Tandem Cleavage of Hydrogenated β - and γ -Carbolines – New Practical Synthesis of Tetrahydroazocino[4,5-*b*]indoles and Tetrahydroazocino[5,4-*b*]- indoles Showing Acetylcholinesterase Inhibitory Activity

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Keywords: Acetylcholinesterase inhibition / Azocine / Fused-ring systems / Nitrogen heterocycles / Ring expansion

Hydrogenated γ -carbolines underwent tandem piperidine ring cleavage on treatment with dimethyl acetylenedicarboxylate (DMAD) or ethyl propiolate (EP) in the presence of alcohols, producing 3-alkoxymethyl-substituted indoles in high yields. These compounds were cyclized to tetrahydroazocino[4,5-*b*]indoles in the presence of AlCl₃. Hydrogenated β carbolines produced tetrahydroazocino[5,4-*b*]indoles directly upon treatment with EP in ethanol. The resulting azocinoindole derivatives were subjected to a preliminary evaluation of their in vitro acetylcholinesterase (AChE) inhibitory activities. Most of them were found to inhibit AChE with IC_{50} values in the micromolar range, compound **17** being the most potent ($IC_{50} = 8.7 \mu m$).

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Introduction

Because of their wide occurrence in many biologically active compounds,^[1,2] eight-membered (azocine) and larger *N*-containing fused ring systems deserve more investigation efforts from the point of view of organic chemistry. The azocine fragment, for example, occurs in many indole alkaloids,^[3-6] but one barrier to their exploitation is a general lack of satisfactory preparative methods. Known synthetic pathways toward this class of indole derivatives include ring expansion of azetopyridoindoles through the [1,2]-Meisenheimer rearrangement of the corresponding Noxides,^[7] Fisher indolization of the appropriate azacyclanones.^[8] cationic domino processes.^[9] and catalytic hydrogenation of the 2-(2-benzylaminoethyl)-3-cyanoethylindoles.^[10] These procedures are either based on poorly available (and thus very expensive) starting materials or require relatively inefficient, multi-step procedures. There is therefore a need for new synthetic approaches to produce azocine-containing heterocycles from readily available compounds.

We have recently reported tandem piperidine ring cleavage in tetrahydropyrrolopyridines (THPPs) on treatment with dimethyl acetylenedicarboxylate (DMAD) in aprotic solvents, resulting in the formation of α - and β -vinylpyrroles^[11,12] or parallel pyrroloazocine formation in low yields.^[13] THPP derivatives were also demonstrated to undergo cleavage in alcohols on treatment with DMAD, producing β -alkoxy(hydroxy)alkylpyrroles in moderate to high yields.^[14] Here we report on a new practical synthesis of tetrahydroazocino[5,4-*b*]indole and tetrahydroazocino[4,5-*b*]indole derivatives, starting from readily obtainable hydrogenated β - and γ -carbolines and DMAD or ethyl propiolate. The preliminary results from an evaluation of their in vitro activities as acetylcholinesterase inhibitors (AChE-Is) are also reported.

Results and Discussion

The starting γ -carbolines 1-4 were obtained by the well known Fischer indolization of the appropriate *N*-alkyl-4-piperidones (see Scheme 1, Table 1).



Scheme 1

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Table 1. R substituents for carbolines 1-8, alkoxymethylindoles 9-15, and azocinoindoles 17-22

Carbolines			Alkoxymethylindoles				Azocinoindoles			
	R	\mathbb{R}^1		R	\mathbb{R}^1		R	\mathbb{R}^1	\mathbb{R}^2	R ³
1	Et	Н	9	Et	F	17	Et	F	COOMe	COOMe
2	Et	F	10	iPr	F	18	iPr	F	COOMe	COOMe
3	iPr	F	11	Et	OMe	19	Et	OMe	COOMe	COOMe
4	Et	OMe	12	Et	Н	20	Et	Н	COOMe	COOMe
5	Me	_	13	Et	Н	21	Et	Н	Н	COOEt
6	<i>i</i> Pr	_	14	Et	OMe	22	Et	F	Н	COOEt
7	Me	_	15	Et	F					
8	iPr	_								

N-Ethyl- β -carbolines **7** and **8** were synthesized by Pictet–Spengler condensation of tryptamine with acetaldehyde or isobutyraldehyde, followed by *N*-ethylation of the intermediate tetrahydro- β -carbolines **5** and **6** (Scheme 2) (Table 1). All reactions proceeded smoothly, giving the desired products in good to high yields, except for the *N*-ethylation of **6**; the isolation of the target product required column chromatography in this case. The main by-product isolated was the N(9)–Et isomeric carboline **8a**. The structure of **8a** was assigned from its IR and ¹H NMR spectroscopic data as well as its mass spectrum. The proton NMR spectrum of **8a** lacks the N(9)–H signal, which in the case of **8** resonates at $\delta = 8.21$ ppm (br. s, 1 H). We relate this unusual alkylation to the presence of the bulky isopropyl group in the position α to the piperidine NH in **6**.



Scheme 2

 γ -Carbolines, unlike THPP derivatives, are cleaved by DMAD in aprotic solvents, producing multicomponent mixtures of products, which were not identified. Compounds 1-4 and 7 and 8 were treated with DMAD or ethyl propiolate (EP) at room temperature in absolute methanol or ethanol. In the case of γ -carbolines 1-4 a tandem cleavage process involving one molecule of solvent took place (Scheme 3).



Scheme 3

In the case of carboline 1, the target azocinoindoles 20 and 21 were isolated as almost the only products; the formation of the corresponding β -alkoxymethylindoles 12 and 13 in the course of the reaction was demonstrated with the aid of LCMS analysis of the crude reaction mixture. We presume that the reaction proceeds via the intermediate zwitterion A, the result of Michael addition of the tertiary nitrogen to DMAD or EP. The cleavage of the C(1)-N bond occurs through the formation of the six-membered transition state **B**, in which a molecule of alcohol facilitates the S_N reaction. The electronic effects of R^1 define the symmetry of the transition state (loose or tight) and the direction of further transformations: the formation of alkoxymethylindoles or azocinoindoles. EP is more active than DMAD in the reaction with carbolines; this may be explained in terms of different delocalization of the negative charge in the anionic part of **B**. The proton NMR spectra of indoles 9-11 and 14 and 15 have similar characteristics: the enamine protons of 9–11 resonate at $\delta = 4.50-4.70$ ppm, whereas the enamine protons of 14 and 15 appear as two doublets at $\delta = 4.70 - 4.82$ ppm and $\delta = 7.45 - 7.62$ ppm with J = 13.0 - 13.8 Hz, such high vicinal coupling constant values indicating the trans configuration of the enamine fragment. This observation is in agreement with the data on cleavage of tetrahydropyrrolopyridines (THPPs) on treatment with DMAD in protic solvents.^[14]

Compounds 9-11 and 15 underwent intramolecular cyclization on treatment with AlCl₃ in acetonitrile to give

the target azocinoindoles 17-19 and 22 in good (except for 19) yields (Scheme 4).



