Contents lists available at ScienceDirect

European Journal of Medicinal Chemistry

journal homepage: http://www.elsevier.com/locate/ejmech



Original article

Synthesis and anticonvulsant activity of N-3-arylamide substituted 5, 5-cyclopropanespirohydantoin derivatives

Xianran He^a, Min Zhong^b, Tao Zhang^a, Wen Wu^a, Zhongyuan Wu^a, Jin Yang^a, Yuling Xiao^a, Yuanhu Pan^a, Guofu Qiu^a, Xianming Hu^{a,*}

^a State Key Laboratory of Virology, College of Pharmacy, Wuhan University, Luojiashan Road, Wuhan 430072, China ^b Hubei Research Institute of Chemistry, Wuhan 430074, China

A R T I C L E I N F O

Article history: Received 27 May 2010 Received in revised form 8 August 2010 Accepted 23 September 2010 Available online 1 October 2010

Keywords: N-3-arylamide substituted 5,5-cyclopropanespirohydantoin Anticonvulsant activity MES test scPTZ test

ABSTRACT

In the present study on the development of new anticonvulsants, twenty new N-3-arylamide substituted 5,5-cyclopropanespirohydantoin derivatives were synthesized and tested for anticonvulsant activity using the maximal electroshock (MES), subcutaneous pentylenetetrazole (*sc*PTZ) screens, which are the most widely employed seizure models for early identification of candidate anticonvulsants. Their neurotoxicity was determined applying the rotorod test. Three compounds **5d**, **5j** and **5t** showed promising anticonvulsant activities in both models employed for anticonvulsant evaluation. The most active compound **5j** showed the MES-induced seizures with ED₅₀ value of 9.2 mg/kg and TD₅₀ value of 421.6 mg/kg after intraperitoneally injection to mice, which provided compound **5j** with a protective index (TD₅₀/ED₅₀) of 45.8 in the MES test.

Crown Copyright © 2010 Published by Elsevier Masson SAS. All rights reserved.

1. Introduction

Epilepsy is a common neurologic affection characterized by excessive temporary neuronal discharge that affects about 1% of the world's population [1]. Despite the availability of many antiepileptic drugs (AEDs), there is still an urgent need for the development of more effective and safer AEDs, since about 30% of epileptic patients are not seizure-free with the existing AEDs [2]. Besides, many AEDs such as phenobarbital, phenytoin, carbamazepine, vigabatrin, valproate, felbamate, and lamotrigine, which are effective toward only 60–80% of patients have some undesirable side effects, such as headache, nausea, anorexia, ataxia, hepatotoxicity, drowsiness, gastrointestinal disturbance, gingival hyperplasia, and hirsutism [3–6]. Thus, there is an enormous need for the development of novel AEDs with fewer side effects and more effectives.

The SAR studies of clinically available AEDs and other anticonvulsant active compounds showed that most of these compounds included 5- or 6-member cyclic imides moiety in their molecules (Fig. 1). Hydantoins, a class of cyclic imides, have been demonstrated to possess good anticonvulsant property [7,8]. Depending on the nature of substitution on the hydantoin ring, a wide range of other pharmacological properties, e.g., antihypertensive [9], herbicidal [10], antitumor [11], anti-HIV [12], antibacterial [13] and antiviral [14] activities, have also been identified. Spirohydantoin analogs, such as 5,5-cyclohexanespirohydantoins [15], 5,5-cycloheptanespirohydantoins [15], and 5,5-cyclooctanespirohydantoins [15], have been synthesized and shown to have anticonvulsant activity. Previous studies from our laboratory have demonstrated the potent anticonvulsant activity among the N-3-substituted 5,5-cyclo-propanespirohydantoin derivatives, from which the compound 1,1-dimethyl-6-(4-fluorophenyl)-4,6-diazaspiro[2.4]heptane-5,7-dione showed more potent anticonvulsant activity at the dose of 7.5 mg/kg than the standard drug phenytoin sodium [16].

In this paper, we report on the synthesis, structural characterization, and preliminary evaluation of the anticonvulsant properties of a number of N-3-arylamide substituted 5,5-cyclopropanespirohydantoin derivatives (5a-t). Compounds **5d**, **5j** and **5t** which displayed the remarkable activity, were chosen for quantification of the pharmacological parameters (ED₅₀ and TD₅₀). It was interesting to see, compound with a small lipophilic group like fluorine substituents at the para-position of the phenyl ring of N-3-arylamide (**5h**) showed promising anticonvulsant activities, which is in agreement with the reported SAR for spirohydantoins [17]. It was also noteworthy that, compound with a CF₃ group at position-4 of the phenyl ring (**5j**) resulted in the most activity of this series. This observation indicated that CF₃ group is recognized

^{*} Corresponding author. Tel.: +86 27 68753532; fax: +86 27 68754629. *E-mail address*: hexianran@yahoo.com.cn (X. Hu).

^{0223-5234/\$ –} see front matter Crown Copyright © 2010 Published by Elsevier Masson SAS. All rights reserved. doi:10.1016/j.ejmech.2010.09.052

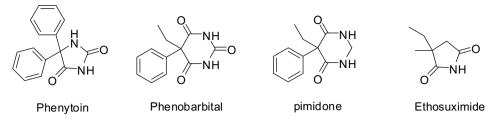


Fig. 1. Structure of clinically available AEDs include cyclic imides moiety.

as pharmacophore and plays a significant role in new drug design, including anticonvulsant active molecules [18]. The pharmacological activity of various N-3-arylamide substituted 5,5-cyclopropane-spirohydantoin derivatives can help better establish their structure-activity relationship (SAR).

2. Chemistry

The synthesis of the compounds 5a-t were accomplished according to the reaction sequence illustrated in Scheme 1. Diethyl 2,2-dimethylcyclopropane-1,1-dicarboxylate 1 was synthesized by a Michael initiated ring closure (MIRC) reaction according to our previous studies [19,20]. Then monoester 2 was obtained after monosaponification in a 1 N NaOH/ethanol (1.1 equiv) solution at room temperature for 12 h [20,21]. This was then converted to corresponding acyl azide by using ethyl chloroformate in the presence of N-methyl morpholine (NMM) followed by reaction with sodium azide in a one-pot synthesis. α-Carboethoxy isocyanate 3 was successfully generated by a Curtius reaction in situ on heating the acyl azide in toluene solution at 75 °C. Isocyanate 3 was allowed to react directly with various arylacethydrazide without isolation. The desired semicarbazides 4 were readily obtained. Finally, those semicarbazides (4a-t) cyclized on treatment with NaOH (1 equiv) in EtOH and provided N-3-arylamide substituted spirohydantoins 5 with a cyclopropane ring in good yields (yield >80%). Their chemical structures were characterized using ¹H-NMR, MS and elemental analysis techniques. The detailed physical and analytical data are listed in Section 5.

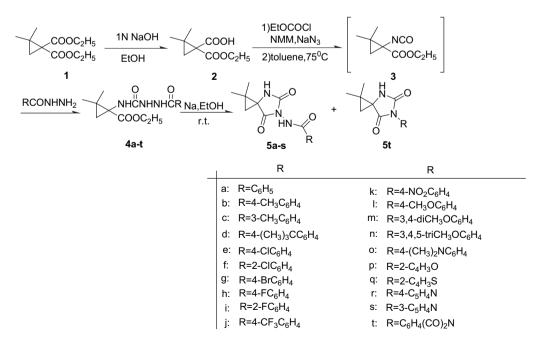
All of these compounds were prepared as racemic mixtures and no attempt was made to resolve the enantiomers.

3. Pharmacology results and discussion

The anticonvulsant activity and neurotoxicity of the synthesized compounds were evaluated following the standard procedures proposed by the NIH anticonvulsant drug development (ADD) program, via the anticonvulsant screening project (ASP). The initial evaluation (Phase I) included the maximal electroshock seizure (MES), the subcutaneous pentylenetetrazole (*sc*PTZ) and neurotoxicity.

The compounds **5a**–**t** were administrated intraperitoneally (ip) into the mice using dose of 30, 100 and 300 mg/kg and the observations were taken at two different time intervals (0.5 h and 4.0 h). Neurotoxicity was measured by the rotorod test. The calculated Log *P* (*C* log *p*) values were calculated using the software in ACD Labs 8.0 version. The results are shown in Table 1.

The initial anticonvulsant evaluation indicated that all the compounds were effective in ip MES and/or *sc*PTZ screens. In the MES test, all of the compounds showed protection in half or more of the tested mice after 0.5 h except **5p**, **5q**, **5r** and **5s**, indicative of their ability to prevent seizure spread. Compounds which were active at 100 mg/kg after 0.5 h in MES test included **5b**, **5c**, **5e**, **5h**, **5i** and **5l**



Scheme 1. General method for the synthesis of compounds 5a-t.

