

EPIPHORELLIC ACIDS 1 AND 2, TWO DIARYL ETHERS FROM THE LICHEN *CORNICULARIA EPIPHORELLA**

PETER FIEDLER, VICENTE GAMBARO, JUAN A. GARBARINO and WANDA QUILHOTT†

Departamento de Química, Facultad de Ciencia, Universidad Federico Santa María, Casilla 110-V, Valparaíso, Chile; †Escuela de Química y Farmacia, Facultad de Medicina, Universidad de Valparaíso, Casilla 92-V, Valparaíso, Chile

(Received 6 June 1985)

Key Word Index—*Cornicularia epiphorella*; Usneaceae; lichen; diaryl ethers; epiphorellic acid 1; epiphorellic acid 2.

Abstract—From the lichen *Cornicularia epiphorella* two new diaryl ethers, epiphorellic acids 1 and 2, were isolated besides the known compound atranorin. The structures of the new compounds were established by spectroscopic evidence and chemical transformations. The structure of epiphorellic acid 1 was also confirmed by correlation with a known compound.

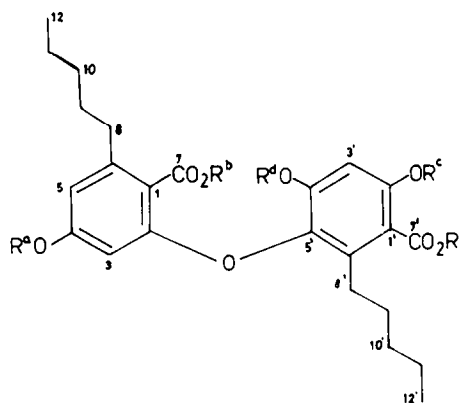
INTRODUCTION

In the course of systematic research on lichen substances from the Chilean flora, we have examined *Cornicularia epiphorella*. This lichen is distributed in the southern part of South America and in continental Chile it is associated with *Nothofagus* spp. and *Araucaria araucana* forests [2, 3]. In previous work on this species, using specimens collected in Tierra del Fuego, protolichesterinic acid was reported [4]. This paper describes the isolation and characterization of two new diaryl ethers, epiphorellic acid 1 (1) and epiphorellic acid 2 (2), present in *C. epiphorella*.

RESULTS AND DISCUSSION

A chloroform extract of *C. epiphorella* revealed a mixture of two main compounds by TLC. Chromatography of this extract afforded 1 and 2.

The pure compound 1, white crystals, mp 158–160°, analysed for $C_{26}H_{34}O_8$ by both mass spectral and ^{13}C NMR methods. The nature of this lichen substance was evident from the 1H NMR spectrum, which showed signals due to the presence of two C_3 -side chains (δ 0.90, 6H; 1.37, 12H; 2.57, 2H; and 2.90, 2H), two methoxy groups (δ 3.70 and 3.90), and three aromatic protons (δ 5.83, d , J = 2.5 Hz; 6.40, d , J = 2.5 Hz; and 6.50, s). Since a doublet aromatic proton occurs at an unusually upfield position [5], a depside or depsidone structure for 1 was tentatively discarded. The ^{13}C NMR spectrum (Table 1) confirmed the presence of two C_3 units, established the two methoxy groups as $Ar-COOMe$ (168.7, s ; 51.7, q and $Ar-OMe$, 56.0, q) and a carbon singlet at δ 173.6 was assigned to an $Ar-COOH$ group. Treatment of epiphorellic acid 1 (1) with ethereal diazomethane gave 1a, whose 1H NMR spectrum showed the characteristics of a new carbomethoxyl function (δ 3.90, s) with an *ortho*-hydroxy group (δ 11.60, s). The presence of this system was also evident



	R ^a	R ^b	R ^c	R ^d	R ^e
1	H	Me	H	Me	H
1a	H	Me	H	Me	Me
1c	Me	Me	Me	Me	Me
1d	Me	Me	H	Me	Me
1e	Me	H	H	H	Me
1f	Me	Me	H	H	Me

from the mass spectrum of 1 and 1a which exhibited peaks at m/z 456 due to loss of water and methanol, respectively. Because the carboxy groups of epiphorellic acid 1 (1) and its methyl derivative 1a do not participate in the linkage between the two aromatic moieties, it was suggested that 1 possessed a diaryl ether structure.

