ARTICLE IN PRESS

Bioorganic & Medicinal Chemistry Letters xxx (2016) xxx-xxx





Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl

Amino(oxo)acetate moiety: A new functional group to improve the cytotoxicity of betulin derived carbamates

Lucie Heller^a, Vincent Perl^a, Jana Wiemann^a, Ahmed Al-Harrasi^b, René Csuk^{a,*}

^a Martin-Luther-University Halle-Wittenberg, Organic Chemistry, Kurt-Mothes-Str. 2, D-06120 Halle (Saale), Germany ^b University of Nizwa, Chair of Oman's Medicinal Plants and Marine Natural Products, PO Box 33, Birkat Al-Mauz, Nizwa, Oman

ARTICLE INFO

Article history: Received 4 March 2016 Revised 17 April 2016 Accepted 20 April 2016 Available online xxxx

Keywords: Betulin Cytotoxicity Triterpene

Cancers are a family of diseases, and one of the hallmarks of cancer is a continuous and unregulated growth of cells leading either to a solid tumor or to diffusely spread cells. In Europe, North America and parts of Asia nearly half of people diagnosed with cancer and receiving a treatment die of cancer or because of the treatment. Cancer can be treated and even cured either by surgery, application of radiation, immunotherapy or by chemotherapy. Although it is indisputable that notable progress and major achievements in the therapy of all kinds of cancer have been made by chemotherapy, but there remains the need to search for new chemotherapeutics. Starting several decades ago, natural product derived compounds made their way into modern anticancer chemotherapy, and nowadays they play a major role in developing new cytotoxic agents. Among these natural products, many terpenoids¹⁻⁸ have shown to act as excellent starting points for finding new therapeutic drugs^{2,3,5,6,8} and even as potential chemopreventive agents^{1,4-7} During our search for new cytotoxic compounds derived from secondary natural products, we became interested in triterpenoic acids (cf. betulinic acid⁹ (**BA**), boswellic acid,^{10,11} glycyrrhetinic acid,¹² maslinic acid,¹³ oleanolic acid,¹⁴ ursolic acid,¹⁴ tormentic acid¹⁵ and others) and other triterpenes (cf. betulin, BN, Fig. 1). Interestingly enough, while there are many reports on the cytotoxicity of BA or derivatives,16-22 BN is of reduced cytotoxicity, and the number of betulin derived cytotoxic derivatives remained limited.23-25

Apart from the fact that **BN** is a cheap, renewable resource usually obtained from agro-industrial waste. Thus, **BN** is easily to

http://dx.doi.org/10.1016/j.bmcl.2016.04.055 0960-894X/© 2016 Elsevier Ltd. All rights reserved.

ABSTRACT

While 3-O-acetylated betulin derivatives carrying a carbamate moiety at position C-28 are of rather low cytotoxicity for human tumor cell lines, the corresponding C-3 amino(oxo) acetates show good cytotoxicity. For example, an EC₅₀ as low as 2.0 μ M was found for (3 β) 28-{[(hexylamino)carbonyl]oxy}lup-20 (29)-en-3-yl amino(oxo)acetate (**16**) employing the ovarian cancer cell line A2780.

© 2016 Elsevier Ltd. All rights reserved.



Figure 1. Structure of betulin (BN) and betulinic acid (BA).

obtain in huge amounts from extracting birch bark, and it is an interesting scaffold for accessing derivatives displaying various biological properties. For example, in HeLa cancer cells BN, unlike BA, seems to trigger apoptosis through an intrinsic pathway by a sequential activation of caspase-9 and caspase 3/-7 and the cleavage of poly-(ADP-ribose)-polymerase.²⁶ Recently, **BN** derived carbamates came into the focus of scientific interest because of their increased cytotoxic activity^{27–31} and an improved therapeutic index.^{30,32} Hence, we decided to have a closer look onto this class of compounds, and to explore their cytotoxic properties.³²

Following well known procedures, **BN** was acetylated and diacetate **1** was obtained in excellent yields (Scheme 1). A regioselective and partial deacetylation of **1** with KOH in a mixture of THF and

^{*} Corresponding author.

