Design, synthesis, and *in vitro* and *in vivo* anti-angiogenesis study of a novel vascular endothelial growth factor receptor-2 (VEGFR-2) inhibitor based on 1,2,3-triazole scaffold

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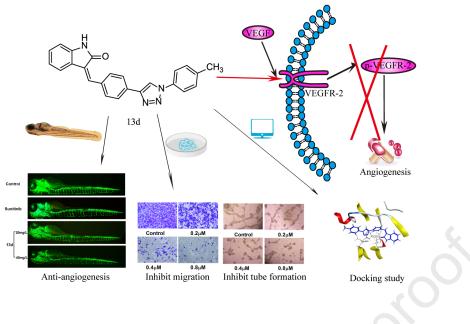
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4	scaffold
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3 Abstract

In the past five years, our team had been committed to click chemistry 4 biological activity of 1,2,3-triazole by research, exploring the 5 synthesizing different target inhibitors. In this study, a series of novel 6 1,2,3-triazole scaffolds were indole-2-one derivatives based on 7 synthesized for the first time, and their inhibitory activity on vascular 8 endothelial growth factor receptor-2 (VEGFR-2) was tested. Most of the 9 compounds had shown promising activity in the VEGFR-2 kinase assay 10 and had low toxicity to human umbilical vein endothelial cells 11 (HUVECs). The compound 13d (IC₅₀ = 26.38 nM) had better kinase 12 activity inhibition ability than sunitinib ($IC_{50} = 83.20$ nM) and was less 13 toxic to HUVECs. Moreover, it had an excellent inhibitory effect on 14 HT-29 and MKN-45 cells. On the one hand, by tube formation assay, 15 transwell, and western blot analysis, compound 13d could inhibit 16 VEGFR-2 protein phosphorylate on HUVECs, thereby inhibiting 17 HUVECs migration and tube formation. In vivo study, the zebrafish 18 model with VEGFR-2 labeling also verified that compound 13d had more 19 anti-angiogenesis ability than sunitinib. On the other hand, molecular 20 docking and molecular dynamics (MD) simulation results showed that 21 compound 13d could stably bind to the active site of VEGFR-2. Based on 22

Keywords: 1,2,3-Triazole; Anti-angiogenesis; VEGFR-2; Zebrafish. 3 **1. Introduction** 4 Angiogenesis is the formation of new blood vessels by sprouting or 5 splitting from pre-existing blood vessels [1-4]. It plays a critical role in 6 the pathogenesis of various disorders, most notably growth and metastasis 7 of solid tumors [5]. The growth and metastasis of tumors require new 8 blood vessels to transport nutrients and oxygen [6]. Vascular endothelial 9 growth factor (VEGF) is a highly specific vascular endothelial growth 10 factor that is overexpressed in most solid tumors [7]. Excessive VEGF in 11 solid tumors promotes the angiogenesis of adjacent blood vessels, and the 12 excessive production of blood vessels can cause the imbalance of the 13 metabolic microenvironment and accelerate tumor invasion and 14 metastasis [6,8,9]. Therefore, anti-angiogenesis is an essential form of 15 inhibiting tumor growth and metastasis [10-12]. 16

Vascular endothelial growth factor receptor-2 (VEGFR-2), as the vital receptor of VEGF, which activated by VEGF initiates downstream signal transduction, ultimately leads to angiogenesis, tumor proliferation, and migration [7,13]. Therefore, a long-held concept that inhibition of VEGFR-2 would cause efficient anti-angiogenesis and antitumor response [14,15]. At present, the small molecule VEGFR-2 inhibitors

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the above findings, compound 13d could be considered an effective

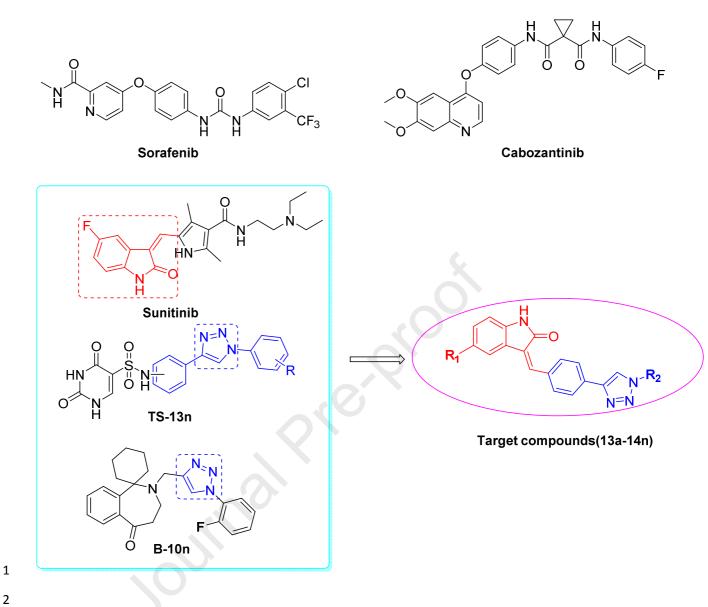
2 anti-angiogenesis drug and has more development value than sunitinib.

have emerged as promising anti-angiogenesis agents against a wide
variety of cancers, such as sorafenib, cabozantinib, and sunitinib, and
applied in clinical cancer therapy (Fig. 1) [16-18]. However,
chemotherapy drugs can cause specific adverse side effects and affect
patients' health [19]. Therefore, it is necessary to find low toxicity
VEGFR-2 inhibitors with a novel structure to treat cancer and enrich
VEGFR-2 inhibitors [7].

In the course of identifying various chemical fragments that could 8 serve as a scaffold for novel anti-angiogenesis agents, our programs 9 began with the indolin-2-one derivative sunitinib, a potent multitargeted 10 kinase inhibitor of VEGFR-2, PDGFR, and c-KIT kinases [20-22]. The 11 12 privileged indolin-2-one scaffold (red dashed rectangle in Fig. 1) was regarded as the most promising pharmacophore binding to the active site 13 of VEGFR-2 kinase [23,24]. Moreover, it should be pointed out that 14 numerous indolin-2-one derivatives have been reported as potent 15 antitumor agents [23,25-28]. Therefore, the indolin-2-one scaffold was 16 selected as a nucleus to develop a new VEGFR-2 inhibitor. The 17 combination principle of multiple pharmacophores for developing a 18 single molecule usually may enhance biological activity and improve 19 bioavailability [29]. In our previous work, inspired by various biological 20 properties of 1,2,3-triazole (blue dashed rectangle in **Fig. 1**), which could 21 bind with biomolecular targets to fulfill its envisaged role as potential 22

pharmacophore [30-32], two series of 1,2,3-triazole derivatives (TS-13n,
B-10n in Fig. 1) had been independently reported as potent antitumor
agents in our laboratory [33-35].
Based on the findings mentioned earlier, we have been inspired to
design and synthesize a series of novel derivatives derived by insertion of
a 1,4-diphenyl-1*H*-1,2,3-triazole moiety on the active nucleus of
indolin-2-one through an ethylene bridge (Fig. 1). Herein, twenty-eight

novel indolin-2-one derivatives bearing 1,2,3-triazole moiety and their VEGFR-2 kinase inhibitory activity were reported. These derivatives exhibited potent inhibition of VEGFR-2 tyrosine kinase activity and efficiently inhibited angiogenesis with less toxicity *in vitro* and *in vivo*, suggesting that these derivatives could be considered promising lead compounds for further development of novel anti-angiogenesis agents.



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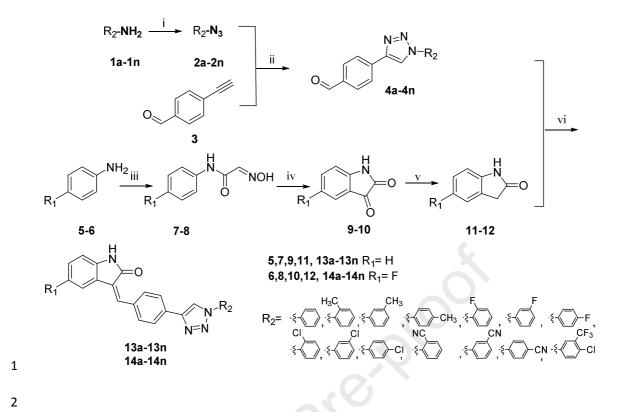
Fig. 1. Structures of sorafenib, cabozantinib, sunitinib, TS-13n, B-10n 3 and design of the target compounds. 4

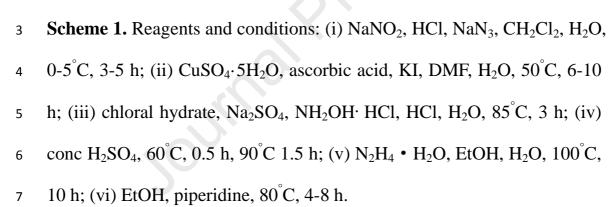
2. Results and discussion 5

2.1. Chemistry 6

adopted The synthetic pathway to prepare novel 7 indole-2-one-1,2,3-triazole derivatives (13a-14n) was depicted in 8 Scheme 1. First, the preparation of different substituted azide benzene 9 (2a-2n) involved diazo-reaction and displacement reaction with sodium 10

1	azide (NaN ₃) (synthetic detail procedure is given in the Experimental
2	section) [35]. Click chemistry is a practical approach for generating a
3	1,2,3-triazole unit [34]. The novel intermediates (4a-4n) were obtained in
4	high yields via Cu (I)-catalyzed azide-alkyne cycloaddition (CuAAC)
5	between aryl-azides $(2a-2n)$ and commercial 4-ethynylbenzaldehyde (3)
6	in the presence of sodium ascorbate and $\text{CuSO}_4{\cdot}5\text{H}_2\text{O}$ as a catalyst in
7	DMF and water mixture as the solvent system [35]. The compounds 9-10
8	were prepared from compounds 5-6 via Sandmeyer's method described in
9	the literature [37,38], reduced which using hydrazine hydrate to give
10	compound 11-12 [39]. Finally, the title compounds (13a-14n) were
11	accomplished by employing Claisen-Schmidt condensation reaction
12	between indolin-2-ones (11-12) and various 1,2,3-triazole aromatic
13	aldehydes (4a-4n) with a catalytic amount of piperidine as a base [40,
14	41].



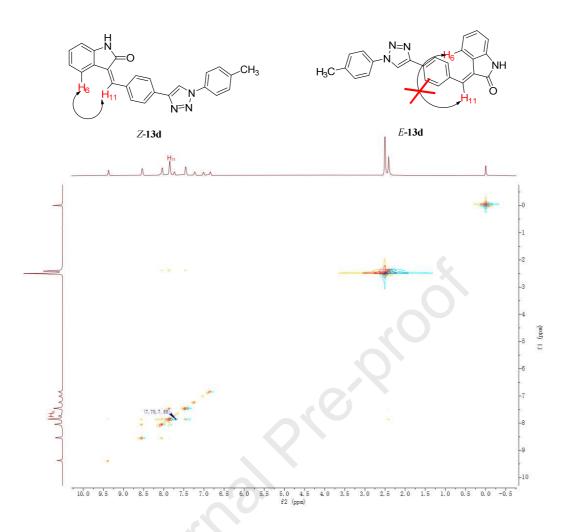


The known intermediates **9-12** and novel intermediates **4a-4n** were characterized by melting points and spectroscopic techniques (¹H NMR, ESI-MS), and the target compounds (**13a-14n**) also were characterized by melting points and spectroscopic techniques (¹H NMR, ¹³C NMR, HRMS, and FT-IR). The melting points of intermediates **9-12** approximately matched with melting points data in existing literatures [42,43].

14 Since the indole-2-one derivative has an exocyclic double bond, the

target compounds could exist in the form of *E* or *Z* isomers. As for compound **13m**, **14a**, **14b**, **14j**, **14k**, and **14m**, the ¹H NMR spectra revealed the existence of their inseparable E/Z mixtures, even the E/Zratios using the corresponding chemical shifts and integrals. However, because we could not separate them from the silica gel column chromatography or conventional method, the ¹³C NMR spectra of these mixtures were not shown.

Although several documents had reported that the analogs of title 8 similar obtained in synthetic compounds the method were 9 Z-configurations [44,45], confirmation of the stereochemistry of 10 indole-2-one-1,2,3-triazole derivatives still be requisite. Thus, the most 11 12 potency compound 13d, as the representative compound, was chosen to undergo the NOESY experiment. The NOE correlation (Fig. 2) between 13 H_6 (δ =7.73 ppm) and H_{11} (δ =7.85 ppm) suggested that the isomer 14 obtained correspond to Z-configuration (Z-13d), which should not exist in 15 *E*-configuration (*E*-13d). This result also illustrated that other compounds 16 (except 13m, 14a, 14b, 14j, 14k, and 14m) also were Z isomers. 17



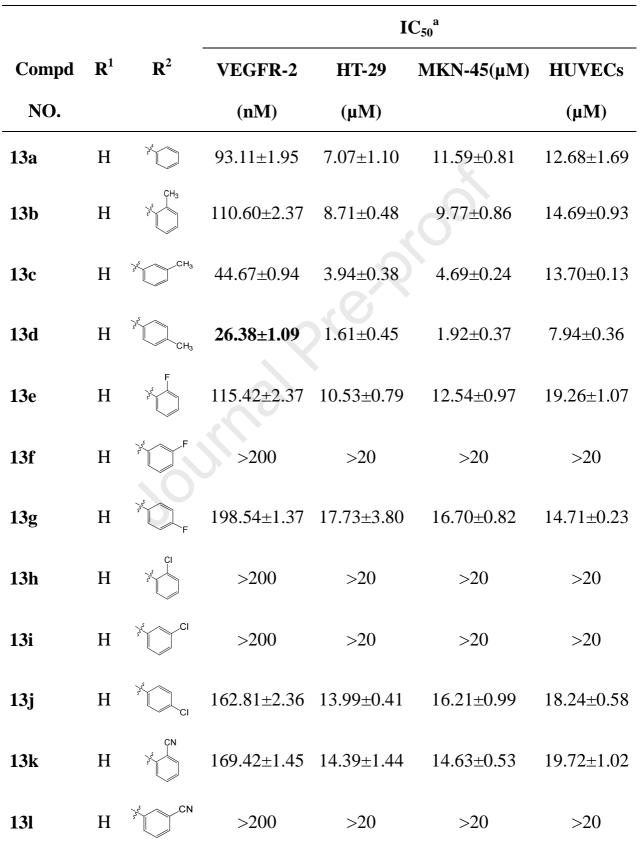
2 Fig. 2. NOESY of the representative compound 13d.

3 2.2. Biological evaluation

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4 2.2.1. VEGFR-2 inhibitory assay in vitro and structure-activity
5 relationship study (SAR)

To evaluate the inhibitory effects of sunitinib and all indole-2-one derivatives based on 1,2,3-triazole scaffolds (**13a-14n**) on VEGFR-2 kinase activity, VEGFR-2 inhibitory assay was carried out and the results were summarized in **Table 1**. Based on the VEGFR-2 kinase activity evaluation, the preliminary structure-activity relationship (SAR) in this work was summarized.



