

Ruthenium(II)-Arene Metallacycles: Crystal Structures, Interaction with DNA and Cytotoxicity

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Abstract: A series of 24, 26-membered Ru(II)₂ metallamacrocycles containing 1-(3-((1H-imidazol-1-yl)methyl)benzyl)-1H-imidazole (mbib) and 1-(4-((1H-imidazol-1-yl)methyl)benzyl)-1H-imidazole (p-bib) ligands have been synthesized and characterized. X-ray crystal structures of $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2Cl_2](CF_3SO_3)_2$ (2), $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2(m-bib)_2Cl_2](CF_3SO_3)_2$ (2), $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2(m-bib)_2(m-bib)_2Cl_2](CF_3SO_3)_2$ (2), $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2(m-b$ cymene)₂(*m*-bib)₂Cl₂](SbF₆)₂ (**3**), $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2l_2]l_2$ (**6**), and $[Ru_2(\eta^6-p-cymene)_2(p-bib)_2Cl_2](CF_3SO_3)_2$ (8) were determined and found to exhibit chair-like conformations. In general, the complexes exhibited little or moderate anti-proliferative activity towards cancer cells (human lung cancer cells (A549), breast adenocarcinoma cells (MCF-7), cervical epithelioid carcinoma cells (HeLa)) as well as normal liver cells (L02), except [Ru₂(η^6 -pcymene)₂(p-bib)₂Cl₂](NO₃)₂ (10) (IC₅₀ = 16.7 μ M) which had activity comparable with the anticancer drug cisplatin (IC₅₀ = 15.8 μ M). Gel electrophoresis studies suggested that the complexes can interacted with DNA and induce DNA condensation.

Introduction

Transition metal complexes play an important role in the design of metal-based drugs,[1] providing a variety of coordination numbers and geometries,^[2] the diversity of linking ligands,^[3] and structures.^[4] The square-planar complex cisplatin in particular is now widely used for the treatment of ovarian, head and neck, bladder, cervical and lymphomas cancers.^[5] However, the clinical use of cisplatin still has some limitations including drug resistance and side effects, e.g. neurotoxicity and gastrointestinal toxicity.^[6] Ruthenium-based complexes, are potential anticancer metal-based candidates,^[7] showing promising activity.^[8] KP1019^[9] has been in phase II clinical trials. Ruthenium(II)-arene complexes have been shown promising in vitro or in vivo biological activity in various studies.^[10] Halfsandwich ruthenium(II)-arene complexes of the general type $[Ru(\eta^6-arene)(YZ)X]^{n+}$ have also attracted attention as potential anticancer drugs.^[11] The monodentate ligand X and chelated YZ of these "piano-stool" structures have an important influence on the antiproliferative activity.^[12] The leaving group X affects the

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DNA in cancer cells is a target for some classes of ruthenium(II)-arene complexes.[19] These complexes can bind directly to DNA bases especially guanine, and also intercalate into DNA when the arene is extended and generate adducts which inhibit cell division^[17] and proliferation.^[12,20-21] Linking Ru(II) arene centres can endow multinuclear complexes have new modes of interaction with DNA. Modification of the ligands or bridging linkers can lead to significant differences in anticancer activity and different types of DNA binding modes.^[22] For example, the ligands containing N-donors (e.g. polypyridine ligands) and their derived complexes of general type $[(n^{6}$ arene)RuX(K²-N.N-L)]Y display promising anticancer activity.^[23] We report herein the synthesis and characterization of a series novel metallacvclic diruthenium(II)-arene complexes of containing flexible bidentate imidazole ligands *m*-bib or *p*-bib. and their antiproliferative activities have been evaluated. The binding of these diruthenium metallamacrocycles to DNA has been investigated by gel electrophoresis. Metallacyclic dinuclear ruthenium complexes offer the potential for new anticancer agents with novel modes of binding to DNA.

Results and Discussion

Design and Synthesis of Macrocyclic Diruthenium Complexes (1-12)

Two flexible ligands 1-(3-((1H-imidazol-1-yl)methyl)benzyl)-1Himidazole (*m*-bib), 1-(4-((1H-imidazol-1-yl)methyl)benzyl)-1Himidazole (*p*-bib) were prepared according to previous reports.^[24] Macrocyclic diruthenium complexes (1, 2, 7 and 8) were obtained from the direct reaction of the appropriate ligands (*m*bib or *p*-bib) with the dimer [Ru(η^6 -*p*-cymene)Cl₂]₂ (Scheme 1), complexes (3-6 and 9-12) were prepared by treating complexes [Ru₂(η^6 -*p*-cymene)₂(*m*-bib)₂Cl₂](Cl)₂ (1) or [Ru₂(η^6 -*p*-cymene)₂(*p*bib)₂Cl₂](Cl)₂ (7) with appropriate metal salt.^[25] All complexes were obtained in good yields and characterized by ¹H NMR spectroscopy, mass spectrometry, and elemental analysis (see Experimental section and supporting information for full details). X-ray Crystallography

The crystal structures of 4 diruthenium-arene complexes (2, 3, 6 and 8) were determined by X-ray crystallographic method. These complexes contain two ruthenium(II) centers linked by two flexible ligands (Figures 1-4). Details of data collection,

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10.1002/ejic.201601226

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Scheme 1. Structures of metallamacrocyclic diruthenium complexes.

structure solutions, and refinement are given in Table S1, and selected bond lengths and angles are shown in Table S2.

