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Tetrahedron

Tetrahedron 61 (2005) 3473-3481

Convenient synthesis of 2-alkylamino-6-carboxy-5,7-diarylcyclopenteno[1,2-b]pyridines via direct acylamination with imidoyl chlorides

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Received 27 December 2004; revised 1 February 2005; accepted 2 February 2005

Abstract—A robust synthetic method for 2-alkylamino-6-carboxy-5,7-diarylcyclopenteno[1,2-*b*]pyridines via acylamination at the alpha position of the functionalized pyridine system has been developed. The key step in this method was achieved by treatment of the corresponding pyridine *N*-oxides with 2.5 equiv of imidoyl chlorides in the presence of triethylamine, thus producing the desired 2-acylaminopyridines in good yields (74–96%). © 2005 Elsevier Ltd. All rights reserved.

1. Introduction

Endothelin-1 (ET-1)¹ and its closely related isopeptides (ET-2, ET-3) have been identified as potent vasoconstrictor peptides. The endothelins exert diverse biological actions through distinct cell surface G-protein coupled receptors (GPCR) termed ET_{A} and ET_{B} .² Elevated levels of endothelins have been observed in numerous disease states including hypertension, congestive heart failure and renal diseases.³ Therefore, non-peptide endothelin receptor antagonists are currently being evaluated by a number of pharmaceutical companies as potential therapeutic agents for the treatment of these disease states.

Previously, we reported that potent and selective ET_A antagonists (1 and 2) were identified in a series of 6-carboxy-5,7-diarylcyclopenteno[1,2-*b*]pyridines.^{4,5} It is of significant interest to further clarify the structure–activity relationships (SARs) of the alkylamino group (Fig. 1, R¹) at the 2-position of the pyridine ring and the substituent (Fig. 1, R²) at the 2'-position of the 7-(4-methoxyphenyl) ring. However, incorporation of the alkylamino group by the previous synthetic method is tedious and the subsequent functionalization on the 7-aryl ring was limited (Route A in

Scheme 1). Therefore, the development of a efficient and convenient alternative method for this highly functionalized 2-alkylaminocyclopentenopyridine system is necessary.



Figure 1.

A number of substituents (Fig. 1, \mathbb{R}^2) are easily incorporated in the 7-aryl ring at an early stage in the synthesis,⁴ therefore, route B is more available than route A in terms of the synthesis of 2-alkylamino-6-carboxy-5,7-diarylcyclopenteno[1,2-*b*]pyridine derivatives. However, the incorporation of the alkylamino group (Fig. 1, \mathbb{R}^1) is a crucial step in this route. A previous work introduced an alkylamino group at the 2-position of a pyridine ring by reacting a readily available pyridine *N*-oxide with an imidoyl chloride.⁶ This reaction is thought to be applicable to our highly functionalized pyridine system. Unfortunately, the reaction

Keywords: Acylamination; Imidoyl chloride; Cyclopenteno[1,2-b]pyridines; ETA antagonist.

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^{0040–4020/\$ -} see front matter @ 2005 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2005.02.003



Scheme 1. Possible synthetic route for compound 3.

of the pyridine *N*-oxide **5** with one equivalent of *N*-isopropylbenzimidoyl chloride afforded the desired compound **6a** in a very low yield (4.5%). Optimization of the detailed reaction conditions is necessary in order to apply this reaction to the current system.

Herein, we describe the results of the optimization of the direct acylamination at the 2-position of functionalized pyridine derivatives with an imidoyl chloride, as well as a flexible and convenient synthesis of 2-alkylamino-6-carboxy-5,7-diarylcyclopenteno[1,2-*b*]pyridine derivatives.

CO₂^tBu

OBn

2. Results and discussion

The optimized acylamination reaction conditions utilizing pyridine *N*-oxide **5** and *N*-isopropylbenzimidoyl chloride are shown in Table 1. Pyridine *N*-oxide **5** was readily prepared from **4** by treatment with *m*-CPBA at 0 °C under a nitrogen atmosphere. The initial conditions (entry 1) afforded the desired amide **6a** in a 4.5% yield.⁶ Under these conditions, half of the starting material *N*-oxide **5** was recovered together with by-products. The structure of the major by-product was estimated by Mass spectral analysis

CO₂^tBu

OBn

Βż

CO₂^tBu

OBn



CO₂^tBu

OBn

Ò



Additive

Solvent, temp

^a Total yield of 7, 8, 9 and 10.

^b 4 was isolated as a byproduct



Scheme 2. Identification of side products.

Among the inorganic bases tested, cesium fluoride (CsF) was found to dramatically enhance the desired reaction pathway resulting in a 69% yield of the 2-acylaminated pyridine (6a), however the starting material was still recovered (14%). In evaluating the organic bases, unfortunately, DBU was reacted with the imidoyl chloride to produce an ammonium salt rapidly; resulting in a 79% recovery of the starting material (entry 8). In contrast, Et₃N worked very well affording **6a** in an excellent yield (96%, entry 9). With regard to a solvent, chloroform is the best choice for this reaction. Using toluene or tetrahydrofuran instead of chloroform, Et₃N did not work well and a considerable amount of the starting material was recovered (entries 10 and 11). Further experiments around these conditions revealed that 2.5 equiv of imidoyl chlorides in the presence of 5 equiv of Et₃N (entry 12) afforded the product 6a in 94% yield. These conditions were determined to be the optimal conditions.



Scheme 3. Proposed reaction mechanism.

to be 7 in the reaction mixture. However, compound 7 was labile and easily changed by usual work-up and following isolation using silica-gel column chromatography to a mixture of 7, 8, 9 and 10 (Scheme 2).⁷ Lower reaction temperature with one equivalent of imidoyl chloride (entry 2) or with an excess of imidoyl chloride (entry 3) resulted in an improved yield of 6a, however, a considerable amount of the starting material and by-products still recovered.

Based on these results, we speculated that the formation of intermediate **11** may be a critical step in this reaction. Further, two reaction pathways were expected to exist for this reaction (Scheme 3). In this reaction, additive(s) are thought to facilitate the formation of the intermediate and the subsequent desired reaction steps, while formation of the by-product **7** can be controlled to trap chloride ions. Moreover, it was reported that addition of a base, such as DBU or triethylamine (Et₃N), increased the yields of 2-acylaminated pyridines in this type of reaction.^{6f} However, simple pyridine substrates were used in the optimization of this reaction. Thus, we reexamined the effect of several additional bases in order to accelerate the desired reaction pathway in this highly functionalized pyridine system.

In an effort to examine the applicability of these reaction conditions, pyridine *N*-oxide **5** was reacted with other imidoyl chlorides (Table 2). As expected, the reaction conditions were well tolerated, and the corresponding 2-benzoylaminopyridines (**6a–6d**) were obtained in good to excellent yields.