Table 1

Anticonvulsant activity and neurotoxocity of compounds 5a-t administered intraperitoneally to mice.

Compounds	Intraperiton	$C \log P^{b}$					
	MES ^c		scPTZ ^d		Neurotoxocity ^e		
	0.5 h	4.0 h	0.5 h	4.0 h	0.5 h	4.0 h	
5a	300	_	300	_	300	_	1.15 ± 0.69
5b	100	_	_	300	_	300	1.61 ± 0.69
5c	100	-	-	-	-	-	1.61 ± 0.69
5d	30	100	100	_	_	_	2.84 ± 0.70
5e	100	300	300	-	-	-	1.91 ± 0.70
5f	300	-	300	-	_	300	1.32 ± 0.70
5g	300	300	-	300	300	300	2.09 ± 0.73
5h	100	300	300	-	_	300	1.37 ± 0.73
5i	100	-	-	_	_	300	$\textbf{0.78} \pm \textbf{0.74}$
5j	30	30	300	300	-	-	2.12 ± 0.72
5k	300	300	300	-	-	-	1.11 ± 0.70
51	100	-	-	-	-	300	1.32 ± 0.70
5m	300	300	-	300	-	300	1.39 ± 0.70
5n	300	-	-	-	100	300	1.34 ± 0.70
50	300	300	100	-	_	_	1.57 ± 0.70
5p	_	300	-	_	100	300	-0.06 ± 0.71
5q	-	300	-	-	100	300	$\textbf{0.75} \pm \textbf{0.70}$
5r	-	300	-	-	100	300	-0.11 ± 0.69
5s	-	300	_	-	100	300	$\textbf{0.08} \pm \textbf{0.70}$
5t	30	100	300	300	-	-	$\textbf{0.68} \pm \textbf{0.82}$
Phenytoin ^f	30	30	_	-	100	100	2.52 ± 0.38
Ethosuximide ^g	_	_	100	300	_	_	$\textbf{0.38} \pm \textbf{0.46}$

^a 30,100, and 300 mg/kg of doses were administered ip. The figures in the table indicate the minimal dose whereby bioactivity was demonstrated in half or more of the mice. The animals were examined at 0.5 h and 4.0 h after injection were administered. A dash indicates an absence of activity at maximum dose administered (300 mg/kg). ^b Clog P was calculated using software ACD Labs 8.0 version.

^c Maximal electroshock test.

^d Subcutaneous pentylenetetrazole test.

^e Neurotoxocity screening(rotorod test).

^f Data from Ref. [31].

^g Data from Ref. [32].

indicative of their good ability to protect from seizure spread at a higher dose. Among these compounds, **5e** and **5h** were also active at the same dose after 4.0 h. This showed that these compounds have quick onset and long duration of action at relatively higher dose. From these series **5d**, **5j** and **5t** showed anti-MES activity at the dose of 30 mg/kg at time periods 0.5 h, the most active compound **5j** were active in the MES test both at 0.5 h and 4.0 h that was equivalent to phenytoin used as reference anticonvulsant drug.

The *sc*PTZ screen showed that compounds **5a**, **5b**, **5d**, **5e**, **5f**, **5g**, **5h**, **5j**, **5k**, **5m**, **5o** and **5t** were found to be active after 0.5 h or/and 4.0 h, the other derivatives devoid of anticonvulsant activity. Compounds **5b**, **5g**, **5j**, **5m** and **5t** were active after 4.0 h at the dose of 300 mg/kg, the other compounds showed no activity.

In the neurotoxicity screen, Compounds **5c**, **5d**, **5e**, **5j**, **5k**, **5o** and **5t** did not show any neurotoxicity in the maximum dose administered (300 mg/kg). Compounds **5n**, **5p**, **5q**, **5r** and **5s** revealed neurotoxicity at a dose of 100 mg/kg. The majority of these compounds exhibited less neurotoxic than phenytoin.

Compounds **5d**, **5j** and **5t** were selected for quantification of the pharmacological parameters (ED_{50} and TD_{50}). Results of the quantitative test for these compounds, along with the data on the standard drugs (phenytoin, carbamazepine, phenobarbital, and valproate), are reported in Table 2. In the mice MES screen, the tested compounds showed a higher protective index (PI) than all the standard drugs. In the mice ip *sc*PTZ screen, compound **5j** gave an ED_{50} of 62.1 mg/kg and a TD_{50} of 421.6 mg/kg, resulting in a high protection index (PI), that is, TD_{50}/ED_{50} , of 6.7 when compared to phenobarbital and valproate.

The results of the preliminary anticonvulsant screening revealed that N-3-arylamide substituted 5,5-cyclopropanespirohydantoin derivatives exhibit a remarkable anticonvulsant activity. The structure of this series fulfilled all the pharmacophoric structural requirements,

i.e., the hydantoin nucleus and phenyl ring provided the basic structural requirement for anticonvulsant activity [22]. Previous studies from our laboratory have demonstrated a good anticonvulsant activity of N-3-substituted 5,5-cyclopropane-spirohydantoin derivatives, the cyclopropanespirohydantoin may provided to be the pharmacophoric group of anticonvulsant property [16].

In the present studies, we have synthesized a library of compounds with cyclopropanespirohydantoin as a core fragment and at the position-3 of hydantoin ring we have introduced different arylamide substituents. The results of bioevaluation led to an understanding of the structure-activity relationship (SAR) of these compounds. The phenyl ring substituted with Cl or Br in paraposition (5e, 5g) lead to compounds with more anticonvulsant activity than ortho-position (5f, 5i). On the other hand, the 4-fluoro substituent derivative (5h) was more potent and less neurotoxic than chloro, bromo and methyl derivatives. It may be proved that substitution of a small lipophilic group like fluorine at the paraposition of the phenyl ring of this type of compounds resulted in increased activity. The inclusion of more methoxy groups to phenyl ring (51, 5m, 5n) resulted in compounds have long duration of action at relatively higher dose (300 mg/kg). Furthermore, comparison of results obtained previously for the N-3-substituted 5,5-cyclopropanespirohydantoins and compounds described herein, which may be showed that introduction of amide between cyclopropanespirohydantoin nucleus and the aromatic ring did not influence the anticonvulsant activity. It was noteworthy that the introduction of an aryl group other than phenyl in the 5,5-cyclopropanespirohydantoin structure had caused a significant reduction or the complete loss of activity (compounds **5p**, **5q**, **5r**, **5s**), it seems that phenyl ring play a fundamental role in anti-MES protection [23]. It was also interested to note that compound 5j with CF₃ group in para-position of phenyl ring exhibited the most

Table 2 Phase-II quantitative anticonvulsant evaluation in mice.(test drug administered i.p.).							
Compound	FD _{co} a						

Compound	ED ₅₀ ^a		TD ₅₀ ^b	PI ^C MES scPTZ	
	MES	scPTZ			
5d	25.3 (22.8–29.5) ^d	208.3 (182.8–232.9)	382.3 (363.9-417.3)	15.5	1.8
5j	9.2 (5.4–12.3)	62.1 (43.7–92.3)	421.6 (398.3-454.5)	45.8	6.7
5t	13.7 (10.1–15.7)	136.6 (113.5-155.1)	387.2 (364.3-405.2)	17.8	2.8
Phenytoin ^e	9.5 (8.1–10.4)	>300	65.5 (52.5-72.9)	6.9	< 0.22
Carbamazepine ^e	8.8 (5.5-14.1)	>100	71.6 (45.9–135)	8.1	< 0.22
Phenobarbital ^e	21.8 (21.8-25.5)	13.2 (5.8–15.9)	69 (62.8-72.9)	3.2	5.2
Valproate ^e	272 (247-338)	149 (123–177)	426 (369-450)	1.6	2.9

Number of animals used: 10; solvent used: polyethylene glycol (0.1 mL, i.p.).

^a Dose in milligrams per kilogram body mass.

^b Minimal toxicity which was determined by rotorod test 30 min after the test drug was administered.

^c Protection index (TD₅₀/ED₅₀).

^d Data in parentheses are the 95% confidence limits.

^e Data from Ref. [33].

activity in this series, the *C* log *P* value of **5j** was also near 2.0, which is considered to be the optimum lipophilicity for the congeners that act on the central nervous system [24]. This observation indicated the importance of lipophilicity as well as electronic properties of the substituents on the activity of N-3-arylamide substituted 5,5-cyclopropanespirohydantoin derivatives. Another noteworthy observation was that compound **5t** exhibited marked anticonvulsant activity in both MES and *sc*PTZ tests. The structure of **5t** contains the two dicarboximide functions (CONRCO), which may contribute to increase activity.

