# Journal of Medicinal Chemistry



Article

Subscriber access provided by University of Winnipeg Library

# Design and Optimization of Sulfone Pyrrolidine Sulfonamide Antagonists of Transient Receptor Potential Vanilloid-4 (TRPV4) with In Vivo Activity in a Pulmonary Edema Model

Joseph E. Pero, Jay Matthews, David J Behm, Edward Brnardic, Carl A Brooks, Brian W Budzik, Melissa Costell, Carla Donatelli, Stephen Eisennagel, Karl F Erhard, Michael Fischer, Dennis A. Holt, Larry Jolivette, Huijie Li, Peng Li, John McAtee, Brent W McCleland, Israil Pendrak, Lorraine Posobiec, Katrina Rivera, Ralph A. Rivero, Theresa J Roethke, Matthew Sender, Arthur Shu, Lamont Terrell, Kalindi Vaidya, Xiaoping Xu, and Brian G. Lawhorn

J. Med. Chem., Just Accepted Manuscript • DOI: 10.1021/acs.jmedchem.8b01344 • Publication Date (Web): 30 Nov 2018

Downloaded from http://pubs.acs.org on December 1, 2018

# **Just Accepted**

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

# Design and Optimization of Sulfone Pyrrolidine Sulfonamide Antagonists of Transient Receptor Potential Vanilloid-4 (TRPV4) with In Vivo Activity in a Pulmonary Edema Model

Joseph E. Pero<sup>\*, \operatornambda</sup>, Jay M. Matthews<sup>†</sup>, David J. Behm<sup>†</sup>, Edward J. Brnardic<sup>†</sup>, Carl Brooks<sup>†</sup>, Brian W. Budzik<sup>\operatornambda</sup>, Melissa H. Costell<sup>†</sup>, Carla A. Donatelli<sup>†</sup>, Stephen H. Eisennagel<sup>†</sup>, Karl Erhard<sup>\operatornambda</sup>, Michael C. Fischer<sup>I</sup>, Dennis A. Holt<sup>†</sup>, Larry J. Jolivette<sup>†</sup>, Huijie Li<sup>\operatornambda</sup>, Peng Li<sup>\operatornambda</sup>, John J. McAtee<sup>†</sup>, Brent W. McCleland<sup>\operatornambda</sup>, Israil Pendrak<sup>\operatornambda</sup>, Lorraine M. Posobiec<sup>I</sup>, Katrina L.K. Rivera<sup>†</sup>, Ralph A. Rivero<sup>\operatornambda</sup>, Theresa J. Roethke<sup>†</sup>, Matthew R. Sender<sup>\operatornambda</sup>, Arthur Shu<sup>\operatornambda</sup>, Lamont R. Terrell<sup>†</sup>, Kalindi Vaidya<sup>I</sup>, Xiaoping Xu<sup>†</sup>, Brian G. Lawhorn<sup>†</sup>

<sup>¢</sup>Flexible Discovery Unit, <sup>†</sup>Heart Failure Discovery Performance Unit and <sup>1</sup>Platform Technology and

Sciences

GlaxoSmithKline, 1250 Collegeville Road, Collegeville, Pennsylvania, 19426, United States

### **ABSTRACT:**

Pulmonary edema is a common ailment of heart failure patients and has remained an unmet medical need due to dose-limiting side effects associated with current treatments. Preclinical studies in rodents have suggested that inhibition of TRPV4 cation channels may offer an alternative - and potentially superior - therapy. Efforts directed toward small molecule antagonists of the TRPV4 receptor have led to the discovery of a novel sulfone pyrrolidine sulfonamide chemotype exemplified by lead compound **6**. Design elements toward the optimization of TRPV4 activity, selectivity and pharmacokinetic properties are

described. Activity of leading exemplars **19** and **27** in an *in vivo* model suggestive of therapeutic potential is highlighted herein.

Despite numerous medical advancements and increased awareness of risk factors, heart disease remains one of the leading causes of death.<sup>1</sup> Specifically, approximately 2% of adults in developed countries have experienced congestive heart failure (HF), with this number rising to 6-10% for those exceeding 65 years of age.<sup>2</sup> Over a 12-month period post-diagnosis, the risk of death is approximately 35%, which is comparable to that of severe cancers.<sup>3</sup> HF and its related side effects also pose a significant economic burden on society, with total associated costs in the United States alone projected to increase from \$31 billion in 2012 to \$70 billion in 2030.<sup>4</sup>

HF is caused by the inability of an impaired left ventricle to pump sufficient blood into peripheral circulation. This results in elevated pulmonary venous pressure (PVP) and disrupts the septal barrier, which separates the circulatory aqueous environment from the alveolar airspaces.<sup>5,6</sup> With an increased flow of fluid from pulmonary circulation into the alveolar space, HF patients frequently suffer from pulmonary edema, which significantly compromises their quality of life and can prove fatal if left untreated.<sup>7</sup>

TRPV4 receptors are members of the Transient Receptor Potential (TRP) superfamily of cation channels.<sup>8</sup> These tetrameric voltage-gated channels are highly expressed in lung endothelial cells<sup>9,10</sup> and are up-regulated in HF patients.<sup>11</sup> Their direct association with pulmonary edema has been implicated by studying the effects of selective TRPV4 antagonist GSK2193874 (Compound 1, Figure 1a) in rodents.<sup>11</sup> In both acute and chronic preclinical models, pre-treatment with GSK2193874 inhibited HF-induced pulmonary edema and increased arterial oxygenation.<sup>11</sup> In addition, GSK2193874 was effective at reversing HF-induced pulmonary edema in mice subjected to myocardial infarction (MI) for one week prior to treatment.<sup>11</sup>

#### Journal of Medicinal Chemistry

Importantly, strong evidence indicates that these effects may be relevant to humans. GSK2193874 was found to be potent in recombinant hTRVP4 channels ( $IC_{50} = 50$  nM), and prevented agonist-driven cellular contraction in human umbilical vein endothelial cells (HUVEC's).<sup>11</sup>

These experimental findings suggest that TRPV4 blockade has the potential to lead to a novel treatment for HF-induced pulmonary edema that could prove more effective than current standards-ofcare (i.e. diuretics, nitroglycerin, angiotensin antagonists), which have been associated with unfortunate side effects including hypotension and deteriorating renal function.<sup>12,13</sup> As such, a number of selective TRPV4 antagonists have been disclosed in the recent literature with superior drug-like properties (i.e. reduced molecular weight, lower lipophilicity) to GSK2193874. Examples from GlaxoSmithKline include 1-(4-piperidinyl)-benzimidazoles<sup>14</sup> and spirocarbamates (**2**, Figure 1b),<sup>15,16</sup> while Pfizer has disclosed azetidine sulfonamides (**3**, Figure 1c) derived from a high-throughput screen.<sup>17</sup>

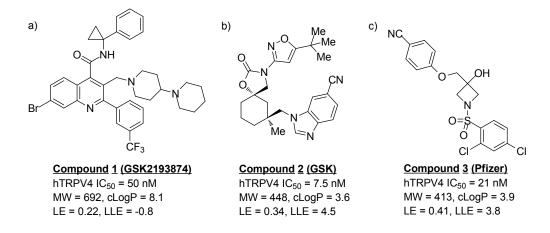


Figure 1. Previously-reported TRPV4 antagonists from GSK (a, b) and Pfizer (c).

With its promising ligand efficiency  $(LE)^{18}$  and lipophilic ligand efficiency  $(LLE)^{19}$ , **3** was previously utilized as a starting point in efforts directed toward discovering novel TRPV4 antagonists.<sup>20</sup> This led to the discovery of pyrrolidine sulfonamide lead **4**, which displayed encouraging *in vitro* activity  $(IC_{50} = 32 \text{ nM})$  at recombinant hTRPV4 channels<sup>21</sup> with comparable LE (0.40) and LLE (3.9) relative to **3** (Figure 2). Subsequent scaffold optimization<sup>20</sup> led to diol **5**, which featured excellent LLE (5.3) due to enhanced TRPV4 activity (IC<sub>50</sub> = 5.0 nM) in conjunction with reduced lipophilicity ( $cLogP^{22} = 3.0$ ).

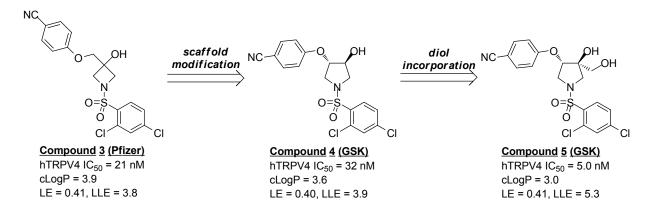


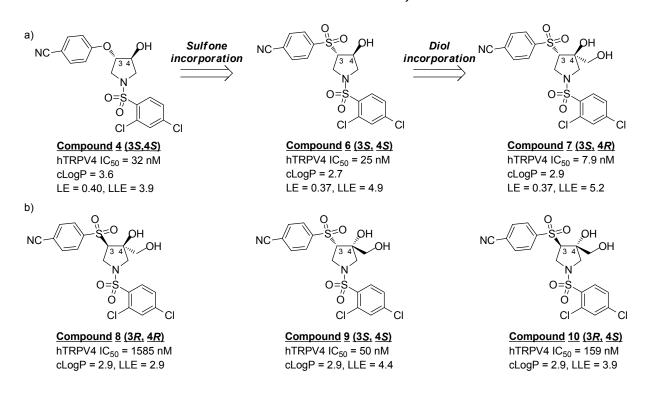
Figure 2. Scaffold modification of 3 to give pyrrolidines 4 and 5.

The present work describes an analogous structural class exemplified by sulfone **6**, which was viewed as a lead compound superior to ether **4** due to reduced lipophilicity (cLogP = 2.7) and improved LLE (4.9) (Figure 3a). Design elements toward an optimal balance of TRPV4 activity, selectivity and pharmacokinetic (PK) properties are detailed. Evaluation of leading compounds in an *in vivo* rat pulmonary edema assay indicative of therapeutic potential is also provided.

#### ■ RESULTS AND DISCUSSION:

At the outset of our optimization efforts, the potency-enhancing diol of the ether structural class<sup>20</sup> was incorporated to afford Compound 7, which displayed improvements in both TRPV4 activity (IC<sub>50</sub> = 7.9 nM) and LLE (5.2) with only a slight increase in lipophilicity (cLogP = 2.9). Notably, the stereoisomers of 7 were also explored (Figure 3b, Compounds 8-10), with each exhibiting considerably weaker activity in the primary FLIPR<sup>21</sup> assay.

#### Journal of Medicinal Chemistry



**Figure 3.** (a) Replacement of phenoxy ether functionality with sulfone, leading to 7. (b) Additional stereoisomers of sulfone pyrrolidine diol 7.

An investigation of the PK properties of **7** further substantiated the diol chemotype (Table 1). Following intravenous (i.v.) dosing in rat (1 mpk), **7** exhibited reduced clearance adjusted for unbound fraction<sup>23</sup> (CL/ $f_u = 9$ ) relative to parent compound **6** (CL/ $f_u = 30$ ), suggesting that the potency-enhancing diol functionality did not pose an additional metabolic liability. Furthermore, high permeability<sup>24</sup> (450 nm/sec) and 100% oral bioavailability (F, 2 mpk p.o.) were maintained despite an elevated total polar surface area (PSA = 136). This could be rationalized by a reduction in solvent-accessible PSA due to conformational restriction imparted by intramolecular hydrogen bonds, which were confirmed by vibrational circular dichroism (VCD) studies.<sup>25,26,27</sup>. In addition, despite elevated total clearance (25 mL/min/kg), the mean residence time<sup>28</sup> of **7** (MRT = 1.7 h) was nearly equivalent to that of **6** due to its higher volume of distribution (Vdss = 2.3 L/kg).