2 VEGFR-2, HT-29 cells, MKN-45 cells, and HUVECs.

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Table 1. The IC_{50} value of compounds (13a-14n) and sunitinib against

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13m	Н	3-5- CN	178.91±1.36	15.78±2.55	>20	>20
13n	Н	CI CF3	108.36±0.88	9.67±0.39	8.89±0.42	>20
14a	F	And the second s	99.67±0.73	8.12±1.29	11.13±0.17	13.70±0.13
14b	F	CH ₃	71.55±0.16	5.87±0.32	7.21±1.90	8.63±0.27
14c	F	°st ⊂ CH3	89.16±0.63	7.32±0.64	6.81±1.49	9.88±0.76
14d	F	St CH3	36.90±0.54	3.98±0.61	34.62±0.34	13.83±0.42
14e	F	F	139.50±1.65	8.98±0.42	8.51±0.54	14.36±1.99
14f	F	5 ² F	171.12±1.09	12.97±0.72	8.19±1.30	17.25±1.22
14g	F	F	192.51±0.47	>20	>20	>20
14h	F	Sec. Cl	119.93±1.36	5.95±0.27	7.54±0.55	18.24±0.58
14i	F	3 ^c Cl	186.44±1.02	10.46±0.43	14.08±0.88	19.42±0.59
14j	F	3 ² CI	170.18±2.19	11.24±0.86	15.81±0.52	19.58±2.14
14k	F	CN 3 ² 3 ²	187.24±2.88	13.98±0.43	15.21±0.73	>20
141	F	S CN	193.41±0.97	>20	>20	>20
14m	F	5-5-5 CN	190.07±1.98	>20	>20	>20

14n F
$$(CF_3)^{Cl}$$
 168.89±2.34 16.52±0.77 >20 >20

sunitinib - - 83.20±1.36 10.34±0.96 9.25±0.77 6.37±0.59

a: IC₅₀ values are presented as mean values of at least three independent 1 determinations. 2 We found that compared to the positive drug (sunitinib $IC_{50} = 83.20$ 3 nM), most compounds exhibited excellent inhibitory activity against 4 VEGFR-2. The semi-inhibitory concentration IC_{50} value of seven 5 compounds (13a, 13c, 13d, 14a-14d) was less than 100 nM, which all 6 showed excellent inhibition to VEGFR- 2. Furthermore, 13d (IC₅₀ = 7 26.38 nM) and 14d (IC₅₀ = 36.9 nM) exhibited the best activities among 8 9 them.

Moreover, we analyzed the effect of two substitutions of the 10 indolin-2-one, 13a-13n ($R_1 = H$) and 14a-14n ($R_1 = F$) possessed similar 11 IC_{50} values for each comparison. It showed that the substitution effect of 12 the indole-2-one part might not have too obvious value on the 13 structure-activity relationship. Then, concerning the activity of 14 derivatives with an unsubstituted or substituted phenyl group attached to 15 triazole scaffold, incorporation of unsubstituted phenyl group or phenyl 16 substituted by electron-donating groups led to compounds 13a-13d and 17 14a-14d with the better inhibitory action on VEGFR-2 relative to the 18 electron-withdrawing groups substituted analogs (13e-13n and 14e-14n). 19 Furthermore, the activity of tolyl substituted and unsubstituted derivatives 20

 $4-CH_3>3-CH_3>2-CH_3>H$. Besides, there was no significant 1 was difference in the inhibitory activity between electron-withdrawing groups. 2 The results showed that the phenyl side groups substituted with 3 electron-donor groups might significantly influence indole 1,2,3-triazole 4 scaffolds' increased potency. In contrast, electron-withdrawing groups 5 might have a common effect or decrease the inhibitory activities of 6 scaffolds. 7

8 2.2.2. HT-29 cell lines, MKN-45 cell lines, HUVECs proliferation assay
9 in vitro

CCK-8 assay was used to explore the effects of target compounds on 10 the cell proliferation of human colon cancer (HT-29) cell lines, human 11 12 gastric cancer (MKN-45) cell lines, and human umbilical vein endothelial cells (HUVECs). The positive control sunitinib was also conducted to 13 assess the inhibitory abilities of cell proliferation of all compounds, 14 shown as IC_{50} values, and summarized in **Table 1**. Including the most 15 potency compound 13d verified by the VEGFR-2 kinase inhibition assay, 16 the target compounds' overall toxicity to HUVECs was lower than that of 17 sunitinib. Furthermore, compound **13d** had a good effect on inhibiting cell 18 viability for HT-29 and MKN-45 cells. Interestingly, combined with the 19 VEGFR-2 kinase inhibition assay results, the compound **13d** could inhibit 20 the kinase activity of VEGFR-2 more effectively than sunitinib in vitro 21 and simultaneously had less toxic to HUVECs. 22

1 2.2.3. Anti-angiogenesis effect in vitro

Based on the above experimental results, compound 13d had the most 2 potential to become a VEGFR-2 inhibitor. Therefore, the inhibitory effect 3 of compound 13d on angiogenesis was further explored. Transwell was 4 used to explore the inhibitory ability of compound 13d on HUVEC cell 5 migration, and the results were shown in Fig. 3A. As the compound 6 concentration increased, the migration ability of HUVECs decreased, 7 signified that the changes have occurred in a concentration-dependent 8 manner. Since compound 13d was less toxic to HUVECs at low 9 concentrations, the facts excluded the migratory inhibition of compound 10 13d caused by toxic effects. At the same time, HUVECs were treated 11 with the same concentration of compound 13d to explore its effect on 12 HUVECs' ability to form tubules, and the results were shown in Fig. 3B. 13 Similarly, the tube formative length decreased with the concentration of 14 13d increased. 15

These data indicated that compound **13d** could inhibit HUVECs migration and tube formation *in vitro*, which was an essential step in inhibiting angiogenesis.

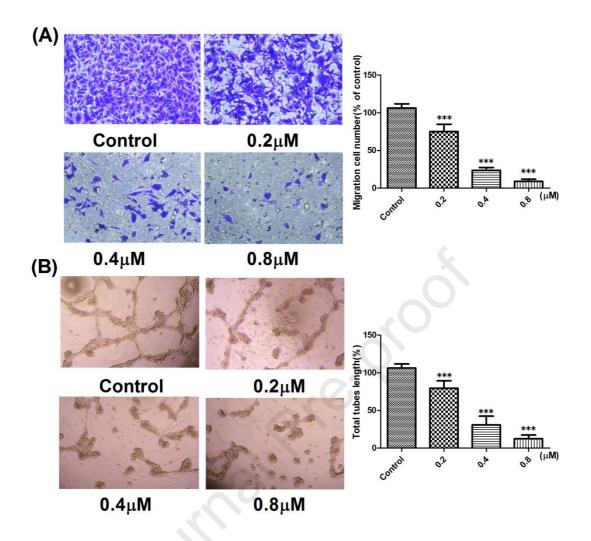


Fig. 3. Compound 13d inhibited HUVECs migration and the tube
formation. (A) Transwell analyzed the inhibitory effects of compound
13d on HUVECs migration. (B) Effects of compound 13d on the tube
formation of HUVECs *in vitro*.

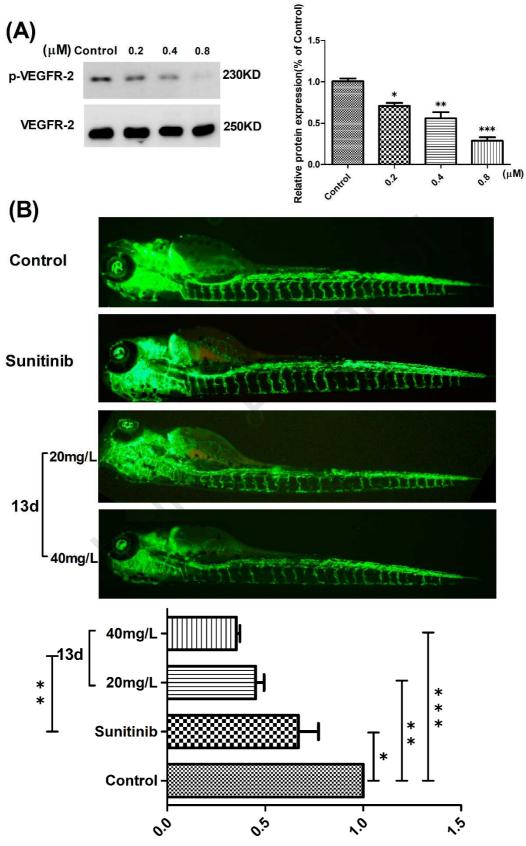
6 2.2.4. Western blot analysis and anti-angiogenesis research in vivo.

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Furthermore, Western blot was carried out to analyze VEGFR-2 protein's phosphorylation on HUVECs cell membranes at different compound concentrations. The results were shown in **Figure 4A**. As the concentration of compound **13d** increased, the phosphorylation of VEGFR-2 in HUVECs decreased. That is, the activation of VEGFR-2

decreased. Simultaneously, to verify the effect of compound 13d on 1 inhibiting angiogenesis in vivo, four groups of blood vessel-specific 2 fluorescent transgenic zebrafish, in a total of 40, were used for 3 anti-angiogenesis research. Zebrafish is currently the ideal vascular 4 biology research and anti-tumor angiogenesis drug evaluation model, 5 which can intuitively observe angiogenesis. The results were shown in 6 Fig. 4B. At a 40 mg/L concentration, compound 13d had a better 7 inhibitory effect on zebrafish internode vascular (ISV) angiogenesis than 8 sunitinib. Also, compound 13d even effectively inhibited zebrafish ISV 9 regeneration at a lower concentration (20 mg/L). 10

In general, compound 13d could effectively inhibit the phosphorylation
of VEGFR-2 with better anti-angiogenesis ability than sunitinib *in vivo*.
These results were significantly different.



Ratio of intact zebrafish ISV roots (% of Control)

Fig. 4. The compound 13d could inhibit the phosphorylation of VEGFR-2
and the angiogenesis of zebrafish microvessels. (A) Western blot analysis
of the phosphorylation changes of VEGFR-2 with increasing compound
13d concentration. (B) The effects of compound 13d and sunitinib on the
neovascularization of zebrafish *in vivo*.

6 2.3. Molecular modeling

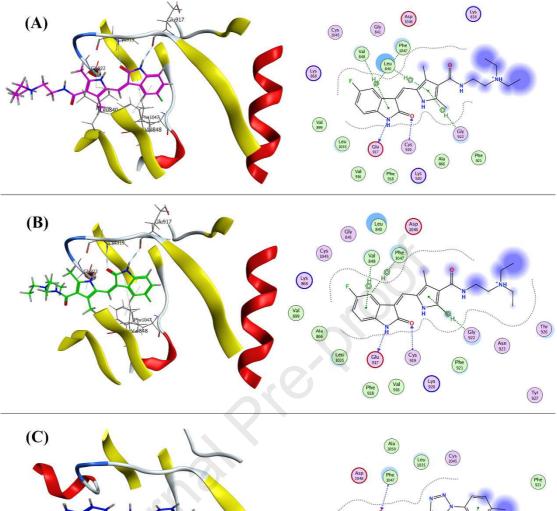
7 2.3.1. Molecular docking study

Molecular docking simulations were performed using MOE (Molecular 8 Operating Environment, version 2016.08, Chemical Computing Group 9 Inc., Canada) software to study the possible binding pattern of target 10 compounds in the active site of VEGFR-2. Moreover, the crystal structure 11 of VEGFR-2 in complex with sunitinib (PDB ID: 4AGD) was adopted in 12 the docking calculations. The best docking conformation of the potent 13 compound 13d based on the compound activity against VEGFR-2 kinase 14 was selected as the most probable binding conformation. The positive 15 drug sunitinib, as the co-crystallized ligand, was re-docked using the 16 same procedure as 13d. 17

The re-docking result demonstrated that sunitinib (**Fig. 5B**) could stack well with the original ligand (**Fig. 5A**), two H-bonds predicted by the simulation of sunitinib were exactly the same to those existed in the crystal structure, and the Arene-H conjugates were roughly the same. These data strongly certified the feasibility of the simulation and

rationalized the predictive interactions of the compound 13d with
 VEGFR-2 (Fig. 5C).

The docking conformation of compound 13d in the binding site 3 showed that carbonyl on the indole fragment formed a strong H-bond 4 with Phe1047, the lateral phenyl and the phenyl group linked to the 5 indole formed Arene-H conjugates with Val848 and Leu840, respectively. 6 By contrast, either H-bonds or the Arene-H interactions of 13d were less 7 than sunitinib. Simultaneously, as a whole molecule, 13d spatially 8 extended more in-depth into the pocket of protein; this provided a 9 reasonable explanation that compound **13d** had better kinase activity 10 inhibition ability than sunitinib, which was confirmed by the VEGFR-2 11 12 kinase assay.



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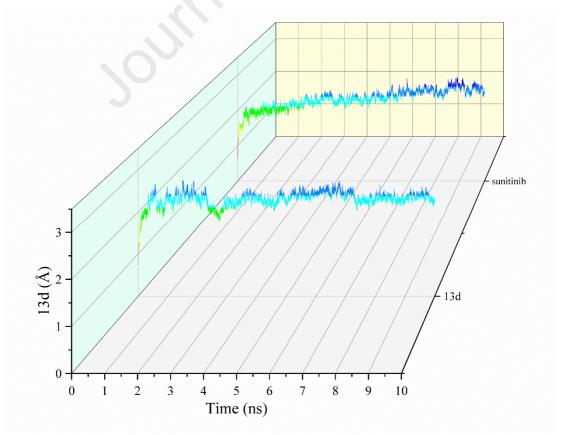
Fig. 5. The 2D diagram and 3D representation of original ligand (A),
sunitinib (B) and compound 13d (C) showing their interactions with the
VEGFR-2 active site (PDB code: 4AGD).

5 2.3.2. Molecular dynamics (MD) simulation

Based on the compound activity against VEGFR-2 kinase, the docking
conformation of the potent compound 13d obtained in MOE (Molecular

Operating Environment) was selected to conduct molecular dynamics (MD) simulation in-depth the binding pose. MD simulation was then carried out in explicit aqueous solution for 10 ns. For comparison, MD simulation of positive control sunitinib in complex with VEGFR-2 was also performed.

The stability of two systems under simulation was evaluated by the root-mean-square deviation (RMSD) of the backbone atoms related to the starting structures (**Fig. 6**). As can be seen in the plots, all systems were stable during the 10-ns MD simulation; the values of RMSD of compound **13d** remained between 2 and 2.5 Å after reaching the summit, which demonstrated that the hit compound **13d** was stabilized in the active site.



- 1 Fig. 6. RMSD tendency of two systems (13d and sunitinib) versus time in
- 2 the 10 ns MD simulation.
- 3 2.4. Toxicity prediction
- 4 **Table 2**. The pharmacokinetic studies of **13d** and sunitinib.

Compound	Pharmacokinetic studies			
NO.	Water solubility ^b	Plasma protei binding ^c	Acute Oral Toxicity ^d	Tetrahymena pyriformis ^e
13d	-3.051	0.831	1.773	1.299
Sunitinib	-3.217	0.914	2.683	1.653
$\mathbf{h} \cdot \mathbf{h} \mathbf{a} \mathbf{c}$	· 1000/ d	····		

5 ^b unit: logS; ^c unit: 100%; ^d unit: kg/mol; ^e unit: pIGC₅₀ (ug/L).

As a comprehensive and open-source tool, admetSAR was used to predict **13d** and sunitinib's toxicity, and the results were shown in **Table 2** [46]. Since the acute oral toxicity units and Tetrahymena pyriformis are kg/mol and pIGC₅₀ (ug/L), respectively (the lower value represents the lower toxicity), compound **13d** had low toxicity than sunitinib in both toxicity prediction assays, which inconsistent with the result of HUVECs proliferation assay.

13 **3. Conclusion**

The newly synthesized indolin-2-one derivatives based 14 on 1,2,3-triazole scaffolds, evaluated as VEGFR-2 inhibitors, showed 15 promising activity in the VEGFR-2 kinase inhibition assay. Among them, 16 compound 13d showed better activity inhibition on VEGFR-2 and 17 lowered toxicity to HUVECs, and had an excellent inhibitory effect on 18 HT-29 and MKN-45 cells than sunitinib. Transwell and tube formation 19 experiments showed that compound 13d inhibited HUVECs migration 20

and tube formation ability in a concentration-dependent manner.
Furthermore, western blot analysis manifested compound 13d could
decrease the phosphorylation of VEGFR-2 in HUVECs. At the same time,
by using VEGFR-2 specific fluorescent transgenic zebrafish, the
inhibitory effect of compound 13d on angiogenesis was verified *in vivo*.
Besides, the rationality and scientificity of molecular design were verified
by docking research and molecular dynamics simulation.

