A Concise Route Towards Isoflavans

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Dedication ((optional))

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Abstract: Isoflavans have gained considerable interest owing to their potential health benefits. Herein, we have presented a straightforward strategy for isoflavans synthesis. The strategy features an intermolecular [Cu]-catalyzed arylation of malonates and an intramolecular [Pd]-catalyzed Buchwald coupling of hydroxy tethered bromoarenes as the key transformations. This protocol enabled the synthesis of a variety of isoflavan analogs.

Introduction

Through the work of a couple of decades, chemists have been able to develop broader transition-metal catalysts based applications that catalyze cross-coupling reactions.¹ Consequently, the implementation of these reactions in synthetic brganic chemistry has led to successful results.² However, there have been concerns regarding target specific cross-coupling eactions, primarily involving C-heteroatom bond forming techniques.³ Challenges in these sorts of transformations would pe due to particular side reactions, particularly, dehalogenation.⁴ In this context, isoflavans have gained much popularity with the synthetic chemists. Isoflavans are oxygen-containing fused bicyclic heterocycles featuring biological properties similar to avones⁵ and are C15 frameworks contemplated as derivatives of 3-phenylchroman.⁶ Also, Isoflavans widely spread and constitute isoflavonoid natural products.7 These compounds showcase prominent biological and pharmacological significance.⁸ Interestingly, the first isoflavan was an accidental liscovery, and it was named Equol. For the first time, Equol isolated during an attempt to isolate estrogen from equine urine.⁹ After that, it was extracted from the urine of other animals and humans, as well.¹⁰ Equol widely used as a dietary phytoestrogen o decrease the prostate weight of rats.¹¹ Apart from this, the compounds belonging to this family have also reportedly seen to possess anti-fungal, anticancer, and antioxidant properties.¹² Also, it revealed that these compounds help prevent osteoporosis, androgen, and inflammation and even slow down the process of ageing.¹³ They also assist in brain mitochondrial activities and prohibit prostrate growth. The various other isoflavans isolated are sativan, vestitol, colutelol, lespedezol G1; lespecyrtin D1, etc.¹⁴

Because of their unique structure and significant biological activities, isoflavans have gained much recognition in synthetic chemistry. Various research groups have put in many efforts to develop numerous chemical routes to synthesize the isoflavan/chroman frameworks.¹⁵ Most synthetic strategies utilized the process of catalytic hydrogenation of isoflavenes, under the action of a palladium catalyst in different solvents and at pH conditions.¹⁶ A few of the approaches have mentioned. The synthesis of Equol achieved from the reaction of the racemic forms of formononetin and daidzein.¹⁷ An enantioselective route to dimethoxy counterpart of (S)-Equol was established by Ferreira and co-workers.¹⁸ Subsequently, Heemstra et al. demonstrated a total synthesis of (S)-Equol by employing enantioselective Evans' alkylation and intramolecular Buchwald-Hartwig etherification, as key steps.¹⁹ Yang et al. shown enantioselective hydrogenation reaction of a-arylcinnamic acids under iridium-catalysis.²⁰ They have extended a similar strategy for the production of (S)-Equol.²¹ Recently, the research group of Jingzhao Xia revealed asymmetric hydrogenation of 2H-chromenes via a catalytic action of Ir/In-BiphPHOX for the synthesis of (S)-Equol with >95% ee.²² Gharpure et al. in his work employed hetero-Diels-Alder reaction between olefin and ortho-quinone methides.23

With this background, herein, we wish to disclose a new protocol towards the synthesis of isoflavans. Carrying forward our work on [TM]-catalysis,²⁴ we have recently reported effective synthetic strategies for flavans.²⁵ In continuation of our interests, we have demonstrated a protocol affording neoflavans.²⁶ Herein; we disclose a new strategy delivering functionalized isoflavans. The synthetic route shows two key transformations *via* transition-metal catalyzed reactions using [Pd] and [Cu] metal catalysts.

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Results and Discussion

From the retrosynthetic analysis, it was conceived that isoflavans **8/9** could be accomplished utilizing an intramolecular [Pd]-catalyzed Buchwald–Hartwig coupling of hydroxy bromoarenes **6/7** (Figure 1). Required primary and tertiary alcohols **6/7** could be synthesized from aryl malonate esters **2** and *ortho*-bromobenzyl bromides **3**. A suitable synthetic path in achieving 6/7 would be benzylation, Krapcho decarboxylation, and reduction/nucleophilic addition.



Figure 1: Retrosynthetic Analysis of Isoflavan 8/9.

The synthetic route to achieve isoflavans was initiated with any malonate esters 2 and benzyl bromides 3. Thus, we attempted the synthessis of aryl malonate esters 2 from iodoarenes 1 and dimethyl malonate. The reaction was carried out using Cul (5 mol%), picolinic acid (10 mol%), Cs₂CO₃ (1.5 equiv), in 1,4-dioxane as a solvent for 24 h at 100 °C. Consequently, aryl malonate esters 2 were isolated in high yields (Table 1).²⁷ Benzylation reaction of aryl malonate esters 2 was performed using ortho-bromobenzyl bromides 3 at ambient temperature, K₂CO₃, and in dry DMF. As a result, excellent yields of the tethered ortho-Bromo-bis-aryl malonate esters 4 were obtained (Table 1). Next, the Krapcho decarboxylation reaction was applied to the above-furnished compounds 4 using NaCl in DMSO and water mixture (10:1) at 160 °C for 20 h. So, ortho-Bromo-bis-aryl monoesters 5 were isolated in acceptable yields, tabulated Table as in 1.





^aIsolated yields of chromatographically pure products 2a-2c, 4aa-4ca and 5aa-5ca.

After achieving aryl benzyl esters **5**, reduction of the ester group was employed to furnish the corresponding alcohol, which is a synthetic precursor for the metal-mediated cyclizations. Consequently, the reduction reaction of the esters **5**

mediated by LiAlH₄, afforded the corresponding primary alcohols **6a-6c** in nearly quantitative yields without affecting the bromosubstitution (Table 2). Furthermore, the reduction of the ester by a Grignard reagent furnished the corresponding tertiary alcohols **7a-7h** in very good to excellent yields (Table 2).

Table 2: Scope for the formation of alcohols 6a-6c/7a-7h.^{a,b,c,d}



^aIsolated yields of chromatographically pure products **6a-6c** and **7a-7h**. Tertiary alcohols **7a-7h** were prepared from bromo-esters **5aa-5ca** using alkyl/aryl Grignard reaction. ^cFor Lithium Aluminium Hydride (LAH) reduction and methyl Grignard reaction, dry diethyl ether (Et₂O) was used as solvent. ^dFor ethyl/propyl/phenyl Grignard reaction dry tetrahydrofuran (THF) was used as a solvent.

After obtaining bromo alcohols **6a-6c/7a-7h**, we went ahead with the key transformation step. It was an intramolecular reaction reading to the construction of a C-O bond facilitated by a transition metal catalyst to generate the corresponding isoflavan. In this context, initially, we tested with our previously established conditions in the synthesis of flavans and neo-flavans [i.e. Cul (20 mol%)/2,2-bipyridyl (20 mol%), base KO^tBu (3 equiv.) in dry DMF (120 °C) for 24 h].²⁵ However, unfortunately, these conditions were not favorable in delivering the expected isoflavan 8a (entries 1, Table 3). At this stage, we reckoned that the use of excess base (KO^tBu) might have hampered the reaction. Thus we thought less amount of base (KO^tBu) would be more effective. Hence, the reaction was conducted using 2 and 1 equiv of KO^tBu (entries 2 & 3, Table 3). However, the attempts were futile in obtaining the isoflavan product 8a. Since copper alone was not an optimum and general catalyst for the C-O bond formation, we turned our focus towards utilizing [Pd]catalyst, which might also serve as more effective. Interestingly, the commencing attempt alone showed effective results [Pd(OAc)₂ (10 mol%), JohnPhos (10 mol%), Cs₂CO₃ (2.0 equiv), in dry toluene, at 90 °C for 24 h (entry 4, Table 3)].²⁸ The final product was predominantly the cyclized compound isoflavan 8a, which isolated in a good yield of 80%. Additionally, to find the most efficient operational conditions, the conditions were modified by altering the temperature (entries 5 & 6, Table 3). The attempt has not found to be effective in increasing the yield. Moving ahead, we increased the base to 3 equiv and observed the undesired products at the expense of the desired product 8a (entry 7, Table 3). While running the reaction in solvent DMF at 90 °C for 14 h, we afforded isoflavan 8a in 60% yield (entry 8, Table 3). Ultimately, by increasing the catalyst loading and decreasing the reaction times to 12 h, we managed to improve the yield to 85% (entry 9, Table 3). Delightfully, the same results obtained by using 3 mol% of a palladium catalyst and 5 mol% of ligand (entry 10, Table 3).