**Table 1.** PK properties (rat) of Compounds 5, 6 and  $7^a$ 

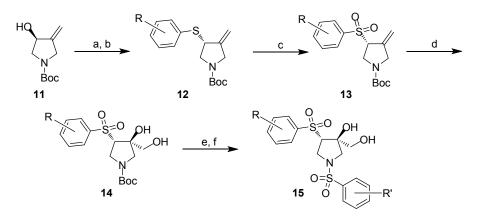
Cmpd	PSA	Perm	CL	$CL/f_u$	Vdss	MRT	F
		(nm/sec)	(mL/min/kg)		(L/kg)	(h)	(%)
5	111	370	74	15	2.4	0.6	46
6	116	780	9	30	1.1	2.0	100
7	136	450	25	9	2.3	1.7	100

<sup>*a*</sup>PSA = total polar surface area; Perm = artificial membrane permeability; CL = total clearance;  $f_u =$  fraction unbound (rat); Vdss = volume of distribution; MRT = mean resonance time; F = oral bioavailability

A comparison of the PK properties of 7 with corresponding ether 5 provided additional validation of the sulfone structural class. Ether 5 featured higher total clearance (74 mL/min/kg) and clearance adjusted for unbound fraction ( $CL/f_u = 15$ ) as well as a shorter MRT (0.6 h), potentially due to oxidative metabolism of the phenoxy ether moiety. Furthermore, ether 5 exhibited reduced oral bioavailability (46%) in relation to sulfone 7.

In order to facilitate analog preparation, a generalized<sup>29</sup> synthetic route toward the sulfonefunctionalized pyrrolidine diols was established (Scheme 1). Chiral resolution of known *tert*-butyl-3hydroxy-4-methylenepyrrolidine-1-carboxylate<sup>30</sup> provided the requisite (*S*)-isomer **11**. Formation of the corresponding mesylate followed by  $S_N$ 2-displacement with an aryl thiol under basic conditions afforded thioether **12**. Oxidation with *m*-CPBA generated sulfone **13** which then underwent a dihydroxylation to give diol **14** in high diastereomeric fidelity. Establishing the sulfone moiety prior to treatment with osmium tetroxide proved critical, as dihydroxylation of thioether **12** proceeded with significantly lower diastereoselectivity.<sup>31</sup> With advanced intermediate **14** in hand, acid-mediated cleavage of the Boccarbamate followed by sulfonamide formation under standard conditions generated targeted compounds of generic structure **15**.

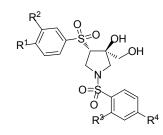
#### Scheme 1. General synthesis of sulfone pyrrolidine diols<sup>a</sup>



<sup>*a*</sup>Reagents and conditions: (a) MsCl, TEA, DCM; (b) arylthiol, K<sub>2</sub>CO<sub>3</sub>, DMF (74-81% over 2 steps); (c) *m*-CPBA, DCM (59-89%); (d) OsO<sub>4</sub>, NMO, THF (81-98%, >95:5 dr); (e) HCl/1,4-dioxane or TFA, DCM; (f) arylsulfonyl chloride, NaHCO<sub>3</sub>, THF (26-85% over 2 steps).

With a superior balance of potency and PK properties being the ultimate goal, the versatility of this synthetic sequence allowed the structure-activity relationships of both the aryl sulfonamide and sulfone moieties to be explored (Table 2). Compound **16** features the installation of a fluoro substituent at R<sup>2</sup> that proved to be a potency enhancer in the aforementioned azetidine<sup>17</sup> and phenoxy ether pyrrolidine<sup>20</sup> structural classes. While Compound **16** indeed demonstrated slight improvements in TRPV4 activity (IC<sub>50</sub> = 2.0 nM) and LLE (5.6) relative to its des-fluoro counterpart **7**, these benefits were counterbalanced by an erosion in PK properties as exemplified by higher clearance adjusted for unbound fraction (CL/f<sub>u</sub> = 14), shorter MRT (0.8 h) and reduced oral bioavailability (51%). One possible rationalization of the higher unbound clearance associated with **16** is that its more electrophilic aryl sulfone functionality was subjected to increased nucleophilic addition by glutathione (GSH), which was suggested by trapping experiments in rat liver microsomes in which only 46% of parent remained after incubation (30 min). Analogous studies with compound **7** showed 89% of parent remaining after incubation.

**Table 2.** Structure-activity relationships of sulfone pyrrolidine diols<sup>a</sup>



Cmp	pd R <sup>1</sup>		R <sup>2</sup>	R <sup>3</sup>	R <sup>4</sup>	IC <sub>50</sub>	LLE	PSA	CL	$CL/f_u$	Vdss	MRT	F
						(nM)			(mL/min/kg)		(L/kg)	(h)	(%)
7	CN	[	Η	Cl	Cl	7.9	5.2	136	25	9	2.3	1.7	100
16	CN	I	F	Cl	Cl	2.0	5.6	136	25	14	1.2	0.8	51
17	CN	ſ	Η	CN	Cl	2.0	6.8	160	82	7	2.6	0.5	25
18	CN	ſ	Η	Cl	CN	3.2	6.9	160	35	4	2.2	1.0	15
19	Cl		Н	Cl	CN	3.2	5.8	136	49	12	4.5	1.6	100

<sup>*a*</sup>human TRPV4 IC<sub>50</sub> values measured by a calcium flux FLIPR assay (see Supporting Information for details); LLE = lipophilic ligand efficiency; PSA = total polar surface area; CL = total clearance;  $f_u$  = fraction unbound (rat); Vdss = volume of distribution; MRT = mean resonance time; F = oral bioavailability

Replacing the *ortho*-chloro substituent of the aryl sulfonamide with a cyano group ( $R^3 = CN$ , Compound 17) resulted in a 4-fold increase in potency ( $IC_{50} = 2.0 \text{ nM}$ ) relative to parent compound 7. While the LLE improved to 6.8, the PK properties in rat diminished. In particular, the MRT decreased to 0.5 h due to a rise in total clearance (CL = 82 mL/min/kg), and the oral bioavailability was only 25%. Metabolite ID studies in rat hepatocytes suggested that this increased metabolism was largely driven by glutathione-mediated sulfonamide cleavage.

In relation to **17**, Compound **18** featured a transposition of the chloro and cyano substituents on the aryl sulfonamide. This modification preserved the excellent LLE of **17** as well as reduced total clearance (CL = 35 mL/min/kg) and clearance adjusted for unbound fraction ( $CL/f_u = 4$ ). While the enhanced metabolic stability was reflected in a longer MRT (1.0 h), oral bioavailability remained poor (F

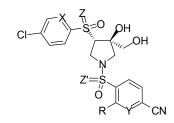
#### Journal of Medicinal Chemistry

= 15%). Given this result, subsequent targets were aimed at reducing PSA in an effort to improve cell permeability and enhance gastrointestinal absorption. To this end, Compound **19** featured a single-point modification involving replacement of the cyano substituent on the aryl sulfone with a chloride (R<sup>1</sup> = Cl). Relative to parent compound **18**, this maintained potency (IC<sub>50</sub> = 3.2 nM) while reducing PSA to 136. Despite an increase in both total CL (49 mL/min/kg) and CL/f<sub>u</sub> (12), MRT was elongated due to an elevated volume of distribution (V<sub>dss</sub> = 4.5 L/kg). Gratifyingly, the oral bioavailability improved to 100%, likely driven by enhanced cell permeability (350 nm/sec) and reduced oxidative (first-pass) metabolism of the aryl sulfone functionality.<sup>32</sup>

Our interest in Compound **19** was further buoyed by its clean off-target profile. It exhibited high TRP selectivity (TRPA1, TRPV1, TRPM2, TRPM8, TRPC3-C6 IC<sub>50</sub> > 30  $\mu$ M) and weak activity (IC<sub>50</sub> > 10  $\mu$ M) against a panel of receptors including CYP's, hERG and Ca<sub>v</sub>1.2. Furthermore, it was inactive up to 100  $\mu$ M in an assay measuring time-dependent inhibition (TDI) of CYP3A4.

Additional structural modifications were investigated to reduce lipophilicity in hopes of further enhancing PK properties. This involved the insertion of heteroatoms into the various subunits of the scaffold (Table 3). Compound **20**, featuring a pyridyl sulfonamide functionality (Y = N), demonstrated reduced total clearance (CL = 22 mL/min/kg), outstanding clearance adjusted for unbound fraction (CL/f<sub>u</sub> = 1.2), and nearly a 2-fold improvement in MRT (3 h). Unfortunately, however, the TRPV4 potency of **20** (IC<sub>50</sub> = 631 nM) was approximately 200-fold weaker than that of parent compound **19**. A significant percentage of this activity could be recovered by the re-introduction of a lipophilic *ortho*-substituent on the aryl sulfonamide moiety. In a representative example, Compound **21** (R = CF<sub>3</sub>) demonstrated an *in vitro* activity (IC<sub>50</sub> = 20 nM) and LLE (6.0) comparable to **19**. However, **21** also featured a startling erosion of PK properties (CL = 310 mL/min/kg, CL/f<sub>u</sub> = 66, MRT = 0.2 h, F = 8%), prompting us to shift our attention to other subunits.

Table 3. Incorporation of heteroatoms into the pyrrolidine diol scaffold<sup>a</sup>



Cmpd	Х	Y	Ζ	Z'	R	IC <sub>50</sub>	LLE	PSA	CL	$CL/f_u$	Vdss	MRT	F
						(nM)			(mL/min/kg)		(L/kg)	(h)	(%)
19	СН	СН	0	0	Cl	3.2	5.8	136	49	12	4.5	1.6	100
20	СН	N	0	0	Н	631	4.9	149	22	1.2	4.1	3.0	62
21	СН	N	0	0	CF <sub>3</sub>	20	6.0	149	310	66	3.6	0.2	8
22	Ν	СН	0	0	Cl	5.0	5.7	149	23	11	2.0	1.4	62
23	СН	СН	NH	Ο	Cl	20	5.0	143	52	4.8	3.2	1.0	38
24	СН	СН	0	NH	Cl	40	4.2	143	48	8.9	2.7	0.9	80

<sup>*a*</sup>human TRPV4 IC<sub>50</sub> values measured by a calcium flux FLIPR assay (see Supporting Information for details); LLE = lipophilic ligand efficiency; PSA = total polar surface area; CL = total clearance;  $f_u$  = fraction unbound (rat); Vdss = volume of distribution; MRT = mean resonance time; F = oral bioavailability

Installation of a heteroaromatic sulfone proved to be better tolerated in regard to both TRPV4 activity and PK properties. The most encouraging results were attained with a 2-pyridyl sulfone, in which Compound **22** displayed nearly identical TRPV4 potency (IC<sub>50</sub> = 5 nM), LLE (5.7) and MRT (1.4 h) relative to direct comparator **19**. Notably, oral bioavailability was reduced (F = 62%), possibly due to limited membrane permeability caused by an increase in total polar surface area (PSA = 149).

Structural diversity was further broadened by the exploration of sulfoximine (Compound  $23^{33}$ ) and sulfonimidamide (Compound  $24^{34}$ ) analogs. While the latter featured excellent oral bioavailability in rat (F = 80%), neither exemplar offered an advantage over parent compound 19 in terms of TRPV4 activity, LLE, or PK properties (CL, MRT, F).

#### Journal of Medicinal Chemistry

With the inability of Compounds 20-24 to improve on the overall profile of Compound 19, focus was subsequently shifted toward modifications of the diol subunit (Table 4). Specifically, amino alcohol 25, a direct comparator of 19, emerged as a benchmark compound with exquisite TRPV4 activity (IC<sub>50</sub> = 0.3 nM) and exceptional LLE (7.6). Its PK properties in rat were also promising, highlighted by reduced total clearance (CL = 12 mL/min/kg) and  $CL/f_{\mu}$  (2.7) in addition to an MRT of nearly 6 hours. Compound 26, the 2-pyridyl sulfone analog of 25, featured improved oral bioavailability (F = 64%) while maintaining sub-nanomolar potency (IC<sub>50</sub> = 0.5 nM) and excellent clearance adjusted for unbound fraction (CL/ $f_u$  = 3.8). The MRT of Compound 26 (1.9 h), while diminished relative to 25, remained comparable to that of leading diol 19. Importantly, Compound 27, the (3R, 4S)-diastereomer of 26, was equipotent in the FLIPR assay ( $IC_{50} = 0.5$  nM), thereby constituting divergent SAR relative to the diol chemotype (Figure 3). Furthermore, it exhibited reduced  $CL/f_{\mu}$  (1.7) with similar MRT (2.2 h) and oral bioavailability (56%) in comparison to 26. The encouraging oral bioavailability of 26 and 27 could be due to facilitated gastrointestinal absorption driven by their enhanced aqueous solubility<sup>35</sup> (500 µM), which exceeded that of 25 (280 µM). Along with a potential reduction in first-pass metabolism, this could also explain the improvement relative to diols with similar PSA such as 17 and 18, which each exhibited lower aqueous solubility (380, 400 µM, respectively).