Scheme 4

Our attempts to isolate the target azocinoindole failed in the case of the cyclization of **14**. The reaction was accompanied by significant tarring, and a multicomponent reaction mixture was formed.

We relate this result (along with the low yield of azocine in the case of 11) to the presence of the OCH₃ group at the C(8) position. This can be alternatively attacked by the molecule of AlCl₃, thus destabilizing the transition state **B** and activating the methoxy group towards intermolecular nucleophilic displacement.

An X-ray crystallographic analysis was carried out on the tricyclic tetrahydroazocino[4,5-*b*]indole **17**, obtained as a suitable monocrystal by recrystallisation from ethyl acetate through slow evaporation at room temperature, to obtain unequivocal structural assignment and to elucidate its three-dimensional structure. The refined X-ray crystal structure of **17** is shown in Figure 1.



Figure 1. X-ray crystal structure of 17

The conformation of the eight-membered ring is a twisted boat, in which the C(1), C(7), N(1), and C(4) atoms are almost coplanar, whilst the C(2), C(3), C(5), and C(6) atoms are located on the same side of this plane within 0.78, 1.26, 0.45, and 0.84 Å, respectively. The N(1)–C(4) bond is

shorter than the N(1)–C(3) (1.364 and 1.469 Å respectively), indicating the presence of conjugation in the enamine [N(1)–C(4)–C(5)] fragment. β -Carbolines 7 and 8, upon treatment with EP in absolute ethanol, directly provided target tetrahydroazocino[5,4-*b*]indoles 23 and 24 in good yield; in the case of 8 the corresponding α -(ethoxy)isobutylindole derivative 16 was also isolated (Scheme 5).



Scheme 5

The three-dimensional structure of compound **24** was also investigated by X-ray diffraction method. The monocrystal was obtained by recrystallisation from an ethyl acetate/hexane mixture by slow evaporation at room temperature, and the refined X-ray crystal structure of **24** is shown in Figure 2. The conformation of the eight-membered ring is a flat boat, in which the C(7), C(13), N(2), and C(10) atoms are coplanar, whilst the C(9), C(11), C(12), and C(19) atoms are located on the same side of this plane, within 1.09, 0.41, 0.90 and 2.42 Å, respectively.

The isopropyl group in azocine moiety is pseudoaxially oriented. The N(2)-C(10) bond is shorter than the N(2)-C(9) (1.364 and 1.469 Å, respectively), once again indicating the presence of conjugation in the enamine [N(2)-C(10)-C(11)] fragment.

We evaluated the synthesized azocinoindoles for their abilities to inhibit AChE, in the context of a research program aimed at testing the neuroprotective potential of new heterocyclic candidates.^[15] AChE inhibition represents a major pharmacological approach to the treatment of Alzheimer's disease (AD).^[16]

Our azocinoindoles were preliminarily tested as inhibitors of bovine AChE, by Ellman's method,^[17] and their mean inhibitory concentrations (IC₅₀s) were estimated from the best-fitting inhibition-concentration curves, as shown in Figure 3 for compound **17**. The biological data showed compound **17** to be the best AChE inhibitor of the tetrahydroazocino[4,5-*b*]indole derivatives tested, its IC₅₀ value being 8.7 μ M. The other derivatives, including tetrahydroazocino[5,4-*b*]indoles **23** and **24**, showed IC₅₀s ranging from 20 to 50 μ M. Detailed kinetic studies and structure-activity relationships will be reported elsewhere in due course.



Figure 2. X-ray crystal structure of 24



Figure 3. AChE inhibition curve for compound 17; interpolation of the best-fitting curve gave an IC₅₀ value of 8.7 μ M; data points and error bars represent means and s.d. (n = 3-5), respectively

Conclusion

In conclusion, we offer a new, two-step procedure for the synthesis of azocino[4,5-*b*]indoles and azocino[5,4-*b*]indoles, starting from readily available γ - and β -carbolines. This reaction could represent a new powerful tool for the construction of fused azocine derivatives, shown to be endowed with different degrees of inhibitory activity toward AChE.

Experimental Section

General Remarks: All solvents were distilled and dried before use; DMAD and EP were purchased from Acros Organics and were used without any additional purification. Column chromatography was performed with alumina oxide 60 from Fluka. ¹H and ¹³C NMR spectra were recorded in CDCl₃ solutions, at 25 °C or in [D₆]DMSO solutions at 40 °C, with a Bruker WM 400 NMR spectrometer operating at 400 and 100 MHz, respectively; peak positions are given in parts per million (δ) with tetramethylsilane as the internal standard. For the protons indexes for compounds 9-11 and 14, 15 see Scheme 3. ¹H-¹H COSY experiments were used for the assignment of the signals in the spectra of compounds 16 and 24. Mass spectra were obtained by the EI technique (Finnigan-MAT 95 XL engine) or the ESI method (Agilent 1100 Series LC/ MSD Trap System VL). IR spectra were recorded with a Perkin-Elmer Spectrum One instrument. Only noteworthy IR absorptions (cm⁻¹) are listed. Melting points were determined in a capillary tube and are uncorrected.

Starting Materials: γ -Carbolines 1–4 were synthesized by the previously described method,^[18] and β -carbolines 5 and 6 were synthesized by a previously described procedure.^[19] The structures of compounds 1–6 were confirmed by ¹H NMR and MS data.

2-Ethyl-1-methyl-1,2,3,4-tetrahydro-\beta-carboline (7): Ethyl iodide (1.3 g, 8 mmol) was added to a stirred solution of **5** (1.0 g, 5.4 mmol) and K₂CO₃ (1.6 g, 16 mmol) in DMF (10 mL), and stir-

ring was continued at 50 °C for 5 h. The crude reaction mixture was poured into water (20 mL) and extracted with CH₂Cl₂ (3 × 70 mL). Solvents were evaporated under reduced pressure to give 7 (600 mg, 52%) as a light brown oil. ¹H NMR (CDCl₃): δ = 1.16 (t, J = 7.3 Hz, 3 H, CH_3 -CH₂), 1.40 (d, J = 6.7 Hz, 3 H, CH_3 -CH), 2.61–2.95 (m, 5 H, CH_2 -CH₃, CH₂-3, 4-H), 3.18–3.20 (m, 1 H, 4-H), 3.85 (q, J = 7.3 Hz, 1 H, 1-H), 7.06–7.1 (m, 2 H, 6-H, 7-H), 7.3 (d, J = 7.9 Hz, 1 H, 5-H), 7.50 (d, J = 7.6 Hz, 1 H, 8-H), 7.75 (br. s, 1 H, NH) ppm. EI MS: m/z (%) = 214 (20) [M⁺], 199 (100), 157 (40), 144 (10), 130 (15). C₁₄H₁₈N₂ (214.31): calcd. C 78.46, H 8.47, N 13.07; found C 78.32, H 8.53, N 12.95.