The previously described encouraging results prompted the

Table 2. Acylamination with various imidoyl chlorides



Entry	Imidoyl chloride	Product	Yield (%)
1	R=iso-Pr	6a	94
2	R = n - Pr	6b	93
3	R=Benzyl	6c	96
4	R=Phenyl	6d	74



Scheme 4. Synthetic route for **3**. Reaction conditions: (a) Methyl Methacrylate, PdCl₂(PPh₃)₂, NaHCO₃, DMF, 130 °C; (b) 10% Pd-C, HCOONH₄, EtOH, 80 °C (**13a**, 37%; **13b**, 28% 2 steps); (c) *m*-CPBA, CHCl₃, 6 °C (**14a**, 96%; **14b**, 83%); (d) *N*-isopropylbenzimidoyl chloride, Et₃N, CHCl₃, 60 °C (**15a**, 72%; **15b**, 74%); (e) TFA, r.t. (f) NaOH, aq-MeOH, 100 °C (**3a**, 74%; **3b**, 67% 2 steps).

synthesis of 3 by route B (Scheme 4). The Heck reaction of triflate 12 with methyl methacrylate $[PdCl_2(PPh_3)_2,$ NaHCO₃, DMF, 130 °C] followed by subsequent hydrogenation of the resultant olefin afforded a mixture of diastereomers (13a and 13b) in a moderate yield.⁵ The diastereomers were easily separated by silica gel column chromatography. Pyridine N-oxide 14a was treated with N-isopropylbenzimidoyl chloride (2.5 equiv) in the presence of Et₃N (5.0 equiv) at 60 °C and afforded 2-acylaminated pyridine 15a in a 72% yield. The stepwise deprotection of the protecting groups (tert-butyl ester, methyl ester, benzamide) on 15a was achieved by treatment with trifluoroacetic acid (TFA), followed by basic hydrolysis (NaOH) to afford the target compound **3a**. The transformation of 13b to 3b was successfully achieved in a manner similar to that described above.

3. Conclusions

A novel synthetic method for 2-alkylamino-6-carboxy-5,7diarylcyclopenteno[1,2-*b*]pyridines via a key acylamination of the corresponding *N*-oxides with imidoyl chlorides has been developed. This mild acylamination is thought to be applicable to the other highly functionalized pyridine derivatives.⁸

4. Experimental

4.1. General

All reagents and solvents were of commercial quality and used without further purification unless otherwise noted. Melting points were determined using a Yanaco MP micromelting point apparatus (Yanaco New Science Inc. Kyoto, Japan) and were not corrected. ¹H NMR and ¹³C NMR spectra were obtained on a Varian MERCURYvx 400 (Varian, Inc. CA, USA) or JEOL JNM-AL 400 (JEOL Ltd. Tokyo, Japan) instrument at 400 MHz. Chemical shifts were reported in parts per million as δ units relative to tetramethylsilane as an internal standard. Mass spectrometry was performed using micromass Q-Tof 2 (Waters Co. MA, USA) (ESI positive). Analytical and Preparative TLC were preformed using E-Merck Kieselgel F₂₅₄ precoated plates (Merck KGaA. Darmstadt, Germany). Silica gel column chromatography was carried out on Wako gel C-300 (Wako Pure Chemical Industries Ltd, Osaka, Japan.

4.1.1. (5RS,6SR,7SR)-7-(2-Benzyloxy-4-methoxyphenyl)-6-tert-butoxycarbonyl-5-(3,4-methylenedioxyphenyl)cyclopenteno[1,2-b]pyridine N-oxide (5). To a solution of (5RS,6SR,7SR)-7-(2-Benzyloxy-4-methoxyphenyl)-6-tertbutoxycarbonyl-5-(3,4-methylenedioxyphenyl)cyclopenteno[1,2-b]pyridine 4 (5.29 g, 9.59 mmol) in CHCl₃ (50 ml) was added *m*-CPBA (3.33 g, 19.2 mmol) at 0 °C and the mixture was stirred for 15 h at cold-room temperature (6 °C) under N₂. The reaction was quenched with 1 M Na₂S₂O₃ solution while stirring at 6 °C for 30 min and subsequently extracted using EtOAc. The organic layer was washed with saturated NaHCO₃ solution, water and brine, dried over Na₂SO₄ and then concentrated. The residue was purified by silica gel column chromatography (CHCl₃-MeOH = 100:0 to 30:1) to give 5 as a pale brown amorphous solid (2.89 g, 53%). ¹H NMR (CDCl₃) δ 7.99 (d, J=6.6 Hz, 1H), 7.25–7.19 (m, 4H), 7.15–6.96 (m, 3H), 6.62 (d, J =7.3 Hz, 1H), 6.55 (d, J = 8.1 Hz, 1H), 6.50–6.43 (m, 2H), 6.38-6.24 (m, 2H), 5.90 (q, J=1.7 Hz, 2H), 4.98-4.68 (m, 3H), 4.38 (d, J=8.8 Hz, 1H), 3.76 (s, 3H), 3.27 (t, J=9.5 Hz, 1H), 1.33 (s, 9H). HRMS calcd for C34H34NO7 (M+1) 568.2335, found 568.2331.

4.1.2. Preparation of *N*-isopropylbenzimidoyl chloride. A solution of isopropylamine (35.0 ml, 410 mmol) and Et₃N (69.0 ml, 490 mmol) in THF (200 ml) was cooled in an ice bath and benzoyl chloride was added to the solution. After the mixture was stirred at the same temperature for 3 h, the mixture was diluted with water and extracted with EtOAc. The organic layer was washed with 2 N HCl, water, saturated NaHCO₃ solution and brine, and subsequently dried over Na₂SO₄ and concentrated to give benzisopropylamide as a white solid (51.9 g, 78%). A mixture of benzisopropylamide (6.09 g, 36.8 mmol) and thionyl chloride (4.50 ml, 61.7 mmol) was stirred at 90 °C under a N₂ atmosphere. After 2.5 h, thionyl chloride was removed in vacuo and distilled under reduced pressure to give N-isopropylbenzimidoyl chloride as a colorless liquid (6.08 g, 91% yield). Bp 82-83 °C/5 mmHg. (lit.,⁹ 52-54 °C/1 mmHg). d = 1.15 (1.15 g/1.0 ml). ¹H NMR (CDCl₃) δ 7.97-7.94 (m, 2H), 7.47-7.34 (m, 3H), 4.14 (sept, J = 6.2 Hz, 1H), 1.27 (d, J = 6.2 Hz, 6H).

