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Synthesis and *in vitro* antimicrobial, antioxidant, and antiproliferative activities of some new pyrano[2,3-c]pyrazoles containing 1,2-azaphospholes, 1,3,2-diazaphosphinines and phosphonate moieties

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ABSTRACT

New 1,2-azaphosphole, 1,3,2-diazaphosphinine, and phosphonate derivatives containing pyrano[2,3-c]pyrazole moiety were achieved. The synthetic pathways depended on the reaction of 6-amino-3-methyl-1,4-diphenyl-1,4-dihydropyrano[2,3-c]pyrazole-5-carbonitrile with some phosphorus reagents such as diethyl phosphite, phosphorus sulfides, and phenyl phosphorus halides under different conditions. Compounds 6 and 7 recorded potent antibacterial and antifungal activities with lower toxicity, while compounds 4 and 6 displayed promising antioxidative properties. Further, compounds 4 and 9 exhibited potent cytotoxic effects against MCF-7, HepG-2, and HCT-116 cancer cells. The early apoptotic cell death was observed by the compounds in all types of the treated cells. Compounds 3, 5, 7, 10, and 14 recorded low to moderate percentages of necrosis and late apoptosis toward all treated cells.

GRAPHICAL ABSTRACT

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Introduction

Organophosphorus compounds are important intermediates in organic synthesis and have been widely used as a pharmaceutical, [1] agricultural, [2] and chemical agents. [3,4] For example, phosphorus heterocycles^[5] have received considerable interest because of their unique biological activities as antimicrobial^[6] and their anticancer effects.^[7,8] Ifosfamide and cyclophosphamide are two important examples of phosphorus heterocycles that were launched on the market more than 30 years ago and are still used in the treatment of cancer. [9,10] Also, phosphonate groups are chemically stable analogs of phosphates and play numerous vital roles in the biochemistry of living organisms. Phosphonates constitute an important class of bioisosteres for medicinal chemists and chemical biologists. [11] The usefulness of this class of molecules has been highlighted by the COVID-19 pandemic. Phosphonate nucleotide analogs, such as tenofovir and cidofovir, can inhibit SARS-CoV-2 RNA polymerase^[12] and may thus represent a much-needed therapeutic weapon against this viral threat. On the other hand, pyrano[2,3-c]pyrazole compounds are the major structural motif in several natural products and synthetic compounds of widely recognized pharmacological properties.^[13] In this context, pyranopyrazole systems have well been recognized for their potent biological activities including antimicrobial, [14] antioxidant, [15] antiviral [16], and antitumor. [17]

In view of the aforementioned details, and in extension to our efforts concerning the design of new polyheterocycles containing phosphorus heterocycles or phosphonate groups with possible biological properties, [18-22] we herein display the synthesis of some novel bioactive polycyclic compounds composed of pyrano[2,3-c]pyrazole skeleton containing phosphorus heterocycles or phosphonate groups in one molecular frame which might show enhanced biological activity. The suggested synthetic pathways depend on the reaction of the 6-amino-3-methyl-1,4-diphenyl-1,4-dihydropyrano[2,3-c]pyrazole-5-carbonitrile with some phosphorus reagents such as diethyl phosphite (in the presence or absence of electrophilic reagents), phosphorus sulfides, and phosphorus halides under different reaction conditions. In addition, the isolated products were screened for their antimicrobial, antioxidant, and antiproliferative activities.

Results and discussion

The known starting material, 6-amino-3-methyl-1,4-diphenyl-1,4-dihydropyrano[2,3-c] pyrazole-5-carbonitrile (3) was synthesized in good yield as reported in the literature^[23] through the reaction of 3-methyl-1-phenyl-5-pyrazolone (1) with 2-benzylidenemalononitrile (2) in distilled water containing a catalytic amount of sodium benzoate (Scheme 1). Substrate 3 possesses one reactive electrophilic center (C≡N) and another nucleophilic center (NH₂) which can be attacked by electrophilic and nucleophilic phosphorus reagents. This provides a wide opportunity for the construction of phosphorus heterocycles fused with pyranopyrazole system.

Phosphorus esters have gained significant interest as versatile reagents in the construction of various novel phosphorus heterocycles. [19,22] Thus, diethyl H-phosphonate reacted with substrate 3 at 80-90 °C in the presence of BF₃·Et₂O as a catalyst to yield the nonisolable intermediate A via removal of the ethanol molecule. The latter

Scheme 1. The reaction of substrate **3** with diethyl *H*-phosphonate.

intermediate underwent an intramolecular heterocyclization through addition of H-phosphorus atom on the nitrile group to give 6-ethoxy- 5-imino-3-methyl-6-oxido-1,4-diphenyl-1,4,5,7-tetrahydropyrazolo[4′,3′:5,6]pyrano[3,2-d][1,2]azaphosphole (4) (Scheme 1). The M⁺ peak for product 4 was recorded at m/z 420 in its mass spectrum, while its ³¹P-NMR spectrum recorded a singlet at δ 16.2 ppm. Moreover, the ¹H-NMR spectrum of compound 4 revealed specific one ethoxy group at δ 1.03 (t, CH₃) and 3.79–3.87 (m, OCH₂) as well as the NH protons resonated as two singlets at δ 9.94 and 11.52 ppm. Furthermore, its ¹³C-NMR spectrum exhibited some specific carbon atoms at δ 16.1 (CH₃), 60.2 (OCH₂), and a doublet signal at δ 142.6 ppm (J_{PC} = 150.9 Hz) for P-C=NH.^[24]

Dimethylformamide dimethylacetal (DMFDMA) is considered one of the very important precursors in the construction of novel bioactive compounds. Thus, heating of substrate 3 with DMFDMA in dry dioxane under reflux for 6 h, followed by addition of diethyl phosphite and BF₃·Et₂O then further heated under reflux for 10 h afforded diethyl{[(5-cyano-3-methyl-1,4-diphenyl-1,4-dihydropyrano[2,3-c]pyrazol-6-yl)imino]methyl}phosphonate (5) in 30% yield (Scheme 2). When the previous reaction was carried out using a two-fold excess of diethyl phosphite and a catalytic amount of sodium metal, it furnished the interesting novel pyrrolo[3',2': 5,6]pyrano[2,3-c]pyrazolyl bis(phosphonate) 6 as a sole product in 47% yield (Scheme 2). The formation of products 5 and 6 was explained through condensation of NH₂ group of substrate 3 with the acetal carbon of the DMFDMA reagent giving the nonisolable *N,N*-dimethylformimidamide intermediate B. The latter intermediate reacted with an equimolar amount of diethyl phosphite in the presence of BF₃·Et₂O or Na metal via a nucleophilic addition of *H*-phosphorus atom at azomethine group to give the nonisolable

i. HP(O)(OEt)2, BF3·Et2O, 80-90 °C ii. HP(O)(OEt)2, Na, 80-90 °C

Scheme 2. The reaction of substrate 3 with DMfDMA followed by diethyl phosphite.

