H₂₃ClN₅O₄ [M+H]⁺ 480.14 and 482.14, found 480.20 and 482.20.

1-(2-Methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-1-(4-chlorophenyl)-5-(4-iodophenyl)-1*H*-3-carboxylate (**6bb**) : yellow solid, mp: 156-158 °C; ¹H NMR (500 MHz, DMSO-*d*⁶): δ = 1.44 (d, *J* = 6.4 Hz, 3H), 2.54 (s, 3H), 4.54 (dd, *J* = 15.0, 9.7 Hz, 1H), 4.69 (dd, *J* = 15.0, 2.7 Hz, 1H), 5.45 (ddd, *J* = 9.4, 6.4, 2.8 Hz, 1H), 7.05 (s, 1H), 7.07 (d, *J* = 1.2, 2H), 7.36 (d, *J* = 8.7, 2H), 7.56 (d, *J* = 8.7, 2H), 7.77 (d, *J* = 8.4, 2H), 8.02 (s, 1H) ppm; ¹³C NMR (125 MHz, DMSO-*d*⁶): δ = 14.60, 17.73, 67.70, 70.31, 96.60, 110.45, 117.46, 127.72, 129.88, 131.11, 131.15, 133.78, 138.03, 138.13, 143.48, 143.64, 144.15, 155.17, 160.89 ppm. *m/z* (MS): calcd for

C₂₃H₂₀ClIN₅O₄ [M+H]⁺ 592.02 and 594.02, found 592.10 and 594.10.

1-(2-Methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-1,5-bis(4-chlorophenyl)-1*H*-pyrazole-3-carboxylate (**6cb**) : yellow solid, mp: 224-226 °C; ¹H NMR (500 MHz, DMSO-*d*⁶): δ = 1.44 (d, *J* = 6.4 Hz, 3H), 2.55 (s, 3H), 4.54 (dd, *J* = 15.0, 9.7 Hz, 1H), 4.69 (dd, *J* = 15.0, 2.6 Hz, 1H), 5.46 (ddd, *J* = 9.4, 6.4, 2.8 Hz, 1H), 7.08 (s, 1H), 7.26-7.31 (m, 2H), 7.33-7.38 (m, 2H), 7.44-7.50 (m, 2H), 7.53-7.60 (m, 2H), 8.02 (s, 1H) ppm; ¹³C NMR (125 MHz, DMSO-*d*⁶): δ = 14.58, 17.74, 49.99, 60.23, 70.33, 100.00, 110.57, 127.75, 127.90, 129.33, 129.89, 131.03, 133.66, 133.80, 134.42, 138.14, 138.98, 143.48, 143.92, 152.35, 160.93 ppm. *m/z* (MS): calcd for C₂₃H₂₀Cl₂N₅O₄ [M+H]⁺ 500.09 and 502.09, found 500.10 and 502.10.

1-(2-Methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-5-(4-iodophenyl)-1-phenyl-1*H*-pyrazole-3-carboxylate (**6bc**) : yellow solid, mp: 182-183 °C; ¹H NMR (500 MHz, DMSO-*d*⁶): δ = 1.44 (d, *J* = 6.4 Hz, 3H), 2.55 (s, 3H), 4.54 (dd, *J* = 15.0, 9.7 Hz, 1H), 4.69 (dd, *J* = 15.0, 2.7 Hz, 1H), 5.46 (ddd, *J* = 9.4, 6.4, 2.8 Hz, 1H), 7.02 (s, 1H), 7.04 (s, 1H), 7.07 (s, 1H), 7.32-7.34 (m, 2H), 7.48-7.49 (m, 3H), 7.73 (d, *J* = 8.4 Hz, 2H), 8.02 (s, 1H) ppm; ¹³C NMR (125 MHz, DMSO-*d*⁶): δ = 14.58, 17.75, 50.01, 70.22, 96.31, 100.00, 110.23, 126.03, 128.75, 129.39, 129.85, 131.04, 133.66, 137.95, 138.98, 139.38, 143.25, 144.02, 152.35, 161.00. *m/z* (MS): calcd for C₂₃H₂₁IN₅O₄ [M+H]⁺ 558.06, found 558.10.

