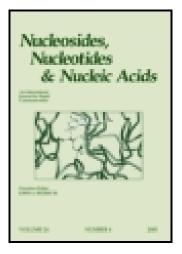
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Nucleosides and Nucleotides

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Synthesis of Novel Isothiazole and Isothiazolo[4,5-d] Pyrimidine Analogues of the Natural C-Nucleosides Pyrazofurin and the Formycins

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SYNTHESIS OF NOVEL ISOTHIAZOLE AND ISOTHIAZOLO[4,5-d] PYRIMIDINE ANALOGUES OF THE NATURAL C-NUCLEOSIDES PYRAZOFURIN AND THE FORMYCINS

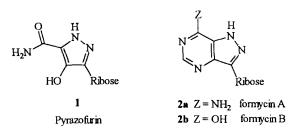
Diederik K. Buffel, Lieven Meerpoel, Suzanne M. Toppet, Georges J. Hoornaert* Laboratorium voor Organische synthese, Departement Scheikunde, Katholieke Universiteit Leuven, Celestijnenlaan 200F, B-3001 Heverlee-Leuven, Belgium

ABSTRACT:

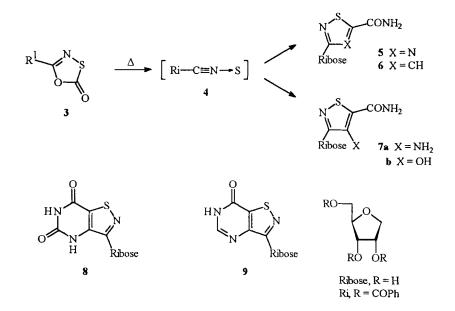
The cycloaddition of tri-O-benzyl-2,5-anhydro-D-allononitrile-N-sulfide with dimethyl acetylenedicarboxylate or with dimethyl fumarate followed by DDQ oxidation was found to give the benzoyl protected dimethyl 3-B-D-ribofuranosylisothiazoledicarboxylate 10. This compound was converted to C-nucleosides 7a, 8 and 9, analogues of pyrazofurin, oxoformycin B and formycin respectively. Despite their structural similarities they did show neither antiviral nor antitumor activity.

Interest in nucleoside analogues relates to their possible antiviral and antitumor activity.¹ Several compounds have been prepared by modifying either the sugar moiety or the heterocycle. Particularly of interest in the latter respect are the C-nucleosides;² due to resistance to hydrolysis of the glycoside linkage these compounds have an enhanced biological stability. An interesting compound in this field is pyrazofurin $1,^3$ a natural C-nucleoside isolated from S. Candidus and synthesized by several groups.⁴ It has a broad spectrum antiviral^{5a,b} and antitumor^{5c} activity. A number of analogues have been synthesized⁶ and some thiophene^{7a} and isoxazole^{7b} analogues were prepared in our laboratory. The formycins 2a and 2b, C-nucleoside analogues of adenosine and inosine, interfere extensively with the nucleic acid metabolism.⁸ Several synthetic routes for these natural products and their analogues have been developed.⁹ Formycin A inhibits the *de novo* purine synthesis¹⁰ and formycin B is a potent inhibitor of purine nucleoside phosphorylase in human erythrocytes.¹¹ Compound 2a has antineoplastic activity and inhibits growth of bacteria, fungi and viruses. Formycin B inhibits the growth of mouse sarcoma 180 cell and of influenza A1 virus. Some extremely toxic analogues have been synthesized.¹²

Dedicated to the memory of Professor R.K. Robins.



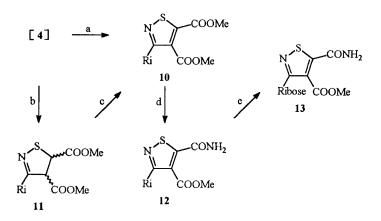




Scheme 1

Some time ago we studied the decomposition of the oxathiazolone 3 and the use of the D-allononitrile-N-sulphide 4 in the synthesis of a thiadiazole (5) and an isothiazole analogue (6) of ribavirin.¹³ Now we wish to report our work on the synthesis of 4-amino-3- β -D-ribofuranosyl-5-isothiazole carboxamide 7a, an analogue of pyrazofurin, and of the isothiazole[4,5-d]pyrimidines 8 and 9, analogues of the formycins (Scheme 1).

Thermolysis of the oxathiazolone 3 with dimethyl acetylenedicarboxylate at 140°C gave rise to low yields of the isothiazole diester 10 (Scheme 2). Side products were the corresponding D-allononitrile, sulphur and benzoic acid (elimination of the

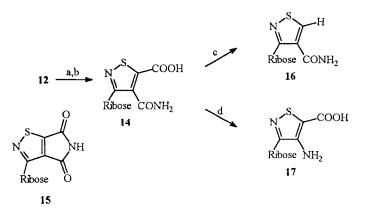


(a) DMAD, 140°C, 6 days (8%); (b) dimethyl fumarate, 200°C, 45 min (58%); (c) DDQ, chlorobenzene, Δ, 4.5h (85%); (d) CHCl₃/CH₃OH, 0°C, NH_{3gas}, 16 h (80%); (e) CHCl₃/CH₃OH, NaOH (79%).