α-aminophosphonate C (Scheme 2). In the case of pathway i, the intermediate C lost one molecule of dimethylamine to yield the isolated product 5, while in case of pathway ii, condensation of the second molecule of diethyl phosphite with the intermediate C gave the bis(phosphonate) intermediate D with the elimination of dimethylamine molecule (Scheme 2). This intermediate D underwent ring closure via the addition of the active CH-P group at the electrophilic C \equiv N function to isolate the final product 6 (Scheme 2). The IR spectrum of product 5 displayed important absorption bands at 2195 (C \equiv N) and 1200 (P \equiv O) cm $^{-1}$, while its mass spectrum exhibited the M $^+$ peak at m/z 476 (27%). The 13 C-NMR spectrum of product 5 exhibited a specific signal relevant to C \equiv N at δ 111.8 ppm and a doublet signal at δ 143.9 ppm (J=140.5 Hz) attributable to CH-P. Also, its 1 H-NMR spectrum affirmed the proposed structure by recording two ethoxy groups and the specific N=CH-P proton as a doublet at δ 8.58 with coupling

Scheme 3. The reaction of substrate 3 with triethyl orthoformate followed by diethyl phosphite.

constant (J=22 Hz). On the other hand, the 1 H-NMR spectrum of compound **6** exhibited four ethoxy groups as two multiplets at δ 0.86–1.31 (4 CH₃) and 3.87–4.15 (4 OCH₂) ppm, besides two broad signals at δ 11.85 and 13.97 (2 NH) ppm. Also, its mass spectrum displayed the expected M⁺ peak at m/z 614 (24%). Additionally, its 13 C-NMR spectrum revealed the specific carbon atom of C–P at δ 53.6 ppm as a doublet signal (J_{PC} =142.6 Hz) and another signal relative to C=NH at δ 144.8 ppm.

Likewise, treatment of substrate **3** with triethyl orthoformate in dry dioxane followed by addition of a solution of diethyl phosphite containing a catalytic amount of sodium metal yielded diethyl {6-ethoxy-5-imino-3-methyl-1,4-diphenyl-4,5,6,7-tetrahydro-1*H*-pyrrolo[3',2':5,6]pyrano[2,3-*c*]pyrazol-6-yl}phosphonate (7) and not the expected product **5** (Scheme **3**). The formation of compound **7** is similar to the formation of compound **6**. The IR spectrum of product **7** recorded the presence of the important bands of two NH and exocyclic C=N functions at 3425, 3363, and 1624 cm⁻¹, respectively.

Scheme 4. The reaction of substrate 3 with benzaldehyde followed by diethyl phosphite.

Moreover, its $^1\text{H-NMR}$ spectrum recorded the chemical shifts of two NH protons as two broad signals at δ 11.82 and 13.98 ppm. In addition, its $^{13}\text{C-NMR}$ spectrum exhibited a doublet signal of C–P at δ 67.0 ppm ($J_{PC}=144.6\,\text{Hz}$), while the carbon atom of C=NH was resonated at δ 146.3 ppm.

In the same manner, treatment of substrate 3 with benzaldehyde and diethyl phosphite in the presence of BF₃·Et₂O as a catalyst in dry dioxane under the Kabachnik-Field reaction^[28] afforded the nonisolable α -amniophosphonate **H**. The latter intermediate underwent an intramolecular cyclization via addition of the active hydrogen of CH–P group at the nitrile function followed by auto-hydrolysis to isolate diethyl {3-methyl-5-oxo-1,4,6-triphenyl-4,5,6,7-tetrahydro-1*H*-pyrrolo[3',2':5,6]pyrano[2,3-*c*]pyrazol-6-yl}phosphonate (8) (Scheme 4). The IR spectrum of product 8 showed absorption bands for NH and C = O functions at 3422 and 1691 cm⁻¹, respectively while its ³¹P-NMR spectrum recorded a singlet at δ 22.0 ppm. Furthermore, its M⁺ peak was

Scheme 5. The reaction of substrate 3 with P_4S_{10} in dioxane.

recorded at m/z 555 (12%) in its mass spectrum. The ¹H-NMR spectrum of the cyclic aminophosphonate **8** recorded additional multiplets for aromatic protons produced from benzaldehyde reagent when compared with the starting material **3**, besides a singlet relevant to NH proton at δ 11.25 ppm. In its ¹³C-NMR spectrum, the carbon atom of C-P moiety resonated as a doublet at δ 68.4 ppm with a coupling constant 143.4 Hz while the C=O resonated as a singlet at δ 165.4 ppm.

Lawesson's reagent and phosphorus decasulfide are convenient thionating agents and powerful platforms to construct novel compounds including bioactive phosphorus and sulfur-based heterocycles. Thus, treatment of substrate 3 with phosphorus decasulfide in dry dioxane afforded 2-sulfanyl-6-methyl-5,8-diphenyl-2-sulfido-4-thioxo-3,5,8-trihydro-pyrazolo[4',3':5,6]pyrano[2,3-d][1,3,2]diazaphosphinine (9) (Scheme 5). The isolated product 9 was formed through the addition of P_4S_{10} reagent on the C \equiv N group to give the nonisolable thioamide intermediate J. The latter intermediate underwent nucleophilic attack of the neighboring NH₂ group at the electrophilic phosphorus atom of the PS₂ fragment, followed by an intramolecular cyclization (Scheme 5). The IR spectrum of compound 9 showed the disappearance of C \equiv N group and displayed broad absorption bands at 3442, 3349 (two NH), 2664 (SH), and 1195 (C \equiv S) cm $^{-1}$. Moreover, its 1 H-NMR spectrum showed a singlet for SH proton at δ 2.40 ppm and two broad signals at δ 10.02 and 11.86 ppm for two NH protons. Further, the 13 C-NMR spectrum of product 9 displayed the carbon atom of C \equiv S at δ 178.4 ppm. In addition, its mass

Scheme 6. The reaction of substrate 3 with P_4S_{10} in ethanol.

spectrum recorded the expected M^+ at m/z 456 which confirmed the suggested structure.

In the same manner, when compound 3 was reacted with P_4S_{10} in absolute ethanol, it furnished the fused triheterocyclic system 10 in 55% yield (Scheme 6). The mass spectrum was great evidence for the formation of compound 10 which recorded its M^+ at m/z 468. Its H-NMR spectrum revealed one ethoxy group at $\delta 1.01$ (t, CH₃) and 3.41 (q, OCH₂) ppm. Furthermore, its ¹³C-NMR spectrum displayed the specific carbon atoms of CH₃, OCH₂, and C=S at δ 17.2, 62.2, and 177.1 ppm, respectively. Besides, the ³¹P-NMR spectrum recorded a singlet at δ 45.2 ppm. The proposed pathway suggested that the chemically active O_iO_i -diethyl dithiophosphoric acid intermediate (formed in *situ* by refluxing P_4S_{10} in absolute ethanol) underwent a nucleophilic attack of SH group at the C=N group of substrate 3 forming the intermediate L. The latter intermediate removed ethanol molecule through an intramolecular cyclization followed by *Dimroth* rearrangement to produce the desired product 10 (Scheme 6).

