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# Free radical scavenging abilities of flavonylthiazolidine-2,4-dione compounds

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ABSTRACT: Free radical scavenging activity of flavonyl-thiazolidine-2,4-dione compounds has been evaluated using chemiluminescence, electron spin resonance spectroscopy with 5,5-dimethyl-1-pyrroline-1-oxide as spin trap and DPPH (2,2'diphenyl-1-picrylhydrazyl) method. The examined compounds exhibited 28–50% scavenging superoxide anion radical  $(O_2^{\bullet})$ , 16.7–76.7% hydroxyl radical (HO<sup>•</sup>) and 9–40% DPPH radical. Compounds containing carbonyl group in their structure can be considered as antioxidants with high relevance and great biological importance. Copyright © 2009 John Wiley & Sons, Ltd.

Keywords: flavone; flavonyl-thiazolidine-2,4-dione compounds; free radicals; scavenging capacity

### Introduction

Free radical-mediated lipid peroxidation, oxidative stress and antioxidants are widely discussed in many current research areas. (1-3) Oxygen-centered free radicals such as hydroxyl radical (HO<sup>•</sup>) and superoxide anion radical ( $O_2^{\bullet}$ ) and other reactive oxygen species (ROS) are the products of normal cellular metabolic processes and pathological stress in human body. (1) They play dual role that is beneficial and harmful in the biology of the cell. (2,3) Under normal conditions ROS and free radicals are effectively eliminated by antioxidant defense systems such as antioxidant enzymes and non-enzymatic factors. (4) However, under pathological conditions, the balance between the generation and elimination of ROS is broken; as a result of these events, biomacromolecules including DNA, membrane lipids and proteins, are damaged by ROS-mediated oxidative stress. The term oxidative stress in essence refers to the situation of a serious imbalance between the production of free radicals and the antioxidant defense mechanisms, leading to potential tissue damage. (1,3) Uncontrolled generation of free radicals that attack membrane lipids, protein and DNA is believed to be involved in many health disorders such as diabetes mellitus, cancer and neurodegenerative and inflammatory diseases. (5-9)

Diabetes mellitus, a state of chronic hyperglycemia, is a major cause of serious micro- and macrovascular diseases, affecting, therefore, nearly every system in the body. Growing evidence indicates that oxidative stress is increased in diabetes due to overproduction of ROS and decreased efficiency of antioxidant defenses, a process that starts very early and worsens over the course of the disease. (5,6,8,10,11)

Hyperglycemia is reported to induce oxidative stress through multiple pathways such as redox imbalances secondary to enhanced aldose reductase activity, increased advanced glycation end products, altered protein kinase C activity, especially  $\beta$ -isoforms, prostanoid imbalances and mitochondrial overproduction of superoxide. (12) All these pathways converge in the production of oxidative stress. (8,12,13) Thiazolidinediones (TZDs) such as pioglitazone and rosiglitazone are a new class of antidiabetic agents that reduce plasma glucose and glucose production and also increase glucose clearance in patients with type 2 diabetes, thus reducing insulin resistance. (10,14) Besides their anti-diabetic potency, these TZDs have been shown to exert antioxidant activity. Pioglitazone inhibits  $O_2^-$  radical production in endothelial cells. Rosiglitazone ameliorates the impaired coronary arteriolar dilation in mice with type 2 diabetes by reducing oxidative stress via a mechanism unrelated to its effect on hyperglycaemia. (14)

Flavonoids are a large family of compounds with common chemical structure that have recently attracted considerable interest due to their broad pharmacological and therapeutical effects. They exist extensively in all parts of plants, including fruits, vegetables, seeds, nuts, herbs, flowers and bark. (15) They have been described as health-promoting, disease-preventing dietary supplements which are extremely safe and associated with low toxicity. (16) In addition, traditional herbal medicines having flavones have been recommended for the treatment of diabetes and the antidiabetic activity of several plant extracts has been correlated with their antioxidant properties. As antioxi-

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dants, these compounds may prevent the progressive impairment of pancreatic  $\beta$ -cell function due to oxidative stress, thus reducing the occurrence of type 2 diabetes. (17) As free radical scavengers, flavonoids (15,16,18,19) can act as powerful antioxidants, even more so than the traditional vitamins. (20,21)

In our previous studies, we reported the synthesis and the *in vitro* antidiabetic and aldose reductase inhibitory activity results of the flavonyl-2,4-thiazolidinediones (FTDs) **1–6**. (22) In this paper, the *in vitro* effect of FTDs (Fig. 1) having antidiabetic and aldose reductase inhibitory activity was examined on the reactions generating  $O_2^-$ , HO and DPPH radicals.