4. Conclusions

In summary, the present studies revealed that number of N-3arylamide substituted 5,5-cyclopropanespirohydantoin derivatives were effective in the MES and/or *sc*PTZ screens. The anticonvulsant activity depended on the kind and position of substituents at the phenyl moiety. In the neurotoxicity studies some of the active compounds were devoid of toxicity. The most active was N-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)-4-fluorobenzamide (**5j**) which showed ED₅₀ value of 9.2 mg/kg and a protective index (TD₅₀/ED₅₀) of 45.8 in the MES test in mice. This compound showed greater ED₅₀ and lower TD₅₀ to the reference drugs.

5. Experimental protocols

5.1. Chemistry

All the chemicals and solvent were purchased from commercial sources. Solvents and reagents were dried and purified according to the literature methods. Melting points were uncorrected and measured on an XT-4 apparatus. ¹H-NMR spectra were obtained on a Varian Mercury VX400 apparatus in DMSO-*d*₆ and CDCl₃ with TMS as internal standard. The elemental analysis (C, H, N) data were obtained from a VarioEL III (German) elemental analyzer. The mass spectra (MS) were recorded on AMD-604 Mass Spectrometer operating at 70 eV.

5.1.1. Synthesis of 1-(ethoxycarbonyl)-2,2dimethylcyclopropanecarboxylic acid **2**

To a solution of diethyl 2,2-dimethylcyclopropane-1,1-dicarboxylate **1** (4.8 g, 22 mmol) in EtOH (25 mL) was added 1 N sodium hydroxide (25 mL, 1.1 equiv, 25 mmol), and the resulting mixture was stirred at room temperature for 12 h. EtOH was removed under reduced pressure, water was added to the residue, and the mixture was acidified by means of a saturated KHSO₄ solution and extracted with ethyl acetate (3×30 mL). The combined extracts were dried over Na₂SO₄ and evaporated to give product **2** (3.75 g, 90%) as a colorless oil: ¹H-NMR (400 MHz, CDCl₃): δ 1.25 (s, 3H, CH₃), 1.33 (t, 3H, *J* = 7.20 Hz, CH₃), 1.38 (s, 3H, CH₃), 1.78 (s, 1H, Cpr-H), 1.85 (s, 1H, Cpr-H), 4.29 (q, 2H, *J* = 7.20 Hz, CH₂), 11.28 (br s, 1H, OH). ¹³C NMR (150 MHz, CDCl₃): δ 13.92, 20.75, 21.66, 26.94, 33.50, 38.54, 62.08, 171.30, 181.29.

5.1.2. General procedure for the synthesis of compounds 4a-t

Compound 2 (10 mmol) was dissolved in dry THF (30 mL) and cooled to -15 °C. After the addition of EtOCOCI (11 mmol) and NMM (12 mmol), the mixture was stirred for 20 min. A solution of NaN₃ (25 mmol) in H₂O was added and stirred for 1 h at -10 °C. The solution was then diluted with H₂O and extracted with EtOAc. The organic layers were washed with brine, dried over Na₂SO₄ and concentrated under reduced pressure to give crude acyl azide. This crude acyl azide could be further purified by a flash column chromatography (PE-EtOAc, 4:1, $R_f = 0.7$). Purified acyl azide was dissolved in toluene (30 mL) and the resulting solution was heated to 75 °C under stirring. After gas evolution had stopped toluene was removed under reduced pressure to afford α -carboethoxy isocyanate **3** as clear oil. This α -carboethoxy isocyanate **3** was directly used in the next step without further purification. Arylhydrazide (10 mmol) was added to a stirred suspension of isocyanate 3 in appropriate solvent (40 mL) at r.t. (when highly reactive arylhydrazides were used, such as 4-methoxybenzhydrazide, 3,4-di methoxybenzhydrazide and 3,4,5-tri methoxybenzhydrazide, they should be dissolved in solvent and added dropwise). The solvent was removed under reduced pressure when the reaction was completed (detected by TLC) and the products 4 were purified by a column chromatography.

5.1.2.1. Ethyl 1-(2-benzoylhydrazinecarboxamido)-2,2-dimethylcyclopropane carboxylate (**4a**). Yield: 85%. White solid. Mp: 173–174 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): δ 0.80 (d, 1H, *J* = 4.92 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.17 (t, 3H, *J* = 7.08 Hz, CH₃), 1.51 (d, 1H, *J* = 4.92 Hz, Cpr-CH), 4.06 (q, 2H, *J* = 7.08 Hz, CH₂), 7.10 (br, 1H, NH), 7.47–7.54 (m, 2H, C_{2,6}-ArH), 7.55–7.57 (m, 1H, C₄-ArH), 7.88–7.90 (m, 2H, C_{3,5}-ArH), 7.98 (br, 1H, NH), 10.17 (br, 1H, NH). ESI-MS: 320.2 ([M + H]⁺). Anal. calc. for C₁₆H₂₁N₃O₄: C 60.17, H 6.63, N 13.61; found: C 60.32, H 6.48, N 13.44.

5.1.2.2. Ethyl 2,2-dimethyl-1-(2-(4-methylbenzoyl)hydrazinecarboxamido) cyclopropanecarboxylate (**4b**). Yield: 85%. White solid. Mp: 150–151 °C. ¹H-NMR (400 MHz, DMSO-d₆): 0.79 (d, 1H, J = 4.00 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.18 (t, 3H, J = 7.04 Hz, CH₃), 1.51 (d, 1H, J = 4.00 Hz, Cpr-CH), 2.36 (s, 3H, CH₃), 4.05 (q, 2H, J = 7.04 Hz, CH₂), 7.08 (brs, 1H, NH), 7.28 (d, 2H, J = 8.00 Hz, C_{3.5}-ArH), 7.79 (d, 2H, J = 8.00 Hz, C_{2.6}-ArH), 7.94 (brs, 1H, NH), 10.09 (brs, 1H, NH). ESI- MS: 334.2 ($[M + H]^+$). Anal. calc. for $C_{17}H_{23}N_3O_4$: C 61.25, H 6.95, N 12.60; found: C 61.17, H 6.76, N 12.83.

5.1.2.3. Ethyl 2,2-dimethyl-1-(2-(3-methylbenzoyl)hydrazinecarboxamido)cyclopropanecarboxylate (**4c**). Yield: 85%. White solid. Mp: 87–88 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 0.78 (d, 1H, *J* = 4.80 Hz, Cpr-CH), 1.12 (s, 6H, 2CH₃), 1.18 (t, 3H, *J* = 7.04 Hz, CH₃), 1.50 (d, 1H, *J* = 4.80 Hz, Cpr-H), 2.35 (s, 3H, CH₃), 4.05 (q, 2H, *J* = 7.04 Hz, CH₂), 7.07 (brs, 1H, NH), 7.32–7.36 (m, 2H, C_{4.5}-ArH), 7.66–7.67 (m, 1H, C₆-ArH), 7.70 (s, 1H, C₂-ArH), 7.96 (brs, 1H, NH), 10.10 (brs, 1H, NH). ESI-MS: 334.2 ([M + H]⁺). Anal. calc. for C₁₇H₂₃N₃O₄: C 61.25, H 6.95, N 12.60; found: C 61.12, H 6.77, N 12.85.

5.1.2.4. Ethyl 1-(2-(4-tert-butylbenzoyl)hydrazinecarboxamido)-2,2dimethylcyclopropanecarboxylate (**4d**). Yield: 80%. White solid. Mp: 115–116 °C. ¹H-NMR (400 MHz, DMSO-d₆): 0.79 (d, 3H, J = 4.88 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.18 (t, 3H, J = 7.04 Hz, CH₃), 1.30 (s, 9H, 3CH₃), 1.51 (d, 1H, J = 4.88 Hz, Cpr-CH), 4.06 (q, 2H, J = 7.04 Hz, CH₂), 7.08 (brs, 1H, NH), 7.48 (d, 2H, J = 8.52 Hz, C_{3.5}-ArH), 7.82 (d, 2H, J = 8.44 Hz, C_{2.6}-ArH), 7.96 (brs, 1H, NH), 10.10 (brs, 1H, NH). ESI-MS: 376.4 ([M + H]⁺). Anal. calc. for C₂₀H₂₉N₃O₄: C 63.98, H 7.79, N 11.19; found: C 63.81, H 7.87, N 11.43.

5.1.2.5. Ethyl 1-(2-(4-chlorobenzoyl)hydrazinecarboxamido)-2,2-dimethylcyclopropanecarboxylate (**4e**). Yield: 80%. White solid. Mp: 95–96 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 0.78 (d, 1H, J = 4.88 Hz, Cpr-CH), 1.12 (s, 6H, 2CH₃), 1.17 (t, 3H, J = 7.08 Hz, CH₃), 1.49 (d, 1H, J = 4.88 Hz, Cpr-CH), 4.05 (q, 2H, J = 7.08 Hz, CH₂), 7.15 (brs, 1H, NH), 7.56 (d, 2H, J = 8.48 Hz, C_{3,5}-ArH), 7.89 (d, 2H, J = 8.48 Hz, C_{2,6}-ArH), 8.00 (brs, 1H, NH), 10.25 (brs, 1H, NH). ESI-MS: 354.2 ([M + H]⁺). Anal. calc. for C₁₆H₂₀ClN₃O₄: C 54.32, H 5.70, N 11.88; found: C 54.53, H 5.61, N 11.95.