Table 4. Profiles of amino alcohols 25, 26 and 27<sup>a</sup>

CI-CI-CN

Cmpd	Х	stereochem.	IC <sub>50</sub>	LLE	PSA	CL	$CL/f_u$	Vdss	MRT	F
			(nM)			(mL/min/kg)		(L/kg)	(h)	(%)
25	СН	3 <i>S</i> , 4 <i>S</i>	0.3	7.6	142	12	2.7	4.0	5.7	22

ACS Paragon Plus Environment

Journal of Medicinal Chemistry	Journal	l of N	1edicinal	C	hemistry
--------------------------------	---------	--------	-----------	---	----------

26	Ν	3 <i>S</i> , 4 <i>S</i>	0.5	7.4	154	12	3.8	1.3	1.9	64
27	Ν	3 <i>R</i> , 4 <i>S</i>	0.5	7.4	154	1.7	1.7	0.2	2.2	56

<sup>*a*</sup>human TRPV4 IC<sub>50</sub> values measured by a calcium flux FLIPR assay (see Supporting Information for details); LLE = lipophilic ligand efficiency; PSA = total polar surface area; CL = total clearance;  $f_u$  = fraction unbound (rat); Vdss = volume of distribution; MRT = mean resonance time; F = oral bioavailability

In contrast to the diol exemplars, the off-target profiles of these amino alcohols featured several areas of concern. For one, compound **25** displayed activity in a phenotypic assay measuring phospholipidosis<sup>36</sup> risk (*p*MEC = 4.7, response = 425%)<sup>37</sup>, which is not uncommon to lipophilic amines containing aromatic residues (Table 5). Cationic amphiphilic drugs (CAD's) have been reported to cause phospholipidosis in humans and activity in the phenotypic screen has correlated with a CAD "likeness" score exceeding 50.<sup>38</sup> With a CAD score defined as the sum of indicators of lipophilicity (CHI IAM)<sup>39</sup> and basicity ( $\Delta$ CHI (pH10.5 – pH7.4))<sup>39</sup>, design elements to address phospholipidosis risk have been well established. Indeed, amino alcohols **26** and **27**, each featuring lower lipophilicity (CHI IAM = 42, 36, respectively) and basicity ( $\Delta$ CHI (pH10.5 – pH7.4) = -2.2, 3.7, respectively) relative to **25** (CHI IAM = 49,  $\Delta$ CHI (pH10.5 – pH7.4) = 9), were gratifyingly inactive in the phenotypic assay (*p*MEC < 4).

**Table 5.** Off-target profiles of amino alcohols 25, 26 and  $27^a$ 

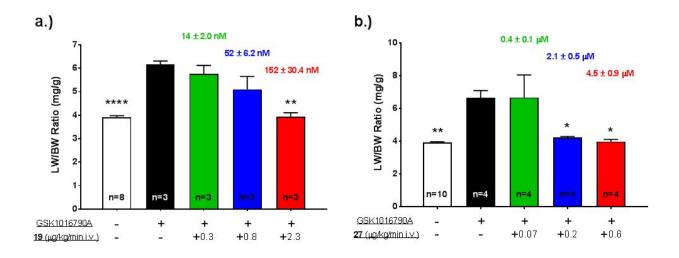
			Phospl	nolipidosis		
Cmpd	hERG IC50	CAD	<i>p</i> MEC	Response <sup>37</sup>	3A4 TDI (100	3A4 RI IC <sub>50</sub>
	(µM)	likeness		(%)	μΜ)	(µM)
25	5.3	58	4.7	425	yes	0.1
26	8.0	40	< 4		no	0.3
27	8.0	40	< 4		no	4.0

#### Journal of Medicinal Chemistry

<sup>*a*</sup>Details for hERG, phospholipidosis, 3A4 TDI and 3A4 RI assays described in Supporting Information; pMEC = -log(minimum effective concentration)

Compounds 26 and 27 also held additional important advantages over 25. Whereas the latter exhibited time-dependent inhibition of human CYP3A4 at 100  $\mu$ M, exemplars 26 and 27 were inactive at the same concentration, potentially a result of their reduced basicity. This may also have contributed to their lower activity (IC<sub>50</sub> = 8  $\mu$ M) against the cardiac hERG channel. Ultimately, compound 27 emerged as the most promising amino alcohol within this structural class, with reduced DDI (drug-drug interaction) risk relative to 25 and 26 as demonstrated by its weaker activity in an assay measuring reversible inhibition of CYP3A4 (IC<sub>50</sub> = 4  $\mu$ M).

Leading diol **19** and amino alcohol **27** were each investigated in a rodent *in vivo* assay suggestive of therapeutic potential for pulmonary edema (Figure 4). In this model, rats were treated with vehicle or TRPV4 antagonist prior to administration of known TRPV4 agonist, GSK1016790A (100 µg/kg total i.v. dose).<sup>11,40,41</sup> Importantly, previous studies have shown that the effects of GSK1016790A correlate with those induced by myocardial infarction in rodents.<sup>11</sup> In the vehicle-treated group, GSK1016790A demonstrated robust pulmonary edema as quantified by an increase in wet lung weight/body weight ratio relative to naïve animals. Gratifyingly, significant attenuation of agonist effects was observed with diol **19** (2.3 µg/kg/min i.v., p < 0.01 vs. vehicle) and amino alcohol **27** (0.2 µg/kg/min i.v., p < 0.05 vs. vehicle).<sup>42</sup>



**Figure 4.** Effects of TRPV4 agonist GSK1016790A (10  $\mu$ g/kg/min i.v. for 10 min) on lung wet weight/body weight ratio (LW/BW) in vehicle-treated rats and rats pre-treated with TRPV4 antagonists **19** (a) and **27** (b). Full inhibition of agonist-mediated LW/BW response was achieved with the 2.3  $\mu$ g/kg/min dose of **19** (99% inhibition) and the 0.2  $\mu$ g/kg/min dose of **27** (88% inhibition). Corresponding steady-state plasma concentrations are indicated above and data are presented as mean  $\pm$  SEM. \**p* < 0.05, \*\**p* < 0.01, and \*\*\*\**p* < 0.0001 vs. vehicle.<sup>43</sup>

Further details regarding the *in vivo* activity of Compounds **19** and **27** are provided in Table 6. Notably, despite slightly weaker *in vitro* potency in rat, Compound **19** featured superior activity ( $IC_{50} = 33 \text{ nM}$ )<sup>44</sup> in the *in vivo* pulmonary edema model in comparison to Compound **27** ( $IC_{50} = 611 \text{ nM}$ ). Correcting for rat unbound fraction ( $rf_u$ ) generated a "free" *in vivo*  $IC_{50}$  of 0.3 nM for diol **19**, a significant improvement over that associated with amino alcohol **27** (8.5 nM).

 Table 6. In vivo activity of Compounds 19 and 27 in the rat pulmonary edema model<sup>a</sup>

Cmpd	rTRPV4	in vivo	rf <sub>u</sub>	"Free" in vivo
	IC <sub>50</sub>	IC <sub>50</sub>	(%)	IC <sub>50</sub> (nM)
	(nM)	(nM)		
19	4.0	33	4.2	0.3

ACS Paragon Plus Environment

 <sup>*a*</sup>rat TRPV4 IC<sub>50</sub> values measured by a calcium flux FLIPR assay (see Supporting Information for details);  $rf_u$  = fraction unbound (rat); conc = concentration

Ultimately, with enhanced *in vivo* activity, less off-target promiscuity and dose-proportional oral exposures in both rat and dog (Table 7), diol **19** was selected over amino alcohol **27** for further progression. To our delight, a 7-day rat safety assessment (100/300 mpk) proceeded with no compound-related macro- or microscopic findings. Based on scaling of preclinical PK data, the exposures at 300 mpk provided minimum rat safety margins of 68-fold and 42-fold over the predicted human  $C_{max}$  and AUC, respectively, with  $C_{min}$  set at the concentration required for full inhibition in the rat pulmonary edema assay (Figure 4a).

Species	p.o. dose (mpk)	$AUC_{0-t} (\mu g \cdot h/mL)$	DNAUC <sub>0-t</sub> (h·kg/L)
rat	2.4	1.1	0.5
rat	98	40	0.4
dog	0.9	2.5	2.9
dog	30	90	3.0

 Table 7. PK studies with Compound 19 in rat and dog<sup>a</sup>

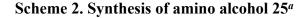
a n = 3 animals; AUC<sub>0-t</sub> = area under the plasma concentration time curve to last time point with quantifiable drug; DNAUC<sub>0-t</sub> = dose-normalized AUC<sub>0-t</sub>.

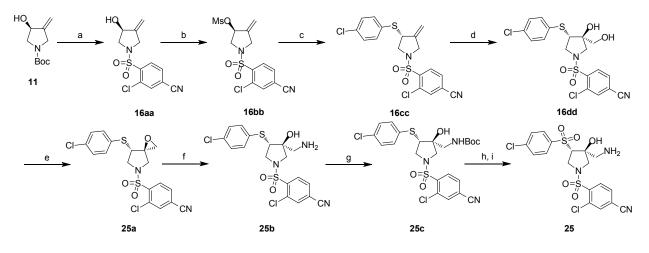
#### ■ CONCLUSIONS:

A novel class of sulfone pyrrolidine TRPV4 antagonists<sup>45</sup> has been discovered, with exemplars featuring excellent potency and tractability with respect to physicochemical and PK properties. Structureactivity relationships within a diol chemotype were investigated in order to balance potency with desirable PK properties, whereas efforts within an amino alcohol sub-series leveraged exquisite *in vitro* activity while addressing off-target concerns. Leading exemplars from both chemotypes (**19** and **27**) demonstrated blockade of TRPV4 agonist effects in an *in vivo* model indicative of therapeutic potential. Of these compounds, **19** emerged as a viable candidate for HF studies which can prove instrumental in assessing the utility of TRPV4 antagonists in the treatment of HF-induced pulmonary edema.

#### ■ EXPERIMENTAL SECTION:

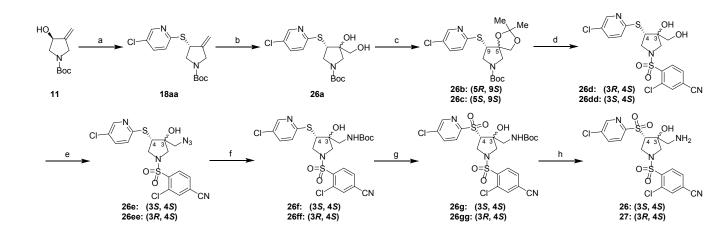
**Chemistry.** The syntheses of diol **19** and amino alcohols **25-27** are described below. Detailed synthetic protocols and spectroscopic characterizations for Compounds **4**, **6-7**, **16-18** and **20-24** are provided in the Supporting Information (along with procedures for biological assays and pharmacokinetic studies). The purity of each compound was determined to be >95% on a Waters Acquity LCMS equipped with a Acquity<sup>TM</sup> UPLC BEH C18 1.7mm, 2.1 x 50 mm column using a gradient of 3% to 100% MeCN, 0.1% formic acid at 1 ml/min flow rate with an Acquity PDA detector at 210 and 350 nm. Mass determinations were conducted using a Waters Acquity SQD with positive/negative switching in ESI mode.