In conclusion, we designed and synthesized compound **13d** as a novel VEGFR-2 inhibitor that is more effective, less toxic than sunitinib. Compound **13d** provided a better choice for drug research and the development of new VEGFR-2 inhibitors.

12 **4. Experimental**

13 *4.1. Chemistry*

14 4.1.1. General methods

Unless otherwise stated, all chemical reagents and solvents were 15 purchased from commercial sources and can be used without further 16 purification. Melting points were measured on a capillary electrothermal 17 melting apparatus without calibration. By point thin layer 18 chromatography (TLC), using GF254 silica gel from Qingdao Ocean 19 Chemical Company (Qingdao, China), the reaction progress was 20 monitored with a fluorescent indicator on a 254nm glass plate and 21 visualized by ultraviolet light. Column chromatography was performed 22

using silica gel (200-300 mesh) from Qingdao Ocean Chemical Company (Qingdao, China), recorded ¹H NMR and ¹³C NMR spectra on a Bruker AMX500 (¹H at 500 MHz, ¹³C at 126 MHz) magnetic resonance spectrometer at ambient temperature. All NMR spectra were recorded using DMSO- d_6 as a solvent, and chemical shifts were reported in ppm (parts per million) relative to tetramethylsilane (TMS) as an internal standard.

8 4.1.2. The synthesis of substituted azido benzenes (2a-2n)

Compound 2a was synthesized by using the following procedure: To a 9 stirred solution of aniline (1a) (3.00 g, 32.21 mmol) in dichloromethane 10 (120 mL) were added 30 mL of aqueous NaNO₂ (2.67 g, 38.66 mmol) 11 solution and 37% concentrated hydrochloric acid (HCl) (3 mL) 12 successively. After 0.5 h of stirring at 0°C, a solution of sodium azide 13 (NaN₃) (2.93 g, 45.10 mmol) dissolved in water (20 mL) was added 14 dropwise, and the reaction mixture was kept stirring at 0-5°C for 3-5 h. 15 The progress of the reaction was monitored through the TLC test. After 16 the reaction was completed, the organic phase was separated and washed 17 with brine, dried over anhydrous Na₂SO₄, filtered, and the solvent was 18 removed under reduced pressure. The residue was purified by column 19 chromatography on silica gel using petroleum ether to afford the title 20 compound as pale yellow oil liquid. 21

22 Compounds **2b-2n** were prepared using the identical synthetic

1 procedure as 2a.

2 4.1.3. The synthesis of substituted azido benzenes (4a-4n)

Compound 4a was synthesized by using the following procedure: 3 Reaction of commercially 4-ethynylbenzaldehyde (3) (1.00 g, 7.68 mmol) 4 with azidobenzene (2a) (1.01 g, 8.45 mmol) in 15 mL of DMF proceeded 5 at 50°C for 5 h in the presence of 4 mL of aqueous copper sulfate 6 pentahydrate (CuSO₄•5H₂O) (0.15 g, 0.60 mmol), ascorbic acid (0.15 g, 7 0.85 mmol) and a catalytic amount of KI. The progress of the reaction 8 was monitored through the TLC test. After the reaction was completed, 9 the reaction was added 100 mL of water and extracted with ethyl acetate 10 (3×100 mL). The combined extracts were washed with brine, dried over 11 anhydrous Na₂SO₄, filtered, and the solvent was removed under reduced 12 pressure. The crude residue was purified by column chromatography on 13 silica gel using ethyl acetate/petroleum ether to afford the title compound 14 as pale yellow solid. 15

Compounds **4b-4n** were prepared using the identical synthetic procedure as **4a**.

18 *4.1.3.1. 4-(1-phenyl-1H-1,2,3-triazol-4-yl)benzaldehyde* (*4a*)

Light yellow solid, yield: 49.52%, mp: 187.3°C. ¹H NMR (500 MHz, DMSO- d_6) δ 10.04 (s, 1H, CHO), 9.50 (s, 1H, H-triazole), 8.18 (d, J = 7.6Hz, 2H, Ar-H), 8.05 (d, J = 7.5 Hz, 2H, Ar-H), 7.97 (d, J = 7.8 Hz, 2H, Ar-H), 7.65 (t, J = 7.4 Hz, 2H, Ar-H), 7.54 (t, J = 7.3 Hz, 1H, Ar-H).

- 1 ESI-MS $[M+H]^+$ m/z: 250.13.
- 2 4.1.3.2. 4-[1-(o-tolyl)-1H-1,2,3-triazol-4-yl]benzaldehyde (**4b**)
- Light yellow solid, yield: 45.15%, mp: 119.4°C. ¹H NMR (500 MHz,
- 4 DMSO- d_6) δ 10.04 (s, 1H, CHO), 9.16 (d, J = 1.5 Hz, 1H, H-triazole),
- 5 8.18 (s, 2H, Ar-H), 8.04 (d, J = 3.1 Hz, 2H, Ar-H), 7.52 (s, 3H, Ar-H),
- 6 7.46 (s, 1H, Ar-H), 2.23 (s, 3H, CH₃). ESI-MS $[M+H]^+$ m/z: 264.15.
- 7 4.1.3.3. 4-[1-(m-tolyl)-1H-1,2,3-triazol-4-yl]benzaldehyde (**4***c*)
- 8 Light brown solid, yield: 44.12%, mp: 174.4°C. ¹H NMR (500 MHz,
- 9 DMSO- d_6) δ 10.04 (s, 1H, CHO), 9.48 (s, 1H, H-triazole), 8.17 (d, J = 7.7
- 10 Hz, 2H, Ar-H), 8.04 (d, J = 7.7 Hz, 2H, Ar-H), 7.80 (s, 1H, Ar-H), 7.75 (d,
- 11 J = 7.5 Hz, 1H, Ar-H), 7.52 (t, J = 7.7 Hz, 1H, Ar-H), 7.35 (d, J = 7.0 Hz,
- 12 1H, Ar-H), 2.44 (s, 3H, CH₃). ESI-MS [M+H]⁺ m/z: 264.15.
- 13 4.1.3.4. 4-[1-(p-tolyl)-1H-1,2,3-triazol-4-yl]benzaldehyde (**4**d)
- Light brown solid, yield: 46.14%, mp: 189.4-193.3°C. ¹H NMR (500 MHz, DMSO- d_6) δ 10.04 (s, 1H, CHO), 9.46 (d, J = 1.8 Hz, 1H, H-triazole), 8.17 (d, J = 6.4 Hz, 2H, Ar-H), 8.04 (d, J = 6.4 Hz, 2H, Ar-H), 7.84 (d, J = 6.4 Hz, 2H, Ar-H), 7.45 (d, J = 6.6 Hz, 2H, Ar-H), 2.40 (s, 3H,
- 18 CH₃). ESI-MS $[M+H]^+$ m/z: 264.16.
- 19 *4.1.3.5.* 4-[1-(2-fluorophenyl)-1H-1,2,3-triazol-4-yl]benzaldehyde (**4***e*)
- Light yellow solid, yield: 46.26%, mp: 126.8°C. ¹H NMR (500 MHz,
- 21 DMSO- d_6) δ 10.05 (s, 1H, CHO), 9.30 (s, 1H, H-triazole), 8.20 (d, J = 7.1
- 22 Hz, 2H, Ar-H), 8.04 (d, J = 7.2 Hz, 2H, Ar-H), 7.93 (t, J = 7.6 Hz, 1H,

Ar-H), 7.64 (d, J = 10.4 Hz, 2H, Ar-H), 7.49 (t, J = 6.8 Hz, 1H, Ar-H).

2	ESI-MS $[M+H]^+$ m/z: 268.10.
3	4.1.3.6. 4-[1-(3-fluorophenyl)-1H-1,2,3-triazol-4-yl]benzaldehyde (4f)
4	Light brown solid, yield: 44.25%, mp: 179.9°C. ¹ H NMR (500 MHz,
5	DMSO- d_6) δ 10.03 (s, 1H, CHO), 9.53 (s, 1H, H-triazole), 8.15 (d, $J = 7.8$
6	Hz, 2H, Ar-H), 8.04 (d, <i>J</i> = 7.8 Hz, 2H, Ar-H), 7.90 – 7.83 (m, 2H, Ar-H),
7	7.72 - 7.66 (m, 1H, Ar-H), 7.39 (t, $J = 8.4$ Hz, 1H, Ar-H). ESI-MS
8	$[M+H]^+ m/z: 268.09.$
9	4.1.3.7. 4-[1-(4-fluorophenyl)-1H-1,2,3-triazol-4-yl]benzaldehyde (4g)
10	Yellow solid, yield: 46.54%, mp: 203.5°C. ¹ H NMR (500 MHz,
11	DMSO- d_6) δ 10.04 (s, 1H, CHO), 9.47 (s, 1H, H-triazole), 8.16 (d, $J = 6.8$
12	Hz, 2H, Ar-H), 8.07 – 7.94 (m, 4H, Ar-H), 7.51 (dd, <i>J</i> = 9.1, 4.8 Hz, 2H,
13	Ar-H). ESI-MS [M+H] ⁺ m/z: 268.10.
14	4.1.3.8. 4-[1-(2-chlorophenyl)-1H-1,2,3-triazol-4-yl]benzaldehyde (4h)
15	Light yellow solid, yield: 42.05%, mp: 121.3°C. ¹ H NMR (500 MHz,
16	DMSO- d_6) δ 10.04 (s, 1H, CHO), 9.26 (s, 1H, H-triazole), 8.19 (d, $J = 7.8$
17	Hz, 2H, Ar-H), 8.04 (d, J = 7.8 Hz, 2H, Ar-H), 7.82 (t, J = 8.0 Hz, 2H,
18	Ar-H), 7.68 (t, J = 7.6 Hz, 1H, Ar-H), 7.63 (t, J = 7.5 Hz, 1H, Ar-H).
19	ESI-MS $[M+H]^+$ m/z: 284.03.
20	4.1.3.9. 4-[1-(3-chlorophenyl)-1H-1,2,3-triazol-4-yl]benzaldehyde (4i)
21	Light yellow solid, yield: 46.15%, mp: 191.4°C. ¹ H NMR (500 MHz,
22	DMSO- d_6) δ 10.04 (s, 1H, CHO), 9.57 (s, 1H, H-triazole), 8.15 (d, $J = 6.1$
	28

1	Hz, 2H, Ar-H), 8.09 (s, 1H, Ar-H), 8.04 (d, <i>J</i> = 4.7 Hz, 2H, Ar-H), 7.98 (d,
2	<i>J</i> = 6.3 Hz, 1H, Ar-H), 7.68 (t, <i>J</i> = 6.2 Hz, 1H, Ar-H), 7.61 (d, <i>J</i> = 6.1 Hz,
3	1H, Ar-H). ESI-MS [M+H] ⁺ m/z: 284.13.
4	4.1.3.10. 4-[1-(4-chlorophenyl)-1H-1,2,3-triazol-4-yl]benzaldehyde (4j)
5	Light brown solid, yield: 38.95%, mp: 202.5°C. ¹ H NMR (500 MHz,
6	DMSO- d_6) δ 10.04 (s, 1H, CHO), 9.53 (s, 1H, H-triazole), 8.16 (d, $J = 8.1$
7	Hz, 2H, Ar-H), 8.04 (d, J = 8.2 Hz, 2H, Ar-H), 8.00 (d, J = 8.8 Hz, 2H,
8	Ar-H), 7.73 (d, <i>J</i> = 8.8 Hz, 2H, Ar-H). ESI-MS [M+H] ⁺ m/z: 284.11.
9	4.1.3.11. 2-[4-(4-formylphenyl)-1H-1,2,3-triazol-1-yl]benzonitrile (4k)
10	Light brown solid, yield: 49.42%, mp: 217.5°C. ¹ H NMR (500 MHz,
11	DMSO- d_6) δ 10.06 (s, 1H, CHO), 9.44 (s, 1H, H-triazole), 8.20 (d, $J = 7.7$
12	Hz, 3H, Ar-H), 8.07 (d, J = 7.8 Hz, 2H, Ar-H), 8.01 (dt, J = 16.1, 7.9 Hz,
13	2H, Ar-H), 7.82 (t, <i>J</i> = 7.5 Hz, 1H, Ar-H). ESI-MS [M+H] ⁺ m/z: 275.12.
14	4.1.3.12. 3-[4-(4-formylphenyl)-1H-1,2,3-triazol-1-yl]benzonitrile (4l)
15	Light brown solid; yield: 43.19%, mp: 102.3°C. ¹ H NMR (500 MHz,
16	DMSO- d_6) δ 10.04 (s, 1H, CHO), 9.60 (s, 1H, H-triazole), 8.48 (s, 1H,
17	Ar-H), 8.34 (d, J = 7.4 Hz, 1H, Ar-H), 8.14 (d, J = 7.4 Hz, 2H, Ar-H),
18	8.03 (dd, $J = 21.2$, 6.6 Hz, 3H, Ar-H), 7.86 (t, $J = 7.8$ Hz, 1H, Ar-H).
19	ESI-MS $[M+H]^+$ m/z: 275.13.
20	4.1.3.13. 4-[4-(4-formylphenyl)-1H-1,2,3-triazol-1-yl]benzonitrile (4m)
21	Light brown solid, yield: 44.05%, mp: 113.5°C. ¹ H NMR (500 MHz,

22 DMSO- d_6) δ 10.04 (s, 1H, CHO), 9.64 (s, 1H, H-triazole), 8.19 (d, J = 8.8

- 1 Hz, 2H, Ar-H), 8.15 (d, J = 6.7 Hz, 4H, Ar-H), 8.04 (d, J = 8.1 Hz, 2H,
- 2 Ar-H). ESI-MS $[M+H]^+$ m/z: 275.10.
- з *4.1.3.14*.
- 4 4-{1-[4-chloro-3-(trifluoromethyl)phenyl]-1H-1,2,3-triazol-4-yl}benzalde
- 5 *hyde* (**4***n*)
- 6 Light yellow solid, yield: 45.64%, mp: 209.4°C. ¹H NMR (500 MHz,
- 7 DMSO- d_6) δ 10.03 (s, 1H, CHO), 9.66 (s, 1H, H-triazole), 8.40 (s, 1H,
- 8 Ar-H), 8.30 (d, J = 8.7 Hz, 1H, Ar-H), 8.13 (d, J = 7.9 Hz, 2H, Ar-H),
- 9 8.03 (t, J = 8.6 Hz, 3H, Ar-H). ESI-MS $[M+H]^+$ m/z: 352.10.
- 4.1.4. The synthesis of substituted 2-(hydroxyimino)- N- phenylacetamide
 (7-8)

Compound 7 was synthesized by using the following procedure: To a 12 stirred solution of anhydrous sodium sulfate (40.00 g, 281.62 mmol) in 13 water (150 mL), chloral hydrate (10.00 g, 60.46 mmol) was added. Then 14 the commercially aniline (5) (3.00 g, 32.21 mmol), 37% concentrated 15 hydrochloric acid (3.6 mL), and hydroxylamine hydrochloride (8.30 g, 16 119.45 mmol) were sequentially added. The mixture was stirred at 80°C 17 for 3 h. After the reaction was completed, the reaction system was cooled 18 to room temperature, filtered and dried to obtain a crude product as a 19 brown solid, which was uesd in the next step without purification. 20

Compound 8 were prepared using the identical synthetic procedure as7.

