

2,6-Di(Arylamino)-3-Fluoropyridine Derivatives as HIV Non-Nucleoside Reverse Transcriptase Inhibitors

Sergey Sergeev, Ashok K. Yadav, Philippe Franck, Johan Michiels, Paul Lewi, Jan Heeres, Guido Vanham, Kevin K. Ariën, Christophe M.L. Vande Velde, Hans De Winter, and Bert U. W. Maes

J. Med. Chem., **Just Accepted Manuscript** • DOI: 10.1021/acs.jmedchem.5b01336 • Publication Date (Web): 19 Jan 2016

Downloaded from <http://pubs.acs.org> on January 22, 2016

Just Accepted

“Just Accepted” manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides “Just Accepted” as a free service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. “Just Accepted” manuscripts appear in full in PDF format accompanied by an HTML abstract. “Just Accepted” manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are accessible to all readers and citable by the Digital Object Identifier (DOI®). “Just Accepted” is an optional service offered to authors. Therefore, the “Just Accepted” Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the “Just Accepted” Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these “Just Accepted” manuscripts.

2,6-Di(Arylamino)-3-Fluoropyridine Derivatives as HIV Non-Nucleoside Reverse Transcriptase Inhibitors

Sergey Sergeev¹, Ashok Kumar Yadav¹, Philippe Franck¹, Johan Michiels², Paul Lewi^{3,†}, Jan Heeres⁴, Guido Vanham^{2,5}, Kevin K. Ariën², Christophe M. L. Vande Velde⁶, Hans De Winter⁷, Bert U. W. Maes^{*,1}

¹ Organic Synthesis Division, Department of Chemistry, University of Antwerp, Groenenborgerlaan 171, B-2020 Antwerp, Belgium

² Virology Unit, Department of Biomedical Sciences, Institute of Tropical Medicine, Nationalestraat 155, B-2000 Antwerp, Belgium

³ Shakturana CV, Pater van Mierlostraat 18, 2300 Turnhout, Belgium

⁴ Heeres Consulting CV, Leemskuilen 18, 2350 Vosselaar, Belgium

⁵ Department of Biomedical Sciences, University of Antwerp, Universiteitsplein 1, 2610 Wilrijk, Belgium

⁶ Faculty of Applied Sciences, University of Antwerp, Salesianenlaan 90, 2660 Hoboken, Belgium

⁷ Medicinal Chemistry, Department of Pharmaceutical Sciences, University of Antwerp, Universiteitsplein 1, 2610 Wilrijk, Belgium

† deceased

ABSTRACT: New non-nucleoside reverse transcriptase inhibitors (NNRTI), which are similar in structure to earlier described di(arylamino)pyrimidines but featuring a 2,6-di(arylamino)-3-fluoropyridine, 2,4-di(arylamino)-5-fluoropyrimidine or 1,3-di(arylamino)-4-fluorobenzene moiety instead of a 2,4-disubstituted pyrimidine moiety, are reported. The short and practical synthesis of novel NNRTI relies on two sequential Pd-catalyzed aminations as the key

1 steps. It is demonstrated through direct comparison with reference compounds, that the presence of a fluorine atom
2 increases the *in vitro* anti-HIV activity, both against the wild type virus and drug-resistant mutant strains.
3
4

5
6
7
8 **Introduction.** At the end of 2012, an estimated 35.3 million people were living with HIV infection worldwide. There
9 were 2.3 million new infections and 1.6 million people died of HIV/AIDS globally that same year. In the Western world,
10 HIV/AIDS is no longer a fatal disease: life expectancy with adequate anti-retroviral treatment and care is more than 24
11 years after HIV-infection. Much of that success is due to the introduction of HAART (highly active antiretroviral
12 treatment) by means of combinations of two or three compounds belonging to different classes of anti-HIV compounds.¹
13
14

15
16
17
18 HAART consists of the combination of several active components belonging to different classes of anti-HIV compounds
19 such as protease inhibitors (PI), integrase inhibitors (INI), nucleoside reverse transcriptase inhibitors (NRTI) and non-
20 nucleoside reverse transcriptase inhibitors (NNRTI). A major problem in the HIV treatment remains the emergence of
21 resistance of the virus against the currently available drugs, whatever class of anti-HIV compounds they belong to.
22
23

24 Application of HAART for the treatment of HIV infection only solves this problem in part, and HAART may become
25 inefficient once resistance to one or more of the drugs used in combination is developed. Finally, none of the currently
26 available anti-HIV drugs or multi-drug therapies allows for the eradication of the virus, causing the need for life-long
27 treatment which possibly results in multidrug resistance. For this reason, there is a continuous need for the development
28 of new anti-HIV combination therapies. In order to treat drug-resistant HIV infection, new components with novel
29 chemical structures for such combination therapies, possessing new modes of action are necessary.
30
31

32
33
34
35
36
37
38 HIV-1 reverse transcriptase (RT) is one of the most important viral enzymes and plays a unique role in the HIV-1 life
39 cycle. It has two known drug-target sites, the substrate catalytic site and an allosteric site that is distinct from, but located
40 closely to, the substrate site.^{2,3} Non-nucleoside reverse transcriptase inhibitors (NNRTIs) interact with the allosteric site
41 in a non-competitive manner to distort the enzyme's active conformation and thus disrupt the function of the enzyme.^{2,4}
42 It is demonstrated by XRD analysis, that many NNRTIs form a hydrogen bond with a K101 amino acid residue in the
43 reverse transcriptase binding site.^{5,6} For example, in Etravirine (**1**, TMC-125, Figure 1a), which was recently approved as a
44 next-generation NNRTI for AIDS therapy, the NH group of the arylamine and the N atom in the pyrimidine ring of the
45 drug are involved in the hydrogen bonding with the enzyme. It was found that **1** is highly potent against wild-type and a
46 number of mutant HIV strains with nanomolar EC₅₀ values and has a high genetic barrier to delay the emergence of drug-
47 resistance.^{7,8} A number of other NNRTIs with an azaheteroaromatic ring C featuring two arylamino substituents were
48
49
50
51
52
53
54
55
56
57

developed, such as di(arylamino)triazine (DATA)⁹⁻¹¹ or di(arylamino)pyrimidine (DAPY)^{9, 10, 12-16} derivatives. Rilpivirine (**2**, TMC-278) was approved by the FDA in May 2011 (Figure 1a).^{14, 17} It showed better potency and pharmacological profiles than **1**, and features the same hydrogen bonding sites as present in **1**.^{6, 13} Another NNRTI, Dapivirine (**3**, TMC-120), is in Phase III clinical trials by the International Partnership for Microbicides for the prevention of HIV infection with aid of a vaginal ring, which can provide women with a protection for a longer period of time.^{18, 19}

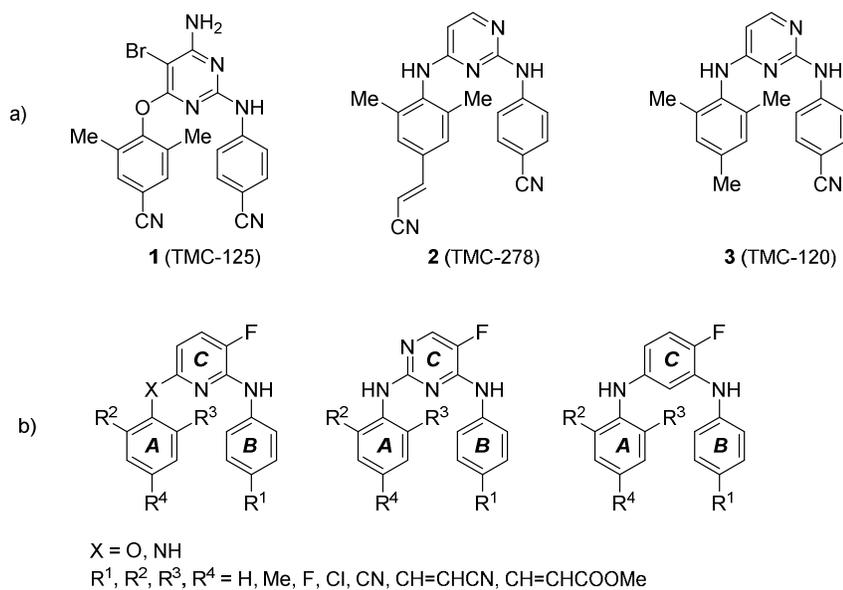


Figure 1. (a) Structures of reported NNRTI **1-3**. (b) General formula of the new NNRTI reported in this paper.

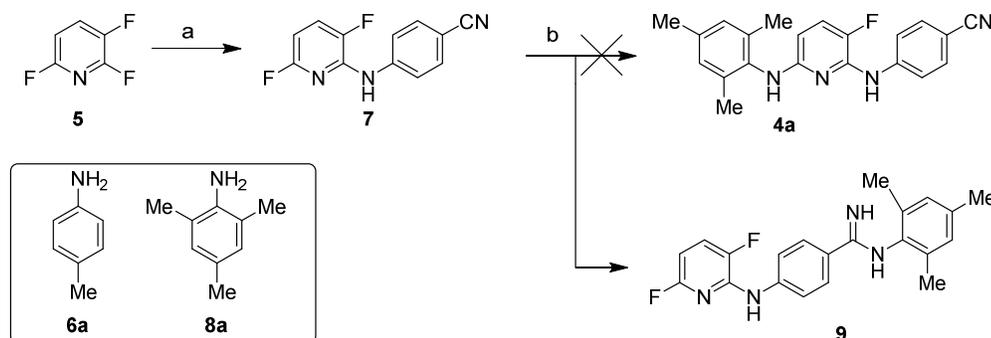
In a quest for novel NNRTI with improved activity, especially towards drug-resistant strains of HIV, we designed 2,6-di(arylamino)-3-fluoropyridine derivatives (Figure 1b). These compounds are structurally similar to **2**, **3** and analogues but feature a 2,6-disubstituted 3-fluoropyridine instead of a 2,4-disubstituted pyrimidine moiety. We hypothesized that a fluorine atom on the ring C can also participate in the interactions with the protein backbone, by way of acting as a hydrogen bond acceptor similarly to a nitrogen atom in the pyrimidine ring of **2**, and/or other weak forces which are unique for the fluorine atom such as orthogonal C=O...F-C interactions.²⁰ To establish the effect of the ring C on the SAR in the fluorinated series, derivatives featuring a pyrimidine and a benzene ring C were synthesized, too (Figure 1b).

Results and discussion.

Synthesis. Our initial target was **4a**, closely resembling the structure of the above-mentioned NNRTI **3**. Our original idea, which was inspired by limited information available in the patent literature,²¹ was to take advantage of the commercial availability of 2,3,6-trifluoropyridine (**5**). The selective nucleophilic substitution in **5** was achieved upon

reaction with deprotonated 4-aminobenzonitrile (**6a**) and gave 46% yield of 4-[(3,6-difluoropyridin-2-yl)amino]benzonitrile (**7**) (Scheme 1). The structure of **7** was established by XRD analysis and confirmed the substitution exclusively in position 2 of the pyridine core. However, the attempted reaction between **7** and 2,4,6-trimethylaniline (**8a**) in NMP did not give the desired **4a**, but instead produced a small amount of the amidine **9** due to the competitive attack of the aniline nucleophile on the CN group.

Scheme 1. Initial attempts to synthesize **4a** from 2,3,6-trifluoropyridine^a



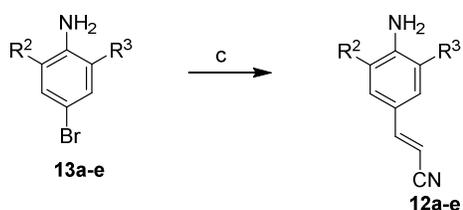
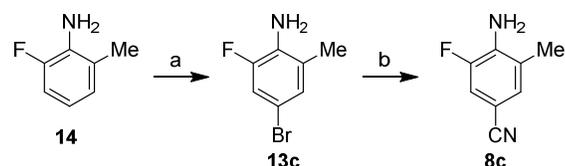
^aReagents and conditions: (a) **6a**, *t*-BuLi, THF, -78°C , then 2 h at 0°C , 46%; (b) **8a**, NMP, TsOH or MsOH, 180°C .

We therefore turned our attention to regioselective Pd-catalyzed amination on commercially available 2,6-dichloro-3-fluoropyridine (**10**), as examples of regioselective Pd-catalyzed aminations involving halogenated pyridines were previously reported.²²⁻²⁸ A reaction between **10** and **6a** in the presence of a Pd source and XPhos ligand afforded the desired 4-[(6-chloro-3-fluoropyridin-2-yl)amino]benzonitrile (**11a**), albeit in modest yield (Table S1₁). Optimization of the catalytic system (Pd source, ligand, base additive) was therefore undertaken. Xantphos ligand in combination with *t*-BuONa as a base and Pd(OAc)₂ as a metal source proved to be the catalytic system of choice. Further, it is essential to stress that the reaction proceeds with complete regioselectivity, the substitution occurred exclusively at position 2 and no substitution at position 6 was observed, as confirmed by the single crystal XRD analysis. A second Pd-catalyzed amination reaction of the intermediate **11a** with 2,4,6-trimethylaniline (**8a**) finally provided the target compound **4a** (Table S1₁ and Scheme 3). It was found that in this case, XPhos ligand provided an acceptable yield of the amination product.

With the optimized synthetic protocol in hand, we proceeded with the synthesis of a library of 2,6-di(arylamino)-3-fluoropyridine derivatives **4a-u** starting from **10** and various anilines (**6a-d**, **8a-c**, **12a-f**, Scheme 3). Based on the knowledge about SAR of di(arylamino)pyrimidine NNRTI^{10,12} we envisaged variation of *para*-substituents (R¹) on the ring B and *ortho*- (R², R³) and *para*-substituents (R⁴) on the ring A of 2,6-di(arylamino)-3-fluoropyridine derivatives (Figure 1b).

Particular attention was given to compounds with $R^4 = (E)\text{-CH=CHCN}$, based on the high activity of **2** (Figure 1a). The anilines **12a–f** necessary for the synthesis of those compounds were prepared via a Heck reaction starting from the corresponding 4-bromoanilines **13a–e**. The latter were commercially available with the exception of 4-bromo-2-fluoro-6-methylaniline (**13c**), which was prepared by bromination of commercially available 2-fluoro-6-methylaniline (**14**) (Scheme 2). 4-Amino-3-fluoro-5-methylbenzonitrile (**8c**) was prepared following the published general method (Scheme 2).²⁹

Scheme 2. Synthesis of the aniline reagents **12a–g**^a



13a $R^2 = R^3 = \text{Me}$

13b $R^2 = \text{H}, R^3 = \text{Me}$

13c $R^2 = \text{F}, R^3 = \text{Me}$

13d $R^2 = R^3 = \text{H}$

13e $R^2 = R^3 = \text{F}$

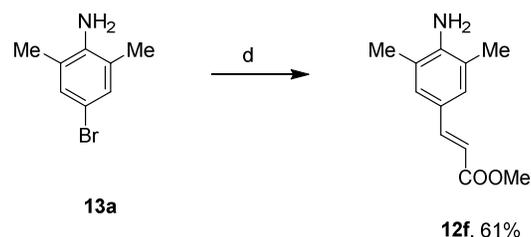
12a $R^2 = R^3 = \text{Me}$, 80%

12b $R^2 = \text{H}, R^3 = \text{Me}$, 74%

12c $R^2 = \text{F}, R^3 = \text{Me}$, 89%

12d $R^2 = R^3 = \text{H}$, 63%

12e $R^2 = R^3 = \text{F}$, 69%

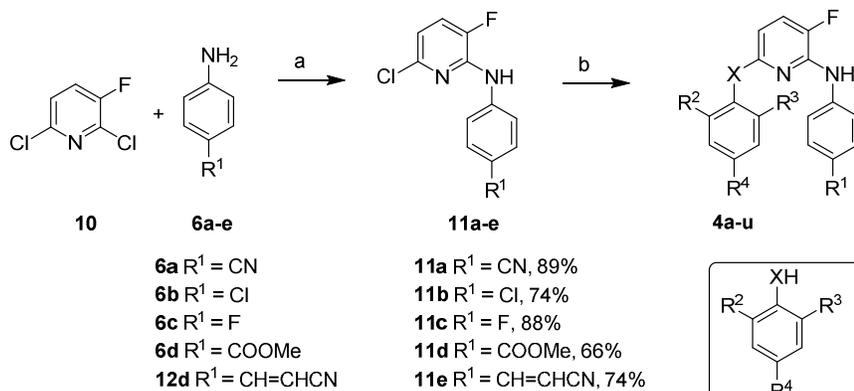


^aReagents and conditions: (a) Br_2 , AcOH, rt, 2 h, 82%; (b) CuCN, DMF, CuI, KI, *N,N'*-dimethyl-1,2-ethanediamine, 110 °C, 16 h, 74%; (c) acrylonitrile, Pd(OAc)₂, P(*o*-tolyl)₃, AcONa·3H₂O, DMA, 140 °C, 48 h; (d) methyl acrylate, Pd(OAc)₂, P(*o*-tolyl)₃, AcONa·3H₂O, DMA, 140 °C, 48 h, 29%.

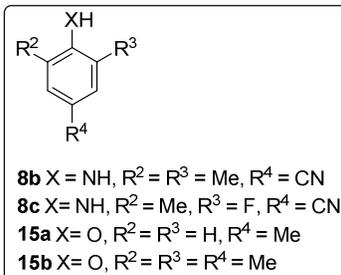
With the anilines in hand a series of 2,6-di(arylamino)-3-fluoropyridine derivatives **4a–k,n–u** was synthesized using our optimized two-step Pd-catalyzed amination protocol (Scheme 3). Similarly, two analogues (**4l,m**) featuring an O rather than an NH linkage between the ring A and the ring C were prepared using a Pd-catalyzed C–O bond formation reaction of **11a** with phenols **15a,b** instead of an aniline (Scheme 3).

