# A New Series of PDGF Receptor Tyrosine Kinase Inhibitors: 3-Substituted Quinoline Derivatives

Martin P. Maguire,<sup>\*,†</sup> Kimberly R. Sheets,<sup>†</sup> Karen McVety,<sup>‡</sup> Alfred P. Spada,<sup>†</sup> and Asher Zilberstein<sup>‡</sup> Rhône-Poulenc Rorer Central Research, 500 Arcola Road, Collegeville, Pennsylvania 19426

Received February 4, 1994\*

A series of 63 3-substituted quinoline derivatives has been prepared and tested for inhibition of cell-free platelet derived growth factor receptor tyrosine kinase (PDGF-RTK) activity. The compounds were generally prepared either by a Friedlander condensation between an aryl-acetaldehyde and an o-aminobenzaldehyde or by a palladium-catalyzed coupling between an aryl bromide or triflate and an organostannane or organozinc chloride. The presence of 6,7-dimethoxy groups on the quinoline ring was found to be advantageous although not essential for potent inhibition of PDGF-RTK. A lipophilic group attached to the quinoline 3-position contributed substantially to activity. The lipophilic groups generally consisted of monocyclic aromatics or small alkynyl, alkenyl, and alkyl groups. Optimum activity of ca.  $\leq 20$  nM (IC<sub>50</sub>) was observed when 6,7-dimethoxyquinoline was substituted in the 3-position with 4-methoxyphenyl (15d), 3-fluoro-4-methoxyphenyl (17m), 3-fluorophenyl (17b), 4-hydroxyphenyl (24), 6-methoxypyridin-3-yl (15o), 5-pyridin-2(1H)-one (23), trans- $\beta$ -styryl (15e), thiophene-3-yl (2e), 5-chlorothiophene-2-yl (15f), or cyclopentenyl (17n) groups. Most of the compounds in the series were tested for inhibition of cell-free epidermal growth factor receptor tyrosine kinase activity and found to be inactive.

Platelet-derived growth factor (PDGF) receptors are integral, transmembrane glycoproteins whose expression is generally limited to cells of mesodermal origin.<sup>1</sup> Abnormal expression of PDGF and/or PDGF receptors has been linked to a number of pathophysiological processes which include various forms of neoplasia, atherosclerosis, rheumatoid arthritis, pulmonary fibrosis, myelofibrosis, and abnormal wound healing.<sup>2</sup> Upon ligand binding, the intrinsic tyrosine kinase activity of the PDGF receptor is activated. This leads to tyrosine phosphorylation of numerous proteins, including the receptor itself. Receptor autophosphorylation is essential for all subsequent steps of signal transduction. Importantly, the signaling cascade that follows from the action of PDGF on smooth muscle cell (SMC) PDGF receptors is believed to result in a significant contribution to the process of SMC proliferation and chemotaxis.<sup>3</sup> This process is a crucial component of the sequence of events leading to restenosis after percutaneous transluminal coronary angioplasty procedures.<sup>4</sup> An inhibitor of the PDGF receptor tyrosine kinase activity would block its action. Such an inhibitor could potentially be utilized in the treatment of proliferative disorders linked to PDGF receptors, including postangioplasty restenosis.

A series of 2,3-diarylacrylonitrile compounds has been prepared in our laboratories as epidermal growth factor receptor tyrosine kinase (EGF-RTK) inhibitors.<sup>5</sup> With several exceptions, these compounds were generally poor PDGF-RTK inhibitors. Compound 1 (RG13022),<sup>6</sup> however, displayed modest inhibitory activity in both the EGF-RTK<sup>7</sup> and PDGF-RTK cell-free assays (IC<sub>50</sub> = 0.5–3 and 0.7–4  $\mu$ M, respectively). As part of an effort to identify lead structures as PDGF-RTK inhibitors, we decided to

0022-2623/94/1837-2129\$04.50/0

synthesize compounds derived from conformationally restricted analogues of 1. Two such compounds were quinolines 2a and 2b, which can be envisioned simply by attachment of the nitrile nitrogen of 1 to either of the two open positions of the adjacent dimethoxybenzene ring.<sup>8</sup> The quinolines differ from the parent in several important respects. The polarized double bond of the acrylonitrile has been deleted. We have observed that 1 and similar compounds have a propensity to undergo light-induced cis/trans isomerization and 2 + 2 cycloadditions. The acrylonitrile system also has the potential to act as a Michael acceptor. In addition, the quinoline nitrogen of 2a and 2b introduces a new basic center which can serve as a potential hydrogen-bond acceptor.



In vitro screening of 2a and 2b revealed a fortuitous result. Quinoline 2a showed good inhibition of PDGF-RTK (IC<sub>50</sub> = 0.1-0.8  $\mu$ M) and excellent selectivity against EGF-RTK (IC<sub>50</sub> > 25  $\mu$ M) while 2b showed poor inhibition of both EGF-RTK (IC<sub>50</sub> > 25  $\mu$ M) and PDGF-RTK (IC<sub>50</sub> > 50  $\mu$ M). Thus, 2a served as the starting point for an extensive chemical analogue program. In this paper, we

© 1994 American Chemical Society

Department of Medicinal Chemistry.

<sup>&</sup>lt;sup>‡</sup> Department of Inflammation/Bone Metabolism.

<sup>•</sup> Abstract published in Advance ACS Abstracts, May 15, 1994.

will present the chemistry and in vitro PDGF-RTK structure-activity relationships for a series of 3-substituted quinolines.

#### Chemistry

Our initial approach to 3-substituted quinoline synthesis was via a simple Friedlander condensation of o-aminobenzaldehydes and arylacetaldehydes as shown in Scheme 1, method A (structures defined in Table 1). By condensation of 3a,<sup>9</sup>, 3b,<sup>10</sup> 3g, or  $3h^{11}$  with aldehydes 4a,<sup>12</sup> 4c,<sup>13</sup> 4e,<sup>14</sup> or 4f, 3-arylquinolines 2a-h were obtained. While this method provided a rapid entry into the series, its usefulness with regard to analogue synthesis is limited by the availability and stability of the prerequisite oaminobenzaldehydes or arvlacetaldehydes. For example. indole-3-acetaldehyde is unstable and must be used quickly after preparation. Other arylacetaldehydes are unstable in basic solution. Likewise, the availability of substituted o-aminobenzaldehydes is variable, and their stability and/ or reactivity is highly dependent upon the nature of the substituents. We therefore required a more direct approach that could be used to quickly construct a larger variety of analogues.

The use of palladium-catalyzed coupling reactions for the construction of substituted heterocycles has been welldocumented<sup>15</sup> and has simplified the synthesis of many important pharmaceutical intermediates. Our need to prepare a series of 3-substituted quinoline derivatives was well-suited to take advantage of this chemistry. Appropriately substituted quinoline-3-triflates could be used to couple with organostannane<sup>16</sup> or organozinc<sup>17</sup> compounds to provide rapid access to the desired analogues.

The sequence of reactions for the preparation of 3-hydroxyquinolines 9a-i from anilines 5a-h makes use a previously described<sup>18</sup> protocol and is shown in Scheme 2. The overall yields in the sequence are quite variable and depend mainly on the success (or lack thereof) of the polyphosphoric acid-induced cyclization of nitromalonaldehyde adducts 6a-h to give 3-nitroquinolines 7a-i. The (methylenedioxy)quinoline compound 7j was prepared by an alternative literature<sup>19</sup> route which we found to be operationally inferior to the one depicted. Triflates 10a-j were obtained by reaction of 9a-j with triflic anhydride in methylene chloride/pyridine.

The organostannanes used to couple with 10a-j were prepared by the methods shown in Schemes 3 and 4. All of the bromides indicated were purchased from commercial sources except for  $11f.^{20}$  Arylstannanes 12a-f (Scheme 3, method B) were synthesized by palladium-catalyzed coupling of triflate 11a or bromides 11b-f with hexamethylditin.<sup>16</sup> The remaining organostannanes 14a-kwere prepared by reacting the corresponding organolithium compounds (from 13a-k, Scheme 4, method C) with trimethyltin chloride. Method C is procedurally more convenient than method B but is obviously unsuitable for use with compounds unstable to organolithium reagents.