		OH Br	catalyst (mol%) ligand (mol%) base(equiv) solvent, Temp (°C) time (h)				
		6a	unic (ii)	8a			
Entry	Catalyst (mol%)	Ligand (mol%)	Base (equiv)	Solvent (mL)	Temp (°C)	Time (h)	% of Yield (8a) ^b
1	Cul (20)	2,2'-bipyridyl (20)	KO [/] Bu (3)	DMF	120	24	_ ^c
2	Cul (20)	2,2'-bipyridyl (20)	KO ^t Bu (2)	DMF	120	24	SM
3	Cul (20)	2,2'-bipyridyl (20)	KO ^t Bu (1)	DMF	120	24	SM
4	Pd(OAc) ₂ (10)	JohnPhos (20)	Cs ₂ CO ₃ (2)	Toluene	90	24	79
5	Pd(OAc) ₂ (10)	JohnPhos (20)	Cs ₂ CO ₃ (2)	Toluene	110	12	80
6	Pd(OAc) ₂ (10)	JohnPhos (20)	Cs ₂ CO ₃ (2)	toluene	70	20	73
7	Pd(OAc) ₂ (10)	JohnPhos (10)	$Cs_2CO_3(3)$	toluene	90	12	64
8	Pd(OAc) ₂ (10)	JohnPhos (20)	Cs ₂ CO ₃ (2)	DMF	90	14	60
9	Pd(OAc) ₂ (20)	JohnPhos (20)	Cs ₂ CO ₃ (2)	toluene	90	12	85
10	Pd(OAc) ₂ (3)	JohnPhos (5)	Cs ₂ CO ₃ (2)	toluene	90	12	85

^aUnless then stated, all reactions were conducted by using 58.0 mg (0.20 mmol) of **6a**, in 2 mL dry solvent. ^bYields of isolated product **8a**. ^bNo significant spot was observed on TLC.

Moving forward, we examined the generality of the scheme by implementing it on other substrates and applying the above optimal conditions (entry 10, Table 3). The various precursor alcohols **6a-6c** subjected to the C–O bond formation intramolecularly. To our delight, the experimental conditions verified to be practical in furnishing cyclized isoflavans **8a-8c** efficiently (Table 4). Furthermore, to test the scope and imitations of the strategy, we employed the same protocol for the production of tertiary alcohols. Thus, concluding the scheme, we investigated the critical step constructing C–O bond intramolecularly from bromo tertiary alcohols **7a-7h** catalyzed using palladium. As expected, the synthetic strategy worked well bn tertiary alcohols **7a-7h** and afforded functionalized isoflavans **9a-9h** in good to excellent yields (Table 4).

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Table 4: Scope for the formation of isoflavans 8a-8c/9a-9h.^{a,b}



^aChromatographically isolated yields of pure compounds **8a-8c & 9a-9h**. ^bReaction carried out on alcohols **6a-7h** (0.20 mmol) in dry toluene (2 mL).

Conclusion

In conclusion, a straightforward synthetic route about the synthesis of functionalized isoflavans devised. An intermolecular C–C and intramolecular C–O bonds formation were facilitated by [Cu] and [Pd], as the key transformations of the strategy, respectively. This method is practical and flexible in enabling the synthesis of different isoflavan derivatives.

Experimental Section

General procedure:

A Bruker Tensor 37 (FT-IR) spectrophotometer was used to record the IR spectra. Bruker Avance 400 (400 MHz) pectrometer was used to record ¹H-NMR spectra at 295 K in CDCl₃; chemical shifts (δ in ppm) and coupling constants (J in Hz) are reported in standard manner with reference to either internal standard tetramethylsilane (TMS) ($\delta H = 0.00$ ppm) or CHCl₃ (δ H = 7.25 ppm). Bruker Avance 400 (100 MHz) spectrometer was used to record ¹³C-NMR spectra at RT in CDCl₃; chemical shifts (δ in ppm) are reported relative to CDCl₃ $\delta C = 77.00$ ppm (central line of triplet)]. The various initials notations used in the ¹H-NMR are: s = singlet, d = doublet, t = 1triplet, q = quartet, m = multiplet. ¹H, ¹³C CPD and DEPT spectra vere useful to validate the assignment of signals. For accurate mass measurements, Agilent 6538 UHD Q-TOF was used to ecord High-resolution mass spectra (HR-MS) through the multimode source. TLC monitored the progress of reactions on silica gel coated on alumina plate or glass plate. The eluents mployed were a mixture of petroleum ether and ethyl acetate. Reactions were conducted under nitrogen atmosphere in order o maintain the inert condition.

Materials: Solvents were distilled/dried before use. Petroleum ether, dichloromethane (DCM), ethyl acetate and dry DMSO were employed as solvents. Petroleum ether has a boiling range of 60 to 80 °C and dry DMSO has a boiling range of 160 to 170 °C with purity 99%. They were bought from Sigma Aldrich as well as local commercial sources. For chromatography, Acme's silica gel (100-200 mesh) was used.

General Procedure-1 for the synthesis of aryl malonate esters (2a-2c): To an oven-dried round-bottomed flask charged with a Teflon-coated magnetic stirrer bar, were added aryl iodides 1 (5.0 mmol), Cul (5.0 mol%), 2-picolinic acid (10.0 mol%), Cs₂CO₃ (10.0 mmol), and distilled diethyl malonate (7.5 mmol) and purged with nitrogen gas (3 cycles). Anhydrous 1,4dioxane (10 mL) was added to the reaction mixture heated in a preheated oil bath at 80 °C. After the designated time, the completed reaction (judged by TLC analysis) was allowed to reach room temperature. The mixture was separated using ethyl acetate (3 × 20 mL) and saturated aqueous NH₄Cl (40 mL). The separated organic layers were dried using Na₂SO₄, filtered and concentrated by rotary evaporator under reduced pressure. Purification of the crude mixture using silica gel column chromatography, furnished α -aryl malonate products **2a-2c** (89-92%).

General Procedure-2 for the synthesis of alkylated malonate esters (4aa-4ca): Potassium carbonate (1.14 gm, 8.18 mmol, 3.0 equiv) was suspended in dry DMF (10 mL) at room temperature. A solution containing aryl malonates 2 (2.72 mmol, 1.0 equiv) in 5 mL dry DMF was added to it dropwise. After 30 min stirring at room temperature, a solution of orthobromobenzyl bromides 3 (3.27 mmol, 1.2 equiv) in dry DMF (5 mL) was added, and stirring was continued for further 12-15 h. The reaction mixture was quenched with saturated aqueous NH₄Cl solution (40 mL), and extracted with EtOAc (3 × 20 mL). Then the organic layers were washed with brine solution (30 mL) and dried using anhydrous Na₂SO₄, filtered and the organic layer was concentrated in rotary evaporator under reduced pressure. Silica gel column chromatography of the residue, generated the products, ortho-bromobenzyl malonates 4aa-4ca (87-95%).

General Procedure-3 for the synthesis of aryl mono esters (5aa-5ca): A mixture of the alkylated malonate esters 4 (1.53 mmol), sodium chloride (178.0 mg, 3.07 mmol, 2.0 equiv) in DMSO:H₂O (10:1) (12 mL) was placed in a sealed tube. The reaction mixture was heated at a temperature of 160 °C for 18-20 h. The mixture was cooled to room temperature and poured into EtOAc (60 mL), washed using brine solution (2 × 30 mL), dried by Na₂SO₄ and concentrated using rotary evaporator under reduced pressure. The crude reaction mixture was purified by silica gel column chromatography to afford the products (5aa-5ca) as an oil (69-75%).

General Procedure-4 for reduction of malonate esters (5aa-5ca): Under N₂ atmosphere, in a dry round-bottomed flask bromo malonate esters 5 (0.50 mmol) in dry Et₂O (25 mL) was taken, to which LiAlH₄ (1.50 mmol) was slowly added in portion wise at 0 °C. Then the reaction mixture was allowed to stirrer for 2 h at 0 °C. Then quenched with ethyl acetate (30 mL), and the resultant mixture was separated using ethyl acetate (2 × 30 mL) and brine solution (60 mL). The separated organic layers were dried by Na₂SO₄ and concentrated using rotary evaporator under reduced pressure. Then the reaction mixture was purified by silica gel column chromatography to furnish the primary alcohols **6a-6c** (93-97%). General Procedure-5 for the synthesis of tertiary alcohol (7a-7h): In a round-bottomed flask magnetically stirred esters 5 (0.50 mmol) in 5 mL of dry ether/THF was taken, to which methylmagnesium iodide (2.5 mmol) or ethyl/ propyl/ phenyl magnesium bromide (2.5 mmol) [prepared from magnesium (2.5 mmol) and methyl iodide (4.00 mmol) in dry ether or ethyl/propyl/phenyl bromide (4.00 mmol) in dry THF and a catalytic amount of iodine] was added at -10 °C. The reaction mixture was stirred for 3-5 h -10 °C to rt. Then quenched by aqueous NH₄Cl (50 mL) and extracted using ethyl acetate (2 × 30 mL). The organic layers were dried by Na₂SO₄ and concentrated in rotary evaporator. The crude products were burified by using silica gel column chromatography to furnish the tertiary alcohols **7a-7h** (71–95%).