Complex 2 crystallized in a triclinic Pi space group, and showed a classic "chair-like" metallamacrocyclic structure with CF₃SO₃ counter anions (Figure 1). Two *m*-bib ligands link the two ruthenium centres. Each Ru was coordinated by two N atoms from two ligands (m-bib) and one Cl atom. The two imidazole groups are located on the opposite faces of the central aromatic ring forming a trans conformation. The bond lengths of Ru-N and Ru-Cl are 2.111(6) Å, 2.135(6) Å, and 2.4315(17) Å, respectively, and angles N1a-Ru1-Cl1, N1a-Ru1a-N4, N4-Ru1-Cl1 are $85.32(16)^{\circ}$, $83.3(2)^{\circ}$, $88.01(16)^{\circ}$, respectively. The CF_3SO_3 anion is located outside of the $"Ru_2L_2"$ metallamacrocycle. The fluorine atoms of CF_3SO_3 form hydrogen bonds (C-H···F) with a C atom on the methylene of the *m*-bib ligand, with C---F distance of 3.10 Å (Table S3). π - π interactions are present between two paralleled phenyl rings from two adjacent units with a "center-to-center" distance of 3.71 Å. The packing of "Ru₂L₂" units is shown in Figure 1d.



Figure 1. (a) X-ray crystal structure of **2**. (b) Hydrogen bonds (C–H···F) between $CF_3SO_3^-$ and the unit "Ru₂L₂". (c) The π - π stacking interactions between two parallel phenyl rings of two adjacent units, with a "centre-to-centre" distance of 3.71 Å. (d) The packing of the "Ru₂L₂" units of **2**. Counter anions, solvent molecules and hydrogen atoms have been omitted for clarity.

Complex **3** showed a similar structure to that of complex **2**, in which each Ru atom coordinated by two N atoms from two *m*-bib ligands and one CI atom, forming a slightly distorted metallamacrocyclic structure (Figure 2). The SbF₆⁻ anions are located outside of metallamacrocycle. There are different types of H-bonds between F atoms of SbF₆⁻ and η^6 -*p*-cymene ligand. One type of H-bond (C1-H1...F1) is between F1 atom and a C atom on the methyl group of η^6 -*p*-cymene ligand, another type of H-bond (C2-H2...F2) is between F2 atom and a C atom from benzene ring of η^6 -*p*-cymene ligand. The C1---F1 and C2---F2 distances are 3.28 Å, and 3.38 Å, respectively. Hydrogen bonds between the F3 atom and η^6 -*p*-cymene ligand are also observed

(C3-H3...F3, C4-H4...F3) with bond lengths of 3.48 Å or 3.30 Å, respectively (Table S3). It is noteworthy that 3 fluorine atoms of SbF_6^- connect two Ru_2L_2 units by hydrogen bonds, forming a two-dimensional network, in which the Ru_2L_2 units have two different orientations.

Complex **6** was obtained by treating **1** with KI in methanol and its yellow crystals were suitable for X-ray analysis. Complex **6** also crystallized in the monoclinic $P2_1/c$ space group, each ruthenium centre is coordinated by an lodine atom and two N atoms from two *m*-bib ligands, forming a "chair-like" metallamacrocyclic (Figure 3).



Figure 2. (a) X-ray crystal structure of 3. (b) Hydrogen bond $(C-H\cdots F)$ between SbF₆ and the unit "Ru₂L₂". (c) The packing of the "Ru₂L₂" units of 3. Counter anions, solvent molecules and hydrogen atoms have been omitted for clarity.

The Ru_2L_2 units of **6** show two different orientations in the packing structure (Figure 3b).