Organostannanes 12a-f and 14a-k were coupled to the indicated quinoline-3-triflates (Scheme 5, method D) using prescribed conditions<sup>16b</sup> to give 15a-w and 15y. Similar coupling of 3-bromoquinoline (13e) with 14j, and of 14e with trans- $\beta$ -bromostyrene (13i), gave 15x and 15z, respectively. Isolated reaction yields were generally very good, ranging from 51 to 99% in most cases. The low yield in the coupling of 10j with 14j ( $\rightarrow$ 15t) was due in part to difficulties in purifying 10j and its preceding intermediates.<sup>19</sup> The remaining coupling products 17a-p were synthesized by palladium-catalyzed reaction of the organozinc chlorides obtained from 16a-p with 10a (Scheme 6, method E). The bromides are commercially available except for  $16h^{21}$  and  $16n.^{22}$  Method E was more convenient in comparison to method D and afforded the desired coupling products in shorter time, using lower temperatures and without isolation of the intermediate organometallics. Alkynes 160,p were subjected to the same procedure to give 170,p in excellent yields. Compounds 18-27 were prepared from the appropriate precursors as indicated in Table 1.

## **Results and Discussion**

It became obvious early in our studies that the placement of certain substituents in the quinoline 8-position was deleterious to PDGF-RTK inhibitory activity as can be seen from the comparison of 2a and 2b. A very similar result was observed with the 3-(indol-3-yl)quinolines 2c and 2d (see Table 1), which were also conceptually derived from a nonselective (EGF/PDGF-RTK) 2.3-diarylacrylonitrile-based inhibitor (RG13291)<sup>23</sup> analogous to 1. This point was further supported by comparing the effect of varying methyl substitution on the quinoline ring. The 5,7-dimethylquinoline 15y is a very potent inhibitor; however, movement of one of the methyl groups from the 7- to the 8-position (15p) results in a dramatic loss of activity. In addition, the presence of the quinoline nitrogen was found to be essential for activity in the series as was illustrated by the lack of activity (IC<sub>50</sub> > 50  $\mu$ M) associated with the naphthalene analogue<sup>24</sup> of 2e.

It also became clear that changes in the quinoline 3-aromatic group had a marked effect on potency. Replacement of the pyridine of 2a with phenyl (2f) improved activity somewhat while replacement with a thiophene-3-yl group (2e) had a dramatic effect by decreasing the  $IC_{50}$  to 1-20 nM. The importance of the lipophilic or aromatic group attached to the quinoline 3-position can be seen by noting the absence of activity observed in compounds where such groups are lacking. For example, the 3-nitro-, 3-amino-, and 3-hydroxyquinolines 7a, 8a, and 9a were inactive. We also tested the parent 6,7-dimethoxyquinoline<sup>25</sup> (21) and found it to lack inhibitory activity. When 8a or 9a were benzylated (i.e., compounds 19 and 20), activity was restored such that  $<100 \text{ nM IC}_{50}$  values were obtained. The addition of a methoxy group to the pyridine of 2a (15o) increased potency by ca. 10-fold, and similar activity was observed with the related pyridone 23. Minor changes to the thiophene of 2e also influenced potency. Comparison of the 3-(thiophene-2-yl)quinoline compound 151 with 2e reveals a weakening effect on the  $IC_{50}$  value. The 5-chlorothiophene-2-yl compound 15f was ca. 10-fold more active than the analogous 5-methoxy compound 17e. Movement of the thiophene 5-chloro substituent of 15f to the 3-position (15g) reduced activity by 100-fold.

Substituent changes to the phenyl ring of 2f significantly varied inhibitory potency. The 4-methoxyphenyl compound 15d was at least 10-fold more active than 2f, and addition of an o-fluoro substituent (17m) produced a compound of comparable potency. Methoxy substitution in either the 2- or 3-phenyl positions (17c and 15h) resulted in much less active compounds relative to 15d. Both the 3-fluorophenyl and the 4-fluorophenyl compounds 17b and 17a were potent inhibitors. Relative to the parent 2f, it Table 1



no.	$R_1$	$\mathbf{R}_2$	$R_3$	$R_4$	$\mathbf{R}_{5}$	method	% yield°	$IC_{50} (\mu M)^{b}$
20	<u>и</u>	004	0CH	u	2 miniday]	٨	70	01_09
28 01	п				3-pyridyl	A	19	0.1-0.0
ZD	н	H	OCH <sub>3</sub>	UCH <sub>3</sub>	3-pyridyl	A	62	>50
2c	н	OCH <sub>3</sub>	OCH3	н	3-indolyl	A	23	0.1-0.6
2d	н	Н	OCH3	OCH <sub>3</sub>	3-indolyl	Α	14	>50
2e	н	$OCH_3$	OCH₃	н	3-thienyl	Α	75	0.001 - 0.02
2f	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	CeH5	Α	61	0.07 - 0.15
20	н	н	H	н	3-indolvl	A	32	2-8
- <b>B</b> 2h	ŭ	осн.	ü	ŭ	3-thionyl	Å	62	0.05-0.07
211	11		004	11	NO	л	00	0.00-0.07
18	n			п 	NO <sub>2</sub>	с	-	>00
8a.	н	$OCH_3$	OCH3	н	$NH_2$	с	-	>50
9a	н	$OCH_3$	OCH₃	н	OH	с	-	≥20
15a	н	OCH <sub>3</sub>	OCH <sub>3</sub>	Н	2-naphthyl	D	62	≥20
15b	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	$C_eH_4(4-C_eH_s)$	D	66	>50
150	н	OCH.	OCH.	н	CeHe(3 4-Cle)	ā	83	1-7
150	ŭ	OCH.	OCH.	u ii	$C_{13}(0, -C_{2})$	ñ	74	0.001 0.015
150	п И			п Л	C6H4(4-OCH3)	U U	14	0.001-0.015
156	н	OCH3		<u> </u>	trans-p-styryl	D	91	0.001-0.005
15f	н	$OCH_3$	OCH3	Н	2-thienyl(5-Cl)	D	56	0.007-0.03
15g	н	OCH₃	$OCH_3$	H	2-thienyl(3-Cl)	D	68	1
15 <b>h</b>	н	OCH <sub>3</sub>	$OCH_3$	н	$C_6H_4(3-OCH_3)$	D	91	1-2
15 <b>i</b>	н	OCH	OCH	н	CeHe[3.4.5-(OCHe)e]	D	51	>50
151	ਸ	OCH.	OCH.	ਸ	$C_{1}H_{2}(0,1,0)$	ñ	97	0 07-0 15
157	11 11	OCH		u II	$C \mathbf{H} (2 \mathbf{NO} \mathbf{A} \mathbf{OCH})$	D	00	0.07-0.10
15K	п т			n	$C_{6}\Pi_{3}(3-NO_{2},4-OC\Pi_{3})$	D	99	0.10-0.5
151	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	2-thienyl	D	82	0.04-0.08
15m	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	$C_6H_3(3,4-F_2)$	D	90	0.09-0.2
15 <b>n</b>	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	$C_6H_4(4-CO_2Et)$	D	93	0.1-0.2
150	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	3-pyridyl(6-OCH <sub>3</sub> )	D	86	0.008-0.07
15n	CH.	н	н	CH.	3-thienvl	Ē	86	>50
150	й Н	F	л. Т	н.	3-thionyl	กั	86	2-4
154	11 11		r Cl	11 11	9 thismal	D D	75	2 <sup></sup>
151	п			п	3-thienyl	D D	10	1-2
158	н	н	OCH3	н	3-thienyl	D	99	0.002-0.06
15t	н	-0Cl	H <sub>2</sub> O-	Н	3-thienyl	D	34	0.2-0.6
15u	OCH <sub>3</sub>	Н	OCH <sub>3</sub>	Н	3-thienyl	D	86	0.025-0.03
15v	F	н	н	н	3-thienvl	D	92	0.02
1550	н	н	F	H	3-thienvl	ñ	92	0.004-0.025
15-	ü	ü	ů	ŭ	2 thionyl	D D	74	0.004 0.020
154		11		11		U U	14	0.2-0.20
159	CH <sub>3</sub>	<u> </u>	Un <sub>3</sub>	<u> </u>	3-thienyl	Ū U	80	0.001-0.005
15z	н	н	н	н	trans-β-styryl	D	33	0.1-0.2
17a	н	OCH₃	OCH₃	Н	$C_6H_4(4-F)$	$\mathbf{E}$	99	0.035-0.06
17b	н	$OCH_3$	OCH <sub>3</sub>	Н	$C_{6}H_{4}(3-F)$	E	65	0.015-0.025
17c	н	OCH.	OCH <sub>2</sub>	н	CeH4(2-OCHa)	Е	90	2
17d	ਸ	OCH.	OCH.	н	CeH4[4-N(CHa)a]	Ē	91	10
170	ü	OCU.	OCH.	Ĥ	2-thionyl(5-004-)	P	Q1	01-095
176	п 11	OCH3		11	$2$ -tillelly ( $(5$ -OCH $_3)$	E	00	0.1-0.20
1/1	п	OCH3	OCH3	п	$0.6\Pi_3[2,4-(0,0\Pi_3)_2]$	ц Г	23	>00
178	н	OCH <sub>3</sub>	OCH3	н	$U_6H_4(4-UUH_2U_6H_5)$	E	75	>30
17 <b>h</b>	Н	OCH3	OCH <sub>3</sub>	Н	$trans-\beta$ -styryl(4-OCH <sub>3</sub> )	E	65	0.03-0.1
17i	н	OCH3	OCH <sub>3</sub>	н	C <sub>6</sub> H <sub>3</sub> (3-CH <sub>3</sub> ,4-OCH <sub>3</sub> )	$\mathbf{E}$	36	0.1-0.4
17j	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	$C_6H_3[3,4-(OCH_3)_2]$	Е	93	2-10
17k	н	OCH.	OCH.	н	$C_6H_4(4-CH_3)$	$\mathbf{E}$	92	0.5-2
173	Ĥ	OCH.	OCH.	н	CeH(3.5-Fe 4-OCH)	Ē	99	0.2-0.4
17m	ŭ	OCH.	OCH.	ü	$C_{1}U_{2}(0,0-12,1-0,0,113)$	Ē	24	0.00 0.1
17-	u II		OCH3	11 11	-6113(0-1, -00003)	E F	04 60	
17 <b>n</b>	н	OCH3	OCH3	н	1-cyclopentenyl	E	62	0.02-0.04
170	н	OCH3	UCH3	н	$etnynyl(2-C_6H_5)$	E	99	0.05 - 0.02
17p	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	ethynyl(2-cyclohexyl)	E	99	0.3–1
18	н	OH	ОН	н	3-pyridyl	d	98	6-10
19	н	OCH <sub>2</sub>	OCH <sub>3</sub>	н	NHCH <sub>2</sub> C <sub>4</sub> H <sub>5</sub>	е	91	0.02-0.08
20	н	OCH.	OCH.	Ĥ	OCH CAH	ŕ	76	0.04-0.07
21	ü	0013	OCH.	ਸ਼ਿੰ	U U	i d	76	20-20
41	11 T		OCH	11 17		Б L	10	20-30
44	л Т	OCH3	OCH3	л Т	$\cup_{6}\Pi_{4}(4-\cup\cup_{2}\Pi)$	n	10	0.7-0
23	н	OCH <sub>3</sub>	OCH3	н	5-pyridin-2(1H)-one	L	55	0.015-0.03
24	н	$OCH_3$	$OCH_3$	н	$C_6H_4(4-OH)$	j1	80	0.02-0.03
25	н	$OCH_3$	$OCH_3$	н	$ethyl[2-C_6H_4(4-OCH_3)]$	j2	99	0.2-0.8
26	н	OCH <sub>3</sub>	$OCH_3$	н	ethyl(2-cyclohexyl)	j3	68	0.3-0.5
27	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	cyclopentyl	j4	92	0.1-0.5
		-	-					