ARTICLE IN PRESS



Scheme 1. Synthesis of target compounds **13–17**: (a) Ac₂O, NEt₃, DMAP, DCM, 12 h, 25 °C, 90%; (b) KOH, MeOH, THF, 0 °C, 30 min, 57%; (c) microwave-assisted, 7 h, 120 °C, THF, RNCO, **3** (81%), **4** (83%), **5** (81%), **6** (73%), **7** (87%); (d) KOH, MeOH, THF, 25 °C, **8** (2 d, 95%), **9** (1 d, 93%), **10** (2 d, 94%), **11** (1 d, 88%), **12** (2 d, 89%); (e) (COCl₂)₂, DCM, 25 °C, 1 h; (f) NH₃, DCM, 25 °C, 12 h, **13** (72%), **14** (63%), **15** (78%), **16** (76%), **17** (60%).

methanol advanced smoothly at 0 °C and provided monoacetate **2** in 57% yield. Microwave assisted reaction of **2** with alkyl isocyanates or phenyl isocyanate gave 3-O-acetylated carbamates **3–7**. They were deacetylated with KOH in methanol³² to yield carbamates **8–12**.

Previous screening of compounds **3**–**7** in photometric sulforhodamine B (SRB)³³ assays showed these compounds of low cytotoxicity (EC₅₀ >30 µmol = cut-off of the assay)—these compounds were of the same low cytotoxicity as parent **BN** (Table 1). Interestingly enough, on deacetylation of **3–7**, for compounds **8–12** cytotoxicity was restored, and EC_{50} values in the same magnitude as observed for **BA** were measured.

The reaction of secondary alcohols with oxalyl chloride is known to yield alkyl chloro(oxo) acetates,^{34–38} but their reactions with ammonia or amines leading to alkyl amino(oxo)acetates have only scarcely been applied.^{39–44} As an alternative, decomposing halonitro-acetic acid derivatives has been suggested.⁴⁵

Table 1

Cytotoxicity of selected compounds, betulin (**BN**) and betulinic acid (**BA**, standard) (EC₅₀ values in μ M from SRB assays after 96 h of treatment; the values are averaged from three independent experiments performed each in triplicate; confidence interval CI = 95 %; cut-off of the assay 30 μ M)

EC ₅₀	518A2	A2780	HT29	MCF-7	A549	HeLa
BA	11.9 ± 1.7	11.0 ± 1.9	14.4 ± 1.5	14.8 ± 1.9	14.9 ± 1.6	8.8 ± 0.7
BN	>30	>30	>30	>30	>30	>30
1	>30	20.3 ± 2.4	25.7 ± 2.9	>30	21.5 ± 3.0	>30
2	>30	28.5 ± 1.9	>30	20.2 ± 2.0	>30	>30
3	10.2 ± 1.3	11.1 ± 1.4	>30	20.4 ± 2.6	14.1 ± 1.2	17.0 ± 1.6
4-7	>30	>30	>30	>30	>30	>30
8	12.5 ± 3.1	4.6 ± 1.0	>30	12.1 ± 2.1	12.0 ± 2.6	>30
9	15.9 ± 2.3	10.6 ± 1.1	16.3 ± 2.0	15.5 ± 3.0	14.2 ± 1.7	9.4 ± 0.6
10	8.0 ± 1.1	7.0 ± 0.9	7.5 ± 1.3	12.5 ± 1.4	8.6 ± 1.4	10.1 ± 1.7
11	5.6 ± 0.7	7.6 ± 1.3	>30	>30	12.7 ± 2.6	7.4 ± 1.9
12	23.9 ± 3.0	7.2 ± 0.7	16.3 ± 1.8	24.6 ± 2.1	>30	11.5 ± 1.5
13	11.4 ± 1.1	10.0 ± 0.9	15.3 ± 2.0	11.2 ± 0.9	13.7 ± 1.1	14.1 ± 2.4
14	9.9 ± 0.9	7.0 ± 0.9	12.8 ± 2.7	12.5 ± 2.5	10.5 ± 1.7	12.8 ± 1.9
15	7.9 ± 1.2	6.4 ± 1.0	10.1 ± 1.2	10.0 ± 1.5	9.1 ± 1.8	11.2 ± 1.8
16	6.5 ± 1.1	2.0 ± 0.4	7.3 ± 1.6	5.6 ± 1.1	7.4 ± 1.3	10.1 ± 1.5
17	15.3 ± 1.0	17.3 ± 0.7	21.4 ± 0.6	17.6 ± 0.6	20.4 ± 0.8	10.5 ± 0.7