1 With the aim to study the effect of the ring C on the biological activity, we prepared two analogues of **4a-u** featuring a
2 pyrimidine or benzene ring C. Following a synthetic route similar to that used for the preparation of **4a-u**, a Pd-catalyzed
3 amination reaction of 2,4-dichloro-5-fluoropyrimidine (**16**) with 4-aminobenzonitrile (**6a**) produced the intermediate 4-
4 [(2-chloro-5-fluoropyrimidin-4-yl)amino]benzonitrile (**17**) as a single regioisomer. Subsequent amination of **17** with the
5 corresponding anilines (**8b**, **12a**) provided the targets **18a,b** in modest yields (Scheme 4). Furthermore, compounds **19a-c**
6 were prepared starting from commercially available 2-bromo-4-chloro-1-fluorobenzene (**20**) as shown in Scheme 5. The
7 bromine atom in **20** was selectively substituted to produce 4-[(5-chloro-2-fluorophenyl)amino]benzonitrile (**21**), which
8 was then converted to **19a-c** via Pd-catalyzed amination with anilines **12a,b,d** (Scheme 5).
9
10
11
12
13
14
15
16

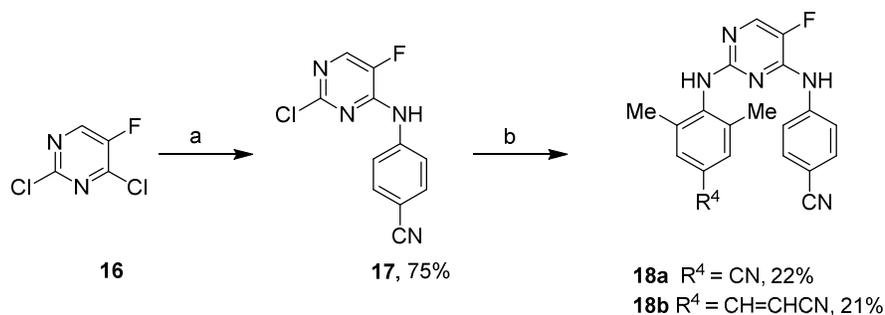
17 Finally, in order to unambiguously evaluate the effect of the fluorine atom in the 3-position of the ring C, reference
18 compounds **22a-d** lacking a fluorine atom were synthesized. Starting from the commercially available 2,6-
19 dichloropyridine (**23**) the intermediate 4-[(6-chloropyridin-2-yl)amino]benzonitrile (**24**) was prepared and then used in a
20 second Pd-catalyzed amination step to obtain **22a-d** (Scheme 6).
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Scheme 3. Synthesis of 2,6-di(arylamino)-3-fluoropyridine derivatives 4a-u^a

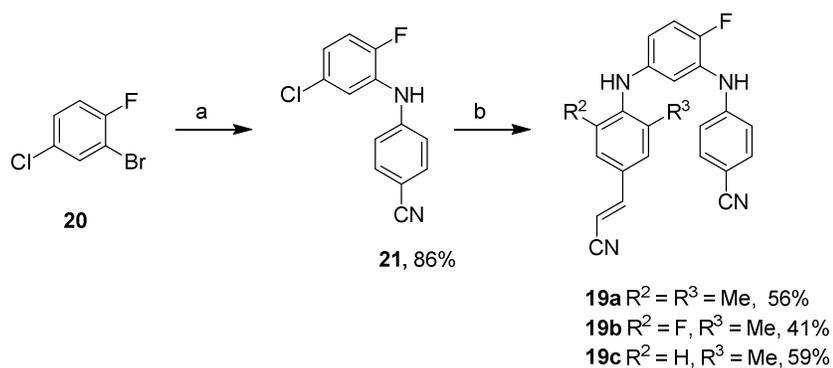
4a X = NH, R¹ = CN, R² = R³ = R⁴ = Me, 53%
4b X = NH, R¹ = CN, R² = R³ = Me, R⁴ = CN, 61%
4c X = NH, R¹ = CN, R² = R³ = Me, R⁴ = CH=CHCN, 62%
4d X = NH, R¹ = CN, R² = R³ = Me, R⁴ = CH=COOMe, 71%
4e X = NH, R¹ = F, R² = R³ = Me, R⁴ = CH=CHCN, 66%
4f X = NH, R¹ = CH=CHCN, R² = R³ = Me, R⁴ = CH=CHCN, 63%
4g X = NH, R¹ = F, R² = R³ = Me, R⁴ = CN, 50%
4h X = NH, R¹ = CN, R² = R³ = H, R⁴ = CH=CHCN, 68%
4i X = NH, R¹ = Cl, R² = R³ = Me, R⁴ = CN, 54%
4j X = NH, R¹ = Cl, R² = R³ = Me, R⁴ = CH=CHCN, 65%
4k X = NH, R¹ = CN, R² = Me, R³ = F, R⁴ = CH=CHCN, 79%
4l X = O, R¹ = CN, R² = R³ = H, R⁴ = Me, 89%
4m X = O, R¹ = CN, R² = R³ = R⁴ = Me, 43%
4n X = NH, R¹ = Cl, R² = Me, R³ = H, R⁴ = CH=CHCN, 73%
4o X = NH, R¹ = F, R² = R³ = Me, R⁴ = CH=COOMe, 55%
4p X = NH, R¹ = Cl, R² = R³ = Me, R⁴ = CH=COOMe, 71%
4q X = NH, R¹ = F, R² = Me, R³ = H, R⁴ = CH=CHCN, 75%
4r X = NH, R¹ = CN, R² = Me, R³ = H, R⁴ = CH=CHCN, 70%
4s X = NH, R¹ = CN, R² = Me, R³ = F, R⁴ = CN, 42%
4t X = NH, R¹ = CN, R² = R³ = F, R⁴ = CH=CHCN, 72%
4u X = NH, R¹ = COOMe, R² = R³ = Me, R⁴ = CH=CHCN, 72%



^aReagents and conditions: (a) Pd(OAc)₂, XantPhos, *t*-BuONa, dioxane, 110 °C, 16 h; (b) **8a-c** or **12a-f** or **15a,b**, Pd(OAc)₂, XPhos, Cs₂CO₃, dioxane, 110 °C, 16 h.

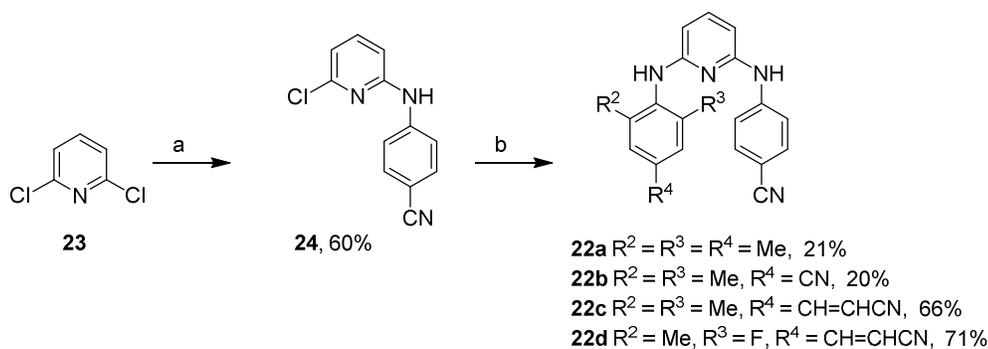
Scheme 4. Synthesis of 2,4-di(arylamino)-5-fluoropyrimidine derivatives **18a,b**^a

^aReagents and conditions: (a) **6a**, Pd₂(dba)₃, XantPhos, Cs₂CO₃, dioxane, 110 °C, 16 h; (b) **8b** or **12a**, Pd(OAc)₂, XPhos, Cs₂CO₃, dioxane, 110 °C, 16 h.

Scheme 5. Synthesis of 1,3-di(arylamino)-4-fluorobenzene derivatives **19a-c**^a

^aReagents and conditions: (a) **6a**, Pd(OAc)₂, XantPhos, *t*-BuONa, dioxane, 110 °C, 16 h; (b) **12a-c**, Pd(OAc)₂, XPhos, Cs₂CO₃, dioxane, 110 °C, 16 h

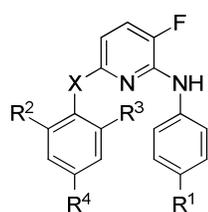
Scheme 6. Synthesis of 2,6-di(arylamino)pyridine derivatives 22a–d



^aReagents and conditions: (a) $\text{Pd}(\text{OAc})_2$, *rac*-BINAP, K_2CO_3 , toluene, 120 °C, 24 h; (b) **8a,b** or **12a,c**, $\text{Pd}(\text{OAc})_2$, XPhos, Cs_2CO_3 , dioxane, 110 °C, 16 h.

In vitro studies. The details on the activity against wild type HIV-1 and cytotoxicity for the new NNRTI are summarized in Tables 1–4. Reference data for **2** and **3** are included for comparison. It should be noted that for molecules containing $\text{CH}=\text{CHCN}$ moiety, in some instances we were able to isolate pure *E* isomers, while a mixture of *E*- and *Z*- isomers was obtained in some other cases, *E*-isomer always being a major one (see Tables 1–4). It was previously reported by Janssen and co-workers that for **2** and a few structural analogues, in vitro activity of *E*-isomers was much higher than that of the corresponding *Z*-isomers. Furthermore, also in this study by Janssen¹⁵ mixtures of *E* and *Z* isomers were examined in some cases. In case of **4c**, we performed activity analysis both on the 9/1 mixture of *E*- and *Z*- isomers and on the pure *E*-isomer, which gave results within the experimental error (Table 1). Based on these observations, we performed screening on the mixtures of *E*- and *Z*- isomers for majority of compounds and assumed that minor amounts of less active *Z*-isomer do not significantly affect the results.

Table 1. Activity and selectivity index of 2,6-di(arylamino)-3-fluoropyridine derivatives 4a–u^a



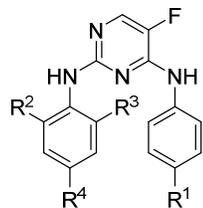
Compound	X	R ¹	R ²	R ³	R ⁴	E/Z	EC ₅₀	SI
							[nM]	

2							100/0	0.72	10700
3								1.4	2000
4a	NH	CN	Me	Me	Me			8.3	2000
4b	NH	CN	Me	Me	CN			4.7	7100
4c	NH	CN	Me	Me	CH=CHCN	100/0	1.5	62500	
4c	NH	CN	Me	Me	CH=CHCN	90/10	1.6	37500	
4d	NH	CN	Me	Me	CH=CHCOOMe	100/0	4.4	13000	
4e	NH	F	Me	Me	CH=CHCN	86/14	6.2	8100	
4f	NH	CH=CHCN	Me	Me	CH=CHCN	88/12 ^b	4.6	19000	
4g	NH	F	Me	Me	CN		51	830	
4h	NH	CN	H	H	CH=CHCN	70/30	480	14	
4i	NH	Cl	Me	Me	CN		15	2000	
4j	NH	Cl	Me	Me	CH=CHCN	93/7	6.3	6600	
4k	NH	CN	Me	F	CH=CHCN	100/0	1.0	>100000	
4l	O	CN	H	H	Me		260	390	
4m	O	CN	Me	Me	Me		8.6	6400	
4n	NH	Cl	Me	H	CH=CHCN	100/0	63	380	
4o	NH	F	Me	Me	CH=CHCOOMe	100/0	84	610	
4p	NH	Cl	Me	Me	CH=CHCOOMe	100/0	53	950	
4q	NH	F	Me	H	CH=CHCN	100/0	400	210	
4r	NH	CN	Me	H	CH=CHCN	87/13	11	24000	
4s	NH	CN	Me	F	CN		8.8	4600	
4t	NH	CN	F	F	CH=CHCN	79/21	3.3	4600	
4u	NH	COOMe	Me	Me	CH=CHCN	100/0	23	2100	

^aThe selectivity index (SI) is defined as CC_{50}/EC_{50} , where EC_{50} is the 50%-effective concentration vs. HIV virus, and CC_{50} is the concentration causing death of 50% of the cells; The EC_{50} value was determined in TZMbl cells against the subtype B R5-tropic

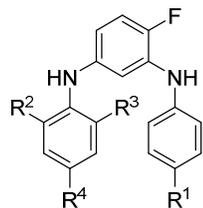
Bal virus and the CC_{50} value was determined in the same cells using the WST-1 assay. ^bRatio (*E,E*)/sum of (*E,Z*) and (*Z,E*) isomers.

Table 2. Activity and selectivity index of 2,4-di(arylamino)-5-fluoropyrimidine derivatives 18a,b



Compound	R ¹	R ²	R ³	R ⁴	E/Z	EC ₅₀ [nM]	SI
18a	CN	Me	Me	CN		2.8	> 36000
18b	CN	Me	Me	CH=CHCN	86/14	0.9	> 42000

Table 3. Activity and selectivity index of 1,3-di(arylamino)-4-fluorobenzene derivatives 19a-c



Compound	R ¹	R ²	R ³	R ⁴	E/Z	EC ₅₀ [nM]	SI
19a	CN	Me	Me	CH=CHCN	81/19	4.5	7400
19b	CN	Me	F	CH=CHCN	75/25	11	6100
19c	CN	Me	H	CH=CHCN	89/11	58	89

From the data obtained it is clear the two ortho-substituents on the ring A (R², R³) are essential, and replacement of one or both substituents by a hydrogen atom results in a significant drop in activity (e.g. **4c** vs. **4h,r**; **4j** vs. **4n**; **4m** vs. **4l**; **4e** vs. **4q**, Table 1). Substitution of one or both methyl groups by fluorine atoms in the ring A provided only an insignificant effect (slight increase in activity and selectivity for **4k** vs **4c**; slight decrease for **4t** vs. **4c** and for **4s** vs **4b**, Table 1). A

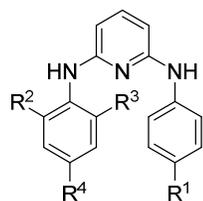
1 cyanovinyl group in the para-position of the ring A produced the best results (**4c** vs. **4b,d**), as was expected from the
 2 published results on **2** and analogues (Table 1).^{10, 12, 16} Finally, the nature of the linker between the ring A and the ring C
 3 had no effect (**4m** vs. **4a**, Table 1).
 4
 5

6
 7 Concerning the variation of the ring B, a cyano group was clearly superior to other alternatives (**4b** vs **4g,i**, **4c** vs
 8 **4e,f,j,u**, and **4d** vs **4o,p**, Table 1). Finally, variation of the ring C provided similar results for pyridine and pyrimidine
 9 derivatives (**4b** vs. **18a**, **4c** vs. **18b**, Table 2), while derivatives with the benzene ring C systematically show a somewhat
 10 lower activity (**4c** vs **19a**, **4k** vs. **19b**, **4r** vs **19c**, Table 3).
 11
 12

13
 14 Overall, most successful were compounds closely resembling the known NNRTI **2**, namely **4c,k** and **18b**, all displaying
 15 EC₅₀ close to 1.0 nM and a very high selectivity index (> 40000).
 16
 17

18
 19 In order to unambiguously compare the effect of the fluorine atom on the anti-HIV activity of 2,6-di(arylamino)-3-
 20 fluoropyridine derivatives, four analogues **22a-d** lacking a fluorine atom on the pyridine ring were evaluated (Table 4). In
 21 every case (**4a** vs **22a**, **4b** vs **22b**, **4c** vs **22c** and **4k** vs **22d**) removal of a fluorine atom decreased the activity and selectivity
 22 (Table 4). In case of **4k** vs **22d** this effect was the most pronounced; an increase of activity by ca. a factor of 6 and of the SI
 23 by ca. a factor of 20 due to the presence of a F atom.
 24
 25
 26
 27
 28
 29

30 **Table 4. Activity and selectivity index of compounds 22a-d.**



41
 42
 43
 44
 45
 46
 47
 48
 49
 50
 51
 52

Compound	R ¹	R ²	R ³	R ⁴	E/Z	EC ₅₀ [nM]	SI
22a	CN	Me	Me	Me		30	1400
22b	CN	Me	Me	CN		25	930
22c	CN	Me	Me	CH=CHCN	80/20	3.5	11000
22d	CN	Me	F	CH=CHCN	92/8	5.5	5500

53
 54
 55
 56
 57
 58
 59
 60

For selected molecules which displayed a high activity against WT HIV-1, data were collected for mutant HIV-1 strains, which have shown resistance against Nevirapine (Y181C) and Efavirenz (L100I, K103N), and site-directed mutants (pNL4.3-K103N, pNL4.3-K103N-Y181C) (Table 5). The activity of **4c** and **4k** is close to or slightly higher than that of earlier reported **3**, but lower than that of **2** for all strains (WT, single and double mutant). Further comparison of three 2,6-di(arylamino)-3-fluoropyridine derivatives with their analogues lacking fluorine clearly reveals for every case (**4b** vs. **22b**, **4c** vs. **22c** and **4k** vs. **22d**) higher activity of the fluorinated derivatives and thus further underlines the advantages of a F atom on the ring C.