<sup>a</sup> Indicates unoptimized isolated yields. <sup>b</sup> Concentration of compound required to inhibit phosphorylation of isolated human PDGF receptor by 50% (minimum of two dose-response assays). For a detailed description of the assay see the Experimental Section. <sup>c</sup> Refer to Scheme 2 and ref 18. <sup>d</sup> By reaction of refluxing 48% aqueous HBr with **2a**. <sup>e</sup> By reaction of benzaldehyde with **8a** and NaCNBH<sub>3</sub>. <sup>f</sup> By reaction of benzyl bromide with **9a** and sodium hydride. <sup>g</sup> By Skraup reaction with **5a**, ref 25. <sup>h</sup> By reaction of NaOH/EtOH with 15n. <sup>i</sup> By reaction of pyridine-HCl at 160 °C, 5 min with 150. <sup>j</sup> By reaction of H<sub>2</sub>, Pd/C with: <sup>1</sup>17g; <sup>2</sup>17h; <sup>3</sup>17p; <sup>4</sup>17n.

appears that optimum activity can be obtained by introducing methoxy (15d), hydroxy (24), or fluoro (17a)

substituents at the 4-position of the phenyl ring. Substitution of the phenyl ring at the 4-position with either



<sup>a</sup> Key: (a) EtOH, dilute aqueous NaOH. <sup>b</sup>  $R_2$ - $R_5$  substituents analogous to derivatives of 2, Table 1.

Scheme 2<sup>a</sup>



<sup>a</sup> Key: (a) Sodium nitromalonaldehyde, EtOH, H<sub>2</sub>O; (b) PPA, 160 °C; (c) H<sub>2</sub>, Pd/C; (d) NaNO<sub>2</sub>, HCl; (e) Tf<sub>2</sub>O, pyridine, CH<sub>2</sub>Cl<sub>2</sub>. <sup>b</sup> Unoptimized overall yield of **9a-i** from **5a-h**. <sup>c</sup> See ref 19.

0-CH2-O

н

F

н

н

11

c

Scheme 3. Method  $B^a$ 

Т

н

н



<sup>a</sup> Key: (a) [(CH<sub>3</sub>)<sub>3</sub>Sn]<sub>2</sub>, Pd(PPh<sub>3</sub>)<sub>4</sub>, dioxane, LiCl for 11a; [(CH<sub>3</sub>)<sub>3</sub>Sn]<sub>2</sub>, Pd(PPh<sub>3</sub>)<sub>4</sub>, toluene for 11b-f.

a nitro (15j) or ethoxycarbonyl (15n) group gave compounds ca. equipotent to 2f while the 4-methyl (17k), 4-carboxy (22), and 4-dimethylamino (17d) compounds were less active.

The size and shape of the 3-substituent can also substantially influence the degree of inhibition. The naphthylquinoline 15a, the biphenylylquinoline 15b, and the [(benzyloxy)phenyl]quinoline 17g had no appreciable

Scheme 4. Method  $C^a$ 

13 13	R <sub>s</sub> — X 3a-j: X = Br 3k: X = H	a,b		R <sub>5</sub> ─ Sn(CH <sub>3</sub> ) <sub>3</sub> 1 <b>4a-k</b>		
analog	R <sub>5</sub>	%yieid	analog	R <sub>5</sub>	%yield	
a	2-naphthyl	99	g	2-thienyl(3-Cl)	93	
b	C <sub>6</sub> H <sub>4</sub> (4-C <sub>6</sub> H <sub>5</sub> )	99	h	C <sub>6</sub> H <sub>4</sub> (3-OCH <sub>3</sub> )	90	
c	C <sub>6</sub> H <sub>3</sub> (3,4-Cl <sub>2</sub> )	90	1	( <i>trans</i> )-ß-styryl	51	
d	C <sub>6</sub> H <sub>4</sub> (4-OCH <sub>3</sub> )	82	J	3-thienyl	97	
	3-quinolyl	50	k	2-thienyl	56	
f	2-thienyl(5-Cl)	73				

<sup>a</sup> Key: (a) t-BuLi 220 M% -78 °C, Et<sub>2</sub>O for 13a-h; t-BuLi 200 M%, -115 °C for 13i (ref 29); n-BuLi 100 M%, -78 °C, Et<sub>2</sub>O for 13j; n-BuLi 120 M%, 22 °C, Et<sub>2</sub>O for 13k; (b) (CH<sub>3</sub>)<sub>3</sub>SnCl.





<sup>a</sup> Key: (a)  $Pd(PPh_3)_4$ , dioxane, LiCl for 10a-j;  $Pd(PPh_3)_4$ , DMF for 13e and 14e. <sup>b</sup> Reaction of 14e with trans- $\beta$ -bromostyrene (13i).