1 4.1.5. The synthesis of substituted indoline-2,3-dione (**9-10**)

Compound 9 was synthesized by using the following procedure: A 2 round bottom flask was charged with compound 7 (2.00 g, 12.18 mmol) 3 and concentrated sulfuric acid H₂SO₄ (30 mL). After 0.5 h of stirring at 4 60° C, the temperature of the reaction mixture was raised to 90° C for 1.5 h. 5 After the reaction was completed, the reaction system was cooled to room 6 temperature and poured into ice water (80 mL). The solid precipitate was 7 filtered with suction, washed with water (4×50 mL) and dried. The crude 8 residue was purified by column chromatography on silica gel using ethyl 9 acetate/petroleum ether to obtain an orange solid (mp: 194°C). 10

11 Compound **10** (mp: 225°C) were prepared using the identical synthetic 12 procedure as **9**.

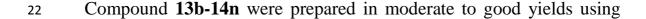
13 *4.1.6. The synthesis of substituted indolin-2-one* (*11-12*)

Compound 11 was synthesized by using the following procedure: To a 14 solution of compound 9 (2.00 g, 14.00 mmol) in ethanol (50 mL) was 15 added 80% hydrazine hydrate (12.78 g, 203.90 mmol) and water (30 mL). 16 The reaction mixture was stirred at 100°C for 10 h. The progress of the 17 reaction was monitored through the TLC test. After the reaction was 18 completed, the solvent was removed under reduced pressure. The crude 19 residue was purified by column chromatography on silica gel using ethyl 20 acetate/petroleum ether to afford the title compound as pale yellow solid. 21 Compound 12 were prepared using the identical synthetic procedure as 22

- 1 **11.**
- 2 4.1.6.1. indolin-2-one (11)
- Light yellow solid, yield: 72.06%, mp: 122.4°C. ¹H NMR (500 MHz,
- 4 DMSO- d_6) δ 10.38 (s, 1H, N-H), 7.22 7.11 (m, 2H, Ar-H), 6.91 (t, J =
- 5 7.4 Hz, 1H, Ar-H), 6.81 (d, J = 7.7 Hz, 1H, Ar-H), 3.45 (s, 2H, CH₂).
- 6 ESI-MS $[M+H]^+ m/z$: 134.03.
- 7 4.1.6.2. 5-fluoroindolin-2-one (12)
- 8 Light yellow solid, yield: 62.14%, mp: 142.4°C. ¹H NMR (500 MHz,
- 9 DMSO- d_6) δ 10.35 (s, 1H, N-H), 7.08 (d, J = 8.4 Hz, 1H, Ar-H), 6.97 (t, J
- 10 = 9.4 Hz, 1H, Ar-H), 6.77 (d, J = 4.1 Hz, 1H, Ar-H), 3.48 (s, 2H, CH₂).
- 11 ESI-MS $[M+H]^+ m/z$: 152.06.

12 4.1.7. The synthesis of target compounds (13a-14n)

Compound 13a was synthesized by using the following procedure: 13 Reaction of indolin-2-ones (11) (1.00 g, 7.51mmol) with compound 4a 14 (2.06g, 8.26 mmol) in ethanol (35 mL) was proceeded at 80°C for 4-8 h in 15 the presence of catalytic amount of piperidine. The progress of the 16 reaction was monitored through the TLC test. After the reaction was 17 completed, the reaction system was cooled to room temperature. The 18 precipitate thus formed were collected by filtration, washed with ethanol 19 $(2 \times 15 \text{ mL})$, and dried to yield pure target compound 13a in moderate 20 yield. 21



- 1 the identical synthetic procedure as **13a**.
- 2 *4.1.7.1*.

(Z)-3-[4-(1-phenvl-1H-1,2,3-triazol-4-vl)benzylidene]indolin-2-one (13a) 3 Yellow powder, yield: 62.44%, mp: 243°C. FI-IR (KBr, v_{max} cm⁻¹): 4 3431 (NH), 1704 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.64 (s, 1H, 5 N-H), 9.43 (s, 1H, H-triazole), 8.10 (d, J = 7.8 Hz, 2H, Ar-H), 7.98 (d, J =6 7.7 Hz, 2H, Ar-H), 7.86 (d, J = 7.7 Hz, 2H, H-vinylic, Ar-H), 7.68 – 7.62 7 (m, 4H, Ar-H), 7.54 (t, J = 7.2 Hz, 1H, Ar-H), 7.25 (t, J = 7.5 Hz, 1H, 8 Ar-H), 6.88 (t, J = 7.8 Hz, 2H, Ar-H). ¹³C NMR (126 MHz, DMSO- d_6) δ 9 168.48 (C=O), 146.59 (C-triazole), 142.88, 136.45 (C-vinylic), 135.06, 10 134.09, 131.24 (C-triazole), 130.07, 130.03 (2C), 129.82 (2C), 128.67, 11 127.57, 125.37 (2C), 122.34, 121.03, 120.74, 120.13 (C-vinylic), 119.92 12 (2C), 110.02. ESI-HRMS calcd for $C_{23}H_{17}N_4O$ [M+H]⁺365.1324, found: 13 365.1398. 14

15 *4.1.7.2*.

16 (Z)-3-{4-[1-(o-tolyl)-1H-1,2,3-triazol-4-yl]benzylidene}indolin-2-one

17 (**13b**)

Orange powder, yield: 53.42%, mp: 239°C. FI-IR (KBr, v_{max} cm⁻¹): 3441 (NH), 1711 (C=O). ¹H NMR (500 MHz, DMSO-*d*₆) δ 10.65 (s, 1H, N-H), 9.08 (s, 1H, H-triazole), 8.11 (d, *J* = 7.8 Hz, 2H, Ar-H), 7.85 (d, *J* = 7.7 Hz, 2H, H-vinylic, Ar-H), 7.67 (s, 1H, Ar-H), 7.63 (d, *J* = 7.6 Hz, 1H, Ar-H), 7.56 – 7.50 (m, 3H, Ar-H), 7.46 (s, 1H, Ar-H), 7.24 (t, *J* = 7.6 Hz,

1	1H, Ar-H), $6.94 - 6.84$ (m, 2H, Ar-H), 2.24 (s, 3H, CH ₃). ¹³ C NMR (126)
2	MHz, DMSO-d ₆) δ 168.50 (C=O), 145.77 (C-triazole), 142.88, 136.07
3	(C-vinylic), 135.11, 133.97, 132.91 (C-triazole), 132.53, 131.29, 129.99
4	(2C), 129.78, 127.55, 126.90, 125.83, 125.35 (2C), 124.76, 123.52,
5	122.33, 121.01 (C-vinylic), 120.77, 110.01, 17.31 (CH ₃). ESI-HRMS
6	calcd for $C_{24}H_{19}N_4O [M+H]^+$ 379.1481, found: 379.1560.
7	4.1.7.3.
8	(Z)-3-{4-[1-(m-tolyl)-1H-1,2,3-triazol-4-yl]benzylidene}indolin-2-one
9	(13c)
10	Brown powder, yield: 64.61%, mp: 240°C. FI-IR (KBr, v_{max} cm ⁻¹): 3405
11	(NH), 1700 (C=O). ¹ H NMR (500 MHz, DMSO- d_6) δ 10.64 (s, 1H, N-H),
12	9.39 (s, 1H, H-triazole), 8.09 (d, J = 7.2 Hz, 2H, Ar-H), 7.85 (d, J = 7.3
13	Hz, 2H, H-vinylic, Ar-H), 7.81 (s, 1H, Ar-H), 7.76 (d, J = 7.5 Hz, 1H,
14	Ar-H), 7.66 (s, 1H, Ar-H), 7.63 (d, J = 7.5 Hz, 1H, Ar-H), 7.51 (t, J = 7.4
15	Hz, 1H, Ar-H), 7.33 (d, J = 6.9 Hz, 1H, Ar-H), 7.24 (t, J = 7.2 Hz, 1H,
16	Ar-H), 6.88 (t, $J = 8.3$ Hz, 2H, Ar-H), 2.44 (s, 3H, CH ₃). ¹³ C NMR (126
17	MHz, DMSO-d ₆) δ 168.49 (C=O), 146.51 (C-triazole), 142.88, 139.55
18	(C-vinylic), 136.40, 135.06, 134.06, 131.28 (C-triazole), 130.04, 130.01
19	(2C), 129.58, 129.22, 127.56, 125.34 (2C), 122.33, 121.01, 120.75,
20	120.30, 120.04 (C-vinylic), 116.98, 110.01, 20.79 (CH ₃). ESI-HRMS
21	calcd for $C_{24}H_{19}N_4O [M+H]^+$ 379.1481, found: 379.1550.
22	4.1.7.4.

1 (Z)-3-{4-[1-(p-tolyl)-1H-1,2,3-triazol-4-yl]benzylidene}indolin-2-one

2 (*13d*)

Yellow powder, yield: 51.64%, mp: 248°C. FI-IR (KBr, v_{max} cm⁻¹): 3 3427 (NH), 1692 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.63 (s, 1H, 4 N-H), 9.35 (s, 1H, H-triazole), 8.53 (d, J = 8.2 Hz, 2H, Ar-H), 8.03 (d, J =5 8.2 Hz, 2H, Ar-H), 7.85 (d, J = 7.3 Hz, 3H, H-vinylic, Ar-H), 7.73 (d, J =6 7.4 Hz, 1H, Ar-H), 7.45 (d, J = 7.9 Hz, 2H, Ar-H), 7.23 (t, J = 7.6 Hz, 1H, 7 Ar-H), 7.01 (t, J = 7.5 Hz, 1H, Ar-H), 6.84 (d, J = 7.7 Hz, 1H, Ar-H), 2.41 8 (s, 3H, CH₃). ¹³C NMR (126 MHz, DMSO- d_6) δ 166.99 (C=O), 146.58 9 (C-triazole), 140.67, 138.34 (C-vinylic), 135.86, 134.23, 133.65, 132.50 10 (2C), 131.75 (C-triazole), 130.13 (2C), 128.85, 126.72, 125.37, 124.76 11 (2C), 120.93, 120.11 (C-vinylic), 119.85 (2C), 119.65, 109.23, 20.44 12 (CH₃). ESI-HRMS calcd for $C_{24}H_{19}N_4O$ [M+H]⁺ 379.1481, found: 13 379.1550. 14

4.1.7.5. (Z)-3-{4-[1-(2-fluorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}
indolin-2-one (13e)

Yellow powder, yield: 57.81%, mp: 234°C. FI-IR (KBr, v_{max} cm⁻¹): 3504 (NH), 1693 (C=O). ¹H NMR (500 MHz, DMSO-*d*₆) δ 10.63 (s, 1H, N-H), 9.19 (s, 1H, H-triazole), 8.12 (d, *J* = 7.9 Hz, 2H, Ar-H), 7.93 (t, *J* = 7.8 Hz, 1H, Ar-H), 7.85 (d, *J* = 8.0 Hz, 2H, H-vinylic, Ar-H), 7.68 – 7.59 (m, 4H, Ar-H), 7.48 (t, *J* = 7.6 Hz, 1H, Ar-H), 7.24 (t, *J* = 7.7 Hz, 1H, Ar-H), 6.88 (dd, *J* = 13.5, 7.3 Hz, 2H, Ar-H). ¹³C NMR (126 MHz,

1	DMSO- <i>d</i> ₆) δ 168.47 (C=O), 154.69, 152.70, 146.21 (C-triazole), 142.88,
2	135.02 (C-vinylic), 134.17, 132.50 (C-triazole), 130.99, 130.01 (d, $J =$
3	6.0 Hz, 2C, C-fluorobenzene), 127.60, 125.83 (d, $J = 3.3$ Hz,
4	C-fluorobenzene), 125.44 (d, $J = 4.6$ Hz, 2C, C-fluorobenzene), 124.86
5	(C-vinylic), 123.25 (d, $J = 4.1$ Hz, C-fluorobenzene), 122.33, 121.00,
6	120.74, 117.11, 116.96, 110.00. ESI-HRMS calcd for C ₂₃ H ₁₆ FN ₄ O
7	[M+H] ⁺ 383.1230, found: 383.1307.
8	4.1.7.6. (Z)-3-{4-[1-(3-fluorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}
9	indolin-2-one (13f)
10	Yellow powder, yield: 63.46%, mp: 246°C. FI-IR (KBr, v_{max} cm ⁻¹):
11	3286 (NH), 1687 (C=O). ¹ H NMR (500 MHz, DMSO- d_6) δ 10.66 (s, 1H,
12	N-H), 9.45 (s, 1H, H-triazole), 8.53 (d, J = 8.0 Hz, 2H, Ar-H), 8.02 (d, J =
13	7.9 Hz, 2H, Ar-H), 7.90 – 7.81 (m, 3H, H-vinylic, Ar-H), 7.70 (dd, J =
14	19.4, 7.4 Hz, 2H, Ar-H), 7.38 (t, <i>J</i> = 8.3 Hz, 1H, Ar-H), 7.22 (t, <i>J</i> = 7.4 Hz,
15	1H, Ar-H), 7.00 (t, <i>J</i> = 7.4 Hz, 1H, Ar-H), 6.83 (d, <i>J</i> = 7.6 Hz, 1H, Ar-H).
16	¹³ C NMR (126 MHz, DMSO- d_6) δ 166.98 (C=O), 163.28, 161.33, 146.80
17	(C-triazole), 140.69, 135.78 (C-vinylic), 133.79 (C-triazole), 132.52 (2C),
18	131.75 (d, $J = 9.4$ Hz, C-fluorobenzene), 131.41, 130.05 (d, $J = 4.1$ Hz,
19	C-fluorobenzene), 128.87, 126.81, 125.39, 124.78 (2C), 120.92, 120.38
20	(C-vinylic), 119.66, 115.83 (d, $J = 3.4$ Hz, C-fluorobenzene), 109.23,
21	107.29 (d, $J = 3.3$ Hz, C-fluorobenzene). ESI-HRMS calcd for
22	$C_{23}H_{16}FN_4O [M+H]^+ 383.1230$, found: 383.1298.

1 4.1.7.7. (Z)-3-{4-[1-(4-fluorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}

2 *indolin-2-one* (**13g**)

Yellow powder, yield: 63.91%, mp: 238°C. FI-IR (KBr, v_{max} cm⁻¹): 3 3448 (NH), 1686 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.66 (s, 1H, 4 N-H), 9.35 (s, 1H, H-triazole), 8.52 (d, J = 7.7 Hz, 2H, Ar-H), 8.01 (d, J =5 7.6 Hz, 4H, Ar-H), 7.81 (s, 1H, H-vinylic), 7.71 (d, J = 7.1 Hz, 1H, Ar-H), 6 7.49 (t, J = 8.1 Hz, 2H, Ar-H), 7.21 (t, J = 7.1 Hz, 1H, Ar-H), 6.99 (t, J = 7 7.1 Hz, 1H, Ar-H), 6.83 (d, J = 7.3 Hz, 1H, Ar-H). ¹³C NMR (126 MHz, 8 DMSO- d_6) δ 166.96 (C=O), 162.55, 160.60, 146.69 (C-triazole), 140.66, 9 135.82 (C-vinylic), 133.70 (C-triazole), 133.00 (d, J = 3.0 Hz, 10 C-fluorobenzene), 132.51 (2C), 131.58, 130.02, 128.86, 126.74, 125.35, 11 124.76 (d, J = 3.2 Hz, 2C, C-fluorobenzene), 122.32 (d, J = 8.9 Hz), 12 120.92 (C-vinylic), 120.51, 119.65, 116.65 (d, J = 23.3 Hz, 13 C-fluorobenzene), 109.21. ESI-HRMS calcd for $C_{23}H_{16}FN_4O [M+H]^+$ 14 383.1230, found: 383.1313. 15

4.1.7.8. (Z)-3-{4-[1-(2-chlorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}
indolin-2-one (13h)

Yellow powder, yield: 64.14%, mp: 229°C. FI-IR (KBr, v_{max} cm⁻¹): 3431 (NH), 1710 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.63 (s, 1H, N-H), 9.17 (s, 1H, H-triazole), 8.11 (d, J = 7.9 Hz, 2H, Ar-H), 7.89 – 7.78 (m, 4H, H-vinylic, Ar-H), 7.71 – 7.60 (m, 4H, Ar-H), 7.24 (t, J = 7.7 Hz, 1H, Ar-H), 6.88 (t, J = 9.0 Hz, 2H, Ar-H). ¹³C NMR (126 MHz,

1	DMSO- d_6) δ 168.47 (C=O), 145.81 (C-triazole), 142.89, 135.86
2	(C-vinylic), 135.05, 134.32, 132.55, 131.70, 131.11 (C-triazole), 130.47,
3	130.02 (2C), 128.39, 128.29, 127.63, 125.41 (2C), 124.80, 124.08, 122.35,
4	121.03 (C-vinylic), 119.66, 110.02. ESI-HRMS calcd for $C_{23}H_{16}ClN_4O$
5	[M+H] ⁺ 399.0934, found: 399.1011.
6	4.1.7.9. (Z)-3-{4-[1-(3-chlorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}
7	indolin-2-one (13i)
8	Orange powder, yield: 60.19%, mp: 227°C. FI-IR (KBr, v_{max} cm ⁻¹):
9	3430 (NH), 1685 (C=O). ¹ H NMR (500 MHz, DMSO- d_6) δ 10.66 (s, 1H,
10	N-H), 9.48 (s, 1H, H-triazole), 8.53 (d, J = 8.0 Hz, 2H, Ar-H), 8.10 (s, 1H,
11	Ar-H), 8.00 (dd, J = 18.4, 8.0 Hz, 3H, Ar-H), 7.84 (s, 1H, H-vinylic), 7.72
12	(d, J = 7.6 Hz, 1H, Ar-H), 7.68 (t, J = 8.1 Hz, 1H, Ar-H), 7.60 (d, J = 8.0
13	Hz, 1H, Ar-H), 7.22 (t, J = 7.5 Hz, 1H, Ar-H), 7.00 (t, J = 7.3 Hz, 1H,
14	Ar-H), 6.84 (d, J = 7.6 Hz, 1H, Ar-H). ¹³ C NMR (126 MHz, DMSO- d_6) δ
15	166.97 (C=O), 146.80 (C-triazole), 140.68, 137.46 (C-vinylic), 135.80,
16	134.10, 133.79, 132.53 (2C), 131.55, 131.42 (C-triazole), 128.89, 128.44,
17	126.80, 124.77 (3C), 120.93, 120.42 (C-vinylic), 119.71, 119.67, 118.50,
18	109.23. ESI-HRMS calcd for $C_{23}H_{16}ClN_4O$ [M+H] ⁺ 399.0934, found:
19	399.1000.

