ORIGINAL RESEARCH



Synthesis and anti(myco)bacterial activity of novel 5,5diphenylpyrrolidine *N*-aroylthiourea derivatives and a functionalized hexahydro-1*H*-pyrrolo[1,2-*c*]imidazole

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Abstract In this paper, five novel 5,5-diphenylpyrrolidine N-aroylthiourea derivatives were synthesized by stereoselective cycloaddition of N-diphenylmethylene-protected glycine methyl ester and methyl acrylate, and subsequent coupling with aroylisothiocyanates. The cis-stereochemistry of one of the heterocyclic thiourea derivatives was characterized by single crystal X-ray diffraction studies. The compounds showed antibacterial activity against Staphylococcus aureus, Bacillus subtilis, Aeromonas hydrophila, Escherichia. coli and Acinetobacter baumannii with minimum inhibitory concentration values in the range of 62.5-1000 µg/mL against these bacterial strains. Antimycobacterial activity of the compounds was investigated against the M. tuberculosis H37Rv strain and all compounds exhibited antimycobacterial activity with a minimum inhibitory concentration value of 80 µg/mL. Additionally, methyl 5,5-diphenylhexahydro-1-oxo-3thioxo-1H-pyrrolo[1,2-c]imidazole-6-carboxylate was synthesized by cyclization reaction of the 5,5-diphenylpyrrolidine N-aroylthiourea derivatives in the presence of hydrazine monohydrate and exhibited antibacterial activity

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with a minimum inhibitory concentration value of $62.5 \,\mu g/$ mL against the same bacterial strains and exhibited antimycobacterial activity with a minimum inhibitory concentration value of $80 \,\mu g/mL$ against the *M. tuberculosis* H37Rv strain.

Keywords 5,5-Diphenylpyrrolidine · Thiohydantoin · Antibacterial activity · Antimycobacterial activity · *M. tuberculosis* H37Rv

Introduction

Nitrogen and sulfur are among the most important elements, which are present in the structure of biologically important compounds. Many drug molecules, such as amoxicillin, quetiapine, olanzapine, pioglitazone, and duloxetine contain at least one nitrogen and sulfur atom in their structures (Ilardi et al. 2014). Thiohydantoins, containing nitrogen and sulfur atoms, represent an important class of compounds within pharmaceutical chemistry and are present in a wide range of pharmacologically active compounds exhibiting antimicrobial (Khurana et al. 2014), antiviral (Khodair et al. 2001), anti-inflammatory (Blanc et al. 1992), antitumor (Al-Obaid et al. 1996), and hypoglycemic (Yildiz and Bozdag-Dundar 2010) activities. Thiohydantoins have also been used as intermediates for the synthesis of biologically active compounds (Abdelaziz et al. 2015; Metwally and Abdel-Latif 2012).

N-Substituted-*N*'-aroyl(acyl)thiourea derivatives are gaining significance in pharmaceutical, coordination and analytical chemistry (Saeed et al. 2014b; Koch 2001). *N*-Substituted-*N*'aroyl(acyl)thiourea derivatives are known to show a wide range of biological properties, such as

15-lipoxygenase inhibitory activity (Mahdavi et al. 2014), carbonic anhydrase inhibitor (Saeed et al. 2014a), antimicrobial (Saeed et al. 2013; Döndaş et al. 2006), antitumor (Saeed et al. 2010), and analgesic activity (Shoaib et al. 2014). In addition, some N-benzoyl-N'-ferrocenylthiourea derivatives exhibited cytokinin and auxin activities (Zhang et al. 2015). Furthermore, N-substituted-N'-aroyl(acyl) thioureas are also useful compounds for the synthesis of biologically important heterocyclic scaffolds, such as thiohydantoins (Nural et al. 2011) and thiazoles (Saeed et al. 2008). Ni(II) (Yang et al. 2012; Nural et al. 2009; Del Campo et al. 2004), Pt(II) (Cîrcu et al. 2009; Del Campo et al. 2004) and Co(III) (Tan et al. 2014; Del Campo et al. 2004) chelate complexes of *N*-Substituted-*N*'-aroyl(acyl) thioureas have been synthesized. In addition, these types of ligands are useful for analytical applications, such as potentiometric sensors for heavy metals (Otazo-Sanchez et al. 2002) or chemosensors for anions (Li et al. 2012; Hu et al. 2009). Moreover, some N-aroyl(acyl)thioureas were used for extraction, separation, and determination of heavy metals (Alkherraz et al. 2014; Luckay et al. 2009).