Scheme 2

latter is a major fragmentation process in the mass spectra of benzoylated nucleosides). Better results were obtained with dimethyl fumarate as the dipolarophile. Upon heating of **3** in neat dimethyl fumarate at 200°C two isomeric isothiazolines **11a,b** were formed in about equal amounts. They were readily separated on a silica gel column with 5% ethyl acetate in toluene; they showed very similar mass, ¹H-NMR and ¹³C-NMR spectra. From the ¹H-NMR spectrum it appeared that both compounds had the expected trans configuration at the C₄-C₅ isothiazoline bond : the corresponding coupling constants were 4 Hz as in the analogous trans 3-phenyl-2-isothiazolinecarboxylate;¹⁴ the cis isoxazolines show a ³J_{4,5} of 11.5 Hz.¹⁵ Oxidation of either compound with DDQ (2,3-dicyano-5,6-dichloro-1,4-benzoquinone) gave the isothiazole dicarboxylate **10**. These data clearly indicate that both compounds were the 4R,5S- and 4S,5R-diastereomers; so, in further reactions the crude reaction mixture obtained after heating **3** in dimethyl fumarate was only subjected to flash chromatography to remove tarry material. After oxidation with DDQ in refluxing chlorobenzene and chromatography compound **10** was obtained in 60% overall yield starting from the oxathiazolone **3**.

Careful ammonolysis (monitored on TLC) of this diester 10 with a saturated solution of anhydrous ammonia in a 1:1 mixture of chloroform and methanol cooled in ice water, gave the monoamide 12 : of both ester functions only one, most probably the isothiazole-5-position, had reacted. This position is presumably more activated (by the



(a) CH_3OH/CH_3ONa , rt., 1.5 h (89%); (b) Dowex 50W-X2, H⁺; (c) 130°C; (d) (1) Ba(OH)₂, Br₂, 5-10°C, (2) H₂SO₄ (95%)

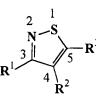
Scheme 3

conjugated ring nitrogen) than the 4-position (see for example the analogous regioselective nucleophilic reactions on isothiazole¹⁶ and pyrazole dicarboxylate esters 9a). Debenzoylation of 12 to free nucleoside 13 could be effected with a catalytic amount of sodium hydroxide in methanol-chloroform. Treatment of 12 with excess sodium methoxide followed by work-up with Dowex 50W-X2(H⁺) yielded a new compound which was assigned the 4-carbamoyl-3- β -D-ribofuranosyl-5-isothiazolecarboxylic acid structure 14 (Scheme 3). After decarboxylation by heating, a new compound was isolated and shown to be identical (TLC, spectra) with the previously obtained 3- β -D-ribofuranosyl-4-isothiazolecarboxamide 16.¹³

We believe that compound 14 is formed via a cyclic imide 15 as it is known that phtalic imide can be hydrolysed to phtalic acid amide by acid and by base.¹⁷ Ring opening of 15 should again occur by nucleophilic attack at the more electrophilic 5-position to give compound 14. The ease of decarboxylation of 14 is also in agreement with Adams and Slack's observations that isothiazole-4,5-dicarboxylic acid is decarboxylated at the 5-position. Removal of the second carboxyl group occurred much less readily.¹⁶

Hofmann rearrangement of the 4-carbamoyl function in 14 to the amine 17 was effected with barium hypobromite¹⁸ at 60°C. After neutralization with sulphuric acid, filtering of the barium sulphate and purification on Dowex 50W-X2(H⁺), the pure

TABLE 1.	¹³ C-NMR chemical shifts (ppm) for substituted isothiazole derivatives
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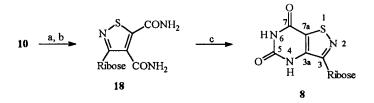
R ¹	R ²	R ³	C ₃	C ₄	C ₅
н	Н	Ha	157	123	150
Ribose	NH ₂	COOH(<u>17</u>)	159	147	126
Ribose	NH ₂	CONH ₂ (<u>7a</u>)	159	147	128
Methyl	COOMe	NH2 ^b	168	106	180

^a from ref. 19.

^b material kindly provided by Prof. Dr. J. Goerdeler

compound 17 was isolated as an oil in excellent yield. The ¹³C-NMR spectrum is in agreement with the proposed structure. The isothiazole-C absorptions are presented in Table 1, together with data for some related compounds. The C₄ atom is deshielded with respect to the parent isothiazole,¹⁹ whereas C₅ is shielded as expected from the substituent effects on the carbon absorptions in benzene²⁰ : the amino group induces an α -shift of +18 ppm and an ortho shift of -13 ppm whereas the carbomethoxy group gives an α -shift of +1.8 ppm and an ortho shift of +1 ppm. The spectrum recorded for methyl 5-amino-3-methyl-4-isothiazole-carboxylate (kindly provided by Prof.dr.J. Goerdeler) shows analogous effects of the 5-amino group clearly distinguishing between the 4- and the 5-amino-isothiazoles. (Table 1)