20 4.1.7.10. (Z)-3-{4-[1-(4-chlorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}

21 *indolin-2-one* (*13j*)

22 Yellow powder, yield: 59.06%, mp: 235°C. FI-IR (KBr, v_{max} cm⁻¹):

1	3432 (NH), 1689 (C=O). ¹ H NMR (500 MHz, DMSO- d_6) δ 10.66 (s, 1H,
2	N-H), 9.45 (s, 1H, H-triazole), 8.53 (d, J = 7.9 Hz, 2H, Ar-H), 8.02 (t, J =
3	8.4 Hz, 4H, Ar-H), 7.84 (s, 1H, H-vinylic), 7.74 (d, <i>J</i> = 7.7 Hz, 3H, Ar-H),
4	7.22 (t, <i>J</i> = 7.4 Hz, 1H, Ar-H), 7.01 (t, <i>J</i> = 7.4 Hz, 1H, Ar-H), 6.84 (d, <i>J</i> =
5	7.6 Hz, 1H, Ar-H). ¹³ C NMR (126 MHz, DMSO- d_6) δ 166.98 (C=O),
6	146.81 (C-triazole), 140.67, 135.83 (C-vinylic), 135.23, 133.76, 132.95,
7	132.53 (2C), 131.49 (C-triazole), 129.80 (2C), 128.89, 126.78, 125.61,
8	124.78 (2C), 121.61 (2C), 120.94, 120.33 (C-vinylic), 119.68, 109.23.
9	ESI-HRMS calcd for $C_{23}H_{16}CIN_4O [M+H]^+$ 399.0934, found: 399.1018.
10	4.1.7.11.

11 (Z)-2-(4-{4-[(2-oxoindolin-3-ylidene)methyl]phenyl}-1H-1,2,3-triazol-1-y
12 l)benzonitrile (13k)

Brown powder, yield: 49.58%, mp: 239°C. FI-IR (KBr, v_{max} cm⁻¹): 3432 13 (NH), 1693 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.65 (s, 1H, N-H), 14 9.36 (s, 1H, H-triazole), 8.20 (d, J = 7.8 Hz, 1H, Ar-H), 8.12 (d, J = 7.715 Hz, 2H, Ar-H), 8.06 - 7.96 (m, 2H, Ar-H), 7.88 (d, J = 7.8 Hz, 2H, Ar-H), 16 7.82 (t, J = 7.5 Hz, 1H, H-vinylic), 7.67 (s, 1H, Ar-H), 7.62 (d, J = 7.7 Hz, 17 1H, Ar-H), 7.25 (t, J = 7.6 Hz, 1H, Ar-H), 6.88 (t, J = 7.7 Hz, 2H, Ar-H). 18 ¹³C NMR (126 MHz, DMSO- d_6) δ 168.47 (C=O), 146.49 (C-triazole), 19 142.91, 137.65 (C-vinylic), 134.95, 134.74, 134.65, 134.39, 132.55 20 (C-triazole), 130.76, 130.29, 130.06 (2C), 127.73, 125.56, 125.52 (2C), 21 122.90, 122.38, 121.04, 120.74 (C-vinylic), 115.62, 110.02, 106.97. 22

- ESI-HRMS calcd for $C_{24}H_{16}N_5O [M+H]^+$ 390.1277, found: 390.1349. 1 4.1.7.12. 2 (Z)-3-(4-{4-[(2-oxoindolin-3-ylidene)methyl]phenyl}-1H-1,2,3-triazol-1-y 3 *l)benzonitrile* (131) 4 Yellow powder, yield: 60.12%, mp: 240°C. FI-IR (KBr, v_{max} cm⁻¹): 5 3433 (NH), 1696 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.62 (s, 1H, 6 N-H), 9.52 (s, 1H, H-triazole), 8.49 (s, 1H, Ar-H), 8.35 (d, J = 8.2 Hz, 1H, 7 Ar-H), 8.07 (d, J = 8.0 Hz, 2H, Ar-H), 8.01 (d, J = 7.7 Hz, 1H, Ar-H), 8 7.87 (d, J = 7.5 Hz, 3H, H-vinylic, Ar-H), 7.66 (s, 1H, Ar-H), 7.62 (d, J =9 7.7 Hz, 1H, Ar-H), 7.25 (t, J = 7.7 Hz, 1H, Ar-H), 6.88 (t, J = 7.9 Hz, 2H, 10 Ar-H). ¹³C NMR (126 MHz, DMSO- d_6) δ 168.46 (C=O), 146.85 11 (C-triazole), 142.92, 136.87 (C-vinylic), 134.93, 134.33, 132.53, 132.18, 12 131.24 (C-triazole), 130.06 (2C), 127.71, 125.41 (2C), 124.79, 124.49, 13 123.26, 122.36, 121.02, 120.74, 120.38 (C-vinylic), 117.62, 112.74, 14 110.02. ESI-HRMS calcd for $C_{24}H_{16}N_5O$ [M+H]⁺ 390.1277, found: 15 390.1355. 16
- 17 *4.1.7.13*.
- 18 (Z/E)-4- $(4-\{4-[(2-oxoindolin-3-ylidene)methyl]phenyl\}$ -1H-1,2,3-triazol-1
- 19 -yl)benzonitrile (**13m**)

20 Yellow powder, yield: 53.41%, mp: 241°C. FI-IR (KBr, v_{max} cm⁻¹): 21 3430 (NH), 1684 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.63 (s, 1H,

22 N-H), 9.56 (s, 0.3H, H-triazole), 9.53 (s, 0.7H, H-triazole), 8.52 (d, J =

1	7.8 Hz, 1.3H, Ar-H), 8.20 (d, $J = 7.7$ Hz, 2H, Ar-H), 8.14 (d, $J = 8.3$ Hz,
2	2H, Ar-H), 8.08 (d, J = 7.6 Hz, 0.7H, Ar-H), 8.02 (d, J = 7.9 Hz, 1.3H,
3	Ar-H), 7.86 (d, J = 7.7 Hz, 0.6H, H-vinylic, Ar-H), 7.82 (s, 0.7H,
4	H-vinylic), 7.71 (d, <i>J</i> = 7.4 Hz, 0.7H, Ar-H), 7.65 (s, 0.3H, Ar-H), 7.61 (d,
5	<i>J</i> = 7.7 Hz, 0.3H, Ar-H), 7.23 (q, <i>J</i> = 9.2, 7.6 Hz, 1H, Ar-H), 7.00 (t, <i>J</i> =
6	7.4 Hz, 0.7H, Ar-H), 6.89 (d, <i>J</i> = 8.0 Hz, 0.7H, Ar-H), 6.84 (d, <i>J</i> = 7.7 Hz,
7	0.7H, Ar-H). ESI-HRMS calcd for $C_{24}H_{16}N_5O$ [M+H] ⁺ 390.1277, found:
8	390.1349.

9 4.1.7.14.

10 (Z)-3-(4-{1-[4-chloro-3-(trifluoromethyl)phenyl]-1H-1,2,3-triazol-4-yl}be
 11 nzylidene)indolin-2-one (13n)

Yellow powder, yield: 63.92%, mp: 219°C. FI-IR (KBr, v_{max} cm⁻¹): 12 3432 (NH), 1677(C=O). ¹H NMR (500 MHz, DMSO-*d*₆) δ 10.66 (s, 1H, 13 N-H), 9.59 (d, J = 2.1 Hz, 1H, H-triazole), 8.53 (d, J = 6.6 Hz, 2H, Ar-H), 14 8.43 (s, 1H, Ar-H), 8.32 (d, J = 8.4 Hz, 1H, Ar-H), 8.03 (t, J = 7.5 Hz, 3H, 15 Ar-H), 7.84 (s, 1H, H-vinylic), 7.72 (d, J = 6.5 Hz, 1H, Ar-H), 7.22 (t, J = 16 6.5 Hz, 1H, Ar-H), 7.00 (t, J = 6.4 Hz, 1H, Ar-H), 6.83 (d, J = 6.1 Hz, 1H, 17 Ar-H). ¹³C NMR (126 MHz, DMSO- d_6) δ 166.94 (C=O), 146.98 18 (C-triazole), 140.67, 135.73 (C-vinylic), 135.37, 133.86, 133.26, 132.53 19 (2C), 131.23 (C-triazole), 130.06, 128.89, 127.81 (d, J = 31.6 Hz, 20 C-trifluoromethylphenyl), 126.84, 125.38, 125.06, 124.75 (d, J = 3.6 Hz, 21 2C, C-trifluoromethylphenyl, Ar-C), 123.23, 120.92, 120.64 (C-vinylic), 22

1	119.67, 119.15 (d, $J = 5.2$ Hz, C-trifluoromethylphenyl), 109.22.
2	ESI-HRMS calcd for $C_{24}H_{15}ClF_{3}N_{4}O$ [M+H] ⁺ 467.0808, found:
3	467.0887.
4	4.1.7.15. (Z/E)-5-fluoro-3-[4-(1-phenyl-1H-1,2,3-triazol-4-yl)benzylidene]
5	indolin-2-one (14a)
6	Orange powder, yield: 69.92%, mp: 229°C. FI-IR (KBr, v_{max} cm ⁻¹):
7	3432 (NH), 1687 (C=O). ¹ H NMR (500 MHz, DMSO- d_6) δ 10.67 (s, 1H,
8	N-H), 9.44 (s, 0.5H, H-triazole), 9.42 (s, 0.5H, H-triazole), 8.54 (d, J =
9	7.8 Hz, 1H, Ar-H), 8.12 (d, <i>J</i> = 8.0 Hz, 1H, Ar-H), 8.06 (d, <i>J</i> = 7.9 Hz, 1H,
10	Ar-H), 7.97 (d, J = 7.6 Hz, 2H, Ar-H), 7.92 (s, 0.5H, Ar-H), 7.86 (d, J =
11	7.7 Hz, 1H, H-vinylic), 7.74 (s, 0.5H, Ar-H), 7.65 (t, J = 7.7 Hz, 2.5H,
12	Ar-H), 7.53 (t, J = 7.4 Hz, 1H, Ar-H), 7.33 (d, J = 9.0 Hz, 0.5H, Ar-H),
13	7.11 (t, <i>J</i> = 9.0 Hz, 0.5H, Ar-H), 7.05 (t, <i>J</i> = 9.1 Hz, 0.5H, Ar-H), 6.88 (dd,
14	J = 8.4, 4.5 Hz, 0.5H, Ar-H), 6.81 (dd, $J = 8.3, 4.3$ Hz, 0.5H, Ar-H).
15	ESI-HRMS calcd for $C_{23}H_{16}FN_4O [M+H]^+$ 383.1230, found: 383.1313.
16	4.1.7.16. (Z/E)-5-fluoro-3-{4-[1-(o-tolyl)-1H-1,2,3-triazol-4-yl]
17	benzylidene} indolin-2-one (14b)
18	Yellow powder, yield: 55.15%, mp: 221°C. FI-IR (KBr, v_{max} cm ⁻¹):
19	3432 (NH), 1701 (C=O). ¹ H NMR (500 MHz, DMSO- d_6) δ 10.68 (s, 1H,

20 N-H), 9.09 (d, J = 1.1 Hz, 1H, H-triazole), 8.54 (d, J = 8.2 Hz, 1H, Ar-H),

21 8.12 (d, J = 8.0 Hz, 1H, Ar-H), 8.06 (d, J = 8.2 Hz, 1H, Ar-H), 7.93 (s,

22 0.4H, Ar-H), 7.85 (d, J = 8.0 Hz, 1H, H-vinylic), 7.74 (s, 0.6H, Ar-H),

1	7.67 (d, J = 9.0 Hz, 0.4H, Ar-H), 7.56 – 7.51 (m, 3H, Ar-H), 7.48 – 7.43
2	(m, 1H, Ar-H), 7.32 (d, J = 9.2 Hz, 0.6H, Ar-H), 7.10 (t, J = 8.9 Hz, 0.6H,
3	Ar-H), 7.04 (t, J = 9.0 Hz, 0.4H, Ar-H), 6.88 (dd, J = 8.0, 4.7 Hz, 0.6H,
4	Ar-H), 6.81 (dd, $J = 7.8$, 4.4 Hz, 0.4H, Ar-H), 2.24 (s, 3H, CH ₃).
5	ESI-HRMS calcd for $C_{24}H_{18}FN_4O [M+H]^+$ 397.1386, found: 397.1463.
6	4.1.7.17. (Z)-5-fluoro-3-{4-[1-(m-tolyl)-1H-1,2,3-triazol-4-yl]benzylidene}
7	indolin-2-one (14c)
8	Orange powder, yield: 65.29%, mp: 217°C. FI-IR (KBr, v_{max} cm ⁻¹):
9	3432 (NH), 1686 (C=O). ¹ H NMR (500 MHz, DMSO- d_6) δ 10.66 (s, 1H,
10	N-H), 9.39 (s, 1H, H-triazole), 8.54 (d, J = 8.0 Hz, 2H, Ar-H), 8.05 (d, J =
11	8.0 Hz, 2H, Ar-H), 7.92 (s, 1H, Ar-H), 7.81 (s, 1H, H-vinylic), 7.75 (d, J
12	= 8.2 Hz, 1H, Ar-H), 7.66 (d, J = 9.3 Hz, 1H, Ar-H), 7.52 (t, J = 7.8 Hz,
13	1H, Ar-H), 7.34 (d, <i>J</i> = 7.6 Hz, 1H, Ar-H), 7.05 (t, <i>J</i> = 9.4 Hz, 1H, Ar-H),
14	6.81 (dd, $J = 8.6$, 4.4 Hz, 1H, Ar-H), 2.45 (s, 3H, CH ₃). ¹³ C NMR (126
15	MHz, DMSO-d ₆) δ 166.99 (C=O), 158.74, 156.87, 146.52 (C-triazole),
16	139.55, 137.61 (C-vinylic), 136.85, 136.37, 133.39, 132.75 (2C), 132.08
17	(C-triazole), 129.58, 129.25, 126.31 (d, $J = 8.6$ Hz, C-fluorobenzene),
18	124.79 (2C, C-vinylic, Ar-C), 120.30 (d, $J = 6.3$ Hz, C-fluorobenzene),
19	117.03, 114.98 (d, $J = 24.0$ Hz, C-fluorobenzene), 109.94 (d, $J = 8.3$ Hz,
20	C-fluorobenzene), 107.20, 107.00, 20.79 (CH ₃). ESI-HRMS calcd for
21	$C_{24}H_{18}FN_4O [M+H]^+ 397.1386$, found: 397.1460.