1-(2-Methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-1,5-diphenyl-1*H*-pyrazole-3-carboxylate (**6dc**) : yellow solid, mp: 183-185 °C; ¹H NMR (500 MHz, DMSO-*d*⁶): δ = 1.45 (d, *J* = 6.4 Hz, 3H), 2.56 (s, 3H), 4.54 (dd, *J* = 15.0, 9.7 Hz, 1H), 4.70 (dd, *J* = 15.0, 2.7 Hz, 1H), 5.46 (ddd, *J* = 9.3, 6.3, 2.8 Hz, 1H), 7.03 (s, 1H), 7.22-7.27 (m, 2H), 7.33-7.29 (m, 2H), 7.34-7.39 (m, 3H), 7.44-7.51 (s, 3H), 8.02 (s, 1H) ppm; ¹³C NMR (125 MHz, DMSO-*d*⁶): δ = 14.59, 17.75, 50.01, 70.19, 110.05, 126.01, 129.08, 129.13, 129.23, 129.39, 129.74, 133.67, 138.96, 139.54, 143.13, 144.93, 152.35, 161.07 ppm. *m/z* (MS): calcd for C₂₃H₂₂N₅O₄ [M+H]⁺ 432.17, found 432.20.

1-(2-Methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-5-(4-methoxyphenyl)-1-phenyl-1*H*-pyrazole-3-carboxylate (**6ec**) : yellow solid, mp: 218-220 °C; ¹H NMR (500 MHz, DMSO-*d*⁶): δ = 1.45 (d, *J* = 6.1 Hz, 3H), 2.55 (s, 3H), 3.75 (s, 3H), 4.54 (dd, *J* = 14.5, 10.0 Hz, 1H), 4.69 (d, *J* = 14.5 Hz, 1H), 5.45 (br, 1H), 6.97-6.85 (m, 3H), 7.17 (d, *J* = 8.4, 2H), 7.32 (d, *J* = 5.6, 2H), 7.46 (s, 3H), 8.02 (s, 1H) ppm; ¹³C NMR (125 MHz, DMSO-*d*⁶): δ = 14.58, 17.76, 50.03, 55.69, 70.11, 109.47, 114.60, 121.49, 126.00, 129.15, 129.72, 130.49, 133.64, 138.99, 139.72, 143.06, 144.86, 152.36, 160.08, 161.15. *m/z* (MS): calcd for C₂₄H₂₄N₅O₅ [M+H]⁺ 462.20, found 462.30.

1-(2-Methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-1-phenyl-5-(*m*-tolyl)-1*H*-pyrazole-3-carboxylate (**6fc**) : yellow solid, mp: 158-160 °C; ¹H NMR (500 MHz, DMSO-*d*⁶): δ = 1.45 (d, *J* = 6.4 Hz, 3H), 2.25 (s, 3H), 2.55 (s, 3H), 4.54 (dd, *J* = 15.0, 2.7 Hz, 1H), 4.70 (dd, *J* = 15.0, 2.7 Hz, 1H), 5.46 (ddd, *J* = 9.3, 6.4, 2.8 Hz, 1H), 6.96 (m, *J* = 7.2, 1H), 7.01 (s, H), 7.14 (s, H), 7.18-7.24 (m, 2H), 7.30-7.33 (m, 2H), 7.50-7.48 (m, 3H), 8.02 (s, 1H) ppm; ¹³C NMR (125 MHz, DMSO-*d*⁶): δ = 14.60, 17.74, 50.02, 70.22, 110.35, 126.03, 128.10, 129.21, 129.39, 129.84, 130.91, 133.69, 134.24, 139.32, 143.18, 143.74, 161.00 ppm. *m/z* (MS): calcd for C₂₄H₂₄N₅O₄ [M+H]⁺ 446.18, found 446.19.

