Synthesis of Insoluble Polystyrene-Supported Flavins and Their Catalysis in Aerobic Reduction of Olefins

Yukihiro Arakawa,¹ Risa Kawachi,¹ Yoshihiko Tezuka,² Keiji Minagawa,^{1,3} Yasushi Imada ^[]

¹Department of Applied Chemistry, Tokushima University, Minamijosanjima, Tokushima 770-8506, Japan ²Department of Optical Science and Technology, Tokushima University, Minamijosanjima, Tokushima 770-8506, Japan ³Institute of Liberal Arts and Sciences, Tokushima University, Minamijosanjima, Tokushima 770-8502, Japan Correspondence to: Y. Imada (E-mail: imada@tokushima-u.ac.jp)

Received 5 January 2017; accepted 27 January 2017; published online 00 Month 2017 DOI: 10.1002/pola.28536

ABSTRACT: 2',4'-p-Vinylbenzylideneriboflavin (2',4'-PVBRFI) was prepared as a flavin-containing monomer and copolymerized with divinylbenzene and styrene or its *p*-substituted derivatives such as 4-acetoxystyrene, 4-vinylbenzyl alcohol, and 4vinylbenzoic acid to give the corresponding non-functionalized and functionalized PS-DVB-supported flavins PS(H)-DVB-FI, PS(OAc)-DVB-FI, PS(CH₂OH)-DVB-FI, and PS(COOH)-DVB-FI, respectively. PS(OH)-DVB-FI was also prepared by hydrolysis of PS(OAc)-DVB-FI under basic conditions. These novel flavincontaining insoluble polymers exhibited characteristic fluorescence in solid state, except PS(OH)-DVB-FI, and different catalytic activities in aerobic reduction of olefins by *in situ* generated diimide from hydrazine depending on their pendant

INTRODUCTION Flavoenzymes such as monooxygenase and DNA photolyase containing flavin cofactors play a pivotal role in biological redox systems in nature.¹ Catalytic functions of these flavoenzymes have inspired chemists to use the active center, the isoalloxazine ring of flavin cofactors, for developing simple flavin molecule-catalyzed metal-free redox reactions for organic synthesis.² Although various biomimetic flavin-catalyzed organic reactions have been reported, they have not yet reached the stage of practical application. One of the unsolved issues responsible for such limitation is efficient recovery and reuse of the flavin catalysts. In general, immobilizing catalysts on an insoluble support can be highly effective since it allows for not only easy recovery and reusability of the catalysts as well as facile product isolation but also the development of continuous flow synthesis.³ Nevertheless, it is surprising that the immobilization of flavin catalysts onto an insoluble support via covalent bond has remained unexplored so far, although there have been several reports on the synthesis and application of linear soluble polymer-supported flavins.⁴⁻⁷ König and coworkers reported that flavin molecules bearing

functional group. For example, **PS(H)-DVB-FI** was found to be particularly effective for neutral hydrophobic substrates, which could be readily recovered by a simple filtration and reused more than 10 times without loss in catalytic activity. On the other hand, **PS(OH)-DVB-FI** and **PS(COOH)-DVB-FI** proved to be highly active for phenolic substrates known to be less reactive in the reaction with conventional non-supported flavin catalysts. © 2017 Wiley Periodicals, Inc. J. Polym. Sci., Part A: Polym. Chem. **2017**, *00*, 000–000

KEYWORDS: aerobic oxidation; catalysis; flavin; fluorescence; heterogeneous polymers

fluorinated or hydrophobic aliphatic chains could be immobilized onto solid supports such as fluorous silica gel, reversed phase silica gel and polyethylene pellet and used them as reusable heterogeneous photocatalysts for oxidation of benzyl alcohols in water.⁸ However, since they are noncovalent immobilizations, it is probably not so easy to apply them to other reactions requiring organic solvents or additional reagents without catalyst leaching. Thus, a readily reusable insoluble polymer-supported flavin catalyst potentially available for comprehensive reactions has so far not been developed.

Reduction of olefins is among the most fundamental molecular transformations in organic synthesis. The most commonly used method for olefin reduction today is arguably that with molecular hydrogen catalyzed by a transition metal such as Pd and Pt because of its high efficiency and reliability, even though noble metal catalysts and risky H_2 gas are required.⁹ Meanwhile, olefin reduction by diimide, NH = NH, is known as an alternative method where in principle NH = NH has to be generated *in situ* due to its high lability.¹⁰ In this respect, there is no doubt that hydrazine, NH_2NH_2 , is the most

Additional Supporting Information may be found in the online version of this article.

© 2017 Wiley Periodicals, Inc.



1

attractive NH = NH generator, because it is much cheaper than other candidates, such as anthracene diimide and arylsulfonylhydrazide,¹⁰ and H₂O is the sole byproduct of oxidation of NH_2NH_2 by O_2 . We previously introduced an efficient way for the generation of NH = NH by the aerobic oxidation of NH₂NH₂ with synthetic flavin catalysts, which led us to the development of a convenient and safe method for the flavin-catalyzed aerobic reduction of olefins.¹¹ Combination of the inherent chemoselectivity of diimide reduction¹⁰ and the redox catalytic activity of flavin molecules has allowed for efficient reduction of various olefins including those not applicable in typical metal-based hydrogenation under mild conditions, as demonstrated by us¹¹ and others,¹² which is highly environmentally benign because of utilizing metal-free organocatalyst (flavin) and nonhazardous terminal oxidant (O_2) and producing nontoxic wastes (N₂ and H₂O). The synthetic utility of the flavin-catalyzed olefin reduction further led us to develop readily reusable insoluble polymersupported flavin catalysts to render it even more practical.