<sup>a</sup>Reagents and conditions: (a) 1. TFA, DCM; 2. 2-chloro-4-cyanobenzene-1-sulfonyl chloride, TEA, DCM (63%); (b) MsCl, TEA, DCM, 0 °C (88%); (c) 4-chlorobenzenethiol, K<sub>2</sub>CO<sub>3</sub>, DMF, (59%); (d) OsO<sub>4</sub>, NMO, THF (37%); (e) MsCl, TEA, K<sub>2</sub>CO<sub>3</sub>, DCM, MeOH (52%;) (f) NH<sub>3</sub>, EtOH, 70 °C, 99%; (g) Boc<sub>2</sub>O, TEA, THF (82%); (h) *m*-CPBA, DCM (99%); (i) TFA, DCM (66%).

#### Scheme 3. Synthesis of amino alcohols 26 and 27<sup>a</sup>



<sup>a</sup>Reagents and conditions: (a) 1. MsCl, TEA, DCM; 2. 5-chloropyridine-2-thiol, K<sub>2</sub>CO<sub>3</sub>, DMF (91%); (b) OsO<sub>4</sub>, NMO, THF (78% (diastereomeric mixture)); (c) propane-2,2-diol, PPTS, DCM (38% (26b), 38% (26c)); (d) 1. TFA, DCM; 2. NaHCO<sub>3</sub>, THF, H<sub>2</sub>O, 2-chloro-4-cyanobenzene-1-sulfonyl chloride (82% (26d), 85% (26dd)); (e) 1. MsCl, TEA, DCM; 2. NaN<sub>3</sub>, DMF (39% (26e), 83% (26ee)); (f) 1. PMe<sub>3</sub>, THF; 2. Boc<sub>2</sub>O, TEA, THF (67% (26f), 77% (26ff)); (g) *m*-CPBA, DCM (98% (26g), 77% (26gg)); (h) TFA, DCM (54% (26), 50% (27)).

(R)-tert-Butyl 3-((4-chlorophenyl)thio)-4-methylenepyrrolidine-1-carboxylate (12, R = para**chloro):** To a stirred mixture of (S)-tert-butyl 3-hydroxy-4 methylenepyrrolidine-1-carboxylate (11, 12.33) g, 61.9 mmol, 1 equiv) in DCM (120 mL) was added TEA (12.94 mL, 93 mmol, 1.5 equiv) in one portion. The mixture was subsequently cooled in an ice bath to give an internal temperature of 5 °C. Methanesulfonyl chloride (8.51 g, 74.3 mmol, 1.2 equiv) was added dropwise via a syringe over 40 min such that the internal temperature did not exceed 2 °C. Following the addition, the reaction vessel was removed from the bath and the mixture was allowed to stir at ambient temperature for 45 min. Following this duration, the mixture was diluted with 50 mL of DCM and washed with 40 mL of water. The layers were separated and the organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure to provide the crude mesylate as a light brown oil (19.9 g), which was immediately carried forward without further purification. To a stirred mixture of the crude mesylate and 4chlorobenzenethiol (9.40 g, 65.0 mmol) in DMF (125 mL) in an ice bath was added K<sub>2</sub>CO<sub>3</sub> (powder form, 12.83 g, 93 mmol) portion-wise. Following the addition, the reaction vessel was removed from the bath and the mixture was allowed to stir at ambient temperature for 1 h. Following this duration, the stirring was stopped and the mixture was allowed to age at RT for 40 min. To this mixture was added 1:1

EtOAc/hexane (400 mL), and ice water (200 mL) sequentially such that the internal temp remained below 20 °C. The resulting mixture was filtered and the filtrate was phase-separated. The organic extract was washed with water (200 mL, then 2 x 150 mL) dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure to give a yellow oil. The crude product was subsequently dissolved in DCM (50 mL), absorbed onto Isolute and concentrated to remove the residual DCM. Purification by silica gel chromatography (0-30% EtOAc:Hexanes) afforded **12** as a clear oil (14.9 g, 74%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm 1.40 (br. s., 9 H) 3.39 (dd, *J*=11.80, 2.51 Hz, 1 H) 3.69 (dd, *J*=11.80, 6.27 Hz, 1 H) 3.89 (d, *J*=12.05 Hz, 1 H) 4.00 - 4.11 (m, 1 H) 4.47 (br. s., 1 H) 5.01 (br. s., 1 H) 5.11 (s, 1 H) 7.35 - 7.51 (m, 4 H); *m/z* 269.9 [MH-*t*-Bu]<sup>+</sup>.

(*R*)-*tert*-Butyl 3-((4-chlorophenyl)sulfonyl)-4-methylenepyrrolidine-1-carboxylate (13, R = para-chloro): To a cooled solution of (*R*)-*tert*-butyl 3-((4-chlorophenyl)thio)-4-methylenepyrrolidine-1-carboxylate (14.57 g, 44.7 mmol, 1 equiv) in 250 mL of DCM was added *m*-CPBA (25 g, 112 mmol, 2.5 equiv) portion-wise over 7 min such that the internal temperature stayed  $\leq$  -5 °C. After 1 h, to the cold mixture was added 10% wt aqueous Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> solution (140 mL) portion-wise over 40 min such that the internal temperature was added 325 mL of saturated aqueous NaHCO<sub>3</sub> over 30 min such that the internal temperature remained  $\leq$  1 °C. The pH of the resulting mixture was 9. The cold mixture was phase-separated. The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure to approximately 50 mL total volume. The solution was adsorbed onto Isolute and concentrated to remove residual DCM. Purification by silica gel chromatography (0-70% EtOAc:Hexanes) afforded **13 (R** = *para*-chloro) as a white foam (15.9 g, 89% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm 1.41 (s, 4 H) 1.36 (s, 5 H) 3.53 - 3.72 (m, 2 H) 3.72 - 3.85 (m, 1 H) 3.86 - 3.99 (m, 1 H) 4.67 (d, *J*=7.28 Hz, 1 H) 5.18 (br.s., 1 H) 5.39 (br. s., 1 H) 7.76 (d, *J*=8.28 Hz, 2 H) 7.85 (d, *J*=7.53 Hz, 2 H); *m/z* 380.0 [M+Na]<sup>+</sup>.

(3R,4S)-tert-Butyl-4-((4-chlorophenyl)sulfonyl)-3-hydroxy-3-(hydroxymethyl)pyrrolidine-1carboxylate (14, R = para-chloro): To a stirred, cooled mixture of (R)-tert-butyl 3-((4-

chlorophenvl)sulfonvl)-4-methylenepyrrolidine-1-carboxylate (15.9 g, 44.4 mmol) and NMO (10.41 g, 89 mmol, 2 equiv) in 300 mL of THF was added OsO<sub>4</sub> solution in *t*-butanol (25 mL, 2.5% wt, 1.99 mmol, 0.045 equiv) via syringe over 4 min such that the internal temperature did not exceed 15 °C. After completion of addition, the internal temperature was allowed to gradually rise to 22-23 °C within 45 min and remained at 23 °C. After 3 h, an additional 2.8 mL of  $OsO_4$  solution (total 0.05 equiv added) was added. After 5 h, stirring was stopped and the mixture was aged at RT over 14 h. The yellow solution was subsequently chilled in an ice bath and saturated aqueous Na<sub>2</sub>SO<sub>3</sub> (100 mL) was added over 10 min (keeping internal temperature at approximately 9 °C), bringing the pH of the reaction mixture to 9-10. The mixture was stirred in the ice bath for 1 h. Following this duration, the ice bath was removed. To the cold mixture was added 100 mL of water and the resulting suspension was filtered through Celite. The cake was washed with 300 mL of EtOAc. The filtrate was phase-separated. The organic layer was washed with brine (30 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and DARCO (activated charcoal), filtered, and concentrated under reduced pressure. The wet residue was taken up in 200 mL of DCM and washed with 20 mL of brine. The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure to afford an off-white foamy residue. The residue was re-dissolved in DCM and purified by silica gel chromatography (5-70% (3:1 EtOAc:EtOH):Hexanes) to provide 14 ( $\mathbf{R} = para$ -chloro) as an off-white solid (14.1 g, 81% vield). <sup>1</sup>H NMR (400MHz, DMSO-*d*<sub>6</sub>) δ ppm 7.91 (d, *J*=8.5 Hz, 2H), 7.77 (d, J=8.3 Hz, 2H), 5.53 (d, J=9.0 Hz, 1H), 4.85 (d, J=4.3 Hz, 1H), 3.98 (d, J=4.5 Hz, 1H), 3.82 - 3.42 (m, 5H), 3.28 - 3.16 (m, 1H), 1.40 (s, 9H); m/z 414.1 [M+Na]+.

(3*R*,4*S*)-4-((4-Chlorophenyl)sulfonyl)-3-(hydroxymethyl)pyrrolidin-3-ol, hydrochloride: (3*R*,4*S*)*tert*-Butyl-4-((4-chlorophenyl)sulfonyl)-3-hydroxy-3-(hydroxymethyl)pyrrolidine-1-carboxylate (14, R = *para*-chloro, 14.0 g, 35.7 mmol) was dissolved in 4N HCl/1,4-dioxane solution (200 mL, 800 mmol) and the resulting reaction mixture was stirred vigorously for 1 h. The slurry was diluted with TBME (50 mL), filtered, and the solid was washed with TBME (50 mL). The solid was subsequently dried under vacuum to afford 10.8 g (92% yield) of (3*R*,4*S*)-4-((4-chlorophenyl)sulfonyl)-3(hydroxymethyl)pyrrolidin-3-ol, hydrochloride as a white solid. <sup>1</sup>H NMR (400MHz, Methanol-*d*<sub>4</sub>) δ ppm 8.00 (d, *J*=8.5 Hz, 2H), 7.75 (d, *J*=8.5 Hz, 2H), 4.18 - 4.11 (m, 1H), 4.10 - 3.96 (m, 2H), 3.81 (dd, *J*=8.0, 13.6 Hz, 1H), 3.70 - 3.55 (m, 3H), 3.44 (d, *J*=11.8 Hz, 1H); *m/z* 292.0 [MH]<sup>+</sup>.

(S)-3-Chloro-4-((3-hydroxy-4-methylenepyrrolidin-1-yl)sulfonyl)benzonitrile (16aa): То а solution of (S)-tert-butyl 3-hydroxy-4-methylenepyrrolidine-1-carboxylate (11, 10 g, 50.2 mmol) in DCM (40 mL) was added TFA (58.0 mL, 753 mmol) and the resulting reaction mixture was stirred at RT for 1 h. Following this duration, the reaction contents were concentrated under reduced pressure to give a tan oil, which was suspended in DCM (80 mL) and cooled in an ice bath. To this reaction mixture was added TEA (35.0 mL, 251 mmol) cautiously, followed by 2-chloro-4-cyanobenzene-1-sulfonyl chloride (11.85 g, 50.2 mmol). The reaction mixture was subsequently warmed to RT and stirred at 23 °C for 1 h. The mixture was then diluted with water (80 mL) and extracted with DCM (80 mL). The combined organics were washed with brine and concentrated. Purification by silica gel chromatography (0-60%) EtOAc:Hexane) provided 16aa as a pale yellow solid (9.5 g, 63% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ ppm 3.12 (dd, J=9.91, 5.65 Hz, 1 H) 3.64 (dd, J=10.04, 6.27 Hz, 1 H) 4.04 - 4.10 (m, 1 H) 4.49 (d, J=5.27 Hz, 1 H) 5.11 (d, J=1.51 Hz, 1 H) 5.18 (d, J=1.76 Hz, 1 H) 5.56 (d, J=5.02 Hz, 1 H) 8.03 - 8.07 (m, 1 H) 8.11 - 8.14 (m, 1 H) 8.36 (d, J=1.26 Hz, 1 H); m/z 280.8 [MH-H<sub>2</sub>O<sup>+</sup>].