2-Ethyl-1-isopropyl-1,2,3,4-tetrahydro-B-carboline (8) and 9-Ethyl-1isopropyl-1,2,3,4-tetrahydro-β-carboline (8a): Ethyl iodide (1.1 g, 7 mmol) was added to a stirred solution of 6 (1.0 g, 4.7 mmol) and K₂CO₃ (1.4 g, 14 mmol) in DMF (10 mL), and stirring was continued at 50 °C for 6 h. The crude reaction mixture was poured into water (20 mL) and extracted with CH_2Cl_2 (3 × 70 mL). Solvents were evaporated under reduced pressure, and the residue (750 mg) was purified by column chromatography with ethyl acetate/heptane (1:3) as eluent. The first fraction provided 8 (590 mg, 53%) as a yellow oil. ¹H NMR (CDCl₃): $\delta = 1.05$ (d, J = 6.8 Hz, 3 H, CH_3 -CH), 1.13 (t, J = 7.3 Hz, 3 H, CH_3 -CH₂), 1.21 (d, J = 6.8 Hz, 3 H, CH_3 -CH), 2.0 (m, 1 H, CH iPr), 2.45-2.60 (m, 3 H, CH₂-CH₃, 4-H), 2.90 (m, 1 H, 4-H), 3.05 (m, 1 H, 3-H), 3.20-3.30 (m, 2 H, 1-H, 3-H), 7.10 (t, J = 7.6 Hz, 1 H, 6-H), 7.15 $(t, J = 7.6 \text{ Hz}, 1 \text{ H}, 7-\text{H}), 7.25 (d, J = 7.6 \text{ Hz}, 1 \text{ H}, 5-\text{H}), 7.50 (d, J = 7.6 \text{ Hz}, 1 \text$ J = 7.6 Hz, 1 H, 8-H), 7.71 (br. s, 1 H, NH) ppm. ESI MS: m/z =243 [M⁺ + 1]. C₁₆H₂₂N₂ (242.36): calcd. C 79.29, H 9.15, N 11.56; found C 78.98, H 9.13, N 11.32.

Further elution provided **8a** (100 mg, 9%) as light yellow crystals, m.p. 142–143 °C. ¹H NMR (CDCl₃): $\delta = 0.85$ (d, J = 6.6 Hz, 3 H, CH_3 –CH), 1.01 (t, J = 7.1 Hz, 3 H, CH_3 –CH₂), 1.08 (d, J = 6.6 Hz, 3 H, CH_3 –CH), 1.85 (td, J = 13.2, 4.2 Hz, 1 H, 4-H_a), 2.27–2.28 (m, 2 H, CH_2 –N), 2.45 (m, 1 H, CH *i*Pr), 2.60–2.71 (m, 2 H, 4-H_e, 3-H_a), 3.07 (d, J = 11.0 Hz, 1 H, 1-H), 3.51 (td, J = 12.3, 2.8 Hz, 1 H, 3-H_e), 7.20 (t, J = 7.4 Hz, 1 H, 6-H), 7.30–7.40 (m, 2 H, 5-H, 7-H), 7.52 (d, J = 7.6 Hz, 1 H, 8-H) ppm. ESI MS: m/z = 243 [M⁺ + 1]. $C_{16}H_{22}N_2$ (242.36): calcd. C 79.29, H 9.15, N 11.56; found C 79.13, H 9.18, N 11.64.

General Synthetic Procedure for the Synthesis of Alkoxyindoles 9-11, 14, and 15: DMAD or EP (1.2 mmol) was added to a solution of the carboline derivative 2-4 (1 mmol) in methyl (ethyl) alcohol (10 mL). The reaction mixture was stirred for 4-6 h at room temperature (TLC monitoring). The solvent was evaporated under reduced pressure. The resulting residue was purified by column chromatography with ethyl acetate/hexane (1:2) as eluent. The first fraction provided the corresponding β -alkoxy derivatives 9-11 and 14 and 15.

Dimethyl (2*E*)-2-(Ethyl{2-[5-fluoro-3-(methoxymethyl)-1*H*-indol-2yl]ethyl}amino)but-2-enedioate (9): Colorless oil. Yield 270 mg (70%). ¹H NMR (CDCl₃): $\delta = 1.07$ (t, J = 7.3 Hz, 3 H, CH_3 -CH₂) 2.95 (q, J = 7.3 Hz, 2 H, CH_2 -CH₃), 2.95 (t, J = 6.7 Hz, 2 H, CH₂-α), 3.39 (s, 3 H, OCH₃), 3.42 (t, J = 6.7 Hz, 2 H, N-CH₂- β), 3.67 (s, 3 H, OCH₃), 3.95 (s, 3 H, OCH₃), 4.56 (s, 2 H, O-CH₂), 4.74 (s, 1 H, H-ε), 6.90 (ddd, J = 11.7 Hz, 9.2 Hz, 2.4 Hz, 1 H, CH_{6'-indolyl}), 7.20-7.27 (m, 2 H, CH_{4'-indolyl}, CH_{7'-indolyl}), 8.32 (br. s, 1 H, NH) ppm. ¹³C NMR: $\delta = 12.5$, 22.1, 46.3, 51.2, 52.3, 55.0, 57.1, 66.3, 86.2, 103.2 (d, ² $J_{C,F} = 26$ Hz), 107.9 (d, ² $J_{C,F} = 26$ Hz), 110.4 (d, ³ $J_{C,F} = 9$ Hz), 110.5, 131.4, 135.2, 136.8, 150.7, 157.3 (d, ¹ $J_{C,F} = 230$ Hz), 168.2, 169.1 ppm. EI MS: m/z (%) = 392 (6) [M⁺], 333 (21), 301 (79), 271 (18), 241 (10), 200 (100), 185 (15), 133 (11), 59 (15). $C_{20}H_{25}FN_2O_5$ (392.42): calcd. C 61.21, H 6.42, N 7.14; found C 61.30, H 6.51, N 7.35.