The diester 10 was also subjected to more vigorous ammonolysis conditions (overnight at room temperature). The resulting complex mixture of partially deblocked material was treated with excess sodium methoxide to remove the benzoyl protection completely. The mixture containing the diamide 18 (Scheme 4) and 20% of the monoamide 14, was subjected to Hofmann conditions (barium hypobromite) without

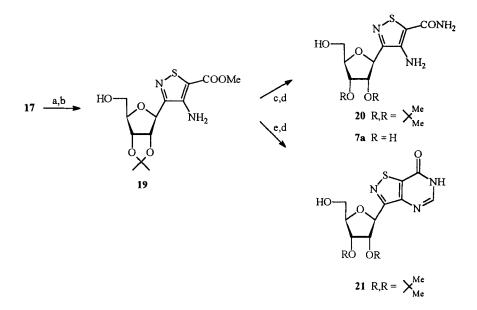


(a) $CHCl_3/CH_3OH$, NH_{3gas} , overnight, rt.; (b) $CH_3OH/NaOCH_3$, rt., 1.5h (73%); (c) (1) $Ba(OH)_2/Br_2$, 60°C, 1h, (2) H_2SO_4 (74%).

Scheme 4

prior purification. After work up the 3-B-D-ribofuranosyl-isothiazole[4,5-d]pyrimidine-5-(4H)-7(6H)-dione 8, an analogue of oxoformycin B, was crystallized from water in 74% yield. The structure of this compound could be proved by 13 C-NMR data : in DMSO-d₆ five quaternary carbon signals appear, C₃ at 158.2, C_{3a} at 140.1, C₅ at 151.4, C7 at 157.4 and C7a at 132.9 ppm. The proton coupled spectrum shows long range coupling between the sugar protons, C₃ and C_{3a}. The spectrum of a partially H/D exchanged sample shows apparent doublets at 140.1 and 157.4 ppm and an apparent triplet at 151.4 ppm, caused by an isotope effect from NH/ND on the neighbouring carbon atoms. This type of isotope effects on amides and peptides has been used by Feeney et al. to assign the amide carbon absorptions in oligopeptides.²¹ Thus the C_{3a} atom shows both a long range C-H coupling with the ribose protons and an isotope effect from the neighbouring NH group. The long range coupling and the isotope splitting excludes the isomeric [5,4-d] structure. This regioselectivity can be explained by the orientation effect of the isothiazole nucleus favouring the Hofmann rearrangement of the more electron rich amide function at position 4.2^2 The same type of selectivity was reported for 1-methyl-1,2,3-triazole-dicarboxamide and 3,4-pyrimidinedicarboxamide,²² although 3,4-isothiazoledicarboxamide did not react in a Hofmann rearrangement²³.

In order to get the 3- β -D-ribofuranosyl-4-amino-5-isothiazolecarboxamide 7a, compound 17 was treated with methanol and hydrogen chloride followed by reaction with dimethoxypropane and acid to yield the ester 19. This compound was then reacted with liquid ammonia in a sealed tube at 75 °C overnight, giving the protected amide 20 in 22% overall yield starting from compound 17. Due to the 4-amino function, ammonolysis of the 5-ester group in 19 occurred much less readily than with the dicarboxylate 10. The ¹H-NMR spectra of the ester 19 and the amide 20 showed a δ



(a) CH₃OH/HCl, 24 h, rt.; (b) CH₃C(OCH₃)₂CH₃, HCl/CH₃OH, 4 h (56%); (c) NH_{3liq}, 75°C, 8 h (39%); (d) diox./H₂O, Dowex 50W-X2 H⁺, 6 h, (87%); (e) ethyl formimidate, CH₃OH/NaOCH₃, 150°C, 2.5 h (61%).

Scheme 5

value for the isopropylidene methyl groups of 0.22 and 0.20 respectively²⁴ and a multiplet absorption for $H_{4'}$.²⁵ This is consistent with a β -configuration at $C_{1'}$. Finally the free nucleoside **7a** was obtained as an oil (87%) after deprotection with Dowex 50W-X2(H⁺).

Several attempts to get compounds of type 7b from compounds 17 and 19, the deprotected ester or its tri-O-(*tert*.-butyl dimethylsilyl)derivative, were not successful. By treatment with sodium nitrite at pH 4, in acetic acid or in hydrochloric acid^{26a} (with or without photolysis), or with nitrosyl tetrafluoroborate and acid,^{26b} either unprotected material, starting material or complex mixtures were found.

Of course the nucleoside 7a could be obtained from 17 without using the isopropylidene protection. We preferred however to use the protection in order to facilitate purification and to prove the β -configuration. Moreover, the protected ester 19 was further used in the synthesis of the formycin B analogue 9 : sodium methoxide catalysed cyclisation of 19 with ethyl formimidate^{27,28} in a sealed tube at 150°C gave the protected 3- β -D-ribofuranosyl-isothiazole[4,5-d]pyrimidine-(6H)-one 21 in 61% yield. Its ¹H-NMR spectrum showed a new singlet absorption at 8.1 ppm for H₅ (the

compd.	P _N	P _S	Ν _{τm}	s _{īm}	X N
1 ^b	41.1	141.9	42.0	42.4	0.32
7a	40.0	143.3	41.4	43.0	0.40
2a ^c	29.0	154.0	37.0	35.5	0.27
9	30.0	153.0	39.7	36.7	0.39

TABLE 2.Calculated a conformational parameters for the ribofuranose moiety of
Pyrazofurin 1, Formycin 2a and the analogues 7a and 9.