Table 5. Antiviral activity of selected compounds vs mutant HIV-1^a

Compound	Y181C	L100I, K103N	pNL4.3-K103N	pNL4.3-K103N-Y181C
	EC ₅₀ [nM]	EC ₅₀ [nM]	EC ₅₀ [nM]	EC ₅₀ [nM]
2	2.5	3.6	0.7	3.1
3	11	> 1000	2.8	92
4a	130	> 10000		
4b	91	> 10000		
22b	1500	6600		
4c	8.1	540	7.8	554
22c	91	> 1000		
4k	25	272	36	> 1000
22d	255	1100		

^aThe EC₅₀ values were determined in TZMbl cells against Nevirapine-resistant VI829 (carrying Y181C) and Efavirenz-resistant VI829 (carrying L100I and K103N). Both viruses are subtype C and R5-tropic. Site-directed mutants (SDMs) carrying K103N and K103N+Y181C were used to further evaluate **2**, **3**, **4c** and **4k**. Construction of the SDMs is described elsewhere.³⁰

Docking studies. Docking of the **4a** and **22a** ligands in the crystal structure of RT in complex with **3**¹³ was performed using the Gromacs molecular dynamics and mechanics software.³¹ A comparison between the docked poses of **3**, **4a** and **22a** shows little variation between the three ligands. In all cases, a hydrogen bond between the backbone oxygen of Lys-101 of the RT and the aniline nitrogen bridging the central aromatic ring with the nitrile-substituted ring is observed. The

1 main difference between the binding of **4a** on the one hand, and **3** and **22a** on the other hand, is the presence of a weak
2
3 interaction between the fluorine substituent of **4a** and the C ϵ hydrogen of Lys-103 (Figure SI5, see Supporting
4
5 Information). A corresponding interaction is not found in **3** due to the fact that this compound does not contain a
6
7 fluorine atom at this position and that the corresponding pyrimidine nitrogen of **3** is located too far away from the
8
9 corresponding C ϵ hydrogen of Lys-103. Similarly, in **22a** this fluorine atom is substituted with a hydrogen atom, bearing a
10
11 partially positive charge (Figure B, supporting information) and therefore incapable of forming a favorable electrostatic
12
13 interaction with the C ϵ hydrogen of Lys-103. These docking experiments corroborate the experimental findings, as the
14
15 differences in EC₅₀ between the three molecules in question are comparatively small.
16

17 Conclusions

18
19 In summary, we have developed a short and practical synthesis of new NNRTI, which are similar in structure to earlier
20
21 described di(arylamino)pyrimidines but feature a 2,6-disubstituted 3-fluoropyridine instead of a 2,4-disubstituted
22
23 pyrimidine moiety. The in vitro studies of a synthesized library confirmed a SAR similar to that observed earlier for the
24
25 di(arylamino)pyrimidines series, as well as for the recently published 3-aminopyridine derivatives.³² An important
26
27 observation is that the presence of a fluorine atom on the central pyridine ring is essential for the antiviral activity, both
28
29 against the wild type HIV and the drug-resistant mutant strains. The activity of the best molecules **4c,k** was, for all
30
31 studied HIV strains, close to or slightly higher than that of a well-known NNRTI API **3**.
32
33
34
35

36 Experimental part

37
38 **Instrumentation and Chemicals.** All reagents, chemicals and solvents were obtained from commercial sources and
39
40 used without extra purification unless stated otherwise. Melting points were determined on a Büchi apparatus and are
41
42 uncorrected. The ¹H NMR and ¹³C NMR spectra were recorded in CDCl₃, CD₃COCD₃, CD₃OD or DMSO-*d*₆ on a Bruker
43
44 Avance II 400 spectrometer with TMS as the internal standard. Coupling constants are given in Hertz and the chemical
45
46 shifts are given in ppm. For mass spectrometric analysis, samples were dissolved in CH₃OH containing 0.1% formic acid
47
48 and diluted to a concentration of approximately 10⁻⁵ mol/L. Injections (1 μ L) were directed to the mass spectrometer at a
49
50 flow rate of 5 μ L/min (CH₃OH and 0.1% formic acid), using a CapLC HPLC system (Waters-Micromass). High resolution
51
52 mass data were acquired on a Q-TOF 2 mass spectrometer (Waters-Micromass) equipped with a standard electrospray
53
54 ionisation (ESI) interface. Column chromatography was performed on Kieselgel 60 (ROCC SI 1721, 40-60 mm), or on an
55
56 automated chromatography system with silica flash cartridges (Grace). Purities of final compounds were determined with
57
58
59
60

1 a UHPLC system based on MS and UV detection. A Waters Aquity UPLC system coupled to a Waters PDA detector and a
2
3 Waters Micromass ZQ ESI mass spectrometer was used. A Halo C₁₈ fused core 2.7 μ m, 2.1mm \times 30mm column was used.
4
5 Solvent A: water with 0.1% formic acid; solvent B: methanol with 0.1% formic acid. Method: 0.5 mL/min, 0.5 min 99% A,
6
7 1% B then in 5.6 min from 99% A, 1% B to 5% A, 95% B then 0.4min, 5% A, 95% B, then in 0.1min to 99% A, 1% B holding
8
9 this for 4.4min before the next analysis. The wavelength for UV detection was 254 nm. All final products had a purity of at
10
11 least 95%, with the exception of compounds **4r** (94%), **4t** (94%) and **18a** (93%).
12

13 **Structural data.** X-ray data were collected on a Bruker platform goniometer equipped with sealed Mo ($\lambda = 0.71073$ Å) X-
14
15 ray tube, pyrrolithic graphite monochromator, and Smart 1000 CCD detector. For data collection and reduction, the
16
17 Bruker SAINT software, V7.66A (Bruker Corporation, Madison, USA, 2010) was employed. Data were corrected for
18
19 absorption with the multi-scan method with SADABS 2008/1 (Bruker Corporation, Madison, USA, 2008). Structures were
20
21 solved with direct methods with SHELXS-97,³³ and refined with SHELXL-2014/7 (George M. Sheldrick, Universität
22
23 Göttingen, 2014) and the shelXle graphical interface.³⁴ CIF files were deposited at the CCDC, reference numbers: 1410424
24
25 (**7**), 1410427 (**11a**), 1410425 (**4a**), 1410426 (**4b**). Ellipsoid plots of all structures were made with ORTEP-3 for Windows³⁵ and
26
27 can be found in the Supporting Information.
28

29 **Biological Assays. Cells.** The TZM-bl cell line (NIH AIDS Research and Reference Reagent Program, Germantown,
30
31 USA) was used for the evaluation of drug susceptibility and cytotoxicity. TZM-bl cells were cultured in Dulbecco's
32
33 Minimum Essential Medium (DMEM) (Lonza) containing 10% heat-inactivated FBS and 50 μ g gentamycin/mL at 37 °C in
34
35 a humidified 5% CO₂, 95% air environment. Twice a week the cells were treated with 0.25% trypsin – 1 mM EDTA (Lonza)
36
37 for 10 minutes. The resulting cell suspension was washed with an equivalent amount of TZM-bl medium and subsequently
38
39 seeded in a T75 culture flask (Greiner Bio-One, Germany) at 10⁶ cells in 20 mL medium.
40

41 **Antiviral assay.** The antiviral activity of the newly designed compounds was measured by pre-incubating 10⁴ TZM-bl
42
43 cells (at 10⁵ cells/mL in culture medium supplemented with 30 μ g/mL DEAE dextran) in a 96-well plate for 30 minutes at
44
45 37 °C, 5% CO₂ in the presence or absence of serial dilutions of the each compound. Subsequently, 200 TCID₅₀ of wild type
46
47 (Bal) or NNRTI-resistant HIV-1 (Y181C, L100I + K103N) was added to each well and cultures were incubated for 48 hours
48
49 before quantifying luciferase activity. Each condition was evaluated in triplicate wells and in at least three independent
50
51 experiments. The antiviral activity of the compound was expressed as the percentage of viral inhibition compared to the
52
53 untreated controls and subsequently plotted against the compound concentration. Non-linear regression analysis was
54
55
56
57
58
59
60

1 used to calculate the 50% effective concentration (EC_{50}) based on at least three independent measurements and using
2
3 GraphPad Prism version 5.03 for Windows (GraphPad Software, San Diego, CA, USA).
4

5 **WST-1 cytotoxicity assay.** The Water Soluble Tetrazolium-1 (WST-1) Cell Proliferation Assay is a colorimetric assay for
6 the measurement of cell proliferation and viability. The assay is based on the cleavage of the tetrazolium salt WST-1 ((4-
7 [3-(4-iodophenyl)-2-(4-nitrophenyl)-2H-5-tetrazolio]-1,3-benzene disulfonate)) to a formazan dye by a complex cellular
8 mechanism. This bioreduction is largely dependent on the glycolytic production of NAD(P)H in viable cells. Therefore,
9 the amount of formazan dye formed correlates directly to the number of viable cells in the culture, and can be quantified
10 by measuring the absorbance at 450nm in a multiwell plate reader. The greater the number of viable cells, the greater the
11 amount of formazan dye produced following the addition of WST-1. Cytotoxicity of each compound was evaluated using
12 this WST-1 viability assay, according to the manufacturer's instructions (Roche, Vilvoorde, Belgium). Briefly, 10^4 TZM-bl
13 cells were seeded in a 96-well plate and cultured for 2 days in the presence of a serial dilution of each compound. After
14 48h exposure, Cell Proliferation Reagent was added and absorbance at 450 nm was quantified after 90 min using a
15 microplate reader (BioRad, Tokyo, Japan). Each compound was tested in three replicate wells and in at least three
16 independent experiments. The percentage cell viability, compared to untreated controls, was plotted against the
17 compound concentration and non-linear regression analysis was performed using GraphPad Prism version 5.02 for
18 Windows (GraphPad Software, San Diego, CA, USA) to calculate the 50% cytotoxic concentration (CC_{50}).
19
20
21
22
23
24
25
26
27
28
29
30
31
32

33 **Computational methods. Docking of 4a and 22a.** The crystal structure of RT in complex with **3**³³ was taken as the
34 starting structure and both compounds were positioned in the pocket by superimposing their aromatic ring atoms onto
35 the corresponding atoms of **3**. Positions were refined by energy minimization of both ligand and protein using the
36 GROMOS 53A6 force-field³⁶ within the Gromacs software suite³¹ and until the maximum force dropped below 10 kJ/mol.
37 Force-field parameters and atomic partial charges for **4a** and **22a** were generated with the online ATB tool (version 2.2).³⁷
38
39
40
41
42
43

44 **Electrostatic potential surface of 4a and 22a.** The electrostatic potential surfaces of both compounds after geometry
45 optimization were calculated with Spartan 14 using Hartree-Fock 6-31G** basis set.³⁸
46
47

48 **Synthetic procedures.**

49
50 **4-[(3,6-Difluoropyridin-2-yl)amino]benzotrile (7).** To a solution of 4-aminobenzotrile (0.156 g, 1.320 mmol) in
51 THF (4.5 mL) a solution of *t*-BuLi (1.6 M in pentane; 0.763 mL, 1.22 mmol) was added dropwise at -78 °C. Next, a solution
52 of 2,3,6-trifluoropyridine (0.266 g, 2.0 mmol) in THF (1.5 mL) was added and the mixture was stirred for 5 min at -78 °C.
53
54 The reaction mixture was warmed to room temperature and stirred for 2 h, then aq. NH_4Cl (10 mL) was added and the
55
56
57
58
59
60

1 mixture was extracted with EtOAc (2 x 20 mL). The combined organic fractions were dried over MgSO₄, filtered and
2 concentrated. The resulting oil was purified by column chromatography to give **7** as a white solid (128 mg, 46%); mp 120–
3 122 °C (CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃): δ 6.40 (ddd, *J* = 8.5 Hz, 3.3 Hz, 2.3 Hz, 1H), 6.86 (br, 1 H), 7.43 (ddd, *J* = 9.6
4 Hz, 8.5 Hz, 6.1 Hz, 1H), 7.62 (d, *J* = 8.9 Hz, 2 H), 7.77 (d, *J* = 8.9 Hz, 2 H); ¹³C NMR (100 MHz, CDCl₃): δ 99.1 (dd, *J* = 40.8
5 Hz, 3.6 Hz), 105.2, 118.4 (2 C), 119.2, 126.0 (dd, *J* = 19.1 Hz, 9.4 Hz), 133.5 (2 C), 141.6 (dd, *J* = 19.1 Hz, 9.4 Hz), 143.0, 144.2 (dd,
6 *J* = 243.6 Hz, 5.4 Hz), 157.2 (d, *J* = 236.8 Hz). HRMS (ESI) *m/z* calcd. for C₁₂H₇F₂N₃ ([M+H]⁺): 231.0608, found: 231.0618.
7 Single crystals for XRD analysis were grown by the slow evaporation from a dichloromethane solution. CCDC Nr.
8 1410424.
9

10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

4-[(3,6-Difluoropyridin-2-yl)amino]-*N*-mesitylbenzimidamide (9). A mixture of nitrile **7** (0.231 g, 1.0 mmol), MeSO₃H (0.065 mL, 1.0 mmol) and 2,4,6-trimethylaniline (0.281 mL, 2 mmol) in *N*-methyl-2-pyrrolidinone (0.5 mL) was placed in a 10 mL microwave vial and heated in the microwave oven (300 W output) for 1 h at 180 °C. The mixture was cooled to room temperature, a solution of Na₂CO₃ (10% in H₂O, 10 mL) was added and the mixture was extracted with EtOAc (3 x 10 mL). The residue was purified by column chromatography to give **9** (33 mg, 9%) as a brownish solid; mp >240 °C (dec). ¹H NMR (400 MHz, CDCl₃): δ 2.09 (s, 6H), 2.25 (s, 3H), 4.53 (br, 2 H), 6.26 (ddd, *J* = 8.5 Hz, 3.1 Hz, 2.4 Hz, 1H), 6.85 (s, 2 H), 6.88 (br, 1 H), 7.33 (ddd, *J* = 9.5 Hz, 8.5 Hz, 6.2 Hz, 1H), 7.64 (d, *J* = 8.9 Hz, 2 H), 7.84 (d, *J* = 8.9 Hz, 2 H); ¹³C NMR (100 MHz, CDCl₃): δ 17.7 (2 C), 20.7, 97.5 (dd, *J* = 41.2 Hz, 3.4 Hz), 118.6 (2 C), 125.4 (dd, *J* = 18.1 Hz, 9.4 Hz), 127.7 (2 C), 128.77 (2 C), 128.84, 129.9, 131.9, 140.9 (2 C), 142.5 (dd, *J* = 18.1 Hz, 12.9 Hz), 143.7, 144.1 (dd, *J* = 252.5 Hz, 5.5 Hz), 153.0, 157.3 (dd, *J* = 235.0 Hz, 1.5 Hz); HRMS (ESI) *m/z* calcd. for C₂₁H₂₁F₂N₄ ([M+H]⁺): 367.1734, found: 367.1720.

4-Bromo-2-fluoro-6-methylaniline (13c).³⁹ A round bottomed flask was charged with 2-fluoro-6-methylaniline (12 mmol, 1.5 g) and acetic acid (20 mL). Bromine (12 mmol, 0.616 mL) was added dropwise, and the reaction mixture was stirred for 2 h at room temperature. The reaction mixture was evaporated, neutralized with saturated aq. Na₂CO₃ solution, and extracted with CH₂Cl₂ (3 x 50 mL). Combined organic phases were dried over Mg₂SO₄ and evaporated in vacuo. The residue was purified with an automated chromatography system using Silica Flash Cartridges applying a heptane-ethyl acetate gradient (from 100% heptane to 100% ethylacetate in 35 min, 35 mL/min). Obtained as red liquid; yield 82% (2.0 g); ¹H NMR (400 MHz, CDCl₃): δ 2.16 (s, 3H), 3.63 (s, 2H), 6.98 (s, 1H), 7.02 (dd, *J* = 10.1 Hz, 2.1 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 17.4, 108.8 (d, *J* = 10.1 Hz), 116.7 (d, *J* = 23.1 Hz), 126.7 (d, *J* = 4.1 Hz), 128.8 (d, *J* = 2.9 Hz), 132.7 (d, *J* = 12.4 Hz), 151.8 (d, *J* = 240.5 Hz).

1 **4-Amino-3-fluoro-5-methylbenzotrile (8c).**⁴⁰ A 50 mL round bottomed flask was charged with CuCN (2.95 mmol,
2 264 mg), CuI (0.25 mmol, 47 mg), KI (0.52 mmol, 86 mg), *N,N'*-dimethyl-1,2-ethanediamine (2.46 mmol, 217 mg), 4-
3 bromo-2-fluoro-6-methylaniline (2.45 mmol, 500 mg) and dry DMF (20 mL). The resulting mixture was heated to 110 °C
4 (oil bath temperature) for 16 h. After the reaction, DMF was evaporated and aqueous 28% NH₃ solution (50 mL) was
5 added. The resulting mixture was extracted with dichloromethane, washed with brine solution and dried over MgSO₄.
6 The product was purified with an automated chromatography system using Silica Flash Cartridges applying a heptane-
7 ethyl acetate gradient (from 100% heptane to 80% ethylacetate in 35 min, 35 mL/min). Obtained as white solid; yield 74%
8 (0.272 g); mp 119–120 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.19 (s, 3H), 4.20 (brs, 2H), 7.10–7.16 (m, 2H); ¹³C NMR
9 (100 MHz, CDCl₃): δ 17.0 (d, *J* = 3.1 Hz), 99.3 (d, *J* = 9.7 Hz), 116.7 (d, *J* = 22.0 Hz), 119.3 (d, *J* = 3.0 Hz), 124.6 (d, *J* = 4.2 Hz),
10 130.2 (d, *J* = 2.4 Hz), 138.1 (d, *J* = 11.9 Hz), 149.8 (d, *J* = 240.6 Hz).

11 **General Procedure for synthesis of anilines 12a–f (General Procedure A):** A solution of Pd catalyst in DMA was
12 prepared first. A flask was charged with Pd(OAc)₂ (0.2 mmol, 45 mg) and tri(*o*-tolyl)phosphine (0.4 mmol, 0.122 g), dry
13 DMA (4 mL) was added and the resulting solution was stirred for 15 min under an argon atmosphere. Next, a 100 mL
14 round bottomed flask was charged with a 4-bromoaniline **9a–e** (4 mmol), acrylonitrile (6.0 mmol, 0.32 g),
15 tetrabutylammonium chloride (4.0 mmol, 1.12 g) and CH₃COONa·3H₂O (4.0 mmol, 0.54 g). The freshly prepared Pd
16 catalyst solution was added, the resulting mixture was stirred for 2 min under an argon atmosphere, and then heated to
17 reflux (oil bath temperature 140 °C) for 48 h. The reaction mixture was allowed to cool down to room temperature and
18 filtered through Celite. The Celite cake was washed with toluene (100 mL), combined filtrates were washed with water
19 (100 mL), brine (50 mL), dried with MgSO₄ and evaporated in vacuo. The residue was separated with an automated
20 chromatography system using Silica Flash Cartridges applying a heptane-ethyl acetate gradient (from 100% heptane to
21 80% ethylacetate in 35 min, 35 mL/min). Compounds **12a**,⁴¹ **12b**,⁴² and **12d**⁴¹ were reported earlier. Data for compounds
22 **12c,e,f** are given below.

23 **3-(4-Amino-3-fluoro-5-methylphenyl)acrylonitrile (12c).** Prepared according to the General Procedure A from
24 Pd(OAc)₂ (0.16 mmol, 36 mg), tri(*o*-tolyl)phosphine (0.32 mmol, 97 mg), 4-bromo-2-fluoro-6-methylaniline (3.19 mmol,
25 0.65 g), acrylonitrile (4.78 mmol, 0.25 g), tetrabutylammonium chloride (3.19 mmol, 0.86 g) and AcONa·3H₂O (3.19 mmol,
26 0.43 g). Obtained as a mixture of geometrical isomers (*Z*:*E* 1:3.13), light yellow solid; yield 89% (0.0.49 g); mp 98–100 °C
27 (AcOEt); ¹H NMR (400 MHz, CDCl₃): signals of major (*E*)-isomer: δ 2.19 (s, 3H), 4.03 (brs, 2H), 5.60 (d, *J* = 16.5 Hz, 1H),
28 6.94 (s, 1H), 6.99 (dd, *J* = 11.4 Hz, 1.6 Hz, 1H), 7.19 (d, *J* = 16.5 Hz, 1H); signals of minor (*Z*)-isomer: δ 2.21 (s, 3H), 5.19 (d, *J* =
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

12.1 Hz, 1H), 6.88 (d, $J = 12.1$ Hz, 1H), 7.29 (s, 1H), 7.47 (dd, $J = 11.8$ Hz, 1.6 Hz, 1H), signal of NH overlapped with the signal of major isomer; ^{13}C NMR (100 MHz, CDCl_3): signals of major (*E*)-isomer: δ 16.9 (d, $J = 3.2$ Hz), 92.3, 111.3 (d, $J = 19.8$ Hz), 118.9, 123.2 (d, $J = 7.7$ Hz), 124.2 (d, $J = 3.8$ Hz), 125.9 (d, $J = 2.0$ Hz), 136.2 (d, $J = 12.5$ Hz), 149.9 (d, $J = 2.8$ Hz), 151.0 (d, $J = 239.5$ Hz); signals of minor (*Z*)-isomer: δ 90.7, 113.3 (d, $J = 20.1$ Hz), 118.1, 123.2 (d, $J = 7.7$ Hz), 123.9 (d, $J = 3.9$ Hz), 127.7 (d, $J = 2.1$ Hz), 136.1 (d, $J = 12.5$ Hz), 147.7 (d, $J = 2.8$ Hz), 150.6 (d, $J = 239.2$ Hz), signal of CH_3 overlapped with the signal of major isomer. HRMS (ESI) m/z calcd. for $\text{C}_{10}\text{H}_{10}\text{FN}_2$ ($[\text{M}+\text{H}]^+$): 177.0828, found: 177.0832.