On the other hand, treatment of compound 3 with Lawesson's reagent (LR) in dry dioxane gave the nonisolable intermediate N which underwent ring closure through a nucleophilic attack of SH group at phosphorus atom of -SP(S)Ar fragment to form the nonisolable intermediate O. *Dimroth* rearrangement of the latter intermediate led to the formation of 2-(4-methoxyphenyl)-6-methyl-5,8-diphenyl-2-sulfido-4-thioxo-3,5,8-trihydropyrazolo[4',3':5,6]pyrano[2,3-d][1,3,2]diazaphosphinine (11) (Scheme 7). The ³¹P-NMR spectrum of product 11 displayed a signal at δ 50.1 ppm. Also, its ¹³C-NMR spectrum showed specific signals at δ 54.4 and 175.9 ppm relative to MeO and C=S groups, respectively. In addition, the ¹H-NMR spectrum recorded a singlet at δ 3.86 ppm relevant to MeO protons and two broad signals at δ 11.56 and 13.16 ppm for two NH protons. The mass and IR spectra of product 11 were in accordance with the suggested structure (see experimental section).

Scheme 7. The reaction of substrate 3 with LR in dioxane.

Furthermore, compound 3 reacted with P-(4-methoxyphenyl)-N,N'-bis(4-methylphenyl) phosphonothioic diamide P (formed by the reaction of LR with p-toluidine in molar ratio 1:4)^[34] to produce the nonisolable intermediate Q with the removal of p-toluidine molecule. Ring closure of the latter intermediate by attacking of NH group at the $C\equiv N$ group led to the sole product 12 in 33% yield (Scheme 8). The IR spectrum of product 12 showed characteristic absorption bands due to two NH and P=S functions at 3289, 3191, and $798\,\mathrm{cm}^{-1}$, respectively. A strong confirmation of the structure 12 was its 1 H-NMR spectrum which exhibited three singlets at δ 2.02, 2.31, and 3.89 ppm relevant to two Me and one MeO groups. Also, other signals appeared at δ 10.36 and 11.19 ppm for the two NH protons. In addition, its 13 C-NMR spectrum displayed characteristic signals for carbon atoms of 2 Me, MeO, and C=NH at δ 12.1, 13.9, 55.0, and 154.9 ppm, respectively. The mass spectrum of product 12 recorded m/z 496 (M^+ -MeOC₄H₆, 12%).

Phenyl phosphorus halides play vital roles in the construction of bioactive organophosphorus compounds. This encouraged us to study the reactivity of substrate 3 toward some phenyl phosphorus halides. Thus, compound 3 condensed with P,P-dichloro(phenyl)phosphine in dry pyridine to furnish 3-methyl-1,4-diphenyl-6-[(phenyl-phosphanylidene)amino]-1,4-dihydropyrano[2,3-c]pyrazole-5-carbonitrile (13) (Scheme 9). The IR spectrum of the latter product showed a characteristic band at 2223 cm⁻¹ relevant to the C \equiv N function confirming the suggested structure. In addition, the carbon atom of C \equiv N resonated at δ 107.7 in its 13 C-NMR spectrum. The mass spectrum of compound 13 exhibited its M⁺ peak at m/z 434 (6%).

Next, P-chlorodiphenylphosphine condensed with compound 3 in dry pyridine to give the nonisolable intermediate S. The latter intermediate underwent ring closure through the addition of the nucleophilic phosphorus atom at the electrophilic carbon atom of $C\equiv N$ group giving the intermediate T which was rearranged into 5-imino-3-

Scheme 8. The reaction of substrate 3 with the intermediate P (formed in situ from LR and *p*-toluidine).

Scheme 9. The reaction of substrate **3** with PhPCl₂ in pyridine.

Scheme 10. The reaction of substrate 3 with (Ph)₂PCl in pyridine.

methyl-1,4,6,6-tetraphenyl-1,4-dihydro-5H-6 λ^5 -pyrazolo[4',3':5,6]pyrano[3,2-d][1,2]aza-phosphole (**14**) (Scheme 10). The IR spectrum of product **14** displayed absorption bands for NH and exocyclic C=N functions at 3354 and 1603 cm⁻¹, respectively. Its 1 H-NMR spectrum displayed a singlet relevant to NH proton at δ 13.01 ppm, beside multiplet signals for twenty aromatic protons at δ 6.98–7.62 ppm. In addition, the 13 C-NMR spectrum revealed the existence of a doublet at δ 150.5 ppm for C=NH due to coupling with phosphorus atom with J_{PC} =60 Hz.

Finally, addition of phenyl phosphonic dichloride to substrate 3 in dry pyridine furnished 2,5,8-triphenyl-3,5,8-trihydropyrazolo[4',3':5,6]pyrano[2,3-d][1,3,2]diazaphosphinine 15 in 72% yield (Scheme 11). The suggested pathway for this product based on condensation between compound 3 and phosphorus reagent to give the nonisolable intermediate U. The nucleophilic addition of chlorine atom at C \equiv N group in the latter intermediate afforded the 4-chloro-1,3,2-diazaphosphinine intermediate V. When the intermediate V was treated with ice-water for 30 minutes, it gave the final product 15 in the form of 15A and not the other tautomeric form 15B according to its spectroscopic data (Scheme 11). The IR spectrum confirmed the suggested structure and showed characteristic absorption bands at 3425, 3363, and 1679 cm $^{-1}$ for two NH and C=O functions, respectively. The expected M $^+$ peak of compound 15 was recorded at m/z 468 in its mass spectrum while its 31 P-NMR spectrum showed a singlet at δ 34.31 ppm. The two NH protons resonated as two singlets at δ 12.68 and 14.63 ppm in its 14 H-NMR spectrum. Also, the 13 C-NMR spectrum exhibited the important signal of C=O at δ 164.8 ppm.

Biological evaluations

Antimicrobial activity

The in vitro antimicrobial activities of the synthesized products were screened toward three bacterial strains, namely Streptococcus pyogenes, Staphylococcus aureus, and

Scheme 11. The reaction of substrate 3 with PhP(O)Cl₂ in pyridine.