In this article, we present the first insoluble organic polymersupported flavins and their use as an efficient and reusable catalyst for aerobic reduction of olefins with hydrazine. We have chosen poly(styrene-*co*-divinylbenzene) (PS-DVB) as a support, because the corresponding polymeric catalysts can be designed from comonomers and easily prepared by radical copolymerization.³ We initially show the synthesis of new flavin-containing styrene monomer, 2',4'-O-*p*-vinylbenzilidene riboflavin (2',4'-**PVBRFI**), and then its radical copolymerization with divinylbenzene and styrene or *p*-functionalized styrene derivatives to prepare poly(styrene-*co*-divinylbenzene*co*-2',4'-**PVBRFI**) (**PS(R)-DVB-FI**) bearing a different pendant group (R). Finally, we detail the catalytic activities as well as reusability of **PS(R)-DVB-FI** in aerobic reduction of olefins with hydrazine.

EXPERIMENTAL

General

Melting points were measured on an AS ONE ATM-01. NMR spectra were recorded using a JEOL JNM-ECX-400 (¹H, 400 MHz) and JNM-ECA-400 spectrometers (¹H, 400 MHz) or a Varian Unity-Inova 600 spectrometer (¹H, 600 MHz and ¹³C, 151 MHz). The chemical shifts of ¹H NMR and ¹³C NMR signals are quoted relative to tetramethylsilane. IR spectra were recorded on a JASCO IR-460 spectrometer with ATR unit. UV spectra were recorded on JASCO V-550 spectrometer. Steadystate emission and excitation spectra were recorded using a Hitachi F-7000 spectrometer. The solid-state emission quantum yield was measured using a BaSO₄-coated integrating sphere (69 mm internal diameter) with an exit port (20 mm diameter) connected to a polychromator (Photal MCPD-1000), image intensifier (Hamamatsu Photonics, C7039-02), and dual-cooled CCD camera (Hamamatsu Photonics, C4880-17). Excitation ray for solid-state emission was a diode laser (Coherent, Radius 405) or a high stability mercury xenon lamp (Hamamatsu Photonics, L2422) coupled with a monochromator (Photon Technology International, Model 101). All emission spectra were corrected for the spectral response of the optical system using quinine bisulfate and *N*,*N*-dimethylaminonitrobenzene as a standard.¹³ GC analyses were carried out on a Shimazu GC-2010 by using a DB-1 glass capillary column (0.25 mm \times 30 m).

Materials

Riboflavin (Kanto Chemical), hydroquinone (nacalai tesque), *p*toluenesulfonic acid monohydrate (TsOH•H₂O, TCI), dimethyl 2,2'-azobis(2-methylpropionate) (MAIB, Wako Chemical), hydrazine monohydrate (Wako Chemical, nacalai tesque), 4phenyl-1-butene (Kanto Chemical), *o*-allylphenol (Kanto Chemical), DMF (nacalai tesque), and acetonitrile (nacalai tesque) were commercially available and used without further purification. 2,2'-Azobis(isobutyronitrile) (AIBN, Wako Chemical) and 4-vinylbenzoic acid (TCI) were purified by recrystallization from methanol. 4-Acetoxystyrene (Sigma Aldrich, Japan) was purified by washing with 0.1 N sodium hydride and water. Styrene (nacalai tesque) and divinylbenzene (DVB, Wako Chemical) were purified by distillation under reduced pressure. 4-Vinylbenzaldehyde diethyl acetal¹⁴ and 4-vinylbenzyl alcohol¹⁵ were prepared according to the reported procedure.

Synthesis of 2',4'-PVBRFl

Riboflavin (225 mg, 0.60 mmol), p-TsOH•H₂O (114 mg, 0.60 mmol), and hydroquinone (0.4 mg, 3.6 μ mol) were placed in a heat-gun-dried 50 mL round bottom flask under a nitrogen atmosphere. DMF (12 mL) and 4-vinylbenzaldehyde diethyl acetal (309 mg, 1.5 mmol) were added, and the solution was stirred for 3 h at 80 °C. The reaction mixture was diluted with ethyl acetate (50 mL) and washed with water (3 \times 30 mL). The aqueous layer was extracted with ethyl acetate (3 \times 30 mL), and the combined organic layers were dried over MgSO₄, which was filtered and concentrated under reduced pressure. The resulting crude product was purified by flash column chromatography on silica gel using a mixture of MeOH:EtOAc 6:94 as eluent to afford 163 mg of 2',4'-PVBRFl as yellow powder (56%): M.p. 180 °C (decomp.); ¹H NMR (600 MHz, DMSO-*d*₆, δ): 2.38 (s, 3H, 7-CH₃), 2.45 (s, 3H, 8-CH₃), 3.45-3.53 (m, 1H, 3'-CH), 3.55 (dt, *J* = 12, 5.9 Hz, 1H, 5'-CH), 3.64 (ddd, *J* = 9.6, 6.0, 1.6 Hz, 1H, 4'-CH), 3.75 (ddd, *J* = 11, 6.0, 1.5 Hz, 1H, 5'-CH), 4.16 (ddd, J = 9.6, 7.5, 2.6 Hz, 1H, 2'-CH), 4.68 (t, J = 5.8 Hz, 1H, 5'-OH), 4.76-4.94 (br, 1H, 1'-CH), 5.11-5.23 (br, 1H, 1'-CH), 5.23 (dd, J = 10.8, 0.6 Hz, 1H, -C = CH), 5.50 (s, 1H, -OCHO-), 5.64 (d, *J* = 5.4 Hz, 1H, 3'-OH), 5.81 (dd, *J* = 18, 0.6 Hz, 1H, -C = CH), 6.68 (dd, J = 10.8, 17.4 Hz, 1H, -CH = C), 7.21 (d, J = 8.4 Hz, 2H, m-H), 7.35 (d, J = 7.8 Hz, 2H, o-H), 7.81 (s, 1H, 6-ArH), 8.01 (s, 1H, 9-ArH), 11.35 ppm (s, 1H, NH); ¹³C NMR (151 MHz, DMSO-d₆, δ): 18.7, 20.6, 46.2, 60.7, 63.3, 78.5, 81.8, 99.0, 114.7, 118.1, 125.4, 126.3, 130.4, 132.0, 133.7, 135.7, 136.1, 137.1, 137.2, 137.4, 145.6, 150.6, 155.3, 159.8 ppm; IR (ATR): v = 3491, 3322, 3162, 3012, 2829, 1716, 1650, 1574, 1532, 1497, 1421, 1402, 1349, 1259, 1226, 1180, 1132, 1082, 1011, 833 cm⁻¹; UV-vis (ethanol): λ_{max} $(\varepsilon) = 264$ (34344), 360 (6351), 444 nm (8898); Fluorescence emission: λ_{max} (solid) = 556 nm (λ_{ex} = 442 nm), λ_{max} (ethanol) = 516 nm (λ_{ex} = 442 nm); MS (DART): m/z 491.1

([M + H] ⁺); Anal. calcd. for $C_{26}H_{26}N_4O_6$: C 61.41, H 5.55, N 11.02; found: C 61.58, H 5.42, N 11.09.