3-(4-Amino-3,5-difluorophenyl)acrylonitrile (12e). Prepared according to the General Procedure A from $\text{Pd}(\text{OAc})_2$ (0.25 mmol, 56 mg), tri(*o*-tolyl)phosphine (0.5 mmol, 152 mg), 4-bromo-2,6-difluoroaniline (5.0 mmol, 1.04 g), acrylonitrile (7.5 mmol, 0.398 g), tetrabutylammonium chloride (5.0 mmol, 1.39 g) and $\text{AcONa}\cdot 3\text{H}_2\text{O}$ (5.0 mmol, 0.680 g). Obtained as a mixture of geometrical isomers (*Z*:*E* 1:3.13), white solid; yield 69% (0.62 g); mp 173-174 °C (AcOEt); ^1H NMR (400 MHz, CDCl_3): signals of major (*E*)-isomer: δ 4.09 (brs, 2H), 5.64 (d, $J = 16.5$ Hz, 1H), 6.95 (dd, $J = 7.0$ Hz, 2.2 Hz, 2H), 7.18 (d, $J = 16.5$ Hz, 1H); signals of minor (*Z*)-isomer: δ 5.29 (d, $J = 12.1$ Hz, 1H), 6.87 (d, $J = 12.1$ Hz, 1H), 7.36 (dd, $J = 7.3$ Hz, 2.2 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3): signals of major (*E*)-isomer: δ 94.2, 110.3 (dd, $J = 14.9$ Hz, 7.4 Hz, 2 C), 118.2, 122.2 (t, $J = 8.7$ Hz), 127.1 (t, $J = 16.4$ Hz), 148.6 (t, $J = 3.0$ Hz), 151.4 (dd, $J = 242.0$ Hz, 8.5 Hz, 2 C); signals of minor (*Z*)-isomer: δ 92.6, 112.1 (dd, $J = 15.2$ Hz, 7.4 Hz, 2 C), 117.5, 146.5 (t, $J = 3.0$ Hz), 150.93 (dd, $J = 241.3$ Hz, 8.7 Hz, 2 C); other signals are overlapped with signals of major isomer. HRMS (ESI) m/z calcd. for $\text{C}_9\text{H}_7\text{F}_2\text{N}_2$ ($[\text{M}+\text{H}]^+$): 181.0577, found: 181.0574.

(*E*)-Methyl 3-(4-amino-3,5-dimethylphenyl)acrylate (12f). Prepared according to the General Procedure A from $\text{Pd}(\text{OAc})_2$ (0.2 mmol, 45 mg), tri(*o*-tolyl)phosphine (0.4 mmol, 122 mg), 4-bromo-2,6-dimethylaniline (4.0 mmol, 0.800 g), methyl acrylate (6.0 mmol, 0.52 g), tetrabutylammonium chloride (4.0 mmol, 1.11 g) and $\text{AcONa}\cdot 3\text{H}_2\text{O}$ (4.0 mmol, 0.544 g). Analytical data correspond to those found in literature. White solid; yield 61% (0.49 g); mp 83-84 °C (AcOEt); ^1H NMR (400 MHz, CDCl_3): δ 2.17 (s, 3H), 3.77 (s, 3H), 3.86 (brs, 2H), 6.23 (d, $J = 15.9$ Hz, 1H), 7.14 (s, 2H), 7.57 (d, $J = 15.9$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 17.5 (2 C), 51.4, 112.8, 121.5 (2 C), 124.1, 128.8 (2 C), 145.4, 145.6, 168.2. HRMS (ESI) m/z calcd. for $\text{C}_{12}\text{H}_{16}\text{NO}_2$ ($[\text{M}+\text{H}]^+$): 206.1181, found: 206.1188.

General Procedure B for the Synthesis of intermediates 11a–e. A solution of Pd catalyst in dioxane was prepared first. A flask was charged with $\text{Pd}(\text{OAc})_2$ (0.025 mmol, 5.6 mg) and Xantphos (0.03 mmol, 17 mg), dry dioxane (5 mL) was added and the resulting solution was stirred for 15 min under an argon atmosphere. Next, a 50 mL round bottomed flask was charged with 2,6-dichloro-3-fluoropyridine (0.5 mmol, 83 mg), an aniline **2a–d** or **8d** (0.6 mmol) and *t*-BuONa (0.7 mmol, 67 mg). The freshly prepared solution of Pd catalyst was added, the resulting mixture was stirred for 2 min under

1 an argon atmosphere, and then heated to reflux (oil bath temperature 110 °C) for 16 h. The reaction mixture was allowed
2
3 to cool down to room temperature and filtered through Celite. The Celite cake was washed with CH₂Cl₂ (100 mL),
4
5 combined organic phases were evaporated in vacuo. The residue was separated with an automated chromatography
6
7 system using Silica Flash Cartridges applying a heptane-ethyl acetate gradient (from 100% heptane to 100% ethylacetate in
8
9 35 min, 35 mL/min).

11 **4-[(6-Chloro-3-fluoropyridin-2-yl)amino]benzotrile (11a)**. Prepared according to the General Procedure B from
12
13 Pd(OAc)₂ (0.025 mmol, 5.61 mg), Xantphos (0.03 mmol, 17 mg), 2,6-dichloro-3-fluoropyridine (0.5 mmol, 83 mg), 4-
14
15 aminobenzotrile (0.6 mmol, 71 mg) and *t*-BuONa (0.7 mmol, 67 mg). Obtained as white solid, yield 89% (0.11 g); mp 171–
16
17 172 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 6.83 (dd, *J* = 8.4 Hz, 2.8 Hz, 1H), 6.87 (brs, 1H), 7.31 (dd, *J* = 10.0 Hz, 8.4 Hz,
18
19 1H), 7.62 (d, *J* = 8.8 Hz, 2H), 7.77 (d, *J* = 8.8 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 105.3, 115.5 (d, *J* = 3.2 Hz), 118.4 (2 C),
20
21 119.2, 124.2 (d, *J* = 18.0 Hz), 133.4 (2 C), 143.0, 143.1 (d, *J* = 3.5 Hz), 143.2 (d, *J* = 11.4 Hz), 146.0 (d, *J* = 253.6 Hz); HRMS (ESI)
22
23 *m/z* calcd. for C₁₂H₈ClFN₃ ([M+H]⁺): 248.0391, found: 248.0397. Single crystals for XRD analysis were grown by the slow
24
25 evaporation from a dichloromethane solution. CCDC Nr. 1410427.

27 **6-Chloro-*N*-(4-chlorophenyl)-3-fluoropyridin-2-amine (11b)**. Prepared according to the General Procedure B from
28
29 Pd(OAc)₂ (0.30 mmol, 68 mg), Xantphos (0.36 mmol, 0.21 g), 2,6-dichloro-3-fluoropyridine (6.02 mmol, 1.0 g), 4-
30
31 chloroaniline (7.23 mmol, 0.92 g) and *t*-BuONa (8.43 mmol, 0.81 g). Obtained as white solid; yield 74% (1.12 g); mp 89–90
32
33 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 6.63 (brs, 1H), 6.74 (dd, *J* = 8.0 Hz, 2.7 Hz, 1H), 7.26 (dd, *J* = 10.0 Hz, 8.0 Hz, 1H),
34
35 7.32 (d, *J* = 8.9 Hz, 2H), 7.61 (d, *J* = 8.9 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 113.9 (d, *J* = 3.2 Hz), 120.2 (2 C), 123.5 (d, *J* =
36
37 17.9 Hz), 127.7 (2 C), 129.0, 137.5, 142.9 (d, *J* = 3.5 Hz), 144.1 (d, *J* = 11.5 Hz), 145.7 (d, *J* = 252.6 Hz); HRMS (ESI) *m/z* calcd. for
38
39 C₁₁H₈Cl₂FN₂ ([M+H]⁺): 257.0049, found: 257.0038.

42 **6-Chloro-3-fluoro-*N*-(4-fluorophenyl)pyridin-2-amine (11c)**. Prepared according to the General Procedure B from
43
44 Pd(OAc)₂ (0.25 mmol, 56 mg), Xantphos (0.30 mmol, 0.17 g), 2,6-dichloro-3-fluoropyridine (5.0 mmol, 0.83 g), 4-
45
46 fluoroaniline (6.0 mmol, 0.58 mL) and *t*-BuONa (7.0 mmol, 0.67 g). Obtained as white solid; yield 88% (1.06 g); mp 87–88
47
48 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 6.56 (brs, 1H), 6.69 (dd, *J* = 8.2 Hz, 2.7 Hz, 1H), 7.06 (t, *J* = 9.1 Hz, 2H), 7.22 (dd, *J*
49
50 = 10.2 Hz, 8.2 Hz, 1H), 7.58 (dd, *J* = 9.1 Hz, 4.7 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃+CD₃COCD₃): δ 113.5 (d, *J* = 3.2 Hz), 115.7
51
52 (d, *J* = 22.6 Hz, 2 C), 121.0 (d, *J* = 7.8 Hz, 2 C), 123.4 (d, *J* = 17.9 Hz), 134.9 (d, *J* = 2.6 Hz), 143.0 (d, *J* = 3.4 Hz), 144.6 (d, *J* = 11.7
53
54 Hz), 145.7 (d, *J* = 252.2 Hz), 158.7 (d, *J* = 242 Hz); HRMS (ESI) *m/z* calcd. for C₁₁H₈ClF₂N₂ ([M+H]⁺): 241.0344, found:
55
56 241.0335.

Methyl 4-[(6-chloro-3-fluoropyridin-2-yl)amino]benzoate (11d). Prepared according to the General Procedure B from Pd(OAc)₂ (0.1 mmol, 22 mg), Xantphos (0.120 mmol, 0.069 g), 2,6-dichloro-3-fluoropyridine (2.0 mmol, 0.332 g), methyl 4-aminobenzoate (2.0 mmol, 0.302 g) and Cs₂CO₃ (10.0 mmol, 3.26 g). white solid; yield 66% (0.367 g); mp 159–160 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 3.89 (s, 3H), 6.78 (dd, *J* = 8.2 Hz, 2.8 Hz, 1H), 6.83 (brs, 1H), 7.27 (dd, *J* = 8.2 Hz, 9.9 Hz, 1H), 7.71 (d, *J* = 8.8 Hz, 2H), 8.02 (d, *J* = 8.8 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 51.9, 114.8 (d, *J* = 2.9 Hz), 117.7 (2 C), 123.8 (d, *J* = 18.0 Hz), 124.0, 131.0 (2 C), 143.0 (d, *J* = 3.6 Hz), 143.2, 143.6 (d, *J* = 11.3 Hz), 145.9 (d, *J* = 253.7 Hz), 166.8; HRMS (ESI) *m/z* calcd. for C₁₃H₁₁ClFN₂O₂ ([M+H]⁺): 281.0493, found: 281.0497.

3-{4-[(6-Chloro-3-fluoropyridin-2-yl)amino]phenyl}acrylonitrile (11e). Prepared according to the General Procedure B from Pd(OAc)₂ (0.05 mmol, 11.0 mg), Xantphos (0.06 mmol, 34 mg), 2,6-dichloro-3-fluoropyridine (1.0 mmol, 166 mg), 3-(4-aminophenyl)acrylonitrile (1.0 mmol, 144 mg) and *t*-BuONa (1.4 mmol, 135 mg). Obtained as a mixture of geometrical isomers (*Z*:*E* 1:9), light yellow solid; yield 74% (0.15 g); mp 169–170 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): signals of major (*E*)-isomer: δ 5.76 (d, *J* = 16.6 Hz, 1H), 6.78 (dd, *J* = 8.2 Hz, 2.8 Hz, 1H), 6.83 (brs, 1H), 7.28 (dd, *J* = 9.9 Hz, 8.2 Hz, 1H), 7.34 (d, *J* = 16.6 Hz, 1H), 7.44 (d, *J* = 8.7 Hz, 2H), 7.71 (d, *J* = 8.7 Hz, 2H); signals of minor (*Z*)-isomer: δ 5.32 (d, *J* = 12.1 Hz, 1H), 7.05 (d, *J* = 12.1 Hz, 1H), 7.83 (d, *J* = 8.8 Hz, 2H); other signals are overlapped with signals of major isomer. ¹³C NMR (100 MHz, CDCl₃): signals of major (*E*)-isomer: δ 93.8, 114.8 (d, *J* = 3.1 Hz), 118.67, 118.71 (2 C), 123.8 (d, *J* = 18.0 Hz), 127.9, 128.6 (2 C), 141.8, 143.0 (d, *J* = 3.5 Hz), 143.6 (d, *J* = 11.4 Hz), 145.9 (d, *J* = 252 Hz), 149.9; HRMS (ESI) *m/z* calcd. for C₁₄H₁₀ClFN₃ ([M+H]⁺): 274.0547, found: 274.0540.

4-[(2-Chloro-5-fluoropyrimidin-4-yl)amino]benzonitrile (17). Prepared according to the General Procedure B from Pd(OAc)₂ (0.25 mmol, 56 mg), Xantphos (0.3 mmol, 0.17 g), 2,4-dichloro-5-fluoropyrimidine (5 mmol, 0.84 g), 4-aminobenzonitrile (5 mmol, 0.59 g) and Cs₂CO₃ (25 mmol, 0.84 g). Obtained as white solid; yield 75% (0.93 g); mp 235–236 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 7.12 (s, 1H), 7.69 (d, *J* = 8.8 Hz, 2H), 7.83 (d, *J* = 8.8 Hz, 2H), 8.18 (d, *J* = 2.5 Hz, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 106.0, 119.5, 121.3 (2 C), 133.5 (2 C), 142.9, 143.2 (d, *J* = 21.2 Hz), 145.9 (d, *J* = 260 Hz), 150.1 (d, *J* = 12.4 Hz), 153.1 (d, *J* = 3.5 Hz); HRMS (ESI) *m/z* calcd. for C₁₁H₇ClFN₄ ([M+H]⁺): 249.0343, Found: 249.0341.

4-[(5-Chloro-2-fluorophenyl)amino]benzonitrile (21). Prepared according to the General Procedure B from Pd(OAc)₂ (0.025 mmol, 5.61 mg), Xantphos (0.03 mmol, 17 mg), 2-bromo-4-chloro-1-fluorobenzene (0.5 mmol, 110 mg), 4-aminobenzonitrile (0.5 mmol, 59 mg) and *t*-BuONa (0.7 mmol, 67 mg). Obtained as white solid; yield 86% (107 mg); mp 141–142 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 6.04 (brs, 1H), 6.96 (ddd, *J* = 8.8 Hz, 4.3 Hz, 2.5 Hz, 1H), 7.00–7.10 (m, 3H), 7.35 (dd, *J* = 7.2 Hz, 2.5 Hz, 1H), 7.54 (d, *J* = 8.8 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 103.9, 116.5 (2 C), 117.0 (d, *J* =

21.2 Hz), 119.3, 120.0 (d, $J = 1.5$ Hz), 123.2 (d, $J = 7.4$ Hz), 129.7 (d, $J = 3.5$ Hz), 129.9 (d, $J = 12.7$ Hz), 133.9 (2 C), 145.9, 152.7 (d, $J = 243.7$ Hz); HRMS (ESI) m/z calcd. for $C_{13}H_9ClFN_2$ ($[M+H]^+$): 247.0438, found: 247.0432.

4-[(6-Chloropyridin-2-yl)amino]benzonitrile (24). A solution of Pd catalyst in dioxane was prepared first. A flask was charged with Pd(OAc)₂ (0.1 mmol, 22 mg) and *rac*-BINAP (0.1 mmol, 62 mg), dry toluene (10 mL) was added and the resulting solution was stirred for 15 min under an argon atmosphere. Next, a 100 mL round bottomed flask was charged with 2,6-dichloropyridine (5 mmol, 0.74 g), 4-aminobenzonitrile (5 mmol, 0.71 g) and K₂CO₃ (0.1 mol, 13.8 g). The freshly prepared Pd catalyst solution was added, the resulting mixture was stirred for 2 min under an argon atmosphere, and then heated to reflux (oil bath temperature 120 °C) for 24 h. The mixture was allowed to cool down to room temperature and filtered through Celite. Celite cake was washed with dichloromethane (100 mL), combined organic phases were evaporated in vacuo and the residue was separated with an automated chromatography system using Silica Flash Cartridges applying a heptane-ethyl acetate gradient (from 100% heptane to 100% ethylacetate in 35 min, 35 mL/min). Obtained as white solid; yield 60% (0.28 g); mp 182-183 °C (AcOEt); ¹H NMR (400 MHz, DMSO-*d*₆): δ 6.87 (d, $J = 8.0$ Hz, 1H), 6.91 (d, $J = 8.0$ Hz, 1H), 7.65 (t, $J = 8.0$ Hz, 1H), 7.69 (d, $J = 8.9$ Hz, 2H), 7.77 (d, $J = 8.9$ Hz, 2H), 9.85 (brs, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 102.5, 110.9, 115.4, 118.1 (2 C), 120.0, 133.7 (2 C), 141.2, 145.5, 148.1, 155.1; HRMS (ESI) m/z calcd. for $C_{13}H_{10}ClN_2$ ($[M+H]^+$): 229.0533, found: 229.0533.