Escherichia coli, and three fungal strains, namely Aspergillus niger, Aspergillus clavatus, and Candida albicans. [36,37] Minimum inhibitory concentration (MIC) of all synthesized compounds was determined and given in Table 1. Ketoconazole and Ciprofloxacin as standard drugs were used for the antifungal and antibacterial activities, respectively. Variable antimicrobial activities toward the used microorganisms were recorded for the synthesized compounds. Compounds 5, 9, and 13 did not record any acceptable inhibitory effects toward all bacteria and fungi organisms in comparison with substrate 3 and standard drugs. Compound 12 exhibited relatively moderate effects against all organisms. On the other hand, products 10, 11, 14, and 15 showed good antibacterial and antifungal effects except compound 11 that recorded excellent antifungal activities equal to the standard drug. Furthermore, compounds 4, 6, 7, and 8 displayed excellent antimicrobial activities toward most of the microorganisms except compound 8 that showed moderate to good effects against the fungi organisms. Generally, the relationship between the structure and activity indicated that the introduction of phosphorus heterocycles and phosphonate groups to pyranopyrazole system can increase the antimicrobial properties. Therefore, the presence of 1,2-azaphosphole ring fused with the pyranopyrazole system in compounds 4 and 14 exhibited extremely good to excellent antibacterial and antifungal activities. In addition, the existence of 1,3,2-diazphosphinine rings

Table 1. The in *vitro* antimicrobial activities as minimum inhibitory concentration $(MIC, \mu g/mL)^a$ for the synthesized compounds.

Compound	Bacterial strains			Fungal strains		
	Streptococcus pyogenes	Staphylococcus aureus	Escherichia coli	Aspergillus niger	Candida albicans	Aspergillus clavatus
3	250	250	250	250	250	500
4	31.25	31.25	31.25	31.25	31.25	31.25
5	250	250	250	125	125	125
6	31.25	31.25	31.25	31.25	31.25	31.25
7	31.25	31.25	31.25	31.25	31.25	31.25
8	31.25	31.25	31.25	62.5	62.5	62.5
9	250	250	250	250	250	250
10	62.5	62.5	62.5	62.5	125	62.5
11	62.5	62.5	62.5	31.25	31.25	31.25
12	125	125	125	125	125	125
13	250	250	250	500	250	250
14	62.5	62.5	62.5	62.5	62.5	62.5
15	62.5	62.5	62.5	62.5	62.5	62.5
Ciprofloxacin	31.25	31.25	31.25			
Ketoconazole				31.25	31.25	31.25

^aAll results are expressed as mean \pm SD from three experiments (n = 3).

Table 2. The cytotoxcity of the products **4**, **6**, **7**, and **8** using the Brine Shrimp bioassay.

Compound	Lethal Concentration (LC ₅₀ , μg/mL) ^a
4	1.65
6	7.32
7	5.67
8	2.36
Gallic acid	4.51

^aAll results are expressed as mean \pm SD from three experiments (n = 3).

possess P=S and P=O groups as in products 10, 11, and 15 exhibited significant improvements in the antimicrobial effects in comparison with the starting material 3. Interestingly, the design of skeleton pyrrolo[3',2':5,6]pyrano[2,3-c]pyrazole bearing diethyl phosphonate groups in compounds 6, 7, and 8 caused excellent inhibitory effects nearly to the standard drugs toward all the microorganisms except compound 8 which recorded good inhibitory effects against only fungal strains.

The cytotoxicity activity

The LC₅₀ values of the most bioactive synthesized compounds **4**, **6**, **7**, and **8** against brine shrimp death were evaluated by the reported method using Gallic acid as a standard. The results are summarized in Table 2. The lowest LC₅₀ values were found in the case of compounds **4** and **8** at 1.65 and 2.36 μ g/mL, respectively, indicating their higher cytotoxicity. On the other hand, compounds **6** and **7** displayed brine shrimp death at 7.32 and 5.67 μ g/mL, respectively. Therefore, both compounds **6** and **7** can be active potent antibacterial and antifungal agents with lower cytotoxicity. So, we can conclude that skeleton pyrrolo[3',2':5,6]pyrano[2,3-c] pyrazole bearing diethyl phosphonate groups is the most active potent antimicrobial agent with lower toxicity in host cells.

Table 3. The *in vitro* antioxidant activities as inhibitory concentration (IC₅₀, μg/mL) for the synthesized compounds by using DPPH and H₂O₂ methods.

	Inhibitory concentration (IC ₅₀ , μg/mL)		
Compound	DPPH method	H ₂ O ₂ method	
3	16.21 ± 0.29	22.53 ± 0.51	
4	11.86 ± 0.31	17.05 ± 0.51	
5	35.12 ± 0.29	40.23 ± 0.54	
6	10.12 ± 0.52	16.69 ± 0.31	
7	13.25 ± 0.23	19.36 ± 0.62	
8	14.67 ± 0.66	21.49 ± 0.34	
9	25.87 ± 0.00	30.53 ± 0.00	
10	18.38 ± 0.33	26.13 ± 0.26	
11	15.63 ± 0.23	24.65 ± 0.29	
12	24.36 ± 0.39	28.52 ± 0.27	
13	19.35 ± 0.21	29.14 ± 0.50	
14	12.43 ± 0.21	20.84 ± 0.33	
15	36.42 ± 0.76	50.46 ± 0.25	
Ascorbic acid	10.23 ± 0.23	18.62 ± 0.52	

Antioxidant activity

The synthesized compounds were investigated for their in vitro antioxidant properties by DPPH^[40,41] and H₂O₂^[42] methods. The ascorbic acid was used as standard control. The lower IC₅₀ values indicated to a higher antioxidant activity. The measured antioxidative properties of the synthesized compounds showed promising radical scavenging abilities (Table 3). The results revealed that compounds 5, 9, 12, and 15 exhibited poor radical scavenging abilities in comparison with ascorbic acid. However, remarkable activities were observed with products 3, 8, 10, 11, and 13. Interestingly, compounds 4 and 14 exhibited significant antioxidant activities due to the presence of 1,2-azaphosphole or 1,3,2-diazaphosphinine ring, two acidic NH groups, and extended conjugation. Besides, the connection of the pyrrole ring with NH and phosphonate group in compounds 6 and 7 caused promising antioxidative properties.

Antiproliferative activity

The in vitro antiproliferative activities of the synthesized compounds were determined by using the SRB assay^[43] against three human cancer cell lines, mammary gland breast cancer (MCF-7), liver cancer (HepG-2), and human colon cancer (HCT-116) in comparison with doxorubicin as reference drug. The IC50 values of cytotoxic activities of the synthesized compounds 3-15 are presented in Table 4. It is obvious that the thirteen tested compounds displayed variable degrees of inhibitory effects on the tested human tumor cell lines. Compounds 3, 5, 12, 13, and 14 revealed no acceptable cytotoxic activities on all tumor cell lines. For action on MCF-7, the highest cytotoxic effects were demonstrated by compounds 4, 6, 8, 9, 10, and 15 which showed the IC50 values in ranges from 3.7 to $8.4 \,\mu\text{g/mL}$ in comparison with doxorubicin (IC₅₀ = $1.4 \,\mu\text{g/mL}$). Moderate cytotoxic effects were also recorded by compounds 7 and 11 with values of $IC_{50} = 16.9$ and $14.5 \,\mu g/mL$, respectively. The HepG-2 cell line was demonstrated to be

Table 4. The IC_{50} ($\mu g/mL$) of the synthesized compounds against different tumor cell lines.

