# Syntheses and Antibacterial Activities of Tizoxanide, an *N*-(Nitrothiazolyl)salicylamide, and its *O*-Aryl Glucuronide<sup>†</sup>

J. Chem. Research (S), 1999, 44–45<sup>†</sup>

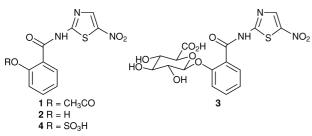
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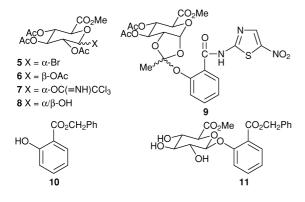
Mild hydrolysis of the broad-spectrum anaerobic antibacterial and antiparasitic agent nitazoxanide **1** affords tizoxanide **2**, which is a major metabolite of **1** retaining most of its activity; further metabolism of **2** leads to the *O*-aryl glucuronide **3**, efficiently synthesised in four steps from benzyl salicylate and showing slight antibacterial activity.

The thiazole derivative nitazoxanide 1, first described by Rossignol,<sup>1</sup> is a broad-spectrum antibacterial and antiparasitic agent, particularly efficacious against anaerobic bacteria<sup>2</sup> and as an anthelmintic and antiprotozoal agent.<sup>3,4</sup> The desacetyl metabolite of 1, tizoxanide 2, is itself a potent antibacterial and antiparasitic agent.<sup>2</sup> Further metabolism of 2 leads to the glucuronide 3 and sulfate 4 conjugates. In this paper, we describe the conversion of 1 to 2 and a convenient synthesis of 3, both to test for any remaining bioactivity and as an analytical standard.

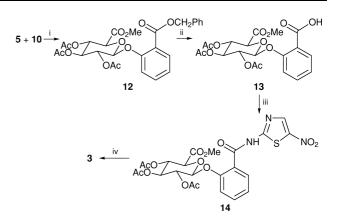


Tizoxanide 2 was readily obtained from nitazoxanide 1 in excellent yield by hydrolysis with aqueous HCl at 50 °C as previously given in the patent literature.<sup>5</sup> However, 2 could not be coupled to the bromosugar 5 using either the Koenigs–Knorr or lithium phenolate<sup>6</sup> methods, and acidcatalysed reactions with the tetraacetate  $6^{7,8}$  or imidate  $7^9$  were also unavailing, the low organic solubility of 2 being a major problem.

A conjugate was obtained when **2** was reacted with the 1-hydroxysugar **8**<sup>10</sup> under Mitsunobu conditions.<sup>11</sup> The <sup>1</sup>H NMR spectrum [in particular  $\delta_{\rm H}$  1.8 (s) and 5.9 (d)] was consistent with the orthoester **9**<sup>11</sup> rather than the desired glucuronide.



†This is a **Short Paper** as defined in the Instructions for Authors, Section 5.0 [see *J. Chem. Research* (*S*), 1999, Issue 1]; there is therefore no corresponding material in *J. Chem. Research* (M).



Acid-catalysed condensation of benzyl salicylate **10** with **6** or **7** also proved quite ineffective: by contrast a 2,6dimethylphenol has been successfully glucuronidated<sup>12</sup> using the classical Helferich procedure (**6** + tosic acid). Coupling of the lithium phenolate of **10** with **5** in methanol gave a low yield (24%) of a more polar product which proved to be the partially deprotected ester **11**. Rather than try to progress **11**, whose unprotected OH groups were likely to cause problems, a literature procedure for the glucuronidation of methyl salicylate<sup>13</sup> was very satisfactorily adapted (Scheme 1).

Koenigs-Knorr reaction of 5 with 10 gave the conjugate 12 in 61% yield after chromatography. The chemical shift and coupling constant (J = ca. 8) of the anomeric proton in 12 were consistent with a  $\beta$ -glucuronide. Debenzylation of 12 using catalytic transfer hydrogenation gave acid 13 in 80% yield: the condensation of 13 with 2-amino-5-nitrothiazole was performed using the water-soluble carbodiimide method shown.<sup>14</sup> Chromatography afforded product 14 in excellent purity and 67% yield. The esters were cleaved using aq. NaOH, and after acidification to pH 6 the sodium salt of glucuronide 3 precipitated in 80% yield. By high-performance liquid chromatographic analysis this material appeared identical with the authentic metabolite.

#### **Biological Data**

The antibacterial activities of compounds 1, 2 and 3 were compared. Minimum inhibitory concentrations (MICs) in the range  $1-10 \ \mu g \ cm^{-3}$  were observed for all three compounds against *Helicobacter pylori*, 3 being about tenfold

less effective than 1 or 2. Against Sarcocystis neurona, 1 and 2 showed MICs of  $2 \mu g \text{ cm}^{-3}$  while the MIC of 3 was  $40 \,\mu g \,\mathrm{cm}^{-3}$ . Against strains of the aerobic Gram-positive and Gram-negative bacteria Staphylococcus aureus, Enterococcus faecalis, Morganella morganii, Escherichia coli and Pseudomonas aeruginosa all three compounds were inactive at up to 512  $\mu$ g cm<sup>-3</sup>.