# Materials and methods

The compounds **1–3** were synthesized by Knoevenagel reaction of ethyl 2,4-dioxothiazolidine-3-ylacetate with flavone-3'/4'/6-carboxaldehydes using acetic acid-sodium acetate mixture. The compounds **4–6** were obtained with acidic hydrolysis of compounds **1–3**. (22)

#### Chemicals

Ammonium ferrous sulphate hexahydrate was from Fluka (Buchs, Switzerland); catalase (19,900 U mg<sup>-1</sup>) from bovine liver and *p*-nitroblue-tetrazolium chloride (NBT) were from Sigma (St Louis, MO, USA). All remaining reagents were purchased from E. Merck (Darmstadt, Germany) and Aldrich (Milwaukee, MI, USA). The reagents were propared fresh daily; dimethylsulfoxide (DMSO) was prepared as a 0.1 mol L<sup>-1</sup> aqueous solution immediately before use and stored in darkness at  $-15^{\circ}$ C under nitrogen to avoid any oxidation of the spin trap.

#### Superoxide anion radical scavenging activity

Superoxide anion radical was generated in a KO<sub>2</sub>–DMSO system. (23) 18-Crown-6 (1,4,7,10,13,16-hexaoxacyclo-octadecane) (60 mg) in 10 mL anhydrous dimethylsulfoxide (DMSO) was mixed quickly to avoid contact with air humidity with 7 mg of KO<sub>2</sub>. The mixture was stirred for 1 h to give a pale yellow solution of  $O_2^{\overline{2}}$ , which was stable at room temperature for at least 1 h. The  $O_2^{\overline{2}}$  concentration was determined using a UV spectrum ( $\lambda_{max} = 251$  nm,  $\varepsilon = 2686 \pm 29$  mol cm<sup>-1</sup>).

Chemiluminescence (CL) measurements were made using an EMI9553Q photomultiplier with an S20 cathode sensitive in the range of 200–800 nm, interfaced with a computer for data acquisition and handling. Reagents were introduced to a thermostated glass cuvette placed in a light-tight chamber. The cuvette was exhausted and washed using a B-169 vacuum system (Büchi, Flawill, Switzerland). The quenching activity was calculated as follows:

Quenching ratio  $Q(\%) = [(\Sigma I_0 - \Sigma I_x)/\Sigma I_0] \times 100\%$ , where  $\Sigma I_0$  is the integrated light intensity (CL sum) measured during

3 min  $\left( \Sigma I = \int_{t=0}^{t=3} I(t) dt \right)$  in the absence of a inhibitor (an FTD)

compound),  $\Sigma I_x$  is the light sum measured in the presence of the inhibitor. Because FTDs were dissolved in DMSO and added to the reaction mixture caused a short-lasting small 'flash' followed by a very small increase in the light emission (data not shown); this reaction was considered as the control reaction.

#### Hydroxyl radical scavenging activity

Electron spin resonance (ESR) and the spin trap agent 5,5-dimethyl-1-pyrroline-*N*-oxide (DMPO) were used to determine the scavenging hydroxyl radical generated in the Fenton reaction: (24)

$$H_2O_2 + Fe(II) \rightarrow HO^{\bullet} + Fe(III) + HO^{-}$$
(1)

This reaction is accompanied by two other reactions

$$Fe(II) + HO^{\bullet} \rightarrow Fe(III) + HO^{-}$$
<sup>(2)</sup>

$$H_2O_2 + HO^{\bullet} \rightarrow HOO^{\bullet} + H_2O$$
 (3)

In the presence of the DMPO trap, HO' is rapidly trapped by this spin trap at high rate constant of  $3.4 \times 10^9$  mol<sup>-1</sup> s<sup>-1</sup>: (25)

$$\mathsf{DMPO} + \mathsf{HO}^{\bullet} \to \mathsf{DMPO} - \mathsf{OH} \tag{4}$$

The rate constant for reaction of the HOO<sup>•</sup> with this nitrone spin trap is very low (~10 mol<sup>-1</sup> s<sup>-1</sup>).