4.1.3. Reaction of (5RS,6SR,7SR)-7-(2-Benzyloxy-4methoxyphenyl)-6-tert-butoxycarbonyl-5-(3,4-methylenedioxyphenyl)cyclopenteno[1,2-b]pyridine N-oxide (5) with *N*-isopropylbenzimidoyl chloride (Table 1). In cases in which inorganic bases were employed as additives, the bases were used before being dried under heat. In cases in which organic bases were used as additives, bases were dried over NaOH. Imidoyl chloride (1, 2.5, 5 or 10 equiv) was added to a mixture of 5 (170 mg, 0.30 mmol) and additive (none, 5 or 10 equiv) in either ClCH₂CH₂Cl or $CHCl_3$ (1.0 ml) and stirred in oil bath at a prescribed temperature under an Ar atmosphere overnight. After cooling the reaction mixture, saturated NaHCO₃ solution was added to the mixture and extracted using EtOAc. The organic layer was washed with brine, dried over Na₂SO₄ and concentrated. The residue was purified by silica gel column chromatography (Hexane-EtOAc-MeOH as eluent) and if needed, further isolation was performed by preparative TLC (CHCl₃–MeOH as eluent). **6a**. ¹H NMR (CDCl₃) δ 7.27– 7.19 (m, 6H), 7.17–7.11 (m, 2H), 7.08–7.04 (m, 2H), 6.97 (d, J = 8.2 Hz, 1H), 6.94 (dd, J = 7.8, 1.2 Hz, 1H), 6.57-6.53(m, 2H), 6.53 (d, J=2.3 Hz, 1H), 6.49 (dd, J=8.4, 2.3 Hz, 1H), 6.39–6.35 (m, 2H), 5.89 (br s, 2H), 5.01–4.91 (m, 1H), 4.94 (d, J = 11.3 Hz, 1H), 4.90 (d, J = 11.3 Hz, 1H), 4.82 (d, J = 11.3 Hz, 1H)J = 10.0 Hz, 1 H), 4.40 (d, J = 10.0 Hz, 1 H), 3.82 (s, 3H), 3.40 (t, J = 10.0 Hz, 1H), 1.28 (s, 9H), 1.25 (d, J = 6.8 Hz, 3H), 1.15 (d, J = 6.8 Hz, 3H). The configuration of **6a** were determined by NOE experiments.7 NOEs were observed between δ 3.40 (H^a) and δ 6.36 (H^d), δ 3.40 (H^a) and δ 6.37 (H^e), δ 4.40 (H^b) and δ 4.82 (H^c), δ 4.40 (H^b) and δ 6.36 $(H^{d}), \delta 4.40 (H^{b}) \text{ and } \delta 6.37 (H^{e}), \delta 4.82 (H^{c}) \text{ and } \delta 6.97 (H^{f}).$



¹³C NMR (CDCl₃) δ 20.6, 21.5, 28.1, 48.9, 51.1, 51.2, 55.4, 61.29, 70.0, 80.7, 99.8, 100.8, 104.5, 108.1, 108.4, 121.3, 121.5, 122.6, 127.1, 127.5, 127.7, 128.2, 128.3, 129.1, 131.4, 133.0, 135.4, 135.9, 136.3, 137.2, 146.3, 147.5, 153.8, 157.3, 159.7, 164.1, 170.0, 172.4. HRMS calcd for C₄₄H₄₅N₂O₇ (M+1) 713.3227, found 713.3232. Anal. Calcd for C₄₄H₄₄N₂O₇: C, 74.14; H, 6.22; N, 3.93. Found C, 73.93; H, 6.23; N, 3.83.

Compound **7** (mixture of 2 isomers). ¹H NMR (CDCl₃) δ 8.51 and 8.43 (dd, J=4.9, 1.5 Hz, 1H), 7.06–7.48 (m, 8H), 6.82–6.44 (m, 5H), 5.98 and 5.87 (br s, 2H), 5.18–4.68 (m, 4H), 3.83 and 3.76 (s, 3H), 1.21 and 1.17 (s, 9H). HRMS calcd for C₃₄H₃₂NO₆ (M+1) 550.2230, found 550.2229. **8**. ¹H NMR (CDCl₃) δ 9.44 (s, 1H), 8.47 (dd, J=4.9, 1.6 Hz, 1H), 7.61 (dd, J=7.5, 1.6 Hz, 1H), 7.37 (d, J=9.0 Hz, 1H), 7.23 7.17 (m, 5H), 7.11 (dd, J=7.5, 4.9 Hz, 1H), 6.68 (dd, J= 1.8 Hz, 1H), 6.63 (br s, 1H), 6.56 (d, J=8.2 Hz, 1H), 5.90 (br s, 2H), 5.00 (d, J=11.2 Hz, 1H), 4.95 (d, J= 11.2 Hz, 1H), 3.85 (s, 3H), 1.21 (s, 9H). HRMS calcd for C₃₄H₃₂NO₈ (M+1) 582.2128, found 582.2134. **9**. ¹H NMR

(CDCl₃) δ 8.46 (dd, J=5.1, 1.5 Hz, 1H), 7.50 (dd, J=7.3, 1.5 Hz, 1H), 7.34 (br s, 1H), 7.24–7.19 (m, 5H), 7.11–7.05 (m, 1H), 7.07 (dd, J=7.3, 5.1 Hz, 1H), 6.94–6.82 (m, 1H), 6.67–6.63 (m, 2H), 5.88 (s, 2H), 5.05 (s, 2H), 4.39 (br s, 1H), 3.84 (s, 3H), 1.17 (s, 9H). HRMS calcd for C₃₄H₃₂NO₇ (M+1) 566.2179, found 566.2176. **10**. ¹H NMR (CDCl₃) δ 8.39 (dd, J=5.1, 1.5 Hz, 1H), 8.01 (d, J=8.8 Hz, 1H), 7.22 (d, J=7.3 Hz, 1H), 7.19–7.12 (m, 3H), 7.07 (dd, J=7.3, 5.1 Hz, 1H), 6.85–6.81 (m, 2H), 6.73 (d, J=8.1 Hz, 1H), 6.65 (dd, J=8.8, 2.2 Hz, 1H), 6.43 (dd, J=8.1, 1.5 Hz, 1H), 6.39 (d, J=1.5 Hz, 1H), 6.36 (d, J=2.2 Hz, 1H), 5.98 (br s, 2H), 4.66 (d, J=11.0 Hz, 1H), 4.57 (d, J=11.0 Hz, 1H), 4.20 (s, 1H), 3.78 (s, 3H), 1.24 (s, 9H). HRMS calcd for C₃₄H₃₂NO₇ (M+1) 566.2179, found 566.2182.

4.1.4. Determination of 8 (Scheme 5). To a solution of **8** (32.0 mg, 0.0552 mmol) in MeOH (0.50 ml) was added 10% Na₂SO₃ solution (0.50 ml). After stirring at room temperature for 0.5 h, the mixture was diluted using water and extracted using EtOAc. The organic layer was washed with brine, dried over Na₂SO₄ and then concentrated in vacuo. The residue was purified by silica gel column chromatography (Hexane–EtOAc=10:0 to 3:1) to yield **9** as a white amorphous solid (14.0 mg, 64%).



Scheme 5. Conversion of 8 to 9.

4.1.5. Preparations of imidoyl chlorides (Table 2). All imidoyl chlorides were prepared from the corresponding amines by a procedure similar to that described for the preparation of *N*-isopropylbenzimidoyl chloride. *N*-*n*-propylbenzimidoyl chloride. Colorless liquid. Bp 72–73 °C/1.1 mmHg. ¹H NMR (CDCl₃) δ 8.03–8.00 (m, 2H), 7.49–7.39 (m, 3H), 3.69 (t, *J*=7.0 Hz, 2H), 1.78 (qt, *J*=7.4, 7.0 Hz, 2H), 1.04 (t, *J*=7.4 Hz, 3H). *N*-benzylbenzimidoyl chloride. Colorless liquid. Bp 139–141 °C/1.1 mmHg. ¹H NMR (CDCl₃) δ 8.10–8.07 (m, 2H), 7.51–7.27 (m, 8H), 4.95 (s, 2H). *N*-phenylbenzimidoyl chloride. White solid. Bp 120–129 °C/1.1 mmHg. (lit. ^{6c} 100 °C/0.05 mmHg). Mp 35–37 °C (lit. ^{6c} 34–35 °C). ¹H NMR (CDCl₃) δ 8.20–8.17 (m, 2H), 7.59–7.40 (m, 5H), 7.25–7.20 (m, 1H), 7.04–7.01 (m, 2H).