<sup>a</sup> Key: (a) t-BuLi 210 M%, -78 °C for 16a-n; n-BuLi 100 M%, -78 °C for 16o,p; (b) ZnCl<sub>2</sub>, Et<sub>2</sub>O; (c) Pd(PPh<sub>3</sub>)<sub>4</sub>, dioxane, 10a. <sup>b</sup> R<sub>5</sub> substituent analogous to derivatives of 17, Table 1.

inhibitory effect. Placement of a styryl group in the quinoline 3-position (15e), however, resulted in one of the most potent inhibitors synthesized. This observation is interesting in light of the comparison of 15a with 15e. The naphthalene group of 15a can be viewed as a ring-constrained version of the styryl group of 15e. The electronic and/or steric variation imposed by this conformational restriction produces a dramatic decrease in potency.

The contribution to activity of the 6,7-dimethoxy groups can be seen clearly by comparison of 2c, 2e, and 15e with their 6,7-unsubstituted quinoline counterparts 2g, 15x, and 15z. Most of the activity was retained when only one of the two methoxy groups of 2e is removed (i.e., 2h and 15s). Constraining the 6,7-dimethoxy groups in the form of a methylenedioxy unit (i.e., 15t) substantially reduced inhibitory potency. Conversion of **2a** to its 6.7-dihydroxy analogue 18 also resulted in a significant reduction in PDGF-RTK inhibition. Unlike 2a, compound 18 displayed inhibition of EGF-RTK (IC<sub>50</sub> =  $0.5-2 \mu$ M). This was not unexpected as it is known that compounds containing ortho or para hydroquinone/quinone functionality frequently inhibit EGF-RTK.<sup>26</sup> With the exception of 15n, 15q, 15v, and 22, which were not tested, all the remaining compounds in Table 1 had EGF-RTK  $IC_{50's}$ of  $>20 \,\mu$ M. Compounds 2e, 15d, and 15u were also tested for inhibition of protein kinase A<sup>27</sup> and found to be inactive  $(IC_{50's} > 200 \ \mu M).$ 

Several quinoline derivatives were prepared in which the 3-substituent was held constant in the form of 3-thienyl and the 5-, 6-, and 7-substituents were varied (15p-y).

### PDGF Receptor Tyrosine Kinase Inhibitors

The 6,7-dimethoxy groups of 2e can also be replaced with 5,7-dimethoxy groups (15u) without loss of activity. Replacement of methoxy with methyl (15y) is also acceptable. The 5- and 7-monofluoro compounds 15v and 15w had activities comparable to their 6,7-dimethoxy counterpart 2e, but the 6,7-difluoro analogue 15q was significantly less active.

The tyrosine kinase inhibition realized by placing substituents in the quinoline 3-position is not limited to aryl groups. As can be seen in Table 1, certain alkynyl, alkenyl, and alkyl groups are also well-tolerated. Of particular note in this regard is the strong inhibition observed with the 3-cyclopentenyl compound 17n.

### Summary

A new inhibitor (2a) of cell-free PDGF-RTK with selectivity versus EGF-RTK was identified by imposing a conformational restriction on the diarylacrylonitrile 1. Although 2a is a valence bond isomer of 1, it differs considerably in terms of electronics, polarity, and shape. From 2a, a series of potent inhibitors of PDGF-RTK was developed.<sup>28</sup> Construction of the series relied heavily on the use of palladium-catalyzed organostannane or organozinc chloride coupling reactions. These compounds are characterized by a lipophilic appendage attached to the quinoline 3-position. The appendage consisted of a variety of substituted aryl, alkynyl, alkenyl, or alkyl groups. Maximum activity was observed when the 3-substituent was 4-methoxyphenyl, 3-fluoro-4-methoxyphenyl, 3-fluorophenyl, 4-hydroxyphenyl, 6-methoxypyridin-3-yl, 5-pyridin-2(1H)-one, trans- $\beta$ -styryl, thiophene-3-yl, 5-chlorothiophene-2-yl, or cyclopentenyl. Activity was particularly enhanced by the presence of 6,7-dimethoxy or 5,7-dimethyl functionality on the quinoline ring. Substitution at the quinoline 8-position abolished inhibitory activity. Structure-activity relationships were also determined for other substituted quinolines and quinoline related heterocycles and will be reported in future disclosures. The compounds are presently being evaluated in intact-cell mitogenesis assays as well as in animal models of fibroproliferative disorders associated with abnormal activation of PDGF receptors.

## **Experimental Section**

Melting points were determined in open-ended capillaries on a Thomas-Hoover apparatus and are uncorrected. Recrystallization solvents are reported in parentheses following melting points. Combustion analyses were obtained on a Perkin-Elmer 2400 CHN elemental analyzer and are within  $\pm 0.40\%$  of theoretical values unless otherwise indicated. Analyses obtained for all of the compounds reported are for CHN. All reported NMR data is for <sup>1</sup>H NMR at 300 MHz and was obtained on either a Bruker ACF-300 or a Bruker ARX-300 instrument. <sup>1</sup>H NMR chemical shifts are reported in  $\delta$  (ppm) downfield of the TMS internal standard calibrated at 0.00 ppm. Column chromatography was conducted on EM 60 silica gel, particle size 0.040-0.063 mm, 230-400 mesh. Anhydrous solvents were used in the palladium coupling reactions and were purchased from Aldrich and used directly. All coupling reactions were conducted under nitrogen or argon. IUPAC nomenclature was generated by AUTONOM Version 1.0.

Method A. 6,7-Dimethoxy-3-thiophene-3-ylquinoline, 2e. A solution of 12.0 g (66.3 mmol) of 3a in 80 mL of ethanol and 5 mL of aqueous 1 M NaOH was heated to 65 °C under nitrogen. To this was added, dropwise, 14.8 g (117 mmol) of freshly prepared neat 4e (prepared by DIBAL/toluene reduction of the methyl ester at -78 °C and chromatography eluting with 9:1 pentane/ ether) over a 40-min period via syringe. The solution was heated for an additional 20 min, concentrated to 1/3 volume on the rotovap, and partitioned between CH<sub>2</sub>Cl<sub>2</sub> (200 mL) and water (100 mL). The aqueous layer was back extracted with  $CH_2Cl_2$  $(3 \times 30 \text{ mL})$ , the combined organic extracts were washed with brine (75 mL), dried (MgSO<sub>4</sub>), and evaporated, and the residue was chromatographed on silica gel (eluting with 7:3 hexane/ EtOAc) to give 11.83 g of pure (by NMR) 2e. Additionally, 1.40 g of unreacted 3a was obtained along with ca. 2.0 g of impure 2e. The impure 2e was suspended in 100 mL of 15% aqueous H<sub>3</sub>PO<sub>4</sub> and the suspension washed with ether (100 mL). The aqueous suspension was then made basic with concentrated NH<sub>4</sub>OH and extracted with CH<sub>2</sub>Cl<sub>2</sub> (100 mL). The organic extract was dried (MgSO<sub>4</sub>) and evaporated and the residue chromatographed as before to give an additional 1.59 g of pure 2e (75% total yield of 2e; 85% based on recovered 3a): mp 116-118 °C (hexane/ EtOAc); NMR (CDCl<sub>3</sub>) 4.03 (s, 3 H), 4.05 (s, 3 H), 7.07 (s, 1 H), 7.42 (s, 1 H), 7.49 (m, 2 H), 7.59 (dd, 1 H, J = 1.6, 2.8 Hz), 8.14(d, 1 H, J = 2.2 Hz), 9.00 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>15</sub>H<sub>13</sub>- $NO_2S$ ).

Compounds 2a-h were similarly prepared by method A. Arylacetaldehydes 4c, 4e, and 4f were added neat or in ethanol solution. Pyridine-3-acetaldehyde (4a) was used directly by basification (with 50% aqueous NaOH) of the acidic solution from which it was prepared<sup>12</sup> after addition of ethanol and either 3a or 3b.

Method B. 1,2-Difluoro-4-(trimethylstannanyl)benzene 12d. To a solution of 1.00 g (5.18 mmol) of 1-bromo-3,4difluorobenzene (11d) and 1.70 g (5.18 mmol) of hexamethylditin in 20 mL of toluene was added 300 mg (0.259 mmol) of Pd-(PPh<sub>3</sub>)<sub>4</sub>. The reaction was heated to 90 °C under argon for 9 h at which time the solution turned black. The mixture was partitioned between 10% NH<sub>4</sub>OH (90 mL) and ether (100 mL) and stirred for 15 min. The organic layer was washed with brine (50 mL), dried (MgSO<sub>4</sub>) and evaporated. The residue was chromatographed on silica gel (eluting with hexane) to give 1.42 g (99%) of 12d as a colorless liquid which was used directly in the next step without further characterization. TLC  $R_f = 0.69$ (hexane).