22 4.1.7.18. (Z)-5-fluoro-3-{4-[1-(p-tolyl)-1H-1,2,3-triazol-4-yl]benzylidene}

1 *indolin-2-one* (**14d**)

Orange powder, yield: 56.25%, mp: 204°C. FI-IR (KBr, v_{max} cm⁻¹): 2 3433 (NH), 1683 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.67 (s, 1H, 3 N-H), 9.36 (s, 1H, H-triazole), 8.54 (d, J = 7.9 Hz, 2H, Ar-H), 8.05 (d, J =4 7.8 Hz, 2H, Ar-H), 7.92 (s, 1H, Ar-H), 7.84 (d, J = 7.6 Hz, 2H, H-vinylic, 5 Ar-H), 7.67 (d, J = 9.0 Hz, 1H, Ar-H), 7.45 (d, J = 7.8 Hz, 2H, Ar-H), 6 7.05 (t, J = 9.1 Hz, 1H, Ar-H), 6.81 (dd, J = 7.8, 4.2 Hz, 1H, Ar-H), 2.40 7 (s, 3H, CH₃). ¹³C NMR (126 MHz, DMSO- d_6) δ 167.01 (C=O), 158.76, 8 156.89, 146.50 (C-triazole), 138.35, 137.65, 136.86 (C-vinylic), 134.19, 9 133.38, 132.76 (2C), 132.13 (C-triazole), 130.14 (2C), 126.35 (d, J = 3.4 10 Hz, C-fluorobenzene), 124.80 (2C), 120.22 (C-vinylic), 119.82 (2C), 11 114.99 (d, J = 24.1 Hz, C-fluorobenzene), 109.96 (d, J = 8.1 Hz, 12 C-fluorobenzene), 107.12 (d, J = 25.4 Hz, C-fluorobenzene), 20.44 (CH₃). 13 ESI-HRMS calcd for $C_{24}H_{18}FN_4O [M+H]^+$ 397.1386, found: 397.1464. 14 4.1.7.19. 15

16 (Z)-5-fluoro-3-{4-[1-(2-fluorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}i

17 *ndolin-2-one* (**14e**)

Yellow powder, yield: 64.18%, mp: 248°C. FI-IR (KBr, v_{max} cm⁻¹): 3432 (NH), 1699 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.66 (s, 1H, N-H), 9.21 (s, 1H, H-triazole), 8.14 (d, J = 7.9 Hz, 2H, Ar-H), 7.93 (t, J =7.6 Hz, 1H, Ar-H), 7.84 (d, J = 7.8 Hz, 2H, H-vinylic, Ar-H), 7.73 (s, 1H,

22 Ar-H), 7.68 - 7.58 (m, 2H, Ar-H), 7.48 (t, J = 7.4 Hz, 1H, Ar-H), 7.31 (d,

1	<i>J</i> = 9.0 Hz, 1H, Ar-H), 7.09 (t, <i>J</i> = 8.6 Hz, 1H, Ar-H), 6.90 – 6.84 (m, 1H,
2	Ar-H). ¹³ C NMR (126 MHz, DMSO- d_6) δ 168.44 (C=O), 157.95, 156.08,
3	154.71, 152.71, 146.14 (C-triazole), 139.21, 136.73 (C-vinylic), 133.72,
4	131.34 (C-triazole), 130.03 (2C), 127.32 (d, <i>J</i> = 2.7 Hz, C-fluorobenzene),
5	125.83, 125.57 (2C, C-vinylic, Ar-C), 125.45 (d, $J = 3.5$ Hz,
6	C-fluorobenzene), 123.38 (d, $J = 4.2$ Hz, C-fluorobenzene), 121.68 (d, J
7	= 8.7 Hz, C-fluorobenzene), 117.06 (d, $J = 19.4$ Hz, C-fluorobenzene),
8	116.41, 110.68 (d, $J = 8.4$ Hz, C-fluorobenzene), 109.23. ESI-HRMS
9	calcd for $C_{23}H_{15}F_2N_4O [M+H]^+ 401.1136$, found: 401.1201.
10	4.1.7.20.

11 (Z)-5-fluoro-3-{4-[1-(3-fluorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}i
 12 ndolin-2-one (14f)

Orange powder, yield: 51.56%, mp: 241°C. FI-IR (KBr, v_{max} cm⁻¹): 13 3432 (NH), 1685 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.65 (s, 1H, 14 N-H), 9.45 (s, 1H, H-triazole), 8.53 (d, J = 8.1 Hz, 2H, Ar-H), 8.03 (d, J =15 8.2 Hz, 2H, Ar-H), 7.93 – 7.82 (m, 3H, H-vinylic, Ar-H), 7.74 – 7.62 (m, 16 2H, Ar-H), 7.39 (t, J = 8.5 Hz, 1H, Ar-H), 7.04 (t, J = 8.9 Hz, 1H, Ar-H), 17 6.81 (dd, J = 8.1, 4.4 Hz, 1H, Ar-H). ¹³C NMR (126 MHz, DMSO- d_6) δ 18 168.43 (C=O), 163.27, 161.32, 156.08, 146.62 (C-triazole), 139.23, 19 137.63 (d, J = 10.4 Hz, C-fluorobenzene), 136.70 (C-vinylic), 133.76, 20 132.75, 131.78 (d, J = 9.3 Hz, C-fluorobenzene), 131.34 (C-triazole), 21 130.08 (2C), 127.35 (d, J = 2.8 Hz, C-fluorobenzene), 125.47 (2C), 22

- 1 124.83 (C-vinylic), 120.46 (d, J = 8.8 Hz, C-fluorobenzene), 116.34 (d, J
- 2 = 23.6 Hz, C-fluorobenzene), 115.80 (d, J = 2.9 Hz, C-fluorobenzene),
- 3 110.70 (d, J = 8.1 Hz, C-fluorobenzene), 107.50 (d, J = 3.8 Hz,
- 4 C-fluorobenzene). ESI-HRMS calcd for $C_{23}H_{15}F_2N_4O [M+H]^+ 401.1136$,
- 5 found: 401.1212.
- 6 *4.1.7.21*.
- 7 (Z)-5-fluoro-3-{4-[1-(4-fluorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}i
 8 ndolin-2-one (14g)

Orange powder, yield: 66.81%, mp: 252°C. FI-IR (KBr, v_{max} cm⁻¹): 9 3440 (NH), 1689 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.66 (s, 1H, 10 N-H), 9.36 (s, 1H, H-triazole), 8.52 (d, J = 8.1 Hz, 2H, Ar-H), 8.04 – 7.97 11 (m, 4H, Ar-H), 7.88 (s, 1H, H-vinylic), 7.64 (d, J = 8.5 Hz, 1H, Ar-H), 12 7.49 (t, J = 8.5 Hz, 2H, Ar-H), 7.03 (t, J = 8.8 Hz, 1H, Ar-H), 6.80 (dd, J 13 = 8.2, 4.3 Hz, 1H, Ar-H). ¹³C NMR (126 MHz, DMSO- d_6) δ 167.00 14 (C=O), 162.54, 160.59, 158.74, 156.87, 146.62 (C-triazole), 137.56, 15 136.84 (C-vinylic), 133.42, 132.96 (d, J = 2.8 Hz, C-fluorobenzene), 16 132.74 (2C), 131.95 (C-triazole), 126.36 (d, J = 3.7 Hz, C-fluorobenzene), 17 124.78 (2C), 122.25 (d, J = 9.0 Hz, C-fluorobenzene), 120.53 (C-vinylic), 18 116.62 (d, J = 23.1 Hz, 2C, C-fluorobenzene), 114.95 (d, J = 23.8 Hz, 19 C-fluorobenzene), 109.92 (d, J = 8.3 Hz, C-fluorobenzene), 107.09 (d, J 20 = 25.4 Hz, C-fluorobenzene). ESI-HRMS calcd for $C_{23}H_{15}F_2N_4O [M+H]^+$ 21 401.1208, found: 401.1215. 22

- 1 4.1.7.22.
- 2 (Z)-3-{4-[1-(2-chlorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}-5-fluoroi
 3 ndolin-2-one (14h)
- Yellow powder, yield 57.36%, mp: 235°C. FI-IR (KBr, v_{max} cm⁻¹): 3432 4 (NH), 1708 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.68 (s, 1H, N-H), 5 9.19 (s, 1H, H-triazole), 8.13 (d, J = 7.8 Hz, 2H, Ar-H), 7.89 – 7.78 (m, 6 4H, H-vinylic, Ar-H), 7.74 (s, 1H, Ar-H), 7.71 – 7.61 (m, 2H, Ar-H), 7.32 7 (d, J = 9.1 Hz, 1H, Ar-H), 7.10 (t, J = 8.7 Hz, 1H, Ar-H), 6.88 (dd, J = 8.0),8 4.5 Hz, 1H, Ar-H). ¹³C NMR (126 MHz, DMSO- d_6) δ 168.44 (C=O), 9 157.96, 156.08, 145.73 (C-triazole), 139.21, 136.77 (C-vinylic), 134.29, 10 133.67, 131.70, 131.45, 130.46 (C-triazole), 130.07 (2C), 128.47, 128.33 11 (d, J = 14.0 Hz, C-fluorobenzene), 127.32 (d, J = 3.0 Hz, 12 C-fluorobenzene), 125.49 (2C), 124.20 (C-vinylic), 121.69 (d, J = 8.7 Hz, 13 C-fluorobenzene), 116.34 (d, J = 23.8 Hz, C-fluorobenzene), 110.70 (d, J14 = 8.4 Hz, C-fluorobenzene), 109.33 (d, J = 25.7 Hz, C-fluorobenzene). 15 ESI-HRMS calcd for $C_{23}H_{15}ClFN_4O [M+H]^+ 417.0840$, found: 417.0921. 16 4.1.7.23. 17
- 18 (Z)-3-{4-[1-(3-chlorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}-5-fluoroi
 19 ndolin-2-one (14i)
- Orange powder, yield: 64.52%, mp: 248°C. FI-IR (KBr, v_{max} cm⁻¹): 3432 (NH), 1685 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.66 (s, 1H, N-H), 9.47 (s, 1H, H-triazole), 8.53 (d, J = 8.0 Hz, 2H, Ar-H), 8.08 (s, 1H,

1	Ar-H), 8.02 (d, $J = 8.0$ Hz, 2H, Ar-H), 7.97 (d, $J = 8.1$ Hz, 1H, Ar-H),
2	7.90 (s, 1H, H-vinylic), 7.66 (q, J = 7.9 Hz, 2H, Ar-H), 7.59 (d, J = 8.0
3	Hz, 1H, Ar-H), 7.04 (t, J = 9.0 Hz, 1H, Ar-H), 6.80 (dd, J = 8.2, 4.3 Hz,
4	1H, Ar-H). ¹³ C NMR (126 MHz, DMSO- d_6) δ 166.99 (C=O), 158.74,
5	156.88, 146.72 (C-triazole), 137.49 (d, J = 14.2 Hz, C-fluorobenzene),
6	136.87 (C-vinylic), 134.10, 133.52, 132.76 (2C), 131.79, 131.53
7	(C-triazole), 128.44, 126.44 (d, J = 2.9 Hz, C-fluorobenzene), 126.29 (d,
8	J = 8.9 Hz, C-fluorobenzene), 124.81 (2C), 120.49 (C-vinylic), 119.68,
9	118.47, 115.00 (d, $J = 23.9$ Hz, C-fluorobenzene), 109.95 (d, $J = 8.1$ Hz,
10	C-fluorobenzene), 107.12 (d, $J = 25.4$ Hz, C-fluorobenzene). ESI-HRMS
11	calcd for $C_{23}H_{15}ClFN_4O [M+H]^+ 417.0840$, found: 417.0914.
12	4.1.7.24.

13 (Z/E)-3-{4-[1-(4-chlorophenyl)-1H-1,2,3-triazol-4-yl]benzylidene}-5-fluo
 14 roindolin-2-one (14j)

Orange powder, yield: 48.67%, mp: 233°C. FI-IR (KBr, v_{max} cm⁻¹): 15 3439 (NH), 1687 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.67 (s, 1H, 16 N-H), 9.45 (d, J = 11.2 Hz, 1H, H-triazole), 8.53 (d, J = 8.3 Hz, 1H, 17 Ar-H), 8.10 (d, J = 8.6 Hz, 1H, Ar-H), 8.02 (dd, J = 14.0, 8.5 Hz, 3H, 18 Ar-H), 7.91 (s, 0.5H, Ar-H), 7.86 (d, *J* = 8.0 Hz, 1H, H-vinylic), 7.73 (d, *J* 19 = 7.4 Hz, 2.5H, Ar-H), 7.66 (d, J = 9.0 Hz, 0.5H, Ar-H), 7.31 (d, J = 9.2 20 Hz, 0.5H, Ar-H), 7.11 (t, J = 8.8 Hz, 0.5H, Ar-H), 7.04 (t, J = 8.9 Hz, 21 0.5H, Ar-H), 6.88 (dd, J = 8.4, 4.6 Hz, 0.5H, Ar-H), 6.81 (dd, J = 8.3, 4.3 22

- 1 Hz, 0.5H, Ar-H). ESI-HRMS calcd for $C_{23}H_{15}ClFN_4O [M+H]^+ 417.0840$,
- 2 found: 417.0909.
- з 4.1.7.25.
- 4 (Z/E)-2- $(4-\{4-[(5-fluoro-2-oxoindolin-3-ylidene)methyl]phenyl\}$ -1H-1,2,3
- 5 *-triazol-1-yl)benzonitrile* (**14k**)
- 6 Brown powder, yield: 58.26%, mp: 218° C. FI-IR (KBr, v_{max} cm⁻¹): 3439
- 7 (NH), 1690 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.68 (s, 1H, N-H),
- 8 9.36 (s, 1H, H-triazole), 8.55 (d, J = 7.7 Hz, 1.7H, Ar-H), 8.19 (d, J = 7.2
- 9 Hz, 1H, Ar-H), 8.14 (d, J = 7.2 Hz, 0.3H, Ar-H), 8.07 (d, J = 7.6 Hz, 1.7H,
- 10 Ar-H), 7.99 (t, J = 9.3 Hz, 2H, Ar-H), 7.93 (s, 0.8H, Ar-H), 7.88 (d, J =
- 11 6.8 Hz, 0.3H, Ar-H), 7.81 (t, J = 6.7 Hz, 1H, H-vinylic), 7.74 (s, 0.2H,
- 12 Ar-H), 7.67 (d, J = 8.4 Hz, 0.8H, Ar-H), 7.32 (d, J = 8.8 Hz, 0.2H, Ar-H),
- 13 7.10 (d, J = 8.7 Hz, 0.2H, Ar-H), 7.05 (t, J = 8.8 Hz, 0.8H, Ar-H), 6.88 (s,
- 14 0.2H, Ar-H), 6.81 (d, J = 4.6 Hz, 0.8H, Ar-H). ESI-HRMS calcd for
- 15 $C_{24}H_{15}FN_5O[M+H]^+$ 408.1182, found: 408.1260.
- 16 *4.1.7.26*.
- 17 $(Z)-3-(4-\{4-[(5-fluoro-2-oxoindolin-3-ylidene)methyl]phenyl\}-1H-1,2,3-tr$
- 18 *iazol-1-yl)benzonitrile* (14*l*)
- 19 Yellow powder, yield: 63.43%, mp: 246°C. FI-IR (KBr, v_{max} cm⁻¹):
- 20 3430 (NH), 1679 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.68 (s, 1H,
- N-H), 9.53 (s, 1H, H-triazole), 8.54 (d, J = 7.9 Hz, 2H, Ar-H), 8.49 (s, 1H,
- 22 Ar-H), 8.35 (d, J = 8.3 Hz, 1H, Ar-H), 8.02 (t, J = 7.6 Hz, 3H, Ar-H), 7.92

1	(s, 1H, Ar-H), 7.87 (t, J = 8.0 Hz, 1H, H-vinylic), 7.67 (d, J = 8.8 Hz, 1H,
2	Ar-H), 7.05 (t, <i>J</i> = 9.0 Hz, 1H, Ar-H), 6.81 (dd, <i>J</i> = 8.3, 4.3 Hz, 1H, Ar-H).
3	¹³ C NMR (126 MHz, DMSO- d_6) δ 166.99 (C=O), 158.75, 156.88, 146.85
4	(C-triazole), 137.52, 136.83 (C-vinylic), 133.61, 132.79 (2C), 132.23,
5	131.68, 131.25 (C-triazole), 130.27, 126.52 (d, $J = 3.2$ Hz,
6	C-fluorobenzene), 124.84 (2C), 124.55, 123.31, 120.62 (C-vinylic),
7	117.64, 115.05 (d, J = 24.3 Hz, C-fluorobenzene), 112.71, 109.98 (d, J =
8	8.3 Hz, C-fluorobenzene), 107.16 (d, $J = 25.4$ Hz, C-fluorobenzene).
9	ESI-HRMS calcd for $C_{24}H_{15}FN_5O [M+H]^+ 408.1182$, found: 408.1265.
10	4.1.7.27.