4.1.6. Reaction of (5RS,6SR,7SR)-7-(2-Benzyloxy-4methoxyphenyl)-6-*tert*-butoxycarbonyl-5-(3,4-methylenedioxyphenyl)cyclopenteno[1,2-*b*]pyridine *N*-oxide (5) with various imidoyl chlorides (Table 2). Imidoyl chloride (0.75 mmol) was added to a mixture of **5** (170 mg, 0.30 mmol) and Et₃N (0.21 ml, 1.50 mmol) in CHCl₃ (1.0 ml) and stirred in oil bath (60 °C) under an Ar atmosphere overnight. After cooling the reaction mixture, saturated NaHCO₃ solution was added to the mixture and extracted using EtOAc. The organic layer was washed with brine, dried over Na₂SO₄ and then concentrated. The residue was purified by silica gel column chromatography (Hexane-EtOAc as eluent) to give 6b-6d. 6b. white amorphous solid (201 mg, 93% yield). ¹H NMR (CDCl₃) δ 7.32–7.15 (m, 8H), 7.07 (d, J = 8.2 Hz, 1H), 6.99–6.95 (m, 2H), 6.90 (dd, J=8.0, 1.2 Hz, 1H), 6.56–6.48 (m, 4H), 6.35 (d, J=1.7 Hz, 1H), 6.31 (dd, J=7.9, 1.7 Hz, 1H), 5.89 (br s,2H), 4.92 (d, J=11.4 Hz, 1H), 4.85 (d, J=11.4 Hz, 1H), 4.75 (d, J = 10.1 Hz, 1H), 4.39 (d, J = 10.1 Hz, 1H), 3.98– 3.84 (m, 1H), 3.82 (s, 3H), 3.42 (t, J = 10.1 Hz, 1H), 1.71 -1.45 (m, 2H), 1.29 (s, 9H), 0.79 (t, J=7.4 Hz, 3H). ¹³C NMR (CDCl₃) δ 11.4, 21.2, 28.1, 50.4, 51.3, 51.9, 55.4, 61.2, 70.0, 80.8, 100.0, 100.9, 104.5, 108.2, 108.6, 120.7, 121.1, 121.7, 127.3, 127.9, 128.0, 128.5, 128.6, 129.8, 132.1, 133.4, 135.4, 135.7, 136.4, 136.7, 146.5, 147.7, 155.6, 157.6, 160.2, 164.5, 170.5, 172.8. HRMS calcd for $C_{44}H_{45}N_2O_7$ (M+1) 713.3227, found 713.3226. Anal. Calcd for C44H44N2O7: C, 74.14; H, 6.22; N, 3.93. Found C, 74.05; H, 6.38; N, 3.75. **6c**. White amorphous solid (219 mg, 96% yield). ¹H NMR (CDCl₃) δ 7.37–7.05 (m, 14H), 6.76 (d, J = 8.0 Hz, 1H), 6.74–6.68 (m, 2H), 6.57– 6.52 (m, 2H), 6.48-6.41 (m, 2H), 6.25 (d, J = 1.4 Hz, 1H),6.15 (dd, J=7.9, 1.4 Hz, 1H), 5.86 (s, 2H), 5.23 (s, 2H), 4.77 (d, J = 11.0 Hz, 1H), 4.71 (d, J = 10.2 Hz, 1H), 4.67 (d, J = 10.2 Hz, 1H)J=11.0 Hz, 1H), 4.34 (d, J=10.2 Hz, 1H), 3.85 (s, 3H), 3.41 (t, J=10.2 Hz, 1H), 1.29 (s, 9H). ¹³C NMR (CDCl₃) δ 28.1, 51.3, 51.7, 52.1, 55.5, 60.9, 70.0, 80.8, 100.0, 100.9, 104.4, 108.3, 108.7, 120.5, 121.1, 121.6, 127.1, 127.6, 127.9, 128.0, 128.3, 128.7, 130.2, 132.2, 133.1 135.1, 135.6, 136.2, 136.2, 138.0, 146.4, 147.6, 155.2, 157.7, 160.3, 164.4, 171.0, 172.9. HRMS calcd for $C_{48}H_{45}N_2O_7$ (M+1) 761.3227, found 761.3214. Anal. Calcd for C₄₈H₄₄N₂O₇: C, 75.77; H, 5.83; N, 3.68. Found C, 75.75; H, 5.96; N, 3.55. 6d. White amorphous solid (167 mg, 74% yield). ¹H NMR (CDCl₃) δ 7.45–7.41 (m, 2H), 7.31–7.09 (m, 12H), 7.00 (dd, J = 8.0, 0.8 Hz, 1H), 6.90–6.85 (m, 3H), 6.52 (d, J = 8.0 Hz, 1H), 6.44 (d, J = 2.4 Hz, 1H), 6.40 (dd, J = 8.2, 2.4 Hz, 1H), 6.33 (d, J = 1.7 Hz, 1H), 6.26 (dd, J = 8.0, 1.7 Hz, 1H), 5.89(d, J=1.4 Hz, 1H), 5.89 (d, J=1.4 Hz, 1H), 4.75 (d, J=11.2 Hz, 1H), 4.64 (d, J=10.0 Hz, 1H), 4.63 (d, J=11.2 Hz, 1H), 4.38 (d, J = 10.0 Hz, 1H), 3.80 (s, 3H), 3.33 (t, J = 10.0 Hz, 1H), 1.27 (s, 9H). ¹³C NMR (CDCl₃) δ 28.1, 51.3, 51.8, 55.4, 61.3, 70.0, 80.7, 99.9, 100.9, 104.5, 108.3, 108.7, 120.1, 121.1, 121.7, 126.7, 127.7, 127.8, 127.9, 128.4, 129.0, 129.0, 130.1, 131.8, 134.2, 135.8, 136.1, 136.4, 136.5, 142.9, 146.5, 147.7, 155.9, 157.6, 160.0, 164.6, 170.9, 172.7. HRMS calcd for $C_{47}H_{43}N_2O_7$ (M+1) 747.3070, found 747.3078. Anal. Calcd for C₄₇H₄₂N₂O₇: C, 75.58; H, 5.67; N, 3.75. Found C, 75.26; H, 5.71; N, 3.61.