Preparation of PS(H)-DVB-Fl

2',4'-PVBRFI (39 mg, 0.08 mmol), styrene (367 mg, 3.52 mmol), DVB (52 mg, 0.40 mmol), and MAIB (18 mg, 0.08 mmol) was dissolved in DMF (2 mL) in a screw tube (18 mmID). The mixture was bubbled with nitrogen for 1 min and stirred at 65 °C for 25 h under nitrogen atmosphere. The resulting yellow gel was poured into ether and crushed, then washed with ether, THF, and methanol by means of Soxhlet extractor and dried in *vacuo* to give 214 mg of poly (2',4'-PVBRFI-*co*-St-*co*-DVB) (**PS(H)-DVB-FI**) as yellow solid (58%). The content of flavin unit in polymer was calculated to be 0.12 mmol g⁻¹ based on the elemental analysis data: Anal. Found: C 88.82, H 7.81, N 0.69; M.p. 180 °C (decomp.); Fluorescence emission (solid): $\lambda_{max} = 539$ nm ($\lambda_{ex} = 405$ nm); IR (ATR): v = 3060, 3026, 2921, 2852, 1601, 1584, 1547, 1493, 1451, 756, 696, 541 cm⁻¹. Particle size of the polymer was controlled by grinding and sieving prior to use as catalyst.

Preparation of PS(OAc)-DVB-Fl

2',4'-PVBRFI (78 mg, 0.16 mmol), 4-acetoxystyrene (1.14 g, 7.04 mmol), DVB (104 mg, 0.80 mmol) and AIBN (26 mg, 0.16 mmol) was dissolved in DMF (4 mL) in a screw tube (18 mmID). The mixture was bubbled with nitrogen for 1 min and stirred at 65 °C for 18 h under nitrogen atmosphere. The resulting yellow gel was poured into ether and crushed, then washed with ether, THF, and methanol by means of Soxhlet extractor and dried in *vacuo* to give 869 mg of poly(2',4'-PVBRFI-*co*-4-acetoxystyrene-*co*-DVB) (**PS(OAc)-DVB-FI**) as yellow solid (65%). The content of flavin unit in polymer was calculated to be 0.18 mmol g⁻¹ based on the elemental analysis data: Anal. Found: C 74.73, H 6.46, N 0.98; Fluorescence emission (solid): $\lambda_{max} = 537$ nm ($\lambda_{ex} = 405$ nm); IR (ATR): v = 2923, 2849, 1754, 1505, 1367, 1130, 1186, 1164, 1014, 909, 844, 552 cm⁻¹.

Preparation of PS(OH)-DVB-Fl

A mixture of **PS(OAc)-DVB-FI** (900 mg) and NH₂NH₂•H₂O (1.02 g, 20.3 mmol) in DMF (20 mL) was stirred at 30 °C for 3 h in a screw tube (18 mmID). The resulting orange gel was poured into water and crushed, then washed with water, methanol, and ether by means of Soxhlet extractor and dried in *vacuo* to give 321 mg of poly(2',4'-PVBRFI-*co*-4-hydroxystyrene-*co*-DVB) (**PS(OH)-DVB-FI**) as orange solid. The content of flavin unit in polymer was estimated to be 0.18 mmol g⁻¹ because of quantitative conversion judged by FTIR analysis: IR (ATR): v = 3329, 3024, 2915, 1611, 1540, 1509, 1444, 1363, 1225, 1171, 1013, 905, 825 cm⁻¹. Particle size of the polymer was controlled by grinding and sieving prior to use as catalyst.

Preparation of PS(CH₂OH)-DVB-Fl

2',4'-PVBRFI (39 mg, 0.08 mmol), 4-vinylbenzyl alcohol (477 mg, 3.56 mmol), DVB (52 mg, 0.40 mmol) and MAIB (18 mg, 0.08 mmol) was dissolved in DMF (2 mL) in a screw tube (18 mmID). The mixture was bubbled with nitrogen for 1 min and stirred at 65 °C for 36 h under nitrogen atmosphere. The resulting yellow gel was poured into ether and crushed, then washed with ether, THF, and methanol by means of Soxhlet

extractor and dried *in vacuo* to give 207 mg of poly(2',4'-PVBRFl*co*-4-vinylbenzyl alcohol-*co*-DVB) (**PS(CH₂OH)-DVB-Fl**) as yellow solid (48%). The content of flavin unit in polymer was calculated to be 0.15 mmol g⁻¹ based on the elemental analysis data: Anal. Found: C 77.36, H 7.40, N 0.85; Fluorescence emission (solid): $\lambda_{max} = 553$ nm ($\lambda_{ex} = 405$ nm); IR (ATR): v = 3322, 2921, 1540, 1511, 1419, 1211, 1011, 806 cm⁻¹. Particle size of the polymer was controlled by grinding and sieving prior to use as catalyst.