The structure of complex 8 containing the p-bib ligands was determined by single crystal X-ray diffraction (Figure 4). 8 crystallized in an orthorhombic P212121 space group, forming a metallamacrocyclic structure with two p-bib ligands and two Ru atoms. Each Ru atom is coordinated by one CI atom and two N atoms from two ligands. The Ru-N bond lengths range from 2.091(13) Å to 2.129(13) Å, and the Ru-Cl bond lengths from 2.411(4) Å to 2.426(4) Å. These bond lengths are slightly shorter than those in the Ru(II) arene *m*-bib structures (2, 3 and 6). The angles N-Ru-N range from 86.6(5)° to 89.7(5)°, and N-Ru-Cl from $84.3(4)^{\circ}$ to $88.3(4)^{\circ}$, similar to those of the *m*-bib complexes. H-bonds are formed between CF₃SO₃ anion and three surrounding Ru₂L₂ units. As shown in the Figure 4b, the representative H-bonds around CF₃SO₃⁻ anion include those between the C-H from the methylene of p-bib ligand and F1 atom (C1-H1...F1), the C-H from the isopropyl group of η^6 -pcymene and F2 atom (C2-H2···F2), between the C-H from the imidazole of p-bib ligand and F3 atom (C3-H3...F3), with C---F distances of 3.24 Å, 3.36 Å or 3.25 Å, respectively (Table S3). The Ru₂L₂ units of 8 also show two different orientations in the packing structure (Figure 4c). These results indicate that Ru₂L₂ metallamacrocyclic complexes can be obtained by reaction of dimer $[Ru(\eta^6-p-cymene)Cl_2]_2$ with flexible bidentate imidazole ligands *m*-bib or *p*-bib. These metallamacrocyclic complexes adopt the "chair-like" conformation with the two coordinated halogen atoms X (CI, or I) located on the opposite sides of the macrocyclic ring.



Figure 3. (a) X-ray crystal structure of 6. (b) The packing of the "Ru₂L₂" units of 6. Solvent molecules and hydrogen atoms have been omitted for clarity.



Figure 4. (a) X-ray crystal structure of **8**. (b) 3 different types of hydrogen bonds between $CF_3SO_3^-$ and the " Ru_2L_2 " unit. (c) Two packing modes for **8**. Counter anions, solvent molecules and hydrogen atoms have been omitted for clarity.

Anticancer activity assays

The anti-proliferative activities of complexes 1-12 were investigated in three cancer cell lines: human lung cancer cell line (A549), human breast adenocarcinoma cell line (MCF-7), and human cervical epithelioid carcinoma (HeLa) as well as a human normal liver cell line (L02) (Table S4). The cytotoxicity was determined by the MTT assay after 44 h of exposure to the complexes. The IC_{50} values are summarized in Table S4. Complexes 1-5, 7-9, 11-12 were nontoxic (IC₅₀>100 μ M) towards the human normal liver cell line (L02). 10 shows much higher anti-proliferative activity to the cancer cells than the other complexes, with IC₅₀ values ranging from 16.7 μ M to 30.1 μ M, and 10 is more active towards cancer cells than towards the human normal L02 cell line (35.6 µM). Potency towards HeLa cells decreased in the order 4>3>5>1>6>2 for the m-bib ligand and in the order 10>9>7>12>8>11 for the p-bib ligand. Additionally, the data obtained from MTT assay clearly indicate that all the complexes except 10 show low cytotoxicity (IC₅₀>100 µM) towards MCF-7, and A549 cell lines. Complex 11 displays good cytotoxicity towards the A549 cell line (43.9 \pm 1.4 μ M) compared with other cell lines (IC₅₀>200 μ M). The antiproliferative activity of **10** towards the HeLa cell line (IC₅₀=16.7 μ M) is comparable with that of the anticancer drug cisplatin (IC₅₀=15.8 μ M). Based on these results, it would appear that the specific molecular shape, the chemical structure, and the nature of anion may play important roles in the cytotoxicity of these complexes.

Solubility studies

All the complexes can be dissolved in DMSO, CH₃OH, 5% DMSO/95% H₂O, and complexes can also be dissolved in CH₂Cl₂, CHCl₃ except that complexes with iodide ligand are only slightly soluble. However, the solubility of the complexes in water differs, decreasing in the order $1>2>4>5>6\approx3$ for the *m*-bib ligand and in the order 7>8>10>11>12>9 for the *p*-bib ligand (Figure S1 in Supporting Information).

DNA electrophoresis studies

Gel electrophoresis assays were used for further studies of the interaction between DNA and these ruthenium(II) macrocycles. Positively-charged ruthenium complexes can usually bind to negatively charged DNA, [7b, 26] so we expected that the binding of closed circular DNA to these macrocyclic ruthenium complexes would result in decreasing the migration rate of supercoiled plasmid (Form II) in agarose gel electrophoresis and formation of a new band for condensed DNA (Form I), as reported in other studies.^[26a] As shown in the Figures 5 and S2, it was clear that the condensation of supercoiled pBR322 DNA occurs in the presence of complexes 1, 4, 5, 6, 7, 10, 11 and 12. With the increasing mol ratio of complexes, the extent of condensation Form I bands increased and the density of supercoiled bands (Form II) decreased. These results indicate that dinuclear ruthenium complexes 1, 4, 5, 6, 7, 10, 11 and 12 can significantly affect the structure of the supercoiled DNA, that probably could lead to distortions of nuclear DNA and cell death.[7b,27,28]



Figure 5. (a) Agarose gel electrophoresis bands for supercoiled pBR322 plasmid DNA (10 μ M) after incubation with complexes **1** and **4**, and (b) complexes **7** and **10** in PBS buffer (100 mM) at 310 K for 12 h. Complex concentrations are 10, 20, 30, 40, 50 and 60 μ M.