^a from ¹H NMR coupling constants, based on the graphs of ref.30.

The parameters P_N and P_S describe the phase angles of the N and S conformer in the pseudorotational model of ref.31. ${}^{N}\tau_m$ and ${}^{S}\tau_m$ are the ring pucker values and X_N is the molar fraction of N in the N,S-equilibrium.

^b based on the spectrum reported in ref.4c.

^c based on the spectrum reported in ref.32.

corresponding proton in quinazoline absorbs at 8.2 ppm.²⁹ Again the δ value for the isopropylidene methyls and the multiplicity for the H₄ · absorption confirmed the β -configuration. Deprotection with Dowex 50W-X2(H⁺) gave white crystals (89%) of the free nucleoside 9. The ¹³C-NMR showed five aromatic carbon atoms which could be assigned using the proton decoupled spectrum : C₃ has a multiplet absorption due to long range coupling with the ribose protons and C_{3a} shows a doublet of doublets (coupling with H₁ and H₅); C₅ obviously is a doublet, C₇ has a long range coupling with H₅ and C_{7a} is a singlet.

The conformational equilibrium for the ribofuranosyl moiety in the nucleoside analogues 7a and 9 was calculated from their 250 MHz ¹H-NMR spectra in D₂0 solution at room temperature. The first order constants $J_{H_{1'2'}}$, $J_{H_{2'3'}}$ and $J_{H_{3'4'}}$ were subjected to the graphical analysis developed by Davies and Danyluk³⁰ based on the pseudorotational model of Altona and Sundaralingam.³¹ The results are collected in Table 2. For comparison we have included the figures for pyrazofurin and formycin, calculated from the literature spectra.^{4c,32} From this table it is clear that the compound 7a is very similar to pyrazofurin as for phase angle and ring puckering of the ribose moiety. The N/S equilibrium shows a small shift towards the north conformer. The comparable conclusions apply to the isothiazolopyrimidine 9 with respect to formycin 2a.

PYRAZOFURIN AND FORMYCIN ANALOGUES

		· · · · · · · · · · · · · · · · · · ·	
compd.	%gg	%gt	%tg
1 ^b	60	40	00
7a	62	32	06
7a 2a ^c	67	25	08
9	62	30	08

TABLE 3.	Calculated a rotameric distribution at the C_4 '- C_5 ' bond in Pyrazofurin 1,
	Formycin $2a$ and the analogues $7a$ and 9 .

^a from ¹H NMR coupling constants, according to Haasnoot's method.

^b based on the reported spectrum in ref.4c.

^c based on the reported spectrum in ref.32.

We have also calculated the rotameric distribution at the $C_4 \cdot C_5 \cdot exocyclic$ bond using Haasnoot's method.³³ The AB part of the $H_{4'} \cdot H_{5'} \cdot H_{5''}$ ABX spectrum was subjected to second order analysis³⁴ to obtain the accurate coupling constants. The rotameric populations are presented in table 3 together with the parameters for pyrazofurin and formycin obtained from the literature spectra 5c.³² The similarity between the natural nucleosides and the analogues is again striking.

The biological activities of compounds 7a, 8, 9, 13 and 17 were tested. Notwithstanding the conformational similarities to pyrazofurin and formycin, no significant inhibition of either in vitro L1210 cell growth or of viral replication (HSV-1, measles, polio-1, VSV, vaccinia, reovirus-1, parainfluenzavirus-3 or cocksackievirus-B4) could be detected.

EXPERIMENTAL

Materials and Methods

Melting points, determined with a Leitz melting point microscope are uncorrected. The ¹H-NMR data are presented in ppm downfield from Me₄Si used as an internal standard; the spectra were taken on a Varian EM 390 spectrometer at 90MHz and a Bruker WM 250 at 250 MHz; for the ¹³C-NMR spectra a Bruker WM 250 and a Bruker WP 80 spectrometer were used. The mass spectra were recorded on a Kratos-MS-907-S apparatus with direct insertion and an ionization energy of 70 eV. Infrared spectra were obtained from a Perkin-Elmer 250 grating apparatus. For the chromatographic separations Merck silicagel 60 (0.063-0.200 mm) was used unless otherwise stated.