General Procedure-6 for the Synthesis of Isoflavans (8a-8c/9a-9h): Alcohol 6 or 7 (0.20 mmol), $Pd(OAc)_2$ (3 mol%), JohnPhos (5 mol%), Cs_2CO_3 (0.40 mmol) were added to a dried Schlenk tube under N₂ atmosphere. Then 2 mL dry toluene was added to the mixture and stirred at 90 °C for 12-14 h. After completion of the reaction, the reaction mixture was quenched by aq. NH₄Cl (30 mL) and extracted with ethyl acetate (3 × 20 mL). The organic layers were dried by Na₂SO₄ and concentrated in rotary evaporator under reduced pressure. The purification of crude mixture using silica gel column chromatography, furnished the isoflavan products 8a-8c/9a-9h (79-94%).

Dimethyl 2-(2-bromobenzyl)-2-phenylmalonate (4aa): GP-2 followed with potassium carbonate (1.14 gm, 8.18 mmol, 3.0 equiv) in dry DMF (10 mL), aryl malonate **2a** (566 mg, 2.72 mmol, 1.0 equiv), *ortho*-bromobenzyl bromides **3a** (814 mg, 3.27 mmol, 1.2 equiv) in 5 mL dry DMF (5 mL), and stirred for 12 h. The column chromatography purification (PE/EA, 95:05 to 90:10), furnished the isolflavan **4aa** (975 mg, 95%) as colourless viscus liquid. [TLC monitor (PE/EA 95:05), R_t (**2a**)= 0.30, R_t (**3a**) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.44 (dd, J = 7.9, 1.4 Hz, 1H), 7.29 – 7.19 (m, 5H), 7.09 (td, J = 7.5, 1.4 Hz, 1H), 7.01 (ddd, J = 16.9, 7.6, 1.8 Hz, 2H), 3.87 (s, 2H), 3.76 (s, 6H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 170.5, 136.0, 135.8, 132.6, 131.0, 128.3, 128.3, 128.0, 127.6, 127.0, 126.6, 63.4, 52.9, 41.2 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₈H₁₈BrO₄⁺: 377.0383, found: 377.0378.

Dimethyl2-(2-bromo-5-methoxybenzyl)-2-phenylmalonate

(4ab): GP-2 was followed with potassium carbonate (1.14 gm, 8.18 mmol, 3.0 equiv) in dry DMF (10 mL), aryl malonate 2a (566 mg, 2.72 mmol, 1.0 equiv), *ortho*-bromobenzyl bromides 3b (915 mg, 3.27 mmol, 1.2 equiv) in 5 mL dry DMF, and stirred for 15 h. The column chromatography purification (PE/EA, 90:10 to

85:15), furnished **4ab** (964 mg, 87%) as colourless viscus liquid. [TLC monitor (PE/EA 90:10), $R_{\rm f}(2a) = 0.30$, $R_{\rm f}(3b) = 0.70$, UV detection]. ¹H NMR (400 MHz, CDCl₃) $\delta = 7.34 - 7.21$ (m, 6H), 6.60 (dd, J = 8.8, 3.1 Hz, 1H), 6.48 (d, J = 3.1 Hz, 1H), 3.82 (s, 2H), 3.77 (s, 6H), 3.57 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) $\delta = 170.4$, 158.2, 136.6, 136.1, 132.9, 128.3, 128.0, 127.6, 117.0, 116.2, 114.7, 63.4, 55.1, 52.9, 41.5 ppm. HRMS (ESI) calculated [M+Na]⁺ for C₁₉H₁₉BrNaO₅⁺: 429.0308, found: 429.0305.

Dimethyl 2-(2-bromobenzyl)-2-(*m***-tolyl)malonate (4ba): GP-2** was followed with potassium carbonate (1.14 gm, 8.18 mmol, 3.0 equiv) in dry DMF (10 mL), aryl malonate **2b** (604 mg, 2.72 mmol, 1.0 equiv) was added with *ortho*-bromobenzyl bromides **3a** (814 mg, 3.27 mmol, 1.2 equiv) in 5 mL dry DMF, and stirred for 14 h. The column chromatography purification (PE/EA, 95:05 to 90:10), furnished **4ba** (968 mg, 91%) as colourless viscus liquid. [TLC monitor (PE/EA 95:05), R_t (**2b**) = 0.30, R_t (**3a**) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.45 (dd, J = 7.8, 1.3 Hz, 1H), 7.15 – 7.11 (m, 2H), 7.11 – 7.04 (m, 3H), 7.00 (ddd, J = 14.3, 7.6, 1.8 Hz, 2H), 3.85 (s, 2H), 3.73 (s, 6H), 2.32 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 170.6, 137.5, 136.0, 135.9, 132.6, 131.0, 129.0, 128.4, 128.2, 127.9, 126.9, 126.6, 125.2, 63.4, 52.9, 41.2, 21.6 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₉H₂₀BrO₄⁺: 391.0539, found: 391.0535

Dimethyl 2-(2-bromobenzyl)-2-(*p***-tolyl)malonate (4ca): GP-2** was followed with potassium carbonate (1.14 gm, 8.18 mmol, 3.0 equiv) in dry DMF (10 mL), aryl malonate **2c** (604 mg, 2.72 mmol, 1.0 equiv) was added with *ortho*-bromobenzyl bromides **3a** (814 mg, 3.27 mmol, 1.2 equiv) in 5 mL dry DMF, and stirred for 14 h. The column chromatography purification (PE/EA, 95:05 to 90:10), furnished **4ca** (947 mg, 89%) as colourless viscus liquid. [TLC monitor (PE/EA 95:05), R_t (**2c**) = 0.30, R_t (**3a**) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.45 (dd, J = 7.8, 1.3 Hz, 1H), 7.16 – 7.11 (m, 2H), 7.11 – 7.05 (m, 3H), 7.01 (ddd, J = 13.1, 7.5, 1.8 Hz, 2H), 3.86 (s, 2H), 3.73 (s, 6H), 2.32 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 170.6, 137.4, 136.0, 133.1, 132.6, 131.0, 128.7, 128.2, 128.1, 126.9, 126.5, 63.0, 52.8, 41.0, 21.0 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₉H₂₀BrO₄⁺: 391.0539, found: 391.0533

Methyl 3-(2-bromophenyl)-2-phenylpropanoate (5aa): GP-3 was followed with alkylated malonate esters **4aa** (1.53 mmol), sodium chloride (178.0 mg, 3.07 mmol, 2.0 equiv) in DMSO:H₂O (10:1) (12 mL) in sealed tube and stirred at 160 °C for 18 h. The column chromatography purification (PE/EA, 97:03 to 95:05), afforded **5aa** (366 mg, 75%) as colourless viscus liquid. [TLC monitor (PE/EA 95:05), R_{l} (**4aa**) = 0.30, R_{l} (**5aa**) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.52 (dd, J = 7.8, 1.2

Hz, 1H), 7.34 – 7.22 (m, 5H), 7.13 – 7.01 (m, 3H), 4.04 (dd, J = 13.0, 10.3 Hz, 1H), 3.60 (s, 3H), 3.50 (dd, J = 13.6, 9.0 Hz, 1H), 3.13 (dd, J = 13.6, 6.3 Hz, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃) $\delta = 173.6, 138.5, 138.1, 132.8, 131.5, 128.7, 128.2, 127.8, 127.4, 127.2, 124.6, 52.0, 51.0, 40.2 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₆H₁₆Br⁷⁹O₂⁺: 319.0328, found: 319.0324 and [M+H]⁺ for C₁₆H₁₆Br⁸¹O₂⁺: 321.0308, found: 321.0305.$

Methyl-3-(2-bromo-5-methoxyphenyl)-2-phenylpropanoate

(5ab): GP-3 was followed with alkylated malonate esters 4ab 11.53 mmol), sodium chloride (178.0 mg, 3.07 mmol, 2.0 equiv) in DMSO:H₂O (10:1) (12 mL) in sealed tube and stirred at 160 IC for 20 h. The column chromatography purification (PE/EA, 97:03 to 93:07), afforded 5ab (379 mg, 71%) as colourless viscus liquid. [TLC monitor (PE/EA 93:07), R_i (4ab) = 0.30, R_i (5ab) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.39 (d, J = 8.5 Hz, 1H), 7.34 – 7.22 (m, 5H), 6.64 – 6.58 (m, 2H), 4.00 (dd, J = 8.7, 6.5 Hz, 1H), 3.65 (s, 3H), 3.62 (s, 3H), 3.47 (dd, J = 13.6, 8.8 Hz, 1H), 3.08 (dd, J = 13.6, 6.5 Hz, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 173.5, 158.5, 139.0, 138.4, 133.2, 128.7, 127.8, 127.4, 116.7, 115.0, 114.3, 55.3, 52.0, 51.0, 40.3 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₇H₁₈Br⁸¹O₃⁺: 349.0434, found: 349.0421 and [M+H]⁺ for C₁₇H₁₈Br⁸¹O₃⁺: 351.0413, found: 351.0408.