11 $(Z/E)-4-(4-\{4-[(5-fluoro-2-oxoindolin-3-ylidene)methyl]phenyl\}-1H-1,2,3$

12 *-triazol-1-yl)benzonitrile* (**14m**)

Yellow powder, yield: 59.91%, mp: 215°C. FI-IR (KBr, v_{max} cm⁻¹): 13 3432 (NH), 1684 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.68 (s, 1H, 14 N-H), 9.10 (s, 1H, H-triazole), 8.54 (d, J = 8.2 Hz, 1H, Ar-H), 8.12 (d, J =15 8.0 Hz, 1H, Ar-H), 8.06 (d, J = 8.2 Hz, 1H, Ar-H), 7.93 (s, 0.4H, Ar-H), 16 7.85 (d, J = 7.9 Hz, 1H, H-vinylic), 7.74 (s, 0.6H, Ar-H), 7.67 (d, J = 8.817 Hz, 0.4H, Ar-H), 7.56 – 7.50 (m, 3H, Ar-H), 7.46 (s, 1H, Ar-H), 7.32 (d, J 18 = 9.2 Hz, 0.6H, Ar-H), 7.11 (t, J = 8.8 Hz, 0.6H, Ar-H), 7.05 (t, J = 8.919 Hz, 0.4H, Ar-H), 6.88 (dd, J = 8.3, 4.6 Hz, 0.6H, Ar-H), 6.81 (dd, J = 8.3, 20 4.3 Hz, 0.4H, Ar-H). ESI-HRMS calcd for C₂₄H₁₅FN₅O [M+H]⁺ 408.1182, 21 found: 408.1259. 22

4.1.7.28. 1

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(Z)-3- $(4-{1-[4-chloro-3-(trifluoromethyl)phenyl]-1H-1,2,3-triazol-4-yl}be$ 2 nzylidene)-5-fluoroindolin-2-one (14n) 3

Orange powder, yield: 47.50%, mp: 251°C. FI-IR (KBr, v_{max} cm⁻¹): 4 3432 (NH), 1684 (C=O). ¹H NMR (500 MHz, DMSO- d_6) δ 10.66 (s, 1H, 5 N-H), 9.58 (s, 1H, H-triazole), 8.52 (d, J = 7.5 Hz, 2H, Ar-H), 8.41 (s, 1H, 6 Ar-H), 8.31 (d, J = 8.7 Hz, 1H, Ar-H), 8.01 (d, J = 6.9 Hz, 3H, Ar-H), 7 7.89 (s, 1H, H-vinylic), 7.64 (d, J = 8.7 Hz, 1H, Ar-H), 7.03 (t, J = 8.8 Hz, 8 1H, Ar-H), 6.80 (s, 1H, Ar-H). ¹³C NMR (126 MHz, DMSO- d_6) δ 166.96 9 (C=O), 158.73, 156.86, 146.91 (C-triazole), 137.48, 136.87 (C-vinylic), 10 135.35, 133.60, 133.01 (d, J = 64.2 Hz, 2C, C- trifluoromethylphenyl), 11 12 131.61 (C-triazole), 130.30, 127.81 (d, J= 31.7 Hz, Ctrifluoromethylphenyl), 126.49 (d, J = 3.0 Hz, C-trifluoromethylphenyl), 13 126.26 (d, J = 8.8 Hz, C-trifluoromethylphenyl), 124.94 (d, J = 29.6 Hz, 14 2C, C-fluorobenzene, C- trifluoromethylphenyl), 123.23, 121.06, 120.73 15 (C-vinylic), 119.15 (d, J = 5.5 Hz, C-fluorobenzene), 115.03 (d, J = 24.016 Hz, C-fluorobenzene), 109.96 (d, J = 8.4 Hz, C-fluorobenzene), 107.13 (d, 17 J = 25.1 Hz, C-fluorobenzene). ESI-HRMS calcd for $C_{24}H_{14}ClF_4N_4O$ 18 [M+H]⁺ 485.0787, found: 485.0793. 19 4.2. VEGFR-2 kinase assay in vitro

The ADP-Glo kinase assay kit (Promega, Madison) was used for 21 VEGFR-2 kinase analysis. According to manufacturer's instructions, The 22

general procedure is as follows: 20 µL mix VEGFR-2 kinase (Invitrogen, 1 USA), 30 µL substrate (ADP-Glo, 50 µM), 30 µL ATP (50 µM) and 20 2 µL different concentrations (0 nM, 25 nM, 50 nM, 100 nM, 200 nM) of 3 test compounds, and the total volume in a 96-well light-proof microtiter 4 plate are 100µL of final buffer Incubate in liquid. Wells containing 5 substrate and compound-free kinase were used as overall reaction 6 controls. The assay plate was incubated at 37°C for 30 min in the dark. 7 Use a full-wavelength microplate reader to perform detection at dual 8 wavelengths to obtain fluorescence values, which are further used to 9 calculate IC_{50} values. The obtained data were compared with sunitinib as a 10 standard inhibitor for VEGFR-2. 11

12 4.3. Cell culture and CCK-8 assay in vitro

Human colon cancer (HT-29) cells, human gastric cancer (MKN-45) 13 cells, human umbilical vein endothelial cells (HUVECs) were purchased 14 from the China Cell Resource Bank and recorded as F0 generation cells. 15 Standardized training according to the protocol provided by the supplier 16 dilute fibronectin to 1 mg/mL with sterile water. Add 25 mL Fetal Bovine 17 Serum (FBS), 5 mL Endothelial Cell Growth Supplement (ECGS), and 5 18 mL P/S solution to 500 mL Endothelial Cell Medium (ECM) (ScienCell, 19 SC-1001). These operations are protected from light. All compounds were 20 dissolved in DMSO as a 1 mM stock solution. Further dilutions of all 21 compounds were performed in DMEM (GIBCO, 21127-022) incomplete 22

medium. The compound was serially diluted to the final concentration:
1.25 μM, 2.5 μM, 5 μM, 10 μM, 20 μM. The concentration of DMSO is
0.5% [47].

The antiproliferative activity was determined using the CCK-8 assay 4 [48]. HT-29, MKN-45, and HUVECs of F4 generation were cultured in 5 culture flasks. When the number of cells reached 80%, the cells were 6 harvested and transferred to a 96-well plate (treated with fibronectin at a 7 concentration of 2 mg/cm^2) and cultured at 3000 cells per well overnight. 8 ECM complete medium was aspirated in DMEM complete medium, and 9 50 ng/mL VEGF was added. 100 µL of the diluted compound was added 10 to the corresponding experimental wells. The mixture was incubated for 11 24 h, and 10 mL of CCK-8 reagent was added to each well and incubated 12 for 4h. The optical density (OD) of each well was detected at 490 nm. 13

14 4.4. Transwell assay

A transwell chamber (pore size 8 µm; Corning Costar, Cambridge, 15 Massachusetts, USA) was used to measure the migration capacity of 16 HUVEC. For transwell analysis, HUVECs were treated with serum-free 17 DMEM medium containing different concentrations of compound 13d (0 18 μ M, 0.2 μ M, 0.4 μ M or 0.8 μ M) for 24 hours, and then collected and 19 counted. 1×10^4 HUVECs were inoculated into the upper chamber, and the 20 lower chamber contained DMEM medium supplemented with 20% serum. 21 After 24 hours of incubation, the filter was fixed in methanol and stained 22

with 0.1% crystal violet. Gently wipe the upper surface of the filter,
image under the microscope, and count the total number of HUVECs that
have migrated through the filter's lower surface [49].

4 *4.5. Tube formation*

5 After treated with different concentrations (0 μ M, 0.2 μ M, 0.4 μ M or 6 0.8 μ M) of compound **13d**, the HUVECs were collected. 1×10⁴ cells 7 HUVECs were seeded on Matrigel's surface (#354234, Corning, USA), 8 then cultured for 6 hours. The tube formation was then observed and 9 photographed with a microscope. Use image pro plus software to 10 calculate the total length of tubes.

11 4.6 Western blot analysis

HUVECs were treated with different concentrations (0 μ M, 0.2 μ M, 0.4 μ M or 0.8 μ M) of compound **13d** for 24 hours. Primary antibody (100 μ L/cm2): VEGFR-2 (26415-1-AP, Proteintech Group), p-VEGFR-2 (Tyr951) (#2471, Cell Signaling Technology). The cellular level of protein was determined by standard western blotting.

17 4.7. Zebrafish labeling and culture

The research group cooperated with China Zebrafish Resource Center (CZRC) to construct a blood vessel marker zebrafish (Tg(kdrl:EGFP)) with a genotype of s843Tg/+, which was recorded as the F0 generation. Collect the fish eggs of the F2 generation, and add different concentrations of target compounds to the culture solution after 24 hours

to make the final concentration of the culture solution 40 mg/L. After 24
hours of action, the zebrafish were anesthetized *in vivo*, and the internode
angiogenesis and sprouting were observed under a confocal microscope.

4 4.8. *Molecular docking study*

Molecular docking study was carried out with MOE (Molecular 5 Operating Environment, version 2016.08, Chemical Computing Group 6 Inc., Canada) [50]. Two mol2 format files of compound 13d and sunitinib 7 were subjected to energy minimization with Amber10: EHT force-field in 8 MOE. The crystal structure of VEGFR-2 in complex with sunitinib (PDB 9 ID: 4AGD) downloaded from Protein Data Bank was 10 and successively optimized with Structure (https://www.rcsb.org/) 11 12 Preparation and Protonate 3D [51]. The ligand atoms were chosen as the docking site in the docking procedure, which was conducted with default 13 parameters. 14

15 4.9. Molecular dynamics (MD) simulations

MD simulations of two systems in PDB format were conducted using NAMD software (version 2.14), and configuration files were generated in VMD (visual molecular dynamics) [52]. Energy minimization and equilibration with Gasteiger–Huckel charges used Boltzmann's initial velocity in CHARMM 22 force field file, which performed in a 15 Å3 size water box. Ultimately, 10 ns MD simulations for three proteins at constant temperature (300 K) and pressure (1 atm) were carried out to 1 analyze three systems' binding affinity and stability.

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15 References

[1] E. Perspicace, V. Jouan-Hureaux, R. Ragno, F. Ballante, S. Sartini, C
La Motta, F. Da Settimo, B Chen, G Kirsch, S. Hesse, Design, synthesis
and biological evaluation of new classes of thieno[3,2-d]pyrimidinone
and thieno[1,2,3]triazine as inhibitor of vascular endothelial growth factor
receptor-2 (VEGFR-2), Eur. J. Med. Chem. 63 (2013) 765-781.
[2] T. Jiang, J. Zhuang, H. Duan, Y. Luo, Q. Zeng, K. Fan, H. Yan, D. Lu,

22 Z. Ye, J. Hao, J. Feng, D. Yang, X. Yan, CD146 is a coreceptor for

1 VEGFR-2 in tumor angiogenesis, Blood. 120 (2012) 2330-2339.

[3] H. K. Mahmoud, T. A. Farghaly, H. G. Abdulwahab, N. T. Al-Qurashi,
M. R. Shaaban, Novel 2-indolinone thiazole hybrids as sunitinib
analogues: Design, synthesis, and potent VEGFR-2 inhibition with
potential anti-renal cancer activity, Eur. J. Med. Chem. 208 (2020)
112752.

[4] S. Sana, V. G. Reddy, S. Bhandari, T. S. Reddy, R. Tokala, A. P. Sakla,
S. K. Bhargava, N. Shankaraiah, Exploration of carbamide derived
pyrimidine-thioindole conjugates as potential VEGFR-2 inhibitors with
anti-angiogenesis effect, Eur. J. Med. Chem. 200 (2020) 112457.

- [5] P. M. Hoff, K. K. Machado, Role of angiogenesis in the pathogenesis
 of cancer, Cancer Treat. Rev. 38 (2012) 825–833.
- [6] P. Carmeliet, R. K. Jain, Molecular mechanisms and clinical
 applications of angiogenesis, Nature. 473 (2011) 298-307.
- 15 [7] C. Fontanella, E. Ongaro, S. Bolzonello, M. Guardascione, G. Fasola,
- G. Aprile, Clinical advances in the development of novel VEGFR-2
 inhibitors, Ann. Transl. Med. 2 (2014) 123-132.
- 18 [8] G. M. Saidel, Quantitative Relationships of Intravascular Tumor Cells,
- 19 Tumor Vessels, and Pulmonary Metastases following Tumor Implantation,
- 20 Cancer Res. 34 (1974) 997-1004.
- [9] J. Folkman, Angiogenesis: An organizing principle for drug
 discovery?, Nat. Rev. Drug Discov. 6 (2007) 273–286.

1	[10] D.	Hanahan,	R.	A.	Weinberg,	Hallmarks	of	cancer:	The	next
2	generatio	on, Cell. 14	4 (2	011) 646-674.					

3 [11] J. Q. Wu, R. Y. Fan, S. R. Zhang, C. Y. Li, L. Z. Shen, P. Wei, Z. H.

4 He, M. F. He, A systematical comparison of anti-angiogenesis and
5 anti-cancer efficacy of ramucirumab, apatinib, regorafenib and
6 cabozantinib in zebrafish model, Life Sci. 247 (2020) 117402.

[12] S. M. Chang, V Jain, T. L. Chen, A. S. Patel, H. B. Pidugu, Y. W. Lin,
M. H. Wu, J. R. Huang, H. C. Wu, A. Shah, T. L. Su, T. C. Lee, Design
and Synthesis of 1, 2-Bis (hydroxymethyl)pyrrolo[2,1- a]phthalazine
Hybrids as Potent Anticancer Agents that Inhibit Angiogenesis and
Induce DNA Interstrand Cross-links, J. Med. Chem, 62 (2019)
2404-2418.