5.1.2.6. Ethyl 1-(2-(2-chlorobenzoyl)hydrazinecarboxamido)-2,2-dimethyl cyclopropanecarboxylate (**4f**). Yield: 85%. White solid. Mp: 113–114 °C. ¹H-NMR (400 MHz, DMSO-d₆): 0.80 (d, 1H, J = 4.92 Hz, Cpr-CH), 1.13 (s, 3H, 2CH₃), 1.17 (t, 3H, J = 7.04 Hz, CH₃), 1.52 (d, 1H, J = 4.92 Hz, Cpr-CH), 4.06 (q, 2H, J = 7.04 Hz), 6.95 (brs, 1H, NH), 7.39–7.47 (m, 5H, ArH), 8.13 (brs, 1H, NH), 10.07 (brs, 1H, NH). ESI-MS: 354.2 ([M + H]⁺). Anal. calc. for C₁₆H₂₀ClN₃O₄: C 54.32, H 5.70, N 11.88; found: C 54.42, H 5.51, N 11.93.

5.1.2.7. Ethyl 1-(2-(4-bromobenzoyl)hydrazinecarboxamido)-2,2-dimethyl cyclopropanecarboxylate (**4g**). Yield: 82%. White solid. M.p. 101–102 °C. ¹H-NMR (400 MHz, DMSO-d₆): 0.77 (d, 1H, J = 4.84 Hz, Cpr-CH), 1.12 (s, 6H, 2CH₃), 1.17 (t, 3H, J = 7.12 Hz, CH₃), 1.49 (d, 1H, J = 4.84 Hz, Cpr-CH), 4.05 (q, 2H, J = 7.12 Hz, CH₂), 7.15 (brs, 1H, NH), 7.69 (d, 2H, J = 8.44 Hz, C_{3,5}-ArH), 7.82 (d, 2H, J = 8.44 Hz, C_{2,6}-ArH), 8.00 (brs, 1H, NH), 10.25 (brs, 1H, NH). ESI-MS: 398.1 ([M + H]⁺). Anal. calc. for C₁₆H₂₀BrN₃O₄: C 48.25, H 5.06, N 10.55; found: C 48.32, H 5.23, N 10.32.

5.1.2.8. Ethyl 1-(2-(4-fluorobenzoyl)hydrazinecarboxamido)-2,2-dimethyl cyclopropanecarboxylate (**4h**). Yield: 83%. White solid. Mp: 169–170 °C. ¹H-NMR (400 MHz, DMSO- d_6): 0.78 (d, 1H, *J* = 4.68 Hz, Cpr-CH), 1.12 (s, 6H, 2CH₃), 1.17 (t, 3H, *J* = 7.08 Hz, CH₃), 1.50 (d, 1H, *J* = 4.68 Hz, Cpr-CH), 4.05 (q, 2H, *J* = 7.08 Hz), 7.13 (brs, 1H, NH), 7.29–7.33 (m, 2H, C_{3,5}-ArH), 7.93–7.97 (m, 2H, C_{2,6}-ArH), 7.99 (brs, 1H, NH), 10.18 (brs, 1H, NH). ESI-MS: 338.3 ([M + H]⁺). Anal. calc. for C₁₆H₂₀FN₃O₄: C 56.97, H 5.98, N 12.46; found: C 57.02, H 6.13, N 12.38.

5.1.2.9. Ethyl 1-(2-(2-fluorobenzoyl)hydrazinecarboxamido)-2,2-dimethyl cyclopropanecarboxylate (**4i**). Yield: 88%. White solid. Mp: 117–118 °C. ¹H-NMR (400 MHz, DMSO- d_6): 0.79 (d, 1H, *J* = 4.88 Hz,

Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.19 (t, 3H, J = 7.04 Hz, CH₃), 1.51 (d, 1H, J = 4.88 Hz, Cpr-CH), 4.06 (q, 2H, J = 7.04 Hz, CH₂), 7.01 (brs, 1H, NH), 7.27–7.32 (m, 2H, ArH), 7.53–7.54 (m, 1H, ArH), 7.65–7.66 (m, 1H, ArH), 8.10 (brs, 1H, NH), 9.99 (brs, 1H, NH). ESI-MS: 338.2 ([M + H]⁺). Anal. calc. for C₁₆H₂₀FN₃O₄: C 56.97, H 5.98, N 12.46; found: C 57.12, H 6.11, N 12.23.

5.1.2.10. Ethyl 2,2-dimethyl-1-(2-(4-(trifluoromethyl)benzoyl)hydrazinecarboxamido) cyclopropanecarboxylate (**4j**). Yield: 80%. White solid. Mp: 138–139 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 0.86 (d, 1H, J = 4.84 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.18 (t, 3H, J = 7.20 Hz, CH₃), 1.51 (d, 1H, J = 4.84 Hz, Cpr-CH), 4.05 (q, 2H, J = 7.20 Hz, CH₂), 7.21 (brs, 1H, NH), 7.87 (d, 2H, J = 8.28 Hz, C_{3,5}-ArH), 8.07 (d, 2H, J = 8.28 Hz, C_{2,6}-ArH), 8.10 (brs, 1H, NH), 10.41 (brs, 1H, NH). ESI-MS: 388.2 ([M + H]⁺). Anal. calc. for C₁₇H₂₀F₃N₃O₄ C 52.71, H 5.20, N 10.85; found : C 52.72, H 5.31, N 10.81.

5.1.2.11. Ethyl 2,2-dimethyl-1-(2-(4-nitrobenzoyl)hydrazinecarboxamido) cyclopropanecarboxylate (**4k**). Yield: 75%. Yellow solid. Mp: 155–156 °C. ¹H-NMR (400 MHz, DMSO-d₆): 0.79 (d, 1H, *J* = 4.88 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.19 (t, 3H, *J* = 7.04 Hz, CH₃), 1.51 (d, 1H, *J* = 4.88 Hz, Cpr-CH), 4.06 (q, 2H, *J* = 7.04 Hz, CH₂), 7.23 (brs, 1H, NH), 8.10 (brs, 1H, NH), 8.12 (d, 2H, *J* = 8.72 Hz, C_{2,6}-ArH), 8.33 (d, 2H, *J* = 8.72 Hz, C_{3,5}-ArH), 10.51 (brs, 1H, NH). ESI-MS: 365.3 ([M + H]⁺). Anal. calc. for C₁₆H₂₀N₄O₆ : C 52.74, H 5.53, N 15.38; found : C 52.66, H 5.37, N 15.46.

5.1.2.12. Ethyl 1-(2-(4-methoxybenzoyl)hydrazinecarboxamido)-2,2dimethyl cyclopropanecarboxylate (**41**). Yield: 85%. White solid. Mp: 153–154 °C. ¹H-NMR (400 MHz, DMSO-d₆): 0.79 (d, 1H, J = 4.92 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.19 (t, 3H, J = 7.06 Hz, CH₃), 1.49 (d, 1H, J = 4.92 Hz, Cpr-CH), 3.81 (s, 3H, OCH₃), 4.02 (q, 2H, J = 7.06 Hz, CH₂), 5.37 (brs, 1H, NH), 7.00 (d, 2H, J = 8.88 Hz, C_{3,5}-ArH), 7.86 (d, 2H, J = 8.88 Hz, C_{2,6}-ArH), 10.03 (brs, 1H, NH), 11.72 (brs, 1H, NH). ESI-MS: 350.1 ([M + H]⁺). Anal. calc. for C₁₇H₂₃N₃O₅ : C 58.44, H 6.64, N 12.03; found : C 58.62, H 6.52, N 12.22.

5.1.2.13. Ethyl 1-(2-(3,4-dimethoxybenzoyl)hydrazinecarboxamido)-2,2- dimethylcyclopropanecarboxylate (**4m**). Yield: 85%. White solid. Mp: 134–135 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 0.78 (d, 1H, J = 4.80 Hz, Cpr-CH), 1.16 (s, 6H, 2CH₃), 1.18 (t, 3H, J = 7.00 Hz, CH₃), 1.50 (d, 1H, J = 4.80 Hz, Cpr-CH), 3.80 (s, 6H, 2OMe), 4.05 (q, 2H, J = 7.00 Hz, CH₂), 7.01–7.07 (m, 2H, C4,5-ArH), 7.47 (brs, 1H, NH), 7.49–7.51 (m, 1H, C₂-ArH), 7.92 (brs, 1H, NH), 10.04 (brs, 1H, NH). ESI-MS: 380.1 ([M + H]⁺). Anal. calc. for C₁₈H₂₅N₃O₆ : C 56.98, H 6.64, N 11.08; found : C 56.82, H 6.42, N 11.12.