Methyl 3-(2-bromophenyl)-2-(m-tolyl)propanoate (5ba): GP-3 vas followed with alkylated malonate esters 4ba (1.53 mmol), sodium chloride (178.0 mg, 3.07 mmol, 2.0 equiv) in DMSO:H₂O (10:1) (12 mL) in sealed tube and stirred at 160 °C for 19 h. The column chromatography purification (PE/EA, 97:03 to 95:05), andrded 5ba (403 mg, 79%) as colourless viscus liquid. [TLC monitor (PE/EA 95:05), R(4ba) = 0.30, R(5ba) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.53 (dd, J = 7.9, 1.0 Hz, 1H), 7.20 (t, J = 7.5 Hz, 1H), 7.17 – 7.02 (m, 6H), 4.00 (dd, J = 9.2, 5.9 Hz, 1H), 3.60 (s, 3H), 3.49 (dd, J = 13.6, 9.2 Hz, 1H), B.12 (dd, J = 13.6, 5.9 Hz, 1H), 2.33 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 173.6, 138.4, 138.3, 138.3, 132.8, 131.5, 128.5, 128.4, 128.2, 128.2, 127.2, 124.8, 124.7, 52.0, 51.0, 40.1, 21.4 ppm. HRMS (ESI) calculated $[M+H]^+$ for $C_{17}H_{18}Br^{79}O_2^+$: 333.0485, found: 333.0484 and [M+H]⁺ for C₁₇H₁₈Br⁸¹O₂⁺: 335.0464, found: 335.0463.

Methyl 3-(2-bromophenyl)-2-(*p*-tolyl)propanoate (5ca): GP-3 was followed with alkylated malonate esters 4ca (1.53 mmol), sodium chloride (178.0 mg, 3.07 mmol, 2.0 equiv) in DMSO:H₂O (10:1) (12 mL) in sealed tube and stirred at 160 °C for 18 h. The column chromatography purification (PE/EA, 97:03 to 95:05), afforded 5ca (352 mg, 69%) as colourless viscus liquid. [TLC monitor (PE/EA 95:05), R_t (4ca) = 0.30, R_t (5ca) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.53 (dd, *J* = 7.9, 1.1, 1H), 7.22 (d, J = 8.1, 2H), 7.18 – 7.08 (m, 4H), 7.05 (ddd, J = 7.9, 3.0, 2.3 Hz, 1H), 4.01 (dd, J = 13.0, 6.2 Hz, 1H), 3.60 (s, 3H), 3.50 (dd, J = 13.6, 9.0 Hz, 1H), 3.12 (dd, J = 13.7, 6.2 Hz, 1H), 2.33 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) $\delta = 173.7$, 138.3, 137.1, 135.5, 132.8, 131.5, 129.3, 128.1, 127.6, 127.2, 124.7, 51.9, 50.7, 40.1, 21.0 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₇H₁₈Br⁷⁹O₂⁺: 333.0485, found: 333.0482 and [M+H]⁺ for C₁₇H₁₈Br⁸¹O₂⁺: 335.0464, found: 335.0460.

3-(2-Bromophenyl)-2-phenylpropan-1-ol (6a): GP-4 was followed with bromo malonateester 5aa (0.50 mmol) in dry ether (25 mL), was added slowly LiAlH₄ (1.50 mmol) in portion wise 0 °C and the resultant reaction mixture was stirred for at 0 °C for 2 h to get compound 6a. The column chromatography purification (PE/EA, 93:07 to 90:10), gave 6a (138 mg, 95%) as colourless viscus liquid. [TLC monitor (PE/EA 95:05), R_f(5aa) = 0.70, R_f(6a) = 0.30, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.50 (d, J=8.4 Hz, 1H), 7.31 – 7.25 (m, 2H), 7.20 (t, J = 6.6 Hz, 3H), 7.10 - 7.05 (m, 1H), 6.98 (ddd, J = 14.6, 7.5, 1.8 Hz, 2H), 3.77 (dd, J = 5.9, 2.9 Hz, 2H), 3.21 - 3.14 (m, 2H), 2.93 (ddd, J = 16.5, 9.1, 4.3 Hz, 1H), 1.63 (br.s, 1H) ppm. 13 C NMR (100 MHz, CDCl₃) δ = 141.6, 139.1, 132.7, 131.3, 128.5, 128.0, 127.7, 127.0, 126.8, 124.6, 65.9, 48.1, 38.8 ppm. HRMS (ESI) calculated [(M+H)+(- $H_2O)$]⁺ for $C_{15}H_{14}Br^{79+}$: 273.0273, found: 273.0271 and $[(M+H)+(-H_2O)]^+$ for C₁₅H₁₄Br⁸¹⁺: 275.0254, found: 275.0251.

3-(2-Bromophenyl)-2-(m-tolyl)propan-1-ol (6b): GP-4 was followed with bromo malonateester 5ba (0.50 mmol) in dry ether (25 mL), was added slowly LiAlH₄ (1.50 mmol) in portion wise 0 °C and the resultant reaction mixture was stirred for at 0 °C for 2 h. The column chromatography purification (PE/EA, 93:07 to 90:10), gave 6b (148 mg, 97%) as colourless viscus liquid. [TLC monitor (PE/EA 95:05), $R_{f}(5ba) = 0.70$, $R_{f}(6b) = 0.30$, UV detection]. ¹H NMR(400 MHz, CDCI₃) δ = 7.52 (dd, J = 8.4, 1.2 Hz, 1H), 7.24 – 7.17 (m, 1H), 7.12 (td, J = 7.4, 1.3 Hz, 1H), 7.08 - 6.99 (m, 5H), 3.86 - 3.74 (m, 2H), 3.23 - 3.13 (m, 2H), 3.03 -2.92 (m, 1H), 2.33 (s, 3H), 1.39 (br.s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 141.5, 139.3, 138.2, 132.8, 131.3, 128.8, 128.5, 127.8, 127.7, 127.1, 125.0, 124.7, 66.0, 48.1, 38.8, 21.5 ppm. HRMS (ESI) calculated [M+NH₄]⁺ for C₁₆H₂₁Br⁷⁹NO⁺: 322.0801, found: 322.0799 and [M+NH₄]⁺ for C₁₆H₂₁Br⁸¹NO⁺: 324.0781, found: 324.0780.

3-(2-Bromophenyl)-2-(p-tolyl)propan-1-ol (6c): GP-4 was followed with bromo malonate ester **5ca** (0.50 mmol) in dry ether (25 mL), was added slowly LiAlH₄ (1.50 mmol) in portion wise 0 °C and the resultant reaction mixture was stirred for at 0 °C for 2 h. The column chromatography purification (PE/EA, 93:07 to 90:10), gave **6c** (142 mg, 93%) as colourless viscus liquid. [TLC control PE/EA 95:05), $R_{\rm f}$ (**5ba**) = 0.70, $R_{\rm f}$ (**6c**) = 0.30, UV

detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.52 (d, *J* = 8.0 Hz, 1H), 7.15 – 7.09 (m, 5H), 7.03 (dd, *J* = 12.1, 4.7 Hz, 2H), 3.79 (s, 2H), 3.18 (dt, *J* = 11.1, 6.2 Hz, 2H), 3.02 – 2.90 (m, 1H), 2.32 (s, 3H), 1.42 (br. s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 139.3, 138.4, 136.4, 132.8, 131.3, 129.3, 127.9, 127.7, 127.1, 124.7, 66.1, 47.7, 38.9, 21.0 ppm. HRMS (ESI) calculated [M+NH₄]⁺ for C₁₆H₂₁Br⁷⁹NO⁺: 322.0801, found: 322.0797 and [M+NH₄]⁺ for C₁₆H₂₁Br⁸¹NO⁺: 324.0782, found: 324.0777.

4-(2-Bromophenyl)-2-methyl-3-phenylbutan-2-ol (7a): GP-5 vas followed to obtain title compound 7a from ester 5aa (0.50 mmol). The column chromatography purification (PE/EA, 93:07 o 90:10), delivered 7a (152 mg, 95%) as colourless viscus liquid. [TLC monitor PE/EA 95:05), R_f(5aa) = 0.70, R_f(7a) = 0.30, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.48 – 7.37 (m, 1H), 7.24 – 7.12 (m, 5H), 6.96 – 6.84 (m, 2H), 6.78 – 6.70 (m, H), 3.46 (dd, J = 12.8, 2.2 Hz, 1H), 3.17 – 3.00 (m, 2H), 1.49 (br. s, 1H), 1.36 (s, 3H), 1.24 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 140.3, 139.9, 132.6, 131.4, 129.7, 127.9, 127.3, 126.7, 126.6, 124.5, 73.1, 56.6, 36.4, 28.4, 28.2 ppm. HRMS (ESI) calculated [(M+H)+(-H₂O)]⁺ for C₁₇H₁₈Br⁷⁹⁺: 301.0586, ound: 301.0582 and $[(M+H)+(-H_2O)]^+$ for $C_{17}H_{18}Br^{81+}$: 303.0567, found: 303.0562.