General Procedure C for the Synthesis of Compounds 4a-u. A solution of Pd catalyst in dioxane was prepared first. A flask was charged with Pd(OAc)₂ (0.025 mmol, 5.61 mg) and XPhos (0.03 mmol, 14 mg), dry dioxane (5 mL) was added and the resulting solution was stirred for 15 min under an argon atmosphere. Next, a 50 mL round bottomed flask was charged with the intermediate **7a-e** (0.5 mmol, 124 mg), an aniline **8a-c** or **12a-f** or a phenol **15a,b** (0.6 mmol) and Cs₂CO₃ (1.25 mmol, 0.41 g). The freshly prepared solution of Pd catalyst was added, the resulting mixture was stirred for 2 min under an argon atmosphere, and then heated to reflux (oil bath temperature 110 °C) for 16 h. The mixture was allowed to cool down to room temperature and filtered through Celite. Celite cake was washed with dichloromethane (100 mL), combined organic phases were evaporated in vacuo and the residue separated with an automated chromatography system using Silica Flash Cartridges applying a heptane-ethyl acetate gradient (from 100% heptane to 100% ethylacetate in 35 min, 35 mL/min).

4-[[6-(Mesitylamino)-3-fluoropyridin-2-yl]amino]benzonitrile (4a). Synthesized according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.6 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a** (0.5 mmol, 124 mg), aniline **8a** (0.6 mmol, 81 mg) and Cs₂CO₃ (1.25 mmol, 408 mg). Obtained as white solid; yield 53% (91 mg); mp 201-202 °C

(AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.18 (s, 6H), 2.33 (s, 3H), 5.59 (dd, *J* = 8.6 Hz, 2.0 Hz, 1H), 5.72 (s, 1H), 6.69 (br, 1H), 6.94 (s, 2H), 7.09 (dd, *J* = 10.2 Hz, 8.6 Hz, 1H), 7.47 (d, *J* = 8.7 Hz, 2H), 7.64 (d, *J* = 8.7 Hz, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 18.6, 21.0, 98.1 (br), 101.4, 118.2 (2 C), 120.2, 125.4 (d, *J* = 18.0 Hz), 129.0 (2 C), 132.8 (2 C), 135.1, 136.1, 136.4 (2 C), 140.0 (d, *J* = 240.0 Hz), 141.5 (d, *J* = 11.2 Hz), 146.1, 153.3; HRMS (ESI) *m/z* calcd. for C₂₁H₂₀FN₄ ([M+H]⁺): 347.1672, found: 347.1688. HUPLC: *t*_r 5.26 min. Single crystals for XRD analysis were grown by the slow evaporation of an acetone solution. CCDC Nr. 1410425.

4-[[6-(4-Cyanophenyl)amino-5-fluoropyridin-2-yl]amino]-3,5-dimethylbenzonitrile (4b). Synthesized according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.6 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a** (0.5 mmol, 124 mg), aniline **8b** (0.6 mmol, 88 mg) and Cs₂CO₃ (1.25 mmol, 408 mg). Obtained as white solid; yield 61% (109 mg); mp 235–236 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.27 (s, 6H), 5.70 (dd, *J* = 8.6 Hz, 2.2 Hz, 1H), 5.91 (s, 1H), 6.78 (brs, 1H), 7.19 (dd, *J* = 10.2 Hz, 8.6 Hz, 1H), 7.45 (s, 2H), 7.49 (d, *J* = 8.9 Hz, 2H), 7.62 (d, *J* = 8.9 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 18.5, 97.6 (d, *J* = 2.5 Hz), 104.1, 109.7, 117.9 (2 C), 118.8, 119.4, 124.3 (d, *J* = 18.0 Hz), 132.2 (2 C), 133.1 (2 C), 137.1 (2 C), 141.4 (d, *J* = 242.7 Hz), 141.8, 142.0 (d, *J* = 10.8 Hz), 143.9, 150.7 (d, *J* = 2.4 Hz); HRMS (ESI) *m/z* calcd. for C₂₁H₁₇FN₅ ([M+H]⁺): 358.1468, found: 358.1486. HUPLC: *t*_r 4.60 min. Single crystals for XRD analysis were grown by the slow evaporation from an acetone solution. CCDC Nr. 1410426.

(E)-4-((6-[4-(2-Cyanovinyl)-2,6-dimethylphenylamino]-3-fluoropyridin-2-yl)amino)benzonitrile (4c). Synthesized according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.6 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a** (0.5 mmol, 124 mg), aniline **12a** (0.6 mmol, 103 mg) and Cs₂CO₃ (1.25 mmol, 408 mg). Obtained as white solid; yield 62% (0.119 g); mp 211–212 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.28 (s, 6H), 5.67 (dd, *J* = 8.6 Hz, 2.2 Hz, 1H), 5.86 (brs, 1H), 5.88 (d, *J* = 16.6 Hz, 1H), 6.75 (brs, 1H), 7.18 (dd, *J* = 10.2 Hz, 8.6 Hz, 1H), 7.26 (s, 2H), 7.39 (d, *J* = 16.6 Hz, 1H), 7.51 (d, *J* = 8.9 Hz, 2H), 7.67 (d, *J* = 8.9 Hz, 2H); ¹³C NMR (100 MHz, CD₃COCD₃): δ 18.9, 96.5, 99.2 (d, *J* = 2.0 Hz), 103.6, 118.9 (2 C), 119.4, 120.1, 125.5 (d, *J* = 18.2 Hz), 128.5 (2 C), 132.5, 133.4 (2 C), 138.0 (2 C), 141.4 (d, *J* = 240.8 Hz), 142.5, 142.7 (d, *J* = 11.2 Hz), 146.3, 151.2, 153.3 (d, *J* = 2.0 Hz); HRMS (ESI) *m/z* calcd. for C₂₃H₁₉FN₅ ([M+H]⁺): 384.1624, found: 384.1629. HUPLC: *t*_r 4.76 min.

(E)-Methyl 3-(4-((6-[(4-cyanophenyl)amino]-5-fluoropyridin-2-ylamino)-3,5-dimethyl-phenyl)acrylate (4d). Synthesized according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.6 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a** (0.5 mmol, 124 mg), aniline **12f** (0.6 mmol, 123 mg) and Cs₂CO₃ (1.25 mmol, 408 mg). Obtained as white solid; yield 71% (0.15 g); mp 189–190 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.25 (s, 6H), 3.82 (s, 3H), 5.68 (dd, *J* = 8.6

1 Hz, 2.2 Hz, 1H), 5.86 (s, 1H), 6.43 (d, $J = 16.0$ Hz, 1H), 6.76 (d, $J = 3.2$ Hz, 1H), 7.15 (dd, $J = 10.2$ Hz, 8.6 Hz, 1H), 7.31 (s, 2H),
2 7.46 (d, $J = 8.8$ Hz, 2H), 7.64 (d, $J = 8.9$ Hz, 2H), 7.67 (d, $J = 16.0$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 18.6 (2C), 51.8, 97.1
3 (d, $J = 2.0$ Hz), 103.7, 117.5, 117.8 (2 C), 119.6, 124.3 (d, $J = 17.9$ Hz), 128.3 (2 C), 132.4, 133.1 (2 C), 136.7 (2 C), 139.2, 141.0 (d, $J =$
4 10.8 Hz), 141.8 (d, $J = 241.4$ Hz), 144.1, 144.4, 151.5 (d, $J = 2.0$ Hz), 167.5; HRMS (ESI) m/z calcd. for $\text{C}_{24}\text{H}_{22}\text{FN}_4\text{O}_2$ ($[\text{M}+\text{H}]^+$):
5 417.1727, found: 417.1732. HUPLC: t_r 5.04 min.

11 **3-(4-{6-[(4-Fluorophenyl)amino]-5-fluoropyridin-2-ylamino}-3,5-dimethylphenyl)acrylo-nitrile (4e).**

12 Synthesized according to the general procedure C using $\text{Pd}(\text{OAc})_2$ (0.025 mmol, 5.6 mg), XPhos (0.03 mmol, 14 mg),
13 intermediate **11c** (0.5 mmol, 120 mg), aniline **12a** (0.6 mmol, 103 mg) and Cs_2CO_3 (1.25 mmol, 408 mg). Obtained as a
14 mixture of geometrical isomers ($Z:E$ 1:6.3), white solid; yield 66% (0.123 g); mp 189–190 °C (AcOEt); ^1H NMR (400 MHz,
15 CDCl_3): signals of major (E)-isomer: δ 2.24 (s, 6H), 5.51 (dd, $J = 8.5$ Hz, 2.1 Hz, 1H), 5.78 (brs, 1H), 5.84 (d, $J = 16.6$ Hz, 1H),
16 6.43 (brs, 1H), 6.90 (t, $J = 8.8$ Hz, 2H), 7.08 (dd, $J = 10.4$ Hz, 8.5 Hz, 1H), 7.21 (s, 2H), 7.35 (d, $J = 16.6$ Hz, 1H), 7.47 (dd, $J =$
17 9.1 Hz, 4.7 Hz, 2H); signals of minor (Z)-isomer: δ 5.39 (d, $J = 12.1$ Hz, 1H), 7.57 (s, 2H), other signals are overlapped with
18 signals of major isomer; ^{13}C NMR (100 MHz, CDCl_3) of major (E)-isomer: δ 18.6 (2C), 95.2 (d, $J = 2.1$ Hz), 95.5, 115.3 (d, $J =$
19 22.3 Hz, 2C), 118.4, 120.2 (d, $J = 7.5$ Hz, 2C), 123.5 (d, $J = 17.5$ Hz), 127.6 (2C), 131.1, 136.1 (d, $J = 2.5$ Hz), 136.8 (2 C), 140.6, 140.9
20 (d, $J = 240.7$ Hz), 143.3 (d, $J = 11.1$ Hz), 150.2, 151.1 (d, $J = 2.2$ Hz), 158.1 (d, $J = 240.8$ Hz); HRMS (ESI) m/z calcd. for
21 $\text{C}_{22}\text{H}_{19}\text{F}_2\text{N}_4$ ($[\text{M}+\text{H}]^+$): 377.1578, found: 377.1585. HUPLC: t_r 4.94 min, 5.03 min.

33 **3-[4-(6-{4-[(2-Cyanovinyl)-2,6-dimethylphenyl]amino}-3-fluoropyridin-2-yl)aminophenyl]acrylonitrile (4f).**

34 Synthesized according to the general procedure C using $\text{Pd}(\text{OAc})_2$ (0.025 mmol, 5.6 mg), XPhos (0.03 mmol, 14 mg),
35 intermediate **11e** (0.5 mmol, 137 mg), aniline **12a** (0.6 mmol, 103 mg) and Cs_2CO_3 (1.25 mmol, 408 mg). Obtained as a
36 mixture of geometrical isomers ($Z,E:E,Z:E$ 1:1:14.7), yellow solid; yield 63% (0.129 g); mp 269–270 °C (AcOEt); ^1H NMR
37 (400 MHz, CD_3COCD_3): signals of major (E,E)-isomer: δ 2.26 (s, 6H), 5.97 (d, $J = 16.6$ Hz, 1H), 6.02 (dd, $J = 8.5$ Hz, 2.0 Hz,
38 1H); 6.30 (d, $J = 16.7$ Hz, 1H), 7.30 (dd, $J = 10.9$ Hz, 8.5 Hz, 1H), 7.33 (d, $J = 8.8$ Hz, 2H), 7.41 (d, $J = 16.6$ Hz, 1H), 7.46–7.52 (m,
39 3H), 7.59 (d, $J = 16.7$ Hz, 1H), 7.68 (d, $J = 8.8$ Hz, 2H), 8.16 (brs, 1H); signals of minor (Z,E)- and (E,Z)-isomers: δ 5.47 (d, $J =$
40 12.1 Hz, 1H), 5.73 (d, $J = 12.1$ Hz, 1H), 6.25 (d, $J = 16.6$ Hz, 1H), 7.15 (d, $J = 12.1$ Hz, 1H), other signals are overlapped with
41 signals of major isomer; ^{13}C NMR (100 MHz, CD_3COCD_3) of major (E,E)-isomer: δ 18.9, 93.3, 96.5, 101.0, 118.9, 119.0, 119.4,
42 119.7, 125.2 (d, $J = 18.2$ Hz), 127.3, 128.5, 129.1, 132.4, 138.0, 142.6, 145.0, 144.0 (d, $J = 240.6$ Hz), 151.0, 151.2, 153.3 (d, $J = 1.8$ Hz);
43 HRMS (ESI) m/z calcd. for $\text{C}_{25}\text{H}_{21}\text{FN}_5$ ($[\text{M}+\text{H}]^+$): 410.1781, found: 410.1793. HUPLC: t_r 4.87 min, 4.92 min.

1 4-{6-[(4-Fluorophenyl)amino]-5-fluoropyridin-2-ylamino}-3,5-dimethylbenzonitrile (**4g**). Synthesized according
2 to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.6 mg), XPhos (0.03 mmol, 14 mg), intermediate **11c** (0.5 mmol,
3 120 mg), aniline **8b** (0.6 mmol, 88 mg), and Cs₂CO₃ (1.25 mmol, 408 mg). Obtained as colorless solid; yield 50% (88 mg);
4 mp 135-136 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.23 (s, 6H), 5.58 (dd, *J* = 8.4 Hz, 2.1 Hz, 1H), 5.86 (brs, 1H), 6.47 (brs,
5 1H), 6.91 (t, *J* = 8.7 Hz, 2H), 7.11 (dd, *J* = 10.4 Hz, 8.4 Hz, 1H), 7.39-7.45 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ 18.5 (2 C),
6 95.6 (d, *J* = 2.3 Hz), 109.1, 115.3 (d, *J* = 22.3 Hz, 2 C), 119.1, 120.2 (d, *J* = 7.5 Hz, 2 C), 123.5 (d, *J* = 18.2 Hz), 132.1 (2 C), 135.9 (d, *J*
7 = 2.5 Hz), 136.9 (2 C), 141.1 (d, *J* = 241.6 Hz), 142.3, 143.3 (d, *J* = 11.1 Hz), 150.5 (d, *J* = 2.3 Hz), 158.1 (d, *J* = 241 Hz); HRMS (ESI)
8 *m/z* calcd. for C₂₀H₁₇F₂N₄ ([M+H]⁺): 351.1421, found: 351.1404. HUPLC: *t*_r 4.89 min.

9 4-{6-[4-(2-Cyanovinyl)phenylamino]-3-fluoropyridin-2-ylamino}benzonitrile (**4h**). Synthesized according to the
10 general procedure C using Pd(OAc)₂ (0.025 mmol, 5.6 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a** (0.5 mmol, 124
11 mg), aniline **12d** (0.6 mmol, 87 mg) and Cs₂CO₃ (1.25 mmol, 408 mg). Obtained as a mixture of geometrical isomers (*Z*:*E*
12 1:2.33), white solid; yield 68% (0.12 g); mp 285-286 °C (AcOEt); ¹H NMR (400 MHz, DMSO-*d*₆): signals of major (*E*)-isomer:
13 δ 6.22 (d, *J* = 16.6 Hz, 1H), 6.44-6.52 (m, 1H), 7.50-7.60 (m, 6H), 7.69 (d, *J* = 8.4 Hz, 2H), 7.84 (d, *J* = 8.4 Hz, 2H), 9.31 (s, 1H),
14 9.39 (s, 1H), signals of minor (*Z*)-isomer: δ 5.60 (d, *J* = 12.0 Hz, 1H), 7.28 (d, *J* = 12.0 Hz, 1H), 7.62 (d, *J* = 8.6 Hz, 2H), 7.75 (d,
15 *J* = 8.6 Hz, 2H), 9.45 (s, 1H), other signals are overlapped with the signals of major (*E*)-isomer; ¹³C NMR (100 MHz, DMSO-
16 *d*₆): signals of major (*E*)-isomer: δ 92.7, 102.4, 103.6 (br), 117.7 (2 C), 118.9 (2 C), 120.2, 125.7 (d, *J* = 18.1 Hz), 126.2, 129.3 (2 C),
17 133.3 (2 C), 141.1 (d, *J* = 11.7 Hz), 141.9 (d, *J* = 244 Hz), 144.7, 145.6, 149.6 (d, *J* = 2.2 Hz), 150.8, 1 signal is missing because of
18 overlap; signals of minor (*Z*)-isomer: δ 91.2, 102.5, 119.1 (2 C), 119.2, 120.1, 126.2, 130.4 (2 C), 133.2 (2 C), 144.5, 148.9, other
19 signals are overlapped with the signals of major isomer; HRMS (ESI) *m/z* calcd. for C₂₁H₁₅FN₅ ([M+H]⁺): 356.1311, found:
20 356.1308. HUPLC: *t*_r 4.60 min, 4.65 min.

21 4-[6-(4-Chlorophenyl)amino-5-fluoropyridin-2-ylamino]-3,5-dimethylbenzonitrile (**4i**). Synthesized according to
22 the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.1 mg), XPhos (0.03 mmol, 14 mg), intermediate **11b** (0.5 mmol, 129
23 mg), aniline **8b** (0.6 mmol, 88 mg), and Cs₂CO₃ (1.25 mmol, 408 mg). Obtained as colorless solid; yield 54% (99 mg); mp
24 197-198 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.25 (s, 6H), 5.60 (dd, *J* = 8.5 Hz, 2.1 Hz, 1H), 5.85 (brs, 1H), 6.52 (brs,
25 1H), 7.13 (dd, *J* = 10.4 Hz, 8.5 Hz, 1H), 7.17 (d, *J* = 8.8 Hz, 2H), 7.43 (s, 2H), 7.44 (d, *J* = 8.8 Hz, 2H); ¹³C NMR (100 MHz,
26 CDCl₃): δ 18.5 (2 C), 96.0 (d, *J* = 2.3 Hz), 109.3, 119.0, 119.7 (2 C), 123.6 (d, *J* = 18.2 Hz), 126.7, 128.7 (2 C), 132.1 (2 C), 136.9 (2
27 C), 138.5, 141.2 (d, *J* = 241.9 Hz), 142.2, 143.0 (d, *J* = 11.0 Hz), 150.6 (d, *J* = 2.3 Hz); HRMS (ESI) *m/z* calcd. for C₂₀H₁₇ClFN₄
28 ([M+H]⁺): 367.1126, found: 367.1142. HUPLC: *t*_r 5.16 min.