Heterocycles containing one or more nitrogen atoms are a priority area of drug research (Hernandez-Toribio et al. 2012). Pyrrolidines incorporate one of the most important scaffolds, possessing various pharmacological activities, such as antimicrobial (Schwartz et al. 1998), antiviral (Martín-Rodríguez et al. 2011; Nájera and Sansano 2009) andantitumor (Fujimoto et al. 1987) activity. Furthermore, the pyrrolidine ring is present in many synthetic or natural bioactive molecules, such as procyclidine (Ettinger et al. 2003), bepridil (Fujiki et al. 2003), elacomine (Pellegrini et al. 1996), anisomycin (Stocker et al. 2010), and hygrine (Rubioa et al. 2013). Although pyrrolidine derivatives have been intensively studied in the literature (Verma et al. 2016; Chen et al. 2013; Pandey et al. 2006; Dondas et al. 1996 2004 Döndaş et al. 2003), there are only a limited number of studies about their 5,5-diphenylpyrrolidine derivatives (Hut'ka et al. 2013; Kim et al. 2011; Tsubogo et al. 2008; Xue et al. 2008; Wang et al. 2008, Khlebnikov et al. 1997, 2001). Noteworthy, the gem-diphenyl group is present in various physiologically active nitrogen-containing compounds, such as methadone (Hassan et al. 2009), diphenhydramine (Ahmadi et al. 2012), clotrimazole (Prasanna et al. 2014), loperamide (Geyer et al. 2015), cinnarizine (Sassene et al. 2015), dextromoramide (Bye 1976), benzatropine (Othman et al. 2008), darifenacin (Pramanik et al. 2012), doxapram (Krasikovs 2015), modafinil (Okunola-Bakare et al. 2014), and phenytoin (Soman et al. 2014).

In this study, we designed and synthesized five novel 5,5-diphenylpyrrolidine *N*-aroylthioureas as precursors of a compound with a thiohydantoin moiety fused to a pyrrolidine and investigated their antibacterial and antimycobacterial activities.

Material and methods

Chemicals and general technical data

All chemicals used were high-grade commercial products purchased from Merck or Aldrich and all solvents provided from commercial suppliers as having reagent grade quality, and used without further purification.

IR spectra were recorded in the range of $4000-600 \text{ cm}^{-1}$ by a Varian Scimitar Series 1000 FT-IR spectrophotometer, using horizontal Attenuated Total Reflectance. Nuclear magnetic resonance spectra and decoupling experiments were determined at 400 MHz on a Bruker Ultrashield Plus Biospin GmbH. Chemical shifts are given in parts per million (δ) downfield from tetramethylsilane as internal standard. Spectra were determined in deuterochloroform. The following abbreviations are used; s = singlet, d =doublet, m = multiplet and brs = broad singlet. Flash column chromatography was performed using silica gel 60 (230-400 mesh). Kieselgel columns were packed with silica gel GF254. Melting points were determined on Stuart SMP3 hot stage apparatus and are uncorrected. Mass spectra were recorded by an Agilent 6460 Triple Quad LC/ MS/MS mass spectrometer. High resolution mass spectra were recorded by an LC-MS TOF electrospray ionization technique. The X-ray crystal structure data were collected by a Rigaku R-Axis Rapid-S model X-ray diffractometer.

Synthesis of cis-dimethyl 5,5-diphenylpyrrolidine-2,4dicarboxylate (3)

The compound 3 was prepared by modification of literature methods (Dondas et al. 1996, 2004, Döndas et al. 2003). To a stirred solution of methyl 2-(diphenylmethyleneamino) acetate 1 prepared according to the literature (De Brabandere et al. 2014) (0.25 g, 1 mmol) in dry toluene (10 mL) was added a solution of Et₃N (0.17 g, 1.7 mmol) in dry toluene (10 mL) and Ag₂O (0.06 g, 0.25 mmol). After 5 min, methyl acrylate 2 (0.17 g, 2 mmol) in dry toluene (20 mL) was added. The mixture was stirred at room temperature for 18 h. After completion of the reaction, the mixture was quenched sequentially with saturated aqueous sodium chloride and aqueous ammonium chloride, and extracted with ethyl acetate. The crude mixture was purified by column chromatography (Et₂O:hexane/1:8) to give **3** (0.3 g, 89%) as colorless crystals. m.p.: 127-129 °C. IR (cm⁻¹): v_{max} 3296, 3060, 2950, 1743, 1724. ¹H NMR (400 MHz, CDCl₃): δ 7.67–7.11 (m, 10H, Ar-H), 4.00 (dd, 1H, J = 6.7Hz, 3.2 Hz, 2-H), 3.84 (dd, 1H, J = 9.0 Hz, 6.6 Hz, 4-H), 3.78 (s, 3H, OCH₃), 3.34 (s, 3H, OCH₃), 2.30-2.28 (m, 2H, 3-H, 3-H'). ¹³C NMR (100 MHz, CDCl₃): δ 174.5 (C=O), 173.5 (C=O), 144.6, 143.8, 128.6 (2 × C), 128.2 (2 × C), 127.0, 126.9, 126.1 (2 × C), 126.0 (2 × C), 76.2, 58.4, 52.3