Method C. 1-Methoxy-3-(trimethylstannyl)benzene, 14h. A solution of 1.50 g (8.02 mmol) of 3-bromoanisole (13h) in 30 mL of ether was cooled, under argon to -78 °C, and 10.4 mL (17.6 mmol) of t-BuLi, 1.7 M in pentane, was added dropwise via syringe. The solution was stirred at -78 °C for 15 min, and 3.80 mL (10.3 mmol) of trimethyltin chloride, 2.71 M in dioxane, was injected. The mixture was then warmed to 22 °C, stirred for 45 min, quenched with 5 mL of methanol, partitioned between 10% NH<sub>4</sub>OH (90 mL) and ether (100 mL), and stirred for 15 min. A diagonal (MgSO<sub>4</sub>), and evaporated. The residue was chromatographed on silica gel (eluting with hexane) to give 1.96 g (90%) of 14h as a colorless liquid which was used directly in the next step without further characterization. TLC  $R_f = 0.48$  (hexane).

Method D. 6,7-Dimethoxy-3-(3-methoxyphenyl)quinoline, 15h. A solution of 300 mg (0.888 mmol) of 10a, 313 mg (115 mmol) of 14h, 113 mg (2.66 mmol) of anhydrous LiCl, and 50 mg (0.043 mmol) of Pd(PPh<sub>3</sub>)<sub>4</sub> in 15 mL of dioxane was refluxed under argon for 16 h. The mixture was partitioned between 10% NH<sub>4</sub>OH (60 mL) and CH<sub>2</sub>Cl<sub>2</sub> (100 mL) and stirred for 30 min. The organic layer was separated, washed with brine, dried (MgSO<sub>4</sub>), and evaporated. The residue was chromatographed (eluting with 7:3 hexane/EtOAc) to give 237 mg (91%) of 15h: mp 122.5-124 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  3.91 (s, 3H), 4.05 (s, 3 H), 4.06 (s, 3 H), 6.97 (dd, 1 H, J = 1.8, 6.0 Hz), 7.11 (s, 1 H), 7.22 (apparent t, 1 H, J = 1.8 Hz), 7.29 (s, 1 H), 7.43 (m, 2 H), 8.16 (d, 1 H, J = 2.1 Hz), 8.98 (d, 1 H, J = 2.1 Hz). Anal. (C<sub>18</sub>H<sub>17</sub>NO<sub>3</sub>).

Method E. 3-(3-Fluorophenyl)-6,7-dimethoxyquinoline, 17b. A solution of 400 mg (2.29 mmol) of 1-bromo-3-fluorobenzene (16b) in 6 mL of THF under nitrogen was cooled to -78 °C, and 2.83 mL (4.81 mmol) of t-BuLi, 1.7 M in pentane, was added dropwise. The solution was stirred for 10 min, and 2.52 mL (2.52 mmol) of ZnCl<sub>2</sub>, 1.0 M in ether, was injected. The solution was then allowed to warm to 22 °C and stirred for 1 h at which time a second solution of 300 mg (0.890 mmol) of 10a and 51 mg (0.044 mmol) of Pd(PPh<sub>3</sub>)<sub>4</sub> in 4 mL of THF was injected via canula. The reaction mixture was heated to a gentle reflux for 2 h and then partitioned between 60 mL of NH<sub>4</sub>OH and 100 mL of CH<sub>2</sub>Cl<sub>2</sub>. After stirring for 15 min the organic layer was separated, washed with brine (50 mL), dried (MgSO<sub>4</sub>), and evaporated. The residue was chromatographed (eluting with CHCl<sub>3</sub>) to give 218 mg (65%) of 17b: mp 156–158 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.05 (s, 3 H), 4.07 (s, 3 H), 7.11 (m, 2 H), 7.37–7.50 (m, 4 H), 8.15 (d, 1 H, J = 2.1 Hz), 8.95 (d, 1 H, J = 2.1 Hz). Anal. (C<sub>17</sub>H<sub>14</sub>NO<sub>2</sub>F).

**6,7-Dimethoxy-3-pyridin-3-ylquinoline, 2a,** was obtained from **3a** and **4a** by method A: mp 131–132 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.05 (s, 3 H), 4.07 (s, 3 H), 7.13 (s, 1 H), 7.44 (m, 1 H), 7.46 (s, 1 H), 7.99 (m, 1 H), 8.18 (d, 1 H, J = 2.2 Hz), 8.66 (br d, 1 H, J = 4.5 Hz), 8.95 (m, 2 H). Anal. (C<sub>16</sub>H<sub>14</sub>N<sub>2</sub>O<sub>2</sub>).

**7,8-Dimethoxy-3-pyridin-3-ylquinoline, 2b,** was obtained from **3b** and **4a** by method A: mp 158-159 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.06 (s, 3 H), 4.13 (s, 3 H), 7.43 (d, 1 H, J = 9.0Hz), 7.44 (m, 1 H), 7.66 (d, 1 H, J = 9.0 Hz), 7.99 (m, 1 H), 8.27 (d, 1 H, J = 2.2 Hz), 8.67 (br d, 1 H, J = 4.3 Hz), 8.96 (br s, 1 H), 9.16 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>16</sub>H<sub>14</sub>N<sub>2</sub>O<sub>2</sub>).

3-(1*H*-Indol-3-yl)-6,7-dimethoxyquinoline, 2c, was obtained from 3a and 4c by method A: mp 204-205 °C (EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.05 (s, 6 H), 7.11 (s, 1 H), 7.28 (m, 2 H), 7.48 (m, 3 H), 8.01 (br d, 1 H, J = 7.4 Hz), 8.27 (d, 1 H, J = 2.0 Hz), 8.77 (br s, 1 H), 9.06 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>19</sub>H<sub>16</sub>N<sub>2</sub>O<sub>2</sub>).

**3-(1***H***-Indol-3-yl)-7,8-dimethoxyquinoline, 2d,** was obtained from **3b** and **4c** by method A: mp 254-255 °C (ethanol); NMR (CDCl<sub>3</sub>)  $\delta$  4.06 (s, 3 H), 4.18 (s, 3 H), 7.27 (m, 2 H), 7.39 (d, 1 H, J = 9.0 Hz), 7.49–7.56 (m, 2 H), 7.63 (d, 1 H, J = 9.0 Hz), 7.99 (br d, 1 H, J = 7.4 Hz), 8.35 (d, 1 H, J = 2.2 Hz), 8.64 (br s, 1 H), 9.28 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>19</sub>H<sub>16</sub>N<sub>2</sub>O<sub>2</sub>).

**6,7-Dimethoxy-3-phenylquinoline, 2f,** was obtained from **3a** and **4f** by method A: mp 126.5–128 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.04 (s, 3 H), 4.06 (s, 3 H), 7.11 (s, 1 H), 7.48 (m, 4 H), 7.70 (m, 2 H), 8.16 (d, 1 H, J = 2.2 Hz), 8.98 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>17</sub>H<sub>18</sub>NO<sub>2</sub>).

**3-(1***H***-Indol-3-yl)quinoline, 2g**, was obtained from **3g** and **4c** by method A: mp 173-175 °C (hexane/EtOAc then hexane/ethanol); NMR (CDCl<sub>3</sub>)  $\delta$  7.22-7.36 (m, 2 H), 7.47-7.61 (m, 3 H), 7.69 (m, 1 H), 7.88 (m, 1 H), 8.02 (m, 1 H), 8.14 (br d, 1 H, J = 8.3 Hz), 8.42 (d, 1 H, J = 2.2 Hz), 8.62 (br s, 1 H), 9.26 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>17</sub>H<sub>12</sub>N<sub>2</sub>·0.15H<sub>2</sub>O).

**6-Methoxy-3-thiophene-3-ylquinoline, 2h**, was obtained from **3h** and **4e** by method A: mp 126–128 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  3.95 (s, 3 H), 7.10 (d, 1 H, J = 2.7 Hz), 7.35 (dd, 1 H, J = 2.8, 9.1 Hz), 7.50 (m, 2 H), 7.64 (dd, 1 H, J = 1.4, 2.8 Hz), 8.00 (d, 1 H, J = 9.2 Hz), 8.18 (d, 1 H, J = 2.1 Hz), 9.04 (d, 1 H, J = 2.1 Hz). Anal. (C<sub>14</sub>H<sub>11</sub>NOS).