		IC ₅₀ (μg/mL)	
Compound	MCF-7	HepG-2	HCT-116
3	23.6 ± 0.5	120.8 ± 6.6	46.8 ± 2.4
4	8.3 ± 0.4	10.5 ± 0.5	3.7 ± 0.7
5	73.5 ± 1.6	102.5 ± 4.1	27.9 ± 0.9
6	8.4 ± 0.3	18.8 ± 0.8	7.7 ± 0.6
7	16.9 ± 0.5	20.3 ± 0.3	10.7 ± 1.3
8	5.04 ± 0.3	14.5 ± 2.2	22.1 ± 1.5
9	3.7 ± 0.7	6.4 ± 0.2	10.6 ± 0.7
10	5.6 ± 1.03	34.4 ± 2.5	30.7 ± 1.5
11	14.5 ± 0.4	47.5 ± 2.1	7.6 ± 0.5
12	85.9 ± 3.1	45 ± 3.2	52.8 ± 1.5
13	46.3 ± 5.2	1732.3 ± 6.6	361.1 ± 5.9
14	388.4 ± 9.9	97.3 ± 5.2	536.1 ± 4.2
15	8.1 ± 0.9	87.1 ± 3.8	38.2 ± 3.6
Doxorubicin	1.4 ± 0.07	1.6 ± 0.04	2.0 ± 0.03

the least sensitive among the tested carcinoma cells. However, the highest sensitivity to compound **9** as its growth inhibitory effect was the displayed value of IC₅₀ at 6.4 μ g/mL. The moderate activity was shown by compounds **4**, **6**, **7**, and **8** which have IC₅₀ values at 10.5, 18.8, 20.3, and 14.5 μ g/mL, respectively. Compounds **10** and **11** revealed weak cytotoxicity properties. In addition, the HCT-116 cancer cell line was weakly affected by compounds **8**, **10**, and **15**. However, a significant growth inhibitory effect was noticed by compounds **4**, **6**, and **11** with IC₅₀ values 3.7, 7.7, and 7.6 μ g/mL, respectively, compared with doxorubicin (IC₅₀=2.0 μ g/mL). The residual compounds **7** and **9** showed moderate antiproliferative activities on HCT-116 cell line with IC₅₀ values 10.7 and 10.6 μ g/mL, respectively. These results indicated a specific therapeutic index for those phosphorus compounds that have an auspicious perspective to be substances for the candidate for anticancer.

In this research, the following relationships of structure with activity were assumed by comparing the structures of the synthesized phosphorus compounds to their experimental cytotoxicity. In general, the study revealed that the new molecular frames exhibited cytotoxicity properties more slightly than the starting substrate 3. The presence of phosphorus heterocycles or phosphonate groups combined with the pyranopyrazole moiety may be causing a broad spectrum of antiproliferative effects on various carcinoma cells (MCF-7, HepG-2, and HCT-116). The presence of phosphoryl moiety such as P(O)(OEt)₂ and N = P-Ph as directly attached on the pyranopyrazole system in compounds 5 and 13 reduced the cytotoxic effects in comparison with the substrate 3. The construction of 1,3,2-diazaphosphinine ring fused with pyranopyrazole moiety in products 9, 10, 11, 12, and 15 exhibited acceptable antiproliferative properties. Remarkably, the existence of C=S and P=S group in the 1,3,2-diazaphosphinine unit as compounds 9, 10 and 11, enhanced the cytotoxic activity more than C=NH and C=O groups of 1,3,2-diazaphosphinine in compounds 12 and 15. Interestingly, the design of 1,2-azaphosphole ring fused with pyranopyrazole core as in compounds 4 and 14 led to variable toxic activities. This indicated that the presence of cyclic NH-P(O)OEt system increased the cytotoxic activity more than N=P(Ph)₂ system. On the other hand, the introduction of pyrrole ring bearing diethyl phosphonate groups to pyranopyrazole core

as in compounds 6, 7, and 8, increased the antiproliferative activities. Further, the presence of two diethyl phosphonate groups improved the antiproliferative potency of more than one diethyl phosphonate group.

Apoptotic effect

Apoptosis is a form of programmed cell death that occurs in multicellular organisms. Biochemical events lead to characteristic cell changes (morphology) and death. These changes include blebbing, cell shrinkage, nuclear fragmentation, chromatin condensation, chromosomal DNA fragmentation, and global mRNA decay.

The treated tumor cell lines MCF-7, HepG-2, and HCT-116 were stained with acridine orange and ethidium bromide for 48 h. After, their examination under a fluorescent microscope, the cells appeared in the form of four colors as follows: living cells (normal green nucleus with normal, round, intact nuclei, and cytoplasm that indicates the viability of the cell control), early apoptotic (early-stage apoptosis is represented by changes and ultimate loss of the mitochondrial membrane potential, they appeared as a bright green nucleus with fragmented chromatin), necrotic cells (necrosis is caused by factors external to the cell or tissue, such as infection, or trauma which result in the unregulated digestion of cell components, they appeared as uniformly bright red-stained cell nuclei) and late apoptotic (late-stage apoptosis that can be identified by looking at the defragmentation of DNA, they appeared as red-stained nuclei with chromatin fragmentation) (Supplementary Figures 1-3). [44,45] The highly early apoptotic cell death was observed in all types of treated tumor cells in all compounds. The considerable necrotic activity compared with other compounds was displayed only in compound 10, while the other compounds did not cause the death of the necrosis pathway in all cancer cells. The late apoptosis was obviously observed with compounds 3, 5, and 7 especially against MCF-7 and HepG-2 cell lines, while compounds 3, 7, and 14 were observed in the case of HCT-116 cell line (Supplementary Figures 4-6). The obtained results were referenced to the control that absence of any cell death manifestations.

Experimental

The melting points were determined in an open capillary tube on a digital Stuart SMP-3 apparatus. Infrared spectra were measured on FT-IR (Nicolet IS10) spectrophotometer using KBr disks and Perkin-Elmer 293 spectrophotometer using KBr disks. 1H- and ¹³C-NMR spectra were measured on Gemini-300BB spectrometer (400 and 100 MHz), using DMSO- d_6 as a solvent and TMS (δ) as an internal standard. ³¹P-NMR spectra were measured on a Bruker (162 MHz) spectrophotometer using DMSO- d_6 as a solvent, TMS as an internal standard, and 85% H₃PO₄ as an external reference. Mass spectra were recorded on direct probe controller inlet part to single quadrupole mass analyzer in (Thermo Scientific GCMS). Elemental microanalysis was performed on Perkin-Elmer 2400II at the Chemical War department, Ministry of Defense. The purity of the synthesized compounds was checked by thin-layer chromatography (TLC) and elemental microanalysis. The biological evaluations were measured in the King Khalid University, Abha, KSA.