Stability studies

To investigate the stability of complexes, 1, 5, 6, 7, 11, 12 were monitored in 5% DMSO/95% H_2O (v/v) at 298 K by UV-vis spectroscopy. As shown in Figure S3, the absorption spectra of 1, 5, 6, 7, 11, 12 show only little or no change with time, indicating that these complexes are stable in aqueous solution at 298 K.

Cellular uptake studies

FULL PAPER

investigate whether intracellular ruthenium In order to concentration correlate with cytotoxicity, intracellular concentration of ruthenium was determined by inductively coupled plasma mass spectrometry (ICP-MS). After incubation with 20 μ M complexes for 24 h, the intracellular ruthenium concentrations are shown in Figure 6. Lipophilicity, molecular size and many other aspects may influence the cellular uptake as previously suggested.^[29-31] The Ru content from 1, 4, 5, 6, 7, 10, 11 and 12 in HeLa cells decreases in the order 1>5>6=4 and 7>11>12>10 for complexes with the m-bib and p-bib ligand, respectively. Interestingly, the trend of intracellular ruthenium concentrations does not correlate with their cytotoxicity. For instance, the higher intracellular ruthenium concentration of 7 was not matched by higher cytotoxicity compared with other complexes.



Figure 6. The concentration of Ru in HeLa cells after exposing to 20 μ M of complexes 1, 4, 5, 6, 7, 10, 11 and 12 for 24 h in a drug-free medium.

Conclusions

In summary, 12 new metallamacrocyclic Ru₂L₂ complexes containing two either *m*-bib or *p*-bib imidazole ligands have been synthesized and characterized. Single crystal structure analyses showed that these complexes were diruthenium-arene Ru₂L₂ metallacycles with a trans conformation, a "titled chair". Gel electrophoresis studies revealed that they can affect the structure of supercoiled DNA significantly, and induce DNA condensation. Furthermore, complex 10 exhibited antiproliferative activity toward HeLa cancer cell line comparable with that of cisplatin (Table S4). Further studies will focus on the chemical and biochemical reactivity of these metallamacrocycles and the mechanism of anticancer activity.

Experimental Section

Materials and methods

All chemicals used in this work were purchased from commercial sources and used without further purification. The dimer [Ru(η^6 -p-cymene)Cl₂]₂ and two ligands 1-(3-((1H-imidazol-1-yl)methyl)benzyl)-1H-imidazole (*m*-bib), 1-(4-((1H-imidazol-1-yl)methyl)benzyl)-1H-imidazole (*p*-bib) were prepared according to literature procedures.^[24] The ruthenium complexes were prepared according to literature procedures.^[25] ¹H NMR spectra were recorded on a Bruker Avance II 400 spectrometer at 298 K. The chemical shifts are

reported in parts per million (ppm) and referenced to residual signals of deuterated solvents. Electrospray ionization mass spectra (ESI-MS) were obtained on a Mass Lynx operating system. Elemental analyses (C, H, N) were performed with a Perkin-Elmer 240C elemental analyzer. UV/Vis spectra were recorded on a Varian Cary 50 spectrophotometer. The X-ray crystallographic data were collected on a Bruker Smart Apex II CCD Diffractometer using graphitemonochromatized Mo- K_{α} radiation (λ = 0.71073 Å) from a rotatinganode generator. Gel electrophoresis was performed on a DYY-2C gel electrophoresis spectrometer. Agarose gel electrophoresis of pBR322 DNA was visualized using the gel imaging system (RAD ChemiDox XPS, American)

Synthesis of complexes

Synthesis of [Ru₂(*η*6-*p*-cymene)₂(*m*-bib)₂Cl₂]Cl₂(1)

To a stirring solution of $[RuCl_2(\eta^6.p-cymene)]_2$ (61.2 mg, 0.1 mmol) in MeOH (12 mL), a solution of the ligand *m*-bib (47.8 mg, 0.2 mmol) in MeOH (3 mL) was added dropwise under argon. The resulting mixture was stirred at 338 K for 12 h. The solvent was removed under reduced pressure by a rotary evaporator to give a crude product. Purification by flash column chromatography using CH₂Cl₂/MeOH elution mixture afforded the product as a yellow solid (40.6 mg, yield 37.2%). ¹H NMR (CDCl₃, 400 MHz) δ : 9.41 (2H, s, H_{im}), 9.26 (2H, s, H_{im}), 8.15 (2H, s, H_{im}), 7.49 (4H, d, H_{im}), 6.85 (8H, m, H_{bz}, *m*-bib), 6.50 (2H, d, H_{im}), 5.92 (8H, d, H_{bz}, *p*-cymene), 5.39 (4H, d, CH₂), 4.94 (4H, d, CH₂), 2.51 (2H, m, CH), 1.89 (6H, s, CH₃), 1.16 (12H, d, CH₃). ESI-MS(+): calcd for [1-2Cl]²⁺ m/z 509.03, found *m*/z 509.25; calcd for [1-Cl]⁺ m/z: 1053.52, found *m*/z 1054.75. Elemental analysis (%): Calcd for Ru₂C₄₈H₅₆N₈Cl₄·CH₂Cl₂·3H₂O C 47.93, H 5.25, N 9.13; found C 48.11, H 5.31, N 9.02.