<u>4-amino-3-β-D-ribofuranosyl-5-isothiazolecarboxamide</u> 7a. A solution of 20 (79.5 mg, 0.25 mmol) in dioxane (2 ml) was stirred with dowex 50W-X2 (H⁺) for 6 h at room temperature. After filtration and evaporation the residue was lyophilized to give 62 mg of 7a as a slightly coloured powder (87%). IR (KBr), 3500 cm⁻¹; ¹H-NMR (D₂O) δ 4.93 (d, J=6.4 Hz, H₁·), 4.45 (dxd, J=6.4 Hz, J=5.5 Hz, H₂·), 4.22 (dxd, J=5.5 Hz, J=4.6 Hz, H₃·), 4.10 (m, H₄·,) 3.83 (dxd, J=3 Hz,J=-12.1 Hz, H₅·) 3.75 (dxd, J=4.4 Hz, J=-12.1 Hz, H₅··); ¹³C-NMR (D₂O) δ 166.4 (CONH₂), 159.1 (C₃), 146.8 (C₄), 127.8 (C₅), 86.1, 81.5, 74.4, 72.1, 62.5 (ribose C); MS (+5 Me₃Si), m/z 635, 620, 545, 502, 440, 230, 217, 103, 72.

<u>3-β-D-ribofuranosyl-isothiazolo[4,5-d]pyrimidine-5(4H)-7(6H)-dione</u> **8**. A mixture (501 mg) of diamide **18** and acid **14** was dissolved in a barium hypobromite solution (prepared as mentioned for the synthesis of **17**) and was heated at 60°C for 1 h. After cooling the solution was acidified with sulphuric acid (3 N) and filtered over celite. The filter was washed with water (100 ml) and filtrate and washings were brought to pH 9 with K₂CO₃. After evaporation the mixture was dissolved in water (10 ml) and purified on a dowex 50W-X2 column with water, giving 295 mg of **8** (74%) as white crystals; mp 225-227°C; ¹H-NMR (dmso-d₆) δ 11.6 (s_{br}, NH), 11.4 (s_{br}, NH), 4.94 (d, J=8 Hz, H₁'), 4.14 (dxd, J=8 Hz, J=6 Hz, H₂'), 4.05 (m, H₃', H₄'), 3.67 (d, J=1 Hz, H₅', H₅''); ¹³C-NMR (dmso-d₆) δ 158.2 (C₃), 157.4 (C₇), 151.4 (C₅), 140.1 (C_{3a}), 132.9 (C_{7a}), 86.1, 82.0, 81.6, 74.6, 72.2, 61.4 (ribose C); MS (+ 3 Me₃Si) m/z 517, 502, 474, 446, 427, 414, 356, 337, 296, 268, 217, 103, 73.

<u>3-β-D-ribofuranosyl-isothiazolo[4,5-d]pyrimidine-7(6H)-one</u> **9**. The isopropylidene derivative **21** (46.1 mg, 0.14 mmol) was dissolved in water (2 ml) and stirred with Dowex 50W-X2 (H+) for 6.5 h at room temperature. After filtration, washing and evaporation of the filtrates and washings, compound **9** was obtained. Recrystallization from water gave 36 mg (89%) of white crystals; mp 132-134°C; ¹H-NMR (D₂O) δ 8.29 (s, H₅), 5.33 (d, J=6.0 Hz, H₁·), 4.61 (dxd, J=6.0 Hz, J=5.8 Hz, H₂·), 4.46 (dxd, J=5.8 Hz, J=3.9 Hz, H₃·), 4.35 (m, H₄·), 3.92 (dxd, J=3.1 Hz, J=-12.8 Hz, H₅·), 3.83 (dxd, J=4.3 Hz, J=-12.8 Hz, H₅··); ¹³C-NMR (D₂O, assigned using the 1H coupled spectrum) δ 164.3 (C₃), 158.8 (C₇), 151.0 (C_{3a}), 148.5 (C₅), 142.9 (C_{7a}), 86.2, 81.0, 75.6, 72.6, 62.9, (ribose C); MS (+ 3 Me₃Si), m/z 501, 486, 411, 398,

340, 280, 259, 258, 254, 252, 230, 217, 182, 103, 73; Anal. Calcd for C₁₀H₁₁O₅N₃S. ¹/₂ H₂O: C, 40.85; H, 3.97; N, 14.19. Found: C, 40.81; H, 4.09; N, 14.28.

<u>Dimethyl 3-(2,3,5-tri-*O*-benzoyl-*B*-D-ribofuranosyl)-4,5-isothiazoledicarboxylate **10**.</u>

<u>Method A</u> (with dimethyl acetylenedicarboxylate). Oxathiazolone 3^{13} (0.547 g, 1 mmol) was suspended in dimethyl acetylenedicarboxylate (1 ml, 8.1 mmol) and heated at 140°C under argon for 6 days. Excess dipolarophile was removed under reduced pressure and the residue was separated on a preparative silica gel plate (eluent : 10% ethyl acetate in benzene) to give 51 mg (8%) of the isothiazole diester **10** as a slightly yellow oil. This compound was identical (¹H-NMR, mass spectrum and TLC characteristics) with the material obtained in method B.