The Fenton reaction contained 200  $\mu$ L of 0.1 mol<sup>-1</sup> DMPO (aqueous solution), 50  $\mu$ L of 4 mmol L<sup>-1</sup> H<sub>2</sub>O<sub>2</sub>, 50  $\mu$ L of 80 mmol L<sup>-1</sup> sodium trifluoroacetate (pH 6.15) and 50  $\mu$ L of ammonium ferrous sulfate hexahydrate solution at concentration of 0.5 mmol L<sup>-1</sup>. The reaction mixture was placed in the ESR cavity using a quartz flat cell with an optical pathlength of 0.25 mm. The spin-trapped DMPO–OH signal was analyzed 5 min after the addition of ammonium ferrous sulfate hexahydrate on a standard ESR spectrometer operating at 9.3 GHz with 100 kHz modulation of the steady magnetic field.

Tested FTDs were dissolved in DMSO, which is suitable for dissolving water-insoluble samples. (26)

The ability to scavenge the HO radicals was calculated using the following formula:

Hydroxyl radical scavenging ratio  $R(\%) = [H_0 - H) / H_0] \times 100\%$ 

where  $H_{o}$  and H represent the relative height of the second peak in the ESR spectrum of the spin-adduct from the blank (DMSO) and a sample dissolved in DMSO, respectively.

The conditions of ESR measurement were as follows: microwave power 0.63 mW, modulation amplitude 0.2 mT, time constant 0.1 s, receiver gain  $4 \times 10^4$ , and temperature 293 K.

#### **DPPH radical scavenging activity**

Free radical scavenging activity was evaluated according to the method reported by Shimada *et al.* (27) Two milliliters of 0.1 mmol L<sup>-1</sup> DPPH radical in ethanol was mixed with 2 mL of sample solution (2.5 mmol L<sup>-1</sup>) in DMSO and incubated at 24°C for 30 min in the dark. The decrease in absorbance at 517 nm was measured against ethanol using a UV–vis Zeiss M-40 spectrophotometer (Zeiss, Germany). Scavenging capacity *A* (%) was calculated using the following equation:

$$A(\%) = [1 - (A_1 - A_2) : A_0] \times 100\%$$

where  $A_1$  is the absorbance of 2 mL DPPH radical solution and 2 mL sample of a FTD, solution,  $A_2$  is the absorbance of 2 mL sample of a FTD solution and 2 mL ethanol,  $A_0$  is the absorbance of 2 mL DPPH radical solution and 2 mL DMSO.

# **Results and discussion**

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Within the framework of this paper, the superoxide radical scavenging effect of six novel synthesized compounds was estimated employing the CL from  $O_2^-$  radicals generated in DMSO. Superoxide anion radical is a reduced form of molecular oxygen formed by receiving one electron. (28) The radical is known as a precursor of very harmful species to cellular components such as HO radicals as well as singlet oxygen ( $^1O_2$ ) and  $H_2O_2$ . In addition, this radical is a one-electron reductant of transition metal ions. (28) Superoxide anion radical in DMSO elicits strong light emission according to the following reactions: (29–31)

$$O_2^{\overline{\bullet}} \to {}^1O_2 + \text{electron}$$
 (5)

$$2O_2^{\overline{\bullet}} + 2Me_2SO \rightarrow {}^1O_2 + Me_2SO_2 + OH^- + MeSO(CH_2)^-$$
(6)

$$2^{1}O_{2} \rightarrow (^{1}O_{2})_{2} \rightarrow (^{3}O_{2})_{2} + \text{light emission}$$
 (7)

Addition of the  $O_2^-$  radical inhibitors leads to a decrease in the light intensity, allowing the evaluation of their antioxidant activity. This reaction has high sensivity and selectivity and has been applied successfully for evaluation of an antioxidative property of the biologically reactive compounds in nonaqueous media. (32)