In order to determine the position of the carboxylic function in 1, it was partially decarboxylated to afford, after treatment of the mixture with diazomethane and

*Part 9 in the series "Studies on Chilean Lichens". For part 8 see ref. [1].

Table 1. ^{13}C NMR spectra of compounds 1, 1a, 2 and 2a

Carbon	1*	1a†	2*	2a†
1	115.1 s	114.6	115.2	114.3
2	163.8 s	162.2	163.9	162.2
3	99.3 d	98.7	99.5	98.7
4	158.5 s	157.7	158.5	157.4
5	109.6 d	109.2	109.8	109.3
6	143.7 s	143.4	142.8	141.6
7	168.7 s	169.2	168.5	168.7
1'	104.5 s	104.1	104.7	104.2
2'	159.4 s	157.9	159.7	157.4
3'	98.5 d	98.1	98.9	98.7
4'	157.8 s	157.1	158.0	158.5
5'	140.6 s	139.7	140.6	139.6
6'	134.4 s	134.1	134.4	133.8
7'	173.6 s	171.8	173.4	171.6
8	30.3 t	33.6	28.3	28.0
9	29.6 t	30.7	43.9	43.6
10	32.0 t	31.7	209.5 s	211.8 s
11	22.7 t	22.4	35.9	36.0
12	14.0 q	14.0	8.0	7.7
8'	31.3 t	30.4	31.2	30.4
9'	28.8 t	28.5	28.7	28.4
10'	32.5 t	32.2	32.8	32.2
11'	22.7 t	22.4	23.0	22.4
12'	14.0 q	14.0	14.3	14.1
MeO (C-4')	56.0 q	55.7	56.3	55.7
MeO (C-7')	51.7 q	52.1	52.0	52.1
MeO (C-7)	—	52.1 q		52.1

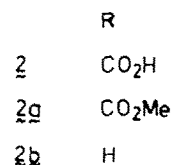
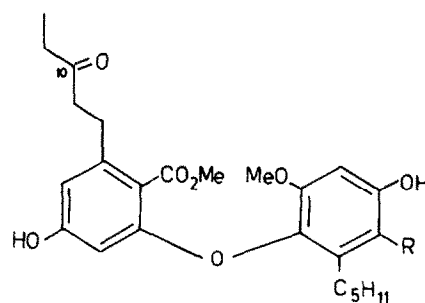
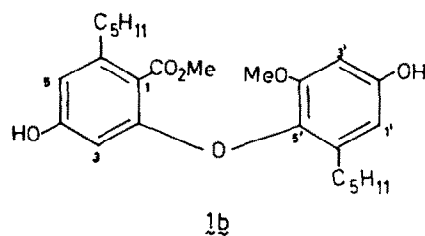
*In $\text{Me}_2\text{CO}-d_6$.†In CDCl_3 .

chromatographic separation, 1a and 1b. From the ^1H NMR spectrum of 1b it was readily deduced that the $\text{Ar}-\text{COOMe}$ group is situated in the same aromatic moiety as the two doublet aromatic protons, since in the new substance (1b) signals were exhibited corresponding to four aromatic protons. A doublet at $\delta 5.80$ (1H, $J = 2.5$ Hz) was assigned to H-3 and a broadened singlet centred at $\delta 6.30$ (3H) was due to H-5, H-1' and H-3' of 1b.

On the basis of data mentioned above and from a biogenetic point of view, methyl epiphorellate 1 (1a) could have either of the alternative structures 1a or 1f. However, the mp of 1f ($85-87^\circ$) [6] is significantly different from 1a ($135-137^\circ$). On the other hand, the ^1H NMR spectrum of 2-(4',6'-dihydroxy-3'-methoxycarbonyl-2'-pentylphenoxy)-4-methoxy-6-pentylbenzoic acid (1e) (sample supplied by M. V. Sargent) showed clearly that the two doublet aromatic protons (H-3 and H-5) are shifted more downfield (ca 0.4 ppm) in comparison with those of 1, thus indicating that the position of methylation was the 4'-hydroxyl group in 1, instead of the 4-hydroxyl group such as occurs in 1e.