Method B (with dimethyl fumarate and oxidation with DDO). Oxathiazolone 3 (10.0 g, 18.2 mmol) was added portionwise to a stirred melt of dimethyl fumarate (40 g, twice recrystallized from cyclohexane) heated in an oil bath at 130°C. After complete dissolution the bath temperature was raised to 200°C; the mixture was then refluxed for 1 h and cooled. After removal of excess fumarate under reduced pressure (15 torr, 100°C), the residue was purified by flash chromatography on a silica gel column with 12% ethyl acetate in toluene. The solvent was then evaporated and the residue was dissolved in chlorobenzene (100 ml). After addition of DDQ (4.0 g, 17.6 mmol), the solution was refluxed for 3.5 h (bath temperature 140°C). The solvent was then removed under vacuum and the residue was subjected to flash chromatography over a silica gel column with 20% ethyl acetate in toluene. The solution was concentrated under reduced pressure and chromatographed on a silica gel column (0.040-0.063 mm with 1 tot 10% ethyl acetate in toluene). The fractions containing compound 10 were pooled and evaporated to give 7.28 g (62%) of the diester as a hygroscopic non crystallizable oil: ¹H-NMR (CDCl₃) δ 8.1-7.8 (m, ArH), 7.6-7.2 (m, ArH), 6.32 (dxd, J=3.4 Hz, J=4.9 Hz, $H_{2'}$), 5.92 (t, J=4.9 Hz, $H_{3'}$), 5.65 (d, J=3.4 Hz, $H_{1'}$), 4.9-4.4 (m, H_{4'}, H_{5'}, H_{5''}), 3.90 (s, CH₃OOC); MS m/z 645, 614, 523, 445, 401, 201, 105 : exact mass calcd. for $C_{33}H_{27}N0_{11}S$: 645.130, found 645.131 \pm 0.001.

Dimethyl 4R, 5S and 4S, 5R-3-(2,3,5-tri-O-benzoyl-ß-D-ribofuranosyl)-isothiazoline-4,5-dicarboxylates 11a,b. Oxathiazolone 3 (5.58 g, 10.2 mmol) and dimethyl fumarate (21.5 g twice recrystallized from cyclohexane) were reacted as above. After work-up the residue was chromatographed on a silica gel column (0.040-0.063 mm) with 5% ethyl acetate in toluene. After evaporation of the homogeneous fractions 1.41 g (21.5%) of isomer A and 2.41 g (37%) of isomer B were obtained as oils. Isomer A : ¹H-NMR (CDCl₃) δ 8.15-7.82 (m, ArH), 7.6-7.1 (m, ArH), 6.05 (dxd, J=3.5 Hz; J=5.5 Hz, H₂), 5.76 (t, J=5.5 Hz, H₃), 5.20 (d, J=3.5 Hz, H₁) 4.86 and 4.79 (2xd, J=5.2 Hz), 4.7 (m, H_{4'}, H_{5'}, H_{5''}), 3.7 (s, CH₃OOC); ¹³C-NMR (CDCl₃, assigned using the ¹H-coupled spectrum and via decoupling at 3.7 ppm in the proton area) δ 170.2, 167.8 (COOCH₃), 166.0 (C₃), 165.2, 165.0, 162.0 (COOPh), 133.3-128.2 (ArC), 81.0 (C₁), 79.3 (C_{4'}), 74.0 (C_{2'}), 71.9 (C_{3'}), 63.6 (C_{5'}), 60.4 and 52.6 (C₄ and C₅), 53.0 (q, CH₃OOC); MS m/z 647, 645, 616, 614, 588, 525, 523, 445, 403, 401, 105. Isomer B : ¹H-NMR (CDCl₃) δ 8.2-7.8 (m, ArH), 7.5-7.2 (m, ArH), 6.0 (dxd, J=4 Hz; J=5 Hz, H_{2'}), 5.83 (t, J=5 Hz), H_{3'}), 5.27 (d, J=4 Hz, H_{1'}), 5.03 (d, J=4.5 Hz, H₄ or H₅), 4.74 (d, J=4.5 Hz, H₅ or H₄), 4.9-4.5 (m, H_{4'}, H_{5'}, H_{5''}), 3.73 (s, CH₃OOC), 3.69 (s, CH₃OOC); ¹³C-NMR (CDCl₃, assigned via the proton coupled spectrum and via decoupling at 3.7 ppm in the proton area) 170.2, 168.2 (2xs, COOCH₃), 166.2 (s, C₃), 165.3, 165.3, 162.8 (s, COOPh), 133.5-128.5 (ArC), 80.5 (C_{1'}), 80.0 (C_{4'}), 74.2 (C_{2'}), 72.7 (C_{3'}), 64.2 (C_{5'}), 59.5, 52.4 (C₄, C₅), 53.2 (CH₃OOC); MS m/z 647, 645, 616, 614, 588, 525, 523, 445, 403, 401, 105.

When either compound was oxidized with DDQ the isothiazole 10 was obtained in 86% and 85% yield.