Dimethyl (2E)-2-[{2-[5-Fluoro-3-(methoxymethyl)-1H-indol-2-y]]ethyl}(isopropyl)amino|but-2-enedioate (10): White crystals, m.p. 100-102 °C (ethyl acetate/hexane), yield 230 mg (56%). ¹H NMR $(CDCl_3)$: $\delta = 0.95$ (d, J = 6.4 Hz, 3 H, CH_3 *i*Pr), 1.01 (d, J = 6.4Hz, 3 H, CH₃ *i*Pr), 3.05 (t, J = 7.2 Hz, 2 H, N-CH₂- α), 3.32 (t, J = 7.2 Hz, 2 H, N-CH₂- β), 3.40 (s, 3 H, OCH₃), 3.54 (sept, J =6.4 Hz, 1 H, CH iPr), 3.65 (s, 3 H, OCH₃), 3.94 (s, 3 H, OCH₃), 4.55 (s, 2 H, O-CH₂), 4.79 (s, 1 H, H- ϵ), 6.87 (ddd, J = 10.5 Hz, 8.9 Hz, 2.3 Hz, 1 H, CH_{6'-indolyl}), 7.23-7.19 (m, 2 H, CH_{4'-indolyl}, CH_{7'-indolvl}), 8.38 (br. s, 1 H, NH) ppm. ¹³C NMR: $\delta = 20.1, 20.1,$ 23.9, 43.6, 50.9, 52.5, 57.8, 65.0, 65.0, 86.2, 103.3 (d, ${}^{2}J_{C,F}$ = 24 Hz), 110.0 (d, ${}^{2}J_{C,F} = 24$ Hz), 110.1, 111.2 (d, ${}^{3}J_{C,F} = 9$ Hz), 128.7, 131.9, 136.6, 153.3, 158.1 (d, ${}^{1}J_{C,F} = 234$ Hz), 166.8, 168.1 ppm. EI MS: m/z (%) = 406 (5) [M⁺], 375 (8), 347 (12), 315 (5), 271 (5), 214 (100), 174 (15), 172 (18), 161 (25), 140 (50), 112 (20), 44 (15). C₂₁H₂₇FN₂O₅ (406.45): calcd. C 62.06, H 6.65, N 6.90; found C 62.30, H 6.51, N 6.45.

Dimethyl (2E)-2-(Ethyl{2-[5-methoxy-3-(methoxymethyl)-1H-indol-2-yl]ethyl}amino)but-2-enedioate (11): Pale yellow oil. Yield 280 mg (70%). ¹H NMR (CDCl₃): $\delta = 1.05$ (t, J = 6.7 Hz, 3 H, CH_3 -CH₂), 2.95 (q, J = 6.7 Hz, 2 H, CH_2 -CH₃), 3.06 (t, J = 7.2 Hz, 2 H, N-CH₂- CH_2), 3.38 (s, 3 H, OCH₃), 3.41 (t, J = 7.2 Hz, 2 H, N-CH₂), 3.66 (s, 3 H, OCH₃), 3.85 (s, 3 H, OCH₃), 3.95 (s, 3 H, OCH₃), 4.58 (s, 2 H, O-CH₂), 4.73 (s, 1 H, H- ϵ), 6.81 (dd, J = 8.7 Hz, 1.9 Hz, 1 H, CH_{6'-indolyl}), 7.04 (d, J = 1.9 Hz, 1 H, CH_{4'-indolyl}), 7.22 (d, J = 8.7 Hz, 1 H, CH_{7'-indolyl}), 8.21 (br. s, 1 H, NH) ppm. ¹³C NMR (CDCl₃): $\delta = 12.8$, 24.9, 46.7, 50.7, 51.2, 53.4, 56.4, 58.1, 65.5, 85.1, 100.1, 110.0, 111.8, 112.2, 129.2, 130.9, 135.5, 153.7, 154.9, 166.9, 168.6 ppm. EI MS: m/z (%) = 404 (5) [M⁺], 373 (8), 372 (20), 345 (15), 313 (25), 200 (100), 173 (85), 158 (20), 96 (15), 45 (15). C₂₁H₂₈N₂O₆ (404.46): calcd. C 62.36, H 6.98, N 6.93; found C 62.02, H 7.12, N 6.90.

Ethyl (2E)-3-[{2-[3-(Ethoxymethyl)-5-methoxy-1H-indol-2-yl]ethyl}-(ethyl)amino]acrylate (14): Pale yellow oil. Yield 230 mg (61%). ¹H NMR: $\delta = 1.05$ (t, J = 7.2 Hz, 3 H, N-CH₂-CH₃), 1.22 (t, J =7.0 Hz, 3 H, $O-CH_2-CH_3$), 1.25 (t, J = 7.1 Hz, 3 H, $O-CH_2-CH_3$, 2.95 (t, J = 7.5 Hz, 2 H, $CH_2-\alpha$), 3.05 (q, J = 7.2Hz, 2 H, $N-CH_2-CH_3$), 3.48 (t, J = 7.5 Hz, 2 H, $N-CH_2-\beta$), 3.52 (q, J = 7.0 Hz, 2 H, $O - CH_2 - CH_3$), 3.85 (s, 3 H, $O - CH_3$), 4.25 (q, J = 7.1 Hz, 2 H, $O-CH_2-CH_3$), 4.52 (s, 2 H, $O-CH_2$ - γ), 4.54 (d, J = 13.2 Hz, 1 H, H- ϵ), 6.80 (dd, J = 8.7 Hz, 1.9 Hz, 1 H, $CH_{6'-indolyl}$), 7.04 (d, J = 2.2 Hz, 1 H, $CH_{4'-indolyl}$), 7.19 (d, J = 8.7 Hz, 1 H, CH_{7'-indolyl}), 7.42 (d, J = 13.2 Hz, 1 H, H- δ), 8.22 (br. s, 1 H, NH) ppm. ¹³C NMR (CDCl₃): $\delta = 13.4, 14.6, 15.4,$ 24.7, 55.9, 55.9, 59.0, 63.2, 63.2, 65.3, 84.5, 100.6, 109.7, 111.3, 111.6, 128.9, 130.5, 135.2, 150.7, 154.3, 169.9 ppm. ESI MS: m/z = 375 [M + 1]. C₂₁H₃₀N₂O₄ (374.47): calcd. C 67.35, H 8.07, N 7.48; found C 67.39, H 8.31, N 7.25.