1-(2-Bromophenyl)-3-ethyl-2-phenylpentan-3-ol (7b): **GP-5** was followed to obtain title compound **7b** from ester **5aa** (0.50 nmol). The column chromatography purification (PE/EA, 93:07 to 90:10), delivered **7b** (160 mg, 92%) as colourless viscus liquid. [TLC monitor PE/EA 95:05), R_i (**5aa**) = 0.70, R_i (**7b**) = 0.30, JV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.45 - 7.36 (m, 1H), 3.40 (dd, *J* = 11.8, 1.5 Hz, 1H), 3.17 - 3.07 (m, 2H), 1.89 - 1.73 (m, 2H), 1.33 - 1.27 (m, 2H), 1.05 - 0.94 (m, 3H), 0.87 - 0.75 (m, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 140.7, 140.0, 132.4, 131.7, 129.9, 127.8, 127.2, 126.5, 126.3, 124.5, 51.6, 36.2, 29.5, 28.2, 8.2, 7.7 ppm. HRMS (ESI) calculated [(M+H)+(-H₂O)]⁺ for C₁₉H₂₂Br⁷⁹⁺: 329.0899, found: 329.0889 and [(M+H)+(-H₂O)]⁺ for C₁₉H₂₂Br⁸¹⁺: 331.0881, found: 331.0875.

3-(2-Bromophenyl)-1,1,2-triphenylpropan-1-ol (7c): **GP-5** was followed to obtain title compound **7c** from ester **5aa** (0.50 mmol). The column chromatography purification (PE/EA, 93:07 to 90:10), delivered **7c** (197 mg, 89%) as colourless viscus liquid. [TLC monitor PE/EA 95:05), R_1 (**5aa**) = 0.70, R_1 (**7c**) = 0.30, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.77 (dd, J = 8.4 and 1.0 Hz, 2H), 7.51 – 7.46 (m, 1H), 7.39 – 7.33 (m, 4H), 7.25 – 7.21 (m, 2H), 7.18 – 7.06 (m, 4H), 7.03 – 6.98 (m, 3H), 6.84 (dddd, J = 26.7, 17.2, 7.4 and 1.6 Hz, 3H), 4.32 (dd, J = 11.6 and 3.8 Hz, 1H), 3.26 (dd, J = 14.2 and 11.7 Hz, 1H), 3.15 (dd, J = 14.3 and 3.7 Hz, 1H), 2.67 (s, 1H) ppm. ¹³C NMR (100 MHz,

$$\begin{split} &\text{CDCl}_3 \ \bar{\pmb{\delta}}\ =\ 146.2,\ 145.4,\ 139.5,\ 139.1,\ 132.5,\ 131.5,\ 130.3,\\ &130.0,\ 128.2,\ 127.7,\ 127.6,\ 127.2,\ 126.8,\ 126.6,\ 126.4,\ 126.3,\\ &126.1,\ 125.7,\ 124.6,\ 80.9,\ 53.2,\ 36.7\ \text{ppm}.\ \text{HRMS}\ (\text{ESI})\\ &\text{calculated}\ [(\text{M+H})+(-\text{H}_2\text{O})]^+\ \text{for}\ \text{C}_{27}\text{H}_{22}\text{Br}^{79+}\!\!:\ 425.0899,\ \text{found}\!\!:\ 425.0896\ \text{and}\ [(\text{M+H})+(-\text{H}_2\text{O})]^+\ \text{for}\ \text{C}_{27}\text{H}_{22}\text{Br}^{81+}\!\!:\ 427.0883,\\ &\text{found}\!\!:\ 427.0879. \end{split}$$

4-(2-Bromo-5-methoxyphenyl)-2-methyl-3-phenylbutan-2-ol

(7d): GP-5 was followed to obtain title compound 7d from ester 5ab (0.50 mmol). The column chromatography purification (PE/EA, 90:10 to 85:15), delivered 7d (157 mg, 90%) as colourless viscus liquid. [TLC monitor PE/EA 90:10), $R_{\rm f}$ (5ab) = 0.70, $R_{\rm f}$ (7d) = 0.30, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.29 (d, *J* = 8.7 Hz, 1H), 7.25 – 7.13 (m, 5H), 6.47 (dd, *J* = 8.7 and 3.1 Hz, 1H), 6.23 (d, *J* = 3.1 Hz, 1H), 3.45 (s, 3H), 3.42 (d, *J* = 1.9 Hz, 1H), 3.12 – 2.87 (m, 2H), 1.46 (s, 1H), 1.36 (s, 3H), 1.24 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 158.1, 140.8, 140.4, 132.9, 129.7, 128.0, 126.7, 116.4, 114.9, 113.8, 73.2, 56.7, 55.1, 36.7, 28.5, 28.2 ppm. HRMS (ESI) calculated [(M+H)+(-H₂O)]⁺ for C₁₈H₂₀Br⁸¹O⁺: 333.0673, found: 333.0668.

4-(2-Bromophenyl)-2-methyl-3-(*m***-tolyl)butan-2-ol (7e): GP-5** was followed to obtain compound **7e** from ester **5ba** (0.50 mmol). The column chromatography purification (PE/EA, 93:07 to 90:10), delivered **7e** (150 mg, 90%) as colourless viscus liquid. [TLC monitor PE/EA 93:07), *R*_f(**5ba**) = 0.70, *R*_f(**7e**) = 0.30, UV detection]. ¹H NMR(400 MHz, CDCl₃) δ = 7.43 (dd, *J* = 7.6 and 1.7 Hz, 1H), 7.11 (t, *J* = 7.7 Hz, 1H), 7.02 – 6.86 (m, 5H), 6.77 (dd, *J* = 7.2 and 2.1 Hz, 1H), 3.44 (dd, *J* = 13.4 and 2.8 Hz, 1H), 3.15 – 3.05 (m, 1H), 3.00 (dd, *J* = 11.8 and 2.8 Hz, 1H), 2.27 (s, 3H), 1.47 (s, 1H), 1.34 (s, 3H), 1.26 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 140.1, 134.0, 137.4, 132.6, 131.3, 130.5, 127.8, 127.4, 127.3, 126.7, 126.7, 124.6, 73.2, 56.5, 36.3, 28.2, 21.5 ppm. HRMS (ESI) calculated [(M+H)+(-H₂O)]⁺ for C₁₈H₂₀Br⁷⁹⁺: 315.0743, found: 315.0738 and [(M+H)+(-H₂O)]⁺ for C₁₈H₂₀Br⁸¹⁺: 317.0724, found: 317.0719.

1-(2-Bromophenyl)-3-ethyl-2-(*m***-tolyl)pentan-3-ol (7f): GP-5** was followed to obtain title compound **7f** from ester **5ba** (0.50 mmol). The column chromatography purification (PE/EA, 93:07 to 90:10), delivered **7f** (161 mg, 89%) as colourless viscus liquid. [TLC monitor PE/EA 93:07), $R_{\rm f}$ (**5ba**) = 0.70, $R_{\rm f}$ (**7f**) = 0.30, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.41 (dd, *J* = 7.4 and 1.7 Hz, 1H), 7.18 – 6.80 (m, 7H), 6.75 – 6.66 (m, 1H), 3.47 – 3.29 (m, 1H), 3.18 – 3.07 (m, 2H), 2.26 (s, 3H), 1.81 (dt, *J* = 13.9 and 7.1 Hz, 2H), 1.31 (dd, *J* = 15.1 and 7.3 Hz, 2H), 0.98 (t, *J* = 7.5 Hz, 3H), 0.82 (t, *J* = 7.4 Hz, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 140.4, 140.1, 137.2, 132.4, 131.6, 130.6, 127.6,

127.1, 127.1, 126.9, 126.5, 124.5, 76.9, 51.5, 36.0, 29.4, 28.1, 21.5, 8.2, 7.8 ppm. HRMS (ESI) calculated $[(M+H)+(-H_2O)]^+$ for $C_{20}H_{24}Br^{79+}$: 343.1056, found: 343.1048 and $[(M+H)+(-H_2O)]^+$ for $C_{20}H_{24}Br^{81+}$: 345.1034, found: 345.1030.

4-(2-BromophenyI)-2-methyI-3-(*p***-tolyI)butan-2-ol (7g): GP-5** was followed to obtain title compound **7g** from ester **5ca** (0.50 mmol). The column chromatography purification (PE/EA, 93:07 to 90:10), delivered **7g** (152 mg, 91%) as colourless viscus liquid. [TLC monitor PE/EA 93:07), R_i (**5ca**) = 0.70, R_i (**7g**) = 0.30, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.43 (dd, J = 7.5 and 1.7 Hz, 1H), 7.05 (dd, J = 23.6 and 7.9 Hz, 4H), 6.95 – 6.87 (m, 2H), 6.78 (dd, J = 7.2 and 2.0 Hz, 1H), 3.43 (dd, J = 13.1 and 2.3 Hz, 1H), 3.16 – 2.93 (m, 2H), 2.27 (s, 3H), 1.33 (s, 3H), 1.25 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 134.0, 137.0, 136.1, 132.5, 131.3, 129.5, 128.7, 127.3, 126.7, 124.5, 73.2, 56.2, 36.3, 28.2, 28.1, 21.0 ppm. HRMS (ESI) calculated [(M+H)+(-H₂O)]⁺ for C₁₈H₂₀Br⁷⁹⁺: 315.0743, found: 315.0738 and (M+H)+(-H₂O)]⁺ for C₁₈H₂₀Br⁸¹⁺: 317.0724, found: 317.0719.