1-(2-Methyl-5-nitro-1*H*-imidazol-1-yl)propan-2-yl-5-(3-methoxyphenyl)-1-phenyl-1*H*-pyrazole-3-carboxylate (**6gc**) : yellow solid, mp: 175-176 °C; ¹H NMR (500 MHz, DMSO-*d*⁶): δ = 1.44 (d, *J* = 6.4 Hz, 3H), 2.55 (s, 3H), 3.75 (s, 3H), 4.54 (dd, *J* = 15.0, 2.7 Hz, 1H), 4.69 (dd, *J* = 15.0, 2.6 Hz, 1H), 5.45 (br, 1H), 6.91-6.95 (m, 3H), 7.17 (d, *J* = 8.8, 2H), 7.31-7.33 (m, 2H), 7.45-7.48 (m, 3H), 8.03 (s, 1H) ppm; ¹³C NMR (125 MHz, DMSO-*d*⁶): δ = 14.59, 17.75, 21.34, 50.02, 70.15, 109.96, 125.97, 126.14, 128.93, 129.15,

129.22, 129.66, 129.70, 130.02, 133.67, 138.46, 139.59, 143.09, 145.02, 152.35, 161.07 ppm. m/z (MS): calcd for C₂₄H₂₄N₅O₅ [M+H]⁺ 462.18, found 462.20.

Acknowledgements

This work was financially supported by the National Natural Science Foundation of China (Nos. 21961008 and 21771043) and the Guangxi Natural Science Foundation (No. 2018GXNSFAA138123).

Conflict of interest

The authors declare no conflict of interest.

Data availability statement

Data available in article supplementary material.

The authors declare no conflict of interest.