Ethyl (2*E*)-3-[(Ethyl){2-[5-fluoro-3-(methoxymethyl)-1*H*-indol-2-yl]ethyl}amino]acrylate (15): Yellow crystals. M.p. 83–84 °C (ethyl acetate). Yield 255 mg (73%). ¹H NMR (CDCl₃): $\delta = 1.07$ (t, J =7.2 Hz, 3 H, N–CH₂–*CH*₃), 1.26 (t, J = 7.0 Hz, 3 H, O–CH₂–*CH*₃), 2.95 (t, J = 8.3 Hz, 2 H, CH₂-α), 3.01 (q, J = 7.3Hz, 2 H, N–*CH*₂–CH₃), 3.39 (s, 3 H, OCH₃), 3.41 (t, J = 8.3 Hz, 2 H, N–CH₂–β), 4.15 (q, J = 7.3 Hz, 2 H, O–*CH*₂–CH₃), 4.55 (s, 2 H, O–CH₂-γ), 4.68 (bd, J = 13.1 Hz, 1 H, H-ε), 6.90 (td, J =8.9, 2.4 Hz, 1 H, CH_{6'-indolyl}), 7.17–7.27 (m, 2 H, CH_{4'-indolyl}, CH_{7'-indolyl}), 7.44 (d, J = 13.1 Hz, 1 H, H-δ), 8.21 (br. s, 1 H, NH) pm. ¹³C NMR (CDCl₃): $\delta = 15.1$, 19.4, 30.8, 52.1, 58.2, 59.5, 60.1, 65.5, 85.2, 97.8, 103.8 (d, $J_{C,F}^2 = 37 \text{ Hz}$), 110.5 (d, ${}^2J_{C,F} = 37 \text{ Hz}$), 111.2 (d, ${}^3J_{C,F} = 14 \text{ Hz}$), 129.3, 132.7, 136.8, 151.2, 158.6 (d, ${}^1J_{C,F} = 240 \text{ Hz}$), 170.4 ppm. EI MS: m/z (%) = 348 (10) [M⁺], 261 (5), 161 (7), 156 (100), 142 (5), 128 (20), 82 (10). C₁₉H₂₅FN₂O₃ (348.41): calcd. C 65.50, H 7.23, N 8.04; found C 65.34, H 7.33, N 8.42.

Use of the same procedure in the case of the carboline 1 resulted in the isolation of the azocinoindoles 20 and 21 (LCMS analysis of the crude reaction mixtures carried out within 2 h of the start of the reaction showed small peaks corresponding to 12 and 13).

Dimethyl 3-Ethyl-2,3,6,11-tetrahydro-1*H***-azocino[4,5-b**]indole-**4,5dicarboxylate** (**20**): Yield 170 mg (50%), m.p. 116–118 °C (ethyl acetate/hexane). ¹H NMR (CDCl₃): $\delta = 1.00$ (t, J = 7.1 Hz, 3 H, CH₂–*CH*₃), 3.00 (q, J = 7.1 Hz, 2 H, *CH*₂–CH₃), 3.17 (t, J = 6.0 Hz, 2 H, CH₂-1), 3.73 (s, 3 H, OCH₃), 3.74 (s, 3 H, OCH₃), 3.94 (t, J = 6.0 Hz, 2 H, CH₂-2), 4.04 (s, 2 H, CH₂-6), 7.07–7.01 (m, 2 H, 8-H, 9-H), 7.25 (m, 1 H, 7-H), 7.41 (m, 1 H, 10-H), 7.95 (br. s, 1 H, NH) ppm. ¹³C NMR (CDCl₃): $\delta = 14.4$, 21.6, 27.9, 45.6, 48.2, 51.7, 52.3, 103.0, 110.0, 110.1, 118.1, 119.2, 121.1, 129.0, 132.5, 135.8, 152.5, 167.8, 169.5 ppm. EI MS: *m/z* (%) = 342 (15) [M⁺], 297 (10), 283 (50), 214 (15), 200 (20), 143 (100). C₁₉H₂₂N₂O₄ (342.39): calcd. C 66.65, H 6.48, N 8.18; found C 66.54, H 6.09, N 8.25.

Ethyl 3-Ethyl-2,3,6,11-tetrahydro-1*H***-azocino[4,5-***b***]indole-5-carboxylate (21): Yield 135 mg (45%), m.p. 122–124 °C (ethyl acetate/ hexane). ¹H NMR: \delta = 1.19 (t,** *J* **= 7.2 Hz, 3 H, N–CH₂–***CH***₃), 1.29 (t,** *J* **= 7.1 Hz, 3 H, O–CH₂–***CH***₃), 3.00 (q,** *J* **= 7.2 Hz, 2 H, N–***CH***₂–CH₃), 3.22 (t,** *J* **= 5.7 Hz,** *J* **= 6.2 Hz, 2 H, CH₂-1), 3.92 (bt, 2 H, CH₂-2), 4.06 (s, 2 H, CH₂-6), 4.14 (q,** *J* **= 7.1 Hz, 2 H, O–CH₂), 7.01–7.11 (m, 2 H, 8-H, 9-H), 7.23 (m, 1 H, 7-H), 7.39 (s, 1 H, 4-H), 7.41 (m, 1 H, 10-H), 7.77 (br. s, 1 H, NH) ppm. ¹³C NMR ([D₆]DMSO): \delta = 15.3, 15.6, 19.1, 30.3, 45.3, 51.6, 59.2, 96.6, 110.7, 112.0, 117.9, 118.6, 120.7, 129.1, 133.7, 136.2, 149.8, 169.5 ppm. EI MS:** *m/z* **(%) = 298 (38) [M⁺], 269 (8), 253 (15), 225 (20), 167 (18), 156 (60), 143 (100). C₁₈H₂₂N₂O₂ (298.38): calcd. C 72.46, H 7.43, N 9.39; found C 73.21, H 7.55, N 9.52.**

General Synthetic Procedure for the Synthesis of Azocinoindoles 17–19 and 22: Anhydrous AlCl₃ (3 mg, 23 µmol) was added to a stirred solution of the alkoxy indole derivative 9–11 or 15 (0.6 mmol) in dry acetonitrile (10 mL), and stirring was continued for 24 h at room temperature (TLC monitoring: alufoil, hexane/ ethyl acetate, 1:1). The reaction mixture was concentrated under reduced pressure, and the resulting dark, oily residue was dissolved in ethyl acetate (200 mL) and percolated through a short column of alumina oxide (h = 3 cm, d = 3 cm). Evaporation of the solvent provided the target azocinoindole derivatives 17–19 and 22.