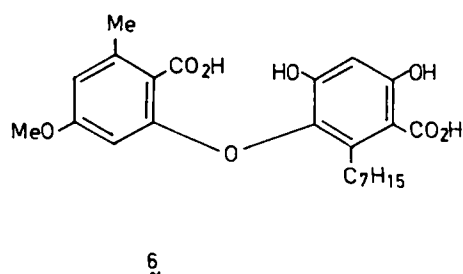
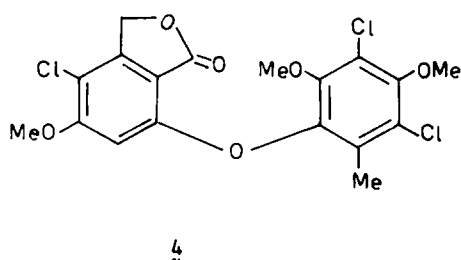
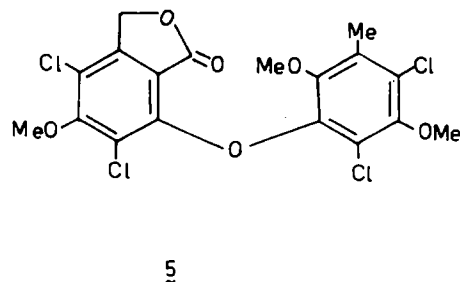
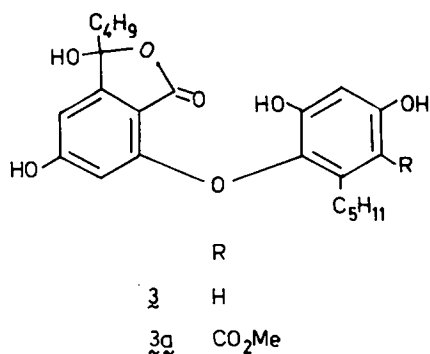
Finally, to confirm the structure 1 previously proposed for epiphorellic acid 1, 1 was converted into its permethylated derivative (1c). The spectral and physical data of 1c were in full agreement with those from the permethylated derivative of 1e.

Epiphorellic acid 2 (2), mp $146-148^\circ$, $\text{C}_{26}\text{H}_{32}\text{O}_9$, exhibited a ^1H NMR spectrum very similar to that of 1. The chemical shifts of the aromatic protons in 2 attached to C-3 ($\delta 5.78$, 1H, d, $J = 2.5$ Hz), C-5 ($\delta 6.36$, 1H, d, J



= 2.5 Hz) and C-3' ($\delta 6.52$, 1H, s) were almost identical to those of 1, which suggested that 2 had the same framework of the diaryl ether structure 1. The proton signals of 2 were markedly different from those of 1 at one C_5 unit, because the ^1H NMR spectrum showed the following signals: $\delta 1.00$ (3H, t, $J = 7.0$ Hz, H-12), 2.45 (2H, q, $J = 7.0$ Hz, H-11) and 2.79 (4H, br s, H-8 and H-9), which were tentatively attributable to an $\text{Ar}-\text{CH}_2\text{CH}_2\text{COEt}$ sequence. This suggestion was consistent with the ^{13}C NMR spectrum of 2 (Table 1), which contained one additional carbonyl function at C-10 (209.5, s) relative to that of 1. The remaining signals of this side chain were assigned as follows: 28.3, C-8; 43.9, C-9; 35.9, C-11; and 8.0, C-12. Additionally, the ^{13}C NMR spectrum exhibited the characteristics of another C_5 -side chain (C_5H_{11}), one $\text{Ar}-\text{OMe}$ group (56.3, q), one carbomethoxy function (168.5, s and 52.0, q) and one carboxylic group (173.4, s). Methylation of 2 gave the corresponding derivative (2a), which showed a new carbomethoxy function with an *ortho*-hydroxy group, such as occurs in 1a. Compound 2 was also partially decarboxylated to afford, after addition of diazomethane and chromatographic separation 2a and 2b. Again, the ^1H NMR spectrum of this new decarboxylated derivative (2b), showed that the carbomethoxy function is situated in the same aromatic moiety as the two doublet aromatic protons. Therefore, the unique difference between epiphorellic acids 1 (1) and 2 (2) is the additional carbonyl group in one of the C_5 -side chains of 2.