Synthesis of 6-ethoxy-5-imino-3-methyl-6-oxido-1,4-diphenyl-1,4,5,7-tetrahydropyrazolo [4',3':5,6]pyrano[3,2-d][1,2]azaphosphole (4)

A mixture of diethyl phosphite (1.5 mL, 10 mmol) and compound **3** (1.64 g, 5 mmol) in the presence of trifluoroboron etherate (0.1 mL) was fused on water bath at 80–90 °C for 6h. The excess of reagent was evaporated under vacuum. The formed semi-solid was treated with n-hexane. The isolated solid was filtered off and crystallized from EtOH to give yellow solid in 42% yield (0.88 g); mp 166–168 °C. IR (KBr), (ν max, cm⁻¹): 3267 (br, 2NH), 3068 (C–H_{arom}), 2915, 2852 (C–H_{aliph}), 1658 (C=NH), 1604, 1581 (C=C), 1213 (P=O), 1081 (P–O–C). 1 H-NMR (400 MHz, DMSO- 4 6): 1.03 (t, 3H, 1 6 Hz, CH₃), 1.97 (s, 3H, CH₃), 3.79–3.87 (m, 2H, OCH₂), 4.95 (s, 1H, H-4), 7.11–7.86 (m, 10H, Ph–H), 9.94 (s, 1H, NH), 11.52 (s, 1H, NH). 13 C-NMR (100 MHz, DMSO- 4 6): 12.0 (CH₃), 16.1 (CH₃), 34.2 (C-4), 60.2 (OCH₂), 81.1 (C-4a), 117.7 (C-3a), 120.4 (C-2', 6 7 phenyl), 124.5 (C-4" phenyl), 126.0 (C-4" phenyl), 127.5 (C-3", 5 7 phenyl), 128.3 (C-2", 6 9 phenyl), 129.3 (C-3', 5 9 phenyl), 135.9 (C-1" phenyl), 140.0 (C-1' phenyl), 142.6 (d, 1 1 = 150.9 Hz, C-5), 144.9 (C-3), 151.2 (C-8a), 159.5 (C-7a). 31 P-NMR (162 MHz, DMSO- 4 6): 16.2 ppm. MS (m 7, 1 %): 420 (M⁺, 16%). Anal. Calcd. for C₂₂H₂₁N₄O₃P (420.21): C, 62.85%; H, 5.03%; N, 13.33%. Found: C, 62.59%; H, 4.88%; N, 13.01%.

Conclusion

We suggested a simple, efficient, one-pot, three-component method for the construction of new functionalized pyrano[2,3-c]pyrazolyl and pyrrolo[3',2':5,6]pyrano[2,3-c]pyrazolyl phosphonates, 2-sulfido(oxido)-pyrazolo[4',3':5,6]pyrano[2,3-d][1,3,2]diazaphosphinines and pyrazolo[4',3':5,6]pyrano[3,2-d][1,2]azaphospholes. The suggested pathway depended on reaction substrate 3 with some phosphorus reagents under different conditions. The synthesized compounds were screened *in vitro* for their antimicrobial, antioxidant, and anticancer activities. Both compounds 6 and 7 recorded potent antibacterial and antifungal activities, while compounds 4 and 6 showed promising antioxidative properties. For antiproliferative activity, compounds 4 and 9 showed the most potent cytotoxic effects.

Full experimental details and spectral data of the synthesized compounds can be accessed on the publisher's website.

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Refernces

- [1] Chen, X.; Alidori, S.; Puschmann, F. F.; Santiso-Quinones, G.; Benkő, Z.; Li, Z.; Becker, G.; Grützmacher, H.-F.; Grützmacher, H. Sodium Phosphaethynolate as a Building Block for Heterocycles. Angew. Chem. Int. Ed. Engl. 2014, 53, 1641-1645. DOI: 10.1002/anie.
- Franz, J. E.; Mao, M. K.; Sikorski, J. A. Gliphosate: A Unique Global Herbicide; American [2] Chemical Society: Washington, DC, 1997.
- Keglevich, G. Newer Developments in the Synthesis of P-Heterocycles. Curr. Org. Chem. [3] 2019, 23, 1342-1355. DOI: 10.2174/1385272823666190726093322.
- Mian, M. R.; Islamoglu, T.; Afrin, U.; Goswami, S.; Cao, R.; Kirlikovali, K. O.; Hall, M. G.; [4] Peterson, G. W.; Farha, O. K. Catalytic Degradation of an Organophosphorus Agent at Zn-OH Sites in a Metal-Organic Framework. Chem. Mater. 2020, 32, 6998-7004. DOI:10. 1021/acs.chemmater.0c02373...
- Mathey, F. Phosphorus-Carbon Heterocyclic Chemistry: The Rise of a New Domain; [5] Pergamon: New York, NY, 2001.
- [6] Golla, M.; Devineni, S. R.; Syed, R.; Rayalacheru, B. K.; Maram, R.; Chamarthi, N. R. New 2-Aminomethyl-2-Oxo- $2\lambda^5$ -Perhydro-[1,3,2]Oxazaphospholo[3,4-a]Pyridine: Design, Synthesis and Antimicrobial Activity. Phosphorus, Sulfur Silicon Relat. Elem. 2017, 192, 794-798. DOI: 10.1080/10426507.2016.1277528.
- [7] Abdou, W. M.; Barghash, R. F.; Khidre, R. E. Antineoplastic Activity of Fused Nitrogen-Phosphorus Heterocycles and Derived Phosphonates. Monatsh. Chem. 2013, 144, 1233-1342. DOI: 10.1007/s00706-013-0950-6.
- [8] Yamada, M.; Asai, K.; Yamashita, J.; Suyama, T.; Niimi, T.; Maddali, K.; Fujie, M.; Nakamura, S.; Yamashita, M. Preparation and Characterization of Phospha Sugar Analogues, 2,3-Dibromo-3-Methyl-1-Phenylphospholane 1-Oxide Derivatives, as Novel Anticancer Agents. Phosphorus, Sulfur Silicon Relat. Elem. 2010, 185, 2286-2291. DOI: 10. 1080/10426501003598622.
- Gilard, V.; Martino, R.; Malet-Martino, M.; Niemeyer, U.; Pohl, J. Chemical Stability and Fate of the Cytostatic Drug Ifosfamide and Its N-Dechloroethylated Metabolites in Acidic Aqueous Solutions. J. Med. Chem. 1999, 42, 2542-2560. DOI: 10.1021/jm980587g.
- [10] Smith, B. R.; Eastman, C. M.; Njardarson, J. T. Beyond C, H, O, and N! Analysis of the Elemental Composition of U.S. FDA Approved Drug Architectures. J. Med. Chem. 2014, 57, 9764–9773. DOI: 10.1021/jm501105n.
- Elliott, T. S.; Slowey, A.; Ye, Y. L.; Conway, S. J. The Use of Phosphonate Bioisosteres in [11] Medicinal Chemistry and Chemical Biology. Med. Chem. Commun. 2012, 3, 735-751. DOI: 10.1039/c2md20079a.
- Jockusch, S.; Tao, C.; Li, X.; Anderson, T. K.; Chien, M.; Kumar, S.; Russo, J. J.; [12] Kirchdoerfer, R. N.; Ju, J. A Library of Nucleotide Analogues Terminate RNA Synthesis Catalyzed by Polymerases of Coronaviruses That Cause SARS and COVID-19. Antiviral. Res. 2020, 180, 104857. DOI: 10.1016/j.antiviral.2020.104857.
- [13] Aslam, N.; White, J. M.; Zafar, A. M.; Jabeen, M.; Ghafoor, A.; Sajjid, N.; Noreen, S.; Khan, M. A. 4H-Pyrano[2,3-c]Pyrazoles: A Review. Arkivoc 2018, 2018, 139-203. DOI: 10. 24820/ark.5550190.p010.622.
- [14] Shahbazi, S.; Ghasemzadeh, M. A.; Shakib, P.; Zolfaghari, M. R.; Bahmani, M. Synthesis and Antimicrobial Study of 1,4-Dihydropyrano[2,3-c]Pyrazole Derivatives in the Presence of Amino-Functionalized Silica-Coated Cobalt Oxide Nanostructures as Catalyst. Polyhedron 2019, 170, 172-179. DOI: 10.1016/j.poly.2019.04.063.
- Dandia, A.; Saini, D.; Bhaskaran, S.; Saini, D. K. Promoted Green Synthesis of [15] Spiro[Pyrano[2,3-c]Pyrazoles] as Antioxidant Agents. Med. Chem. Res. 2014, 23, 725-734. DOI: 10.1007/s00044-013-0671-8.
- Shamroukh, A. H.; Zaki, M. E. A.; Morsy, E. M. H.; Abdel-Motti, F. M.; Abdel-Megeid, [16] F. M. E. Synthesis of Pyrazolo[4',3':5,6]Pyrano[2,3-d]Pyrimidine Derivatives for Antiviral Evaluation. Arch. Pharm. 2007, 340, 236-243. DOI: 10.1002/ardp.200700005.