Further biological results, with a discussion of the mode of action of these compounds, will be published separately.

### Experimental

For general directions, see an earlier paper from these laboratories.<sup>15</sup> Mass spectra were recorded on a Varian-Saturn GC-ITD instrument in the electron-impact (EI) mode for compound 2, on a Kratos MS 25 instrument for chemical ionisation (CI) spectra and on a Kratos Concept 1S instrument for the fast atom bombardment (FAB) mode. Antibacterial screening was performed using either an agar dilution technique in a Wilkens Chalgren medium containing 10% blood at an inoculum of 109 colony forming units (CFU) cm<sup>-3</sup>, for the anaerobic bacteria, or in a Mueller Hinton agar medium at an inoculum of 10<sup>6</sup> CFU cm<sup>-3</sup> in Mueller Hinton broth for the aerobic bacteria.

2-Hydroxy-N-(5-nitrothiazol-2-yl)benzamide (Tizoxanide) 2.--A suspension of 2-acetoxy-N-(5-nitrothiazol-2-yl)benzamide (nitazoxanide, 1, 100 g, 0.326 mol) in 37% w/v HCl (500 cm<sup>3</sup>) was stirred and heated at 50  $^{\circ}$ C for 24 h.<sup>5</sup> The resulting slurry was cooled and filtered, then the filtrate was well washed with deionized water until the washings were neutral and dried at 50 °C to give tizoxanide 2 (85 g, 98%), mp 254 °C;  $\nu_{max}/cm^{-1}$  (Nujol) 1670;  $\delta$  [220 MHz, (CD<sub>3</sub>)<sub>2</sub>SO] 7.00–7.15 (2 H, m, ArH), 7.60 (1 H, t, ArH), 8.00 (1 H, d, 6-H) and 8.75 (1 H, s, 4'-H); m/z (Me<sub>3</sub>Si derivative, EI) 338 [MSi(CH<sub>3</sub>)<sub>3</sub><sup>+</sup>], 193 (100%, cleavage of thiazole fragment).

Methyl 1-[2-(Benzyloxycarbonyl)phenyl]-β-D-glucopyranuronate 11. -The bromosugar 5 (0.60 g, 1.5 mmol) was added in one portion to a solution of benzyl salicylate 10 (0.34 g, 1.5 mmol) and LiOH·H<sub>2</sub>O (0.063 g, 1.5 mmol) in methanol (1.5 cm<sup>3</sup>) which was stirred at 0 °C. After 1 h, the temperature having risen to 20 °C, the solution was diluted with water containing a few drops of acetic acid, then extracted with  $CH_2Cl_2$  (3 × 5 cm<sup>3</sup>). Evaporation gave crude product (0.60 g) which was chromatographed to afford the product 11 as a solid (0.15 g, 24%) which on trituration with diethyl ether and recrystallisation (methanol-diethyl ether) had mp 164-166 °C (Found: C, 60.1; H, 5.3. C<sub>21</sub>H<sub>22</sub>O<sub>9</sub> requires C, 60.3; H, 5.3%); δ [(CD<sub>3</sub>)<sub>2</sub>SO], inter alia, 3.68 (3 H, s, CH<sub>3</sub>O), 4.12 (1 H, d, 5-H), 5.23 (1 H, d, 1-H), 5.34 (2 H, m, ArCH<sub>2</sub>O) and 7.10-7.70 (9 H, m, ArH); *m/z* (CI, NH<sub>3</sub>) 436 (MNH<sub>4</sub><sup>+</sup>, 12%).