The position of the carbonyl C_5 -side chain of 2 and 2a was elucidated by comparison of ^{13}C NMR spectral data (Table 1) with those of 1 and 1a, respectively. The

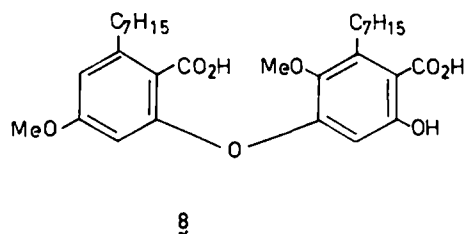
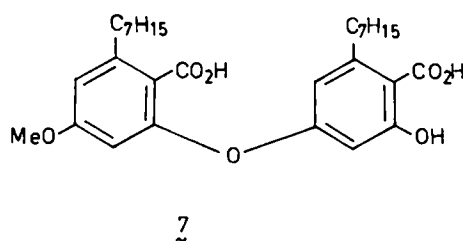


assignments of the ¹³C NMR spectral signals of 1, 1a, 2 and 2a were made on the basis of the observed multiplicities, empirical shift rules [7] and comparison with reported ¹³C NMR spectral data of known derivatives [8, 9]. On going from 1 to 2 and 1a and 2a, it could be appreciated that the signal due to C-6 was displaced upfield by 0.9 and 1.8 ppm, respectively. The differences observed in chemical shift could be rationalized by considering the γ effects of the carbonyl function [7]. The other carbon resonances remained almost unshifted, leading to the assignment of the structure of 2 and 2a as epiphorellic acid 2 and its methyl derivative.

Diphenyl ethers constitute a relatively rare group of lichen metabolites with eight examples known prior to the present work. They are norlobariol (3) [10], loxodinol (3a) [11], buellolide (4) [12], canesolide (5) [12], congrayanic acid (6) [13], micareic acid (7) [14], methoxymicareic acid (8) [14] and leprolomin [15]. Compounds 3, 3a, 4, 5 and 6 have been postulated as biogenetic depsidone cleavage products [12], whereas 7 and 8 have a distinctly different substitution pattern, and presumably, biogenetic origin [14]. On the other hand, leprolomin apparently arises by direct oxidative coupling of two molecules of C-methylphloracetophenone [15]. Therefore, epiphorellic acid 1 (1) and epiphorellic acid 2 (2) probably arise biogenetically by catabolism of their congeneric depsidones.

EXPERIMENTAL

Mps are uncorr. ¹H NMR were recorded at 60 and 100 MHz in Me₂CO-d₆ or CDCl₃ soln with TMS as int. standard. ¹³C NMR were recorded in Me₂CO-d₆ or CDCl₃ with TMS as int. standard. MS were recorded by direct inlet with 70 eV ionization. IR spectra were recorded in KBr pellets.



Cornicularia epiphorella (Nyl.) Du Rietz, collected on *Araucaria araucana*, Parque Nacional Conguillío, IX Región, Chile, in December 1983, was identified by G. Guzmán (Academia Superior de Ciencias Pedagógicas, Valparaíso); voucher specimens are deposited at the herbarium of Universidad F. Santa María.