5.1.2.14. Ethyl 1-(2-(3,4,5-trimethoxybenzoyl)hydrazinecarboxamido)-2,2-dimethylcyclopropanecarboxylate (**4n**). Yield: 85%. White solid. Mp: 187–188 °C. ¹H-NMR (400 MHz, DMSO-d₆): 0.78 (d, 1H, J = 4.96 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.19 (t, 3H, J = 7.08 Hz, CH₃), 1.51 (d, 1H, J = 4.96 Hz, Cpr-CH), 3.71 (s, 3H, C₄-ArOCH₃), 3.82 (s, 6H, C_{3,5}-ArOCH₃), 4.06 (q, 2H, J = 7.08 Hz, CH₂), 7.11 (brs, 1H, NH), 7.22 (s, 2H, C_{2,6}-ArH), 7.97 (brs, 1H, NH), 10.12 (brs, 1H, NH). ESI-MS: 410.1 ([M + H]⁺). Anal. calc. for C₁₉H₂₇N₃O₇ : C 55.74, H 6.65, N 10.26; found : C 55.43, H 6.43, N 10.37.

5.1.2.15. Ethyl 1-(2-(4-(dimethylamino)benzoyl)hydrazinecarboxamido)-2,2-dimethylcyclopropanecarboxylate (40). Yield: 77%. White solid. Mp: 186–187 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 0.78 (d, 1H, J = 4.92 Hz, Cpr-CH), 1.12 (s, 6H, 2CH₃), 1.18 (t, 3H, J = 7.04 Hz, CH₃), 1.50 (d, J = 4.92 Hz, Cpr-CH), 2.97 (s, 6H, 2CH₃), 6.70 (d, 2H, J = 8.92 Hz, C_{3.5}-2H), 6.99 (brs, 1H, NH), 7.75 (d, 2H, J = 8.80 Hz, C_{2.6}-2H), 7.82 (brs, 1H, NH), 9.81 (brs, 1H, NH). ESI-MS: 363.4 ([M + H]⁺). Anal. calc. for $C_{18}H_{26}N_4O_4$: C 59.65, H 7.23, N 15.46; found : C 59.42, H 7.12, N 15.29.

5.1.2.16. Ethyl 1-(2-(furan-2-carbonyl)hydrazinecarboxamido)-2,2dimethylcyclopropane carboxylate (**4p**). Yield: 80%. White solid. Mp: 120–121 °C. ¹H-NMR (400 MHz, DMSO- d_6): 0.79 (d, 1H, *J* = 4.96 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.17 (q, 3H, *J* = 7.08 Hz, Me), 1.49 (d, 1H, *J* = 4.96 Hz, Cpr-CH), 4.05 (q, 2H, *J* = 7.08 Hz, CH₂), 6.64 (m, 1H, C₄-ArH), 7.04 (brs, 1H, NH), 7.21 (brs, 1H, NH), 7.87–7.88 (m, 1H, C₃-ArH), 7.89–7.90 (m, 1H, C₅-ArH), 10.03 (brs, 1H, NH). ESI-MS: 310.2 ([M + H]⁺). Anal. calc. for C₁₄H₁₉N₃O₅ : C 54.36, H 6.19, N 13.58; found : C 54.61, H 6.27, N 13.69.

5.1.2.17. Ethyl 2,2-dimethyl-1-(2-(thiophene-2-carbonyl)hydrazinecarboxamido)-cyclopropanecarboxylate (**4q**). Yield: 77%. White solid. Mp: 180–181 °C. ¹H-NMR (400 MHz, DMSO-d₆): 0.79 (d, 1H, J = 4.96 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.18 (t, 3H, J = 7.04 Hz, CH₃), 1.50 (d, 1H, J = 4.96 Hz, Cpr-CH), 4.03 (q, 2H, J = 7.04 Hz, CH₂), 7.08 (brs, 1H, NH), 7.17–7.19 (m, 1H, C₄-ArH), 7.81–7.82 (m, 2H, C_{2,5}-ArH), 8.00 (brs, 1H, NH), 10.19 (brs, 1H, NH). ESI-MS: 326.2 ([M + H]⁺). Anal. calc. for C₁₄H₁₉N₃O₄ S: C 51.68, H 5.89, N 12.91; found : C 51.43, H 6.07, N 13.14.

5.1.2.18. Ethyl 1-(2-isonicotinoylhydrazinecarboxamido)-2,2-dimethylcyclopropane carboxylate (**4r**). Yield: 81%. White solid. Mp: 188–189 °C. ¹H-NMR (400 MHz, DMSO- d_6): 0.79 (d, 1H, *J* = 4.96 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.17 (t, 3H, *J* = 7.04 Hz, CH₃), 1.50 (d, 1H, *J* = 4.96 Hz, Cpr-CH), 4.06 (q, 2H, *J* = 7.04 Hz, CH₂), 7.22 (brs, 1H, NH), 7.78 (d, 2H, *J* = 6.04 Hz, C_{3,5}-ArH), 8.10 (brs, 1H, NH), 8.74 (d, 2H, *J* = 6.04 Hz, C_{2,6}-ArH), 10.47 (brs, 1H, NH). ESI-MS: 321.3 ([M + H]⁺). Anal. calc. for C₁₅H₂₀N₄O₄ : C 56.24, H 6.29, N 17.49; found : C 56.47, H 6.37, N 17.42.

5.1.2.19. Ethyl 2,2-dimethyl-1-(2-nicotinoylhydrazinecarboxamido) cyclopropane carboxylate (**4s**). Yield: 79%. White solid. Mp: 178–179 °C. ¹H-NMR (400 MHz, DMSO- d_6): 0.79 (d, 1H, *J* = 4.84 Hz, Cpr-CH), 1.13 (s, 6H, 2CH₃), 1.18 (t, 3H, *J* = 7.04 Hz, CH₃), 1.51 (d, 1H, *J* = 4.84 Hz, Cpr-CH), 4.06 (q, 2H, *J* = 7.04 Hz, CH₂), 7.22 (brs, 1H, NH), 7.51–7.56 (m, 1H, ArH), 8.06 (s, 1H, ArH), 8.21–8.23 (m, 1H, ArH), 8.72–8.73 (m, 1H, ArH), 9.03 (brs, 1H, NH), 10.36 (brs, 1H, NH). ESI-MS: 321.3 ([M + H]⁺). Anal. calc. for C₁₅H₂₀N₄O₄ : C 56.24, H 6.29, N 17.49; found : C 56.44, H 6.34, N 17.62.

5.1.2.20. Ethyl 1-(3-(1,3-dioxoisoindolin-2-yl)ureido)-2,2-dimethylcyclopropane carboxylate (**4t**). Yield: 82%. White solid. Mp: 188–189 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 0.81 (d, *J* = 4.72 Hz, Cpr-CH), 1.12 (s, 6H, 2CH₃), 1.21 (t, 3H, *J* = 7.06 Hz, CH₃), 1.48 (d, 1H, *J* = 4.72 Hz, Cpr-CH), 4.06 (q, 2H, *J* = 7.06 Hz, CH₂), 7.62 (brs, 1H, NH), 7.81–7.91 (m, 4H, ArH), 8.58 (brs, 1H, NH). ESI-MS: 346.3 ([M + H]⁺). Anal. calc. for C₁₇H₁₉N₃O₅: C 59.12, H 5.55, N 12.17; found : C 59.34, H 5.77, N 12.08.

5.1.3. General procedure for the synthesis of compounds 5a-t

To a solution of compound **4** (1 mmol) and EtOH (10 mL) was added sodium (1.2 mmol), and the reaction mixture was stirred at room temperature for 2 h. EtOH was removed under reduced pressure. Ethyl acetate and water were added to the residue. The ethyl acetate layer was dried over anhydrous Na_2SO_4 , concentrated, and recrystallized to give compound **5**.

5.1.3.1. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl) benzamide (**5a**). Yield: 91%. White solid. Mp: 125–126 °C. ¹H-NMR (400 MHz, CDCl₃): 1.19 (d, 1H, *J* = 3.12 Hz, Cpr-H), 1.23 (s, 3H, CH₃), 1.27 (s, 3H, CH₃), 1.43 (d, 1H, *J* = 3.12 Hz, Cpr-H), 7.28–7.35 (m, 2H, C_{2.6}-ArH), 7.43–7.47 (m, 1H, C₄-ArH), 7.75 (br, 1H, NH), 7.80–7.82

(m, 2H, C_{3,5}-ArH), 9.76 (br, 1H, NH). ESI-MS: 274.1 ($[M + H]^+$). Anal. calc. for C₁₄H₁₅N₃O₃ : C 61.53, H 5.53, N 15.38; found : C 61.62, H 5.42, N 15.58.