Preparation of PS(COOH)-DVB-Fl

2',4'-PVBRFI (39 mg, 0.08 mmol), 4-vinylbenzoic acid (527 mg, 3.56 mmol), DVB (52 mg, 0.40 mmol) and MAIB (18 mg, 0.08 mmol) was dissolved in DMF (2 mL) in a screw tube (18 mmID). The mixture was bubbled with nitrogen for 1 min and stirred at 65 °C for 10 h under nitrogen atmosphere. The resulting yellow gel was poured into ether and crushed, then washed with ether, THF, and methanol by means of Soxhlet extractor and dried in vacuo to give 466 mg of poly(2',4'-PVBRFl-co-4vinylbenzoic acid-co-DVB) (PS(COOH)-DVB-FI) as yellow solid (76%). The content of flavin unit in polymer was calculated to be 0.13 mmol g^{-1} based on the elemental analysis data: Anal. Found: C 68.92, H 6.03, N 0.74; Fluorescence emission (solid): $\lambda_{max} = 555 \text{ nm}$ ($\lambda_{ex} = 405 \text{ nm}$); IR (ATR): v = 2921, 1684, 1608, 1576, 1540, 1507, 1419, 1240, 1177, 1105, 1017, 853, 803, 775, 705 cm⁻¹. Particle size of the polymer was controlled by grinding and sieving prior to use as catalyst.

Typical Procedure for Aerobic Reduction of Olefins Catalyzed by PS(R)-DVB-Fl

Hydrazine monohydrate (19 mg, 375 μ mol) and acetonitrile (0.7 mL) was mixed for 1 min under vigorous stirring. To the mixture was added **PS(R)-DVB-Fl** (3.75 μ mol, 3 mol %), an ole-fin (125 μ mol), and acetonitrile (0.3 mL), which was stirred at 30 °C for 14 h under air. The reaction yield was determined by either GC analysis with absolute calibration or NMR measurement using 1,3,5-trimethoxybenzene as an internal standard.

Reuse Test of PS(H)-DVB-Fl in Aerobic Reduction of 4-Phenyl-1-Butene

A glass reactor with Teflon filter (MultiSynTech, V050TF118) equipped with a Luer stopper (MultiSynTech, V000LS100) was charged with **PS(H)-DVB-FI** (61 mg, 7.4 μ mol), 4-phenyl-1-butene (33 mg, 0.25 mmol), NH₂NH₂•H₂O (38 mg, 0.75 mmol), and acetonitrile (2 mL). The mixture was agitated using a shaker (125 rpm) at 30 °C under air. The reaction was monitored by GC analysis. The reaction mixture was directly filtered from the glass reactor and the remaining **PS(H)-DVB-FI** was washed with acetonitrile, THF, and ether, which was reused for the next reaction.

RESULTS AND DISCUSSION

Synthesis of Flavin-Containing Styrene Monomer

Riboflavin (Vitamin B₂) is readily available in commerce and its diacetalized derivatives, such as 2',4':3',5'-di-0-methyleneriboflavin (**DMRFI**), have previously proven to be an active catalyst for aerobic reduction of olefins with hydrazine (Fig. 1).^{11(b,c)} Taking this into account, we decided to use riboflavin as a starting material and introduce a styrene moiety to its ribityl group through the 2',4'- or 3',5'-acetal linkage.



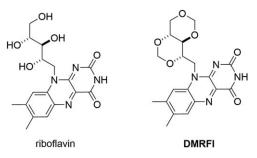
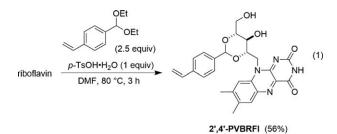


FIGURE 1 Structures of riboflavin and DMRFI.

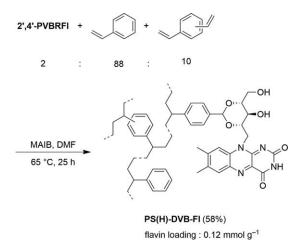
Such partial acetalization of the ribityl group often encounters a synthetic problem even through its protection and deprotection. For example, Yashima and coworkers synthesized 2',4'-*Op*-ethynylbenzilidene riboflavin as a monomer by the reaction of 5'-*O*-trityl riboflavin and 4-ethynylbenzaldehyde dimethyl acetal followed by the deprotection of the trityl group in 11% yield.⁷ By contrast, we found that riboflavin could be readily converted into **2'**,**4'**-**PVBRFI** and its 3',5'-analogue in a ratio of approximately 2:1 with a trace amount of diacetalized product by treating with 2.5 equiv of 4-vinylbenzaldehyde diethyl acetal and 1 equiv of TsOH•H₂O in DMF at 80 °C for 3 hours, which could be separated by a column purification to give **2'**,**4'**-**PVBRFI** in 56% yield (eq 1). The selective formation of **2'**,**4'**-**PVBRFI** can be probably explained by transacetalization–



hydrolysis equilibria that results in thermodynamically more stable **2**',**4**'-**PVBRFI** having a freely rotatable primary alcohol group. The structure of **2**',**4**'-**PVBRFI** was fully characterized by NMR (¹H, ¹³C, COSY, HMBC, HMQC, ROESY), FTIR, mass spectroscopy, and elemental analysis.

Synthesis of Flavin-Containing Cross-Linked Polystyrenes PS-DVB-Supported Flavin

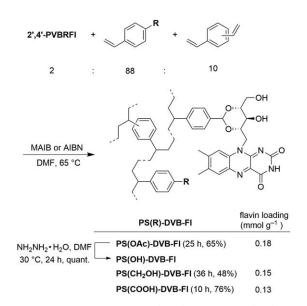
We carried out radical copolymerization of styrene, divinylbenzene, and 2',4'-**PVBRFI** in the molar ratio of 88:10:2 using MAIB as an initiator under heating and inert conditions (Scheme 1). The copolymerization proceeded successfully, which was recognized by gelation of the reaction mixture, to give PS-DVBsupported flavin (**PS(H)-DVB-FI**) in 58% yield after washing with ether, THF, and methanol, followed by drying *in vacuo*. The flavin loading of **PS(H)-DVB-FI** was calculated to be 0.12 mmol g^{-1} from its nitrogen content determined by elemental analysis. In addition, the actual CHN content of **PS(H)-DVB-FI** was in good agreement with the theoretical value, indicating that the present copolymerization took place according to the feed molar ratio.



SCHEME 1 Synthesis of PS(H)-DVB-FI.