 $(2 \times C)$, 51.5, 34.3. MS (ESI, M + H⁺): *m*/z 340.2 (M + H⁺, 100). HRMS (ESI-TOF-MS): calcd. for C₂₀H₂₁NO₄ [MH]⁺ 340.1549; found 340.1548.

General procedure for the synthesis of cis-5,5diphenylpyrrolidine N-aroylthiourea derivatives (5a–e)

The novel 5,5-diphenylpyrrolidine *N*-aroylthiourea derivatives **5a–e** were synthesized by modification of the literature methods (Nural et al. 2011, 2009; Döndaş et al. 2006; Arslan and Külcü 2003) from reaction of **3** and corresponding aroylisothiocyanates **4** in acetonitrile for 36 h. The solvent was evaporated under reduced pressure and the crude mixture was purified by column chromatography (Et₂O:hexane/1:6). The requisite aroylisothiocyanates **4** were synthesized by modification of the literature method (Arslan and Külcü 2003) via reaction of potassium thiocyanate and corresponding acyl chloride in acetone at reflux temperature for 2 h. Upon reaction completion, the solvent was removed and used without any further purification.

Cis-dimethyl 1-((benzoyl)carbamothioyl)-5,5-diphenylpyrrolidine-2,4-dicarboxylate (**5a**) Yellow crystals. Yield, 0.8 g, 95%. m.p.: 178–180 °C. IR (cm⁻¹): v_{max} 3366, 3057, 2982, 1746, 1735, 1710, 1349, 1225, 1210. ¹H NMR (400 MHz, CDCl₃): δ 8.15 (brs, 1H, NH), 7.93–6.86 (m, 15H, Ar-H), 5.39 (dd, 1H, J = 8.3 Hz, 8.3 Hz, 2-H), 4.04 (dd, 1H, J = 8.4 Hz, 8.4 Hz, 4-H), 3.88 (s, 3H, OCH₃), 3.38 (s, 3H, OCH₃), 2.44 (dd, 2H, J = 8.0 Hz, 8.0 Hz, 3-H, 3-H'). ¹³C NMR (100 MHz, CDCl₃): δ 177.9 (C=S), 170.6 (C=O), 169.3 (C=O), 163.6 (C=O), 140.1, 135.7, 135.1, 133.4, 132.2, 130.5, 129.4 (2 × C), 129.0 (3 × C), 128.9 (2 × C), 128.7, 128.3, 127.8, 127.0 (2 × C), 79.8, 64.9, 61.1, 52.7, 52.1, 28.8 MS (ESI, M – H⁺): m/z 501.4 (M – H⁺, 100). HRMS (ESI-TOF-MS): calcd. for C₂₈H₂₆N₂O₅S [MH]⁺ 503.1641; found 503.1657.

Cis-dimethyl 1-((4-tert-butylbenzoyl)carbamothioyl)-5,5diphenylpyrrolidine-2,4-dicarboxylate (5b) Amorphous yellow solid. Yield, 0.5 g, 79%. m.p.: 87–89 °C. IR (cm⁻¹): *v*_{max} 3385, 3062, 2952, 1736, 1685, 1346, 1267, 1195. ¹H NMR (400 MHz, CDCl₃): δ 8.14 (brs, 1H, NH), 7.95–6.78 (m, 14H, Ar-H), 5.41(dd, 1H, J = 8.1 Hz, 8.1 Hz, 2-H), 4.03 (dd, 1H, J = 7.8 Hz, 7.8 Hz, 4-H), 3.88 (s, 3H, OCH₃), 3.39 (s, 3H, OCH₃), 2.43 (dd, 2H, J = 8.4 Hz, 8.1 Hz, 3-H, 3-H'), 1.25 (s, 9H, C(CH₃)₃). ¹³C NMR (100 MHz, CDCl₃): δ 178.0 (C=S), 170.7 (C=O), 169.4 (C=O), 163.4 (C=O), 156.0, 140.0, 135.9, 130.4, 129.4 (2 × C), 129.0 (2 × C), 128.9 (2 × C), 128.6, 127.8 (2 × C), 126.9 (2 × C), 126.0, 125.2 (2 × C), 79.8, 65.0, 61.2, 52.6, 52.0, 34.9, 31.0 (3 × C), 28.7. MS (ESI, $M - H^+$): m/z 557.2 ($M - H^+$, 100). HRMS (ESI-TOF-MS): calcd. for $C_{32}H_{34}N_2O_5S$ [MH]⁺ 559.2267; found 559.2280.