Synthesis of $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2Cl_2](CF_3SO_3)_2$ (2)

To a solution of [RuCl₂(η^6 -p-cymene)]₂ (61.2 mg, 0.1 mmol) in CH₂Cl₂ (15 mL), a solution of AgCF₃SO₃ (51.4 mg, 0.2 mmol) in EtOAc (5 mL) was added. The mixture was further stirred for 2 h at room temperature, the AgCl precipitate was removed by filtration, and *m*-bib (47.8 mg, 0.2 mmol) was added to the resulting solution. The resulting mixture was stirred at RT for another 12 h, and then the solvent was removed on a rotary evaporator. The residue was recrystallized from CHCl₃ to give yellow crystals (41.1 mg, yield 33.1%). ¹H NMR (CDCl₃, 400 MHz) δ : 8.59 (2H, s, H_{im}), 8.42 (2H, s, H_{im}), 7.58 (2H, s, H_{im}), 7.41 (4H, d, H_{im}), 7.07 (2H, m, H_{im}), 6.87 (4H, m, H_{bz}, *m*-bib), 6.73 (2H, d, H_{bz}, *m*-bib), 6.44 (2H, d, H_{bz}, *m*-bib) 5.82 (4H, d, H_{bz}, *p*-cymene), 5.70 (4H, d, H_{bz}, *p*-cymene), 5.24 (4H, d, CH₂), 4.96 (4H, d, CH₂), 2.54 (2H, m, CH), 1.91 (6H, s, CH₃), 1.20 (12H, d, CH₃). ESI-MS(+): calcd for [**2**-CF₃SO₃]^{*} *m/z* 1167.14, found *m/z* 1166.08. Elemental analysis (%): Calcd for Ru₂C₅₀H₅₆N₆Cl₂F₆S₂O₆ C 45.63, H 4.29, N 8.51; found C 45.34, H 4.43, N 8.05.

Synthesis of $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2CI_2](SbF_6)_2$ (3)

To a solution of $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2Cl_2](Cl)_2$ (21.76 mg, 0.02 mmol) in MeOH (5 mL were added 5 mL MeOH solution of AgSbF₆ (13.74 mg, 0.04 mmol). The mixture was stirred at room temperature for 2 h, and then the solvent was removed on a rotary evaporator. The residue was recrystallized from CHCl₃ to give yellow crystals (11.3 mg, yield 37.9%). ¹H NMR (CDCl₃, 400 MHz) δ : 9.41 (2H, s, H_{im}), 9.26 (2H, s, H_{im}), 8.15 (2H, s, H_{im}), 7.49 (4H, d, H_{im}), 6.87 (8H, m, H_{bz}, *m*-bib), 6.53 (2H, d, H_{im}), 5.93 (8H, d, H_{bz}, *p*-cymene), 5.36 (4H, d, CH₂), 5.01 (4H, d, CH₂), 2.52 (2H, m, CH), 1.89 (6H, s, CH₃), 1.17 (12H, d, CH₃). ESI-MS(+): calcd for [3-2SbF₆]²⁺ *m/z* 509.03, found *m/z* 509.17. Elemental analysis (%): Calcd for $Ru_2C_{48}H_{56}N_8Cl_2Sb_2F_{12}C$ 38.70, H 3.79, N 7.52; found C 38.62, H 3.66, N 7.38.

Synthesis of $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2Cl_2](NO_3)_2$ (4)

To a solution of $[Ru_2(\eta^6-\rho-cymene)_2(m-bib)_2Cl_2](Cl)_2$ (21.76 mg, 0.02 mmol) in MeOH (5 mL) were added 5 mL MeOH solution of AgNO₃ (6.79 mg, 0.04 mmol). The mixture was stirred at room temperature for 2 h, and then the solvent was removed under reduced pressure. Purification by flash column chromatography with CH₂Cl₂/MeOH afforded the product as a yellow solid (10.2 mg, yield 44.6%). ¹H NMR (CDCl₃, 400 MHz) δ : 8.85 (2H, s, H_{im}), 8.74 (2H, s, H_{im}), 7.58 (2H, s, H_{im}), 7.41 (4H, d, H_{im}), 6.90 (8H, m, H_{bz}, *m*-bib), 6.61 (2H, d, H_{im}), 5.85 (4H, d, H_{bz}, *p*-cymene), 5.73 (4H, d, H_{bz}, *p*-cymene), 5.27 (4H, d, CH₂), 4.97 (4H, d, CH₂), 2.56 (2H, m, CH), 1.89 (6H, s, CH₃), 1.18 (12H, d, CH₃). ESI-MS(+): calcd for [4-

FULL PAPER

Synthesis of $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2Br_2](Br)_2(5)$