Functionalized PS-DVB-Supported Flavins

The choice of support polymer is crucial for developing efficient polymer-supported catalysts since the catalytic reactions take place in microenvironment created in the polymer network.3 In view of the fact that the catalysis of neutral flavin for the aerobic reduction of olefins involves the nucleophilic reduction of a flavin molecule with hydrazine as a rate-determining step,^{11(c)} PS-DVB was supposed to be too hydrophobic as a support for the target reaction because of the strong hydrophilicity of hydrazine. Thus, we designed additional three polymeric flavins PS(CH2OH)-DVB-Fl, PS(OH)-DVB-Fl, and PS(COOH)-DVB-Fl containing different hydrophilic pendant groups in PS-DVB. 4-Vinylbenzyl alcohol, 4-acetoxystyrene or 4-vinylbenzoic acid was copolymerized with divinylbenzene and 2',4'-PVBRFl in the molar ratio of 88:10:2 under radical conditions to afford PS(CH₂OH)-DVB-Fl, PS(OAc)-DVB-Fl, and PS(COOH)-DVB-Fl, respectively, in moderate to high yields (Scheme 2). The acetyl groups in



SCHEME 2 Synthesis of PS(OAc)-DVB-FI, PS(OH)-DVB-FI, PS(CH₂OH)-DVB-FI, and PS(COOH)-DVB-FI.

ARTICLE

PS(OAc)-DVB-FI were quantitatively hydrolyzed by a treatment with an excess of hydrazine in DMF to give **PS(OH)-DVB-FI**, which was confirmed by IR spectroscopic data that showed the disappearance of C = 0 absorbance at 1754 cm⁻¹ and the appearance of –OH absorbance at 3329 cm⁻¹. The flavin loadings of **PS(CH₂OH)-DVB-FI**, **PS(OAc)-DVB-FI**, and **PS(COOH)-DVB-FI** were estimated to be 0.15, 0.18, and 0.13 mmol g⁻¹, respectively, through elemental analyses.

Photophysical Properties of PS(R)-DVB-Fl

Flavins exhibit a characteristic absorption at \sim 440 nm with high molar absorptivities as well as an intense fluorescence emission at \sim 520 nm in solution state.^{1(g)} Indeed, 0.1 mM solution of 2', 4'-**PVBRFI** in ethanol emitted a bright light under irradiation by a range of UV/Vis lights less than \sim 500 nm, and its fluorescence spectrum measured with an excitation wavelength of 442 nm showed the maximum emission peak (λ_{max}) at 516 nm (Supporting Information Fig. S9). Not surprisingly, the fluorescence peak of 2',4'-PVBRFI in solid state appeared at a slightly longer wavelength $(\lambda_{max} = 556 \text{ nm})$ with much lower intensity, due to concentration quenching (Supporting Information Fig. S9). On the other hand, the flavin-containing polymers including PS(H)-DVB-Fl, PS(OAc)-DVB-Fl, and PS(COOH)-DVB-Fl were found to emit light apparently in solid state under 365 nm UV light, except PS(OH)-DVB-Fl (Fig. 2), which led us to explore their emission properties in more detail. We carried out fluorescence measurements for all kinds of PS(R)-DVB-Fl in solid state and determined their solid-state fluorescence quantum

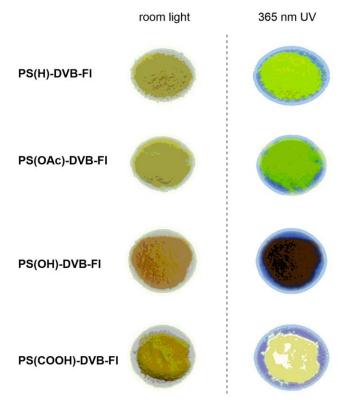


FIGURE 2 Photographs of **PS(R)-DVB-FI** under room light or UV irradiation (365 nm). [Color figure can be viewed at wileyonlinelibrary.com]

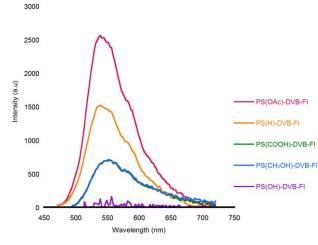
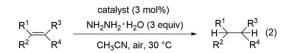


FIGURE 3 Solid-state fluoresence spectra of **PS(R)-DVB-FI** ($\lambda_{ex} = 405$ nm). [Color figure can be viewed at wileyonlinelibrary. com]

yield (ϕ) . As expected, a distinct fluorescence emission at ${\sim}550$ nm was observed for PS(H)-DVB-Fl (λ_{max} =542 nm, $\phi = 10.6\%$), **PS(OAc)-DVB-Fl** ($\lambda_{max} = 542$ nm, $\phi = 16.5\%$), **PS(CH₂OH)-DVB-FI** ($\lambda_{max} = 552$ nm, $\phi = 10.1\%$), and **PS(COOH)-DVB-Fl** ($\lambda_{max} = 552 \text{ nm}, \phi = 9.3\%$) using a 405-nm excitation wavelength (Fig. 3), which would indicate that the immobilized flavin molecules were distributed to the polymer network rather homogeneously. By contrast, PS(OH)-DVB-Fl exhibited almost no fluorescence emission. This is at least not due to heterogeneous dispersion of flavins into the polymer, given the fact that PS(OAc)-DVB-Fl showed the intense fluorescence as mentioned above. Although a clear explanation for the non-fluorescence of PS(OH)-DVB-Fl is not available at the moment, we find it interesting because it may be caused by some interactions between the immobilized flavin molecule and phenolic hydroxyl groups, just like the situation in flavoenzyme.16

Flavin-Catalyzed Aerobic Reduction of Olefins

Aerobic reductions of olefins were carried out with 3 equiv of hydrazine monohydrate in acetonitrile under air at 30 $^{\circ}$ C in the presence of 3 mol % of **PS(R)-DVB-FI** under heterogeneous conditions to evaluate their catalytic activity, in which the polymeric catalysts were ground and sieved to a defined range of particle size prior to use (eq 2).