**6,7-Dimethoxy-3-naphth-2-ylquinoline, 15a,** was obtained from 10a and 14a by method D: mp 162.5–165 °C (hexane/CH<sub>2</sub>-Cl<sub>2</sub>); NMR (CDCl<sub>3</sub>)  $\delta$  4.06 (s, 3 H), 4.07 (s, 3 H), 7.15 (s, 1 H), 7.51 (m, 3 H), 7.82–8.00 (m, 4 H), 8.15 (s, 1 H), 8.29 (s, 1 H), 9.11 (s, 1 H). Anal. (C<sub>21</sub>H<sub>17</sub>NO<sub>2</sub>).

**3-Biphenyl-4-yl-6,7-dimethoxyquinoline, 15b,** was obtained from 10a and 14b by method D: mp 143–145 °C (Et<sub>2</sub>O); NMR (CDCl<sub>3</sub>)  $\delta$  4.06 (s, 3 H), 4.07 (s, 3 H), 7.13 (s, 1 H), 7.39 (d, 1 H, J = 7.2 Hz), 7.48 (m, 2 H), 7.51 (s, 1 H), 7.67 (d, 2 H, J = 7.2 Hz), 7.77 (m, 4 H), 8.22 (d, 1 H, J = 2.0 Hz), 9.04 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>23</sub>H<sub>19</sub>NO<sub>2</sub>·0.9H<sub>2</sub>O).

**3-(3,4-Dichlorophenyl)-6,7-dimethoxyquinoline, 15c,** was obtained from 10a and 14c by method D: mp 184–186 °C (EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.05 (s, 3 H), 4.07 (s, 3 H), 7.11 (s, 1 H), 7.46 (s, 1 H), 7.52 (dd, 1 H, J = 2.0, 8.6 Hz), 7.58 (d, 1 H, J = 8.6 Hz), 7.77 (d, 1 H, J = 2.0 Hz), 8.14 (d, 1 H, J = 2.2 Hz), 8.91 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>17</sub>H<sub>13</sub>NO<sub>2</sub>Cl<sub>2</sub>).

**6,7-Dimethoxy-3-(4-methoxyphenyl)quinoline, 15d,** was obtained from 10a and 14d by method D: mp 162.5-164.5 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  3.87 (s, 3 H), 4.03 (s, 3 H), 4.05 (s, 3 H), 7.04 (d, 2 H, J = 8.6 Hz), 7.08 (s, 1 H), 7.43 (s, 1 H), 7.62 (d, 2 H, J = 8.6 Hz), 8.09 (d, 1 H, J = 2.2 Hz), 8.95 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>18</sub>H<sub>17</sub>NO<sub>3</sub>).

**6,7-Dimethoxy-3-***trans-* $\beta$ **-styrylquinoline, 15e,** was obtained from 10a and 14i by method D: mp 104–105.5 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.03 (s, 3 H), 4.05 (s, 3 H), 7.06 (s, 1 H), 7.24 (d, 2 H, J = 7.3 Hz), 7.30 (m, 1 H), 7.37 (m, 3 H), 7.57 (d, 2 H, J = 7.3 Hz), 8.06 (d, 1 H, J = 2.0 Hz), 8.91 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>19</sub>H<sub>17</sub>NO<sub>2</sub>).

3-(5-Chlorothiophene-2-yl)-6,7-dimethoxyquinoline, 15f, was obtained from 10a and 14f by method D: mp 131.5-132.5 °C (hexane/EtOAc then methanol); NMR (CDCl<sub>3</sub>)  $\delta$  4.04 (s, 3 H), 4.05 (s, 3 H), 6.96 (d, 1 H, J = 4.0 Hz), 7.06 (s, 1 H), 7.20 (d, 1 H, J = 4.0 Hz), 7.41 (s, 1 H), 8.03 (d, 1 H, J = 2.2 Hz), 8.90 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>15</sub>H<sub>12</sub>NO<sub>2</sub>SCl).

**3-(3-Chlorothiophene-2-yl)-6,7-dimethoxyquinoline, 15g,** was obtained from 10a and 14g by method D: mp 168–170 °C (hexane/EtOAc then methanol); NMR (CDCl<sub>3</sub>)  $\delta$  4.03 (s, 3 H), 4.06 (s, 3 H), 7.06 (d, 1 H, J = 5.3 Hz), 7.09 (s, 1 H), 7.34 (d, 1 H, J = 5.3 Hz), 7.44 (s, 1 H), 8.28 (d, 1 H, J = 2.2 Hz), 8.98 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>16</sub>H<sub>12</sub>NO<sub>2</sub>SCl).

**6,7-Dimethoxy-3-(3,4,5-trimethoxyphenyl)quinoline, 15i**, was obtained from **10a** and **12a** by method D: mp 153–155 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  3.93 (s, 3 H), 3.97 (s, 6 H), 4.05 (s, 3 H), 4.07 (s, 3 H), 6.89 (s, 2 H), 7.12 (s, 1 H), 7.46 (s, 1 H), 8.13 (s, 1 H), 8.95 (s, 1 H). Anal. (C<sub>20</sub>H<sub>21</sub>NO<sub>5</sub>).

**6,7-Dimethoxy-3-(4-nitrophenyl)quinoline, 15j,** was obtained from 10a and 12b by method D: mp 222-223.5 °C (EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.06 (s, 3 H), 4.08 (s, 3 H), 7.15 (s, 1 H), 7.48 (s, 1 H), 7.86 (d, 2 H, J = 8.7 Hz), 8.24 (d, 1 H, J = 2.1 Hz), 8.38 (d, 2 H, J = 8.7 Hz), 9.00 (br s, 1 H). Anal. (C<sub>17</sub>H<sub>14</sub>N<sub>2</sub>O<sub>4</sub>·0.25H<sub>2</sub>O).

**6,7-Dimethoxy-3-(4-methoxy-3-nitrophenyl)quinoline, 15k,** was obtained from 10a and 12c by method D: mp 185–187 °C (EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.04 (s, 3 H), 4.05 (s, 3 H), 4.07 (s, 3 H), 7.12 (s, 1 H), 7.26 (d, 1 H, J = 8.8 Hz), 7.46 (s, 1 H), 7.88 (dd, 1 H, J = 2.4, 8.8 Hz), 8.15 (d, 1 H, J = 2.2 Hz), 8.19 (d, 1 H, J= 2.4 Hz), 8.93 (br s, 1 H). Anal. Calcd for C<sub>18</sub>H<sub>16</sub>N<sub>2</sub>O<sub>5</sub>-0.25H<sub>2</sub>O: C, 62.69; H, 4.82; N, 8.12. Found: C, 62.74; H, 4.85; N, 7.32.

**6,7-Dimethoxy-3-thiophene-2-ylquinoline, 15l,** was obtained from 10a and 14k by method D: mp 122.5–124 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.02 (s, 3 H), 4.04 (s, 3 H), 7.06 (s, 1 H), 7.14 (dd, 1 H, J = 3.7, 5.1 Hz), 7.36 (br d, 1 H, J = 5.1 Hz), 7.41 (s, 1 H), 7.43 (br d, 1 H, J = 3.7 Hz), 8.13 (d, 1 H, J = 2.2 Hz), 9.00 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>15</sub>H<sub>13</sub>NO<sub>2</sub>S).

**3-(3,4-Difluorophenyl)-6,7-dimethoxyquinoline, 15m,** was obtained from 10a and 12d by method D: mp 147-149 °C (EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.05 (s, 3 H), 4.07 (s, 3 H), 7.11 (s, 1 H), 7.40 (m, 4 H), 8.11 (s, 1 H), 8.90 (d, 1 H, J = 1.9 Hz). Anal. (C<sub>17</sub>H<sub>13</sub>NO<sub>2</sub>F<sub>2</sub>).

4-(6,7-Dimethoxyquinolin-3-yl)benzoic acid ethyl ester, 15n, was obtained from 10a and 12e by method D: mp 165–166 °C (EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  1.44 (t, 3 H, J = 7.1 Hz), 4.05 (s, 3 H), 4.07 (s, 3 H), 4.42 (q, 2 H, J = 7.1 Hz), 7.13 (s, 1 H), 7.47 (s, 1 H), 7.77 (d, 2 H, J = 8.3 Hz), 8.19 (d, 2 H, J = 8.3 Hz), 8.22 (d, 1 H, J = 2.2 Hz), 9.00 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>20</sub>H<sub>19</sub>NO<sub>4</sub>).