4-(2-(2-Bromophenyl)-1-(*p***-tolyl)ethyl)heptan-4-ol (7h): GP-5** was followed to obtain title compound **7h** from ester **5ca** (0.50 mmol). The column chromatography purification (PE/EA, 93:07 to 90:10), delivered **7h** (138 mg, 71%) as colourless viscus iquid. [TLC monitor PE/EA 93:07), R_t (**5ca**) = 0.70, R_t (**7h**) = 0.30, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.41 (d, *J* = 7.4 Hz, 1H), 7.10 – 7.04 (m, 2H), 6.98 (d, *J* = 7.6 Hz, 2H), 6.89 (q, *J* = 7.2 Hz, 2H), 6.72 (d, *J* = 7.1 Hz, 1H), 3.37 (d, *J* = 8.7 Hz, 1H), 3.17 – 3.03 (m, 2H), 2.26 (s, 3H), 1.83 – 1.60 (m, 2H), 1.39 – 1.18 (m, 6H), 0.96 (t, *J* = 7.2 Hz, 3H), 0.78 (t, *J* = 6.7 Hz, 3H) ppng. ¹³C NMR (100 MHz, CDCl₃) δ = 140.2, 137.3, 135.7, 132.4, 131.6, 128.6, 127.1, 126.6, 124.5, 76.6, 51.9, 39.9, 38.8, 35.9, 21.0, 17.1, 16.7, 14.7, 14.5 ppm.

β-(*m***-Tolyl)chromane (8b): GP-6** was followed with alcohol **6b** (0.20 mmol), Pd(OAc)₂ (3 mol%), JohnPhos (5 mol%), Cs₂CO₃ 0.40 mmol), 2 mL toluene(dry) was added and allowed to stirrer at 90 °C for 12 h. The column chromatography purification (PE/EA, 98:02 to 95:05), furnished **8b** (37 mg, 82%) as colourless viscus liquid. [TLC monitor PE/EA 95:05), *R*_l(**6b**) = 0.20, *R*_l(**8b**) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.28 (t, *J* = 7.5 Hz, 1H), 7.22 – 7.04 (m, 5H), 6.96 – 6.87 (m, 2H), 4.39 (ddd, *J* = 10.6, 3.6 and 2.2 Hz, 1H), 4.05 (t, *J* = 10.6 Hz, 1H), 3.34 – 3.19 (m, 1H), 3.18 – 2.93 (m, 2H), 2.40 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 154.3, 141.2, 138.4, 129.7, 128.6, 128.1, 127.8, 127.4, 124.3, 122.0, 120.3, 116.5, 70.9, 38.5, 32.4, 21.4 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₆H₁₇O⁺: 225.1274, found: 225.1272.

2,2-Dimethyl-3-phenylchromane (9a): GP-6 was carried out with alcohol 7a (0.20 mmol), Pd(OAc)₂ (3 mol%), JohnPhos (5

mol%), Cs₂CO₃ (0.40 mmol), 2 mL toluene(dry) was added and allowed to stirrer at 90 °C for 12 h. The column chromatography purification (PE/EA, 98:02 to 95:05), furnished **9a** (45 mg, 94%) as colourless viscus liquid. [TLC monitor PE/EA 95:05), $R_{\rm f}$ (**7a**) = 0.20, $R_{\rm f}$ (**9a**) = 0.70, UV detection]. ¹H NMR(400 MHz, CDCl₃) δ = 7.33 – 7.24 (m, 5H), 7.17 – 7.07 (m, 2H), 6.91 – 6.80 (m, 2H), 3.20 (dd, *J* = 16.2 and 10.5 Hz, 1H), 3.06 (dd, *J* = 10.5 and 5.3 Hz, 1H), 2.96 (dd, *J* = 16.2 and 5.3 Hz, 1H), 1.30 (s, 3H), 1.22 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 153.5, 141.5, 129.3, 128.8, 128.2, 127.4, 126.9, 121.6, 120.0, 117.2, 77.2, 47.8, 29.1, 28.1, 21.3 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₇H₁₉O⁺: 239.1430, found: 239.1426.

2,2-Diethyl-3-phenylchromane (9b): GP-6 was carried out with alcohol **7b** (0.20 mmol), Pd(OAc)₂ (3 mol%), JohnPhos (5 mol%), Cs₂CO₃ (0.40 mmol), 2 mL toluene(dry) was added and allowed to stirrer at 90 °C for 13 h. The column chromatography purification (PE/EA, 98:02 to 95:05), furnished **9b** (47 mg, 89%) as colourless viscus liquid. [TLC monitor PE/EA 95:05), $R_{\rm f}$ (**7b**) = 0.20, $R_{\rm f}$ (**9b**) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.27 - 7.10 (m, 6H), 7.08 - 7.01 (m, 1H), 6.95 - 6.78 (m, 2H), 3.25 - 3.06 (m, 2H), 2.96 (dd, *J* = 16.9 and 6.1 Hz, 1H), 1.77 - 1.67 (m, 2H), 1.58 (dq, *J* = 14.6 and 7.4 Hz, 1H), 1.38 (dq, *J* = 14.6 and 7.4 Hz, 1H), 0.92 (t, *J* = 7.4 Hz, 3H), 0.84 (t, *J* = 7.5 Hz, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 153.8, 142.5, 129.4, 128.7, 128.3, 127.3, 126.6, 121.5, 120.0, 117.3, 80.3, 43.1, 29.4, 26.6, 25.6, 7.5, 7.3 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₉H₂₃O⁺: 267.1743, found: 267.1742.

2,2,3-Triphenylchromane (9c): GP-6 was followed with alcohol **7c** (0.20 mmol), Pd(OAc)₂ (3 mol%), JohnPhos (5 mol%), Cs₂CO₃ (0.40 mmol), 2 mL toluene(dry) was added and allowed to stirrer at 90 °C for 14 h. The column chromatography purification (PE/EA, 98:02 to 95:05), furnished **9c** (63 mg, 87%) as colourless viscus liquid. [TLC monitor PE/EA 95:05), $R_{\rm f}$ (**7c**) = 0.20, $R_{\rm f}$ (**9c**) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.66 (dd, *J* = 8.4 and 1.1 Hz, 2H), 7.37 – 7.25 (m, 4H), 7.24 – 7.14 (m, 3H), 7.14 – 7.05 (m, 2H), 7.07 – 6.90 (m, 7H), 6.90 – 6.81 (m, 1H), 4.24 (dd, *J* = 7.0 and 1.3 Hz, 1H), 3.32 (dd, *J* = 17.0 and 7.1 Hz, 1H), 2.90 (d, *J* = 16.2 Hz, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 154.1, 144.8, 144.0, 141.5, 129.5, 129.2, 128.5, 127.7, 127.5, 127.3, 127.0, 126.3, 126.3, 126.1, 125.8, 121.4, 121.0, 117.4, 83.4, 44.4, 30.1 ppm. HRMS (ESI) calculated [M+H]⁺ for C₂₇H₂₃O⁺: 363.1743, found: 363.1737.

6-Methoxy-2,2-dimethyl-3-phenylchromane (9d): **GP-6** was followed with alcohol **7d** (0.20 mmol), Pd(OAc)₂ (3 mol%), JohnPhos (5 mol%), Cs₂CO₃ (0.40 mmol), 2 mL toluene(dry) was added and allowed to stirrer at 90 °C for 14 h. The column chromatography purification (PE/EA,, 98:02 to 93:07), furnished

9d (42 mg, 79%) as colourless viscus liquid. [TLC monitor PE/EA 93:07), $R_{\rm f}(7d) = 0.20$, $R_{\rm f}(9d) = 0.70$, UV detection]. ¹H NMR (400 MHz, CDCl₃) $\delta = 7.26 - 7.12$ (m, 5H), 6.71 (d, J = 8.8 Hz, 1H), 6.65 (dd, J = 8.9 and 3.0 Hz, 1H), 6.56 (d, J = 2.9 Hz, 1H), 3.67 (s, 3H), 3.07 (dd, J = 16.1 and 9.9 Hz, 1H), 2.95 (dd, J = 9.9 and 5.4 Hz, 1H), 2.86 (dd, J = 16.1 and 5.4 Hz, 1H), 1.20 (s, 3H), 1.11 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) $\delta = 153.1$, 147.5, 141.5, 128.7, 128.2, 126.9, 122.1, 117.7, 113.7, 113.5, 76.8, 55.6, 47.7, 29.5, 27.9, 21.4 ppm. HRMS (ESI) calculated $M+H]^+$ for C₁₈H₂₁O₂⁺: 269.1536, found: 269.1539.