The reduction of 4-phenyl-1-butene (**1a**) using **PS(H)-DVB-FI** with particle size of 53–100 μ m proceeded smoothly to give butylbenzene (**2a**) in 86% yield and 98% yield after 24 and 48 h, respectively [Fig. 4(b), Run 1], in which a filter-equipped glass reactor was used to operate the following catalyst recycling process as efficient as possible [Fig. 4(a)].¹⁷ Solution phase of the reaction mixture was filtered out and analyzed by GC, whereas the residual solid catalyst was washed with

5

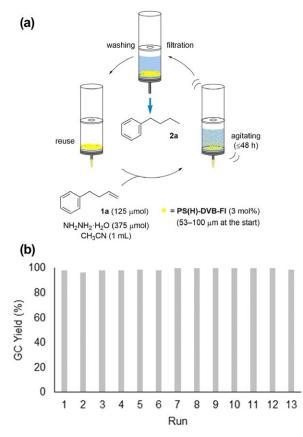


FIGURE 4 Aerobic reduction of **1a** catalyzed by **PS(H)-DVB-FI**: (a) reaction and recycling protocol, (b) reusability of **1a** (Run 1–11, 48 h; Run 12, 24 h; Run 13, 8 h). [Color figure can be viewed at wileyonlinelibrary.com]

acetonitrile, THF, and ether, and dried in vacuo, and reused [Fig. 4(a)]. Using this simple protocol, the excellent chemical robustness and reusability of PS(H)-DVB-Fl could be demonstrated [Fig. 4(b)]. Reactions from the 1st run to the 6th run were carried out successively, in which quantitative yields of 2a were observed without exception. After the 6th run, the reused polymeric flavin catalyst was preserved in a UVshielding amber desiccator for 3 months, and then the reuse test was restarted to evaluate the storage stability of PS(H)-DVB-Fl. The 7th run demonstrated that the catalytic activity was not affected by the storage, which led us to further continue the reuse test until the 11th run under the same conditions. In the 12th run, we were finally aware of complete conversion of 1a to 2a within 24 h, which was much faster than the reaction using a fresh catalyst in the 1st run. Earlier monitoring of the reaction in the 13th run revealed that the reaction was completed already in 8 h. We supposed that this considerable enhancement of the catalytic activity could be attributed to increasing in the surface area of the solid catalyst as the reactions were repeated. To confirm this assumption, we freshly prepared **PS(H)-DVB-FI** with particle size of \leq 53 μ m by careful grinding and sieving, and used it as a catalyst for the same reaction under identical conditions. As expected, nearly quantitative yield was observed in 9 h (Table 1, Entry 1), verifying

the effectiveness of **PS(H)-DVB-Fl** as a readily reusable as well as highly active catalyst for the present reaction.

Next, we explored the catalytic functions of PS(CH2OH)-DVB-Fl, PS(OH)-DVB-Fl, and PS(COOH)-DVB-Fl, which were expected to be more effective than PS(H)-DVB-Fl as a catalyst because of their amphiphilic properties. However, the reduction of 1a to 2a catalyzed by any one of these functionalized polymers with particle size of \leq 53 μ m was not completed even after 24 h (Table 1, Entries 2-4), while PS(H)-**DVB-FI** could efficiently catalyze the reaction (Entry 1) as mentioned. Similar tendency in catalytic activity was observed in the reduction of another simple olefin, ethyl 10undecanoate (1b), using PS(H)-DVB-Fl (Entry 6, 14 h, 96%) and PS(OH)-DVB-Fl (ently 7, 14 h, 61%). The amphiphilicity of the acid-functionalized polymers could actually be recognized by apparent aggregation of the polymer particles in the reaction mixtures containing an excess of hydrophilic hydrazine. Nevertheless, only PS(H)-DVB-Fl, which is strongly hydrophobic and therefore not aggregated, showed extremely high activity, indicating that particle size is a more crucial factor for reaction efficiency in this case. It is important to note that, at least for the reduction of 1a, the catalytic functions of PS(CH2OH)-DVB-Fl, PS(OH)-DVB-Fl, and PS(COOH)-DVB-Fl are not as excellent as PS(H)-DVB-Fl, but still comparable with a homogeneous flavin catalyst, DMRFI (Entry 5), despite their heterogeneity. This fact may be explained by site-isolation effect of the flavin-containing insoluble polymers in which each flavin molecule is isolated from others, that is to say, redoxes of them do not affect each other.

To use the acid-functionalized PS-DVB-supported flavins effectively, we turned our attention to substrates bearing a phenolic hydroxyl group. Given the fact that the ratedetermining step of the neutral flavin-catalyzed aerobic reduction is the nucleophilic addition of hydrazine to flavin,^{11(c)} one can expect that such acidic substrates can be unfavorable to the reaction system due to acid-base interaction that lowers the nucleophilicity of hydrazine. Indeed, in homogeneous reaction system with DMRFI as a catalyst, oallylphenol (1c) and *p*-vinylphenol (1d) underwent reduction very slowly to give o-propylphenol (2c) and p-ethylphenol (2d) in 19% yield (36 h, Entry 11) and in 4% yield (24 h, Entry 15), respectively. On the other hand, our polymersupported flavin catalysts were found to exhibit significantly higher activity for these acidic olefins than DMRFI. Particularly effective were PS(OH)-DVB-Fl as well as PS(COOH)-DVB-Fl over PS(H)-DVB-Fl. For example, the reduction of 1c was catalyzed to furnish 2c in 42% (PS(H)-DVB-Fl, Entry 8), 72% (PS(OH)-DVB-Fl, Entry 9), and 76% yield (PS(COOH)-DVB-Fl, Entry 10), respectively, after 36 h. An even more distinct advantage of these acid-functionalized polymeric flavins was observed in the reduction of 1d, which was converted to 2d in 27% (PS(H)-DVB-Fl, Entry 12), 70% (PS(OH)-DVB-Fl, Entry 13), and >99% yield (PS(COOH)-DVB-Fl, Entry 14), respectively, after 24 h. These results demonstrate that the acid-functionalized polymeric flavins