1-[2-(Benzyloxycarbonyl)phenyl]-2,3,4-tri-O-acetyl-β-D-Methvl glucopyranuronate 12.—Silver(I) oxide (2.12 g, 0.91 mmol) was added in portions to a stirred mixture of bromosugar 5 (3.30 g, 8.31 mmol) and benzyl salicylate 10 (3.78 g, 16.6 mmol) in isoquinoline (4.6 g) at 0 °C, giving a thick slurry. On warming to 20 °C over 1 h no remaining 5 was seen (TLC in 1:1 EtOAc-hexane), so the mixture was diluted with diethyl ether and filtered through Celite, then the filtrate was worked up for a neutral product, followed by evaporation to an orange oil which was washed with hexane  $(2\times)$ , decanting the mother liquors, to remove the bulk of the unreacted 10. Chromatography afforded the product 12 as a foam (2.75 g, 61%) (Found: m/z, 562.1933. C<sub>27</sub>H<sub>32</sub>NO<sub>12</sub> requires MNH<sub>4</sub><sup>++</sup>, 562.1924);  $\nu_{max}$  (CHCl<sub>3</sub>)/cm<sup>-1</sup> 1750 (vs), 1610, 1590 (sh) and 1490; δ (CDCl<sub>3</sub>) 2.09 (9 H, s, 3 × CH<sub>3</sub>CO), 3.77 (3 H, s, CH<sub>3</sub>O), 4.21 (1 H, d, 5-H), 5.20 (1 H, m, 1-H), 5.30–5.40 (3 H, m, 2-H + 3-H + 4-H), 5.37 (2 H, s, Ph*CH*<sub>2</sub>O), 7.10–7.25 (2 H, m, ArH) 7.35–7.55 (6 H, m, ArH) and 7.81 (1 H, dd, ArH); m/z (CI, NH<sub>3</sub>) 562 (MNH<sub>4</sub><sup>+</sup> 65%).

Methvl 1-(2-Carboxyphenyl)-2,3,4,-tri-O-acetyl-β-D-glucopyranuronate 13.—A solution of ester 12 (2.71 g, 4.98 mmol) in propan-2ol (75 cm<sup>3</sup>) and cyclohexene (5 cm<sup>3</sup>) was stirred and heated at gentle reflux for 0.5 h with Pd-C (0.3 g). The catalyst was filtered off, then the filtrate was evaporated to a foam which was dissolved in 4%. aq. NaHCO<sub>3</sub> (25 cm<sup>3</sup>) and washed with diethyl ether (2×). Cautious acidification of the aq. phase then extraction with  $Et_2O$  gave on evaporation the acid 13 as a colourless foam (1.84 g, 80%) (Found: C, 52.4; H, 4.9; m/z, 472.1465. C<sub>20</sub>H<sub>22</sub>O<sub>12</sub> requires C, 52.85; H, 4.8%; MNH<sub>4</sub><sup>+</sup>, 472.1455);  $\nu_{max}$  (Nujol)/cm<sup>-1</sup> 3700–2500(br), 1760 (br, s), 1610 (m) and 1495; δ (220 MHz, CDCl<sub>3</sub>) 2.00-2.10 (9 H, 3 s, 3 × CH<sub>3</sub>CO), 3.73 (3 H, s, CH<sub>3</sub>O), 4.34 (1 H, d, 5-H), 5.35-5.45 (4 H, m, 1-H to 4-H), 7.28 (2 H, m ArH), 7.62 (1 H,

t, ArH) and 8.11 (1 H, d, ArH); m/z (CI, NH<sub>3</sub>) 472 (MNH<sub>4</sub><sup>+</sup>, 100%).

1-[2-N-(5-Nitrothiazol-2-yl)carboxamido]phenyl-2,3,4-tri-Methvl O-acetyl-β-D-glucopyranuronate 14.—1-(3-Dimethylaminopropyl)-3ethylcarbodiimide hydrochloride (0.81 g, 4.25 mmol) was added to a stirred suspension of acid 13 (1.75 g, 3.85 mmol), 4-N,N-dimethylaminopyridine (0.5 g, 4.10 mmol), 1-hydroxybenzotriazole mono-hydrate (0.65 g, 4.25 mmol) and 2-amino-5-nitrothiazole (0.615 g, 4.24 mmol) in DMF (25 cm<sup>3</sup>) at 0 °C. After 2 h at 20 °C, then 16 h at 0 °C the solution was concentrated to near dryness, then extracted with  $CH_2Cl_2$  (2 × 25 cm<sup>3</sup>) and worked up for a neutral product. Evaporation gave a brown solid (2.63 g) which was chromatographed on silica. Appropriate fractions were pooled and evaporated to a sticky solid which on trituration with diethyl ether deposited the product 14 as a flaky yellow solid (1.51 g, 67%), mp 262-264 °C (Kofler block, from CH<sub>2</sub>Cl<sub>2</sub>-methanol-diethyl ether) (Found: C, 47.5; H, 4.15; N, 7.15.  $C_{23}H_{23}N_3O_{13}S$  requires C, 47.5; H, 4.0; N, 7.2%);  $\nu_{max}$  (Nujol)/cm<sup>-1</sup> 3350 (sharp), 1750, 1665, 1625 (w), 1605 (m), 1530 and 1350; δ [(CD<sub>3</sub>)<sub>2</sub>SO] 1.94, 1.98, 2.05 (9 H, 3 s, 3x CH<sub>3</sub>CO), 3.68 (3 H, s, CH<sub>3</sub>O), 4.80 (1 H, d, 5-H), 5.05 (2 H, t) and 5.48 (3 H, m, 2-H + 3-H + 4-H), 5.67 (1 H, d, 1-H), 7.20-7.30 (2 H, m, ArH), 7.55-7.70 (2 H, m, ArH) 8.71 (1 H, s, 4"-H) and 13.39 (1 H, br s, NH); *m*/*z* (CI, NH<sub>3</sub>) 582 (MH<sup>+</sup>, 100%).