2,2-Dimethyl-3-(*m***-tolyl)chromane (9e): GP-6** was followed with alcohol **7e** (0.20 mmol), Pd(OAc)₂ (3 mol%), JohnPhos (5 mol%), Cs₂CO₃ (0.40 mmol), 2 mL toluene(dry) was added and allowed to stirrer at 90 °C for 14 h. The column chromatography purification (PE/EA, 98:02 to 95:05), furnished **9e** (46 mg, 92%) as colourless viscus liquid. [TLC monitor PE/EA 95:05), $R_{\rm f}$ (**7e**) = 0.20, $R_{\rm f}$ (**9e**) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.24 - 7.16 (m, 1H), 7.17 - 7.11 (m, 1H), 7.11 - 7.01 (m, 4H), 6.87 (dd, *J* = 11.8, 4.5 Hz, 2H), 3.20 (dd, *J* = 16.3, 10.9 Hz, 1H), 3.03 (dd, *J* = 10.9, 5.3 Hz, 1H), 2.92 (dd, *J* = 16.3, 5.3 Hz, 1H), 2.35 (s, 3H), 1.30 (s, 3H), 1.22 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 153.5, 141.4, 137.7, 129.6, 129.3, 128.1, 127.6, 127.3, 125.8, 121.7, 119.9, 117.1, 77.3, 47.8, 29.1, 28.2, 21.5, 21.1 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₈H₂₁O⁺: 253.1587, found: 253.1584.

2,2-Diethyl-3-(m-tolyl)chromane (9f): GP-6 was followed with alcohol 7f (0.20 mmol), Pd(OAc)₂ (3 mol%), JohnPhos (5 mol%), Cs₂CO₃ (0.40 mmol), 2 mL toluene(dry) was added and allowed to stirrer at 90 °C for 14 h. The column chromatography purification (PE/EA, 98:02 to 95:05), furnished 9f (48 mg, 85%) as colourless viscus liquid. [TLC monitor PE/EA 95:05), Rf(7f) = $0.20, R_{f}(9f) = 0.70, UV detection].^{1}H NMR (400 MHz, CDCl_{3}) \delta$ = 7.20 - 7.10 (m, 2H), 7.08 - 6.96 (m, 4H), 6.93 - 6.81 (m, 2H), B.17 (t, J = 6.7 Hz, 1H), 3.09 (dd, J = 17.0 and 6.5 Hz, 1H), 2.98 (dd, J = 17.0 and 6.9 Hz, 1H), 2.30 (s, 3H), 1.74 (dqd, J = 14.8, 7.5 and 2.4 Hz, 2H), 1.65 – 1.50 (m, 2H), 1.41 (dq, J = 14.6 and 7.4 Hz, 1H), 0.93 (t, J = 7.4 Hz, 3H), 0.85 (t, J = 7.5 Hz, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 153.8, 142.3, 137.6, 129.5, 129.4, 128.2, 127.3, 127.3, 125.6, 121.6, 119.9, 117.3, 80.3, 43.1, 29.4, 26.7, 25.4, 21.5, 7.5, 7.3 ppm. HRMS (ESI) calculated [M+H]⁺ for C₂₀H₂₅O⁺: 281.1900, found: 281.1899.

2,2-Dimethyl-3-(p-tolyl)chromane (9g): GP-6 was followed with alcohol **7g** (0.20 mmol), Pd(OAc)₂ (3 mol%), JohnPhos (5 mol%), Cs₂CO₃ (0.40 mmol), 2 mL toluene(dry) was added and allowed to stirrer at 90 °C for 14 h. The column chromatography purification (PE/EA, 98:02 to 95:05), furnished **9g** (45 mg, 90%) as colourless viscus liquid. [TLC monitor (PE/EA 95:05), *R*₁(**7g**)

= 0.20, $R_f(9g) = 0.70$, UV detection]. ¹H NMR (400 MHz, CDCl₃) $\delta = 7.18 - 7.08$ (m, 6H), 6.88 (dd, J = 11.6, 4.4 Hz, 2H), 3.19 (dd, J = 16.2 and 10.8 Hz, 1H), 3.04 (dd, J = 10.7 and 5.3 Hz, 1H), 2.93 (dd, J = 16.3 and 5.3 Hz, 1H), 2.35 (s, 3H), 1.31 (s, 3H), 1.22 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) $\delta = 153.5$, 138.4, 136.5, 129.3, 128.9, 128.6, 127.3, 121.7, 119.9, 117.1, 77.3, 47.4, 29.2, 28.1, 21.1, 21.0 ppm. HRMS (ESI) calculated [M+H]⁺ for C₁₈H₂₁O⁺: 253.1587, found: 253.1585.

2,2-Dipropyl-3-(*p***-tolyl)chromane (9h): GP-6** was carried out with alcohol **7h** (0.20 mmol), Pd(OAc)₂ (3 mol%), JohnPhos (5 mol%), Cs₂CO₃ (0.40 mmol), 2 mL toluene(dry) was added and allowed to stirrer at 90 °C for 14 h. The column chromatography purification (PE/EA, 98:02 to 95:05), furnished **9h** (50 mg, 81%) as colourless viscus liquid. [TLC monitor (PE/EA 95:05), $R_{\rm f}$ (**7h**) = 0.20, $R_{\rm f}$ (**9h**) = 0.70, UV detection]. ¹H NMR (400 MHz, CDCl₃) δ = 7.19 – 7.11 (m, 1H), 7.12 – 7.00 (m, 5H), 6.94 – 6.79 (m, 2H), 3.15 (t, *J* = 6.6 Hz, 1H), 3.03 (ddd, *J* = 41.2, 16.7, 6.5 Hz, 2H), 2.32 (s, 3H), 1.73 – 1.59 (m, 2H), 1.54 – 1.28 (m, 6H), 0.88 (t, *J* = 7.1 Hz, 3H), 0.80 (t, *J* = 7.1 Hz, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ = 153.8, 139.3, 136.1, 129.4, 128.9, 128.5, 127.2, 121.5, 119.9, 117.2, 80.2, 43.3, 37.3, 35.7, 29.5, 21.0, 16.5, 16.3, 14.5 ppm. HRMS (ESI) calculated [M+H]⁺ for C₂₂H₂₉O⁺: 309.2213, found: 309.2205.

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Keywords: Isoflavans 1 • Intramolecular arylation 2 • Iodobenzene 3 • malonates 4 • *ortho*-bromobenzylbromide 5

References:

- (a) Q. Shelby, N. Kataoka, G. Mann, J. Hartwig, J. Am. Chem. Soc.,
 2000, 122, 10718–10719; (b) A. Neogi, T. P. Majhi, B. Achari, P. Chattopadhyay, Eur. J. Org. Chem., 2008, 330–336; (c) B. Xu, J. Xue, J. Zhu, Y. Li, Chem. Lett., 2008, 37, 202–203; (d) J. Niu, P. Guo, J. Kang, Z. Li, J. Xu, S. Hu, J. Org. Chem., 2009, 74, 5075–5078; (e) H. Adams, N. J. Gilmore, S. Jones, M. P. Muldowney, S. H. von Reuss, R. Vemula, Org. Lett., 2008, 10, 1457–1460; (f) J. Zhao, Y. Zhao, H. Fu, Angew. Chem., Int. Ed. Engl., 2011, 50, 3769–3773; (g) Y. Fang, C. Li, J. Org. Chem., 2006, 71, 6427–6431; (h) G. Satyanarayana, M. E. Maier, J. Ora. Chem., 2008, 73, 5410–5415.
- [2] (a) B. M. Trost, H. C. Shen, L. Dong, J.-P. Surivet, C. Sylvain, *J. Am. Chem. Soc.*, **2004**, *126*, 11966–11983; (b) J. Barluenga, M. Trincado, E. Rubio, J. M. González, *J. Am. Chem. Soc.*, **2004**, *126*, 3416–3417; (c) Y. Yamamoto, K. Itonaga, *Org. Lett.*, **2009**, *11*, 717–720; (d) T. T. Dang, F. Boeck, L. Hintermann, *J. Org. Chem.*, **2011**, *76*, 9353–9361.