- [17] Sun, X.; Zhang, L.; Gao, M.; Que, X.; Zhou, C.; Zhu, D.; Cai, Y. Nanoformulation of a Novel Pyrano[2,3-c]Pyrazole Heterocyclic Compound AMDPC Exhibits anti-Cancer Activity via Blocking the Cell Cycle through a P53-Independent Pathway. *Molecules* **2019**, 24, 624–633. DOI: 10.3390/molecules24030624.
- [18] Ali, T. E.; Assiri, M. A.; El-Shaaer, H. M.; Fouda, A. M.; Hassan, M. M.; Hassanin, N. M. Reaction of 2-Imino-2*H*-Chromene-3-Carboxamide with Phosphorus Halides: Synthesis of Some Novel Chromeno[2,3-d][1,3,2]Diazaphosphinines and Chromeno[4,3-c][1,2] Azaphosphole and Their Antioxidant and Cytotoxicity Properties. *Heterocycles* **2019**, *98*, 681–692. DOI: 10.3987/COM-19-14062.
- [19] Ali, T. E.; Assiri, M. A.; El-Shaaer, H. M.; Hassan, M. M.; Fouda, A. M.; Hassanin, N. M. Reaction of 2-Imino-2H-Chromene-3-Carboxamide with Some Phosphorus Esters: Synthesis of Some Novel Chromenes Containing Phosphorus Heterocycles and Phosphonate Groups and Their Antioxidant and Cytotoxicity Properties. *Synth. Commun.* **2019**, 49, 2983–2994. DOI: 10.1080/00397911.2019.1652323.
- [20] Hassanin, N. M.; Ali, T. E.; El-Shaaer, H. M.; Hassan, M. M. Reaction of 2-Imino-2*H*-Chromene-3-Carboxamide with Phosphorus Sulfides: Synthesis of Novel 2-Sulfido-2,3-Dihydro-4H-Chromeno[2,3-d][1,3,2]Diazaphosphinines. *Phosphorus, Sulfur Silicon Relat. Elem.* **2018**, *193*, 651–655. DOI: 10.1080/10426507.2018.1487439.
- [21] Assiri, M. A.; Ali, T. E.; Hassanin, N. M.; Yahia, I. S.; Sakr, G. B. Reaction of 2-Imino-2*H*-Chromene-3-Carboxamide with Phosphorus Isothiocyanates: First Synthesis of Novel Chromeno[2,3-d]Pyrimidinyl and Bis(Chromeno[2,3-d]Pyrimidinyl)Phosphines and Chromeno[2,3:4,5]Pyrimido[2,1-d][1,3,5,2]Triazaphosphinine. *J. Heterocyclic Chem.* **2019**, *56*, 1646–1650. DOI: 10.1002/jhet.3552.
- [22] Ali, T. E.; Assiri, M. A.; Hassanin, N. M.; Yahia, I. S.; Hussien, M. S. A Convenient Synthetic Route of Diethyl (4-Oxo-Chromeno[2,3-d] Pyrimidin-2(5)-yl)Phosphonates. J. Heterocyclic Chem. 2019, 56, 1684–1686. DOI: 10.1002/jhet.3550.
- [23] Kiyania, H.; Samimib, H. A.; Ghorbania, F.; Esmaielia, S. One-Pot, Four-Component Synthesis of Pyrano[2,3-c]Pyrazoles Catalyzed by Sodium Benzoate in Aqueous Medium. *Curr. Chem. Lett.* **2013**, *2*, 197–206. DOI: 10.5267/j.ccl.2013.07.002.
- [24] Sravya, G.; Suresh, G.; Zyryanov, G. V.; Balakrishna, A.; Reddy, N. B. K₂CO₃/Al₂O₃: An Efficient and Recyclable Catalyst for One-Pot, Three Components Synthesis of α-Amino-Phosphonates and Bioactivity Evaluation. *Asian J. Chem.* **2019**, *31*, 2383–2388. DOI: 10. 14233/ajchem.2019.22194.
- [25] Abu-Shanab, F. A.; Mousa, S. A. S.; Eshak, E. A.; Sayed, A. Z.; Al-Harrasi, A. Dimethylformamide Dimethyl Acetal (DMFDMA) in Heterocyclic Synthesis: Synthesis of Polysubstituted Pyridines, Pyrimidines, Pyridazine and Their Fused Derivatives. *Int. J. Org. Chem.* **2011**, *1*, 207–214. DOI: 10.4236/ijoc.2011.14030.
- [26] Abdou, W. M.; Shaddy, A. A.; Khidre, R. E.; Awad, G. E. A. Synthesis and Antimicrobial Evaluation of Newly Synthesized N,S-Bisphosphonate Derivatives. *J. Heterocyclic Chem.* **2016**, *53*, 525–532. DOI: 10.1002/jhet.2306.
- [27] Dąbrowska, E.; Burzyńska, A.; Mucha, A.; Matczak-Jon, E.; Sawka-Dobrowolska, W.; Berlicki, Ł.; Kafarski, P. Insight into the Mechanism of Three Component Condensation Leading to Aminomethylenebisphosphonates. *J. Organomet. Chem.* **2009**, *694*, 3806–3813. DOI: 10.1016/j.jorganchem.2009.07.025.
- [28] Ali, T. E.; El-Edfawy, S. M. A Convenient Synthesis and Biological Evaluation of Some Novel Linear and Cyclic α-Aminophosphonic Acid Derivatives Containing a Quinazolinone Ring. *Res. Chem. Intermed.* **2016**, 42, 1329–1347. DOI: 10.1007/s11164-015-2088-0.
- [29] Kozachenko, A. P.; Shablykin, O. V.; Gakh, A. A.; Rusanov, E. B.; Brovarets, V. S. Synthesis of New Heterocyclic System of 4,5,7,8-Tetrahydroimidazo[1,2-c][1,3]Thiazolo[4,5-e][1,3,2] Diazaphosphinine Starting from 2-Aroylamino-3,3-Dichloroacrylonitrile. *Heteroatom Chem.* **2010**, *21*, 492–498. DOI: 10.1002/hc.20638.
- [30] Ali, T. E.; Hassan, M. M. Reaction of 2-Cyano-3-(4-Oxo-4H-Chromen-3-yl)Prop-2-Enamide with Some Phosphorus Reagents: Synthesis of Some Novel Diethyl