A solution of $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2Cl_2](Cl)_2$ (43.52 mg, 0.04 mmol) in MeOH (5 mL) was stirred at room temperature for 30 min. A solution of KBr (50 equiv, 238 mg) in MeOH (15 mL) was added dropwise to the mixture and stirred for another 24 h. The solvent was removed under reduced pressure. The residue was subsequently purified by silica gel chromatography to give yellow solid (21.8 mg, yield 43.0%). ¹H NMR (DMSO, 400 MHz) δ : 8.64 (2H, s, H_{im}), 8.19 (2H, s, H_{im}), 7.51 (2H, s, H_{im}), 7.18 (8H, m, H_{bz}, *m*-bib), 7.09 (4H, d, H_{im}), 6.93 (2H, m, H_{bz}, *m*-bib), 6.03 (4H, d, H_{bz}, *p*-cymene), 5.78 (4H, d, H_{bz}, *p*-cymene), 5.20 (8H, d, CH₂), 2.66 (2H, m, CH), 1.74 (6H, s, CH₃), 1.11 (12H, d, CH₃). ESI-MS(+): calcd for [**5**-2Br]²⁺ *m/z* 553.49, found *m/z* 553.33. Elemental analysis (%): Calcd for Ru₂C₄₈H₅₆N₈Br₄·3H₂O C 43.65, H 4.73, N 8.48; found C 43.76, H 4.95, N 8.44.

Synthesis of $[Ru_2(\eta^6-p-cymene)_2(m-bib)_2I_2](I)_2(6)$

A solution of $[Ru_2(\eta^6$ -p-cymene)_2(*m*-bib)_2Cl_2](Cl)_2 (43.52 mg, 0.04 mmol) in MeOH (5 mL) was stirred at room temperature for 30 min. A solution of KI (50 equiv, 332 mg) in MeOH (15 mL) was added dropwise to the mixture and stirred for another 24 h. The solvent was removed under reduced pressure. The residue was recrystallized with CHCl₃ to give yellow crystals (23.1 mg, yield 39.6%). ¹H NMR (DMSO, 400 MHz) δ : 8.26 (2H, s, H_{im}), 8.19 (2H, s, H_{im}), 7.51 (2H, s, H_{im}), 7.18 (8H, m, H_{bz}, *m*-bib), 7.09 (4H, d, H_{im}), 6.93 (2H, m, H_{bz}, *m*-bib), 6.03 (4H, d, H_{bz}, *p*-cymene), 5.78 (4H, d, H_{bz}, *p*-cymene), 5.20 (8H, d, CH₂), 2.66 (2H, m, CH), 1.74 (6H, s, CH₃), 1.11 (12H, d, CH₃). ESI-MS(+): calcd for [6-21]²⁺ *m/z* 600.48, found *m/z* 601.17. Elemental analysis (%): Calcd for Ru₂C₄₈H₅₆N₈I₄•2H₂O C 38.67, H 4.06, N 7.52; found C 38.78, H 4.30, N 7.37.

Synthesis of [Ru₂(η6-p-cymene)₂(p-bib)₂Cl₂](Cl)₂ (7)

To a solution of $[RuCl_2(\eta^6-p-cymene)]_2$ (61.2 mg, 0.1 mmol) in MeOH (15 mL), the ligand pbib (47.8 mg, 0.2 mmol) was added. The reaction mixture was stirred at 338 K for 6 h, the solvent was then removed under reduced pressure. The residue was subsequently purified by silica gel chromatography to give yellow solid (45.7 mg, yield 41.9%). ¹H NMR (CDCl₃, 400 MHz) δ : 9.74 (4H, d, H_{im}), 7.39 (4H, d, H_m), 7.06 (8H, d, H_{bz}, *p*-bib), 6.58 (4H, s, H_{im}), 5.82 (8H, d, H_{bz}, *p*-cymene), 5.69 (4H, d, CH₂), 4.89 (4H, d, CH₂) 2.44 (2H, m, CH), 1.85 (6H, s, CH₃), 1.17 (12H, m, CH₃). ESI-MS(+): calcd for [7-2Cl]²⁺ m/z 509.03, found m/z 509.33; calcd for [7-Cl]^{*} m/z 1053.52, found m/z 1053.75. Elemental analysis (%): Calcd for Ru₂C₄₈H_{bg}N₈Cl₄+CH₂Cl₂+2H₂O C 49.38, H 5.07, N 9.40; found C 48.94, H 5.37, N 9.36.