**6,7-Dimethoxy-3-(6-methoxypyridin-3-yl)quinoline**, 150, was obtained from 10a and 12f by method D: mp 170.5–171.5 °C (EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.01 (s, 3 H), 4.05 (s, 3 H), 4.06 (s, 3 H), 6.89 (d, 1 H, J = 8.6 Hz), 7.10 (s, 1 H), 7.44 (s, 1 H), 7.88 (dd, 1 H, J = 2.6, 8.6 Hz), 8.09 (d, 1 H, J = 2.2 Hz), 8.49 (d, 1 H, J = 2.6 Hz), 8.91 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>17</sub>H<sub>16</sub>N<sub>2</sub>O<sub>3</sub>).

**5,8-Dimethyl-3-thiophene-3-ylquinoline, 15p,** was obtained from 10b and 14j by method D: mp 109.5-111 °C (hexane/ EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  2.69 (s, 3 H), 2.79 (s, 3 H), 7.28 (br d, 1 H, J = 7.0 Hz), 7.43 (br d, 1 H, J = 7.0 Hz), 7.50 (m, 1 H), 7.56 (m, 1 H), 7.66 (m, 1 H), 8.42 (d, 1 H, J = 2.1 Hz), 9.23 (d, 1 H, J = 2.1 Hz). Anal. (C<sub>15</sub>H<sub>13</sub>NS).

**6,7-Difluoro-3-thiophene-3-ylquinoline, 15q,** was obtained from **10c** and **14j** by method D: mp 141.5–143.5 °C (hexane/ EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  7.26 (s, 1 H), 7.58 (m, 3 H), 7.86 (dd, 1 H, J = 7.9, 11.0 Hz), 8.22 (br s, 1 H), 9.16 (br s, 1 H). Anal. (C<sub>13</sub>H<sub>7</sub>NSF<sub>2</sub>).

**6,7-Dichloro-3-thiophene-3-ylquinoline, 15r,** was obtained from **10d** and **14j** by method D: mp 167-167.5 °C (hexane/ EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  7.50 (m, 2 H), 7.67 (apparent t, 1 H, J = 2.0 Hz), 7.95 (s, 1 H), 8.16 (d, 1 H, J = 2.0 Hz), 8.22 (s, 1 H), 9.19 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>13</sub>H<sub>7</sub>NSCl<sub>2</sub>).

7-Methoxy-3-thiophene-3-ylquinoline, 15s, was obtained from 10e and 14j by method D: mp 122–124 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  3.97 (s, 3 H), 7.22 (dd, 1 H, J = 2.4, 9.0 Hz), 7.43 (d, 1 H, J = 2.4 Hz), 7.49 (m, 2 H), 7.61 (m, 1 H), 7.73 (d, 1 H, J = 9.0 Hz), 8.21 (d, 1 H, J = 2.2 Hz), 9.12 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>14</sub>H<sub>11</sub>NOS).

7-Thiophene-3-yl[1,3]dioxolo[4,5-g]quinoline, 15t, was obtained from 10j and 14j by method D: mp 159–160 °C (EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  6.12 (s, 2 H), 7.08 (s, 1 H), 7.39 (s, 1 H), 7.48 (m,

#### PDGF Receptor Tyrosine Kinase Inhibitors

2 H), 7.59 (m, 1 H), 8.10 (d, 1 H, J = 2.0 Hz), 8.98 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>14</sub>H<sub>9</sub>NO<sub>2</sub>S).

**5,7-Dimethoxy-3-thiophene-3-ylquinoline, 15u,** was obtained from 10g and 14j by method D: mp 113.5–115 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  3.96 (s, 3 H), 4.00 (s, 3 H), 6.54 (d, 1 H, J = 2.0 Hz), 7.03 (d, 1 H, J = 2.0 Hz), 7.46 (dd, 1 H, J = 2.9, 5.0 Hz), 7.52 (dd, 1 H, J = 1.3, 5.0 Hz), 7.61 (dd, 1 H, J = 1.3, 2.9 Hz), 8.59 (d, 1 H, J = 2.2 Hz), 9.09 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>15</sub>H<sub>13</sub>NO<sub>2</sub>S).

**5-Fluoro-3-thiophene-3-ylquinoline, 15v,** was obtained from **10h** and **14j** by method D: mp 87.5-89 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  7.22 (m, 1 H), 7.51 (m, 2 H), 7.59 (m, 1 H), 7.68 (m, 1 H), 7.90 (d, 1 H, J = 8.5 Hz), 8.50 (d, 1 H, J = 1.9 Hz), 9.22 (s, 1 H). Anal. (C<sub>13</sub>H<sub>8</sub>NFS).

**7-Fluoro-3-thiophene-3-ylquinoline**, **15w**, was obtained from **10i** and **14j** by method D: mp 106.5–108.5 °C (hexane/ EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  7.37 (m, 1 H), 7.51 (m, 2 H), 7.65 (m, 1 H), 7.75 (dd, 1 H, J = 2.4, 10.0 Hz), 7.86 (dd, 1 H, J = 6.0, 9.1 Hz), 8.28 (d, 1 H, J = 2.2 Hz), 9.21 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>13</sub>H<sub>8</sub>NSF).

3-Thiophene-3-ylquinoline, 15x, was obtained from 13e and 14j by method D: mp 87-88.5 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  7.54 (m, 3 H), 7.70 (m, 2 H), 7.85 (d, 1 H, J = 8.2 Hz), 8.11 (d, 1 H, J = 8.2 Hz), 8.29 (d, 1 H, J = 2.1 Hz), 9.21 (d, 1 H, J = 2.1 Hz). Anal. (C<sub>13</sub>H<sub>9</sub>NS).

**5,7-Dimethyl-3-thiophene-3-ylquinoline, 15y,** was obtained from **10f** and **14j** by method D: mp 85-86 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  2.52 (s, 3 H), 2.69 (s, 3 H), 7.23 (s, 1 H), 7.47-7.54 (m, 2 H), 7.63 (m, 1 H), 7.74 (s, 1 H), 8.37 (m, 1 H), 9.14 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>15</sub>H<sub>18</sub>NS).

**3**-trans- $\beta$ -styrylquinoline, 15z, was obtained from 14e and 13i by method D: mp 96–98 °C (hexane); NMR (CDCl<sub>3</sub>)  $\delta$  7.27 (s, 1 H), 7.33 (m, 2 H), 7.41 (m, 2 H), 7.58 (m, 3 H), 7.69 (m, 1 H), 7.83 (d, 1 H, J = 8.3 Hz), 8.09 (d, 1 H, J = 8.3 Hz), 8.19 (d, 1 H, J = 1.8 Hz), 9.13 (d, 1 H, J = 1.8 Hz). Anal. (C<sub>17</sub>H<sub>13</sub>N).

**3-(4-Fluorophenyl)-6,7-dimethoxyquinoline, 17a,** was obtained from 10a and 16a by method E: mp 117.5–119.5 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.04 (s, 3 H), 4.06 (s, 3 H), 7.10 (s, 1 H), 7.20 (m, 2 H), 7.45 (s, 1 H), 7.64 (m, 2 H), 8.11 (d, 1 H, J = 2.0 Hz), 8.93 (d, 1 H, J = 2.0 Hz). Anal. Calcd for C<sub>17</sub>H<sub>14</sub>-NO<sub>2</sub>F: C, 72.07; H, 4.98; N, 4.94. Found: C, 71.51; H, 4.75; N, 4.60.

**6,7-Dimethoxy-3-(2-methoxyphenyl)quinoline**, 17c, was obtained from 10a and 16c by method E: mp 110.5–112 °C (hexane/EtOAc twice); NMR (CDCl<sub>3</sub>)  $\delta$  3.86 (s, 3 H), 4.04 (s, 3 H), 4.06 (s, 3 H), 7.08 (m, 3 H), 7.41 (m, 3 H), 8.14 (d, 1 H, J = 2.0 Hz), 8.90 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>18</sub>H<sub>17</sub>NO<sub>8</sub>).