Cis-dimethyl 1-((4-methoxybenzoyl)carbamothioyl)-5.5diphenylpyrrolidine-2,4-dicarboxylate (5c) Amorphous vellow solid. Yield, 0.3 g, 71%. m.p.: 85–87 °C. IR (cm⁻¹): *v*_{max} 3348, 3057, 2955, 1736, 1688, 1354, 1253, 1207. ¹H NMR (400 MHz, CDCl₃): δ 8.07 (s, 1H, NH), 7.94–6.60 (m, 14H, Ar-H), 5.40 (dd, 1H, J = 8.2 Hz, 8.2 Hz, 2-H), 4.04 (dd, 1H, J = 7.9 Hz, 7.9 Hz, 4-H), 3.88 (s, 3H, OCH₃), 3.77 (s, 3H, OCH₃), 3.39 (s, 3H, OCH₃), 2.43 (dd, 2H, J =8.1 Hz, 8.1 Hz, 3-H, 3-H'). ¹³C NMR (100 MHz, CDCl₃): δ 178.2 (C=S), 170.7 (C=O), 169.4 (C=O), 163.1 (C=O), 162.8, 140.09, 135.9, 129.4 (2 × C), 129.1 (2 × C), 129.0 $(2 \times C)$, 128.9 $(2 \times C)$, 128.6 $(2 \times C)$, 127.8 $(2 \times C)$, 125.7, 113.5 (2 × C), 79.7, 64.9, 61.1, 55.4, 52.6, 52.0, 28.7. MS (ESI, $M - H^+$): m/z 531.2 (M - H⁺, 100). HRMS (ESI-TOF-MS): calcd. for $C_{29}H_{28}N_2O_6S$ [MH]⁺ 533.1746; found 533.1745.

Cis-dimethyl 1-((4-chlorobenzovl)carbamothiovl)-5,5diphenylpyrrolidine-2,4-dicarboxylate (5d) Amorphous vellow solid. Yield, 0.5 g, 72%. m.p.: 90–92 °C. IR (cm⁻¹): *v*_{max} 3340, 3060, 2950, 1736, 1684, 1344, 1267, 1206. ¹H NMR (400 MHz, CDCl₃): δ 8.05 (brs, 1H, NH), 7.93-6.77 (m, 14H, Ar-H), 5.38 (dd, 1H, J = 8.2 Hz, 8.2 Hz, 2-H), 4.04 (dd, 1H, J = 8.0 Hz, 8.0 Hz, 4-H), 3.88 (s, 3H, OCH₃), 3.39 (s, 3H, OCH₃), 2.44 (dd, 2H, J = 8.0 Hz, 8.0 Hz, 3-H, 3-H'). ¹³C NMR (100 MHz, CDCl₃): δ 177.8 (C=S), 170.5 (C=O), 169.3 (C=O), 163.0 (C=O), 140.1, 138.6, 135.7, 131.8, 129.5 (2 × C), 129.4, 129.0 (2 × C), 128.8, 128.7, 128.6 $(2 \times C)$, 128.5 $(2 \times C)$, 128.2, 127.8 $(2 \times C)$, 79.8, 65.0, 61.1, 52.6, 52.1, 28.7. MS (ESI, M – H⁺): *m*/*z* 535.5 $(M - H^+, 100), 537.5 (M - H^+, 35).$ HRMS (ESI-TOF-MS): calcd. for $C_{28}H_{25}ClN_2O_5S$ [MH]⁺ 537.1251; found 537.1257.