4.1.7. (5RS,6SR,7SR)-6-tert-Butoxycarbonyl-7-[2-(2methoxycarbonylpropyl)-4-methoxyphenyl]-5-(3,4methylenedioxyphenyl)cyclopenteno[1,2-b]pyridine (13a, 13b). A mixture of PdCl₂(PPh₃)₂ (2.37 g, 3.37 mmol), NaHCO₃ (5.66 g, 67.4 mmol), methyl methacrylate (90 ml, 841 mmol) and 12⁵ (9.99 g, 16.9 mmol) in DMF (240 ml) was heated with stirring at 130 °C for 15 h under a N₂ atmosphere. After cooling to room temperature, the reaction mixture was diluted with EtOAc (200 ml). Insoluble materials were filtered off using Celite pad and the Celite pad was washed with EtOAc. The combined filtrate was washed with saturated NaHCO₃ solution, water and brine, dried over Na₂SO₄ and then concentrated. The residue was purified by silica gel column chromatography (hexane-EtOAc = 4:1 to 3:2) to give mixture of *endo* and *exo* olefins (1:5 by ¹H NMR) as a pale brown amorphous solid (9.48 g, quant). This solid was used in the subsequent reaction without further separation. Pure samples of each exo and endo olefins were isolated by further separation with silica gel column chromatography (hexane-EtOAc = 3:1 to 2:1). The *exo* olefin. white amorphous solid. ¹H NMR (CDCl₃) δ 8.42 (d, J = 4.9 Hz, 1H), 7.27 (d, J = 7.6 Hz, 1H), 7.08 (dd, J = 7.6, 4.9 Hz, 1H), 7.04–6.63 (m, 6H), 6.26 (d, J = 1.1 Hz, 1H), 5.95 (br s, 2H), 5.46 (br s, 1H), 4.79 (d, J = 10.2 Hz, 1H), 4.51 (d, J = 10.2 Hz, 1H), 4.00–3.40 (m, 2H), 3.77 (s, 3H), 3.74 (s, 3H), 3.21 (t, J=10.2 Hz, 1H), 1.35 (s, 9H). HRMS calcd for $C_{32}H_{34}NO_7$ (M+1) 544.2335, found 544.2327. The *endo* olefin. Pale brown amorphous solid. ¹H NMR (CDCl₃) δ 8.42 (d, J=4.9 Hz, 1H), 7.49 (s, 1H), 7.28 (d, J=7.6 Hz, 1H), 7.16–6.67 (m, 6H), 7.10 (dd, J=7.6, 4.9 Hz, 1H), 5.97 (br s, 2H), 4.75 (d, J = 10.4 Hz, 1H), 4.53 (d, J = 10.4 Hz, 1H), 3.80 (s, 3H), 3.73 (s, 3H), 3.19 (t, J =10.4 Hz, 1H), 1.92 (d, J=1.3 Hz, 3H), 1.32 (s, 9H). HRMS calcd for C₃₂H₃₄NO₇ (M+1) 544.2335, found 544.2330.

A quantity of 6.5 g of 10% Pd–C and 20.2 g HCOONH₄ was added to a suspension of the olefin (9.48 g, 16.9 mmol) in EtOH (160 ml). After the mixture was heated with stirring at 80 °C for 24 h, insoluble materials were filtered off using a Celite pad and the Celite pad was washed with EtOAc and MeOH. After the combined filtrate was concentrated, the residue was diluted with EtOAc and water and the organic layer was separated. The organic layer was washed with saturated NaHCO₃ solution and brine, dried over Na₂SO₄ and then concentrated. The residue was purified by silica gel column chromatography (hexane–EtOAc=9:1 to 2:1) to give the more polar diastereomer **13a** as a colorless oil (3.44 g, 37%) and the less polar diastereomer **13b** as a white amorphous solid (2.54 g, 28%).

4.1.8. More polar diastereomer 13a. ¹H NMR (CDCl₃) δ 8.43 (d, J=5.0 Hz, 1H), 7.29 (d, J=7.6 Hz, 1H), 7.09 (dd, J=7.6, 5.0 Hz, 1H), 6.92–6.65 (m, 6H), 6.37 (s, 1H), 5.97 (br s, 2H), 4.91 (d, J=10.1 Hz, 1H), 4.53 (d, J=10.1 Hz, 1H), 3.76 (s, 3H), 3.66 (s, 3H), 3.20 (t, J=10.1 Hz, 1H), 3.76 (s, 3H), 1.34 (s, 9H), 1.22 (d, J=6.6 Hz, 3H). ¹³C NMR (CDCl₃) δ 17.1, 27.9, 36.5, 41.2, 49.5, 51.5, 51.9, 55.0, 64.4, 81.0, 100.8, 108.1, 108.1, 112.6, 115.0, 121.6, 121.7, 129.4, 132.0, 132.3, 135.5, 137.6, 139.6, 146.4, 147.7, 149.2, 157.7, 164.6, 172.1, 176.3. HRMS calcd for C₃₂H₃₆NO₇ (M+1) 546.24292, found 546.2488.

4.1.9. Less polar diastereomer 13b. ¹H NMR (CDCl₃) δ 8.41 (d, J=4.9 Hz, 1H), 7.29 (d, J=7.6 Hz, 1H), 7.08 (dd, J=7.6, 4.9 Hz, 1H), 6.94–6.86 (m, 1H), 6.80 (d, J=7.9 Hz, 1H), 6.79–6.67 (m, 4H), 5.97 (br s, 2H), 4.94 (d, J=9.7 Hz, 1H), 4.55 (d, J=9.7 Hz, 1H), 3.77 (s, 3H), 3.63 (s, 3H), 3.48–3.24 (m, 1H), 3.24 (t, J=9.7 Hz, 1H), 2.95–2.50 (m, 2H), 1.34 (s, 9H), 1.28–1.15 (m, 3H). ¹³C NMR (CDCl₃) δ 17.1, 28.1, 37.3, 41.0, 49.1, 51.7, 52.0, 55.2, 64.6, 81.1, 101.0, 108.2, 108.3, 112.7, 115.6, 121.8, 129.6, 132.2, 132.4, 135.8, 137.7, 139.6, 146.6, 147.9, 149.4, 157.7, 164.9, 172.2, 176.5. HRMS calcd for C₃₂H₃₆NO₇ (M+1) 546.2492, found 546.2485.

4.1.10. (5RS,6SR,7SR)-6-tert-Butoxycarbonyl-7-[2-(2methoxycarbonylpropyl)-4-methoxyphenyl]-5-(3,4methylenedioxyphenyl)cyclopenteno[1,2-b]pyridine *N***-oxide** (14a). To a solution of 13a (3.63 g, 6.65 mmol) in CHCl₃ (50 ml) was added *m*-CPBA (2.47 g, 14.3 mmol) at 0 °C and the mixture was stirred for 15 h at cold room temperature (6 °C) under a N₂ atmosphere. The reaction was quenched with 1 M Na₂S₂O₃ solution, stirred at 6 °C for 30 min and then extracted with EtOAc. The organic layer was washed with saturated NaHCO₃ solution, water and brine, dried over Na₂SO₄ and then concentrated. The residue was purified by silica gel column chromatography (CHCl₃-MeOH=30:1) to give 14a as a pale brown amorphous solid (3.58 g, 96%). ¹H NMR (CDCl₃) δ 8.02 (d, J=6.2 Hz, 1H), 7.14 (dd, J=7.7, 7.0 Hz, 1H), 6.92 (d, J = 7.7 Hz, 1H), 6.82–6.60 (m, 6H), 5.96 (br s, 2H), 5.05 (d, J=7.0 Hz, 1H), 4.56 (d, J=7.7 Hz, 1H), 3.73 (s, 3H), 3.64 (s, 3H), 3.40-2.98 (m, 4H), 1.42 (s, 9H), 1.31 (d, J=7.0 Hz)3H). HRMS calcd for $C_{32}H_{36}NO_8$ (M+1) 562.2441, found 562.2444.