Human cancer cell lines: 518A2 (melanoma), A2780 (ovarian carcinoma), HT29 (colorectal adenocarcinoma), MCF-7 (breast adenocarcinoma), A549 (lung adenocarcinoma), and HeLa (epitheloid cervix carcinoma).

Please cite this article in press as: Heller, L.; et al. Bioorg. Med. Chem. Lett. (2016), http://dx.doi.org/10.1016/j.bmcl.2016.04.055

ARTICLE IN PRESS

L. Heller et al. / Bioorg. Med. Chem. Lett. xxx (2016) xxx-xxx

As a result, the influence of the presence of an amino(oxo)-acetate onto the biological activity of a molecule has never been examined before. Thus, compounds **8–12** were allowed to react with oxalyl chloride followed by reacting the intermediary (and not isolated) chloro(oxo)-acetates with ammonia, and products **13–17** were obtained in good to moderate yields. Screening of these compounds in photometric SRB assays gave surprising results: While acetates **3–7** were of rather low cytotoxicity for a variety of different human tumor cell lines, the amino(oxo) acetates **13–17** showed good cytotoxicity. For example, an EC₅₀ as low as 2.0 µM was found for compound **16** employing the ovarian cancer cell line A2780.

In conclusion: there are numerous reports describing the positive influence of esters, amides, carbamates, hydroxamates and other derivatives onto the biological/cytotoxic activity of triterpenes. The deduction of general rules how to improve cytotoxicity while gaining an high therapeutic index remains difficult inasmuch as small structural modifications may alter the mode of action in a most dramatic way: for example, chloroacetylation of BN seems to improve cytotoxicity⁴⁶ while acetylation of **BN** gave less active derivatives^{46,47} and introducing a long chain alkyl ester at HO-C (28) in BN caused a loss of cytotoxicity.⁴⁶ Carbamates exhibited enhanced selectivity toward different tumor cell lines^{28,31,48} but the effect of introducing an amino(oxo)acetate has never been studied before. Calculations of log P values suggest, that the amino(oxo)acetates 13-17 possess logP values similar to analogs with an unprotected hydroxyl group (8-12), while for acetates 3-7 significantly higher log P values were calculated. This 'new' functional variation might have some positive and significant influence onto the bioactivity of potential drugs thus warranting future investigations.

Acknowledgments

Thanks are due to Dr. R. Kluge for measuring the ESI-MS spectra, and to Dr. D. Ströhl and his team for many NMR spectra. We like to thank J. Wiese, MSc, for measuring optical rotations, UV–Vis and IR spectra, and Ms J. Pech for the microanalyses. The cell lines were kindly provided by Dr. Thomas Müller (Dept. of Haematology/ Oncology, Martin-Luther Universität Halle-Wittenberg). Support by the 'Gründerwerkstatt–Biowissenschaften', the 'WissenschaftsCampus Halle WCH' and the Oman Research Council (ORG/HSS/14/004) is gratefully recognized.

Supplementary data

Supplementary data (experimental procedures and full analytical data of all compounds) associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.bmcl. 2016.04.055.

References and notes

- Bodduluru, L. N.; Kasala, E. R.; Thota, N.; Barua, C. C.; Sistla, R. Toxicol. In Vitro 2014, 28, 1026.
- 2. Chudzik, M.; Korzonek-Szlacheta, I.; Krol, W. Molecules 2015, 20, 1610.
- 3. Krol, S. K.; Kielbus, M.; Muller, A. R.; Stepulak, A. BioMed Res. Int. 2015, 1.
- 4. Olden, K.; Vulimiri, S. V. Cancer Prev. Res. 2014, 7, 648.
- 5. Patlolla, J. M. R.; Rao, C. V. Curr. Pharm. Biotechnol. 2012, 13, 147.