JOURNAL OF Polymer POLYMER SCIENCE Chemistry

Entry	Substrate	Product	Catalyst ^b	Time (h)	Yield (%)
1	1a	2a	PS(H)-DVB-FI	9	95
2	1a	2a	PS(CH ₂ OH)-DVB-FI	24	68
3	1a	2a	PS(OH)-DVB-FI	24	76
4	1a	2a	PS(COOH)-DVB-FI	24	61
5	1a	2a	DMRFI	24	56
6			PS(H)-DVB-FI	14	96
7	1b	2b	PS(OH)-DVB-FI	14	61
8	OH 1c	OH 2c	PS(H)-DVB-FI	36	42
9	1c	2c	PS(OH)-DVB-FI	36	72
10	1c	2c	PS(COOH)-DVB-FI	36	76
11	1c	2c	DMRFI	36	19
12	HO 1d	HO 2d	PS(H)-DVB-FI	24	27
13	1d	2d	PS(OH)-DVB-FI	24	70
14	1d	2d	PS(COOH)-DVB-FI	24	>99
15	1d	2d	DMRFI	24	4

TABLE 1 Flavin-Catalyzed Aerobic Reduction of Olefins^a

^a Reactions were performed using 125 μ mol of the olefin and 375 μ mol of hydrazine monohydrate in 1 mL of acetonitrile in the presence of 3 mol % of the catalyst under air at 30 °C.

^b Polymeric catalysts with particle size of \leq 53 μ m were used.

^c Determined by GC analysis (Entries 1–5) or ¹H NMR measurement (Entries 6–15) with internal standard.

the reaction unlike with the acid-functionalized PS-DVB-sup-

ported flavins, (iii) the acid functionalized PS-DVB-

supported flavins such as PS(OH)-DVB-Fl and PS(COOH)-

DVB-Fl have an enhanced affinity for hydrazine, which

allows for efficiently catalyzing the reduction of phenolic

hydroxyl group-containing olefins known as less reactive

substrates in conventional homogeneous system. Efficient

catalyst recovery and reuse were demonstrated with PS(H)-

DVB-Fl that could be readily recovered by a simple filtration

and reused without loss in activity at least until 13th run. The results are currently leading us to the synthesis of

morphology-controlled flavin-containing insoluble polymers, such as microspheres and mesoporous network polymers,

using other polymerization techniques, and their application

to the development of a continuous flow system for the pre-

This work was supported by Grant-in-Aid for Scientific

Research on Innovative Areas 'Advanced Molecular Transfor-

sent aerobic reduction.

ACKNOWLEDGEMENTS

REFERENCES AND NOTES

mations by Organocatalysts' from MEXT.

are highly effective catalysts for the present aerobic reduction of olefins bearing a phenolic hydroxy group. This finding can be explained by the high affinity of these flavin polymers for hydrazine via acid-base interaction, which can make the concentration of hydrazine on solid phase higher and that in solution phase lower, and as a result, the rate-determining nucleophilic addition of hydrazine to flavin favorable. Finally, it should also be noted that, in the reduction of **1d** to **2d**, **PS(COOH)-DVB-FI** could be reused without loss in the activity at least 3 times (see Supporting Information).

CONCLUSIONS

We introduced a novel flavin-containing vinyl monomer, 2',4'-PVBRFl, and its copolymers, PS(R)-DVB-Fl, as efficient and reusable heterogeneous catalysts for aerobic reduction of olefins with hydrazine. The monomer was easily synthesized from commercially available riboflavin in one step, and the polymeric catalysts were readily prepared by free radical solution copolymerization. Exploring the catalytic activities of PS(R)-DVB-Fl in aerobic reduction of olefins with hydrazine revealed that (i) PS(R)-DVB-Fl exhibit comparable activities to the non-supported counterpart DMRFl, possibly because of flavin molecules dispersedly immobilized onto supports as demonstrated by their characteristic solid-state fluorescence, (ii) the non-functionalized PS-DVBsupported flavin, PS(H)-DVB-Fl, is particularly effective for aprotic substrates, which can be attributed to its strong hydrophobicity not allowing for aggregation of itself during



7

1 (a) T. C. Bruice, Acc. Chem. Res. 1980, 13, 256-262; (b) C.

Walsh, Acc. Chem. Res. 1980, 13, 148-155; (c) D. P. Ballou, In

Flavins and Flavoproteins: Flavoprotein Monooxygenase; V.

Massey, C. H. Williams, Eds.; Elsevier: New York, **1982**; pp 301– 310; (d) Chemistry and Biochemistry of Flavoenzymes; F. Müller, Ed.; CRC Press: Boston, **1991**; (e) R. B. Silverman, *Acc. Chem. Res.* **1995**, *28*, 335–342; (f) N. M. Kamerbeek, D. B. Janssen, W. J. H. van Berkel, M. W. Fraaije, *Adv. Synth. Catal.* **2003**, *345*, 667–678; (g) Flavins—Photochemistry and Photobiology; E. Silva, A. M. Edwards, Eds.; Royal Society of Chemistry: Cambridge, **2006**; (h) M. W. Fraaije, D. B. Janseen, In Modern Biooxidations—Enzymes, Reactions and Applications; R. D. Schmid, V. Urlancher-Kursif, Eds.; Wiley-VCH: Weinheim, **2007**, p 77–97; (i) M. Insińska-Rak, M. Sikorski, *Chem. Eur. J.* **2014**, *20*, 15280–15291.

2 (a) H. lida, Y. Imada, S.-I. Murahashi, *Org. Biomol. Chem.* **2015**, *13*, 7599–7613; (b) R. Cibulka, *Eur. J. Org. Chem.* **2015**, 915–932; (c) G. de Gonzalo, M. W. Fraaije, *ChemCatChem* **2013**, *5*, 403–415; (d) Y. Imada, T. Naota, *Chem. Rec.* **2007**, *7*, 354– 361; (e) F. G. Gelalcha, *Chem. Rev.* **2007**, *107*, 3338–3361; (f) J.-E. Bäckvall, In Modern Oxidation Methods; J.-E. Bäckvall, Ed.; Wiley-VCH: Weinheim, **2004**, p 193–222.