Air dried lichen thalli (530 g) were triturated and extracted at room temp., successively with petrol (40–60°) and CHCl₃. The petrol extract was concd in vacuum and the residue subjected to fractional crystallization from CHCl₃–EtOH (1:1). On standing

a white solid deposited (31 mg), which was removed by filtration and identified as atranorin by direct comparison (mp, TLC, IR, ^1H NMR) with an authentic sample. The CHCl_3 extract (14.3 g) was also concd and showed a mixture of two components by TLC (R_f 0.53 and 0.41, silica gel, toluene–EtOAc– HCO_2H , 81:16:3). This crude material (2 g) was subjected to silica gel CC (80 g, HF_{254} for TLC) using a mixture of C_6H_6 –EtOAc– HCO_2H (45:3:2). Fractions of 40 ml were taken and combined based upon TLC monitoring. Fractions 26–42, containing a pure compound, were mixed and afforded 1. Fractions 48–53, containing another pure compound, were mixed and afforded 2.

Epiphorellic acid 1 (1). Compound 1 (1.15 g) was isolated as white crystals; mp 158–160° (Me_2CO –*n*-heptane). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3420, 2960–2860, 1710, 1700, 1625, 1610, 1430, 1370, 1280, 1250, 1150. ^1H NMR (CDCl_3): δ 0.90 (6H, *m*, H-12 and H-12'), 1.37 (12H, *m*, H-9, H-10, H-11, H-9', H-10' and H-11'), 2.57 (2H, *t*, J = 7.0 Hz, H-8), 2.90 (2H, *t*, J = 7.0 Hz, H-8'), 3.70 (3H, *s*, Ar–OMe), 3.90 (3H, *s*, Ar–COOMe), 5.83 (1H, *d*, J = 2.5 Hz, H-3), 6.40 (1H, *d*, J = 1.5 Hz, H-5), 6.50 (1H, *s*, H-3'). ^{13}C NMR see Table 1. MS m/z (rel. int.): 474 [M] $^+$ (15.3; $\text{C}_{26}\text{H}_{34}\text{O}_6$), 456 [$\text{M} - \text{H}_2\text{O}$] $^+$ (20), 430 [$\text{M} - \text{CO}_2$] $^+$ (100), 399 [430 – OMe] $^+$ (25), 355 (5), 285 (5), 271 (6.5), 193 (10), 192 (9), 137 (5.3).

Methyl epiphorellate 1 (1a). After addition of CH_2N_2 , 1 (200 mg) was transformed to 1a, which was purified by crystallization from Me_2CO –petrol (160 mg). White crystals, mp 135–137°. IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3400, 2950–2860, 1730, 1700, 1650, 1610, 1590, 1440, 1375, 1325, 1240, 1210, 1145. ^1H NMR (CDCl_3): δ 0.90 (6H, *m*, H-12 and H-12'), 1.33 (12H, *m*, H-9, H-10, H-11, H-9', H-10' and H-11'), 2.63 (4H, *m*, H-8 and H-8'), 3.67 (3H, *s*, Ar–OMe), 3.85 (3H, *s*, Ar–COOMe), 3.90 (3H, *s*, Ar–COOMe), 5.40 (1H, *br s*, OH-4), 5.73 (1H, *d*, J = 2.5 Hz, H-3), 6.27 (1H, *d*, J = 2.5 Hz, H-5), 6.40 (1H, *s*, H-3'), 11.60 (1H, *s*, OH-2'). ^{13}C NMR: see Table 1. MS m/z (rel. int.): 488 [M] $^+$ (100), 456 [$\text{M} - \text{MeOH}$] $^+$ (81.3), 355 (7), 353 (5), 251 (9), 250 (8), 219 (10), 191 (8), 163 (5).