(*S*)-1-((2-Chloro-4-cyanophenyl)sulfonyl)-4-methylenepyrrolidin-3-yl methanesulfonate (16bb): To a solution of (*S*)-3-chloro-4-((3-hydroxy-4-methylenepyrrolidin-1-yl)sulfonyl)benzonitrile (16aa, 9.50 g, 31.8 mmol) in DCM (80 mL) in an ice bath was added TEA (7.80 mL, 56.0 mmol) followed by methanesulfonyl chloride (3.10 mL, 39.7 mmol). The resulting reaction mixture was stirred at RT for 1 h. Following this duration, water (80 mL) was added and the resulting layers were separated. The aqueous layer was extracted with DCM (1 x 80 mL). The combined organics were washed with water (2 x), dried over anhydrous MgSO<sub>4</sub> and filtered. The filtrate was concentrated under reduced pressure to give an offwhite solid. Trituration with 15% EtOAc in hexane, filtration and concentration under reduced pressure afforded 16bb as a white solid (10.6 g, 88% yield). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  ppm 3.24 (s, 3 H)

3.66 - 3.81 (m, 2 H) 4.02 (d, *J*=14.56 Hz, 1 H) 4.20 (d, *J*=14.31 Hz, 1 H) 5.45 (s, 1 H) 5.51 (d, *J*=1.76 Hz, 2 H) 7.99 - 8.09 (m, 1 H) 8.11 - 8.20 (m, 1 H) 8.37 (d, *J*=1.51 Hz, 1 H); *m/z* 280.8 [MH-H<sub>2</sub>O]<sup>+</sup>.

(*R*)-3-Chloro-4-((3-chlorophenyl)thio)-4-methylenepyrrolidin-1-yl)sulfonyl)benzonitrile (16cc): To a solution of (*S*)-1-((2-chloro-4-cyanophenyl)sulfonyl)-4-methylenepyrrolidin-3-yl methanesulfonate (16bb, 2.5 g, 6.63 mmol) in chloroform (100 mL) at RT was added 4-chlorobenzenethiol (1.151 g, 7.96 mmol). Triethylamine (2.312 mL, 16.59 mmol) was then added and the resulting mixture was allowed to stir at RT for 18 h. Following this duration, the reaction mixture was concentrated under reduced pressure and purified by silica gel chromatography to give **16cc** as a white solid (1.7 g, 59%). <sup>1</sup>H NMR (400MHz, CHLOROFORM-*d*)  $\delta$  ppm 8.21 (d, *J*=8.0 Hz, 1H), 7.85 (s, 1H), 7.71 (d, *J*=8.3 Hz, 1H), 7.36 - 7.26 (m, 4H), 5.11 (d, *J*=7.3 Hz, 2H), 4.30 - 4.21 (m, 1H), 4.12 (t, *J*=14.2 Hz, 2H), 3.89 (dd, *J*=6.3, 11.0 Hz, 1H), 3.56 (dd, *J*=4.0, 11.0 Hz, 1H); *m/z* 425.0 [MH]<sup>+</sup>.

# 3-Chloro-4-(((3R,4S)-4-((4-chlorophenyl)thio)-3-hydroxy-3-(hydroxymethyl)pyrrolidin-3-

yl)sulfonyl)benzonitrile (16dd): To a solution of (*R*)-3-chloro-4-((3-((4-chlorophenyl)thio)-4methylenepyrrolidin-1-yl)sulfonyl)benzonitrile (16cc, 1.66 g, 3.90 mmol) in tetrahydrofuran (13.0 mL) was added NMO (0.503 g, 4.29 mmol) and osmium tetroxide (2.5% in *t*-BuOH, 2.45 mL, 0.195 mmol) sequentially to give a bright yellow solution. After 30 min, an additional 270 mg (0.6 equiv) of NMO was added. After 30 min, an additional 150 mg (0.3 equiv) of NMO was added. After 10 min, saturated aqueous NaHSO<sub>3</sub> was added (40 mL) followed by DCM (100 mL). The layers were separated and the organic layer was dried over anhydrous sodium sulfate, filtered and concentrated under reduced pressure. Purification by silica gel chromatography (0-50% EtOAc:Hexanes) afforded 16dd as a white solid (660 mg, 37% yield). <sup>1</sup>H NMR (400 MHz, CHLOROFORM-*d*)  $\delta$  ppm 1.63 (br. s., 1 H) 2.03 (br. s., 1 H) 3.09 (s, 1 H) 3.50 (d, *J*=10.79 Hz, 1 H) 3.59 (dd, *J*=10.54, 2.76 Hz, 1 H) 3.70 - 3.77 (m, 2 H) 3.97 (d, *J*=11.29 Hz, 1 H) 4.03 (dd, *J*=10.54, 6.02 Hz, 1 H) 7.27 - 7.39 (m, 4 H) 7.69 (d, *J*=8.03 Hz, 1 H) 7.84 (s, 1 H) 8.19 (d, *J*=8.28 Hz, 1 H); *m/z* 441.0 [M-OH]<sup>+</sup>.

(*R*)-*tert*-Butyl 3-((5-chloropyridin-2-yl)thio)-4-methylenepyrrolidine-1-carboxylate (18aa):

To a solution of (*S*)-*tert*-butyl 3-hydroxy-4-methylenepyrrolidine-1-carboxylate (**11**, 2.0 g, 10.04 mmol) in DCM (30 mL) was added TEA (2.1 mL, 15.07 mmol), followed by mesyl chloride (0.94 mL, 12.06 mmol) dropwise. The resulting reaction mixture was stirred at RT for 30 min. Following this duration, the reaction mixture was diluted with water and extracted with dichloromethane. The organic extract was washed with water, dried over anhydrous magnesium sulfate and filtered. The filtrate was concentrated under reduced pressure to give the desired mesylate. The intermediate was immediately dissolved in *N*,*N*-dimethylformamide (100 mL). 5-Chloropyridine-2-thiol (1.46 g, 10.04 mmol) was then added, followed by potassium carbonate (2.08 g, 15.06 mmol). The reaction mixture was stirred for 1 h at RT, diluted with water and extracted with ethyl acetate. The organic extract was washed with water, dried over anhydrous magnesium sulfate and filtered. The filtrate over anhydrous magnesium sulfate and filtered. The at RT, diluted with water and extracted with ethyl acetate. The organic extract was washed with water, dried over anhydrous magnesium sulfate and filtered. The filtrate was subsequently concentrated under reduced pressure and purified by silica gel chromatography (0-15% EtOAc:Hexanes) to give **18aa** as a clear, colorless oil (3.0 g, 91% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm 1.39 - 1.41 (m, 9 H) 3.53 - 3.59 (m, 1 H) 3.86 - 4.00 (m, 2 H) 4.13 - 4.32 (m, 2 H) 4.84 - 4.97 (m, 1 H) 5.01 - 5.13 (m, 1 H) 7.41 - 7.45 (m, 1 H) 7.71 - 7.76 (m, 1 H) 8.46 - 8.49 (m, 1 H); *m*/z 327.0 [MH]<sup>+</sup>.

#### 3-Chloro-4-(((3R,4S)-4-((4-chlorophenyl)sulfonyl)-3-hydroxy-3-(hydroxymethyl)pyrrolidin-1-

vl)sulfonyl)benzonitrile (19): To а solution of (3R,4S)-4-((4-chlorophenyl)sulfonyl)-3-(hydroxymethyl)pyrrolidin-3-ol, hydrochloride (9.66 g, 29.4 mmol) in THF (100 mL) was added saturated aqueous NaHCO<sub>3</sub> (150 mL) and the resulting reaction mixture was stirred for 5 min, to which was added a suspension of 2-chloro-4-cyanobenzene-1-sulfonyl chloride (7.02 g, 28.8 mmol) in THF (150 mL) portion-wise at 10 °C. The resulting reaction mixture was stirred for 1 h, at which an additional 2-chloro-4cyanobenzene-1-sulfonyl chloride (0.7 g) was added and the reaction mixture was stirred for an additional 1 h. Following this duration, the reaction mixture was diluted with EtOAc and H<sub>2</sub>O, the layers separated, and the aqueous phase extracted with EtOAc. The combined organics were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude oil was purified in two batches, using flash column chromatography eluting with a 5% EtOAc-hexanes to 100% EtOAc gradient. The pure

#### Journal of Medicinal Chemistry

product-containing fractions were combined and the solvent evaporated under reduced pressure to afford an off-white foam (12.7 g). The foam was triturated with heptane (50 mL) and DCM (5 mL) and the solvent was evaporated under reduced pressure and dried under vacuum to afford 11.6 g (80% yield) of **19** as an off-white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm 3.38 (d, *J*=10.04 Hz, 1 H) 3.59 (dd, *J*=11.29, 3.01 Hz, 1 H) 3.63 - 3.72 (m, 2 H) 3.74 - 3.87 (m, 2 H) 4.14 (dd, *J*=7.28, 3.01 Hz, 1 H) 4.94 (t, *J*=5.40 Hz, 1 H) 5.76 (s, 1 H) 7.68 - 7.79 (m, 2 H) 7.82 - 7.90 (m, 2 H) 8.01 - 8.08 (m, 1 H) 8.14 (s, 1 H) 8.36 (s, 1 H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm 48.3, 56.8, 63.4, 68.6, 81.8, 116.9, 117.1, 130.3, 130.4, 131.9, 132.3, 132.4, 136.2, 137.8, 139.9, 140.9; LRMS: *m/z* 491.1 [MH]<sup>+</sup>. HRMS: *m/z* 602.96736 (observed), 602.96717 (calculated) (TFA-adduct). Chiral purity was determined to be 100% by chiral HPLC (CHIRALPAK IC, 5 µ, 30 x 250 mm, 30:70 Heptane:EtOH). [ $\alpha$ ]<sup>D</sup><sub>20</sub> = +53° (c = 0.2, CH<sub>3</sub>OH).

#### 3-Chloro-4-(((3S,7S)-7-((4-chlorophenyl)thio)-1-oxa-5-azaspiro[2.4]heptan-5-yl)sulfonyl-

**benzonitrile (25a):** To a solution of **16dd** (0.700 g, 1.52 mmol) in DCM (20.0 mL) was added TEA (0.380 mL, 2.73 mmol) followed by mesyl chloride (0.140 mL, 1.79 mmol). The resulting reaction mixture was stirred at RT for 0.5 h. Following this duration, the reaction mixture was concentrated under reduced pressure to afford a white solid. The solid was dissolved in MeOH (20 mL) to which was added  $K_2CO_3$  (0.316 g, 2.29 mmol). The resulting reaction mixture was stirred at RT for 3 h, diluted with H<sub>2</sub>O and extracted with EtOAc. The combined organics were dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel chromatography (0-35% EtOAc:Hexanes) to afford **25a** as a white solid (350 mg, 52% yield); *m/z* 441.0 [MH]<sup>+</sup>.

## 4-(((3S,4S)-3-(Aminomethyl)-4-((4-chlorophenyl)thio)-3-hydroxypyrrolidin-1-yl)sulfonyl-3-

**chloro benzonitrile (25b):** A suspension of **25a** (0.100 g, 0.227 mmol) in 2 M NH<sub>3</sub> / EtOH (2.0 mL, 4.00 mmol) was irradiated under microwave irradiation at 70 °C for 0.5 h. The reaction mixture was cooled to ambient temperature and the solvent was evaporated under reduced pressure. The residue was dissolved in MeOH and loaded onto a SCX cartridge (2.0 g), eluting with MeOH, followed by 2 M ammonia in

MeOH solution. The ammonia solution was collected and concentrated under reduced pressure to afford **25b** as an oil (100 mg, 99% yield); m/z 458.0 [MH]<sup>+</sup>.

#### tert-Butyl(((3S,4S)-1-((2-chloro-4-cyanophenyl)sulfonyl)-4-((4-chlorophenyl)thio)-3-

hydroxypyrrolidin-3-yl)methyl)carbamate (25c) To a solution of 25b (0.100 g, 0.218 mmol) in THF (3.0 mL) was added TEA (0.061 mL, 0.436 mmol), followed by Boc<sub>2</sub>O (0.061 g, 0.262 mmol) and the resulting reaction mixture was stirred at RT for 1 h. The reaction mixture was diluted with H<sub>2</sub>O and extracted with EtOAc. The combined organics were dried over MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel chromatography (0-35% EtOAc:Hexanes) to afford **25b** as a colorless semi-solid (105 mg, 82% yield); m/z 558.2 [MH]<sup>+</sup>.