- [5] [6]
- [3] (a) E. N. Pitsinos, V. P. Vidali, E. A. Couladouros, *Eur. J. Org. Chem.*, 2011, 1207–1222; (b) G. Evano, N. Blanchard, M. Toumi, *Chem. Rev.*, 2008, *108*, 3054–3131; (c) B. Schlummer, U. Scholz, *Adv. Synth. Catal.*, 2004, *346*, 1599–1626; (d) F. Monnier, M. Taillefer, *Angew. Chem., Int. Ed. Engl*, 2008, *120*, 3140–3143.
- [4] (a) M. Palucki, J. P. Wolfe, S. L. Buchwald, *J. Am. Chem. Soc.*, **1996**, *118*, 10333–10334; (b) K. E. Torraca, S.-I. Kuwabe, S. L. Buchwald, *J. Am. Chem. Soc.*, **2000**, *122*, 12907–12908; (c) S. Kuwabe, K. E. Torraca, S. L. Buchwald, *J. Am. Chem. Soc.*, **2001**, *123*, 12202–12206.
 [5] N. C. Veitch, *Nat. Prod. Rep.*, **2007**, *24*, 417–464.
 - (a) D. Slade, D. Ferreira, J. P. J. Marais, *Phytochemistry*, **2005**, *66*, 2177–2215;
 (b) J. Justino, *Flavonoids: From Biosynthesis to Human Health*, BoD Books on Demand, **2017**.
 - (a) W. B. Eyton, W. D. Ollis, I. O. Sutherland, O. R. Gottlieb, M. Taveira Magalhâes, L. M. Jackman, *Tetrahedron*, **1965**, *21*, 2683–2696; (b) M. Gregson, W. D. Ollis, B. T. Redman, I. O. Sutherland, H. H. Dietrichs, O. R. Gottlieb, *Phytochemistry*, **1978**, *17*, 1395–1400; (c) M. Gregson, W. D. Ollis, I. O. Sutherland, O. R. Gottlieb, M. T. Magalhães, *Phytochemistry*, **1978**, *17*, 1375–1377.
 - (a) Comprehensive Natural Products II: Chemistry and Biology, Elsevier,
 2010; (b) J. B. Harborne, T. J. Mabry, *The Flavonoids: Advances in Research*, Springer US, 1982. (f) A. Francisco, *Phytochem. Anal.*, 1995,
 6, 55–55; (c) G. M. Boland, D. M. X. Donnelly, J.-P. Finet, M. D. Rea, *J. Chem. Soc., Perkin Trans.* 1, 1996, 2591–2597; (d) D. M. X.
 Donnelly, G. M. Boland, *Nat. Prod. Rep.*, 1995, *12*, 321–338; (e) D. A.
 Horton, G. T. Bourne, M. L. Smythe, *Chem. Rev.*, 2003, *103*, 893–930; (f) H. C. Shen, *Tetrahedron*, 2009, *65*, 3925–3929.
 - (a) G. F. Marrian, G. A. D. Haslewood, *Biochem J*, **1932**, *26*, 1227–1232;
 (b) G. F. Marrian, D. Beall, *Biochem J*, **1935**, *29*, 1586–1589.
 - (a) Nottle MC. *Res Vet Sci.* **1976**; *21*, 309-317; (b) K. D. R. Setchell, A. M. Lawson, F. L. Mitchell, H. Adlercreutz, D. N. Kirk, M. Axelson, *Nature*, **1980**, *287*, 740–742.
 - T. D. Lund, D. J. Munson, M. E. Haldy, K. D. R. Setchell, E. D. Lephart, R. J. Handa, *Biol Reprod*, **2004**, *70*, 1188–1195.
 - H. D. VanEtten, Phytochemistry, 1976, 15, 655-659.
 - X. Hu, J.-W. Wu, M. Wang, M.-H. Yu, Q.-S. Zhao, H.-Y. Wang, A.-J. Hou, *J. Nat. Prod.*, **2012**, *75*, 82–87.
 - (a) J. L. Ingham, R. L. Millar, *Nature*, **1973**, *242*, 125–126;
 (b) R. W. Miller, G. F. Spencer, A. R. Putnam, *J. Nat. Prod.*, **1989**, *52*, 634–636;
 (c) M. R. Bonde, R. L. Millar, J. L. Ingham, *Phytochemistry*, **1973**, *12*, 2957–2959;
 (d) P. W. Grosvenor, D. O. Gray, *J. Nat. Prod.*, **1998**, *61*, 99–101;
 (e) T. Miyase, M. Sano, H. Nakai, M. Muraoka, M. Nakazawa, M. Suzuki, K. Yoshino, Y. Nishihara, J. Tanai, *Phytochemistry*, **1999**, *52*, 303–310;
 (f) M. Mori-Hongo, H. Takimoto, T. Katagiri, M. Kimura, Y. Ikeda, T. Miyase, *J. Nat. Prod.*, **2009**, *72*, 194–203.
 - [15] (a) F. Wessely, F. Prillinger, **1939**, *72*, 629–633; (b) J. A. Lamberton, H. Suares, K. G. Watson, *Aust. J. Chem.*, **1978**, *31*, 455–457; (c) S. J. Gharpure, A. M. Sathiyanarayanan, P. Jonnalagadda, *Tetrahedron Lett.*, **2008**, *49*, 2974–2978.
 - [16] (a) S. Usse, G. Guillaumet, M.-C. Viaud, *Tetrahedron Lett.*, **1997**, *38*, 5501–5502; (b) H. G. Krishnamurty & S. Sathyanarayana. *Synth. Commun.*, **1986**, *16* 1657-1663; (c) Liepa, A. J. *Aust. J. Chem.* **1984**, 37, 2545; (d) Deschamps-Vallet, C.; Ilotse, J-B.; Meyer-Dayan, M. *Tetrahedron lett.* **1983**, *24*, 3993; (e) Burali, C.; Desideri, N.; Stein, M. C.; Conti, C.; Orsi, N. *Eur. J. Med. Chem.*, **1987**, *22*, 119; (f) Gupta, A.; Ray, S. Synthesis, **2008**, *23*, 3783; (g) Setchell, K. D. R.; Sorokin, V. D.

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- [17] (a) R. S. Muthyala, Y. H. Ju, S. Sheng, L. D. Williams, D. R. Doerge, B. S. Katzenellenbogen, W. G. Helferich, J. A. Katzenellenbogen, *Bioorg. Med. Chem.*, 2004, *12*, 1559–1567; (b) X.-L. Wang, H.-G. Hur, J. H. Lee, K. T. Kim, S.-I. Kim, *Appl. Environ. Microbiol.*, 2005, *71*, 214–219; (c) S.-R. Li, P.-Y. Chen, L.-Y. Chen, Y.-F. Lo, I.-L. Tsai, E.-C. Wang, *Tetrahedron Lett.*, 2009, *50*, 2121–2123.
- [18] (a) M. Versteeg, B. C. B. Bezuidenhoudt, D. Ferreira, K. J. Swart, J. Chem. Soc., Chem. Commun., 1995, 1317–1318; (b) M. Versteeg, B. C. B. Bezuidenhoudt, D. Ferreira, *Tetrahedron*, 1999, *55*, 3365–3376.
- [19] J. M. Heemstra, S. A. Kerrigan, D. R. Doerge, W. G. H. mA. Boul, Willia, Org. Lett., 2007, 9, 379–379.
- [20] S. Yang, S.-F. Zhu, C.-M. Zhang, S. Song, Y.-B. Yu, S. Li, Q.-L. Zhou, *Tetrahedron*, **2012**, *68*, 5172–5178.
- [21] Y. Takashima, Y. Kaneko, Y. Kobayashi, *Tetrahedron*, **2010**, *66*, 197–207.
- [22] J. Xia, Y. Nie, G. Yang, Y. Liu, W. Zhang, Org. Lett., 2017, 19, 4884– 4887.
- [23] S. J. Gharpure, A. M. Sathiyanarayanan, P. K. Vuram, RSC Adv., 2013, 3, 18279-18282.
- [24] (a) L. Mahendar, J. Krishna, A. G. K. Reddy, B. V. Ramulu, G. Satyanarayana, Org. Lett. 2012, 14, 628–631; (b) D. R. Kumar, G. Satyanarayana, Org. Lett. 2015, 17, 5894–5897; (c) J. Krishna, A. G. K. Reddy, G. Satyanarayana, Adv. Synth. Catal. 2015, 357, 3597–3610; (d) K. Ramesh, G. Satyanarayana, J. Org. Chem. 2017, 82, 4254-4264; (e) B. Suchand, G. Satyanarayana, J. Org. Chem. 2016, 81, 6409-6423; (f) B. Suchand, G. Satyanarayana, J. Org. Chem. 2017, 82, 372-381; (g) C. Sreenivasulu, D. A. Thadathil, S. Pal, S. Gedu, Synth. Commun., 2020, 50, 112–122. (h) C. Sreenivasulu, G. Satyanarayana, Eur. J. Org. Chem., 2018, 2846–2857.
- [25] B. Suchand, J. Krishna, B. Venkat Ramulu, D. Dibyendu, A. Gopi Krishna Reddy, L. Mahendar, G. Satyanarayana, *Tetrahedron Lett.*, 2012, 53, 3861–3864.
- [26] B. Suchand, J. Krishna, K. Mritunjoy and G. Satyanarayana, RSC Adv., 2014, 4, 13941–13945.
- [27] (a) G. Pandey, M. Karthikeyan, A. Murugan, J. Org. Chem. 1998, 63, 2867–2872; (b) L. Ma, D. Seidel, Chem. Eur. J., 2015, 21, 12908–12913; (c) S. F. Yip, H. Y. Cheung, Z. Zhou, F. Y. Kwong, Org. Lett. 2007, 9, 3469–3472.
- [28] (a) J. M. Heemstra, S. A. Kerrigan, D. R. Doerge, W. G. H. m A. Boul and Willia, Org. Lett. 2007, 9, 379–379; (b) L. Zheng, F. Gao, C. Yang, G.-L. Gao, Y. Zhao, Y. Gao, W. Xia, Org. Lett. 2017, 19, 5086–5089; (c) J. M. Heemstra, S. A. Kerrigan, D. R. Doerge, W. G. Helferich, W. A. Boulanger, Org. Lett. 2006, 8, 5441–5443; (d) Y.-C. Wong, T. T. Jayanth, C.-H. Cheng, Org. Lett. 2006, 8, 5613–5616.

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Insert text for Table of Contents here. We have presented a straightforward strategy for isoflavans synthesis. The strategy features an intermolecular [Cu]-catalyzed arylation of malonates and an intramolecular [Pd]-catalyzed Buchwald coupling of hydroxyl tethered promoarenes, as the key transformations. This method is practical and flexible in enabling the synthesis of different isoflavan derivatives.

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