References

- [1] a) D. C. Blakemore, L. Castro, I. Churcher, D. C. Rees, A. W. Thomas, D. M. Wilson, A. Wood, *Nat. Chem.* **2018**, *10*, 383–394; b) M. D. Eastgate, M. A. Schmidt, K. R. Fandrick, *Nat. Rev. Chem.* **2017**, *1*, 0016; c) D. J. Foley, A. Nelson, S. P. Marsden, *Angew. Chem. Int. Ed.* **2016**, *55*, 13650–13657; d) A. Hamid, A. Elomri, A. Daïch, *Tetrahedron Lett.* **2006**, *47*, 1777–1781; e) L.-L. Wang, T. Jiang, P.-H. Li, R.-J. Sun, and Z. Zuo, *Adv. Synth. Catal.* **2018**, *360*, 4832–4836; f) X.-J. Peng, H.-P. He, Q. Liu, K. She, B.-Q. Zhang, H.-S. Wang, H.-T. Tang, Y.-M. Pan, *Sci. China Chem.* 2021, DOI: 10.1007/s11426-020-9958-6.
- [2] a) V Deiana, G.-C. María, M. R. Pazos, Javier F.-R., B. Asproni, E. Cichero, P. Fossa, M. Eduardo, F. Deligia, G. Murineddu, G.-A. Moisés, G. A. Pinna, *Eur. J. Med. Chem.* **2016**, *112*, 66–80; b) E. Kotsikorou, F. Navas, M. J. Roche, A. F. Gilliam, B. F. Thomas, H. H. Seltzman, P. Kumar, Z.-H. Song, D. P. Hurst, D. L. Lynch, and P. H. Reggio, *J. Med. Chem.* **2013**, *56*, 6593–6612; c) S. Narayanan, R. Maitra, J. R. Deschamps, K. Bortoff, J. B. Thomas, Y. Y. Zhang, K. Warner, V. Vasukuttan, A. Decker, S. P. Runyon, *Bioorg. Med. Chem.* **2016**, *24*, 3758–377.
- [3] Z. Q. Wang, T. Song, Y. G. Feng, Z. W. Guo, Y. D. Fan, W. J. Xu, L. Liu, A. H. Wang, and Z. C. Zhang, *J. Med. Chem.* **2016**, *59*, 3152–3162.
- [4] a) R. V. Ragavan, V. Vijayakumar, N. S. Kumari, *Eur. J. Med. Chem.* **2010**, *45*, 1173–1180; b) I. Bildirici, A. Şener, İ. Tozlu, *Med. Chem. Res.* **2007**, *16*, 418–426; c) I. Bildirici, A. Şener, E. Atalan, A. Battal, H. Genc, *Med. Chem. Res.* **2009**, *18*, 327–340; d) L.-W. Chen, P.-F. Wang, D.-J. Tang, X.-X. Tao, R.-J. Man, H.-Y. Qiu, Z.-C. Wang, C. Xu, H.-L. Zhu, *Chem. Biol. Drug Des.* **2016**, *88*, 592–598.
- [5] a) M. Samet, K. Rahmi, I. Tuba, Ç. Ferdag, A. Ahmet, O. Salim, *Eur. J. Med. Chem.* **2014**, *78*, 86–96;
- [6] H.-V. Eduardo, A.-O. Rodrigo, J. R.-E. Juan, E.-S. Samuel, H.-L. Francisco, *Eur. J. Med. Chem.* **2013**, *69*, 10–21; c) H.-V. Eduardo, S.-B. Sandybel, J. R.-E. Juan, E.-S. Samuel, H.-L. Francisco, *Bioorg. Med. Chem.* **2016**, *24*, 2298–2306.
- [7] a) K. R. A. Abdellatif, M. A. Chowdhury, Y. Dong, E. E. Knaus, *Bioorg. Med. Chem.* **2008**, *16*, 6528–6534.
- [8] N. M. A.-E. Gawad, G. S. Hassan, H. H. Georgey, *Med. Chem. Res.* **2012**, *21*, 983–994.
- [9] H.-V. Eduardo, C.-A. Romina, J. R.-E. Juan, N. M.-C. Omar, H.-L. Francisco, P. C. Jose, E.-S. Samuel, *Eur. J. Med. Chem.* **2015**, *100*, 106–118.
- [10] a) J. Finn, K. Mattia, M. Morytko, S. Ram, Y. F. Yang, X. M. Wu, E. Mak, P. Gallant and D. Keith, *Bioorg. Med. Chem. Lett.* **2003**, *13*, 2231–2234; b) M. A. Tabrizi, P. G. Baraldi, E. Ruggiero, G. Saponaro, S. Baraldi, R. Romagnoli, A. Martinelli, T. Tuccinardi, *Eur. J. Med. Chem.* **2015**, *97*, 289–305; c) M. A. Tabrizi, P. G. Baraldi, S. Baraldi, E. Ruggiero, L. D. Stefano, F. Rizzolio, L. D. C. Mannelli, C. Ghelardini, A. Chicca, M. Lapillo, J. Gertsch, C. Manera, M. Macchia, A. Martinelli, C. Granchi, F. Minutolo, and T. Tuccinardi, *J. Med. Chem.* **2018**, *61*, 1340–1354; d) A. O. Frank, M. D. Feldkamp, J. P. Kennedy, A. G. Waterson, N. F. Pelz, J. D. Patrone, B. Vangamudi, D. V. Camper, O. W. Rossanese, W. J. Chazin, and S. W. Fesik, *J. Med. Chem.* **2013**, *56*, 9242–9250; e) M. M. Ahlström, . Ridderström, I. Zamora, and K. Luthman, *J. Med. Chem.* **2007**, *50*, 4444–4452; f) A. G. Waterson, J. P. Kennedy, J. D. Patrone, N. F. Pelz, M. D. Feldkamp, A. O. Frank, B. Vangamudi, M. S.-F. Elaine, O. W. Rossanese, W. J. Chazin, and S. W. Fesik, *ACS Med. Chem. Lett.* **2015**, *6*, 140–145.
- [11] I. Raji, F. Yadudu, E. Janeira, S. Fathi, L. Szymczak, J. R. Kornacki, K. Komatsu, J.-D. Li, M. Mrksich, A. K. Oyelere, *Bioorg. Med. Chem.* **2017**, *25*, 1202–1218.
- [12] a) F.-Q. Shen, Z.-C. Wan, S.-Y. Wu, S.-Z. Ren, R.-J. Man, B.-Z. Wang, H.-L. Zhu, *Bioorg. Med. Chem. Lett.* **2017**, *27*, 3653–3660; b) Z. Li, Z.-C. Wang, X. Li, M. Abbas, S.-Y. Wu, S.-Z. Ren, Q.-X. Liu, Y. Liu, P.-W. Chen, Y.-T. Duan, P.-C. Lv, H.-L. Zhu, *Eur. J. Med. Chem.* **2019**, *169*, 168–184.
- [13] a) P. Nyirjesy, J. R. Schwebke, *Future Microbiol.* **2018**, *13*, 507–524; b) A. E. Aziz MA, F. Sharifipour, P. Abedi, S. Jahanfar, H. M. Judge, *BMC Womens Health.* **2019**, *1*, 121–133.
- [14] a) E. M. Lord, L. Harwell, C. J. Koch, *Tumor Res.* **1993**, *53*, 5721–5726; b) P. Nyirjesy, J. R. Schwebke, *Future Microbiol.* **2018**, *13*, 507–524; c) C. W. Ang, A. M. Jarrad, M. A. Cooper, M. A. T. Blaskovich, *J. Med. Chem.* **2017**, *60*, 7636–7657; d) M. Kaiser, M. A. Bray, M. Cal, B. B. Trunz, E. Torreele, R. Brun, *Antimicrob. Agents Chemother.* **2011**, *55*, 5602–5608; e) S. Kumar, S. T. Saha, L. Gu, G. Palma, S. Perumal, A. Singh-Pillay, P. Singh, A. Anand, M. Kaur, V. Kumar, *ACS*