4.1.11. A diastereomer of 14a (14b). *Compound* 14b was prepared from 13b (starting material) in a similar procedure to that described for the preparation of 14a. White amorphous solid (83% yield); ¹H NMR (CDCl₃) δ 8.01 (d, J=6.4 Hz, 1H), 7.15 (t, J=7.1 Hz, 1H), 6.94 (d, J=7.9 Hz, 1H), 6.80–6.66 (m, 6H), 5.96 (s, 2H), 5.09 (d, J=7.3 Hz, 1H), 4.61 (d, J=8.2 Hz, 1H), 3.73 (s, 3H), 3.65 (s, 3H), 3.52 (dd, J=13.9, 4.6 Hz, 1H), 3.40–3.29 (m, 1H), 3.15 (dd, J=8.2, 7.3 Hz, 1H), 2.64 (dd, J=13.9, 10.4 Hz, 1H), 1.42 (s, 9H), 1.20 (d, J=6.8 Hz, 3H). HRMS calcd for C₃₂H₃₆NO₈ (M+1) 562.2441, found 562.2441.

4.1.12. (5RS,6SR,7SR)-2-(1-Benzoyl-1-iso-propylamino)-6-tert-butoxycarbonyl-7-[2-(2-methoxycarbonylpropyl)-4-methoxyphenyl]-5-(3,4-methylenedioxyphenyl)cyclopenteno[1,2-b]pyridine (15a). To a solution of 14a (3.40 g, 6.05 mmol) and Et₃N (2.7 ml, 19.4 mmol) in CHCl₃ (30 ml) was added N-isopropylbenzoylimidoyl chloride (2.97 ml, 18.9 mmol) at room temperature. After stirring overnight at 60 °C under N₂, the mixture was cooled to room temperature and diluted with saturated NaHCO₃ solution. The mixture was extracted with ethyl acetate and the organic layer was washed with brine, dried over Na₂SO₄ and then concentrated. The residue was purified by silica gel column chromatography (hexane–EtOAc = 10:0 to 4:1) to give 15a as a pale brown amorphous solid (3.20 g, 72% yield). ¹H NMR (CDCl₃) δ 7.27-7.12 (m, 5H), 7.02-6.97 (m, 1H), 6.79–6.48 (m, 7H), 5.94 (br s, 2H), 4.98 (sept, J = 6.8 Hz, 1H), 4.89 (d, J=9.8 Hz, 1H), 4.46 (d, J=9.8 Hz, 1H), 3.81 (s, 3H), 3.67 (s, 3H), 3.14 (t, J=9.8 Hz, 1H), 3.07–2.96 (m, 2H), 2.92–2.82 (m, 1H), 1.34 (s, 9H), 1.25 (d, J=7.0 Hz, 3H), 1.13 (d, J=7.0 Hz, 6H). ¹³C NMR (CDCl₃) δ 17.0, 20.8, 21.1, 28.0, 36.6, 41.8, 48.2, 49.0, 51.3, 51.7, 55.2, 64.7, 81.3, 101.1, 108.1, 108.3, 113.0, 114.4, 121.8, 123.5, 127.6, 128.4, 129.2, 129.4, 133.0, 133.6, 135.9, 136.3, 137.5, 140.0, 146.8, 148.0, 154.3, 158.0, 164.6, 170.2, 172.4, 176.8. HRMS calcd for $C_{42}H_{47}N_2O_8$ (M+1) 707.3332, found 707.3325. Anal. Calcd for C₄₂H₄₆N₂O₈: C, 71.37; H, 6.56; N, 3.96. Found C, 71.09 H, 6.59; N, 3.83.

4.1.13. A diastereomer of 15a (15b). *Compound* 15b was prepared from 14b (starting material) in a similar procedure

to that described for the preparation of 15a. White amorphous solid (74% yield). ¹H NMR (CDCl₃) δ 7.25– 7.12 (m, 5H), 7.01 (dd, J=7.7, 1.1 Hz, 1H), 6.79–6.69 (m, 4H), 6.64 (dd, J = 7.8, 2.0 Hz, 1H), 6.58–6.51 (m, 2H), 5.96 (br s, 2H), 4.96 (septet, J = 6.8 Hz, 1H), 4.90 (d, J = 9.8 Hz, 1H), 4.49 (d, J=9.8 Hz, 1H), 3.82 (s, 3H), 3.68 (s, 3H), 3.52-3.43 (m, 1H), 3.21 (t, J=9.8 Hz, 1H), 2.99-2.89 (m, 1H), 2.63–2.53 (m, 1H), 1.34 (s, 9H), 1.18 (d, J=7.0 Hz, 3H), 1.12 (d, J = 7.0 Hz, 3H), 1.10 (d, J = 7.0 Hz, 3H). ¹³C NMR (CDCl₃) δ 16.4, 20.7, 21.2, 28.0, 37.4, 40.6, 48.2, 48.8, 51.0, 51.7, 55.3, 64.3, 81.3, 101.1, 108.2, 108.4, 112.8, 115.7, 121.8, 123.4, 127.6, 128.4, 129.3, 129.4, 132.9, 133.7, 136.1, 136.2, 137.4, 139.6, 146.8, 148.0, 154.4, 157.7, 164.7, 170.2, 172.3, 176.8. HRMS calcd for C₄₂H₄₇N₂O₈ (M+1) 707.3332, found 707.3329. Anal. Calcd for C₄₂H₄₆N₂O₈: C, 71.37; H, 6.56; N, 3.96. Found C, 71.15 H, 6.57; N, 3.83.