Typical chemiluminescence intensity-time diagrams in the absence (blank) and in the presence of a antioxidant (sample) were givn in our previous paper. (33)

Four of the six examined FTDs compounds dissolved in DMSO reduced the light sum at least by 28% (range 28–50%) at a concentration of 0.5 mmol L<sup>-1</sup> (Fig. 2). One of these compounds (denoted as **4**) exhibited the O<sub>2</sub> radical scavenging activity comparable to that for specific this radical scavenger NBT (1 mmol L<sup>-1</sup>) and a little lower than (1 mmol L<sup>-1</sup>) Tiron (1,2-dihydroxy benzene-3,5-disulphonic acid) (56%) at the same concentration. The CL sums for these two inhibitors were accounted for by integrating the area under the kinetic curve I = f(t) after 20 min. The sums were compared with those of the control (without examined compound), considered as 90%. In contrast, under the conditions examined, comounds **1** and **2** increased light emission by about 99 and 207%, respectively.

The enhanced chemiluminescence from the  $O_2^-$  radical–DMSO system in the presence of compounds 1 and 2 indicates that both compounds were able to stimulate the  $O_2^-$  radical dismutation as follows:

$$2O_{2}^{\bar{\bullet}} + 2H^{+} \rightarrow {}^{1}O_{2} + H_{2}O_{2}$$
(8)

This means that these compounds might deliver H<sup>+</sup> for reaction (8) that is a source of singlet oxygen. (34) To check if H<sub>2</sub>O<sub>2</sub> and <sup>1</sup>O<sub>2</sub> are generated in reaction (8) the effect of catalase (an enzyme responsible for destruction of H<sub>2</sub>O<sub>2</sub> (28,35)) and histidine (a quencher of <sup>1</sup>O<sub>2</sub> (28)) was examined. An addition of the enzyme at a concentration of 200 µg/mL caused a decrease in the CL sum of about 40%. Also the aqueous solution of histidine (1 mmol L<sup>-1</sup>) inhibited the light emission by 46%, although water alone at the same volume (5%) as these tested antioxidant H<sub>2</sub>O solutions quenched the CL by about 10%. In contrast, an addition of O<sub>2</sub><sup>-</sup> radical into hydrogen peroxide, (35) to the O<sub>2</sub><sup>-</sup> –DMSO solution increased the light sum by about 45%. Direct addition of H<sub>2</sub>O<sub>2</sub> (0.1 mmol L<sup>-1</sup>) elicited a time-dependent increment of light emission by about 20% (data not shown).

The antioxidant activities of the individual compound may depend on the structure of the free-radical inhibitors, the substituents present on the rings, e.g. free carboxylic group and the degree of polymerization.

Compounds containing the  $-CH_2COOH$  group exhibited the strong direct activity towards the  $O_2^-$  radical. This is consistent with a previous finding of Sawyer and Gibian, (36) who reported that weakly acidic  $O_2^-$  radical as a weak base can promote the H<sup>+</sup> transfer from organic compounds.



**Figure 2.** Scavenging effect of the examined flavonyl-thiazolidine-2,4-dione compounds (0.5 mol L<sup>-1</sup>) on the chemiluminescence from 1 mol L<sup>-1</sup>  $O_2^-$  radicals generated in DMSO. The examined scavengers were dissolved in 0.5 mL DMSO. Temperature 294 K. All data represent means  $\pm$  SD of three different experiments. Numbers given at the bottom of the figure refer to compounds listed in Fig. 1. The remaining condictions are reported under Materials and Methods.

Among the oxygen-derived free radicals HO radical is the most reactive unstable oxidizing species and can be produced from  $O_2^$ radical and  $H_2O_2$  in the presence of transition metal ions, such as cooper or iron. (28) This species reacts with a variety of organic compounds, including biomolecules such as lipids, proteins, polypeptides and DNA. When a hydroxyl radical reacts with aromatic compounds, its addition across a double bond is reported, and reaction products such as hydroxycyclo-hexadienyl radical and peroxyl radical are thought to be formed. (37) Except for addition reactions, the HO radical may react by hydrogen abstraction and electron transfer reactions. Indeed the reactions of HO radicals with biomolecules result in generation of other radicals, but they are usually of lower toxicity.