5.1.3.2. *N*-(1,1-*dimethyl*-5,7-*dioxo*-4,6-*diazaspiro*[2.4]*heptan*-6-*yl*)-4-*methylbenzamide*(**5b**). Yield: 88%. White solid. Mp: 132–133 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 1.18 (d, 1H, *J* = 4.84 Hz, Cpr-CH), 1.12 (s, 3H, CH₃), 1.31 (s, 3H, CH₃), 1.41 (d, 1H, *J* = 4.84 Hz, Cpr-CH), 2.36 (s, 3H, Ar-CH₃), 7.34 (d, 2H, *J* = 8.02 Hz, C_{3,5}-ArH), 7.88 (d, 2H, *J* = 8.02 Hz, C_{2,6}-ArH), 8.66 (brs, 1H, NH), 10.42 (brs, 1H, NH). ESI-MS: 288.1 ([M + H]⁺). Anal. calc. for C₁₅H₁₇N₃O₃ : C 62.71, H 5.96, N 14.63; found : C 62.92, H 5.82, N 14.43.

5.1.3.3. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)-3-methylbenzamide (**5c**). Yield: 90%. White solid. Mp: 115–116 °C. ¹H-NMR (400 MHz, DMSO- d_6): 1.17 (d, 1H, *J* = 4.44 Hz, Cpr-CH), 1.25 (s, 3H, CH₃), 1.30 (s, 3H, CH₃), 1.38 (d, 1H, *J* = 4.44 Hz, Cpr-CH), 2.34 (s, 3H, CH₃), 7.41–7.47 (m, 2H, ArH), 7.66–7.71 (m, 2H, ArH), 8.78 (brs, 1H, NH), 10.94 (brs, 1H, NH). ¹ESI-MS: 288.1 ([M + H]⁺). Anal. calc. for C₁₅H₁₇N₃O₃ : C 62.71, H 5.96, N 14.63; found : C 62.86, H 5.72, N 14.54.

5.1.3.4. 4-tert-butyl-N-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4] heptan-6-yl)-benzamide (**5d**). Yield: 84%. White solid. Mp: 169–170 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 1.16 (d, 1H, *J* = 4.92 Hz, Cpr-CH), 1.26 (s, 3H, CH₃), 1.30 (s, 3H, CH₃), 1.32 (s, 9H, 3CH₃), 7.48 (d, 2H, *J* = 8.52 Hz, C_{3,5}-ArH), 7.81 (d, 2H, *J* = 8.52 Hz, C_{2,6} = ArH), 8.76 (brs, 1H, NH), 10.08 (brs, 1H, NH). ESI-MS: 330.2 ([M + H]⁺). Anal. calc. for C₁₈H₂₃N₃O₃ : C 65.63, H 7.04, N 12.76; found : C 65.45, H 6.99, N 12.97.

5.1.3.5. 4-Chloro-N-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)benzamide (**5e**). Yield: 90%. White solid. Mp: 97–98 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 1.18 (d, 1H, *J* = 4.96 Hz, Cpr-CH), 1.23 (s, 3H, CH₃), 1.30 (s, 3H, CH₃), 1.37 (d, 1H, *J* = 4.96 Hz, Cpr-CH), 7.65 (d, 2H, *J* = 8.56 Hz, C_{3,5}-ArH), 7.93 (d, 2H, *J* = 8.56 Hz, C_{2,6}-ArH), 8.81 (brs, 1H, NH), 11.12 (brs, 1H, NH). ESI-MS: 308.3 ($[M + H]^+$). Anal. calc. for C₁₄H₁₄ClN₃O₃ : C 54.64, H 4.59, N 11.52; found : C 54.53, H 4.64, N 11.77.

5.1.3.6. 2-Chloro-N-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)benzamide (**5f**). Yield: 85%. White solid. Mp: 110–111 °C. ¹H-NMR (400 MHz, DMSO- d_6): 1.15 (d, 1H, *J* = 4.92 Hz, Cpr-CH), 1.23 (s, 3H, CH₃), 1.31 (s, 3H, CH₃), 1.36 (d, 1H, *J* = 4.92 Hz, Cpr-CH), 7.44–7.58 (m, 5H, ArH), 8.80 (brs, 1H, NH), 10.94 (brs, 1H, NH). ESI-MS: 308.3 ([M + H]⁺). Anal. calc. for C₁₄H₁₄ClN₃O₃ : C 54.64, H 4.59, N 11.52; found : C 54.41, H 4.79, N 11.75.

5.1.3.7. 4-Bromo-N-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)benzamide (**5g**). Yield: 85%. White solid. Mp: 101–102 °C. ¹H-NMR (400 MHz, DMSO-d₆): 0.91 (d, 1H, J = 4.88 Hz, Cpr-CH), 1.12 (s, 3H, CH₃), 1.17 (s, 3H, CH₃), 1.38 (d, 1H, J = 4.88 Hz, Cpr-CH), 7.69 (d, 2H, J = 8.52 Hz, C_{3.5}-ArH), 7.78 (d, 2H, J = 8.52 Hz, C_{2.6}-ArH), 8.80 (brs, 1H, NH), 10.41 (brs, 1H, NH). ESI-MS: 352.2 ([M + H]⁺). Anal. calc. for C₁₄H₁₄BrN₃O₃ : C 47.74, H 4.01, N 11.93; found : C 47.55, H 3.98, N 12.13.

5.1.3.8. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)-4-fluorobenzamide (**5h**). Yield: 88%. White solid. Mp: 109–110 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 1.16 (d, 1H, *J* = 5.56 Hz, Cpr-CH), 1.25 (s, 3H, CH₃), 1.30 (s, 3H, CH₃), 1.37 (d, 1H, *J* = 5.56 Hz, Cpr-CH), 7.39–7.43 (m, 2H, C_{3,5}-ArH), 7.98–8.01 (m, 2H, C_{2,6}-ArH), 8.80 (brs, 1H, NH), 11.05 (brs, 1H, NH). ESI-MS: 292.3 ([M + H]⁺). Anal. calc. for C₁₄H₁₄FN₃O₃ : C 57.73, H 4.84, N 14.43; found : C 57.65, H 4.62, N 14.63. 5.1.3.9. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)-2-fluorobenzamide (**5i**). Yield: 85%. White solid. Mp: 83–84 °C. ¹H-NMR (400 MHz, DMSO-d₆): 1.15 (d, 1H, J = 5.52 Hz, Cpr-CH), 1.24 (s, 3H, CH₃), 1.31 (s, 3H, CH₃), 1.36 (d, 1H, J = 5.52 Hz, Cpr-CH), 7.41–7.43 (m, 2H, ArH), 7.99–8.02 (m, 2H, ArH), 8.83 (brs, 1H, NH), 11.12 (brs, 1H, NH). ESI-MS: 292.3 ([M + H]⁺). Anal. calc. for C₁₄H₁₄FN₃O₃ : C 57.73, H 4.84, N 14.43; found : C 57.81, H 4.74, N 14.55.

5.1.3.10. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)-4-(trifluoromethyl)benzamide (**5***j*). Yield: 88%. White solid. Mp: 81–82 °C. ¹H-NMR (400 MHz, DMSO-d₆): 1.13 (d, 1H, J = 5.02 Hz, Cpr-CH), 1.15 (s, 3H, CH₃), 1.32 (s, 3H, CH₃), 1.34 (d, 1H, J = 5.02 Hz, Cpr-CH), 7.88–7.90 (d, 2H, J = 8.24 Hz, C_{3,5}-ArH), 8.01–8.03 (d, 2H, J = 8.24 Hz, C_{2,6}-ArH), 8.68 (brs, 1H, NH), 11.22 (brs, 1H, NH). ESI-MS: 342.1 ([M + H]⁺). Anal. calc. for C₁₅H₁₄F₃N₃O₃ : C 52.79, H 4.13, N 12.31; found : C 52.83, H 4.24, N 12.44.

5.1.3.11. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)-4-nitrobenzamide (**5k**). Yield: 92%. Brown solid. Mp: 131–132 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 0.91 (d, 1H, *J* = 4.76 Hz, Cpr-CH), 1.14 (s, 3H, CH₃), 1.16 (s, 3H, CH₃), 1.38 (d, 1H, Cpr-H), 8.10 (d, 2H, *J* = 8.80 Hz, C_{2,6}-ArH), 8.18 (d, 2H, *J* = 8.80 Hz, C_{3,5}-ArH), 8.22 (brs, 1H, NH), 10.62 (brs, 1H, NH). ESI-MS: 319.1 ([M + H]⁺). Anal. calc. for C₁₄H₁₄N₄O₅ : C 52.83, H 4.43, N 17.60; found : C 52.87, H 4.34, N 17.82.