Cis-dimethyl 1-((4-phenylbenzoyl)carbamothioyl)-5,5diphenylpyrrolidine-2,4-dicarboxylate (5e) Yellow crystals. Yield, 0.3 g, 80%. m.p.: 108–110 °C. IR (cm⁻¹): v_{max} 3304, 3035, 2986, 1730, 1696, 1368, 1252, 1211. ¹H NMR (400 MHz, CDCl₃): δ 8.19 (brs, 1H, NH), 7.96-6.92 (m, 19H, Ar-H), 5.41 (dd, 1H, J = 8.2 Hz, 8.2 Hz, 2-H), 4.04 (dd, 1H, J = 8.0 Hz, 8.0 Hz, 4-H), 3.89 (s, 3H, OCH₃), 3.39 $(s, 3H, OCH_3), 2.45 (dd, 2H, J = 8.1 Hz, 8.1 Hz, 3-H, 3-H').$ ¹³C NMR (100 MHz, CDCl₃): δ 178.0 (C=S), 170.6 (C=O), 169.4 (C=O), 163.4 (C=O), 145.0, 140.1, 139.6, 135.8, 132.0, 129.4 (2 × C), 129.1, 129.0, 128.9 (4 × C), 128.7, 128.2, 127.8, 127.6 (2 × C), 127.2 (4 × C), 126.9 (2×C), 79.8, 65.0, 61.1, 52.6, 52.1, 28.7. MS (ESI, $M - H^+$): m/z 577.5 (M - H⁺, 100). HRMS (ESI-TOF-MS): calcd. for $C_{34}H_{30}N_2O_5S$ [MH]⁺ 579.1954; found 579.1975.

The synthesis of cis-methyl 5,5-*diphenylhexahydro-1-oxo-3thioxo-1H-pyrrolo*[1,2-*c*]*imidazole-6-carboxylate* (**6**)

To a stirred solution of 5a (0.502 g, 1 mmol) in dry MeOH (20 mL) was added a solution of hydrazine monohydrate (0.055 g, 1.1 mmol) in dry MeOH (10 mL) and the mixture was stirred at reflux temperature for 12 h. After completion of the reaction, the solvent was evaporated under reduced pressure, quenched with saturated aqueous sodium chloride and extracted with ethyl acetate. The crude mixture was purified by column chromatography (ethyl acetate:hexane/ 1:4) to afford compound 6. Amorphous white solid. Yield, 0.28 g, 84%. m.p.: 172–174 °C. IR (cm⁻¹): v_{max} 3202, 3069, 2921, 1746, 1716,1463, 1345, 1271, 1225.¹H NMR (400 MHz, CDCl₃): δ 8.56 (bs, 1H, NH), 8.01–7.98 (m, 2H, Ar-H), 7.51–6.86 (m, 8H, Ar-H), 4.60 (dd, 1H, J = 12.4 Hz, 6.1 Hz, 7a-H), 4.24 (dd, 1H, J = 12.3 Hz, 7.3 Hz, 6-H), 3.25 (s, 3H, OCH₃), 2.90 (dt, 1H, J = 12.6 Hz, 12.3 Hz, 7-H),2.60 (ddd, 1H, J = 13.1 Hz, 7.0 Hz, 6.0 Hz, 7-H'). ¹³C NMR (100 MHz, CDCl₃): δ 178.5 (C=S), 171.2 (C=O), 169.7 (C=O), 142.6, 137.3, 128.7 (3 × C), 128.5 (3 × C), 128.2, 128.1, 127.6 (2 × C), 76.1, 66.1, 62.6, 52.2, 29.4. MS (ESI, M+H⁺): *m*/*z* 367.1 (M+H⁺, 100). HRMS (ESI-TOF-MS): calcd. for $C_{20}H_{18}N_2O_3S$ [MH]⁺ 367.1116; found 367.1130.

Crystallography

For the crystal structure determination, single-crystal of 5a, obtained from Et₂O:hexane 1:6 as crystallization solvent was used for data collection on a four-circle Rigaku R-AXIS RAPID-S diffractometer (equipped with a twodimensional area IP detector). Graphite-monochromated Mo-K_{α} radiation ($\lambda = 0.71073$ Å) and oscillation scans technique with $\Delta w = 5^{\circ}$ for one image were used for data collection. The lattice parameters were determined by the least-squares methods on the basis of all reflections with F^2 $> 2\sigma(F^2)$. Integration of the intensities, correction for Lorentz and polarization effects and cell refinement was performed using CrystalClear (2005) software. The structures were solved by direct methods using SHELXS-97and refined by a full-matrix least-squares procedure using the program SHELXL-97 (Sheldrick 1997). All non-hydrogen atoms were refined anisotropically and H atoms were positioned geometrically and refined using a riding model. The final difference Fourier maps showed no peaks of chemical significance. Crystal data for 5a: C₂₈H₂₆N₂O₅S, crystal system, space group: triclinic, P-1; (no:2); unit cell dimensions: a = 14.190(2), b = 13.043(2), c = 14.261(2) Å, $\alpha = 90, \beta = 103.451(4), \gamma = 90^{\circ}$; volume: 2567.0(6) Å³; Z = 2; calculated density: 1.30 g/cm^3 ; absorption coefficient: 0.167 mm⁻¹; F(000): 1056; θ -range for data collection 3.1-25.9°; refinement method: full matrix least-square on F^2 ; data/parameters: 5040/328; goodness-of-fit on F^2 : 0.95; final *R*-indices $[I > 2\sigma(I)]$: $R_1 = 0.068$, w $R_2 = 0.088$; largest diff. peak and hole: 0.179 and -0.203 e/Å^3 .