1-[2-N-(5-Nitrothiazol-2-yl)carboxamido]phenyl-β-D-glucopyranosid-uronic Acid 3.—A 2.5 mol dm<sup>-3</sup> NaOH solution (5 cm<sup>3</sup>) was added in one portion to a stirred suspension of the ester 14 (1.45 g, 2.50 mmol) in methanol (17.5 cm<sup>3</sup>) at 0 °C. On warming to 20 °C over 1 h, a yellow solution resulted which was acidified to pH 6.9 with acetic acid, followed by evaporation to dryness. The residue was triturated with aq. ethanol, 1:4 (20 cm<sup>3</sup>) then the yellow solid was filtered to give the sodium salt of the product 3 (1.03 g, 89%), mp >200 °C (decomp.) from aq. ethanol (Found: m/z, 464.0367. C<sub>16</sub>H<sub>15</sub>N<sub>3</sub>O<sub>10</sub>SNa requires MH<sup>+</sup>, 464.0376);  $\nu_{max}$  (Nujol)/cm<sup>-1</sup> 3700-2500 (br), 3540, 3260, 3100 (w), 1645 (sh), 1620, 1600, 1535 and 1350;  $\delta(D_2O)$  3.53 (2 H, m) and 3.67 (1 H, t, 2-H + 3-H + 4-H), 3.83 (1 H, d, 5-H), 5.14 (1 H, d, *J* 8, 1-H), 7.16 (1 H, t, ArH), 7.29 (1 H, d, ArH), 7.54 (1 H, dt, ArH), 7.74 (1 H, dd, ArH) and 8.38 (1 H, s, 4"-H); m/z (FAB +ve ion, glycerol) 442 (MH<sup>+</sup> free acid), 464 (MH<sup>+</sup>) and 486 (MNa<sup>+</sup>). High-performance liquid chromatographic analysis of the product (C<sub>18</sub>  $\mu$ -Bondapak reversephase column, aq. acetonitrile eluent) showed an area purity of 99.25%

Received, 25th August 1998; Accepted, 25th September 1998 Paper E/8/06676K

## References

- 1 J.-F. Rossignol and H. Maisonneuve, Am. J. Trop. Med. Hyg., 1984. 33. 511.
- 2 L. Dubreuil, I. Houcke, Y. Mouton and J.-F. Rossignol, Antimicrob. Agents Chemother., 1996, 40, 2266.
- 3 R. Romerio Cabello, L. Roberto Guerroro, M. R. Muñoz Garcia and A. Geyne Cruz, Trans. R. Soc. Trop. Med. Hyg., 1997. 91. 701.
- 4 O. Doumbo, J.-F. Rossignol, E. Pichard, H. Traore, M. Dembele, M. Diakite, F. Traore and D. A. Diallo, *Am. J.* Trop. Med. Hyg., 1997, 56, 637.
- 5 J.-F. Rossignol and R. Cavier, Chem. Abstr., 1975, 83, 28216.
- 6 B. Berrang, C. E. Twine, G. L. Hennessee and F. I. Carroll, Synth. Commun., 1975, 5, 231.
- 7 G. N. Bellenback, J. W. Long, D. G. Benjamin and J. A. Lindquist, J. Am. Chem. Soc., 1955, 77, 3310. 8 K. Honma, K. Nakazima, T. Uematsu and A. Hamada, Chem.
- Pharm. Bull., 1976, 24, 394.
- 9 B. Fischer, A. Nudelman, M. Ruse, J. Herzig, H. E. Gottlieb and E. Keinan, J. Org. Chem., 1984, 49, 4988.
- A. Nudelman, J. Herzig, H. E. Gottlieb, E. Keinan and J. Sterling, *Carbohydr. Res.*, 1987, 162, 145; N. Pravdic and D. Keglevic, J. Chem. Soc., 1964, 4633.
  11 G. T. Badman, D. V. S. Green and M. Voyle, J. Organomet.
- Chem., 1990, 388, 117.
- 12 T. Yoshioka, Y. Aizawa, T. Fujita, K. Nakamura, K. Sasahara, H. Kuwano, T. Kinoshita and H. Horikoshi, Chem. Pharm. Bull., 1991, 39, 2124.
- 13 C. D. Lunsford and R. S. Murphey, J. Org. Chem., 1956, 21, 580.
- 14 J. C. Sheehan, P. A. Cruickshank and G. L. Boshart, J. Org. Chem., 1961, 26, 2525.
- 15 A. V. Stachulski, D. E. Nichols and F. Scheinmann, J. Chem. Res. (S), 1996, 30.