The ability of the tested compounds to scavenge the HO radical was monitored using the ESR spin-trapping assay, the technique routinely used for the sensitive and specific detection of short-lived hydroxyl radicals. The Fenton reaction, a well-documented HO radical generator, was utilized to test the ability of tested compounds to scavenge this radical. Hydroxyl radicals trapped by DMPO form spin-adducts, for which a spectrum is shown in Fig. 3A(a). The spectrum exhibits four splitting lines with an intensity ratio of 1:2:2:1 and hyperfine splitting constant of  $a_N = a_H = 14.9$  G. The parameters are consistent with those values for the DMPO–OH adduct reported by other authors. (25,38) No ESR signal was measured from the Fenton–DMPO reaction, when one substrate was omitted.

The height of the second peak of the spectrum determined the relative amounts of DMPO- $\dot{OH}$  adduct formed in the reaction. When hydroxyl radical scavenger was added, a decrease in the amount of DMPO- $\dot{OH}$  adduct was observed.

Figure 3A(b and c) shows the ESR spectrum measured in the Fenton–DMPO reaction in the presence of salicylic acid and compound **5**, the OH radical scavenger, respectively. Salicylic acid can remove this radical from the reaction at the reaction rate  $1.2 \times 10^{1-}$ L mol<sup>-1</sup> s<sup>-1</sup> (39) The use of salicylic acid at concentration

7.2 mmol L<sup>-1</sup> led to an 80% inhibition. The specificity of the Fenton probe for  $H_2O_2$  and further HO radicals generation was checked by adding catalase (180 µg mL<sup>-1</sup>). We observed a near-complete prevence of appearance of any ESR signal, whereas in the presence of a heat-denaturateed enzyme the signal was decreased by 5%, indicating that the origin of the DMPO–OH signal was dependent on HO radical generation. This conclusion finds confirmation in a fact that superoxide dismutase (SOD) even at a high concentration did not prevent DMPO–OH formation.

The scavenging activities of the tested compounds dissolved in DMSO to this Fenton reaction system with DMPO on HO radicals are shown in Fig. 3B. The scavenging abilities were, in descending order, **6** > **5** > **4** > **3** > **1** > **2** > DMSO, and the amplitude decrease ranged from 16.7  $\pm$  3.5 to 76.7  $\pm$  4.6%. The scavenging effect of 0.37 mmol L<sup>-1</sup> compound **6** (76.7%) was nearly equal to that of the salicylate at concentrartion of 7.2 mmol L<sup>-1</sup>.

The ability of tested compounds to exhaust their H-donating capacity was examined using 2,2-diphenyl-1-picrylhydrazyl radical (DPPH\*). The compound is a stable free radical that has a proton free radical with an absorption maximum band around 515–528 nm. The characteristic absorption decreases significantly on exposure due to the proton-donating ability of the antioxidants proportional to the concentration and antioxidant activity of the antioxidant. (39) The DMPPH radical can be used to the primary characterization of the scavenging activity of compounds. (40–43)

According to the conclusion of Sanchez-Moreno, (44) compounds able to scavenge the DPPH radical *in vitro* may also scavenge polyaromatic hydrocarbon cations *in vivo* by donating hydrogen ions or electrons.

The DPPH radical scavenging capacities of tested FTDs are showed in Fig. 4A. For compounds **2**, **4**, **5** and **6** the reaction was biphasic, with a faster decay in the absorbance in the first dozen minutes, followed by a slower step. The DPPH radical scavenging activity of the examined compounds was evident (p < 0.05), but