Crystallographic data that were deposited in the Cambridge Structural Database under CCDC-1476027 registration number contain the supplementary crystallographic data for this Letter. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre (CCDC) via www.ccdc.cam.ac.uk/data_request/cif and are available free of charge upon request to CCDC, 12 Union Road, Cambridge, UK (fax: +441223 336033, e-mail: deposit@ccdc.cam.ac.uk).

Antibacterial activity

Stock solutions of 5a-e, 6 were prepared by dissolving the compounds in dimethyl sulfoxide (DMSO) and then diluting in Mueller-Hinton broth and Triptic soy broth to give an initial concentration of 2000 µg/mL. Further dilutions of the compounds and ampicillin, which was used as control drug in the test medium were prepared at the concentrations of 2000, 1000, 500, 250, 125, 62.5, 31.25, 15.62, 7.8, 3.9, and 1.9 µg/mL. To ensure that the solvents had no effect on microbial growth, a control test was performed containing inoculated broth supplemented with DMSO at the same dilutions used for the test compounds, and was determined to be inactive. MIC values for each compound were determined in duplicate against Staphylococcus aureus (ATCC 25925) and Bacillus subtilis (ATCC 6633) as Gram (+) bacterial strains and Aeromonas hydrophila (ATCC 95080), Escherichia coli (ATCC 25923) and A. baumannii (ATCC 02026) as Gram (-) bacterial strains obtained from the Refik Saydam Hıfzıssıhha Institute, Ankara, Turkey. The observed data on the antibacterial activity of 5a-e, 6 and ampicillin are reported in Table 1.

Antimycobacterial activity

Agar proportion method

MIC values of synthesized compounds **5a–e**, **6** were determined by agar dilution in duplicate as recommended by the Clinical Laboratory Standards Institute (CLSI) (NCCLS -M24-A 2003; NCCLS -M24-T 2002). Positive and negative growth controls were run in each assay. Isoniazid (INH) (Sigma I3377) and ethambutol (EMB) (Sigma E4630) were used as control agents. *M. tuberculosis* H37Rv was used as the standard strain and was provided by the Refik Saydam National Public Health Agency, National Tuberculosis Reference Laboratory, Ankara, Turkey. Stock solutions of **5a–e**, **6** and reference compounds were prepared in DMSO at a concentration of 1000 μ g/mL. These solutions were then filtered through a 0.22- μ m membrane filter (Millipore, USA). Middlebrook 7H10 agar medium

Table 1 The MIC values (µg/ mL) of the tested compounds against the (myco)bacterial strains

Compound	S. aureus	B. subtilis	A. hydrophila	E. coli	A. baumanii	M. tuberculosis H37Rv
5a	125	62.5	125	125	62.5	80
5b	1000	250	500	1000	500	80
5c	125	500	250	125	125	80
5d	125	125	62.5	125	62.5	80
5e	500	500	500	500	500	80
6	62.5	62.5	62.5	62.5	62.5	80
Ampicillin	31.25	0.9	31.25	15.62	125	
Isoniazid						0.2
Ethambutol						5

MIC the minimal inhibitory concentrations determined in duplicate with deviations within one two-fold dilution

(BBL, Becton Dickinson and Company, Sparks, MD, USA) was supplemented with oleic acid-albumin-dextrosecatalase (OADC, BBL, Becton Dickinson and Company, Sparks, MD, USA). Compounds **5a–e**, **6** and control agents were added to obtain an appropriate final concentration in the medium. The final concentrations of INH and EMB were 0.2–1 and 5–10 µg/mL, respectively. Compounds **5a– e**, **6** were prepared at final concentrations of 5, 10, 20, 40, 80, and 160 µg/mL.Three milliliters of prepared medium without any references and synthesized compounds **5a–e**, **6** were dispensed into a sterile tube and it was used as a growth control. The DMSO concentration in the final solutions was not above 1% for antimycobacterial activity.