4.1.14. (5RS,6SR,7SR)-7-[2-(2-Carboxypropyl)-4-methoxyphenyl]-5-(3,4-methylenedioxyphenyl)-2-iso-propylaminocyclopenteno[1,2-b]pyridine-6-carboxylic acid (3a). Compound 15a (3.40 g, 4.81 mmol) were dissolved with TFA (50 ml) and stirred for 4 h at room temperature. After TFA was removed in vacuo, water was added to the residue and the compound was extracted with EtOAc. The organic layer was washed with brine, dried over Na₂SO₄ and then concentrated. The residue was purified by silica gel column chromatography (CHCl₃ to CHCl₃–MeOH=10:1) to yield the monoester as a white amorphous solid (2.94 g, 94%). ¹H NMR (CDCl₃) δ 7.25–7.09 (m, 5H), 7.01 (d, J= 7.9 Hz, 1H), 6.76-6.53 (m, 7H), 5.94 (br s, 2H), 4.95 (septet, J=6.8 Hz, 1H), 4.93 (d, J=9.6 Hz, 1H), 4.52 (d, J=9.6 Hz, 1H), 3.79 (s, 3H), 3.61 (s, 3H), 3.28 (t, J=9.6 Hz, 1H), 3.09–2.78 (m, 3H), 1.22 (d, J=6.8 Hz, 3H), 1.12 (d, J=6.8 Hz, 3H), 1.11 (d, J=6.8 Hz, 3H). HRMS calcd for C₃₈H₃₉N₂O₈ (M+1) 651.2706, found 651.2699. A quantity of 50 ml (200 mmol) of 4 M NaOH solution was added to a solution of monoester (2.94 g, 4.52 mmol) in MeOH (70 ml) and the mixture was stirred under reflux overnight. After cooling to room temperature, 4 N HCl was added drop-wise to the mixture until it reached pH 2-3. The mixture was then extracted with EtOAc and the organic layer was washed with brine, dried over Na₂SO₄ and then concentrated. The residue was purified by silica gel column chromatography (hexane-EtOAc = 1:2 to CHCl₃-MeOH = 10:1) to give 3aas a white solid (1.79 g, 74%). Mp 158–160 °C. ¹H NMR (acetone- d^{6}) δ 7.08 (d, J=8.5 Hz, 1H), 7.05 (d, J=8.3 Hz, 1H), 6.88–6.72 (m, 5H), 6.38 (d, J = 8.5 Hz, 1H), 6.00 (s, 2H), 5.20 (br s, 1H), 4.90 (d, J=9.5 Hz, 1H), 4.47 (d, J=8.9 Hz, 1H), 3.87–3.75 (m, 1H), 3.75 (s, 3H), 3.29 (dd, J =9.5, 8.9 Hz, 1H), 3.17-3.10 (m, 1H), 3.00-2.80 (m, 2H), 1.22 (d, J=6.5 Hz, 3H), 1.13 (d, J=6.3 Hz, 3H), 1.10 (d, J = 6.3 Hz, 3H). ¹³C NMR (acetone- d^6) δ 17.8, 22.8, 22.9, 38.6, 42.6, 43.4, 50.3, 52.0, 55.2, 63.7, 101.8, 106.8, 108.7, 108.9, 113.3, 115.8, 122.2, 125.7, 130.3, 133.8, 135.0, 138.4, 140.7, 147.2, 148.6, 158.6, 159.6, 162.8, 174.9, 176.7. HRMS calcd for $C_{30}H_{33}N_2O_7$ (M+1) 533.2288, found 533.2282. Anal. Calcd for $C_{30}H_{32}N_2O_7 \cdot 0.5 H_2O$: C, 66.53; H, 6.14; N, 5.17. Found C, 66.55 H, 6.28; N, 5.01.

4.1.15. A diastereomer of **3a** (**3b**). A volume of 2 ml of 6 M NaOH solution (12 mmol) was added to a solution of **15b** (85 mg, 0.120 mmol) in MeOH (4 ml) and the mixture was

stirred under reflux overnight. After cooling to room temperature, 6 N HCl was added to the mixture until it reached pH 2-3. Insoluble material was collected by filtration and the filtrate was extracted using EtOAc. The organic layer was washed with brine, dried over Na₂SO₄ and then concentrated. The residue and the insoluble material were combined and purified by silica gel column chromatography (CHCl₃–MeOH = 10:0 to 9:1) to give **3b** as a pale brown solid (43 mg, 67%). Mp 137-141 °C. ¹H NMR (CD₃OD) δ 7.33 (d, J=8.8 Hz, 1H), 6.94 (d, J=7.8 Hz, 1H), 6.83–6.75 (m, 5H), 6.68 (d, J=8.8 Hz, 1H), 5.92 (s, 2H), 5.16–5.05 (m, 1H), 4.48 (d, J=8.4 Hz, 1H), 3.90–3.80 (m, 1H), 3.76 (s, 3H), 3.28-3.20 (m, 1H), 3.14-3.06 (m, 1H), 2.85 (dd, J=14.6, 7.0 Hz, 1H), 2.73–2.62 (m, 1H), 1.30–1.23 (m, 3H), 1.21 (d, J=6.3 Hz, 3H), 1.14 (d, J=6.3 Hz, 3H). ¹³C NMR (CD₃OD) δ 18.3, 22.3, 22.6, 37.5, 43.2, 45.1, 52.8, 55.7, 65.4, 102.4, 109.1, 109.2, 110.6, 114.0, 116.0, 122.4, 127.8, 130.3, 132.6, 137.2, 139.9, 141.8, 148.2, 149.4, 156.0, 156.2, 159.9, 176.1, 180.5. HRMS calcd for $C_{30}H_{33}N_2O_7$ (M+1) 533.2288, found 533.2295. Anal. Calcd for C₃₀H₃₂N₂O₇·0.5 H₂CO₃: C, 65.00; H, 5.90; N, 4.97. Found C, 64.99 H, 6.04; N, 4.94.

Acknowledgements

We thank Dr. Shigeru Nakajima and Mr. Hirokazu Ohsawa for analytical support (NMR and Mass spectra). We also thank Toray Research Center, Inc. for elemental analysis.

References and notes

- Yanagisawa, M.; Kurihara, H.; Kimura, S.; Tomobe, Y.; Kobayashi, M.; Mitsui, Y.; Yazaki, Y.; Goto, K.; Masaki, T. *Nature* 1988, *332*, 411–415.
- Hosoda, K.; Nakao, K.; Arai, H.; Nagakawa, O.; Hosoda, K.; Suga, S.; Nakanishi, S.; Imura, H. *FEBS Lett.* **1991**, 287, 23–26.
- 3. For ET_A selective antagonist. (a) Winn, M.; Geldern, T. W.; Opgenorth, T. J.; Jae, H.; Tasker, A. S.; Boyd, S. A.; Kester, J. A.; Mantei, R. A.; Bal, R.; Sorensen, B. K.; Wu-Wong, J. R.; Chiou, W. J.; Dixon, D. B.; Novosad, E. I.; Hernadez, L.; Marsh, K. C. J. Med. Chem. 1996, 38, 1039-1048. (b) Wu, C.; Chan, M. F.; Stavros, F.; Okun, I.; Mong, S.; Keller, K. M.; Brock, T.; Kogan, T. P.; Dixon, R. A. F. J. Med. Chem. 1997, 40, 1690–1697. (c) Liu, G.; Henry, K. J., Jr.; Szczepankiewicz, B. G.; Winn, M.; Kozmina, N. S.; Boyd, S. A.; Wasicak, J.; von Geldern, T. W.; Wu-Wong, J. R.; Chiou, W. J.; Dixon, D. B.; Nguyen, B.; Marsh, K. C.; Opgenorth, T. J. J. Med. Chem. 1998, 41, 3261-3275. (d) Astles, P. C.; Brown, T. J.; Halley, F.; Handscombe, C. M.; Harris, N. V.; Majid, T. N.; McCarthy, C.; McLay, A.; Porter, B.; Roach, A. G.; Sargent, C.; Smith, C.; Walsh, R. J. A. J. Med. Chem. 2000, 43, 900-910. (e) Wu, C.; Decker, E. R.; Blok, N.; Li, J.; Bourgoyne, A. R.; Bui, H.; Keller, K. M.; Knowles, V.; Li, W.; Stavros, F. D.; Holland, G. W.; Brock, T. A.; Dixon, R. A. F. J. Med. Chem. 2001, 44, 1211-1216. (f) Morimoto, H.; Ohashi, N.; Shimadzu, H.; Kushiyama, E.; Kawanishi, H.; Hosaka, T.; Kawase, Y.; Yasuda, K.; Kikkawa, K.; Yamauchi-Kohno, R.; Yamada, K. J. Med. Chem. 2001, 44, 3369-3377. (g) Ishizuka, N.; Matsumura, K.; Sakai, K.; Fujimoto, M.; Mihara, S.;