Decarboxylated epiphorellic acid 1 (1b). A soln of 1 (200 mg) in 20 ml of EtOH was heated under reflux for 96 hr. After evapn of solvent the residue was treated with CH_2N_2 . This crude material showed two components by TLC. The substances were isolated by prep. TLC (silica gel, toluene–EtOAc– HCO_2H , 78:19:3) and yielded 1a (40 mg, R_f 0.77) and 1b (140 mg, R_f 0.45). Compound 1b, prisms, mp 66–67° (petrol). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3320, 2960–2860, 1725, 1715, 1610, 1470, 1280, 1160, 1100. ^1H NMR (CDCl_3): δ 0.90 (6H, *m*, H-12 and H-12'), 1.30 (12H, *m*, H-9, H-10, H-11, H-9', H-10' and H-11'), 2.45 (4H, *m*, H-8 and H-8'), 3.60 (3H, *s*, Ar–OMe), 3.90 (3H, *s*, Ar–COOMe), 5.80 (1H, *d*, J = 2.5 Hz, H-3), 6.30 (3H, *br s*, H-5, H-3' and H-1'). MS m/z (rel. int.): 430 [M] $^+$ (100), 399 [$\text{M} - \text{OMe}$] $^+$ (13), 355 (12), 341 (5), 285 (5), 271 (5), 193 (7), 192 (6), 137 (8), 43 (49), 41 (52).

Methyl epiphorellate 1-2,4-dimethyl ether (1c) and methyl epiphorellate 1-4-methyl ether (1d). A soln of 1 (200 mg) in dry Me_2CO (20 ml) was heated with anhydrous K_2CO_3 (1 g) and MeI (2 ml) for 48 hr. After filtration and evapn of solvent, the residue showed two components by TLC. The substances were isolated by prep. TLC (silica gel; toluene–EtOAc– HCO_2H ; 22:2:1) and yielded 1c (100 mg, R_f 0.53) and 1d (50 mg, R_f 0.76). Compound 1c, white crystals, mp 73–74° (petrol). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 2980–2860, 1730, 1620, 1590, 1490, 1470, 1440, 1380, 1330, 1270, 1160, 1100, 950, 820. ^1H NMR (CDCl_3): δ 0.87 (5H, *m*, H-12 and H-12'), 1.30 (12H, *m*, H-9, H-10, H-11, H-9', H-10' and H-11'), 2.50 (4H, *m*, H-8 and H-8'), 3.71 (3H, *s*, OMe), 3.80 (3H, *s*, OMe), 3.88 (3H, *s*, OMe), 3.93 (6H, *s*, OMe), 5.87 (1H, *d*, J = 2.5 Hz, H-3), 6.40 (1H, *d*, J = 2.5 Hz, H-5), 6.47 (1H, *s*, H-3'). MS m/z (rel. int.): 516 [M] $^+$ (100), 485 [$\text{M} - \text{OMe}$] $^+$ (13), 453 [485 – MeOH] $^+$ (5), 441 (9), 425 (10), 386 (6), 265 (10), 233 (8), 205 (5), 191 (5), 149 (5), 91 (5), 69 (5), 59 (7), 43 (12), 41 (13). Compound 1d, white crystals,

mp 95–96° (petrol). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 2960–2860, 1730, 1650, 1610, 1690, 1490, 1470, 1440, 1380, 1335, 1210, 1160, 1100, 1050, 950, 840, 810. ^1H NMR (CDCl_3): δ 0.87 (6H, *m*, H-12 and H-12'), 1.33 (12H, *m*, H-9, H-10, H-11, H-9', H-10' and H-11'), 2.70 (4H, *m*, H-8 and H-8'), 3.70 (3H, *s*, OMe), 3.78 (3H, *s*, OMe), 3.94 (3H, *s*, OMe), 3.97 (3H, *s*, OMe), 5.83 (1H, *d*, J = 2.5 Hz, H-3), 6.40 (1H, *d*, J = 2.5 Hz, H-5), 6.46 (1H, *s*, H-3'), 11.80 (1H, *s*, OH-2'). MS m/z (rel. int.): 502 [M] $^+$ (100), 470 [$\text{M} - \text{MeOH}$] $^+$ (48), 440 (10), 411 (6), 369 (4), 251 (9), 250 (10), 192 (6), 164 (3), 135 (5), 43 (8), 41 (10).