Figure 3. (A) The ESR spectra of the DMPO-OH spin adduct arising in the Fenton reaction: (a) ESR spectrum produced by the reaction mixture containing 50 mol L<sup>-1</sup> DMPO, 0.5 mol L<sup>-1</sup> H<sub>2</sub>O<sub>2</sub>, 1.0 mol L<sup>-1</sup> sodium trifluoracetate, pH 6.15, 0.5 mL DMSO, and 50  $\mu$ mol L<sup>-1</sup> FeSO<sub>4</sub> (NH<sub>4</sub>)<sub>2</sub> SO<sub>4</sub>. The reaction was started by an addition of the Fe(II) ion. (b) The signal of DMPO–OH radical quenched by the addition of 7.2 mol  $L^{-1}$  salicylic acid dissolved in 0.5 mL DMSO. (c) The DMPO-OH radical signal quenched by the presence of 0.37 mol L<sup>-1</sup> compound 5 dissolved in 0.5 mL of DMSO. (B) The inhibitory effect of FTD compounds exerted on the DMPO-OH radical formation. The composition of the reaction mixtures was the same as in (A, part c). Data are expressed as means  $\pm$  SD of three measurements. The numbers in figure and on the x-axis refer to the compounds given in Fig. 1. Measurement conditions are reported under the Materials and Methods section.

Compound

32.5

3

38.3

4

5

was lower than ascorbic acid and Trolox at concentrations similar to that used during the compounds examination. The compounds' hierarchy of reducing capacity was 6 > 4 > 5 > 2 > 3 > 1, ranging from  $9 \pm 5$  to  $40 \pm 3\%$  (Fig. 4B).

40

30

20

10

0

28.5

1

16.7

2

It can be observed that FTDs possessing a free carboxylic group, denoted as 4-6, exhibited the highest effectiveness in the free radical scavenging power. The -COOH group is capable of readily donating hydrogen electron.

6

The inhibitory effect examined FTDs containing the -CH<sub>2</sub>-COOH group on DPPH,  $O_2^-$  and HO radicals may represent the direct scavenging activity. Compounds 1-3 could not scavenge free radical as effectively as 4-6. Other structural properties





**Figure 4.** DPPH radical scavenging activity of the FTDs compounds, ascorbic acid 1 mol  $L^{-1}$  and Trolox. (A) Kinetic curves of the reaction between DPPH radical and solutions of the examined compounds. (B) The maximal scavenging effect exerted by examined compounds on DPPH<sup>•</sup>. Results are means  $\pm$  SD of triplicate measurements. Numbers given in the figure refer to compounds listed in Fig. 1. Reaction conditions are described in Materials and Methods.

important for antioxidant nature of the examined compounds include the presence of a C2–C3 double bond in conjugation with 4-oxo function in the C ring. (45) It has been reported that the degree of unsaturation of the C2–C3 bond is an important determinant of high bioactivity of flavonoids. (46) The presence of nitrogen atom in the thiazole ring may also be an active center of FTDs, due to its lone pair electrons. In addition, all examined compounds contain the benzene ring; this also may be valid due to HO and  $O_2^-$  radicals abilities for substitution on the ring. (47)

In conclusion, the results obtained in the present study indicated that FTDs have good antioxidant activities. It has been found that among tested compounds those containing the carboxylic group exerted stronger antioxidant effect than FTDs without this group. These findings suggest that there is promise for FTDs to be used successfully in treating diseases that involve free radicals.

## References

- 1. Dröge W. Free radicals in the physiological control of cell function. Physiol Rev 2002;82:47–95.
- 2. Nordberg J, Arner ESJ. Free Radic Biol Med 2001;31:1287-312.

- Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol 2007;39:44–84.
- 4. Halliwell B. Antioxidant characterization: Methodology and mechanism. Biochem Pharmacol 1995;49:1341–8.
- Valko M, Izakovic M, Mazur M, Rhodes CJ, Telser J. Role of oxygen radicals in DNA damage and cancer incidence. Mol Cell Biochem 2004;266:37–56.
- 6. Nakabeppu Y, Sakumi K, Sakamoto K, Tsuchimoto D, Tsuzuki T, Nakatsu Y. Mutagenesis and carcinogenesis caused by the oxidation of nucleic acids. Biol Chem 2006;387:373–9.
- Zhong-Ji Q, Won-Kyo J, Se-Kwon K. Free radical scavenging activity of a novel antioxidative peptide purified from hydrolysate of bullfrog skin, Rana catesbeiana Shaw. Bioresource Technol 2008;99:1690–8.
- 8. Jie J, Zhang X, Hu Y-S, Wu Y, Wang Q-Z, Li NN, Guo Q-C, Dong XC. Evaluation of in vivo antioxidant activities of Ganoderma lucidum polysaccharides in STZ-diabetic rats. Food Chem 2009;115:32–6.
- 9. Amaral S, Oliveira PJ, Ramalho-Santos J. Diabetes and the impairment of reproductive function: possible role of mitochondria and reactive oxygen species. Curr Diabetes Rev 2008;4:46–54.
- 10. Da Ros R, Assaloni R, Ceriello A. Antioxidant therapy in diabetic complications: What is new? Curr Vasc Pharmacol 2004;2:335–41.
- Subhasree B, Baskar R, Laxmi Keerthana R, Lijina Susan R, Rajasekaran P. Evaluation of antioxidant potential in selected green leafy vegetables. Food Chem 2009;115:1213–20.
- 12. Pop-Busui R, Sima A, Stevens M. Diabetic neuropathy and oxidative stress. Diabetes Metab Res Rev 2006;22:257–73.
- Fukui K, Onodera K, Shinkai T, Suzuki S, Urano S. Impairment of learning and memory in rats caused by oxidative stress and aging, and changes in antioxidative defense systems. Ann NY Acad Sci 2001;928:168–75.
- Manning PJ, Sutherland WH, Walker RJ, Williams SM, de Jong SA, Berry EA. The effect of rosiglitazone on oxidative stress and insulin resistance in overweight individuals. Diabetes Res Clin Pract 2008;81: 209–15.
- 15. Sh. Nafisi A, Shadaloi A, Feizbakhsh HA, Tajmir-Riahi HA. RNA binding to antioxidant flavonoids. J Photochem Photobiol B 2009;94:1–7.
- 16. Havsteen BH. The biochemistry and medical significance of the flavonoids. Pharmacol Ther 2002;96:67–202.
- Rauter AP, Martins A, Lopes R, Ferreira J, Serralheiro LM, Araújo ME, Borges C, Justino J, Silva FV, Goulart M, Thomas-Oates J, Rodrigues JA, Edwards E, Noronha JP, Pinto R, Mota-Filipe H. Bioactivity studies and chemical profile of the antidiabetic plant Genista tenera. J Ethnopharmacol 2009;122:384–93.
- Lim HK, Yoo ES, Moon JY, Jeon YJ, Cho SK. Antioxidant activity of extracts from Dangyuja (Citrus grandis Osbeck) fruits produced in Jeju Island. Food Sci Biotechnol 2006;15:312–6.
- Lin K-W, Liu C-H, Tu H-Y, Ko H-H, Wei B-L. Antioxidant prenylflavonoids from Artocarpus communis and Artocarpus elasticus. Food Chem 2009;115:558–62.
- Naczk M, Shahidi F. Phenolics in cereals, fruits and vegetables: occurrence, extraction and analysis. J Pharm Biomed Anal 2006;41:1523– 42.
- 21. Vijayakumar S, Presannakumar G, Vijayalakshmi NR. Antioxidant activity of banana flavonoids. Fitoterapia 2008;79:279–82.
- Bozdağ-Dündar O, Verspohl EJ, Daş-Evcimen N, Kaup RM, Bauer K, Sarıkaya M, Evranos B, Ertan R. Synthesis and biological activity of some new flavonyl-2,4-thiazolidinediones. Bioorg Med Chem 2008;16:6747–51.
- Valentine JS, Miksztal AR, Sawyer DT. Methods for the study of superoxide chemistry in nonaqueous solutions. Meth Enzymol 1984;105: 71–81.
- 24. Goldstein S, Meyerstein D, Czapski G. The Fenton reagents. Free Radic Biol Med 1993;15:435–45.