3-{4-[6-(4-Chlorophenyl)amino-5-fluoropyridin-2-ylamino]-3,5-dimethylphenyl}acrylonitrile (**4j**). Synthesized according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.6 mg), XPhos (0.03 mmol, 14 mg), intermediate **11b** (0.5 mmol, 129 mg), aniline **12a** (0.6 mmol, 103 mg) and Cs₂CO₃ (1.25 mmol, 408 mg). Obtained as a mixture of geometrical isomers (*Z*:*E* 1:13.5), brown solid; yield 65% (128 mg); mp 156-157 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): signals of major (*E*)-isomer: δ 2.23 (s, 6H), 5.56 (dd, *J* = 8.5 Hz, 2.1 Hz, 1H), 5.83 (d, *J* = 16.6 Hz, 1H), 5.84 (brs, 1H), 6.50 (brs, 1H), 7.09 (dd, *J* = 10.4 Hz, 8.5 Hz, 1H), 7.13 (d, *J* = 8.9 Hz, 2H), 7.21 (s, 2H), 7.34 (d, *J* = 16.6 Hz, 1H), 7.45 (d, *J* = 8.9 Hz, 2H); signals of minor (*Z*)-isomer: δ 5.42 (d, *J* = 12.1 Hz, 1H), 7.60 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 18.5 (2 C), 95.4, 95.6 (d, *J* = 2.2 Hz), 118.4, 119.6 (2 C), 123.6 (d, *J* = 17.8 Hz), 126.3, 128.2 (2 C), 129.0 (2 C), 131.1, 136.7 (2 C), 138.6, 140.4, 140.9 (d, *J* = 241.0 Hz), 142.8 (d, *J* = 11.0 Hz), 150.1, 151.1 (d, *J* = 2.2 Hz); HRMS (ESI) *m/z* calcd. for C₂₂H₁₉ClFN₄ ([M+H]⁺): 393.1282, found: 393.1296. HUPLC: *t*_r 5.24 min.

(*E*)-4-[6-[4-(2-Cyanovinyl)-2-fluoro-6-methylphenylamino]-3-fluoropyridin-2-ylamino]benzotrile (**4k**). Synthesized according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.6 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a** (0.5 mmol, 124 mg), aniline **12c** (0.6 mmol, 110 mg) and Cs₂CO₃ (1.25 mmol, 408 mg). Obtained as brown solid; yield 79% (150 mg); mp 235-236 °C (AcOEt); ¹H NMR (400 MHz, CD₃COCD₃): δ 2.29 (s, 3H), 6.26 (dd, *J* = 8.6 Hz, 2.2 Hz, 1H), 6.34 (d, *J* = 16.4 Hz, 1H), 7.33-7.38 (m, 1H), 7.37 (d, *J* = 8.8 Hz, 2H), 7.45-7.48 (m, 2H), 7.59 (d, *J* = 16.6 Hz, 1H), 7.68 (br s, 1H), 7.75 (d, *J* = 8.8 Hz, 2H), 8.32 (brs, 1H); ¹³C NMR (100 MHz, CD₃COCD₃): δ 17.5, 96.9, 99.5 (d, *J* = 2.0 Hz), 102.9, 111.8 (d, *J* = 22.5 Hz), 117.9, 118.1 (2 C), 119.1, 124.5 (d, *J* = 18.3 Hz), 125.8 (d, *J* = 2.7 Hz), 130.4 (d, *J* = 13.1 Hz), 131.8 (d, *J* = 8.6 Hz), 132.5 (2 C), 138.2 (d, *J* = 2.0 Hz), 141.1 (d, *J* = 242.1 Hz), 141.7 (d, *J* = 11.4 Hz), 145.2, 149.1 (d, *J* = 2.7 Hz), 151.5 (d, *J* = 2.1 Hz), 158.5 (d, *J* = 245.2 Hz); HRMS (ESI) *m/z* calcd. for C₂₂H₁₆F₂N₅ ([M+H]⁺): 388.1374, found: 388.1368. HUPLC: *t*_r 4.55 min.

4-[3-Fluoro-6-(*p*-tolylxy)pyridin-2-ylamino]benzotrile (**4l**). Synthesized according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a** (0.5 mmol, 124 mg), phenol **15a** (0.6 mmol, 65 mg) and Cs₂CO₃ (1.25 mmol, 0.41 g). Obtained as white solid; yield 89% (0.142 g); mp 153-154 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.46 (s, 3H), 6.41 (dd, *J* = 8.5 Hz, 2.0 Hz, 1H), 6.76 (brs, 1H), 7.06 (d, *J* = 8.4 Hz, 2H), 7.25 (d, *J* = 8.4 Hz, 2H), 7.36 (d, *J* = 8.6 Hz, 2H), 7.38 (m, 1H), 7.42 (d, *J* = 8.6 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 21.2, 101.0 (d, *J* = 2.9 Hz), 104.2, 118.0 (2 C), 119.7, 122.1 (2 C), 125.5 (d, *J* = 18.9 Hz), 130.3 (2 C), 133.2 (2 C), 134.8, 141.1 (d, *J* = 11.8 Hz), 142.7 (d, *J* = 244.8 Hz), 143.7, 152.3, 158.4 (d, *J* = 1.8 Hz); HRMS (ESI) *m/z* calcd. for C₁₉H₁₅FN₃O ([M+H]⁺): 320.1199, found: 320.1197. HUPLC: *t*_r 5.23 min.

1 **4-[3-Fluoro-6-(mesityloxy)pyridin-2-ylamino]benzotrile (4m)**. Synthesized according to the general procedure C
2
3 using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a** (0.5 mmol, 124 mg), phenol **15b** (0.6
4 mmol, 82 mg) and Cs₂CO₃ (1.25 mmol, 0.41 g). Obtained as yellow solid; yield 43% (0.074 g); mp 201-202 °C (AcOEt); ¹H
5 NMR (400 MHz, CDCl₃): δ 2.08 (s, 6H), 2.38 (s, 3H), 6.39 (dd, *J* = 8.4 Hz, 2.2 Hz, 1H), 6.70 (brs, 1H), 6.95 (s, 2H), 7.28 (s,
6 4H), 7.36 (dd, *J* = 10 Hz, 8.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 16.4 (2 C), 20.9, 99.6 (d, *J* = 2.8 Hz), 103.8, 117.6 (2 C),
7 119.6, 125.5 (d, *J* = 18.1 Hz), 129.2 (2 C), 131.0 (2 C), 132.9 (2 C), 134.6, 140.9 (d, *J* = 11.7 Hz), 142.1 (d, *J* = 244 Hz), 143.6, 148.9,
8 157.3 (d, *J* = 1.4 Hz); HRMS (ESI) *m/z* calcd. for C₂₁H₁₉FN₃O ([M+H]⁺): 348.1512, found: 348.1509. HUPLC: *t*_r 5.61 min.

9
10
11
12
13
14
15 **(E)-3-[4-[6-(4-Chlorophenyl)amino-5-fluoropyridin-2-ylamino]-3-methylphenyl]acrylonitrile (4n)**. Synthesized
16 according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **11b**
17 (0.5 mmol, 129 mg), aniline **12b** (0.6 mmol, 95 mg) and Cs₂CO₃ (1.25 mmol, 0.41 g). Obtained as yellow solid; yield 73%
18 (0.137 g); mp 189-190 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.30 (s, 3H), 5.74 (d, *J* = 16.6 Hz, 1H), 6.16 (brs, 1H), 6.24
19 (dd, *J* = 8.4 Hz, 2.2 Hz, 1H), 6.54 (brs, 1H), 7.19-7.29 (m, 5H), 7.32 (d, *J* = 16.6 Hz, 1H), 7.53 (d, *J* = 8.8 Hz, 2H), 7.67 (d, *J* = 8.4
20 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 18.0, 93.1, 99.4 (d, *J* = 2.4 Hz), 118.8, 118.9, 120.5 (2 C), 123.6 (d, *J* = 17.9 Hz), 126.3,
21 127.13, 127.17, 127.4, 128.8 (2 C), 129.9, 138.1, 141.7 (d, *J* = 243.3 Hz), 142.3, 143.0 (d, *J* = 18.1 Hz), 148.7 (d, *J* = 2.7 Hz), 150.1;
22 HRMS (ESI) *m/z* calcd. for C₂₁H₁₇³⁵ClFN₄ ([M+H]⁺): 379.1126, found: 379.1109. HUPLC: *t*_r 4.65 min.

23
24
25
26
27
28
29
30
31 **(E)-Methyl 3-[4-[6-(4-fluorophenyl)amino-5-fluoropyridin-2-ylamino]-3,5-dimethylphenyl]acrylate (4o)**.
32 Synthesized according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg),
33 intermediate **11c** (0.5 mmol, 120 mg), aniline **12f** (0.6 mmol, 123 mg) and Cs₂CO₃ (1.25 mmol, 0.41 g). Obtained as brown
34 solid; yield 55% (0.113 g); mp 162-163 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.24 (s, 6H), 3.82 (s, 3H), 5.51 (dd, *J* = 8.5
35 Hz, 2.1 Hz, 1H), 5.77 (s, 1H), 6.42 (d, *J* = 16.0 Hz, 1H), 6.43 (brs, 1H), 6.92 (t, *J* = 8.8 Hz, 2H), 7.07 (dd, *J* = 10.5 Hz, 8.5 Hz,
36 1H), 7.30 (s, 2H), 7.49 (dd, *J* = 9.1 Hz, 4.7 Hz, 2H), 7.66 (d, *J* = 16.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 18.6 (2 C), 51.7,
37 95.0 (d, *J* = 2.2 Hz), 115.3 (d, *J* = 22.3 Hz, 2 C), 117.2, 120.1 (d, *J* = 7.4 Hz, 2 C), 123.5 (d, *J* = 17.6 Hz), 128.3 (2 C), 132.1, 136.0
38 (br), 136.6 (2 C), 139.5, 140.8 (d, *J* = 240.2 Hz), 143.2 (d, *J* = 11.2 Hz), 144.5 (2 C), 151.3 (br), 158.0 (d, *J* = 240.6 Hz), 167.5;
39 HRMS (ESI) *m/z* calcd. for C₂₃H₂₂F₂N₃O₂ ([M+H]⁺): 410.1680, found: 410.1686. HUPLC: *t*_r 5.36 min.

40
41
42
43
44
45
46
47
48
49
50 **(E)-Methyl 3-[4-[6-(4-chlorophenyl)amino-5-fluoropyridin-2-ylamino]-3,5-dimethylphenyl]acrylate (4p)**.
51 Synthesized according to the general procedure C using Pd(OAc)₂ (0.018 mmol, 4.10 mg), XPhos (0.022 mmol, 10.46 mg),
52 intermediate **11b** (0.366 mmol, 94 mg), aniline **12f** (0.44 mmol, 90 mg) and Cs₂CO₃ (1.25 mmol, 0.41 g). Obtained as brown
53 solid; yield 71% (0.067 g); mp 162-163 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): δ 2.24 (s, 6H), 3.82 (s, 3H), 5.53 (dd, *J* = 8.6
54
55
56
57
58
59
60

Hz, 2.1 Hz, 1H), 5.77 (brs, 1H), 6.42 (d, $J = 16.0$ Hz, 1H), 6.48 (brs, 1H), 7.09 (dd, $J = 10.5$ Hz, 8.5 Hz, 1H), 7.17 (d, $J = 8.9$ Hz, 2H), 7.30 (s, 2H), 7.49 (d, $J = 8.9$ Hz, 2H), 7.66 (d, $J = 16.0$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 18.6, 51.7, 95.4 (d, $J = 2.1$ Hz), 117.3, 119.6 (2 C), 123.6 (d, $J = 17.9$ Hz), 126.4, 128.3 (2 C), 128.7 (2 C), 132.1, 136.6 (2 C), 138.6, 139.4, 140.9 (d, $J = 234.7$ Hz), 142.8 (d, $J = 11.0$ Hz), 144.5, 151.4 (d, $J = 2.0$ Hz), 167.5; HRMS (ESI) m/z calcd. for $\text{C}_{23}\text{H}_{22}^{35}\text{ClFN}_3\text{O}_2$ ($[\text{M}+\text{H}]^+$): 426.1385, found: 426.1383. HUPLC: t_r 5.52 min.

(E)-3-(4-{5-Fluoro-6-[4-(fluorophenyl)amino]pyridin-2-ylamino}-3-methylphenyl)acrylonitrile (4q). Synthesized according to the general procedure C using $\text{Pd}(\text{OAc})_2$ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **11c** (0.5 mmol, 120 mg), aniline **12b** (0.6 mmol, 95 mg) and Cs_2CO_3 (1.25 mmol, 0.41 g). Obtained as yellow solid; yield 75% (0.135 g); mp 150-151 °C (AcOEt); ^1H NMR (400 MHz, CDCl_3): δ 2.29 (s, 3H), 5.72 (d, $J = 16.6$ Hz, 1H), 6.16 (brs, 1H), 6.21 (dd, $J = 8.4$ Hz, 2.2 Hz, 1H), 6.48 (brs, 1H), 7.00 (t, $J = 8.7$ Hz, 2H), 7.19-7.28 (m, 3H), 7.31 (d, $J = 16.6$ Hz, 1H), 7.50-7.53 (dd, $J = 9.0$ Hz, 4.7 Hz, 2H), 7.69 (d, $J = 8.4$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 18.0, 93.1, 99.1 (d, $J = 2.3$ Hz), 115.4 (d, $J = 22.4$ Hz, 2 C), 118.7, 118.9, 121.4 (d, $J = 7.7$ Hz, 2 C), 123.5 (d, $J = 17.8$ Hz), 126.4, 127.1, 127.2, 129.9, 135.6 (d, $J = 2.6$ Hz), 141.6 (d, $J = 243.8$ Hz), 142.4, 143.5 (d, $J = 11.2$ Hz), 148.8 (d, $J = 2.7$ Hz), 150.2, 158.5 (d, $J = 241.6$ Hz); HRMS (ESI) m/z calcd. for $\text{C}_{21}\text{H}_{17}\text{F}_2\text{N}_4$ ($[\text{M}+\text{H}]^+$): 363.1421, found: 363.1424. HUPLC: t_r 5.02 min.

4-{6-[4-(2-Cyanovinyl)-2-methylphenylamino]-3-fluoropyridin-2-ylamino}benzotrile (4r). Synthesized according to the general procedure C using $\text{Pd}(\text{OAc})_2$ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a** (0.5 mmol, 124 mg), aniline **12b** (0.6 mmol, 0.95 mg) and Cs_2CO_3 (1.25 mmol, 0.41 g). Obtained as mixture of geometrical isomers ($Z:E$ 1:6.9), yellow solid; yield 70% (0.13 g); mp 239-240 °C (AcOEt); ^1H NMR (400 MHz, CD_3COCD_3): signals of major (*E*)-isomer: δ 2.26 (s, 3H), 6.29 (d, $J = 16.6$ Hz, 1H), 6.50 (dd, $J = 8.6$ Hz, $J = 2.2$ Hz, 1H), 7.46 (dd, $J = 8.4$ Hz, $J = 1.9$ Hz, 1H), 7.49-7.59 (m, 5H), 7.71 (d, $J = 8.4$ Hz, 1H), 7.84 (d, $J = 8.8$ Hz, 2H), 8.29 (s, 1H), 9.20 (s, 1H); signals of minor (*Z*)-isomer: δ 5.68 (d, $J = 12.0$ Hz, 1H), 7.31 (d, $J = 12.0$ Hz, 1H), 8.35 (s, 1H); ^{13}C NMR (100 MHz, CD_3COCD_3): signals of major (*E*)-isomer: δ 17.4, 93.5, 101.7 (d, $J = 2.5$ Hz), 103.3, 118.5 (2 C), 118.7, 119.1, 121.1, 124.5 (d, $J = 18.2$ Hz), 126.1, 128.1, 129.9, 132.7 (2 C), 141.5 (d, $J = 243.6$ Hz), 141.6 (d, $J = 11.3$ Hz), 142.6, 145.1, 148.2, 150.1, 150.3 (d, $J = 2.5$ Hz); signals of minor (*Z*)-isomer: 17.5, 91.9, 118.4, 120.9, 121.0, 127.4, 129.4, 131.5, other signals are overlapped with signals of major isomer. HRMS (ESI) m/z calcd. for $\text{C}_{22}\text{H}_{17}\text{FN}_5$ ($[\text{M}+\text{H}]^+$): 370.1468, found: 370.1455. HUPLC: t_r 4.69 min, 4.75 min.

4-[6-(4-Cyanophenyl)amino-5-fluoropyridin-2-ylamino]-3-fluoro-5-methylbenzotrile (4s). Synthesized according to the general procedure C using $\text{Pd}(\text{OAc})_2$ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a** (0.5 mmol, 124), aniline **8c** (0.6 mmol, 90 mg), and Cs_2CO_3 (1.25 mmol, 0.41 g). Obtained as brown solid; yield 42% (75

1 mg); mp 253-254 °C (AcOEt); ¹H NMR (400 MHz, CD₃COCD₃): δ 2.36 (s, 3H), 6.38 (dd, *J* = 8.6 Hz, 2.2 Hz, 1H), 7.43 (dd, *J* =
2 10.7, 8.6 Hz, 1H), 7.47 (d, *J* = 8.8 Hz, 2H), 7.63-7.67 (m, 2H), 7.76 (d, *J* = 8.8 Hz, 2H), 7.93 (s, 1H), 8.44 (s, 1H); ¹³C NMR (100
3 MHz, CD₃COCD₃): δ 17.4 (d, *J* = 2.6 Hz), 100.2 (d, *J* = 2.3 Hz), 103.1, 108.2 (d, *J* = 10.3 Hz), 117.1 (d, *J* = 25.1 Hz), 117.7 (d, *J* = 3.0
4 Hz), 118.1 (2 C), 119.1, 124.6 (d, *J* = 18.3 Hz), 130.1 (d, *J* = 3.2 Hz), 132.5, 133.2 (d, *J* = 12.3 Hz), 138.6 (d, *J* = 2.3 Hz), 141.5 (d, *J* =
5 242.3 Hz), 141.7 (d, *J* = 11.6 Hz), 145.0, 150.8 (d, *J* = 2.2 Hz), 157.5 (d, *J* = 247.3 Hz); HRMS (ESI) *m/z* calcd. for C₂₀H₁₄F₂N₅
6 ([M+H]⁺): 362.1217, found: 362.1233. HUPLC: *t*_r 4.46 min.