Methyl-5-carbamoyl-3-(2,3,5-tri-*O*-benzoyl-β-D-ribofuranosyl)-isothiazole-4carboxylate 12. The diester 10 (13.36 g, 20.7 mmol) was dissolved in a mixture of CHCl₃ (150 ml) and CH₃OH (150 ml), cooled in an ice bath and saturated with ammonia. After 2 h at 0°C the solvent was removed under reduced pressure and the residue was chromatographed over silica gel with 10 to 50% ethyl acetate in toluene. The fractions containing the amide 12 were evaporated, leaving 10.53 g of an amorphous compound (80.2%) : ¹H-NMR (CDCl₃) δ 8.49 (s_{br}, NH), 8.1-7.8 (m, ArH), 7.6-7.2 (m, ArH), 6.54 (s_{br}, NH), 6.33 (dxd, J=2.5 Hz, J=5 Hz, H₂·), 5.92 (dxd, J=5 Hz, J=7.0 Hz, H₃·), 5.76 (d,J=2.5 Hz, H₁.), 4.7 (m, H₄·, H₅·, H₅··), 3.93 (s, CH₃OOC); MS m/z 630, 599, 508, 495, 445, 105. Anal. Calcd. for C₃₂H₂₆N₂0₁₀S : C, 60.95; H, 4.16; N, 4.44; S, 5.08. Found : C, 61.30; H, 4.14, N, 4.33; S, 5.08.

<u>Methyl-5-carbamoyl-3-β-D-ribofuranosyl-isothiazole-4-carboxylate</u> **13**. The protected monoamide **12** (0.15 g; 0.24 mmol) was dissolved in CHCl₃ (2 ml) and NaOH (0.15 mmol) in CH₃OH (2 ml) was added. The mixture was kept at room temperature for 20 min and was then neutralized with Dowex 50 W-X2 (H⁺), filtered and evaporated under reduced pressure. The residue was dissolved in water and extracted with CHCl₃, the water layer was concentrated to give 60 mg (79%) of a clear sirup ; IR (KBr) 3300 (OH, NH₂), 1710 (COOCH₃), 1660, 1590 cm⁻¹ (CONH₂); 1H-NMR (D₂0) δ 5.42 (d, J=3.5 Hz, H₁·), 4.41 (dxd, J=3.5 Hz, J=4.2 Hz, H₂·), 4.22 (t, J=4.2 Hz, H₃·), 4.17 (m, H₄·), 3.95 (s, CH₃OOC), 3.8 (m, H₅·, H₅··); ¹³C-NMR (D₂0), 169.1, 167.2, 164.7, 164.5 (CONH₂, COOCH₃, C₃, C₅), 127.8 (C₄) 84.7 (C₁·), 82.4 (C₄·), 76.2, 71.6 (C₂·, C₃·), 62.7 (C₅·), 54.2 (CH₃OOC); MS (+4 Me₃Si), m/z 606, 591, 503, 217, 215, 73.

<u>4-carbamoyl-3-ß-D-ribofuranosyl-isothiazole-5-carboxylic</u> acid <u>14.</u> The protected monoamide <u>12</u> (10.48 g, 16.6 mmol) was treated with 200 ml absolute methanol containing sodium methoxide (from 2.21 g Na, 96 mmol) at room temperature. The reaction was monitored by TLC (cellulose, 20% 0.2 M NH₄OAc - 80% Me₂CO) and was complete after 75 min. The mixture was treated with Dowex 50 W-X2 (H⁺)-which gave an acid solution - and concentrated under vacuum.

The residue was dissolved in water, extracted with CHCl₃ (3 x 50 ml) decolorized with active carbon, filtered over celite and evaporated to give the acid 14 as an oil (4.50 g, 89%). The compound was crystallized from water, mp 103-107°C (dec at 112°C); IR (KBr) 3300 (OH, NH₂), 1710 (COOH), 1670, 1615 cm⁻¹ (CONH₂); ¹H-NMR (D₂0), δ 5.15 (d, J=6 Hz, H₁·), 4.45 (t, J=6 Hz, H₂·), 4.25 (t, J=6 Hz, H₃·), 4.1 (m, H₄·), 3.8 (m, H₅·, H₅··); MS (+5 Me₃Si) m/z 664, 649, 574, 533, 484, 471, 299, 217, 73.

When this compound was heated at 130° C, a product identical (TLC, ¹H-NMR and mass spectrum) with 3-B-D-ribofuranosylisothiazole-4-carboxamide 16^{13} was obtained.

4-Amino-3-B-ribofuranosyl-isothiazole-5-carboxylic acid 17. Ba(OH)₂.8H₂0 (4.36 g, 13.8 mmol) was dissolved in demineralized water (100 ml) at 60°C; insoluble barium carbonate was removed by filtration. To the cooled (5-10°C) filtrate bromine (0.53 g, 3.3 mmol) was added. The mixture was stirred until all bromine was dissolved and the amide 14 (760 mg, 2.5 mmol) was then added. After 45 min at 60°C all hypobromite had disappeared (KI- H_2SO_4 test). The amide was converted to a single new compound (TLC on cellulose, 30% 0.2 M NH₄OAc-(CH₂)₂CO). The cooled solution was neutralized with 3 N sulphuric acid and the precipitate was removed by cellite filtration. The filtrate was concentrated under reduced pressure to 25 ml and passed through a Dowex column 50W-X4 (200-400 mesh). First a strongly acidic fraction was washed off with water and the amine was eluted with 1 N ammonia. The fractions containing the amine 17 were evaporated and lyophilized to give 655 mg (95%) of an oily compound : IR (KBr) 3300, 1690, 1610 cm⁻¹; ¹H-NMR (D_20) δ 4.95 (d, J=6 Hz, H₁), 4.42 (t, J=4 Hz, H₂), 4.15 (m, H₃, H₄), 3.8 (m, H₅, H₅); ¹³C-NMR (D₂0) δ 165.3 (COOH), 158.9 (C₃), 147.0 (C₄), 125.7 (C₅), 85.9, 81.9, 74.5, 72.1, 62.4 (ribose C); MS (+ 4 Me₃Si) m/z 564, 549, 474, 432, 384, 371, 331, 230, 217, 73.