5.1.3.12. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)-4-methoxybenzamide (**5***I*). Yield: 88%. White solid. Mp: 141–142 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 1.16 (d, 1H, *J* = 4.00 Hz, Cpr-CH), 1.25 (s, 3H, CH₃), 1.30 (s, 3H, CH₃), 1.37 (d, 1H, *J* = 4.00 Hz, Cpr-CH), 3.84 (s, 3H, OCH₃), 7.09 (d, 2H, *J* = 8.80 Hz, C_{3,5}-ArH), 7.90 (d, 2H, *J* = 8.80 Hz, C_{2,6}-ArH), 8.76 (brs, 1H, NH), 10.85 (brs, 1H, NH). ESI-MS: 304.1 ([M + H]⁺). Anal. calc. for C₁₅H₁₇N₃O₄ : C 59.40, H 5.65, N 13.85; found : C 59.55, H 5.51, N 13.92.

5.1.3.13. *N*-(1,1-*dimethyl*-5,7-*dioxo*-4,6-*diazaspiro*[2.4]*heptan*-6-*yl*)-3,4-*dimethoxybenzamide* (**5m**). Yield: 80%. White solid. Mp: 95–96 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 1.17 (d, 1H, *J* = 4.98 Hz, Cpr-CH), 1.26 (s, 3H, CH₃), 1.28 (s, 3H, CH₃), 1.37 (d, 1H, *J* = 4.98 Hz, Cpr-CH), 3.82 (s, 3H, OCH₃), 3.84 (s, 3H, OCH₃), 7.11 (d, 1H, *J* = 8.48 Hz, C₅-ArH), 7.48 (s, 1H, C₂-ArH), 7.57 (d, 1H, *J* = 8.48 Hz, C₆-ArH), 8.77 (brs, 1H, NH), 10.85 (brs, 1H, NH). ESI-MS: 334.1 ([M + H]⁺). Anal. calc. for C₁₆H₁₉N₃O₅ : C 57.65, H 5.75, N 12.61; found : C 57.77, H 5.64, N 12.77.

5.1.3.14. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)-3,4,5-trimethoxybenzamide (**5n**). Yield: 80%. White solid. Mp: 221–222 °C. ¹H-NMR (400 MHz, DMSO- d_6): 1.16 (d, 1H, *J* = 4.86 Hz, Cpr-CH), 1.26 (s, 3H, CH₃), 1.30 (s, 3H, CH₃), 1.36 (d, 1H, *J* = 4.86 Hz, Cpr-CH), 3.74 (s, 3H, OCH₃), 3.85 (s, 6H, 2OCH₃), 7.25 (s, 2H, ArH), 8.80 (brs, 1H, NH), 10.93 (brs, 1H, NH).ESI-MS: 364.1 ([M + H]⁺). Anal. calc. for C₁₇H₂₁N₃O₆ : C 56.19, H 5.83, N 11.56; found : C 56.27, H 5.63, N 11.72.

5.1.3.15. N-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl)-4-(dimethylamino)-benzamide (**50**). Yield: 80%. White solid. Mp: 133–134 °C. ¹H-NMR (400 MHz, CDCl₃): 1.22 (d, 1H, J = 4.78 Hz, Cpr-CH), 1.24 (s, 3H, CH₃), 1.30 (s, 3H, CH₃), 1.35 (d, 1H,, J = 4.78 Hz, Cpr-CH), 3.00 (s, 6H, 2CH₃), 6.75 (d, 2H, J = 8.80 Hz, C_{3,5}-ArH), 7.77 (d, 2H, J = 8.80 Hz, C_{2,4}-ArH), 8.71 (brs, 1H, NH), 10.54 (brs, 1H, NH). ESI-MS: 317.1 ([M + H]⁺).Anal. calc. for C₁₆H₂₀N₄O₃ : C 60.75, H 6.37, N 17.71; found: C 60.54, H 6.54, N 17.63.

5.1.3.16. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl) furan-2-carboxamide (**5p**). Yield: 88%. White solid. Mp: 97–98 °C.

¹H-NMR (400 MHz, DMSO-*d*₆): 1.14 (d, 1H, J = 4.82 Hz, Cpr-CH), 1.13 (s, 3H, CH₃), 1.31 (s, 3H, CH₃), 1.36 (d, 1H, J = 4.82 Hz, Cpr-CH), 6.67–6.68 (m, 1H, ArH), 7.86–7.87 (m, 1H, ArH), 7.95–7.06 (m, 1H, ArH), 8.89 (brs, 1H, NH), 11.12 (brs, 1H, NH). ESI-MS: 264.2 ([M + H]⁺). Anal. calc. for C₁₂H₁₃N₃O₄ : C 54.75, H 4.98, N 15.96; found : C 54.92, H 5.24, N 16.13.

5.1.3.17. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl) thiophene-2-carboxamide (**5q**). Yield: 80%. White solid. Mp: 213–214 °C. ¹H-NMR (400 MHz, DMSO-d₆): 1.18 (d, 1H, *J* = 4.96 Hz, Cpr-CH), 1.25 (s, 3H, CH₃), 1.30 (s, 3H, CH₃), 1.37 (d, 1H, *J* = 4.96 Hz, Cpr-CH), 7.25–7.27 (m, 1H, ArH), 7.89–7.90 (m, 1H, ArH), 7.94–7.95 (m, 1H, ArH), 8.80 (brs, 1H, NH), 11.03 (brs, 1H, NH). ESI-MS: 280.2 ([M + H]⁺). Anal. calc. for C₁₂H₁₃N₃O₃S : C 51.60, H 4.69, N 15.04; found : C 51.51, H 4.88, N 15.24.

5.1.3.18. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl) isonicotinamide (**5r**). Yield: 80%. White solid. Mp: 191–192 °C. ¹H-NMR (400 MHz, DMSO- d_6): 1.15 (d, 1H, *J* = 4.88 Hz, Cpr-CH), 1.23 (s, 3H, CH₃), 1.29 (s, 3H, CH₃), 1.33 (d, 1H, *J* = 4.88 Hz, Cpr-CH), 7.76 (d, *J* = 6.06 Hz, C_{2,6}-ArH), 8.77 (d, *J* = 6.06 Hz, C_{3,5}-ArH), 8.89 (brs, 1H, NH), 10.88 (brs, 1H, NH). ESI-MS: 275.1 ([M + H]⁺). Anal. calc. for C₁₃H₁₄N₄O₃ : C 56.93, H 5.14, N 20.43; found : C 56.96, H 5.30, N 20.61.

5.1.3.19. *N*-(1,1-dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl) nicotinamide(**5s**). Yield: 82%. White solid. Mp: 73–74 °C. ¹H-NMR (400 MHz, DMSO- d_6): 1.16 (d, 1H, *J* = 4.82 Hz, Cpr-CH), 1.24 (s, 3H, CH₃), 1.32 (s, 3H, CH₃), 1.51 (d, 1H, *J* = 4.82 Hz, Cpr-CH), 7.52–7.56 (m, 1H, ArH), 8.04 (s, 1H, ArH), 8.22–8.24 (m, 1H, ArH), 8.73–8.75 (m, 1H, ArH), 8.78 (brs, 1H, NH), 10.92 (brs, 1H, NH). ESI-MS: 275.1 ([M + H]⁺). Anal. calc. for C₁₃H₁₄N₄O₃ : C 56.93, H 5.14, N 20.43; found : C 56.94, H 5.21, N 20.52.

5.1.3.20. 2-(1,1-Dimethyl-5,7-dioxo-4,6-diazaspiro[2.4]heptan-6-yl) isoindoline-1,3-dione (**5t**). Yield: 80%. White solid. Mp: 97–98 °C. ¹H-NMR (400 MHz, DMSO-*d*₆): 1.15 (d, J = 4.94 Hz, Cpr-CH), 1.22 (s, 3H, CH₃), 1.33 (s, 3H, CH₃), 1.52 (d, 1H, J = 4.94 Hz, Cpr-CH), 7.83–8.01 (m, 4H, ArH), 8.88 (brs, 1H, NH). ESI-MS: 300.1 ([M + H]⁺). Anal. calc. for C₁₇H₁₈N₄O₄ : C 59.64, H 5.30, N 16.37; found : C 59.72, H 5.43, N 16.55.

5.2. Anticonvulsant screening

Anticonvulsant activity assays were carried out by the Antiepileptic Drug Development Program, Epilepsy Branch, National Institute of Neurological and Communicative Disorders and Stroke, National Institute of Health in Bethesda, USA [25]. Male Kunming mice $(20 \pm 2.0 \text{ g})$ and male Kunming rats $(250 \pm 5.0 \text{ g})$ were used as experimental animals in this study. Animals of the same age and weight have been selected in order to minimize biological. The animals were approved by the Animal Care Committee of Wuhan University. All the animals were purchased from Wuhan University Laboratory Animal Center (Wuhan, China). The tested compounds were injected intraperitoneally to mice as a suspension in 0.5% methylcellulose at dose of 30, 100, and 300 mg/kg to one to four mice. The anticonvulsant activity of the tested compounds were evaluated by two models namely, MES and scPTZ models. Phenytoin and ethosuximide were used as the standard drugs for the comparison. The neurological toxicity was determined in the rotorod test. Procedures employed for evaluation of anticonvulsant activity and neurotoxicity were described elsewhere [26].