Inoculum preparation

H37Rv was maintained in Lowenstein-Jensen medium. A culture suspension was prepared by subculturing in Middlebrook 7H9 broth (BBL, Becton Dickinson and Company, Sparks, MD, USA) supplemented with 10% OADC at 37 °C for 7–10 days, until a density corresponding to 10^{-2} dilution was obtained from McFarland standard No. 1. Then 0.1 mL of the diluted suspension was inoculated onto the control and the other tubes with compounds 5a-e, 6 in different concentrations. The tubes were incubated at 37 °C in an atmosphere of 5% CO₂ for 3 weeks. The MIC values were defined as the lowest concentration that inhibited more than 90% of the bacterial growth, and the results of INH and EMB were interpreted according to the CLSI. The MIC was considered the lowest concentration that showed no visible colonies in all dilutions. In case duplicates did not give an identical result and indicated different MIC values within one two-fold dilution, the highest of the different values was defined as the MIC.

Results and discussion

Cis-5,5-diphenylpyrrolidine *N*-aroylthiourea derivatives **5a–e** were synthesized from reaction of *cis*-5,5-

diphenylpyrrolidine-2,4-dicarboxylate **3**, which was stereoselectively prepared as the sole and pure diastereomer by 1,3-dipolar *endo*-cycloaddition reaction of methyl 2-(diphenylmethyleneamino)acetate **1** (De Brabandere et al. 2014) and methyl acrylate **2**, and 4-substituted-benzoyl isothiocyanate **4**, prepared by reaction of potassium thiocyanate and the corresponding acyl chloride by modification of the literature method (Arslan and Külcü 2003), in good to excellent yields (Scheme 1). In the cycloaddition reaction, the formation of the *trans*-isomer of 5,5-diphenylpyrrolidine-2,4-dicarboxylate **3** was not observed.

The cis-methyl 5,5-diphenylhexahydro-1-oxo-3-thioxo-1H-pyrrolo[1,2-c]imidazole-6-carboxylate **6** was synthesized by cyclization reaction of the cis-5,5-diphenylpyrrolidine N-aroylthioureas 5a-e with concomitant cleavage of the *N*-acyl group in the presence of hydrazine monohydrate in refluxing dry MeOH for 12 h (Scheme 1). The highest yield (84%) was obtained from the cyclization reaction of 5a and the lowest yield (30%) was obtained from the cyclization reaction of 5c in the presence of hydrazine monohydrate (Table 2). After all of these cyclization reactions, starting pyrrolidine 3 was found in the reaction mixture and could be recovered. The most recovery of pyrrolidine 3 (55%) was observed in the cyclization reaction of 5c. Therefore, the decreasing yield can be explained by cleavage of the 4substituted-N-thioformylbenzamide group. Structures of 3, 5a-e and 6 were fully characterized by ¹H and ¹³C NMR, FT-IR, MS and HRMS studies. The stereochemistry was unambiguously determined for **5a** by single crystal X-ray diffraction studies (Fig. 1). Additionally, analogous ¹H NMR chemical shifts (2-H: $\delta = 5.39-5.41$ ppm, 3-H and 3'-H: $\delta = 2.43 - 2.45$ ppm, 4-H: $\delta = 4.03 - 4.04$ ppm) and the corresponding coupling constants (Table 3) of all derivatives 5 pointed out the *cis*-stereochemistry.

The structure of **5a** was fully characterized by single crystal X-ray diffraction study. Compound **5a** crystallized as yellow prisms and the structure was solved in the triclinic centrosymmetric space group $P\overline{1}$ with one molecule in the asymmetric unit (Fig. 1). There are two stereogenic carbon





Table 2 Synthesis of hexahydro-1*H*-pyrrolo[1,2-c]imidazole 6

Entry	X	Yield (%)	
5a	Н	84	
5b	CMe3	42	
5c	OMe	30	
5d	Cl	82	
5e	Ph	46	

atoms and both have *S*-configuration (C14 and C16). The bond length of N1-C21 = 1.351(5) Å, N2-C21 = 1.393(5)Å and N2-C22 = 1.387(5) Å are shorter than typical N–C single bond values. Whereas, the bond S–C(21) = 1.640(4)Å and O(1)–C(22) = 1.203(5) Å are considerably longer than C=S and C=O double bonds, respectively. The bond values are consistent with the literature and these results show us the existence of a partial electron delocalization in the N–C(S)–NH–C(O) structural fragment (Saeed et al. 2010; Döndaş et al. 2006; Del Campo et al. 2004).