Methylation of 2-(4',6'-dihydroxy-3'-methoxycarbonyl-2'-penylphenoxy)-4-methoxy-6-pentylbenzoic acid (1e) to 1d and 1c. A soln of 1e (100 mg) in CHCl_3 (5 ml) was treated with freshly prepared CH_2N_2 for 5 min. Evapn of solvent and crystallization from petrol provided 1g as white crystals (85 mg). The spectral and physical data (TLC, mp, IR, ^1H NMR, MS) of this compound were in complete agreement with those of 1d. Compound 1g was treated with excess MeI in Me_2CO and K_2CO_3 , as described above. After filtration and evapn of solvent, the residue showed two components by TLC. The substances were isolated by prep. TLC, as described above and provided 1g = 1d (30 mg) and 1h (45 mg). The spectral and physical data (TLC, mp, IR, ^1H NMR, MS) of 1h were in full agreement with those of 1c.

Epiphorellic acid 2 (2). Compound 2 (280 mg) was isolated as white crystals; mp 146–148° (Me_2CO –*n*-heptane). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3360, 2960, 2860, 1740, 1700, 1655, 1625, 1610, 1485, 1450, 1440, 1415, 1370, 1280, 1240, 1215, 1150, 1100, 830, 740. ^1H NMR ($\text{Me}_2\text{CO}-d_6$): δ 0.82 (3H, *t*, J = 7.0 Hz, H-12'), 1.00 (3H, *t*, J = 7.0 Hz, H-12), 1.32 (4H, *m*, H-10' and H-11'), 1.48 (2H, *m*, H-9'), 2.45 (2H, *q*, J = 7.0 Hz, H-11), 2.79 (4H, H-8 and H-9), 2.93 (2H, *m*, H-8'), 3.80 (3H, *s*, Ar–OMe), 3.86 (3H, *s*, Ar–COOMe), 5.78 (1H, *d*, J = 2.5 Hz, H-3), 6.36 (1H, *d*, J = 2.5 Hz, H-5), 6.52 (1H, *s*, H-3'), 9.70 (2H, *m*, OH). ^{13}C NMR: see Table 1. MS m/z (rel. int.): 444 [$\text{M} - \text{CO}_2$] $^+$ (72.9), 413 [444 – OMe] $^+$ (13.5), 381 (33.7), 355 (100), 337 (20.8), 285 (96.6), 271 (17.5), 193 (50.4), 192 (54.8), 163 (22.2), 138 (21.8), 137 (67.7), 57 (58.6).

Methyl epiphorellate 2 (2a). After addition of CH_2N_2 , 2 (100 mg) was transformed to 2a, which was purified by crystallization from CHCl_3 –*n*-heptane (90 mg). White crystals, mp 108–109°. IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3440, 3220, 2960, 2860, 1730, 1685, 1660, 1610, 1440, 1380, 1325, 1270, 1240, 1210, 1150, 1090, 1050, 950, 840. ^1H NMR (CDCl_3): δ 0.84 (3H, *t*, J = 7.0 Hz, H-12'), 1.00 (3H, *s*, Ar–OMe), 3.86 (3H, *s*, Ar–COOMe), 3.91 (3H, *s*, 2H, *q*, J = 7.0 Hz, H-11), 2.75 (6H, *m*, H-8', H-8 and H-9), 3.66 (3H, *s*, Ar, OMe), 3.86 (3H, *s*, Ar–COOMe), 3.91 (3H, *s*, Ar–COOMe), 5.80 (1H, *d*, J = 2.5 Hz, H-3), 6.25 (1H, *d*, J = 2.5 Hz, H-5), 6.36 (1H, *s*, H-3'), 11.73 (1H, *s*, OH-2'). ^{13}C NMR: see Table 1. MS m/z (rel. int.): 502 [M] $^+$ (100), 470 [$\text{M} - \text{MeOH}$] $^+$ (88), 355 (25), 285 (10), 192 (5), 191 (6), 137 (15), 57 (20).