5.2.1. MES-maximal electroshock seizure pattern test

This activity was tested according to the method of Swinyard [27]. In experiments with mice, a 60-Hz current of 50-mA intensity was applied through corneal electrodes for a 0.25 s duration; The procedures caused immediate hindlimb tonic extension. After 0.5 h and 4.0 h of drug administration, electroshocks were via corneal electrodes. Absence of tonic extension suggests that the tested compound was considered as positive criteria.

5.2.2. Pentylenetetrazole (PTZ) induced seizure test

For the chemically induced convulsant test according to the method of Vamecq et al. [28], pentylenetetrazole was dissolved in sufficient 0.9% saline to allow subcutaneous injections to mice or rat. Standard drug in this model was ethosuximide. After 0.5 h and 4.0 h of drug administration the failure to observe even a threshold seizure (a single episode of clonic spasms of at least 5s duration) is defined as protection.

5.2.3. Neurotoxicity screening

Minimal motor impairment was measured in mice or rats by using standardized rotorod test [29]. The mouse was placed on a 1 in. diameter knurled plastic rod rotating at 6 rpm. Trained animals were given an ip injection of the test compounds in doses of 30, 100, and 300 mg/kg. Neurotoxicity was indicated by the inability of the animal to maintain equilibrium on the rod for at least 1 min in each of the four trials.

5.2.4. Quantification studies

Anticonvulsant activity was expressed in terms of the median effective dose (ED_{50}), that is, the dose of drug required to produce the biological responses in 50% of animals, neurotoxicity was expressed as the median toxic dose (TD_{50}). For determination of the ED_{50} and TD_{50} , groups of 10 mice were given a range of ip doses of the test drug until at least three points were established in the range of 10–90% seizure protection or minimal observed neurotoxicity [30]. From the plot of these data, the respective ED_{50} and TD_{50} values, 95% confidence intervals, slope of the regression line, and standard error of the slope were calculated by means of a computer program written at NINDS, NIH.

Acknowledgments

This investigation was made possible through the financial support of Key Project of Science & Technology of HuBei Province, China (Grant NO. 2008CDA 057) and the Important National Science & Technology Specific Projects, China (Grant No. 2009ZX09301-14).

References

- N. Pessah, M. Bialer, B. Wlodarczyk, R.H. Finnell, B. Yagen, J. Med. Chem. 52 (2009) 2233–2242.
- [2] M. Bialer, M.C. Walker, W.S. Josemir, CNS Drugs 16 (2002) 285-289.
- [3] K.J. Meador, J. Clin, Psychiatry 64 (Suppl. 8) (2003) 30-34.
- [4] Z. Lin, P.K. Kadaba, Med. Res. Rev. 17 (1997) 537–572.
- [5] M.L. Wagner, Am. J. Hosp. Pharm. 51 (1994) 1657–1666.
 [6] G. Zaccara, D. Franciotta, E. Perucca, Epilepsia 48 (2007) 1223–1244.
- [7] H.H. Merritt, T.J. Putnam, Arch. Neurol. Psychiatry 39 (1938) 1003–1015.
- [8] T.M. Hassell, M.C. Johnson, K.H. Dudley, Phenytoin Induced Teratology and Gingival Pathology. Raven Press, New York, 1980.
- [9] J.J. Edmunds, S. Klutchko, J.M. Hamby, A.M. Bunker, C.J.C. Connolly, R.T. Winters, J. Quin III, I. Stircar, J.C. Hodges, R.L. Panek, J.A. Keiser, A.M. Doherty, J. Med. Chem. 38 (1995) 3759–3771.
- [10] S. Hanessian, J.Y. Sanceau, P. Chemla, Tetrahedron 51 (1995) 6669-6678.
- [11] K.I. Ahmed, Carbohydr. Res. 306 (1998) 567–573.
- [12] R.N. Comber, R.C. Reynolds, J.D. Friedrich, R.A. Manguikian, R.W. Buckheit,
- J.J.W. Truss, W.M. Shannon, J.A. Secrist, J. Med. Chem. 35 (1992) 3567–3572. [13] C.-H. Oh, H.J. Kim, S.-Y. Hong, Y.-H. Lee, J.K. Cho, J.-H. Cho, Arch. Pharm. 328
- (1995) 385–387. [14] D. Kim, L. Wang, C.G. Caldwell, P. Chen, P.E. Finke, B. Oates, M. MacCoss,
- [14] D. Kini, L. Wallg, C.G. Caldweil, P. Chen, P.E. Finke, B. Oales, M. Maccoss, S.G. Mills, L. Malkowitz, S.L. Gould, J.A. DeMartino, M.S. Springer, D. Hazuda, M. Miller, J. Kessler, R. Danzeisen, G. Carella, K. Holmes, J. Lineberger, W.A. Schleif, E.A. Emini, Bioorg. Med. Chem. Lett. 11 (2001) 3099–3102.
- [15] E. Naydenova, N. Pencheva, J. Popova, N. Stoyanov, M. Lazarova, B. Aleksiev, Farmaco 57 (2002) 189–194.
- [16] Q. Zhu, Y. Pan, Z. Xu, R. Li, G. Qiu, W. Xu, X. Ke, L. Wu, X. Hu, Eur. J. Med. Chem. 44 (2009) 296–302.
- [17] H.J. Patel, J. Sarra, F. Caruso, M. Rossi, U. Doshia, R.A. Stephania, Bioorg. Med. Chem. Lett. 16 (2006) 4644–4647.
- [18] H.A. Schenck, P.M. Lenkowski, I. Choudhury-Mukherjee, S.H. Ho, J.P. Stables, M.K. Patel, M.L. Brown, Bioorg. Med. Chem. 12 (2004) 979–993.
- [19] J. Su, G. Qiu, S. Liang, X. Hu, Synth. Commun. 35 (2005) 1427-1433.
- [20] J. Su, Z. Xiong, S. Liang, G. Qiu, X. Feng, H. Teng, L. Wu, X. Hu, Synth. Commun. 36 (2006) 693–699.
- [21] N.D. Kimpe, M. Boeykens, J. Org. Chem. 59 (1994) 8215-8219.
- [22] A. Camerman, N. Camerman, in: G.H. Glasen, J.K. Penry, D.M. Woodbury (Eds.), Antiepileptic Drugs: Mechanisms of Action, Raven Press, New York, 1980, pp. 223–231.
- [23] J.C. Thenmozhiyal, P.T.-H. Wong, W.-K. Chui, J. Med. Chem. 47 (2004) 1527–1535.
- [24] E.J. Lien, R.C. Liao, H.G. Shinouda, J. Pharm. Sci. 68 (1979) 463–465.
- [25] R.J. Porter, J.J. Cereghino, G.D. Gladding, B.J. Hessie, H.J. Kupferberg, B. Scoville, B. White, Cleve. Clin. Q. 51 (1984) 293–305.
- [26] R.L. Krall, J.K. Penry, B.G. White, H.J. Kupferberg, E.A. Swinyard, Epilepsia 19 (1978) 409-428.
- [27] E.A. Swinyard, Epilepsia 10 (1969) 107-119.
- [28] J. Vamecq, D. Lambert, J.H. Poupaert, B. Masereel, J.P. Stables, J. Med. Chem. 41 (1998) 3307–3313.
- [29] N.W. Dunham, T.A. Miya, J. Am. Pharm. Assoc. Sci. 46 (1957) 206-212.
- [30] H.S. White, J.H. Woodhead, K.S. Wilcox, J.P. Stables, H.J. Kupferberg, H.H. Wolf, in: R.H. Levy, R.H. Mattson, B.S. Meldrum, E. Perucca (Eds.), Antiepileptic Drugs, Lippincott Williams & Wilkins Publishers, New York, 2002, pp. 36–48.
- [31] J.R. Dimmock, S.N. Pandey, J.W. Quail, U. Pugazhenthi, T.M. Allen, G.Y. Kao, J. Balzarini, E. DeClercq, Eur. J. Med. Chem. 30 (1995) 303–314.
- [32] H. Rajak, R. Deshmukh, N. Aggarwal, S. Kashaw, M.D. Kharya, P. Mishra, Arch. Pharm. 342 (2009) 456.
- [33] H. Ucar, V.D. Kim, S. Cacciaguerra, S. Spampinato, J.P. Stables, P. Depovere, M. Isa, B. Masereel, J. Delarge, J.H. Poupaert, J. Med. Chem. 41 (1998) 1138–1145.