- 25. Finkelstein E, Rosen GM, Rauckman EJ. Spin trapping of superoxide and hydroxyl radical: practical aspects. Arch Biochem Biophys 1980;200:1–16.
- 26. Cynshi O, Takashima Y, Katoh Y, Tamura K, Sato M, Fujita Y. Action of phenolic antioxidants on various active oxygen species. J Biolumin Chemilumin 1995;10:261–9.
- 27. Shimada K, Fujikowa K, Yahara K, Nakamura T. Antioxidative properties of xanthan on the autoxidation of soybean oil in cyclodextrin emulsion. J Agric Food Chem 1992;40:945–8.
- Kruk I. Environmental Toxicology and Chemistry of Oxygen Species. The Handbook of Environmental Chemistry. Reactions and Processess, Pt 1 Vol 2 (Eds: Hutzinger O, Kruk I). Springer: New York, 1998.
- 29. Khan AU, Kasha M. Chemiluminescence arising from simultaneous transitions in pairs of singlet oxygen molecules. J Am Chem Soc 1970;92:3293–330.
- 30. Khan UA. Photochem Photobiol 1984;28:615.
- 31. Gampp H, Lippard SJ. Reinvestigation of 18-crown-6 ether/potassium superoxide solutions in Me<sub>2</sub>SO<sub>4</sub>. Inorg Chem 1983;22:357–8.
- 32. Suzuki N, Mashiko S, Hamada M, Nomoto T, Hasegaga M, Yoda BA. Biolominescence and Chemiluminescence Fundamentals and Applied Aspects (Eds: Campbell AK, Kircka LJ, Stanley PE). Wiley: New York 1994.
- Kładna A, Aboul-Enein HY, Kruk I, Lichszteld K, Michalska T. Scavenging of reactive oxygen species by some nonsteroidal anti-inflammatory drugs and fenofibrate. Biopolymers 2006;82:99–105.
- 34. Khan AU, Kasha M. Singlet molecular oxygen in the Haber-Weiss reaction. Proc Natl Acad Sci USA 1994;91:12365–7.
- 35. Halliwell B, Gutteridge JMC. Free Radicals in Biology and Medicine. Clarendon Press: Oxford, 1989.
- 36. Sawyer DT, Gibian MJ. The chemistry of superoxide ion. Tetrahedron 1979;35:1471–81.
- 37. Lee J, Koo N, Min DB. Comprehen Rev Food Sci Food Safety 2004;3: 21.
- Zang LY, Stone K, Pryor WA. Detection of free radicals in aqueous extracts of cigarette tar by electron spin resonance. Free Radic Biol Med 1995;19:161–7.
- Bartosz G. Druga twarz tlenu. Warszawa, Wydawnictwo Naukowe PWN, 2003.
- 39. Brand-Willams W, Cuvelier ME, Berset C. Food Sci Technol Lebensmitt-Wiss Technol 1995;28:25.
- Tung YT, Wu JH, Kuo YH, Chang ST. Antioxidant activities of natural phenolic compounds from Acacia confusa bark. Bioresource Technol 2007;98:1120–3.
- 41. Wang BS, Li BS, Zeng QX, Liu HX. Antioxidant and free radical scavenging activities of pigments extracted from molasses alcohol wastewater. Food Chem 2008;107:1198–204.
- Liu X, Ardo S, Bunning M, Parry J, Zhon K, Stushnoff C, Stoniker F, Yu L, Kendal P. Total phenolic content and DPPHd radical scavenging activity of lettuce (Lactuca sativa L.) grown in Colorado. LWT 2007;40:552–7.
- 43. Sendra JM, Sentandreu E, Navarro JL. Reduction kinetics of the free stable radical 2,2-diphenyl-1-picrylhydrazyl (DPPH•) for determination of the antiradical activity of citrus juices. Eur Food Res Technol 2006;223:615–24.
- 44. Sanchez-Moreno C. Methods used to evaluate the free radical scavenging activity in foods and biological systems. Food Sci Technol Int 2002;8:121–37.
- 45. Rice-Evans CA, Meller NJ, Paganga G. Structure-antioxidant activity relationships of flavonoids and phenolic acids. Free Radic Biol Med 1996;20:933–56.
- 46. Williams RJ, Spencer JPE, Rice-Evans C. Flavonoids: antioxidants or signalling molecules? Free Radic Biol Med 2004;36:838–49.
- 47. Aruoma Ol. Characterization of drugs as antioxidant prophylactics. Free Rad Biol Med 1996;20:675–705.