Synthesis of $[Ru_2(\eta^6-p-cymene)_2(p-bib)_2Cl_2](CF_3SO_3)_2$ (8)

To a solution of $[RuCl_2(\eta^6-p-cymene]_2$ (61.2 mg, 0.1 mmol) in CH₂Cl₂ (15 mL), a solution of AgCF₃SO₃ (51.38 mg, 0.2 mmol) in EtOAc (5 mL) was added. The resulting mixture was stirred at room temperature for 2 h, the AgCl precipitate was then removed by filtration. The ligand *p*-bib (47.8 mg, 0.2 mmol) was added to the resulting solution. The mixture was stirred at RT for another 6 h, and then the solvent was removed on a rotary evaporator. The residue was recrystallized from CHCl₃ to give yellow crystals (43.0 mg, yield 32.6%). ¹H NMR (CDCl₃, 400 MHz) δ : 8.63 (4H, d, H_m), 7.37 (4H, d, H_m), 7.04 (8H, d, H_{b2}, *p*-bib), 6.71 (4H, s, Hi_m), 5.77 (4H, d, H_{b2}, *p*-cymene), 5.65 (4H, d, CH₂), 5.54 (4H, d, CH₂), 4.94 (4H, d, CH₂), 2.49 (2H, m, CH), 1.84 (6H, s, CH₃), 1.13 (12H, m, CH₃). ESI-MS(+): calcd for [8-CF₃SO₃]²* *m/z* 509.03, found *m/z* 509.33; calcd for [8-CF₃SO₃]⁴ *m/z* 1167.14, found *m/z* 1166.00. Elemental analysis (%): Calcd for Ru₂C₅₀H₅₆N₆Cl₂F₆S₂O₆ C 45.63; H 4.29, N 8.51; found C 45.23, H 4.26, N 8.24.

$[Ru_2(\eta^6-p-cymene)_2(p-bib)_2Cl_2](SbF_6)_2$ (9)

To a solution of $[Ru_2(\eta^6-p\text{-cymene})_2(p\text{-bib})_2Cl_2](Cl)_2$ (21.76 mg, 0.02 mmol) in MeOH (5 mL), a solution of AgSbF₆ (13.74 mg, 0.04 mmol) in MeOH (5 mL) was added. The mixture was stirred at room temperature for 2 h, and then the solvent was removed on a rotary evaporator. The residue was recrystallized from CHCl₃ to give yellow solid (12.1 mg, yield 40.6%). ¹H NMR (CDCl₃, 400 MHz) 5: 7.97 (4H, d, H_m), 7.30 (4H, d, H_m), 6.95 (8H, d, H_{bz}, *p*-bib), 6.84 (4H, s, Hi_m), 5.74 (4H, d, H_{bz}, *p*-cymene), 5.54 (4H, d, H_{bz}, *p*-cymene), 5.31 (4H, d, CH₂), 4.97 (4H, d, CH₂), 2.57 (2H, m, CH), 1.86 (6H, s, CH₃), 1.19 (12H, m, CH₃). ESI-MS(+): calcd for [**9**-2SbF₆]^{2*} *m*/z 509.03, found *m*/z 509.25. Elemental analysis (%): Calcd for Ru₂C₄₈H₅₆N₈Cl₂Sb₂F₁₂ C 38.70, H 3.79, N 7.52; found: C 38.45, H 3.67, N 7.42.

Synthesis of $[Ru_2(\eta^6-p-cymene)_2(p-bib)_2Cl_2](NO_3)_2$ (10)

To a solution of $[Ru_2(\eta^6-p-cymene)_2(p-bib)_2Cl_2](Cl)_2$ (21.76 mg, 0.02 mmol) in MeOH (5 mL), a solution of AgNO₃ (6.79 mg, 0.04 mmol) in MeOH (5 mL) was added. The mixture was stirred at room temperature for 2 h, and then the solvent was removed on a rotary evaporator. The residue was purified by flash column chromatography with CH₂Cl₂/CH₃OH as elution mixture to afford yellow solid (10.6 mg, yield 46.4%). ¹H NMR (CDCl₃, 400 MHz) δ : 8.83 (4H, d, H_{im}), 7.34 (4H, d, H_{im}), 6.98 (8H, d, H_{bz}, *p*-bib), 6.68 (4H, d, H_{im}), 5.76 (4H, d, H_{bz}, *p*-cymene), 5.64 (4H, d, H_{bz}, *p*-cymene), 5.48 (4H, d, CH₂), 4.90 (4H, d, CH₂) 2.47 (2H, m, CH), 1.81 (6H, s, CH₃), 1.16 (12H, m, CH₃). ESI-MS(+): calcd for [10-2NO₃]²⁺ *m/z* 510.25, found *m/z* 509.03. Elemental analysis (%): Calcd for Ru₂C₄₈H₅₆N₁₀Cl₂O₆•CH₂Cl₂ C 47.97, H 4.76, N 11.42; found C 47.78, H 4.90, N 11.78.