4-(((3S,4S)-3-(Aminomethyl)-4-((4-chlorophenyl)sulfonyl)-3-hydroxypyrrolidin-1-yl)sulfonyl)-3chlorobenzonitrile (25): To a solution of 25c (0.105 g, 0.188 mmol) in DCM (3.0 mL) was added m-CPBA (0.065 mL, 0.376 mmol), and the reaction mixture was stirred at ambient temperature for 24 h. The reaction mixture was diluted with H<sub>2</sub>O and extracted with DCM. The combined organics were dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel chromatography (0-40% EtOAc: Hexanes) to afford 110 mg (99% yield) of the corresponding sulfone as a semi-solid (m/z 490.0 [MH]<sup>+</sup>). To a solution of the sulfone (110 mg, 0.186 mmol) in DCM (1 mL) was added TFA (0.2 mL, 2.60 mmol) and the reaction mixture was stirred at RT for 0.5 h. Following this duration, the reaction mixture was concentrated under reduced pressure, diluted with  $H_2O_1$ neutralized with saturated aqueous NaHCO<sub>3</sub> and extracted with EtOAc. The combined organics were dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was purified by silica gel chromatography (0-5% MeOH:DCM) and further purified by semi-prep HPLC, eluting with a 10% MeCN-H<sub>2</sub>O to 90% MeCN-H<sub>2</sub>O gradient. The pure product-containing fractions were combined and concentrated under reduced pressure to afford 25 as a white solid (64 mg, 66% yield). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) \delta ppm 3.42 - 3.48 (m, 1 H) 3.56 - 3.62 (m, 1 H) 3.66 - 3.74 (m, 1 H) 3.81 - 3.94 (m, 1

 H) 4.21 - 4.30 (m, 1 H) 6.31 - 6.59 (m, 1 H) 7.17 - 7.36 (m, 1 H) 7.83 (s, 4 H) 8.12 (s, 2 H) 8.39 (s, 1 H); *m/z* 489.9 [MH]<sup>+</sup>.

(3*R*,4*S*) and (3*S*,4*S*)-*tert*-Butyl 4-((5-chloropyridin-2-yl)thio)-3-hydroxy-3-(hydroxymethyl)pyrrolidine-1-carboxylate (26a): To a solution of 18aa (3.0 g, 9.18 mmol) in THF (30 mL) was added 50% aqueous NMO (2.85 mL, 13.8 mmol) followed by 2.5% OsO<sub>4</sub> in *t*-BuOH (4.61 mL, 0.367 mmol) and the reaction mixture was stirred at RT for 2 h. Following this duration, the reaction mixture was quenched with saturated aqueous Na<sub>2</sub>SO<sub>3</sub> and extracted with EtOAc. The organic extracts were washed with brine, dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was purified by silica gel chromatography (0-55% EtOAc:Hexanes) to afford 26a as a colorless oil (2.58 g, 78% yield, diastereomeric mixture). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm 1.36 - 1.42 (m, 9 H) 3.15 - 3.31 (m, 2 H) 3.85 - 3.98 (m, 1 H) 4.00 - 4.07 (m, 2 H) 4.13 - 4.32 (m, 1 H) 4.86 - 5.09 (m, 1 H) 5.33 - 5.49 (m, 1 H) 7.27 - 7.49 (m, 1 H) 7.66 - 7.86 (m, 1 H) 8.36 - 8.61 (m, 1 H); *m/z* 305.0 [MH-Boc]<sup>+</sup>.

(5*R*,9*S*)-*tert*-Butyl 9-((5-chloropyridin-2-yl)thio)-2,2-dimethyl-1,3-dioxa-7-azaspiro[4.4]nonane-7carboxylate (26b) and (5*S*,9*S*)-*tert*-butyl 9-((5-chloropyridin-2-yl)thio)-2,2-dimethyl-1,3-dioxa-7azaspiro[4.4]nonane-7-carboxylate (26c): To a solution of 26a (2.58 g, 7.15 mmol) in DCM (20 mL) was added 2,2-dimethoxypropane (2.64 mL, 21.4 mmol) and PPTS (0.136 g, 0.715 mmol) and the resulting reaction mixture was stirred at RT for 0.5 h. Following this duration, the reaction mixture was diluted with H<sub>2</sub>O and extracted with DCM. The organics were washed with brine, dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was purified by silica gel chromatography (0-25% EtOAc:Hexanes). Pure product-containing fractions of the *trans*-isomer (first eluting peak) were combined and concentrated under reduced pressure to afford **26b** as a clear oil (1.1 g, 38% yield). Pure product-containing fractions of the *cis*-isomer (second eluting peak) were combined and concentrated under reduced pressure to afford **26b** as a clear oil (1.1 g, 38% yield). Pure product-containing fractions of the *cis*-isomer (second eluting peak) were combined and concentrated under reduced pressure to afford **26c** as a clear oil (1.1 g, 38% yield). **26b**: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm 1.30 - 1.56 (m, 15 H) 3.39 (d, *J*=12.80 Hz, 1 H) 3.51 (d, *J*=11.04 Hz, 1 H) 3.70 - 3.93 (m, 1 H) 4.00 - 4.12 (m, 3 H) 4.37 (d, *J*=15.31 Hz, 1 H) 7.46 (d, *J*=8.53 Hz, 1 H) 7.83 (d, *J*=8.53 Hz, 1 H) 8.55 (s, 1 H); *m/z* 345.2 [MH-*t*-Bu]<sup>+</sup>. **26c:** <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ ppm 1.33 - 1.44 (m, 15 H) 3.23 (d, *J*=11.80 Hz, 1 H) 3.45 - 3.61 (m, 2 H) 3.93 - 4.04 (m, 3 H) 4.29 - 4.49 (m, 1 H) 7.44 (d, *J*=8.53 Hz, 1 H) 7.79 (d, *J*=6.53 Hz, 1 H) 8.51 (s, 1 H); *m/z* 345.2 [MH-*t*-Bu]<sup>+</sup>.

#### 3-Chloro-4-(((3R,4S)-4-((5-chloropyridin-2-yl)thio)-3-hydroxy-3-(hydroxymethyl)pyrrolidin-1-

yl)sulfonyl)benzonitrile (26d): To a solution of 26b (0.800 g, 1.99 mmol) in DCM (3.0 mL) was added TFA (7.69 mL, 100 mmol) and the resulting reaction mixture was stirred for 0.5 h. Following this duration, the reaction mixture was concentrated and treated with saturated aqueous NaHCO<sub>3</sub>. The resulting suspension was diluted with THF (10 mL). A mixture of 2-chloro-4-cyanobenzene-1-sulfonyl chloride (0.921 g, 3.90 mmol) in THF (10 mL) was added dropwise and the reaction mixture was allowed to stir at RT for 0.5 h. The reaction mixture was diluted with H<sub>2</sub>O and extracted with EtOAc. The organics were washed with brine, dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was purified by silica gel chromatography (0-60% EtOAc:Hexanes) to afford **26d** as a semi-solid (853 mg, 82% yield); m/z 460.1 [MH]<sup>+</sup>.

# **3-Chloro-4-(((3***S***,4***S***)-4-((5-chloropyridin-2-yl)thio)-3-hydroxy-3-(hydroxymethyl)pyrrolidin-1yl)sulfonyl)benzonitrile (26dd)** was prepared from **26c** analogously to the synthesis of **26d** from **26b** to afford **26dd** as a white solid (390 mg, 85% yield). <sup>1</sup>H NMR (400MHz, DMSO-*d*<sub>6</sub>) δ ppm 8.50 (s, 1H), 8.37 (s, 1H), 8.15 (d, *J*=8.0 Hz, 1H), 8.10 - 8.03 (m, 1H), 7.76 (dd, *J*=2.3, 8.5 Hz, 1H), 7.39 (d, *J*=8.5 Hz, 1H), 5.63 (s, 1H), 5.18 - 5.04 (m, 1H), 4.27 (t, *J*=9.2 Hz, 1H), 4.21 - 4.13 (m, 1H), 3.67 (d, *J*=10.3 Hz, 1H), 3.54 - 3.40 (m, 2H), 3.36 (br. s., 2H); *m/z* 460.3 [MH]<sup>+</sup>.

4-(((3S,4S)-3-(Azidomethyl)-4-((5-chloropyridin-2-yl)thio)-3-hydroxypyrrolidin-1-yl)sulfonyl)-3chlorobenzonitrile (26e): To a solution of 26d (0.280 g, 0.608 mmol) in DCM (8.0 mL) was added TEA (0.153 mL, 1.09 mmol) followed by MsCl (0.057 mL, 0.730 mmol) and the reaction mixture was stirred at RT for 10 min. Following this duration, the reaction mixture was diluted with H<sub>2</sub>O and extracted with DCM. The combined organics were dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under

reduced pressure. The residue was dissolved in DMF (5.0 mL), to which was added NaN<sub>3</sub> (0.099 g, 1.52 mmol) and the reaction mixture was stirred at 80 °C for 40 min. The reaction mixture was subsequently cooled to ambient temperature, diluted with H<sub>2</sub>O and extracted with EtOAc. The combined organics were washed with H<sub>2</sub>O, dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel chromatography (0-25% EtOAc:Hexanes) to afford **26e** as a colorless wax (115 mg, 39% yield); *m/z* 485.1 [MH]<sup>+</sup>.

**4-(((3***R***,4***S***)-3-(Azidomethyl)-4-((5-chloropyridin-2-yl)thio)-3-hydroxypyrrolidin-1-yl)sulfonyl)-3-chlorobenzonitrile (26ee)** was prepared from **26dd** analogously to synthesis of **26e** from **26d** to afford **26ee** as a colorless wax (343 mg, 83% yield). <sup>1</sup>H NMR (400MHz, DMSO-d<sub>6</sub>) δ ppm 8.49 (d, *J*=1.8 Hz, 1H), 8.38 (s, 1H), 8.15 (d, *J*=8.3 Hz, 1H), 8.10 - 8.02 (m, 1H), 7.79 (dd, *J*=2.3, 8.5 Hz, 1H), 7.42 (d, *J*=8.5 Hz, 1H), 6.16 (s, 1H), 4.28 (t, *J*=8.8 Hz, 1H), 4.12 (t, *J*=8.5 Hz, 1H), 3.64 - 3.43 (m, 5H); *m/z* 485.3 [MH]<sup>+</sup>.

*tert*-Butyl (((3*S*,4*S*)-1-((2-chloro-4-cyanophenyl)sulfonyl)-4-((5-chloropyridin-2-yl)thio)-3hydroxypyrrolidin-3-yl)methyl)carbamate (26f): To a solution of 26e (0.115 g, 0.237 mmol) in THF (4.0 mL) was added a 1.0 M solution of PMe<sub>3</sub> in THF (0.474 mL, 0.474 mmol) and the resulting reaction mixture was stirred at RT for 24 h. Following this duration, the reaction mixture was loaded onto a SCX cartridge and flushed with MeOH followed by 2 M NH<sub>3</sub> in MeOH. The filtrate was concentrated under reduced pressure to afford 92 mg (85% yield) of the requisite amine as a colorless wax (m/z 459.1 [MH]<sup>+</sup>). The amine (92 mg, 0.200 mmol) was dissolved in THF (4.0 mL) to which was added Boc<sub>2</sub>O (0.060 g, 0.260 mmol) followed by TEA (0.056 mL, 0.401 mmol) and the reaction mixture was stirred at RT for 0.5 h. Following this duration, the reaction mixture was diluted with H<sub>2</sub>O and extracted with EtOAc. The combined organics were dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was purified by silica gel chromatography (0-30% EtOAc:Hexanes) to afford **26f** as a colorless semi-solid (75 mg, 67% yield); m/z 559.2 [MH]<sup>+</sup>.

# *tert*-Butyl(((*3R*,4*S*)-1-((2-chloro-4-cyanophenyl)sulfonyl)-4-((5-chloropyridin-2-yl)thio)-3hydroxy-pyrrolidin-3-yl)methyl)carbamate (26ff) was prepared from 26ee analogously to the synthesis of 26f from 26e to afford 26ff as a colorless semi-solid (178 mg, 77% yield). <sup>1</sup>H NMR (400 MHz, DMSO*d*<sub>6</sub>) δ ppm 1.38 (d, *J*=13.30 Hz, 9 H) 3.18 (d, *J*=4.77 Hz, 2 H) 3.48 (s, 1 H) 3.65 (d, *J*=10.54 Hz, 1 H) 4.01 - 4.15 (m, 2 H) 4.31 (s, 1 H) 5.35 - 5.61 (m, 1 H) 7.39 (d, *J*=8.53 Hz, 1 H) 7.76 (dd, *J*=8.53, 2.51 Hz, 1 H) 8.01 - 8.09 (m, 1 H) 8.16 (d, *J*=8.03 Hz, 1 H) 8.36 (s, 1 H) 8.49 (s, 1 H); *m/z* 559.5 [MH]<sup>+</sup>.