<u>3-B-D-ribofuranosyl-isothiazole-4,5-dicarboxamide</u> 18. A solution of diester 10 (4.53 g, 7 mmol) in a mixture of methanol (50 ml) and chloroform (50 ml) was saturated with NH_3 gas at 0°C, and kept overnight at room temperature. The solvent was evaporated and the residue was treated with NaOCH₃ in methanol (from 1.00 g

NaH in 30 ml methanol). After 90 min at room temperature the mixture was neutralized with Dowex 50W-X2 (H+) and evaporated. The residue was dissolved in water (50 ml) and extracted with chloroform (3x50 ml). The water layer was lyophilized to give 2.01 g of a 4 to 1 mixture (according to ¹H-NMR) of the diamide **18** and the acid **14**; ¹H-NMR (D₂0) δ 5.10 (d, 5.4 Hz, H₁·), 4.50 (t, 5.4 Hz, H₂·), 4.22 (dxd, J=5.4 Hz, J=5.1 Hz, H₃·), 4.07 (m, H₄·), 3.8 (m, H₅·, H₅··); chemical ionisation MS (+5 Me₃Si), m/z 664 (MH⁺).

Methyl 4-amino-3-(2', 3'-O-isopropylidene)- β -D-ribofuranosyl-isothiazole-5-carboxylate 19. Aminoacid 17 (127 mg, 0.646 mmol) was treated with a saturated solution of HCl gas in dry methanol at room temperature for several days. Then the solvent was removed in vacuo and the residue was dissolved in dimethoxypropane - HCl (0.8 M, 4 ml) and 1 ml of dry methanol was added. After 4 h the mixture was neutralised with 5% aqueous NaHCO₃ and extracted with chloroform. The organic layer was dried on MgSO₄ and evaporated and the residue was purified on a preparative silica gel plate to give 85 mg (56%) of the ester 19 as a colourless oil; ¹H-NMR (CDCl₃) δ 5.85 (s_{br}, NH₂), 5.0 (m, H₁', H₂'), 4.76 (dxd, J=3 Hz, J=6 Hz, H₃'), 4.22 (m, H₄'), 3.87 (s, CH₃OOC), 3.75 (m, H₅',H₅'', OH), 1.60 (s, CH₃), 1.38 (s, CH₃); MS m/z 330, 315, 300, 299, 272, 242, 187, 59, 42.

<u>4-Amino-3-(2',3'-O-isopropylidene)- β -D-ribofuranosyl-isothiazole-5-carbox-</u> amide 20. Compound 19 (106 mg, 0.32 mmol) was dissolved in liquid ammonia in a sealed tube and heated in a bomb for 8 h at 75°C. The ammonia was slowly evaporated and the residue was separated on a preparative silica gel plate (with 10% methanol in chloroform) to give 40 mg of the amide 20 (39%). Another fraction contained 29.6 mg (28%) of the starting material; IR (oil film) 3500-3150 (OH, NH₂CO, NH₂), 1660, 1600 cm⁻¹ (CONH₂); ¹H NMR (CDCl₃) δ 5.93, 5.83 (2 x s_{br}, NH₂ and CONH₂), 5.1 (m, H₁', H₂'), 4.80 (dxd, J=2.5 Hz, J=4.5 Hz, H₃'), 4.30 (m, H₄',), 3.9 - 3.4 (m, H_{5'}, H_{5''}, OH), 1.60 (s, CH₃), 1.40 (s, CH₃); mass spectrum, m/z 315, 300, 297, 285, 257, 59, 43.

<u>3-(2',3'-O-isopropylidene)-B-D-ribofuranosyl-isothiazolo[4,5-d]-pyrimidine-</u> <u>7(6H)-one</u> **21**. A mixture of compound **19** (88 mg, 0.26 mmol), ethyl formimidate (4 ml) and sodium methoxide in methanol (from 0.8 mg NaH in 0.2 ml methanol) was heated at 150°C in a sealed tube for 2.5 h. The reaction mixture was evaporated and purified on a preparative silica gel plate (5% methanol in chloroform) to give 52 mg of **21** as an oil (61%): ¹H NMR (CDCl₃ + CD₃OD) δ 8.1 (s, H₅), 5.46 (d, J=6 Hz, H₁·), 5.04 (m, H₂·, H₃·), 4.47 (m, H₄·), 4.3 (s_{br}, OH and NH), 3.85 (m, H₅·, H₅··), 1.70 (s, CH₃) 1.41 (s, CH₃); MS m/z 325, 310, 307, 292, 267, 237, 182, 151.

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