Synthesis of $[Ru_2(\eta^6-p-cymene)_2(p-bib)_2Br_2](Br)_2(11)$

A solution of $[Ru_2(\eta^6-p\text{-}cymene)_2(p\text{-}bib)_2\text{Cl}_2](\text{Cl})_2$ (43.52 mg, 0.04 mmol) in MeOH (5 mL) was stirred at room temperature for 30 min, a solution of KBr (50 equiv, 238 mg) in MeOH (15 mL) was added dropwise, and the mixture was stirred for another 24 h. The solvent was removed under reduced pressure. The residue was purified by flash column chromatography to give yellow solid (21.6 mg, 42.7%). ¹H NMR (DMSO, 400 MHz) δ : 8.03 (4H, d, H_{im}), 7.34 (4H, d, H_{im}), 7.14 (4H, d, Him), 7.06 (8H, d, H_{bz}, *p*-bib), 5.96 (4H, d, H_{bz}, *p*-cymene), 5.71 (4H, d, H_{bz}, *p*-cymene), 5.22 (8H, d, CH₂), 2.64 (2H, m, CH), 1.80 (6H, s, CH₃), 1.11 (12H, m, CH₃). ESI-MS(+): calcd for [11-2Br]²⁺ m/z 553.49, found m/z 553.75. Elemental analysis (%): Calcd for Ru₂C₄₈H₅₆N₈Br₄•3H₂O C 43.65, H 4.73, N 8.48; found C 43.55, H 4.74, N 8.35.

Synthesis of $[Ru_2(\eta^6-p-cymene)_2(p-bib)_2l_2](l)_2(12)$

A solution of $[Ru_2(\eta^6-p\text{-}cymene)_2(p\text{-}bib)_2Cl_2](Cl)_2$ (43.52 mg, 0.04 mmol) in MeOH (5 mL) was stirred at room temperature for 30 min, a solution of KI (50 equiv, 332 mg) in MeOH (15 mL) was added dropwise, and the mixture was stirred for another 24 h. The solvent was removed under reduced pressure. The residue was purified by flash column chromatography to give yellow solid (23.4 mg, 40.2%). ¹H NMR (DMSO, 400 MHz) δ : 8.03 (4H, d, H_{im}), 7.32 (4H, d, H_{im}), 7.18 (4H, d, Him), 7.08 (8H, d, H_{bz}, *p*-bib), 6.09 (4H, d, H_{bz}, *p*-cymene), 5.75 (4H, d, H_{bz}, *p*-cymene), 5.20 (8H, d, CH₂), 2.71 (2H, m, CH), 1.72 (6H, s, CH₃), 1.12 (12H, m, CH₃). ESI-MS(+): calcd for [12-2I]²⁺ m/z 600.48, found *m*/z 601.25. Elemental analysis (%): Calcd for Ru₂C₄₈H₅₆N₈I₄•2H₂O C 38.67, H 4.06, N 7.52; found C 38.67, H 4.32, N 7.34.

Determination of X-ray Crystal Structure

Single-crystal X-ray diffraction data were measured at 296(2) K for complexes **2**, **3**, **6** and at 298 K for complex **8** on a Bruker Apex II CCD using Mo K α radiation (λ = 0.71073 Å). An empirical absorption correction was applied to the data by using the SADABS program. The structures were solved by direct methods and refined by full-matrix least-squares methods with SHELX.^[32] All non-hydrogen atoms were refined anisotropically and hydrogen atoms of organic molecule were placed in calculated positions, assigned isotropic thermal parameters, and allowed to ride their parent atoms. The following computer programs and hardware were used: structure solution, SHELXS-2014/7; structure refinement, SHELXL-2014/7. Data collection parameters and structure refinement details are given in Table S1 in the supporting information. CCDC 1502211 (for **2**), 1502212 (for **3**), 1502213 (for **6**), 1502214 (for **8**) containing the supplementary crystallographic data for this paper that can be obtained free of charge from The Cambridge Crystallographic Data Centre.

Gel Electrophoresis Experiments

The closed circular supercoiled pBR322 plasmid DNA was used for the gel electrophoresis experiments, which was purchased from Sangon Biotech (Shanghai, China). DNA (1 μ L, 10 μ M) was treated with 4 μ L ultrapure water, 5 μ L PBS buffer (100 mM) and 10 μ L different concentrations of complexes (molar ratio of [M]/[DNA] r_i = 1, 2, 3, 4, 5, 6). The mixture was incubated at 310 K for 12 h in the dark, then 4 μ L loading buffer (0.05% Bromophenol Blue, 30 mM EDTA, 36% glycerol, 0.05% Xylene Cyanol FF) was added. Electrophoresis was carried out in a native agarose gel (1%) in 1×TAE buffer for 2 h at 70 mA. The gel was subsequently stained with 1 μ g/mL ethidium bromide and visualized under a UV transilluminator and photographed using a digital camera.

FULL PAPER

Acknowledgements

We thank the Key International (Regional) Joint Research Program of NSFC (Grant No. 21420102002), NSFC (Project 21171095, 21302094,21401105), the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD), the "Summit of the Six Top Talents" Program of Jiangsu Province China, ERC (Grant 247450), EPSRC (Grant EP/F034210/1), Science City (ERDF/AWM) for their support, our collaborators for their contributions, and members of EC COST Action CM1105 for stimulating discussions.

Keywords: Organometallic • Coordination complexes • Ruthenium arene • Metallamacrocycles • DNA • anticancer

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