*tert*-Butyl (((3S,4S)-1-((2-chloro-4-cyanophenyl)sulfonyl)-4-((5-chloropyridin-2-yl)sulfonyl)-3hydroxypyrrolidin-3-yl)methyl)carbamate (26g): To a solution of 26f (0.075 g, 0.134 mmol) in DCM (4.0 mL) was added *m*-CPBA (0.077 g, 0.335 mmol) and the reaction mixture was stirred at RT for 3 h. Following this duration, the reaction mixture was quenched with 10% aqueous Na<sub>2</sub>S<sub>2</sub>SO<sub>3</sub> and the layers were separated. The organic layer was washed with saturated aqueous NaHCO<sub>3</sub>, dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was purified by silica gel chromatography (0-35% EtOAc:Hexanes) to afford **26g** as a colorless semi-solid (78 mg, 98% yield); *m/z* 491.1 [MH-Boc]<sup>+</sup>.

# tert - Butyl(((3R, 4S) - 1 - ((2 - chloro - 4 - cyanophenyl) sulfonyl) - 4 - ((5 - chloropyridin - 2 - yl) sulfonyl) - 3 - ((2 - chloropyridin - 2 - yl) sulfonyl) - ((2 - chloropyridin - 2 - yl) sulfonyl)

**hydroxypyrrolidin-3-yl)methyl)carbamate (26gg)** was prepared from **26ff** analogously to the synthesis of **26g** from **26f** to afford **26gg** as a white solid (150 mg, 77% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ ppm 1.35 (s, 9 H) 3.38 (br. s., 4 H) 3.94 (d, *J*=9.29 Hz, 1 H) 4.11 (br. s., 1 H) 4.37 (br. s., 1 H) 6.94 (br. s., 1 H) 8.01 (d, *J*=8.53 Hz, 1 H) 8.05 (d, *J*=8.28 Hz, 1 H) 8.14 (d, *J*=8.03 Hz, 1 H) 8.25 - 8.31 (m, 1 H) 8.36 (s, 1 H) 8.82 (s, 1 H); *m/z* 491.4 [MH-Boc]<sup>+</sup>.

#### 4-(((3S,4S)-3-(Aminomethyl)-4-((5-chloropyridin-2-yl)sulfonyl)-3-hydroxypyrrolidin-1-

yl)sulfonyl)-3-chlorobenzonitrile (26): To a solution of 26g (0.078 g, 0.132 mmol) in DCM (1.0 mL) was added TFA (0.305 mL, 3.96 mmol) and the resulting reaction mixture was stirred at RT for 0.5 h. Following this duration, the reaction mixture was concentrated under reduced pressure. The residue was diluted with H<sub>2</sub>O and the resulting solution was basified with saturated aqueous NaHCO<sub>3</sub>. The white

#### Journal of Medicinal Chemistry

precipitate was filtered, washed with H<sub>2</sub>O and dried under vacuum to afford **26** as a white solid (37 mg, 54% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ ppm 2.82 - 3.00 (m, 2 H) 3.43 (d, *J*=10.29 Hz, 1 H) 3.58 (d, *J*=10.04 Hz, 1 H) 3.69 (d, *J*=11.54 Hz, 1 H) 3.91 (d, *J*=4.02 Hz, 1 H) 4.29 (d, *J*=6.27 Hz, 1 H) 5.62 - 5.98 (m, 1 H) 8.04 (dd, *J*=13.05, 8.53 Hz, 2 H) 8.10 - 8.17 (m, 1 H) 8.29 - 8.40 (m, 2 H) 8.90 (s, 1 H); *m/z* 491.3 [MH]<sup>+</sup>.

## 4-(((3R,4S)-3-(Aminomethyl)-4-((5-chloropyridin-2-yl)sulfonyl)-3-hydroxypyrrolidin-1-

yl)sulfonyl)-3-chlorobenzonitrile (27) was prepared from 26gg analogously to the synthesis of 26 from 26g to afford 27 as a white solid (61 mg, 50% yield). <sup>1</sup>H NMR (400MHz, DMSO-d<sub>6</sub>) δ ppm 8.84 (s, 1H), 8.36 (s, 1H), 8.28 (d, *J*=8.5 Hz, 1H), 8.14 (d, *J*=8.0 Hz, 1H), 8.03 (dd, *J*=8.3, 19.8 Hz, 2H), 4.49 (t, *J*=7.5 Hz, 1H), 4.16 - 4.02 (m, 1H), 3.88 (d, *J*=9.3 Hz, 1H), 3.51 (d, *J*=10.0 Hz, 1H), 3.40 (br. s., 2H), 2.81 (d, *J*=8.3 Hz, 2H); <sup>13</sup>C NMR (101 MHz, DMSO-d<sub>6</sub>) δ ppm 44.7, 57.6, 64.3, 73.2, 77.9, 116.8, 117.3, 124.3, 132.6, 136.0, 136.2, 139.1, 140.4, 142.7, 145.2, 148.8, 155.7; LRMS: *m/z* 491.3 [MH]<sup>+</sup>. HRMS: *m/z* 491.0018 (observed), 491.0017 (calculated).

#### ■ ASSOCIATED CONTENT:

The Supporting Information is available free of charge on the ACS Publications website at DOI: \*\*\*

Included are experimental details regarding the calcium flux FLIPR, CLND solubility, hERG, Ca<sub>v</sub>1.2, phospholipidosis, 3A4 TDI, 3A4 RI assays; *in vivo* pharmacokinetics studies; rat pulmonary edema model and *in vivo* IC<sub>50</sub> calculations; complete experimental procedures and compound characterization data (NMR, MS); VCD analyses of Compounds **7** and **10**; crystal structure of Compound **19** (PDF)

Molecular formula strings and associated biological data (CSV)

#### ■ AUTHOR INFORMATION:

#### **Corresponding Author**

\*E-mail for J.E.P.: joseph.e.pero@gsk.com. Phone: (484)-923-3836

## ORCIDID

Joseph E. Pero: 0000-0003-2674-4713

# Notes

The authors declare the following competing financial interest(s): Authors affiliated with GlaxoSmithKline have received compensation in the form of salary and stock.

# ■ ACKNOWLEDGEMENTS:

The authors thank Amy Gibble and Ashley Leister for assistance in the chromatographic separation of compounds, William Clegg, Michael Probert and Roy Copley for X-ray crystallographic studies, and Douglas Minick for VCD analysis.

# ■ ABBREVIATIONS USED:

AUC, area under the (concentration) curve; CAD, cationic amphiphilic drug; CHI, chromatographic hydrophobicity index; CL, total clearance; CLND, Chemiluminescence Nitrogen Detection; C<sub>max</sub>, maximum concentration; C<sub>min</sub>, minimum concentration; Cmpd, compound; CYP, cytochrome P450; DCM, dichloromethane; DDI, drug-drug interaction; DMF, *N*,*N*-dimethylformamide; DNAUC, dosenormalized area under the (concentration) curve; F, oral bioavailability; FLIPR, fluorometric imaging plate reader; f<sub>u</sub>, fraction unbound; h, hour(s); GSH, glutathione; HCl, hydrochloric acid; hERG, human Ether-à-go-go-Related Gene; HF, heart failure; HPLC, high pressure liquid chromatography; HUVEC, human umbilical vein endothelial cells; IAM, immobilized artificial membrane; IC<sub>50</sub>, half-maximal inhibitory concentration; i.v., intravenous administration; LE, ligand efficiency; LLE, lipophilic ligand efficiency; *m*-CPBA, *meta*-chloroperoxybenzoic acid; MEC, minimum effective concentration; MI, myocardial infarction; mpk, milligrams per kilogram; min, minutes; MRT, mean resonance time; MsCl, mesyl chloride; nM, nanomolar; NMO, 4-methylmorpholine *N*-oxide; Perm, permeability; PK, pharmacokinetics; p.o., per os (oral) administration; PSA, polar surface area; PPTS, pyridinium *p*-toluenesulfonate; PVP, pulmonary venous pressure; RI, reversible inhibitor; RT, room temperature;

SAR, structure-activity relationships; SEM, standard error of mean; TBME, *tert*-butyl methyl ether; TDI, time-dependent inhibition; TEA, triethylamine; TFA, trifluoroacetic acid; THF, tetrahydrofuran; TRPV4, Transient Receptor Potential Vanilloid-4; μM, micromolar; VCD, vibrational circular dichroism; V<sub>dss</sub>, volume of distribution.

■ REFERENCES:

- (1) McMurray, J.J.; Pfeffer, M.A. Heart Failure. Lancet. 2005, 365, 1877-1889.
- (2) Dickstein, K.; Cohen-Solal, A.; Filippatos, G. ESC Guideline for the Diagnosis and Treatment of Acute and Chronic Heart Failure 2008: The Task Force for the Diagnosis and Treatment of Acute and Chronic Heart Failure 2008 of the European Society of Cardiology. Developed in Collaboration with the Heart Failure Association of the ESC (HFA) and Endorsed by the European Society of Intensive Care Medicine (ESICM). *Eur. Heart J.* **2008**, *29*, 2388-2442.
- (3) Al-Mohammad, A.; Davis, M.; Dawda, P.; Foley, P.; Fuat, A.; Gilmour, J.; Hardman, S.; Leyva, F.; McIntyre, H.; Mant, J.; Mindham, R.; Price, A.; Collinson, P.; Cowie, A. Chronic Heart Failure: National Clinical Guideline for Diagnosis and Management in Primary and Secondary Care: Partial Update. *NCIE Clinical Guidelines*, **2010**, *108*, 19-24.
- (4) Heidenreich, P.A.; Albert, N.M.; Allen, L.A.; Bluemke, D.A.; Butler, J.; Fonarow, G.C.; Ikonomidis, J.S.; Khavjou, O.; Konstam, M.A.; Maddox, T.M.; Nichol, G.; Pham, M.; Pina, I.L.; Trogdon, J.G. Forecasting the Impact of Heart Failure in the United States: A Policy Statement from the American Heart Association. *Circulation: Heart Failure* 2013, *6*, 606-619.
- (5) Alvarez, D.F.; King, J.A.; Weber, D.; Addison, E.; Liedtke, W.; Townsley, M.I. Transient Receptor Potential Vanilloid 4–Mediated Disruption of the Alveolar Septal Barrier. *Circ. Res.* 2006, 99, 988-995.
- (6) Jian, M.-Y.; King, J.A.; Al-Mehdi, A.-B.; Liedtke, W.; Townsley, M.I. High Vascular Pressure-Induced Lung Injury Requires P450 Epoxygenase-Dependent Activation of TRPV4. Am. J. Respir. Cell Mol. Biol. 2008, 38, 386-392.