- Shanmugam, M. K.; Dai, X.; Kumar, A. P.; Tan, B. K. H.; Sethi, G.; Bishayee, A. Biochem. Pharmacol. 2013, 85, 1579.
- Shanmugam, M. K.; Nguyen, A. H.; Kumar, A. P.; Tan, B. K. H.; Sethi, G. Cancer Lett. 2012, 320, 158.
- 8. Yang, A. Y.; Kim, H.; Li, W.; Kong, A.-N. T. Curr. Top. Med. Chem. 2016, 16, 697.
- Csuk, R. Targeting Cancer by Betulin and Betulinic Acid. In Novel Apoptotic Regulators in Carcinogenesis; Chen, G. G., Lai, P. B. S., Eds.; Springer: Dordrecht, 2012; pp 267–287.
- Csuk, R.; Barthel-Niesen, A.; Barthel, A.; Schäfer, R.; Al-Harrasi, A. Eur. J. Med. Chem. 2015, 100, 98.
- 11. Csuk, R.; Niesen-Barthel, A.; Schäfer, R.; Barthel, A.; Al-Harrasi, A. Eur. J. Med. Chem. 2015, 92, 700.
- 12. Csuk, R. Mini-Rev. Org. Chem. 2014, 11, 253.
- 13. Siewert, B.; Pianowski, E.; Obernauer, A.; Csuk, R. *Bioorg. Med. Chem.* 2014, 22, 594.
- 14. Sommerwerk, S.; Heller, L.; Serbian, I.; Csuk, R. Tetrahedron 2015, 71, 8528.
- 15. Csuk, R.; Siewert, B.; Dressel, C.; Schäfer, R. Eur. J. Med. Chem. 2012, 56, 237.
- 16. Eiznhamer, D. A.; Xu, Z.-Q. IDrugs 2004, 7, 359.
- 17. Fulda, S. Int. J. Mol. Sci. 2008, 9, 1096.
- 18. Fulda, S.; Krömer, G. Drug Discovery Today 2009, 14, 885.
- Mukherjee, R.; Kumar, V.; Srivastava, S. K.; Agarwal, S. K.; Burman, A. C. Anti-Cancer Agents Med. Chem. 2006, 6, 271.
- 20. Paduch, R.; Kandefer-Szerszen, M. Mini-Rev. Org. Chem. 2014, 11, 262.
- 21. Tripathi, L.; Kumar, P.; Singh, R. Curr. Bioact. Compd. 2009, 5, 160.
- 22. Zhang, D.-M.; Xu, H.-G.; Wang, L.; Li, Y.-J.; Sun, P.-H.; Wu, X.-M.; Wang, G.-J.; Chen, W.-M.; Ye, W.-C. Med. Res. Rev. 2015, 35, 1127.
- Bhui, K.; Srivastava, A. K.; Shukla, Y. Cytotoxic Action of Natural Pentacyclic Triterpenes on Cancer Cells: Lupane-type Compounds. In *Pentacyclic Triterpenes* as Promising Agents in Cancer; Salvador, J. A. R., Ed.; Nova Science Publishers Inc, 2010; pp 49–87.
- Sarek, J.; Kvasnica, M.; Vlk, M.; Biedermann, D. Semisynthetic Lupane Triterpenes with Cytotoxic Activity. In *Pentacyclic Triterpenes as Promising Agents in Cancer*; Salvador, J. A. R., Ed.; Nova Science Publishers Inc, 2010; pp 159–189.
- 25. Csuk, R. Expert Opin. Ther. Pat. 2014, 24, 913.
- 26. Li, Y.; Shen, J. T.; Gao, C.; Li, Q.; Jin, Y. H. Chem. Res. Chin. Univ. 2010, 26, 792.