[13] H. A. Mahdy, M. K. Ibrahim, A. M. Metwaly, A. Belal, A. B. M. 13 Mehany, K. M. A. El-Gamal, A. El-Sharkawy, M. A. Elhendawy, M. M. 14 Radwan, M. A. Elsohly, I. H. Eissa, Design, synthesis, molecular 15 modeling, in vivo studies and anticancer evaluation of 16 quinazolin-4(3H)-one derivatives as potential VEGFR-2 inhibitors and 17 apoptosis inducers. Bioorg Chem. 94 (2020) 103422. 18

[14] H. T. Abdel-Mohsen, M. A. Omar, A. M. El Kerdawy, A. E. E.
Mahmoud, M. M. Ali, H. I. El Diwani, Novel potent substituted
4-amino-2-thiopyrimidines as dual VEGFR-2 and BRAF kinase inhibitors,
Eur. J. Med. Chem. 179 (2019) 707–722.

[15] D. H. Dawood, E. S. Nossier, M. M. Ali, A. E. Mahmoud, Synthesis
and molecular docking study of new pyrazole derivatives as potent
anti-breast cancer agents targeting VEGFR-2 kinase, Bioorg. Chem. 101
(2020).
[16] L. Shi, J. Zhou, J. Wu, Y. Shen, X. Li, Anti-Angiogenesis Therapy:

6 Strategies to Develop Potent VEGFR-2 Tyrosine Kinase Inhibitors and
7 Future Prospect, Curr. Med. Chem. 23 (2016) 1000-1040.

8 [17] S. Faivre, G. Demetri, W. Sargent, E. Raymond, Molecular basis for
9 sunitinib efficacy and future clinical development, Nat. Rev. Drug Discov.
10 6 (2007) 734-745.

[18] F. Koinis, P. Corn, N. Parikh, J. Song, I. Vardaki, I. Mourkioti, S. H. 11 Lin, C. Logothetis, T. Panaretakis, G. Gallick, 12 Resistance to MET/VEGFR2 inhibition by cabozantinib is mediated by 13 YAP/TBX5-dependent induction of FGFR1 in castration-resistant 14 prostate cancer, Cancers (Basel). 12 (2020) 244. 15

[19] V. G. Reddy, T. S. Reddy, C. Jadala, M. S. Reddy, F. Sultana, R.
Akunuri, S. K. Bhargava, D. Wlodkowic, P. Srihari, A. Kamal,
Pyrazolo-benzothiazole hybrids: Synthesis, anticancer properties and
evaluation of antiangiogenesis activity using in vitro VEGFR-2 kinase
and in vivo transgenic zebrafish model, Eur. J. Med. Chem. 182 (2019)
111609.

22 [20] S. Mohamady, M. Galal, W. M. Eldehna, D. C. Gutierrez, H. S.

1	Ibrahim, M. M. Elmazar, H. I. Ali, Dual Targeting of VEGFR2 and C-Met
2	Kinases via the Design and Synthesis of Substituted
3	3-(Triazolo-thiadiazin-3-yl)indolin-2-one Derivatives as Angiogenesis
4	Inhibitors, ACS Omega. 5 (2020) 18872-18886.
5	[21] H. M. Roaiah, I. A. Y. Ghannam, I. H. Ali, A. M. El Kerdawy, M. M.
6	Ali, S. E. S. Abbas, S. S. El-Nakkady, Design, synthesis, and molecular
7	docking of novel indole scaffold-based VEGFR-2 inhibitors as targeted
8	anticancer agents, Arch. Pharm. 351 (2018) 1-17.
9	[22] B. Beuselinck, S. Oudard, O. Rixe, P. Wolter, A. Blesius, J. Ayllon, R.
10	Elaidi, P. Schöffski, E. Barrascout, A. Morel, B. Escudier, H. Lang, J.
11	Zucman-Rossi, J. Medioni, Negative impact of bone metastasis on
12	outcome in clear-cell renal cell carcinoma treated with sunitinib, Ann.
13	Oncol.22 (2011)794-800.
14	[23] P. C. Tang, Y. D. Su, J. Feng, J. H. Fu, J. L. Yang, L. Xiao, J. H. Peng,
15	Y. L. Li, L. Zhang, B. Hu, Y. Zhou, F. Q. Li, B. B. Fu, L. G. Lou, A. S.
16	Gong, G. H. She, W. H. Sun, X. T. Mong, Novel potent orally active
17	multitargeted receptor tyrosine kinase inhibitors: Synthesis,
18	structure-activity relationships, and antitumor activities of 2-indolinone

- 19 derivatives, J. Med. Chem. 53 (2010) 8140-8149.
- [24] H. E. Dweedar, H. Mahrous, H. S. Ibrahim, H. A. Abdel-Aziz,
 Analogue-based design, synthesis and biological evaluation of
 3-substituted-(methylenehydrazono)indolin-2-ones as anticancer agents,

1 Eur. J. Med. Chem. 78 (2014) 275-280.

[25] M. Qin, Y. Tian, X. Han, Q. Cao, S. Zheng, C. Liu, X. Wu, L. Liu, Y.
Meng, X. Wang, H. Zhang, Y. Hou, Structural modifications of
indolinones bearing a pyrrole moiety and discovery of a multi-kinase
inhibitor with potent antitumor activity, Bioorganic Med. Chem. 28 (2020)
115486.

7 [26] L. Zhang, Q. Zheng, Y. Yang, H. Zhou, X. Gong, S. Zhao, C. Fan,
8 Synthesis and in vivo SAR study of indolin-2-one-based multi-targeted
9 inhibitors as potential anticancer agents, Eur. J. Med. Chem. 82 (2014)
10 139-151.

[27] A. M. S. El-Sharief, Y. A. Ammar, A. Belal, M. A. M. S. El-Sharief,
Y. A. Mohamed, A. B. M. Mehany, G. A. M. Elhag Ali, A. Ragab, Design,
synthesis, molecular docking and biological activity evaluation of some
novel indole derivatives as potent anticancer active agents and apoptosis
inducers, Bioorg. Chem. 85 (2019) 399-412.

16 [28] J. Guo, F. Zhao, W. Yin, M. Zhu, C. Hao, Y. Pang, T. Wu, J. Wang, D.

17 Zhao, H. Li, M. Cheng, Design, synthesis, structure-activity relationships

- 18 study and X-ray crystallography of
 19 3-substituted-indolin-2-one-5-carboxamide derivatives as PAK4
 20 inhibitors, Eur. J. Med. Chem. 155 (2018) 197-209.
- [29] D. Wu, A. Pusuluri, D. Vogus, V. Krishnan, C. W. Shields, J. Kim, A.
- 22 Razmi, S. Mitragotri, Design principles of drug combinations for

- 1 chemotherapy, J. Control. Release. 323 (2020) 36-46.
- 2 [30] J. J. Yang, W. W. Yu, L. L. Hu, W. J. Liu, X. H. Lin, W. Wang, Q.

3 Zhang, P. L. Wang, S. W. Tang, X. Wang, M. Liu, W. Lu, H. K. Zhang,

- 4 Discovery and Characterization of 1 H-1,2,3-Triazole Derivatives as
- 5 Novel Prostanoid EP4 Receptor Antagonists for Cancer Immunotherapy, J.
- 6 Med. Chem. 63 (2020) 569–590.
- 7 [31] M. Taddei, S. Ferrini, L. Giannotti, M. Corsi, F. Manetti, G. Giannini,
- 8 L. Vesci, F. M. Milazzo, D. Alloatti, M. B. Guglielmi, M. Castorina, M. L.
- 9 Cervoni, M. Barbarino, R. Foderà, V. Carollo, C. Pisano, S. Armaroli, W.
 10 Cabri, Synthesis and evaluation of new Hsp90 inhibitors based on a
- 1,4,5-trisubstituted 1,2,3-triazole scaffold, J. Med. Chem. 57 (2014)
 2258–2274.
- [32] Z. Xu, S. J. Zhao, Y. Liu, 1,2,3-Triazole-containing hybrids as
 potential anticancer agents: Current developments, action mechanisms
 and structure-activity relationships, Eur. J. Med. Chem. 183 (2019)
 111700.
- [33] S. Li, X. Y. Li, T. J. Zhang, M. O. Kamara, J. W. Liang, J. Zhu, F. H.
 Meng, Design, synthesis and biological evaluation of homoerythrina
 alkaloid derivatives bearing a triazole moiety as PARP-1 inhibitors and as
 potential antitumor drugs, Bioorg. Chem. 94 (2020) 103385.
- 21 [34] G. Q. Lu, X. Y. Li, M. O. Kamara, D. Wang, F. hao Meng, Design,
- 22 synthesis and biological evaluation of novel uracil derivatives bearing

1,2,3-triazole moiety as thymidylate synthase (TS) inhibitors and as
 potential antitumor drugs, Eur. J. Med. Chem. 171 (2019) 282-296.
 [35] S. Li, X. Y. Li, T. J. Zhang, J. Zhu, W. H. Xue, X. H. Qian, F. H.
 Meng, Design, synthesis and biological evaluation of erythrina
 derivatives bearing a 1,2,3-triazole moiety as PARP-1 inhibitors, Bioorg.
 Chem. 96 (2020) 103575.
 [36] A. Kamal, S. Prabhakar, M. J. Ramaiah, P. V. Reddy, C. R. Reddy, A.

Mallareddy, N. Shankaraiah, T. L. N. Reddy, S. N. C. V. L. Pushpavalli, 8 Synthesis M. Pal-Bhadra, and anticancer activity of 9 chalcone-pyrrolobenzodiazepine conjugates linked via 1,2,3-triazole ring 10 side-armed with alkane spacers, Eur. J. Med. Chem. 46 (2011) 3820-11 12 3831.

[37] Q. Zhang, Y. Teng, Y. Yuan, T. Ruan, Q. Wang, X. Gao, Y. Zhou, K.
Han, P. Yu, K. Lu, Synthesis and cytotoxic studies of novel 5-phenylisatin
derivatives and their anti-migration and anti-angiogenesis evaluation, Eur.
J. Med. Chem. 156 (2018) 800–814.

[38] N. Kaila, K. Janz, A. Huang, A. Moretto, S. Debernardo, P. W. 17 Bedard. S. Tam, V. Clerin, J. C Keith, D. H. H. Tsao, 18 2-(4-Chlorobenzyl)-3-hydroxy-7,8,9,10-tetrahydrobenzo[H]quinoline-4-c 19 arboxylic acid (PSI-697): identification of a clinical candidate from the 20 quinoline salicylic acid series of P-selectin antagonists, J. Med. Chem. 50 21 (2007) 40-64. 22

1 [39] N. A. Lozinskaya, D. A. Babkov, E. V. Zaryanova, E. N. Bezsonova,

2 A. M. Efremov, M. D. Tsymlyakov, L. V. Anikina, O. Y. Zakharyascheva,

A. V. Borisov, V. N. Perfilova, I. N. Tyurenkov, M. V. Proskurnina, A. A.
Spasov, Synthesis and biological evaluation of 3-substituted 2-oxindole
derivatives as new glycogen synthase kinase 3β inhibitors, Bioorganic
Med. Chem. 27 (2019) 1804–1817.

[40] Y. Z. Zhou, Y. Ju, Y. Yang, Z. Sang, Z. Wang, G. He, T. Yang, Y. Luo,
Discovery of hybrids of indolin-2-one and nitroimidazole as potent
inhibitors against drug-resistant bacteria, J. Antibiot. 71 (2018) 887–897.

[41] W. D. Long, W. L. Ying, W. Y. Ling, S. Shuang, F. J. Tao, Z. Xing,
Natural α-methylenelactam analogues: Design, synthesis and evaluation
of α-alkenyl-γ and δ-lactams as potential antifungal agents against
Colletotrichum orbiculare, Eur. J. Med. Chem. 130 (2017) 286–307.

[42] M. E. Matheus, F. de A. Violante, S. J. Garden, A. C. Pinto, P. D.
Fernandes, Isatins inhibit cyclooxygenase-2 and inducible nitric oxide
synthase in a mouse macrophage cell line, Eur. J. Pharmacol. 556 (2007)
200–206.

[43] B. R. Dinesh, A. R. Baba, K. U. Sankar, D. C. Gowda, Synthesis of
indolones and quinolones by reductive cyclisation of o-nitroaryl acids
using zinc dust and ammonium formate, J. Chem. Res. (2008) 287–288.

21 [44] S. Shah, C. Lee, H. Choi, J. Gautam, H. Jang, G. J. Kim, Y. J. Lee, C.

22 L. Chaudhary, S.W. Park, T. G. Nam, J. A. Kim, B. S. Jeong,

5-Hydroxy-7-azaindolin-2-one, a novel hybrid of pyridinol and sunitinib:
 Design, synthesis and cytotoxicity against cancer cells, Org. Biomol.
 Chem. 14 (2016) 4829–4841.

[45] L. Sun, N. Tran, F. Tang, H. App, P. Hirth, G. McMahon, C. Tang,
Synthesis and biological evaluations of 3-substituted indolin-2-ones: A
novel class of tyrosine kinase inhibitors that exhibit selectivity toward
particular receptor tyrosine kinases, J. Med. Chem. 41 (1998) 2588–2603.
[46] H. Yang, C. Lou, L. Sun, J. Li, Y. Cai, Z. Wang, W. Li, G. Liu, Y.

9 Tang, AdmetSAR 2.0: Web-service for prediction and optimization of

10 chemical ADMET properties, Bioinformatics. 35 (2019) 1067–1069.

11 [47] X. yang Li, S. Li, G. qing Lu, D. pu Wang, K. li Liu, X. hua Qian, W.

han Xue, F. hao Meng, Design, synthesis and biological evaluation of
novel (E)-N-phenyl-4-(pyridine-acylhydrazone) benzamide derivatives as
potential antitumor agents for the treatment of multiple myeloma (MM),
Bioorg. Chem. 103 (2020) 104189.

[48] Z. Gao, M. Shi, Y. Wang, J. Chen, Y. Ou, Apatinib enhanced
anti-tumor activity of cisplatin on triple-negative breast cancer through
inhibition of VEGFR-2, Pathol. Res. Pract. 215 (2019) 152422.

[49] X. Y. Li, T. J. Zhang, M. O. Kamara, G. Q. Lu, H. L. Xu, D. P. Wang,
F. H. Meng, Discovery of
N-phenyl-(2,4-dihydroxypyrimidine-5-sulfonamido) phenylurea-based
thymidylate synthase (TS) inhibitor as a novel multi-effects antitumor

1 drugs with minimal toxicity, Cell Death Dis. 10 (2019) 532.

[50] J. L. Velázquez-Libera, J. Caballero, J. A. Murillo-López, A. F. de la
Torre, Structural requirements of n-alpha-mercaptoacetyl dipeptide
(Namdp) inhibitors of pseudomonas aeruginosa virulence factor lasb:
3Q-QSAR, molecular docking, and interaction fingerprint studies, Int. J.
Mol. Sci. 20 (2019) 6133.

[51] M. McTigue, B. W. Murray, J. H. Chen, Y. L. Deng, J. Solowiej, R. S.
Kania, Molecular conformations, interactions, and properties associated
with drug efficiency and clinical performance among VEGFR TK
inhibitors, P. Natl. Acad. Sci. USA. 109 (2012) 18281–18289.

[52] T. J Zhang, Y. Zhang, S. Tu, Y. H. Wu, Z. H. Zhang, F. H. Meng, 11 and 12 Design, synthesis biological evaluation of N-(3-(1H-tetrazol-1-yl)phenyl)isonicotinamide derivatives as novel 13 xanthine oxidase inhibitors, Eur. J. Med. Chem. 183 (2019) 111717. 14

- A novel VEGFR-2 inhibitor based on 1,2,3-triazole scaffold was identified.
- Compound 13d had better kinase activity inhibition ability than sunitinib.
- Compound **13d** could inhibit angiogenesis more effectively than sunitinib.

Conflict of Interest

The authors declared that they have no conflicts of interest in this work.

We declared that we do not have any commercial or associative interest that represents a conflict of interest in connection with the work submitted.

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