Decarboxylated epiphorellic acid 2 (2b). A soln of 2 (150 mg) in 20 ml of EtOH was heated under reflux for 70 hr. After evapn of solvent, the residue was treated with CH_2N_2 . This crude material showed two components by TLC. The substances were isolated by prep. TLC (silica gel; toluene–EtOAc– HCO_2H , 15:4:1) and yielded 2a (70 mg, 0.65) and 2b (65 mg, R_f 0.40). Compound 2b, white crystals, mp 145–146° (petrol). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3370, 2960–2860, 1725, 1690, 1600, 1490, 1450, 1270, 1200, 1140, 1110, 1090, 1070, 1030, 1010, 990, 950, 840. ^1H NMR (CDCl_3): δ 0.80 (3H, *t*, J = 8.0 Hz, H-12'), 1.00 (3H, *t*, J = 7.0 Hz, H-12), 1.27 (6H, *m*, H-9', H-10' and H-11'), 2.40 (4H, *m*, H-11, H-8'), 2.77 (4H, *br s*, H-8 and H-9), 3.67 (3H, *s*, Ar–OMe), 3.90 (3H, *s*, Ar–COOMe), 5.90 (1H, *d*, J = 2.5 Hz, H-3), 6.30 (1H, *d*, J = 2.5 Hz, H-5), 6.37 (2H, *s*, H-1' and H-3'). MS m/z (rel. int.): 444 [M] $^+$ (100), 413 (10), 381 (8), 355 (100), 285 (60), 193 (45), 192 (55), 137 (38), 57 (65).

Acknowledgements—We are grateful to 'Corporación Nacional Forestal (CONAF)', IX Región, Chile, for help in the collection of the lichen, to G. Guzmán, Academia Superior de Ciencias Pedagógicas de Valparaíso, for identification of the lichen, to Professor W. B. Mors, Núcleo de Pesquisas de Produtos Naturais, Universidade Federal do Rio de Janeiro, for recording the ^1H (100 MHz) and ^{13}C NMR spectra, to Dr. Rinaldo Marini-Bettolo, Università di Roma, for the mass spectra, to Professor M. V. Sargent, for providing a generous gift of 2-(4',6'-dihydroxy-3'-methoxycarbonyl-2'-pentylphenoxy)-4-methoxy-6-pentylbenzoic acid and to Mrs. M. Piovano, M. L. Vera and V. S. Hormaechea for technical assistance. This research was supported by a grant (\neq 841303 and 851303) from DGDCYT, Universidad Federico Santa María.

REFERENCES

1. Piovano, M., Garrido, M. I., Gambaro, V., Garbarino, J. A. and Quilhot, W. (1985) *J. Nat. Prod.* **48** (5).
2. Lindsay, D. C. (1974) *Br. Antarctic Surv. Sci. Rept.* **89**, 1.
3. Lamb, I. M. (1964) *Br. Antarctic Surv. Sci. Rept.* **38**, 1.
4. Hawksworth, D. L. and Moore, D. M. (1969) *The Bryologist* **72**, 247.
5. Huneck, S. and Linscheid, P. (1968) *Z. Naturforsch.* **23B**, 717.
6. Fox, C. H., Klein, E. and Huneck, S. (1970) *Phytochemistry* **9**, 2567.
7. Breitmaier, E. and Voelter, W. (1978) *^{13}C NMR Spectroscopy*, 2nd edn. Verlag Chemie, Weinheim.
8. Scott, K. N. (1972) *J. Am. Chem. Soc.* **94**, 8564.
9. Hylands, P. J. and Ingolfisdottir K. (1985) *Phytochemistry* **24**, 127.
10. Foo, L. Y. and Gwyn, S. A. (1978) *Experientia* **34**, 970.
11. Foo, L. Y. and Galloway, D. J. (1979) *Phytochemistry* **18**, 1977.
12. Sala, T., Sargent, M. V. and Elix, J. A. (1981) *J. Chem. Soc.* **849**.
13. Chester, D. O. and Elix, J. A. (1980) *Aust. J. Chem.* **33**, 1153.
14. Elix, J. A., Jones, A. J., Lajide, L., Coppins, B. J. and James, P. W. (1984) *Aust. J. Chem.* **37**, 2349.
15. Elix, J. A., Engkaninan, V., Jones, A. J., Raston, C. L., Sargent, M. V. and White, A. H. (1978) *Aust. J. Chem.* **31**, 2057.