- Cao, M.; Onyango, E. O.; Williams, C. R.; Royce, D. B.; Gribble, G. W.; Sporn, M. B.; Liby, K. T. Pharmacol. Res. 2015, 100, 135.
- Flekhter, O. B.; Boreko, E. I.; Nigmatullina, L. R.; Tret'yakova, E. V.; Pavlova, N. I.; Baltina, L. A.; Nikolaeva, S. N.; Savinova, O. V.; Galin, F. Z.; Tolstikov, G. A. Russ. J. Bioorg. Chem. 2003, 29, 594.
- Hubbs, J. L.; Fuller, N. O.; Austin, W. F.; Shen, R.; Creaser, S. P.; McKee, T. D.; Loureiro, R. M. B.; Tate, B.; Xia, W.; Ives, J.; Bronk, B. S. J. Med. Chem. 2012, 55, 9270.
- Kommera, H.; Kaluderovic, G. N.; Dittrich, S.; Kalbitz, J.; Dräger, B.; Müller, T.; Paschke, R. Bioorg. Med. Chem. Lett. 2010, 20, 3409.
- Santos, R. C.; Salvador, J. A. R.; Marin, S.; Cascante, M.; Moreira, J. N.; Dinis, T. C. P. Bioorg. Med. Chem. 2010, 18, 4385.
- Wiemann, J.; Heller, L.; Perl, V.; Kluge, R.; Ströhl, D.; Csuk, R. Eur. J. Med. Chem. 2015, 106, 194.
- Skehan, P.; Storeng, R.; Scudiero, D.; Monks, A.; Mcmahon, J.; Vistica, D.; Warren, J. T.; Bokesch, H.; Kenney, S.; Boyd, M. R. J. Natl. Cancer Inst. 1990, 82, 1107.
- 34. Nahar, L.; Sarker, S. D.; Turner, A. B. Nat. Prod. Commun. 2008, 3, 27.
- Piotrkowska, B.; Milewska, M. J.; Gdaniec, M.; Polonski, T. Tetrahedron: Asymmetry 2007, 18, 1486.
- 36. Zhao, Y.; Li, Y.; Wang, S.; Li, Z. ARKIVOC 2009, 152.
- 37. Zhao, Y.; Wang, G.; Li, Y.; Wang, S.; Li, Z. Chin. J. Chem. 2010, 28, 475.
- Zhao, Y.; Wang, G.; Li, Y.-Q.; Wang, S.-H.; Li, Z.-M. Chem. Res. Chin. Univ. 2010, 26, 380.
- Belokon, Y. N.; Clegg, W.; Harrington, R. W.; Ishibashi, E.; Nomura, H.; North, M. Tetrahedron 2007, 63, 9724.
- 40. Belokon, Y. N.; Ishibashi, E.; Nomura, H.; North, M. Chem. Commun. 2006, 1775.
- 41. Clarke, C. T.; Elliott, J. D.; Jones, J. H. J. Chem. Soc., Perkin Trans. 1 1978, 1088.
- 42. Guan, X.; Dwivedi, C.; Li, Y.; Yang, Y.; Lee, J.H.; Miskimins, W.K. US20150315131A1, 2015.
- 43. Surburg, H.; Looft, J.; Oertling, H.; Vossing, T. US20090054520A1, 2009.
- Xu, Y.-J.; McLaughlin, M.; Bolton, E. N.; Reamer, R. A. J. Org. Chem. 2010, 75, 8666.
- 45. Yurtanov, A. I. Izv. Akad. Nauk SSSR, Ser. Khim. 1992, 151.
- Kommera, H.; Kaluderovic, G. N.; Kalbitz, J.; Paschke, R. Invest. New Drugs 2011, 29, 266.
- 47. Ahmad, F. B. H.; Moghaddam, M. G.; Basri, M.; Rahman, M. B. A. *Biosci. Biotechnol. Biochem.* 2010, 74, 1025.
- 48. Wiemann, J.; Heller, L.; Csuk, R. Bioorg. Med. Chem. Lett. 2016, 26, 907.