- (7) Ware, L.B.; Matthay, M.A. Acute Pulmonary Edema. N. Eng. J. Med. 2005, 353, 2788-2796.
- (8) Everaerts, W.; Nilius, B.; Owsianik, G. The Vanilloid Transient Receptor Potential Channel TRPV4: From Structure to Disease. *Prog. Biophys. Mol. Biol.* **2010**, *103*, 2-17.
- (9) Vincent, F.; Duncton, A.J. TRPV4 Agonists and Antagonists. *Curr. Top. Med. Chem.* 2011, *11*, 2216-2226.
- (10) Bakthavatchalam, R.; Kimball, S.D. Modulators of Transient Receptor Potential Ion Channels.
   *Annu. Rep. Med. Chem.* 2010, 45, 37-53.
- (11) Thorneloe, K. S.; Cheung, M.; Bao, W.; Alsaid, H.; Lenhard, S.; Jian, M.-Y.; Costell, M.; Maniscalco-Hauk, K.; Krawiec, J. A.; Olzinski, A.; Gordon, E.; Lozinskaya, I.; Elefante, L.; Qin, P.; Matasic, D. S.; James, C.; Tunstead, J.; Donovan, B.; Kallal, L.; Waszkiewicz, A.; Vaidya, K.; Davenport, E. A.; Larkin, J.; Burgert, M.; Casillas, L. N.; Marquis, R. W.; Ye, G.; Eidam, H. S.; Goodman, K. B.; Toomey, J. R.; Roethke, T. J.; Jucker, B. M.; Schnackenberg, C. G.; Townsley, M. I.; Lepore, J. J.; Willette, R. N. An Orally Active TRPV4 Channel Blocker Prevents and Resolves Pulmonary Edema Induced by Heart Failure. *Sci. Transl. Med.* 2012, *4*, 159ra148.
- (12) McMurray, J.J. Clinical practice. Systolic Heart Failure. N. Engl. J. Med. 2010, 362, 228–238.
- (13) Felker, G.M.; Lee, K.L.; Bull, D.A.; Redfield, M.M.; Stevenson, L.W.; Goldsmith, S.R.; LeWinter, M.M.; Deswal, A.; Rouleau, J.L.; Ofili, E.O.; Anstrom, K.J.; Hernandez, A.F.; McNulty, S.E.; Velazquez, E.J.; Kfoury, A.G.; Chen, H.H.; Givertz, M.M.; Semigran, M.J.; Bart, B.A.; Mascette, A.M.; Braunwald, E.; O'Connor, C.M. NHLBI Heart Failure Clinical Research Network, Diuretic Strategies in Patients with Acute Decompensated Heart Failure. *N. Engl. J. Med.* 2011, *364*, 797–805.
- (14) Hilfiker, M.A.; Hoang, T.H.; Cornil, J.; Eidam, H.S.; Matasic, D.S.; Roethke, T.J.; Klein, M.; Thorneloe, K.S.; Cheung, M. Optimization of a Novel Series of TRPV4 Antagonists with In Vivo Activity in a Model of Pulmonary Edema. *ACS Med. Chem. Lett.* 2013, *4*, 293-296.

- (15) Brooks, C.; Cheung, M.; Eidam, H.S.; Goodman, K.B.; Hammond, M.; Hilfiker, M.A.; Patterson, J.R.; Stoy, P.; Ye, G. Spiro[cyclohexane-oxazolidinone] Derivatives as TRPV4 Antagonists and their Preparation. Patent WO2012174342 A1, 2012.
- (16) Brooks, C.; Cheung, M.; Eidam, H.S.; Goodman, K.B.; Hammond, M.; Hilfiker, M.A.; Hoang, T.H.; Patterson, J.R.; Stoy, P.; Ye, G. Preparation of Spirocarbamate Compounds as TRPV4 Antagonists. Patent WO2013012500 A1, 2013.
- (17) Skerratt, S.E.; Mills, J.E.J.; Mistry, J. Identification of False Positives in "HTS hits to Lead": The Application of Bayesian Models in HTS Triage to Rapidly Deliver a Series of Selective TRPV4 Antagonists. *Med. Chem. Commun.* 2013, *4*, 244-251.
- (18) Ligand efficiency is defined as  $LE = 1.37*(pIC_{50})/HAC$ , in which HAC = heavy atom count.
- (19) Lipophilic ligand efficiency is defined as LLE = *p*IC<sub>50</sub> cLogP. For more details, see: Ryckmans, T.; Edwards, M.P.; Horne, V.A.; Correia, A.M.; Owen, D.R.; Thompson, L.R.; Tran, I.; Tutt, M.F.; Young, T. Rapid Assessment of a Novel Series of Selective CB(2) Agonists Using Parallel Synthesis Protocols: A Lipophilic Efficiency (LipE) Analysis. *Bioorg. Med. Chem. Lett.* 2009, *19*, 4406-4409.
- (20) (a) Brnardic, E.J.; Ye, G.; Brooks, C.; Donatelli, C.; Barton, L.; McAtee, J.; Sanchez, R.M.; Shu, A.; Erhard, K.; Terrell, L.; Graczyk-Millbrandt, G.; He, Y.; Costell, M.H.; Behm, D.J.; Roethke, T.; Stoy, P.; Holt, D.A.; Lawhorn, D.G. Discovery of Pyrrolidine Sulfonamides as Selective and Orally Bioavailable Antagonists of Transient Receptor Potential Vanilloid-4 (TRPV4). *J. Med. Chem.* DOI: 10.1021/acs.jmedchem.8b01317. (b) Brnardic, E.J.; Brooks, C.A.; Lawhorn, B.G.; Ye, G.; Barton, L.S.; Budzik, B.W.; Matthews, J.M.; McAtee, J.J.; Patterson, J.R.; Pero, J.E.; Sanchez, R.; Sender, M.R.; Terrell, L.R.; Behm, D.J.; Thomas, J.V. Preparation of Pyrrolidine Sulfonamide Compounds as TRPV4 Antagonists. Patent WO2018055524 A1, **2018**.
- (21) TRPV4 potency was determined by measuring the blockade of agonist GSK634775-induced calcium immobilization through human, recombinant TRPV4 channels in HEK cells on a fluorescent light imaging plate reader (FLIPR). The reported potencies are an average of at least

two measurements with a SEM  $\pm$  0.1 for the *p*IC<sub>50</sub>. For further details, please see the Supporting Information.

- (22) All cLogP values presented herein are calculated from ChemAxon JChem for Excel software package based on the following method: Viswanadhan, V.D.; Ghose, A.K.K.; Revankar, G.R.; Robins, R.K. Atomic Physicochemical Parameters for Three-Dimensional Structure Directed Quantitative Structure-Activity Relationships. *J. Chem. Inf. Comput. Sci.* **1989**, *29*, 163-172.
- (23) Clearance adjusted for unbound fraction is defined as  $CL/f_u$  in which CL is the total body clearance (mL/min/kg) and  $f_u$  is the unbound fraction as a percent (%).
- (24) Permeability values were provided by an artificial membrane permeability assay. For details, please see the Supporting Information.
- (25) Malon, P.; Kobrinskaya, R.; Keiderling, T.A. Vibrational Circular Dichroism of Polypeptides XII. Re-evaluation of the Fourier Transform Vibrational Circular Dichroism of Poly-gamma-Benzyl-L-Glutamate. *Biopolymers* 1988, 27, 733-746.
- (26) (a) Sebastiano, M.R.; Doak, B.C.; Backlund, M.; Poongavanam, V.; Over, B.; Ermondi, G.; Caron, G.; Matsson, P.; Kihlberg, J. Impact of Dynamically Exposed Polarity on Permeability and Solubility of Chameleonic Drugs Beyond the Rule of 5. *J. Med. Chem.* 2018, *61*, 4189-4202.
  (b) Ertl, P.; Rohde, B.; Selzer, P. Fast Calculation of Molecular Polar Surface Area as a Sum of Fragment-Based Contributions and its Application to the Prediction of Drug Transport Properties. *J. Med. Chem.* 2000, *43*, 3714-3717. (c) Kelder, J.; Grootenhuis, P.D.J.; Bayada, D.M.; Delbressine, L.P.C.; Ploemen, J.-P. Polar Molecular Surface as a Dominating Determinant for Oral Absorption and Brain Penetration of Drugs. *Pharm. Res.* 1999, *16*, 1514-1519. (c) Palm, K.; Stenberg, P.; Luthman, K.; Artursson, P. Polar Molecular Surface Properties Predict the Intestinal Absorption of Drugs in Humans. *Pharm. Res.* 1997, *14*, 568-571.
- (27) For VCD analysis of Compounds 7 and 10, please see the Supporting Information.

- (28) Cheng, H.Y.; Jusko, W.J. Mean Residence Time Concepts for Pharmacokinetic Systems with Nonlinear Drug Elimination Described by Michaelis-Menten Equation. *Pharm. Res.* 1988, 5, 156-164.
- (29) While the synthetic sequence illustrated in Scheme 1 generally proved to be the most reliable and diastereoselective approach toward the desired sulfone pyrrolidine sulfonamides, alternative routes were also utilized for select compounds. Please see the Supporting Information section for further details.
- (30) Alcaraz, L; Crindland, A.; Kinchin, E. Novel Conversion of 1,2-Disubstituted *cis*-Epoxides to One-Carbon Homologated Allylic Alcohols Using Dimethylsulfonium Methylide. *Org. Lett.* 2001, *3*, 4051-4053.
- (31) A number of examples were prepared through this less-diastereoselective approach, which was established prior to the highly-diastereoselective sequence depicted in Scheme 1. For further details, please see the Supporting Information.
- (32) PK studies in rat liver microsomes suggested that oxidative metabolism (intrinsic clearance) of the aryl sulfone functionality decreases as the electron-withdrawing effect(s) of its substituent(s) decreases.
- (33) The sulfoximine functionality of Compound 23 is chiral. For details concerning the elucidation of its stereochemical configuration, please refer to the Supporting Information.
- (34) The sulfonimidamide functionality of Compound **24** is chiral. For details concerning the elucidation of its stereochemical configuration, please refer to the Supporting Information.
- (35) Kinetic aqueous solubility measurements were measured using Chemiluminescence Nitrogen Detection (CLND) quantification. For further details, please refer to Supporting Information.
- (36) Halliwell, W.H. Cationic Amphiphilic Drug-Induced Phospholipidosis. *Toxicol. Pathol.* 1997, 25, 53-60.
- (37) The response in the phospholipidosis assay is the percent-increase in accumulation of a fluorescently-labeled phospholipid in cultured HepG2 cells after 72 hours of incubation with a

test compound relative to the accumulation of the phospholipid in the absence of the test compound. For further details, please refer to the Supporting Information.

- (38) Mortuza, G.B.; Neville, W.A.; Delaney, J.; Waterfield, C.J.; Camilleri, P. Characterisation of a Potential Biomarker of Phospholipidosis from Amiodarone-Treated Rats. *Biochimica et Biophysica Acta 1631* 2003, 136-146.
- (39) Bunally, S.; Young, R.J. The Role and Impact of High Throughput Biomimetic Measurements in Drug Discovery. ADMET & DMPK 2018, 6, 74-84.
- (40) Willette, R. N.; Bao, W.; Nerurkar, S.; Yue, T-I.; Doe, C. P.; Stankus, G.; Turner, G. H.; Ju, H.; Thomas, H.; Fishman, C. E.; Sulpizio, A.; Behm, D. J.; Hoffman, S.; Lin, Z.; Lozinskaya, I.; Cassillas, L. N.; Lin, M.; Trout, R. E. L.; Bartholomew, J. V.; Thorneloe, K.; Lashinger, E. S. R.; Figueroa, D. J.; Marquis, R.; Xu, X. Systematic Activation of the Transient Receptor Potential Vanilloid Subtype 4 Channel Causes Endothelial Failure and Circulatory Collapse: Part 2. *J. Pharmacol. Exp. Ther.* 2011, *338*, 408-409.
- (41) Mendoza, S. A.; Fang, J.; Gutterman, D. D.; Wilcox, D. A.; Bubolz, A. H.; Li, R.; Suzuki, M.; Zhang, D. X. TRPV4-Mediated Endothelial Ca<sup>2+</sup> Influx and Vasodilation in Response to Shear Stress. *Am. J. Physiol. Heart Circ. Physiol.* 2010, *298*, H466.
- (42) All animal studies were conducted in accordance with the GlaxoSmithKline Policy on the Care, Welfare, and Treatment of Laboratory Animals and were reviewed by the Institutional Animal Care and Use Committee at GlaxoSmithKline.
- (43) For details on the % inhibition calculations and statistical analyses, please refer to the Supporting Information.
- (44) For details on the calculation of the *in vivo*  $IC_{50}$  values, please refer to the Supporting Information.
- (45) Brnardic, E.J.; Brooks, C.A.; Lawhorn, B.J.; Li, P.; Matthews, J.M.; McAtee, J.J.; Pero, J.E.; Sender, M.R.; Terrell, L.R.; Behm, D.J. TRPV4 Antagonists. Patent WO 2018055526 A1, 2018.

Table of Contents graphic.