Dimethyl 3-Ethyl-8-fluoro-2,3,6,11-tetrahydro-1*H*-azocino[4,5-*b*]indole-4,5-dicarboxylate (17): Yield 130 mg (60%), white crystals, m.p. 141–143 °C (ethyl acetate/hexane). ¹H NMR (CDCl₃): δ = 1.01 (t, *J* = 7.4 Hz, 3 H, CH₂–*CH*₃), 3.00 (q, *J* = 7.4 Hz, 2 H, *CH*₂–CH₃), 3.19 (t, *J* = 6.7 Hz, 2 H, CH₂-1), 3.75 (s, 6 H, 2 × OCH₃), 3.85 (t, *J* = 6.7 Hz, 2 H, CH₂-2), 3.95 (s, 2 H, CH₂-6), 6.9 (td, *J*_{9H,F} = *J*_{9H,10H} = 8.7, *J*_{9H,7H} = 2.0 Hz, 1 H, 9-H), 7.17 (dd, *J*_{10H,9H} = 8.7, *J*_{10H,F} = 4.5 Hz, 1 H, 10-H), 7.24 (dd, *J*_{7H,F} = 8.7, *J*_{7H,9H} = 2.0 Hz, 1 H, 7-H), 7.90 (br. s, 1 H, NH) ppm. EI MS: *m/z* (%) = 360 (35) [M⁺], 328 (10), 301 (62), 271 (12), 185 (20), 161 (100), 133 (11), 59 (15). C₁₉H₂₁FN₂O₄ (360.38): calcd. C 63.32, H 5.87, N 7.77; found C 63.30, H 6.01, N 7.45. **Dimethyl 8-Fluoro-3-isopropyl-2,3,6,11-tetrahydro-1***H***-azocino**[**4,5-***b*]**indole-4,5-dicarboxylate (18):** Yield 135 mg (60%), yellow crystals, m.p. 160–162 °C (ethyl acetate/hexane). ¹H NMR (CDCl₃): δ = 1.13 (d, J = 6.6 Hz, 6 H, 2 × CH₃ *i*Pr), 3.15 (t, J = 6.0 Hz, 2 H, CH₂-1), 3.40 (m, 1 H, CH *i*Pr), 3.70 (s, 3 H, OCH₃), 3.77 (s, 3 H, OCH₃), 3.90 (t, J = 6.0 Hz, 2 H, CH₂-2), 3.93 (s, 2 H, CH₂-6), 6.83 (td, $J_{9H,F} = J_{9H,10H} = 8.6, J_{9H,7H} = 2.5$ Hz, 1 H, 9-H), 7.30–7.21 (m, 2 H, 7-H, 10-H), 7.76 (br. s, 1 H, NH) ppm. IR (KBr): \tilde{v} = 3395.9, 2952.0, 1715.8, 1669.5, 1546.3 cm⁻¹. ESI MS: m/z = 375 [M⁺ + 1]. C₂₀H₂₃FN₂O₄ (374.41): calcd. C 64.16, H 6.19, N 7.48; found C 64.11, H 6.15, N 7.09.

Dimethyl 3-Ethyl-8-methoxy-2,3,6,11-tetrahydro-1*H***-azocino[4,5-***b***]indole-4,5-dicarboxylate (19):** Yield 60 mg (25%), yellow oil, ¹H NMR (CDCl₃): $\delta = 1.01$ (t, J = 7.1 Hz, 3 H, CH₂-*CH*₃), 3.01 (q, J = 7.1 Hz, 2 H, *CH*₂-CH₃), 3.16 (t, J = 6.2 Hz, 2 H, CH₂-1), 3.73 (s, 3 H, OCH₃), 3.75 (s, 3 H, OCH₃), 3.86 (s, 3 H, OCH₃), 3.93 (t, J = 6.2 Hz, 2 H, CH₂-2), 4.00 (s, 2 H, CH₂-6), 6.76 (dd, J = 8.7 Hz, 2.3 Hz, 1 H, 9-H), 7.09 (d, J = 2.3 Hz, 1 H, 7-H), 7.12 (d, J = 8.7 Hz, 1 H, 10-H), 7.75 (br. s, 1 H, NH) ppm. ESI MS: m/z = 373 [M⁺ + 1]. C₂₀H₂₄N₂O₅ (372.42): calcd. C 64.50, H 6.50, N 7.52; found C 64.87, H 6.35, N 7.53.

Ethyl 3-Ethyl-8-fluoro-2,3,6,11-tetrahydro-1*H*-azocino[4,5-*b*]indole-**5-carboxylate (22):** Yield 125 mg (65%), white crystals, m.p. 218–220 °C (ethyl acetate/hexane). ¹H NMR (CDCl₃) :δ = 1.17 (t, J = 7.2 Hz, 3 H, N–CH₂–*CH*₃), 1.30 (t, J = 7.1 Hz, 3 H, O–CH₂*CH*₃), 3.16 (q, J = 7.2 Hz, 2 H, N–*CH*₂–CH₃), 3.20 (t, J = 5.4 Hz, 2 H, CH₂-1), 3.98 (m, 4 H, CH₂-6, CH₂-2), 4.17 (q, J = 7.1 Hz, 2 H, O–CH₂), 6.9 (td, $J_{9H,F} = J_{9H,10H} = 8.6, J_{9H,7H} = 2.5$ Hz, 1 H, 9-H), 7.09 (dd, $J_{10H,9H} = 8.6, J_{10H,F} = 4.3$ Hz, 1 H, 10-H), 7.32 (dd, $J_{7H,F} = 9.7, J_{7H,9H} = 2.5$ Hz, 1 H, 7-H), 7.35 (s, 1 H, 4-H), 7.87 (br. s, 1 H, NH) ppm. ¹³C NMR (CDCl₃): δ = 14.7, 14.8, 18.9, 30.3, 44.9, 51.6, 59.6, 97.2, 103.4 (d, ² $J_{C,F} = 23$ Hz), 109.3 (d, ² $J_{C,F} = 23$ Hz), 110.4 (d, ³ $J_{C,F} = 9$ Hz), 113.4, 129.3, 132.2, 133.6, 149.2, 157.7 (d, ¹ $J_{C,F} = 240$ Hz), 170.0 ppm. ESI MS: m/z = 317 [M⁺ + 1]. C₁₈H₂₁FN₂O₂ (316.37): calcd. C 68.34, H 6.69, N 8.85; found C 68.31, H 6.42, N 8.63.

Ethyl 3-Ethyl-6-methyl-1,2,3,6-tetrahydroazocino[5,4-b]indole-5-car**boxylate (23):** A solution of β -carboline 7 (200 mg, 0.9 mmol) and ethyl propiolate (110 mg, 1.1 mmol) in absolute ethanol (10 mL) was stirred at room temp. for 5 h. The solvent was evaporated under reduced pressure, and dry diethyl ether (10 mL) was added to the oily residue. White crystals of 23 (150 mg, 52%) were filtered off and dried, m.p. 188–190 °C. ¹H NMR (CDCl₃) : $\delta = 1.16$ (t, J = 7.2 Hz, 3 H, CH_3CH_2-N , 1.28 (t, J = 7.1 Hz, 3 H, CH_3CH_2-O , 1.59 (d, J = 7.3 Hz, 3 H, CH_3-CH), 3.05 (dt, J =14.5 Hz, 2.8 Hz, 1 H, 1-H_e), 3.25 (m, 3 H, N-CH₂CH₃, 1-H_a), 3.57 $(dt, J = 14.5, 2.8 \text{ Hz}, 1 \text{ H}, 2\text{-H}_e), 4.08 (td, J = 14.5, 2.8 \text{ Hz}, 1 \text{ H},$ 2-H_a), 4.18 (q, J = 7.1 Hz, OCH₂CH₃), 4.69 (q, J = 7.3 Hz, 1 H, 6-H), 7.05-7.16 (m, 2 H, 9-H, 10-H), 7.25 (d, J = 7.6 Hz, 1 H, 11-H), 7.45 (d, J = 7.6 Hz, 1 H, 8-H), 7.61 (s, 1 H, 4-H), 7.95 (br. s, 1 H, NH) ppm. ¹³C NMR ([D₆]DMSO): $\delta = 15.4, 15.6, 26.4,$ 27.1, 31.6, 49.8, 51.8, 59.5, 98.6, 107.4, 111.1, 118.0, 118.7, 121.0, 130.0, 134.8, 139.0, 151.6, 169.7 ppm. IR (KBr): $\tilde{v} = 3295.0$, 2974.1, 1647.6, 1598.0 cm⁻¹. EI MS: m/z (%) = 312 (100) [M⁺], 297 (27), 267 (20), 255 (13), 239 (54), 223 (9), 209 (9), 194 (16), 182 (25), 170 (61), 143 (64), 122 (50). C₁₉H₂₄N₂O₂ (312.41): calcd. C 73.05, H 7.74, N 8.97; found C 73.18, H 7.87, N 8.59.