- Phosphonates, 1,2,3-Diazaphosphinanes, 1,2,3-Thiazaphosphinine and 1,2-Azaphospholes Bearing a Chromone Ring. Res. Chem. Intermed. 2018, 44, 173-189. DOI: 10.1007/s11164-017-3096-z.
- [31] Ozturk, T.; Ertas, E.; Mert, O. ChemInform Abstract: A Berzelius Reagent, Phosphorus Decasulfide (P₄S₁₀), in Organic Syntheses. Chem. Rev. **2010**, 110, 3419–3478. DOI: 10. 1021/cr900243d.
- Khalladi, K.; Touil, S. Synthesis of Novel Fused Thienodiazaphosphorine Derivatives from [32] 2-Amino-3-Cyanothiophenes and Lawesson's Reagent. J. Sulfur Chem. 2012, 33, 27-32. DOI: 10.1080/17415993.2011.639021.
- Younes, S. H. H.; Mohamed, S. K.; Albayati, M. R. Studies on Organophosphorus [33] Compounds. Part 1: Synthesis and in Vitro Antimicrobial Activity of Some New Pyrimido[5',4':5,6]Pyrano[2,3-d][1,3,2]Thiazaphosphinine Compounds. *Arch.* Chem. Life Sci. 2013, 346, 727-732. DOI DOI: 10.1002/ardp.201300171...
- Ali, T. E.; Assiri, M. A.; Abdel-Kariem, S. M.; Yahia, I. S. Facile Synthesis of Novel 6-[34] Methyl-5-Phenyl-2-Sulfido-1,2,3,5-Tetrahydro-4H-[1,2]Oxazolo[4',5':5,6]Pyrano[2,3d][1,3,2] Diazaphosphinines. J. Sulfur Chem. 2018, 39, 472-482. DOI: 10.1080/17415993. 2018.1455837.
- [35] Ali, T. E. Synthesis of Some New 1,3,2-Oxazaphosphinine, 1,3,2-Diazaphosphinine, Acyclic, and/or Cyclic α-Aminophosphonate Derivatives Containing the Chromone Moiety. Phosphorus, Sulfur Silicon Relat. Elem. 2009, 185, 88-96. DOI: 10.1080/ 10426500802713309.
- [36] Rahman, A. U.; Choudhary, M. I.; Thomsen, W. J. Bioassay Techniques for Drug Development; Harwood Academic Publishers: Amsterdam, The Netherlands, 2001.
- [37] Balouiri, M.; Sadiki, M.; Ibnsouda, S. K. Methods for in Vitro Evaluating Antimicrobial Activity: A Review. J. Pharm. Anal. 2016, 6, 71-79. DOI: 10.1016/j.jpha.2015.11.005.
- Meyer, B. N.; Ferrigni, N. R.; Putnam, J. E.; Jacobsen, L. B.; Nichols, D. E.; McLaughlin, [38] J. L. Brine Shrimp: A Convenient General Bioassay for Active Plant Constituents. Planta Med. 1982, 45, 31-34. DOI: 10.1055/s-2007-971236...
- [39] Finney, D. J. Probit Analysis, 3rd ed.; Cambridge University Press: Cambridge, UK, 1972.
- [40] Kato, K.; Terao, S.; Shimamoto, N.; Hirata, M. Studies on Scavengers of Active Oxygen Species. 1. Synthesis and Biological Activity of 2-O-Alkylascorbic Acids. J. Med. Chem. 1988, 31, 793-798. DOI: 10.1021/jm00399a019...
- Siddhuraju, P.; Becker, K. The Antioxidant and Free Radical Scavenging Activities of [41]Processed Cowpea (Vigna unguiculata (L.) Walp.) Seed Extracts. Food Chem. 2007, 101, 10-19. DOI: 10.1016/j.foodchem.2006.01.004.
- [42] Winston, G. W.; Regoli, F.; Dugas, A. J.; Fong, J. H.; Blanchard, K. A. A. Rapid Gas Chromatographic Assay for Determining Oxyradical Scavenging Capacity of Antioxidants and Biological Fluids. Free Radic. Biol. Med. 1998, 24, 480-493. DOI: 10.1016/s0891-5849(97)00277-3...
- [43] Skehan, P.; Storeng, R.; Scudiero, D.; Monks, A.; McMahon, J.; Vistica, D.; Warren, J. T.; Bokesch, H.; Kenney, S.; Boyd, M. R. New Colorimetric Cytotoxicity Assay for Anticancer-Drug Screening. J. Natl. Cancer Inst. 1990, 82, 1107-1112. DOI: 10.1093/jnci/ 82.13.1107.
- [44]Albright, F.; Stephenson, R. A.; Agarwal, N.; Teerlink, C. C.; Lowrance, W. T.; Farnham, J. M.; Albright, L. A. Prostate Cancer Risk Prediction Based on Complete Prostate Cancer Family History. Prostate 2015, 75, 390-398. DOI: 10.1002/pros.22925...
- Liu, E. H.; Qi, L. W.; Wu, Q.; Peng, Y. B.; Li, P. Anticancer Agents Derived from Natural [45] Products. Mini Rev. Med. Chem. 2009, 9, 1547-1555. DOI: 10.2174/138955709790361520...