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Omega **2018**, *3*, 12106–12113; D. I. Edwards, *J. Antimicrob. Chemother.* **1979**, *5*, 499–502.

- [15] a) Y. Qian, H.-J. Zhang, H. Zhang, C. Xu, J. Zhao, H.-L. Zhu, *Bioorg. Med. Chem.* **2010**, *18*, 4991–4996; b) X. Lina, Q. Ruan, X. R. Zhang, X. J. Duan, Y. G. Teng, J. B. Zhang, *Appl. Radiat. Isotopes* **2018**, *140*, 289–293; c) A. P. A. Oliveira, J. T. J. Freitas, R. Diniz, C. Pessoa, S. S. Maranhão, J. M. Ribeiro, E. M. Souza-Fagundes, and H. Beraldo, *ACS Omega* **2020**, *5*, 2939–2946.
- [16] H. L. Zhu, T. L. Yan, X. X. Tao, P. F. Wang, Z. C. Wang, Z. Li, CN 104945386, **2015**.
- [17] a) Q.-H. Teng, Y. Yao, W.-X. Wei, H.-T. Tang, J.-R. Li and Y.-M. Pan, *Green Chem.* **2019**, *21*, 6241–6245;

b) Q.-H. Teng, X.-J. Peng, Z.-Y. Mo, Y.-L. Xu, H.-T. Tang, H.-S. Wang, H.-B. Sun and Y.-M. Pan, *Green Chem.* **2018**, *20*, 2007–2012; c) Q.-H. Teng, Y. Sun, Y. Yao, H.-T. Tang, J.-R. Li, and Y.-M. Pan, *ChemElectroChem* **2019**, *6*, 3120–3124; d) Q.-H. Teng, Y.-L. Xu, Y. Liang, H.-S. Wang, Y.-C. Wang, and Y.-M. Pan, *Adv. Synth. Catal.* **2016**, *358*, 1897 – 1902.