Ethyl 3-Ethyl-6-isopropyl-1,2,3,6-tetrahydroazocino[5,4-*b*]indole-5carboxylate (24): A solution of β -carboline 8 (0.20 g, 0.8 mmol) and ethyl propiolate (100 mg, 1 mmol) in absolute ethanol (10 mL) was heated at reflux for 4 h. The solvent was evaporated under

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reduced pressure, and dry diethyl ether (10 mL) was added to the oily residue, causing precipitation of 24 (120 mg, 44%); white crystals, m.p. 222–224 °C. ¹H NMR (CDCl₃): $\delta = 0.89$ (t, J = 6.2 Hz, 3 H, $CH_3 iPr$), 1.06 (t, J = 6.8 Hz, 3 H, $CH_3 iPr$), 1.16 (t, J = 7.2Hz, 3 H, CH_3CH_2 -N), 1.26 (t, J = 7.1 Hz, 3 H, CH_3CH_2 -O), 2.04 (m, 1 H, CH *i*Pr), 2.99 (dt, J = 16.1 Hz, 2.6 Hz, 1 H, 1-H_e), 3.23 (m, 3 H, N- CH_2CH_3 , 1-H_a), 3.46 (ddt, J = 14.1, 2.9, 1.8 Hz, 1 H, 2-H_e), 4.09 (td, J = 14.1, 2.9 Hz, 1 H, 2-H_a), 4.18-4.11 (m, 3 H, 6-H, CH_2 -O), 7.11-7.08 (m, 2 H, 9-H, 10-H), 7.25 (d, J =7.6 Hz, 1 H, 11-H), 7.46 (d, J = 7.5 Hz, 1 H, 8-H), 7.64 (s, 1 H, 4-H), 8.07 (br. s, 1 H, NH) ppm. ¹³C NMR (CDCl₃): $\delta = 14.7$, 15.0, 20.4, 22.6, 26.3, 39.2, 45.2, 48.7, 52.0, 59.8, 96.9, 106.9, 110.4, 117.5, 118.8, 120.9, 130.0, 134.2, 137.9, 151.0, 171.0 ppm. IR (KBr): $\tilde{v} = 3395.9$, 2952.0, 1715.8, 1669.5, 1546.3 cm⁻¹. ESI MS: $m/z = 341 [M^+ + 1]$. C₂₁H₂₈N₂O₂ (340.46): calcd. C 74.08, H 8.29, N 8.23; found C 73.93, H 8.63, N 8.01. The solvent was evaporated from the mother liquor, and the residue was dissolved in ethyl acetate/hexane (1:10) and percolated through a short column with alumina oxide. Evaporation of the solvent provided 16 as white crystals (77 mg, 25%), m.p. 88-90 °C (ethyl acetate/hexane). ¹H NMR $(CDCl_3)$: $\delta = 0.82$ (d, J = 6.6 Hz, 3 H, CH_3 *i*Pr), 1.10 (d, J = 6.6Hz, 3 H, CH₃ *i*Pr), 1.15 (t, J = 7.2 Hz, 3 H, CH_3CH_2N), 1.16 (t, J = 7.1 Hz, 3 H, CH_3CH_2O), 1.27 (t, J = 7.1 Hz, 3 H, CH_3CH_2O), 2.00 (m, 1 H, CH iPr), 3.00 (m, 2 H, CH2-CH2-N), 3.15-3.17 (m, 2 H, N-CH₂CH₃), 3.30-3.42 (m, 4 H, CH₂CH₂N, OCH₂), 4.14 $(d, J = 7.1 \text{ Hz}, 1 \text{ H}, \text{CH} - \text{O}), 4.14 (q, J = 7.1 \text{ Hz}, 2 \text{ H}, \text{O}CH_2\text{CH}_3),$ 4.70 (d, J = 12.2 Hz, 1 H, CH = CH - N), 7.11 (t, J = 8.0 Hz, 1 H, 5-H), 7.15 (t, J = 8.0 Hz, 1 H, 4-H), 7.34 (d, J = 8.0 Hz, 1 H, 6-H), 7.48 (d, J = 12.2 Hz, 1 H, CH=CH-N), 7.53 (d, J = 8.0 Hz, 1 H, 7-H), 8.21 (br. s, 1 H, NH) ppm. ¹³C NMR (CDCl₃): δ = 14.6, 15.2, 15.2, 19.0, 19.2, 20.4, 34.3, 58.8, 58.8, 64.9, 64.9, 79.5, 84.2, 110.4, 111.0, 118.2, 119.4, 121.9, 127.9, 135.2, 135.5, 150.8, 169.9. ESI MS: $m/z = 387 [M^+ + 1]$. C₂₃H₃₄N₂O₃ (386.53): calcd. C 71.45, H 8.87, N 7.25; found C 71.34, H 8.89, N 7.34.

X-ray Crystallographic Study: Crystal data were collected on a Bruker AXS SMART 1000 area detector diffractometer^[20] (three-circle goniometer with 1 K CCD detector, Mo- K_{α} radiation, graphite monochromator).

Crystal Structure Analysis for 17: $C_{19}H_{21}FN_2O_4$, $M_r = 360.38 \text{ g} \cdot \text{mol}^{-1}$, monoclinic, space group $P2_1/c$, a = 8.7731(19), b = 17.636(4), c = 11.738(3) Å, $\beta = 104.458(4)$ Å, V = 1758.6(7) Å³, Z = 4, $\rho = 1.361 \text{ g} \cdot \text{cm}^3$, $\mu = 0.103 \text{ cm}^{-1}$, F(000) = 760. A total of 9800 reflections (2.13 < θ < 30.02°) were collected, of which 4145 were unique [R(int.) = 0.0337]. The structure was solved with the program SHELXS-97^[21] and refined by use of SHELXL-97 ^[22] to $R_1 = 0.0735$ and $wR(F^2) = 0.1830$ for 4145 reflections with $I > 2\sigma(I)$; max.\min. residual electron density 0.930 and $-0.293e \cdot \text{Å}^{-3}$.