Yamamori, T. J. Med. Chem. 2002, 45, 2041-2055. (h) Murugesan, N.; Gu, Z.; Spergel, S.; Young, M.; Chen, P.; Mathur, A.; Leith, L.; Hermsmeier, M.; Liu, E. C.-K.; Zhang, R.; Bird, E.; Waldron, T.; Marino, A.; Koplowitz, B.; Humphreys, W. G.; Chong, S.; Morrison, R. A.; Webb, M. L.; Moreland, S.; Trippodo, N.; Barrish, J. C. J. Med. Chem. 2003, 46, 125-137. For ET_B selective antagonist. (a) Balwierczak, J. L.; Bruseo, C. W.; Del Grande, D.; Jeng, A. Y.; Savage, P.; Shetty, S. S. J. Cardiovasc. Pharmacol. 1995, 26(Suppl. 3), S393-S396. (b) Chan, M. F.; Kois, A.; Verner, E. J.; Raju, B. G.; Castillo, R. S.; Wu, C.; Okun, I.; Stavros, F. D.; Balaji, V. N. Bioorg. Med. Chem. 1998, 6, 2301-2316. (c) Liu, G.; Kozmina, N. S.; Winn, M.; von Geldern, T. W.; Chiou, W. J.; Dixon, D. B.; Nguyen, B.; Marsh, K. C.; Opgenorth, T. J. J. Med. Chem. 1999, 42, 3679-3689. (d) Mederski, W. W. K. R.; Osswald, M.; Dorsch, D.; Christadler, M.; Schmitges, C.-J.; Wilm, C. Bioorg. Med. Chem. Lett. 1999, 9, 619–622. For ET_A/ET_B balanced antagonist. (a) Clozel, M.; Breu, V.; Gray, G. A.; Kalina, B.; Loffler, B.; Burri, K.; Cassal, J.; Hirth, G.; Muller, M.; Neidhart, W.; Ramuz, H. J. Pharmacol. Exp. Ther. 1994, 270, 228-235. (b) Elliot, J. D.; Lago, M. A.; Cousins, R. D.; Gao, A.; Leber, J. D.; Erhard, K. F.; Nambi, P.; Elshourbagy, N. A.; Kumar, C.; Lee, J. A.; Bean, J. W.; Debrosse, C. W.; Eggleston, D. S.; Brooks, D. P.; Feuerstein, G.; Ruffolo, R. R.; Weinstock, J.; Gleason, J. G.; Peishoff, C. E.; Ohlstein, E. J. Med. Chem. 1994, 37, 1553-1557. (c) Walsh, T. F.; Fitch, K. J.; Chakravarty, P. K.; Williams, D. L.; Murphy, K. A.; Nolan, N. A.; O'Brien, J. A.; Lis, E. V.; Pettibone, D. J.; Kivlighn, S. D.; Gabel, R. A.; Zingaro, G. J.; Krause, S. M.; Siegl, P. K. S.; Clineschmidt, B. V.; Greenlee, W. J. Abstracts of American Chemical Society National Meeting, Washington, DC, August 21-25, 1994, American Chemical Society: Washington, DC, 1994; MEDI 145. (d) Jae, H.-S.; Winn, M.; Dixon, D. B.; Marsh, K. C.; Nguyen, B.; Opgenorth, T. J.; von Geldern, T. W. J. Med. Chem. 1997, 40, 3217-3227. (e) Amberg, W.; Hergenröder, S.; Hillen, H.; Jansen, R.; Kettschau, G.; Kling, A.; Klinge, D.; Raschack, M.; Riechers, H.; Unger, L. J. Med. Chem. 1999, 42, 3026-3032.

- Niiyama, K.; Takahashi, H.; Nagase, T.; Kojima, H.; Amano, Y.; Katsuki, K.; Yamakawa, T.; Ozaki, S.; Ihara, M.; Yano, M.; Fukuroda, T.; Nishikibe, M.; Ishikawa, K. *Bioorg. Med. Chem. Lett.* 2002, *12*, 3041–3045.
- Niiyama, K.; Mase, T.; Takahashi, H.; Naya, A.; Katsuki, K.; Nagase, T.; Ito, S.; Hayama, T.; Hisaka, A.; Ozaki, S.; Ihara, M.; Yano, M.; Fukuroda, T.; Noguchi, K.; Nishikibe, M.; Ishikawa, K. *Bioorg. Med. Chem.* **2002**, *10*, 2461–2470.
- 6. (a) Abramovitch, R. A.; Singer, G. M. J. Am. Chem. Soc. 1969, 91, 5672–5673. (b) Abramovitch, R. A.; Rogers, R. B. Tetrahedron Lett. 1971, 22, 1951–1954. (c) Abramovitch, R. A.; Singer, G. M. J. Org. Chem. 1974, 39, 1795–1802. (d) Abramovitch, R. A.; Rogers, R. B. J. Org. Chem. 1974, 39, 1802–1807. (e) Abramovitch, R. A.; Rogers, R. B.; Singer, G. M. J. Org. Chem. 1975, 40, 41–47. (f) Abramovitch, R. A.; Bailey, T. D. J. Heterocycl. Chem. 1975, 12, 1079–1080. (g) Abramovitch, R. A.; Abramovitch, D. A.; Tomasik, P. J. Chem. Soc., Chem. Commun. 1979, 21, 956–957. (h) Abramovitch, R. A.; Pilski, J.; Konitz, A.; Tomasik, P. J. Org. Chem. 1983, 48, 4391–4393. Recently, one pot method for the generation of imidoyl chloride and in situ reaction with pyridine N-oxide was described. Manley, P. J.; Bilodeau, M. T. Org. Lett. 2002, 4, 3127–3129.
- 7. Compound 7 and 9 was synthesized or determined in previous

literature. Niiyama, K.; Yoshizumi, T.; Takahashi, H.; Naya, A.; Ohtake, N.; Fukami, T.; Mase, T.; Hayama, T.; Ishikawa, K. *Bioorg. Med. Chem.* **2002**, *10*, 3437–3444. Compound **10** was synthesized in the unpublished SAR study of 7-aryl part. The structure of **8** was elucidated by conversion of **8** to **9** (10% aqueous Na₂SO₃ solution, Scheme 5).

 (a) Takahashi, H.; Ohtake, N.; Sakamoto, T.; Iino, T.; Kawanishi, N.; Nakamura, M.; Yoshizumi, T.; Niiyama, K.; Ozaki, S.; Okada, H.; Kano, A.; Takahashi, H.; Ishii, Y.; Okada, M.; Saito, M.; Sawazaki, Y.; Hayama, T.; Nishikibe, M. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 1503–1507. (b) Yoshizumi, T.; Takahashi, H.; Ohtake, N.; Jona, H.; Sato, Y.; Kishino, H.; Sakamoto, T.; Ozaki, S.; Takahashi, H.; Shibata, Y.; Ishii, Y.; Saito, M.; Okada, M.; Okada, M.; Hayama, T.; Nishikibe, M. *Bioorg. Med. Chem.* **2004**, *12*, 2139–3150.

 Brindly, J. C.; Caldwell, J. M.; Meakins, G. D.; Plackett, S. J.; Price J. Chem. Soc., Perkin Trans. 1 1987, 1153–1158.