7
8
9
10
11
12
13 **4-{6-[4-(2-Cyanovinyl)-2,6-difluorophenylamino]-3-fluoropyridin-2-ylamino}benzotrile (4t)**. Synthesized
14 according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **11a**
15 (0.5 mmol, 124 mg), aniline **12e** (0.6 mmol, 108 mg) and Cs₂CO₃ (1.25 mmol, 0.41 g). Obtained as mixture of geometrical
16 isomers (*Z:E* 1:6.9), brown solid; yield 72% (140 mg); mp 223-224 °C (AcOEt); ¹H NMR (400 MHz, CD₃COCD₃), signals of
17 major (*E*)-isomer: δ 6.38-6.44 (m, 2H), 7.38-7.46 (m, 3H), 7.51 (d, *J* = 8.8 Hz, 2H), 7.62 (d, *J* = 16.7 Hz, 1H), 7.79 (d, *J* = 8.8
18 Hz, 2H), 7.96 (brs, 1H), 8.38 (brs, 1H); signals of minor (*Z*)-isomer δ 5.87 (d, *J* = 12.1 Hz, 1H), 7.68 (d, *J* = 9.0 Hz, 2H), 8.01
19 (brs, 1H), other signals are overlapped with signals of major isomer; ¹³C NMR (100 MHz, CD₃COCD₃) signals of major (*E*-
20 isomer: δ 99.0, 101.5 (d, *J* = 2.2 Hz), 104.0, 111.9 (dd, *J* = 18.5, *J* = 6.8 Hz), 118.7, 119.0 (2 C), 120.1, 121.9 (t, *J* = 16.3 Hz), 125.5 (d, *J* =
21 18.3 Hz), 132.0 (t, *J* = 9.7 Hz), 133.5 (2 C), 142.6 (d, *J* = 243.2 Hz), 142.7 (d, *J* = 11.6 Hz), 145.9, 149.0 (t, *J* = 2.8 Hz), 151.0 (d, *J*
22 = 2.2 Hz), 158.6 (dd, *J* = 247.2 Hz, *J* = 6.5 Hz). HRMS (ESI) *m/z* calcd. for C₂₁H₁₃F₃N₅ ([M+H]⁺): 392.1123, found: 392.1127.
23 HUPLC: *t*_r 4.35 min, 4.42 min.

24
25
26
27
28
29
30
31
32
33
34
35 **(E)-Methyl 4-{6-[4-(2-cyanovinyl)-2,6-dimethylphenylamino]-3-fluoropyridin-2-ylamino}benzoate (4u)**.
36 Synthesized according to the general procedure C using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg),
37 intermediate **11d** (0.5 mmol, 140 mg), aniline **12a** (0.6 mmol, 103 mg) and Cs₂CO₃ (1.25 mmol, 0.41 g). Obtained as white
38 solid; yield 72% (0.149 g); mp 233-234 °C (AcOEt); ¹H NMR (400 MHz, CD₃COCD₃): δ 2.23 (s, 6H), 3.81 (s, 3H), 6.05 (dd, *J* =
39 8.5 Hz, 2.0 Hz, 1H), 6.27 (d, *J* = 16.7 Hz, 1H), 7.27 (dd, *J* = 10.8 Hz, 8.5 Hz, 1H), 7.45 (brs, 1H), 7.47 (s, 2H), 7.56 (d, *J* = 16.7
40 Hz, 1H), 7.61 (d, *J* = 9.2 Hz, 2H), 7.65 (d, *J* = 9.2 Hz, 2H), 8.14 (brs, 1H); ¹³C NMR (100 MHz, CD₃COCD₃): δ 17.9 (2 C), 51.0,
41 95.6, 97.8 (d, *J* = 2.0 Hz), 117.2 (2 C), 118.4, 121.9, 124.3 (d, *J* = 18.2 Hz), 127.5 (2 C), 130.0 (2 C), 131.5, 137.1 (2 C), 140.6 (d, *J* =
42 240.9 Hz), 141.7, 142.1 (d, *J* = 10.8 Hz), 145.5, 150.2, 152.3 (d, *J* = 1.9 Hz), 166.1; HRMS (ESI) *m/z* calcd. for C₂₄H₂₂FN₄O₂
43 ([M+H]⁺): 417.1727, found: 417.1732. HUPLC: *t*_r 4.96 min.

44
45
46
47
48
49
50
51
52
53
54 **General Procedure D for the Synthesis of Compounds 18a,b**. Same as the General Procedure C but starting from
55 the intermediate **17** (0.5 mmol, 124 mg), and an aniline **8b** or **12a** (0.6 mmol).
56
57

1 4-[4-(4-Cyanophenyl)amino-5-fluoropyrimidin-2-ylamino]-3,5-dimethylbenzotrile (**18a**). Synthesized
2
3 according to the general procedure D using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **17**
4 (0.5 mmol, 124 mg), aniline **8b** (0.6 mmol, 80 mg) and Cs₂CO₃ (2.50 mmol, 0.82 g). Obtained as light yellow solid; yield
5 22% (0.039 g); mp 268-269 °C (AcOEt); ¹H NMR (400 MHz, CD₃COCD₃): δ 2.29 (s, 6H), 7.51 (d, *J* = 8.8 Hz, 2H), 7.58 (s,
6 2H), 7.82 (d, *J* = 8.8 Hz, 2H), 8.00 (m, 2H), 8.85 (s, 1H); ¹³C NMR (100 MHz, CD₃COCD₃): δ 17.7, 105.1, 109.6, 118.6, 118.7,
7 119.5, 119.6, 131.5, 132.6, 138.2, 140.8 (d, *J* = 246 Hz), 142.0 (d, *J* = 11.4 Hz), 143.7, 149.6 (d, *J* = 10.4 Hz), 157.1 (d, *J* = 2.9 Hz);
8
9 HRMS (ESI) *m/z* calcd. for C₂₀H₁₆FN₆ ([M+H]⁺): 359.1420, found: 359.1411. HUPLC: *t*_r 3.96 min.

10 4-{2-[4-(2-Cyanovinyl)-2,6-dimethylphenylamino]-5-fluoropyrimidin-4-ylamino}-benzotrile (**18b**). Synthesized
11 according to the general procedure D using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **17**
12 (0.5 mmol, 124 mg), aniline **12a** (0.6 mmol, 86 mg) and Cs₂CO₃ (2.50 mmol, 0.82 g). Obtained as a mixture of geometrical
13 isomers (*Z:E* 1:6.0), light yellow solid; yield 21% (0.040 g); mp 232-233 °C (AcOEt); ¹H NMR (400 MHz, CD₃COCD₃): signals
14 of major (*E*)-isomer: δ 2.25 (s, 3H), 6.27 (d, *J* = 16.7 Hz, 1H), 7.44-7.49 (m, 4H), 7.56 (d, *J* = 16.7 Hz, 1H), 7.81-7.87 (m, 3H),
15 7.99 (s, 1H), 8.80 (s, 1H); signals of minor (*Z*)-isomer: δ 5.72 (d, *J* = 12.1 Hz, 1H), 7.38 (d, *J* = 12.1 Hz, 1H), other signals are
16 overlapped with signals of major isomer; ¹³C NMR (100 MHz, CD₃COCD₃) of major (*E*)-isomer: δ 17.8, 95.9, 104.9, 118.3,
17 118.7, 119.5, 127.3, 128.6, 131.9, 132.5, 137.2, 140.3, 140.6 (d, *J* = 246 Hz), 142.0 (d, *J* = 10.4 Hz), 149.5 (d, *J* = 10.5 Hz), 150.2, 157.4
18 (d, *J* = 2.7 Hz); HRMS (ESI) *m/z* calcd. for C₂₂H₁₈FN₆ ([M+H]⁺): 385.1577, found: 385.1572. HUPLC: *t*_r 3.82 min, 3.98 min.

19 **General Procedure E for the Synthesis of Compounds 19a–c**. Same as the General Procedure C but starting from
20 the intermediate **21** (0.5 mmol, 123 mg) and an aniline **12a–c** (0.6 mmol).

21 4-{5-[4-(2-Cyanovinyl)-2,6-dimethylphenylamino]-2-fluorophenylamino}benzotrile (**19a**). Synthesized
22 according to the general procedure E using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **21**
23 (0.5 mmol, 123 mg), aniline **12a** (0.6 mmol, 103 mg) and Cs₂CO₃ (2.50 mmol, 0.82 g). Obtained as a mixture of geometrical
24 isomers (*Z:E* 1:4.2), yellow solid; yield 56% (0.11 g); mp 219-220 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): signals of major
25 (*E*)-isomer: δ 2.21 (s, 6H), 5.23 (s, 1H), 5.79 (d, *J* = 16.6 Hz, 1H), 5.95 (s, 1H), 6.15-6.19 (m, 1H), 6.54 (dd, *J* = 6.7 Hz, 2.5 Hz,
26 1H), 6.92-6.97 (m, 3H), 7.18 (s, 2H), 7.31 (d, *J* = 16.6 Hz, 1H), 7.48 (d, *J* = 8.5 Hz, 2H); signals of minor (*Z*)-isomer: δ 5.25 (s,
27 1H), 5.37 (d, *J* = 12.2 Hz, 1H), 7.01 (d, *J* = 12.2 Hz, 1H), other signals are overlapped with signals of major isomer; ¹³C NMR
28 (100 MHz, CD₃COCD₃) of major (*E*)-isomer: δ 18.7, 96.1, 101.9, 109.2, 110.9 (d, 7.0 Hz), 116.0, 117.5 (d, *J* = 21.3 Hz), 119.3, 120.3,
29 128.9, 129.6 (d, *J* = 12.9 Hz), 131.7, 134.2, 135.4, 142.9, 144.1, 148.6, 148.8 (d, *J* = 236.4 Hz), 151.0; HRMS (ESI) *m/z* calcd. for
30 C₂₄H₂₀FN₄ ([M+H]⁺): 383.1672, found: 383.1690. HUPLC: *t*_r 4.85 min.

1 4-{5-[4-(2-Cyanovinyl)-2-fluoro-6-methylphenylamino]-2-fluorophenylamino}benzo-nitrile (**19b**). Synthesized
2 according to the general procedure E using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **21**
3 (0.5 mmol, 123 mg), aniline **12c** (0.6 mmol, 106 mg) and Cs₂CO₃ (2.50 mmol, 0.82 g). Obtained as a mixture of geometrical
4 isomers (*Z*:*E* 1:3.13), light yellow solid; yield 59% (0.12 g); mp 173-174 °C (AcOEt); ¹H NMR (400 MHz, CDCl₃): signals of
5 major (*E*)-isomer: δ 2.20 (s, 3H), 5.49 (br, 1H), 5.77 (d, *J* = 16.6 Hz, 1H), 5.98 (br, 1H), 6.35-6.38 (m, 1H), 6.73 (dd, *J* = 6.8 Hz,
6 2.4 Hz, 1H), 6.97-7.02 (m, 3H), 7.05-7.10 (m, 2H), 7.27 (d, *J* = 16.6 Hz, 1H), 7.49 (d, *J* = 8.6 Hz, 2H); signals of minor (*Z*-
7 isomer: δ 5.40 (d, *J* = 12.1 Hz, 1H), 5.51 (br, 1H), other signals are overlapped with signals of major isomer; ¹³C NMR (100
8 MHz, CD₃COCD₃) of major (*E*)-isomer: δ 18.4 (*J* = 2.7 Hz), 97.0, 102.1, 110.8, 112.4 (*J* = 6.8 Hz), 113.3 (d, *J* = 22.0 Hz), 116.1,
9 117.2 (d, *J* = 21.0 Hz), 119.1, 120.3, 127.3 (d, *J* = 2.5 Hz), 129.5 (d, *J* = 13.1 Hz), 131.0 (d, *J* = 8.5 Hz), 132.2 (d, *J* = 12.1 Hz), 134.3,
10 136.2 (d, *J* = 2.4 Hz), 142.6 (d, *J* = 1.7 Hz), 149.4, 149.9 (d, *J* = 2.4 Hz), 150.5 (d, *J* = 236.7 Hz), 157.8 (d, *J* = 244.8 Hz); HRMS
11 (ESI) *m/z* calcd. for C₂₃H₁₇F₂N₄ ([M+H]⁺): 387.1421, found: 387.1414. HUPLC: *t*_r 4.69 min.

12 4-{5-[4-(2-Cyanovinyl)-2-methylphenylamino]-2-fluorophenylamino}benzonitrile (**19c**). Synthesized according to
13 the general procedure E using Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **21** (0.5 mmol, 123
14 mg), aniline **12b** (0.55 mmol, 87 mg) and Cs₂CO₃ (2.50 mmol, 0.82 g). Obtained as a mixture of geometrical isomers (*Z*:*E*
15 1:8.3), light yellow solid; yield 41% (0.076 g); mp 170-171 °C (AcOEt); ¹H NMR (400 MHz, CD₃COCD₃): signals of major (*E*-
16 isomer: δ 2.28 (s, 3H), 5.99 (d, *J* = 16.6 Hz, 1H), 6.88-6.94 (m, 1H), 7.08 (br, 1H), 7.12 (d, *J* = 8.6 Hz, 2H), 7.15-7.24 (m, 3H),
17 7.36-7.42 (m, 2H), 7.46 (br, 1H), 7.56 (d, *J* = 8.6 Hz, 2H), 7.85 (br, 1H); signals of minor (*Z*)-isomer: δ 5.41 (d, *J* = 12.1 Hz, 1H),
18 7.96 (br, 1H), other signals are overlapped with signals of major isomer; ¹³C NMR (100 MHz, CD₃COCD₃) of major (*E*-
19 isomer: δ 17.2, 92.0, 101.5, 114.5 (d, *J* = 1.6 Hz), 114.9, 115.5, 116.5 (d, *J* = 7.0 Hz), 116.5 (d, *J* = 21.1 Hz), 118.9, 119.2, 126.0, 126.2,
20 126.8, 129.2 (d, *J* = 13.0 Hz), 130.3, 133.4, 139.4 (d, *J* = 1.6 Hz), 145.7, 148.2, 150.1, 150.7 (d, *J* = 240.9 Hz); HRMS (ESI) *m/z*
21 calcd. for C₂₃H₁₈FN₄ ([M+H]⁺): 369.1516, found: 369.1505. HUPLC: *t*_r 4.80 min.

22 **General procedure F for the synthesis of Compounds 22a–d.** Same as the General Procedure C but starting from
23 the intermediate **24** (0.5 mmol, 115 mg) and an aniline **8a,b** or **12a,c** (0.6 mmol).

24 4-(6-Mesitylaminopyridin-2-ylamino)benzonitrile (**22a**). Synthesized according to the general procedure F using
25 Pd(OAc)₂ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **24** (0.5 mmol, 115 mg), aniline **8a** (0.6 mmol, 81
26 mg) and Cs₂CO₃ (1.25 mmol, 0.41 g). Obtained as white solid; yield 21% (35 mg); mp 216-217 °C (AcOEt); ¹H NMR (400
27 MHz, CDCl₃): δ 2.20 (s, 6H), 2.33 (s, 3H), 5.67 (d, *J* = 8.0 Hz, 1H), 5.92 (s, 1H), 6.20 (d, *J* = 7.8 Hz, 1H), 6.59 (s, 1H), 6.96 (s,
28 2H), 7.28 (t, *J* = 7.8 Hz, 1H), 7.44 (d, *J* = 8.9 Hz, 2H), 7.50 (d, *J* = 8.9 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 18.3, 21.0, 95.8,
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

99.2, 103.4, 117.8 (2 C), 119.7, 129.2 (2 C), 133.3 (2 C), 134.4, 136.6 (2 C), 136.7, 140.1, 144.9, 152.2, 156.8; HRMS (ESI) m/z calcd. for $C_{21}H_{21}N_4$ ($[M+H]^+$): 329.1766, found: 329.1765. HUPLC: t_r 5.06 min.

4-{6-[4-(Cyanophenyl)amino]pyridin-2-ylamino}-3,5-dimethylbenzonitrile (22b). Synthesized according to the general procedure F using $Pd(OAc)_2$ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **24** (0.5 mmol, 115 mg), aniline **8b** (0.6 mmol, 88 mg) and Cs_2CO_3 (1.25 mmol, 0.41 g). Brown solid; yield 20% (34 mg); mp 223-224 °C (AcOEt); 1H NMR (400 MHz, $CDCl_3$): δ 2.28 (s, 6H), 5.75 (d, $J = 8.1$ Hz, 1H), 5.93 (s, 1H), 6.28 (d, $J = 7.9$ Hz, 1H), 6.54 (s, 1H), 7.37 (t, $J = 8.0$ Hz, 1H), 7.42-7.46 (m, 4H), 7.50 (d, $J = 8.9$ Hz, 2H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 18.6, 99.1, 101.0, 103.5, 109.7, 117.6 (2C), 118.9, 119.6, 132.2 (2 C), 133.3 (2 C), 137.3 (2 C), 139.8, 141.9, 144.9, 153.2, 155.4; HRMS (ESI) m/z calcd. for $C_{21}H_{18}N_5$ ($[M+H]^+$): 340.1562, found: 340.1579. HUPLC: t_r 4.51 min.

4-{6-[4-(2-Cyanovinyl)-2,6-dimethylphenylamino]pyridin-2-ylamino}benzonitrile (22c). Synthesized according to the general procedure F using $Pd(OAc)_2$ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **24** (0.5 mmol, 115 mg), aniline **12a** (0.6 mmol, 0.10 g) and Cs_2CO_3 (1.25 mmol, 0.41 g). Obtained as a mixture of geometrical isomers ($Z:E$ 1:4.1), light yellow solid; yield 66% (0.12 g); mp 229-230 °C (AcOEt); 1H NMR (400 MHz, $CDCl_3$) of major (E)-isomer: δ 2.27 (s, 6H), 5.73 (d, $J = 8.0$ Hz, 1H), 5.86 (d, $J = 16.6$ Hz, 1H), 5.96 (s, 1H), 6.26 (d, $J = 7.6$ Hz, 1H), 6.61 (s, 1H), 7.24 (s, 2H), 7.32-7.38 (m, 2H), 7.42-7.49 (m, 4H); signals of minor (Z)-isomer: 5.78 (d, $J = 8.0$, 1H), 6.25 (d, $J = 7.8$, 1 H), 7.09 (d, $J = 12.1$, 1 H), other signals are overlapped with signals of the major (E)-isomer. ^{13}C NMR (100 MHz, $CDCl_3$) of major (E)-isomer: δ 18.6, 95.9, 98.9, 100.5, 103.4, 117.6 (2 C), 118.3, 119.6, 127.6 (2 C), 129.2, 131.6, 133.3 (2 C), 136.8, 137.1 (2 C), 139.7, 145.1, 153.2, 156.1; HRMS (ESI) m/z calcd. for $C_{23}H_{20}N_5$ ($[M+H]^+$): 366.1719, found: 366.1731. HUPLC: t_r 4.50 min, 4.61 min.