Crystal Structure Analysis for 24: $C_{21}H_{28}N_2O_2$, $M_r = 340.45 \text{ g}\cdot\text{mol}^{-1}$, monoclinic, space group $P2_1/c$, a = 10.432(2), b = 18.175(4), c = 11.302(2) Å, $\beta = 113.62(3)$ Å, V = 1963.5(7) Å³, Z = 4, $\rho = 1.152 \text{ g}\cdot\text{cm}^3$, $\mu = 0.103 \text{ cm}^{-1}$, F(000) = 736. A total of 4592 reflections (2.13 < $\theta < 27.96$) were collected, of which 4323 were unique [R(int.) = 0.1185]. The structure was solved with the program SHELXS-97 and refined by use of SHELXL-97 to $R_1 = 0.0683$ and $wR(F^2) = 0.1877$ for 4097 reflections with $I > 2\sigma(I)$; max./min. residual electron density 0.324 and $-0.269e\cdot\text{Å}^{-3}$.

CCDC-218170 (for **17**) and -229769 (for **24**) contain the supplementary crystallographic data for this paper. These data can be

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obtained free of charge at www.ccdc.cam.ac.uk/conts/ retrieving.html [or from the Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; Fax: +44-1223/ 336-033; E-mail: deposit@ccdc.cam.ac.uk].

AChE Inhibition Measurements: Enzyme inhibitory activities were determined by Ellman's^[17] general method, with AChE from bovine erythrocytes (EC. 3.1.1.7, C-5021 from Sigma). The AChE activity was determined in a reaction mixture containing a solution of AChE (200 μ L, 0.415 U/mL in 0.1 M phosphate buffer, pH 8.0), a solution of 5,5'-dithiobis(2-nitrobenzoic acid) (100 μ L, 3.3 mM in 0.1 M phosphate-buffered solution pH 7.4), an aqueous solution of the substrate (5 mM, 100 μ L), a solution of the inhibitor (100 μ L) and phosphate buffer (pH 8, 0.1 M, 500 μ L). Acetylthiocholine was used as the substrate, and five to seven concentrations of the examined compounds, ranging from 1×10^{-7} to 1×10^{-4} M, were spectrophotometrically tested for AChE inhibition (measurements of increase in absorbance at 412 nm was taken over 3.0 min at 25 °C).

Acknowledgments

This work was supported by the Russian Foundation for Basic Research (grant no. 02-03-32941).

- ^[1] C. M. Bertha, M. Ellis, J. L. Flippen-Anderson, F. Porreca, R. B. Rothman, P. Davis, H. Xu, K. Becketts, K. C Rice, *J. Med. Chem.* **1996**, *39*, 2081–2086.
- [2] H. Rojas, M. Nidia, P. Diaz, C. Celina, O. Perez, *Revista Cub*ana de Farmacia 1977, 11, 249–255.
- [3] M. Alvarez, J. A. Joule, *Alkaloids* (Academic Press), 2001, 57 (Chemistry and Biology), 235–272.
- [4] F. Abe, R. Fu Chen, T. Yamauchi, N. Marubayashi, I. Ueda, *Chem. Pharm. Bull.* **1989**, *37*, 887–890.
- ^[5] T. Kam, K. Yoganathan, H. Li, N. Harada, *Tetrahedron* 1997, 53, 12661–12670.
- ^[6] T. Kam, K. Yoganathan, Cheng-Hock Chuakh, *Tetrahedron Lett.* 1995, 36, 759–762.
- [7] R. Yoneda, T. Kimura, J. Kinomota, S. Harusawa, T. Kurihara, J. Heterocycl. Chem. 1996, 33, 1909–1914.
- ^[8] J. Bonjoch, N. Casamitjana, J. Gracia, M. Ubeda Carmen, J. Bosch, *Tetrahedron Lett.* **1990**, *31*, 2449–2452.
- ^[9] S. Blechert, R. Knier, H. Schroers, T. Wirth, Synthesis 1995, 592-604.
- [10] K. Diker, M. Döe de Maindreville, J. Levy, *Tetrahedron Lett.* 1995, 36, 3511–3512.
- ^[11] A. V. Varlamov, T. N. Borisova, L. G. Voskressensky, B. Nsabimana, A. I. Chernyshev, *Heterocycl. Commun.* 2001, 7, 461–464.
- ^[12] T. N. Borisova, L. G. Voskressensky, T. A. Soklakova, B. Nsabimana, A. V. Varlamov, *Mendeleev Commun.* 2002, 12, 162–163.
- ^[13] A. V. Varlamov, T. N. Borisova, L. G. Voskressensky, L. N. Kulikova, T. A. Soklakova, A. I. Chernyshev, G. G. Alexandrov, *Tetrahedron Lett.* **2002**, *43*, 6767–6769.
- ^[14] T. N. Borisova, L. G. Voskressensky, L. N. Kulikova, T. A. Soklakova, A. V. Varlamov, *Molecular Diversity* 2003, 6, 207–212.
- ^[15] C. Brühlmann, F. Ooms, P.-A. Carrupt, B. Testa, M. Catto, F. Leonetti, C. Altomare, A. Carotti, J. Med. Chem. 2001, 44, 3195–3198.
- ^[16] E. Scarpini, P. Scheltsen, H. Feldman, *Lancet Neurol.* 2003, 2, 539–547.

- ^[17] G. L. Ellman, D. Courtney, V. Andres Jr., R. M. Featherstone, Biochem. Pharmacol. 1961, 7, 88-95.
- ^[18] C. J. Cattanach, A. Cohen, B. J. Heath-Brown, J. Chem. Soc., *C* **1968**, *10*, 1235–1242.
- ^[19] T. Hino, A. Hasegawa, J. Liu, M. Nakagawa, *Chem. Pharm. Bull.* **1990**, *38*, 59–64.
- ^[20] SMART (control) and SAINT (integration) software, version 5.0 Bruker AXS Inc., Madison, WI, **1997**. ^[21] G. M. Sheldrick, SHELXS, *Program for crystal structure solu*-
- ¹² G. M. Sheldrick, SHELXS, Program for crystal structure solution, University of Göttingen, Germany, 1997.
 ¹²² G. M. Sheldrick SHELXL, Program for crystal structure refine-
- ment, University of Göttingen, Germany, 1997.

Received February 17, 2004