3 (a) Polymeric Chiral Catalyst Design and Chiral Polymer Synthesis; S. Itsuno, Ed.; Wiley: Hoboken, **2011**; (b) A. F. Trindade, P. M. P. Gois, C. A. M. Afonso, *Chem. Rev.* **2009**, *109*, 418–514.

4 For the first linear soluble polymer-supported flavin, see: H. Kamogawa, J. Polym. Sci. Part A: Polym. Chem. **1962**, 7, 409–413.

5 For soluble polymer-supported flavins as oxidation catalysts, see: (a) S. Shinkai, S. Yamada, T. Kunitake, *Macromolecules* **1978**, *11*, 65–68; (b) S. Shinkai, R. Ando, T. Kunitake, *Biopolymers* **1978**, *17*, 2757–2760; (c) W. J. Spetnagel, I. M. Klotz, *Biopolymers* **1978**, *17*, 1657–1668; (d) S. Shinkai, K. Mori, Y. Kusano, O. Manabe, *Bull. Chem. Soc. Jpn.* **1979**, *52*, 3606–3610; (e) J. P. C. Bootsma, G. Challa, F. Müller, *J. Polym. Sci: Polym. Chem. Ed.* **1984**, *22*, 705–719; (f) H. F. M. Schoo, G. Challa, *Polymer* **1990**, *31*, 1559–1563; (g) H. F. M. Schoo, G. Challa, *Macromolecules* **1992**, *25*, 1633–1638; (h) H. Iida, S. Iwahana, T. Mizoguchi, E. Yashima, *J. Am. Chem. Soc.* **2012**, *134*, 15103–15113.

6 For soluble polymer-supported flavins as molecular recognition platforms, see: (a) J. B. Carroll, B. J. Jordan, H. Xu, B. Erdogan, L. Lee, L. Cheng, C. Tiernan, G. Cooke, V. M. Rotello, *Org. Lett.* **2005**, *7*, 2551–2554; (b) B. J. Jordan, G. Cooke, J. F. Garety, M. A. Pollier, N. Kryvokhyzha, A. Bayir, G. Rabani, V. M. Rotello, *Chem. Commun.* **2007**, 1248–1250; (c) D. Patra, C. Pagliuca, C. Subramani, B. Samanta, S. S. Agasti, N. Zainalabdeen, S. T. Caldwell, G. Cooke, V. M. Rotello, *Chem.*

Commun. **2009**, 4248–4250; (d) C. Subramani, G. Yesilbag, B. J. Jordan, X. Li, A. Khorasani, G. Cooke, A. Sanyal, V. M. Rotello, *Chem. Commun.* **2010**, *46*, 2067–2069; (e) T. P. Thomas, S. K. Choi, M.-H. Li, A. Kotlyar, J. R. Baker Jr., *Bioorg. Med. Chem. Lett.* **2010**, *20*, 5191–5194; (f) A. B. Witte, C. M. Timmer, J. J. Gam, S. K. Choi, M. M. Banaszak Holl, B. G. Orr, J. R. Baker, K. Sinniah, *Biomacromolecules* **2012**, *13*, 507–516.

7 For a flavin-containing helical polymer, see: H. lida, T. Mizoguchi, S. D. Oh, E. Yashima, *Polym. Chem.* **2010**, *1*, 841–848.

8 H. Schmaderer, P. Hilgers, R. Lechner, B. König, *Adv. Synth. Catal.* 2009, *351*, 163–174.

9 (a) Handbook of Homogeneous Hydrogenation; J. G. de Vries, C. J. Elsevier, Eds.; Wiely-VCH: Weinheim, **2007**; (b) S. Nishimura, Handbook of Heterogeneous Catalytic Hydrogenation for Organic Synthesis; John Wiley & Sons: New York, **2001**.

10 (a) S. Hünig, H. R. Müller, W. Thier, *Angew. Chem. Int. Ed. Engl.* **1965**, *4*, 271–280; (b) D. J. Pasto, R. T. Taylor, *Org. React.* **1991**, *40*, 91–155.

11 (a) Y. Imada, H. Iida, T. Naota, *J. Am. Chem. Soc.* **2005**, *127*, 14544–14545; (b) Y. Imada, T. Kitagawa, T. Ohno, H. Iida, T. Naota, *Org. Lett.* **2010**, *12*, 32–35; (c) Y. Imada, H. Iida, T. Kitagawa, T. Naota, *Chem. Eur. J.* **2011**, *17*, 5908–5920.

12 (a) C. Smit, M. W. Fraaije, A. J. Minnaard, *J. Org. Chem.* **2008**, *73*, 9482–9485; (b) J. F. Teichert, T. den Hartog, M. Hanstein, C. Smit, B. ter Horst, V. Hernandez-Olmos, B. L. Feringa, A. J. Minnaard, *ACS Catal.* **2011**, *1*, 309–315; (c) B. J. Marsh, E. L. Heath, D. R. Carbery, *Chem. Commun.* **2011**, *47*, 280–282.

13 K. Suzuki, A. Kobayashi, S. Kaneko, K. Takehira, T. Yoshihara, H. Ishida, Y. Shiina, S. Oishi, S. Tobita, *Phys. Chem. Chem. Phys.* **2009**, *11*, 9850–9860.

14 R. J. Mancini, J. Lee, H. D. Maynard, *J. Am. Chem. Soc.* **2012**, *134*, 8474–8479.

15 O. Shimomura, B. S. Lee, S. Meth, H. Suzuki, S. Mahajan, R. Nomura, K. D. Janda, *Tetrahedron* **2005**, *61*, 12160–12167.

16 (a) L. L. Poulsen, D. M. Ziegler, *J. Biol. Chem.* **1979**, *254*, 6449–6455; (b) V. Massey, P. Hemmerich, *Biochem. Soc. Trans.* **1980**, *8*, 246–257; (c) N. B. Beaty, D. P. Ballou, *J. Biol. Chem.* **1980**, *255*, 3817–3819.

17 Y. Arakawa, M. Wiesner, H. Wennemers, *Adv. Synth. Catal.* **2011**, *353*, 1201–1206.