4-{6-[4-(2-cyanovinyl)-2-fluoro-6-methylphenylamino]pyridin-2-ylamino}benzonitrile (22d). Synthesized according to the general procedure F using $Pd(OAc)_2$ (0.025 mmol, 5.61 mg), XPhos (0.03 mmol, 14 mg), intermediate **24** (0.5 mmol, 115 mg), aniline **12c** (0.6 mmol, 0.106 g) and Cs_2CO_3 (1.25 mmol, 0.41 g). Obtained as a mixture of geometrical isomers ($Z:E$ 1:10.8), white solid; yield 71% (0.132 g); mp 261-262 °C (AcOEt); 1H NMR (400 MHz, CD_3COCD_3): signals of major (E)-isomer: δ 2.30 (s, 3H), 6.26 (d, $J = 7.9$ Hz, 2H), 6.34 (d, $J = 16.7$ Hz, 1H), 7.32 (d, $J = 8.8$ Hz, 2H), 7.41 (t, $J = 7.9$ Hz, 1H), 7.44-7.50 (m, 2H), 7.56-7.68 (m, 4H), 8.57 (s, 1H); signals of minor (Z)-isomer: δ 5.78 (d, $J = 12.1$ Hz, 1H), 7.84 (s, 1H), other signals are overlapped with signals of major isomer; ^{13}C NMR (100 MHz, CD_3COCD_3) of major (E)-isomer: δ 17.6 (d, $J = 2.7$ Hz), 96.9, 100.0, 101.1, 101.8, 111.8 (d, $J = 22.5$ Hz), 117.3, 118.0, 119.3, 125.8 (d, $J = 2.6$ Hz), 130.3 (d, $J = 13.3$ Hz), 132.0 (d, $J = 8.5$ Hz), 132.5, 138.5, 139.1, 146.1, 149.1 (d), 154.0, 156.0, 158.6 (d, $J = 245.4$ Hz); HRMS (ESI) m/z calcd. for $C_{22}H_{17}FN_5$ ($[M+H]^+$): 370.1468, found: 370.1481. HUPLC: t_r 4.33 min, 4.44 min.

Associated Content

SUPPORTING INFORMATION

Supporting Information Available: spectra of synthesized compounds; experimental data on the optimization of synthetic protocols; crystallographic data; figures illustrating docking studies. This material is available free of charge via the Internet at <http://pubs.acs.org>.

Author Information

Corresponding author

* Phone: +32 3 265 3205. Fax: +32 3 265 3233. E-mail: bert.maes@uantwerpen.be

Notes

The authors declare no competing financial interest.

Acknowledgements

This work was financially supported by the University of Antwerp (IOF POC project) and the Hercules Foundation. The authors thank Matthias Zeller (Youngstown State University, USA) for collecting the X-ray data sets. The X-ray diffractometer was funded by NSF Grant 0087210, Ohio Board of Regents Grant CAP-491, and by Youngstown State University.

Abbreviations used

HAART, highly active antiretroviral treatment; PI, protease inhibitor; INI, integrase inhibitor; NRTI, nucleoside reverse transcriptase inhibitor; NNRTI, non-nucleoside reverse transcriptase inhibitor; RT, reverse transcriptase; DATA, di(arylamino)triazine; DAPY, di(arylamino)pyrimidine; SDM, site-directed mutant; WST, water-soluble tetrazolium; TCID₅₀, tissue culture infective dose; CC, cytotoxic concentration; EC, effective concentration; DEAE, diethylaminoethyl.

References

1. 2013 UNAIDS report
http://www.unaids.org/sites/default/files/en/media/unaids/contentassets/documents/epidemiology/2013/gr2013/UNAIDS_Global_Report_2013_en.pdf.
2. Das, K.; Arnold, E. HIV-1 reverse transcriptase and antiviral drug resistance. Part 1. *Curr. Opin. Virol.* **2013**, *3*, 111-118.
3. Arts, E. J.; Hazuda, D. J. HIV-1 antiretroviral drug therapy. *Cold Spring Harbor Perspect. Med.* **2012**, *2*, a007161/1-a007161/23.
4. Das, K.; Arnold, E. HIV-1 reverse transcriptase and antiviral drug resistance. Part 2. *Curr. Opin. Virol.* **2013**, *3*, 119-128.

- 1 5. Das, K.; Lewi, P. J.; Hughes, S. H.; Arnold, E. Crystallography and the design of anti-AIDS drugs: conformational flexibility and
2 positional adaptability are important in the design of non-nucleoside HIV-1 reverse transcriptase inhibitors. *Prog. Biophys. Mol. Biol.*
3 **2005**, *88*, 209-231.
- 4
5
6 6. Das, K.; Bauman, J. D.; Clark, A. D.; Frenkel, Y. V.; Lewi, P. J.; Shatkin, A. J.; Hughes, S. H.; Arnold, E. High-resolution
7 structures of HIV-1 reverse transcriptase/TMC278 complexes: strategic flexibility explains potency against resistance mutations. *Proc.*
8 *Natl. Acad. Sci. U. S. A.* **2008**, *105*, 1466-1471.
- 9
10
11 7. Vingerhoets, J.; Azijn, H.; Fransen, E.; De Baere, I.; Smeulders, L.; Jochmans, D.; Andries, K.; Pauwels, R.; de Bethune, M.-P.
12 TMC125 displays a high genetic barrier to the development of resistance: Evidence from in vitro selection experiments. *J. Virol.* **2005**, *79*,
13 12773-12782.
- 14
15
16 8. Sluis-Cremer, N. The emerging profile of cross-resistance among the nonnucleoside HIV-1 reverse transcriptase inhibitor.
17 *Viruses* **2014**, *6*, 2960-2973.
- 18
19
20 9. Lewi, P.; Arnold, E.; Andries, K.; Bohets, H.; Borghys, H.; Clark, A.; Daeyaert, F.; Das, K.; de Bethune, M.-P.; de Jonge, M.;
21 Heeres, J.; Koymans, L.; Leempoels, J.; Peeters, J.; Timmerman, P.; Van den Broeck, W.; Vanhoutte, F.; van't Klooster, G.; Vinkers, M.;
22 Volovik, Y.; Janssen, P. A. J. Correlations between factors determining the pharmacokinetics and antiviral activity of HIV-1 non-
23 nucleoside reverse transcriptase inhibitors of the diaryltriazine and diarylpyrimidine classes of compounds. *Drugs in R&D* **2004**, *5*, 245-
24 257.
- 25
26
27 10. van Herrewege, Y.; Vanham, G.; Michiels, J.; Fransen, K.; Kestens, L.; Andries, K.; Janssen, P.; Lewi, P. A series of diaryltriazines
28 and diarylpyrimidines are highly potent nonnucleoside reverse transcriptase inhibitors with possible applications as microbicides.
29 *Antimicrob. Agents Chemother.* **2004**, *48*, 3684-3689.
- 30
31
32 11. Ludovici, D. W.; Kavash, R. W.; Kukla, M. J.; Ho, C. Y.; Ye, H.; De Corte, B. L.; Andries, K.; de Bethune, M. P.; Azijn, H.;
33 Pauwels, R.; Moereels, H. E. L.; Heeres, J.; Koymans, L. M. H.; de Jonge, M. R.; Van Aken, K. J. A.; Daeyaert, F. F. D.; Lewi, P. J.; Das, K.;
34 Arnold, E.; Janssen, P. A. J. Evolution of anti-HIV drug candidates. Part 2: diaryltriazine (DATA) analogues. *Bioorg. Med. Chem. Lett.*
35 **2001**, *11*, 2229-2234.
- 36
37
38 12. Ludovici, D. W.; De Corte, B. L.; Kukla, M. J.; Ye, H.; Ho, C. Y.; Lichtenstein, M. A.; Kavash, R. W.; Andries, K.; de Bethune, M. P.;
39 Azijn, H.; Pauwels, R.; Lewi, P. J.; Heeres, J.; Koymans, L. M. H.; de Jonge, M. R.; Van Aken, K. J. A.; Daeyaert, F. F. D.; Das, K.; Arnold, E.; Janssen,
40 P. A. J. Evolution of anti-HIV drug candidates. Part 3: diarylpyrimidine (DAPY) analogues. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 2235-2239.
- 41
42
43 13. Das, K.; Clark, A. D., Jr.; Lewi, P. J.; Heeres, J.; De Jonge, M. R.; Koymans, L. M. H.; Vinkers, H. M.; Daeyaert, F.; Ludovici, D.
44 W.; Kukla, M. J.; De Corte, B.; Kavash, R. W.; Ho, C. Y.; Ye, H.; Lichtenstein, M. A.; Andries, K.; Pauwels, R.; De Bethune, M.-P.; Boyer, P.
45 L.; Clark, P.; Hughes, S. H.; Janssen, P. A. J.; Arnold, E. Roles of conformational and positional adaptability in structure-based design of
46 TMC125-R165335 (Etravirine) and related non-nucleoside reverse transcriptase inhibitors that are highly potent and effective against
47 wild-type and drug-resistant HIV-1 variants. *J. Med. Chem.* **2004**, *47*, 2550-2560.
- 48
49
50
51
52
53
54
55
56
57
58
59
60

14. Janssen, P. A. J.; Lewi, P. J.; Arnold, E.; Daeyaert, F.; de Jonge, M.; Heeres, J.; Koymans, L.; Vinkers, M.; Guillemont, J.; Pasquier, E.; Kukla, M.; Ludovici, D.; Andries, K.; de Bethune, M.-P.; Pauwels, R.; Das, K.; Clark, A. D., Jr.; Frenkel, Y. V.; Hughes, S. H.; Medaer, B.; De Knaep, F.; Bohets, H.; De Clerck, F.; Lampo, A.; Williams, P.; Stoffels, P. In Search of a novel anti-HIV Drug: multidisciplinary coordination in the discovery of 4-[[4-[[4-(1e)-2-cyanoethenyl]-2,6-dimethylphenyl]amino]-2-pyrimidinyl]amino]benzotrile (R278474, Rilpivirine). *J. Med. Chem.* **2005**, *48*, 1901-1909.
15. Guillemont, J.; Pasquier, E.; Palandjian, P.; Vernier, D.; Gaurrand, S.; Lewi, P. J.; Heeres, J.; de Jonge, M. R.; Koymans, L. M. H.; Daeyaert, F. F. D.; Vinkers, M. H.; Arnold, E.; Das, K.; Pauwels, R.; Andries, K.; de Bethune, M.-P.; Bettens, E.; Hertogs, K.; Wigerinck, P.; Timmerman, P.; Janssen, P. A. J. Synthesis of novel diarylpyrimidine analogues and their antiviral activity against human immunodeficiency virus type 1. *J. Med. Chem.* **2005**, *48*, 2072-2079.
16. Heeres, J.; Lewi, P. J. The medicinal chemistry of the data and DAPY series of HIV-1 non-nucleoside reverse transcriptase inhibitors (NNRTIS). *Adv. Antiviral Drug Des.* **2007**, *5*, 213-242.
17. Azijn, H.; Tirry, I.; Vingerhoets, J.; de Bethune, M.-P.; Kraus, G.; Boven, K.; Jochmans, D.; Van Craenenbroeck, E.; Picchio, G.; Rimsky, L. T. TMC278, a next-generation nonnucleoside reverse transcriptase inhibitor (NNRTI), active against wild-type and NNRTI-resistant HIV-1. *Antimicrob. Agents Chemother.* **2010**, *54*, 718-727.
18. Devlin, B.; Nuttall, J.; Wilder, S.; Woodson, C.; Rosenberg, Z. Development of dapivirine vaginal ring for HIV prevention. *Antiviral Res.* **2013**, *100*, S3-S8.
19. Arien, K. K.; Jespers, V.; Vanham, G. HIV sexual transmission and microbicides. *Rev. Med. Virol.* **2011**, *21*, 110-133.
20. Paulini, R.; Müller, K.; Diederich, F. Orthogonal multipolar interactions in structural chemistry and biology. *Angew. Chem., Int. Ed.* **2005**, *44*, 1788-1805.
21. Davies, A.; Lamb, M.; Lyne, P.; Mohr, P.; Wang, B.; Wang, T.; Yu, D. Preparation of pyrazolylaminopyridines as kinase inhibitors. WO2006082392, 2006.
22. Jonckers, T. H. M.; Maes, B. U. W.; Lemièrre, G. L. F.; Dommissie, R. Selective palladium-catalyzed aminations on dichloropyridines. *Tetrahedron* **2001**, *57*, 7027-7034.
23. Hikawa, H.; Yokoyama, Y. Cross-coupling reaction on N-(3,5-dibromo-2-pyridyl)piperazines: regioselective synthesis of 3,5-disubstituted pyridylpiperazines. *Tetrahedron* **2010**, *66*, 9552-9559.
24. Fairlamb, I. J. S. Regioselective (site-selective) functionalisation of unsaturated halogenated nitrogen, oxygen and sulfur heterocycles by Pd-catalysed cross-couplings and direct arylation processes. *Chem. Soc. Rev.* **2007**, *36*, 1036-1045.
25. Loones, K. T. J.; Maes, B. U. W.; Meyers, C.; Deruytter, J. Orthogonal and auto-tandem catalysis: synthesis of dipyrido[1,2-a:2',3'-d]imidazole and its benzo and aza analogues via inter- and intramolecular C-N bond formation. *J. Org. Chem.* **2006**, *71*, 260-264.
26. Maes, B. U. W.; Loones, K. T. J.; Jonckers, T. H. M.; Lemièrre, G. L. F.; Dommissie, R. A.; Haemers, A. Selective palladium-catalyzed aminations on 2-chloro-3-iodo- and 2-chloro-5-iodopyridine. *Synlett* **2002**, 1995-1998.

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
27. Kuethe, J. T.; Wong, A.; Davies, I. W. Synthesis of disubstituted imidazo[4,5-b]pyridin-2-ones. *J. Org. Chem.* **2004**, *69*, 7752-7754.
28. Maes, B. U. W.; Verbeeck, S.; Verhelst, T.; Ekonomie, A.; von Wolff, N.; Lefevre, G.; Mitchell, E. A.; Jutand, A. Oxidative addition of haloheteroarenes to palladium(o): concerted versus S_NAr-type mechanism. *Chem. Eur. J.* **2015**, *21*, 7858-7865.
29. Komatsu, T.; Hirano, T.; Songkram, C.; Kawachi, E.; Kagechika, H. Novel thyroid hormone receptor antagonists with an N-alkylated diphenylamine skeleton. *Bioorg. Med. Chem.* **2007**, *15*, 3115-3126.
30. Arien, K. K.; Venkatraj, M.; Michiels, J.; Joossens, J.; Vereecken, K.; Van der Veken, P.; Heeres, J.; De Winter, H.; Heyndrickx, L.; Augustyns, K.; Vanham, G. Resistance and cross-resistance profile of the diaryltriazine non-nucleoside reverse transcriptase inhibitor and candidate microbicide UAMC01398. *J. Antimicrob. Chemother.*, in press, DOI: 10.1093/jac/dkv501.
31. Van Der Spoel, D.; Lindahl, E.; Hess, B.; Groenhof, G.; Mark, A. E.; Berendsen, H. J. C. GROMACS: Fast, flexible, and free. *J. Comput. Chem.* **2005**, *26*, 1701-1718.
32. Wu, Z.-Y.; Liu, N.; Qin, B.; Huang, L.; Yu, F.; Qian, K.; Morris-Natschke, S. L.; Jiang, S.; Chen, C. H.; Lee, K.-H.; Xie, L. Optimization of the antiviral potency and lipophilicity of halogenated 2,6-diarylpyridinamines as a novel class of HIV-1 NNRTIS. *ChemMedChem* **2014**, *9*, 1546-1555.
33. Sheldrick George, M. A short history of SHELX. *Acta Crystallogr A* **2008**, *64*, 112-122.
34. Huebschle, C. B.; Sheldrick, G. M.; Dittrich, B. ShelXle: a Qt graphical user interface for SHELXL. *J. Appl. Crystallogr.* **2011**, *44*, 1281-1284.
35. Farrugia, L. J. ORTEP-3 for windows - a version of ORTEP-III with a graphical user interface (GUI). *J. Appl. Crystallogr.* **1997**, *30*, 565.
36. Oostenbrink, C.; Villa, A.; Mark, A. E.; van Gunsteren, W. F. A biomolecular force field based on the free enthalpy of hydration and solvation: The GROMOS force-field parameter sets 53A5 and 53A6. *J. Comput. Chem.* **2004**, *25*, 1656-1676.
37. Koziara, K. B.; Stroet, M.; Malde, A. K.; Mark, A. E. Testing and validation of the Automated Topology Builder (ATB) version 2.0: prediction of hydration free enthalpies. *J. Comput.-Aided Mol. Des.* **2014**, *28*, 221-233.
38. Ditchfield, R.; Hehre, W. J.; Pople, J. A. Self-consistent molecular-orbital methods. IX. Extended Gaussian-type basis for molecular-orbital studies of organic molecules. *J. Chem. Phys.* **1971**, *54*, 724-728.
39. Wilkening, R. R.; Parker, D. L., Jr.; Wildonger, K. J.; Meng, D.; Ratcliffe, R. W. Preparation of 1,2,9,9a-tetrahydro-3H-fluoren-3-ones and related compounds as estrogen receptor modulators. WO2002041835, 2002.
40. Aguilar, N.; Fernandez, J. C.; Terricabras, E.; Carceller Gonzalez, E.; Salas Solana, J. Preparation of substituted tricyclic compounds with activity towards EP1 receptors. WO2013149997, 2013.
41. Schils, D.; Stappers, F.; Solberghe, G.; van Heck, R.; Coppens, M.; Van den Heuvel, D.; Van der Donck, P.; Callewaert, T.; Meeussen, F.; De Bie, E.; Eersels, K.; Schouteden, E. Ligandless Heck coupling between a halogenated aniline and acrylonitrile catalyzed

1 by Pd/C: development and optimization of an industrial-scale Heck process for the production of a pharmaceutical intermediate. *Org.*

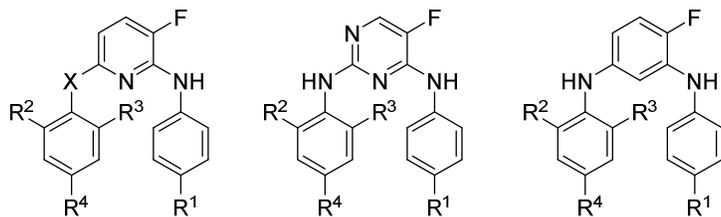
2
3 *Process Res. Dev.* **2008**, *12*, 530-536.

4 42. Janssen, P. A. J.; Guillemont, J. E. G.; Paugam, M.; Delest, B. F. M.; Heeres, J.; Lewi, P. J. Preparation of HIV inhibiting bicyclic
5 pyrimidine derivatives. WO 2006045828, 2006.
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Insert Table of Contents artwork here

Graphical entry for the Table of Contents



X = O, NH

R¹, R², R³, R⁴ = H, Me, F, Cl, CN, CH=CHCN, CH=CHCOOMe

4k (R¹ = CN, R² = F, R³ = H, R⁴ = CH=CHCN)

EC₅₀ = 1.0 [nM] (WT HIV)