Antibacterial and antimycobacterial activities

Antibacterial activity studies of the compounds **5a–e**, **6** were performed against five standard bacterial strains. The compounds **5a–e** inhibited the growth of bacteria at minimum

inhibitory concentrations (MIC) of 62.5 to 1000 µg/mL (Table 1). 5a, 5c, and 5d, containing H, OMe and Cl as substituent at the 4-position of the aromatic ring, respectively, exhibited antibacterial activity against the S. aureus strain with a MIC value of 125 µg/mL. For the same bacterial strain, 5b containing CMe3 and 5e containing Ph as substituent at the 4position of the aromatic ring exhibited antibacterial activity with a MIC value of 1000 and 500 µg/mL, respectively. Ampicillin exhibited activity with a MIC value of 31.25 µg/ mL against the S. aureus strain. Compared to ampicillin, 5a-e showed moderate antibacterial activity against the B. subtilis strain. Compared to the other compounds, 5d showed the best activity against the A. hydrophila strain with a MIC value of 62.5 µg/mL, while MIC values of the other compounds were observed in the range of 125-500 µg/mL. Compounds 5a, 5c, and 5d exhibited antibacterial activity against the E. coli strain with a MIC value of 125 µg/mL, whereas the other compounds 5b and 5e exhibited bioactivity with a MIC value of 1000 and 500 µg/mL, respectively. The compounds 5a, 5d showed higher activity than the control against the A. baumanii strain, with a MIC value of 62.5 µg/mL. Compound 5c showed activity with a MIC value of 125 µg/mL similar to ampicillin. Finally, derivatives 5b and 5e showed activity with a MIC value of 500 µg/mL against the A. baumanii strain.

Compound 6 exhibited antibacterial activity against the tested ATCC strains with a MIC value of $62.5 \,\mu\text{g/mL}$. Thus,

Fig. 1 Crystal structure of 5a. Selected bond lengths (Å) and angles: S–C21 1.640(4), O(1)–C (22) = 1.203(5), N1–C7 1.511 (5), N1–C16 1.458(5), N1–C21 1.351(5), N2–C21 1.393(5), N2–C22 1.387(5), S–C21–N1 123.6(3), S–C21–N2 123.1(3), C22–N2–C21 128.4(3), N1–C7–C14 99.1(5)



 Table 3
 Selected ¹H NMR chemical shifts and coupling constants of pyrrolidines 5

Compound	2-Н		3-н, 3'-н		4-H	
	δ (ppm)	J (Hz)	δ (ppm)	J (Hz)	δ (ppm)	J (Hz)
5a	5.39	8.3	2.44	8.0	4.04	8.4
5b	5.41	8.1	2.43	8.4, 8.1	4.03	7.8
5c	5.40	8.2	2.43	8.1	4.04	7.9
5d	5.38	8.2	2.44	8.0	4.04	8.0
5e	5.41	8.2	2.45	8.1	4.04	8.0

the 5,5-diphenylpyrrolidine *N*-aroylthiourea derivatives possessed a broad spectrum of antibacterial activity, and it can be said that 5a and 5d showed a better pharmacological activity than the other derivatives. The pyrrolidine-fused thiohydantoin 6 showed better antibacterial activity than the 5,5-diphenylpyrrolidine *N*-aroylthiourea derivatives **5** against the tested ATCC strains.

Antimycobacterial activity studies of the compounds **5a**e, **6** were performed against the *M. tuberculosis* H37Rv strain, and all compounds exhibited activity with a MIC value of 80 µg/mL against the *M. tuberculosis* H37Rv strain (Table 1). It can be said that the presence of the *tert*-butyl, chloro, methoxy, and phenyl substituent on para position of the phenyl ring of the benzoyl group has no effect on bioactivity.

Conclusion

In summary, we have demonstrated the synthesis of a pyrrolidine-fused thiohydantoin as a single stereoisomer by

cyclization reaction of the 5,5-diphenylpyrrolidine *N*aroylthiourea derivatives in the presence of hydrazine monohydrate. These type of structures are very useful in pharmaceutical studies because the different functional groups of the synthesized hexahydro-1*H*-pyrrolo[1,2-*c*] imidazole compound can be easily modified. Although all compounds **5a–e** and **6** showed antibacterial activity, the final compound **6** exhibited better activity and its antibacterial activity is promising for future studies. Furthermore, the synthesized novel 5,5-diphenylpyrrolidine, *N*aroylthioureas and the hexahydro-1*H*-pyrrolo[1,2-*c*]imidazole compounds showed a considerable antimycobacterial activity against the *M. tuberculosis* H37Rv strain.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no competing interests.

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