[4-(6,7-Dimethoxyquinolin-3-yl)phenyl]dimethylamine, 17d, was obtained from 10a and 16d by method E: mp 157–159.5 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  3.03 (s, 6 H), 4.04 (s, 3 H), 4.05 (s, 3 H), 6.86 (d, 2 H, J = 8.7 Hz), 7.08 (s, 1 H), 7.43 (s, 1 H), 7.60 (d, 2 H, J = 8.7 Hz), 8.09 (d, 1 H, J = 1.9 Hz), 8.98 (d, 1 H, J = 1.9 Hz). Anal. (C<sub>19</sub>H<sub>20</sub>N<sub>2</sub>O<sub>2</sub>).

**6,7-Dimethoxy-3-(5-methoxythiophene-2-yl)quinoline**, 17e, was obtained from 10a and 16e by method E: mp 111-113 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  3.95 (s, 3 H), 4.02 (s, 3 H), 4.04 (s, 3 H), 6.24 (d, 1 H, J = 4.0 Hz), 7.03 (s, 1 H), 7.08 (d, 1 H, J = 4.0 Hz), 7.39 (s, 1 H), 7.96 (d, 1 H, J = 2.2 Hz), 8.90 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>16</sub>H<sub>16</sub>NO<sub>3</sub>S).

**3-(2,4-Dimethoxyphenyl)-6,7-dimethoxyquinoline, 17f,** was obtained from 10a and 16f by method E: mp 122-123.5 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  3.84 (s, 3 H), 3.88 (s, 3 H), 4.03 (s, 3 H), 4.05 (s, 3 H), 6.84 (m, 2 H), 7.07 (s, 1 H), 7.33 (d, 1 H, J = 8.0 Hz), 7.43 (s, 1 H), 8.09 (d, 1 H, J = 2.0 Hz), 8.87 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>19</sub>H<sub>19</sub>NO<sub>4</sub>).

**3-[4-(Benzyloxy)phenyl]-6,7-dimethoxyquinoline, 17g,** was obtained from **10a** and **16g** by method E: mp 163.5–165 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.03 (s, 3 H), 4.05 (s, 3 H), 5.14 (s, 2 H), 7.10 (m, 3 H), 7.41 (m, 6 H), 7.62 (d, 2 H, J = 8.7 Hz), 8.10 (d, 1 H, J = 2.2 Hz), 8.95 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>24</sub>H<sub>21</sub>-NO<sub>3</sub>).

**6,7-Dimethoxy-3-[2-(4-methoxyphenyl)vinyl]quinoline, 17h,** was obtained from **10a** and **16h** by method E: mp 145.5–147 °C (EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  3.84 (s, 3 H), 4.02 (s, 3 H), 4.03 (s, 3 H), 6.93 (d, 2 H, J = 8.7 Hz), 7.03 (s, 1 H), 7.05 (d, 1 H, J = **16.4** Hz), 7.20 (d, 1 H, J = 16.4 Hz), 7.40 (s, 1 H), 7.50 (d, 2 H, J = 8.7 Hz), 8.01 (br s, 1 H), 8.88 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>20</sub>H<sub>19</sub>NO<sub>3</sub>).

**6,7-Dimethoxy-3-(4-methoxy-3-methylphenyl)quinoline, 17i, was obtained from 10a and 16i by method E: mp 124.5-**127 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  2.32 (s, 3 H), 3.89 (s, 3 H), 4.02 (s, 3 H), 4.04 (s, 3 H), 6.94 (d, 1 H, J = 8.5 Hz), 7.08 (s, 1 H), 7.47 (m, 3 H), 8.09 (d, 1 H, J = 2.2 Hz), 8.95 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>19</sub>H<sub>19</sub>NO<sub>3</sub>).

**3-(3,4-Dimethoxyphenyl)-6,7-dimethoxyquinoline, 17j,** was obtained from 10a and 16j by method E: mp 120.5–122 °C (hexane/EtOAc twice); NMR (CDCl<sub>3</sub>)  $\delta$  3.96 (s, 3 H), 4.00 (s, 3 H), 4.04 (s, 3 H), 4.06 (s, 3 H), 7.01 (d, 1 H, J = 8.4 Hz), 7.11 (s, 1 H), 7.19 (d, 1 H, J = 2.0 Hz), 7.25 (dd, 1 H, J = 2.0, 8.4 Hz), 7.45 (s, 1 H), 8.11 (d, 1 H, J = 2.1 Hz), 8.95 (d, 1 H, J = 2.1 Hz). Anal. (C<sub>19</sub>H<sub>19</sub>NO<sub>4</sub>).

**6,7-Dimethoxy-3-***p***-tolylquinoline, 17k,** was obtained from **10a** and **16k** by method E: mp 157–159 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  2.43 (s, 3 H), 4.04 (s, 3 H), 4.06 (s, 3 H), 7.10 (s, 1 H), 7.32 (d, 2 H, J = 8.0 Hz), 7.45 (s, 1 H), 7.60 (d, 2 H, J = 8.0 Hz), 8.14 (d, 1 H, J = 2.1 Hz), 8.97 (d, 1 H, J = 2.1 Hz). Anal. (C<sub>18</sub>H<sub>17</sub>NO<sub>2</sub>).

**3-(3,5-Difluoro-4-methoxyphenyl)-6,7-dimethoxyquino**line, 171, was obtained from 10a and 16l by method E: mp 165– 167.5 °C (EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.04 (s, 3 H), 4.06 (s, 6 H), 7.10 (s, 1 H), 7.22 (s, 1 H), 7.25 (s, 1 H), 7.44 (s, 1 H), 8.08 (d, 1 H, J = 1.9 Hz), 8.88 (d, 1 H, J = 1.9 Hz). Anal. (C<sub>18</sub>H<sub>18</sub>-NO<sub>3</sub>F<sub>2</sub>·0.2H<sub>2</sub>O).

**3-(3-Fluoro-4-methoxyphenyl)-6,7-dimethoxyquinoline, 17m,** was obtained from 10a and 16m by method E: mp 165.5– 167 °C (hexane/EtOAc); NMR (CDCI<sub>3</sub>)  $\delta$  3.96 (s, 3 H), 4.04 (s, 3 H), 4.06 (s, 3 H), 7.10 (m, 2 H), 7.48 (m, 2 H), 7.68 (m, 1 H), 8.09 (d, 1 H, J = 2.1 Hz), 8.92 (d, 1 H, J = 2.1 Hz). Anal. (C<sub>18</sub>H<sub>16</sub>-NO<sub>3</sub>F).

**3-Cyclopent-1-enyl-6,7-dimethoxyquinoline, 17n,** was obtained from **10a** and **16n** by method E: mp (HCl salt) 213–215 °C (methanol/Et<sub>2</sub>O); NMR (CDCl<sub>3</sub>, free base)  $\delta$  2.08 (m, 2 H), 2.61 (m, 2 H), 2.81 (m, 2 H), 4.02 (s, 3 H), 4.04 (s, 3 H), 6.38 (t, 1 H, J = 2.0 Hz), 7.04 (s, 1 H), 7.40 (s, 1 H), 7.86 (d, 1 H, J = 2.0 Hz), 8.93 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>16</sub>H<sub>17</sub>NO<sub>2</sub>·HCl-0.6H<sub>2</sub>O).

**6,7-Dimethoxy-3-(phenylethynyl)quinoline**, 170, was obtained from 10a and 160 by method E: mp 147.5-149 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.03 (s, 3 H), 4.05 (s, 3 H), 7.02 (s, 1 H), 7.39 (m, 4 H), 7.58 (m, 2 H), 8.15 (d, 1 H, J = 2.0 Hz), 8.83 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>19</sub>H<sub>15</sub>NO<sub>2</sub>).

**3-(Cyclohexylethynyl)-6,7-dimethoxyquinoline, 17p,** was obtained from 10a and 16p by method E: mp 66–67.5 °C (hexane); NMR (CDCl<sub>3</sub>)  $\delta$  1.37–1.94 (m, 10 H), 2.64 (m, 1 H), 4.01 (s, 3 H), 4.03 (s, 3 H), 6.96 (s, 1 H), 7.39 (s, 1 H), 8.01 (d, 1 H, J = 1.8 Hz), 8.70 (d, 1 H, J = 1.8 Hz). Anal. (C<sub>19</sub>H<sub>21</sub>NO<sub>2</sub>).

**3-Pyridin-3-ylquinoline-6,7-diol**, 18, was obtained by reaction of **2a** with refluxing 48% aqueous HBr: mp >300 °C dec (methanol); NMR (DMSO- $d_6$ )  $\delta$  7.21 (s, 1 H), 7.27 (s, 1 H), 7.53 (dd, 1 H, J = 4.8, 7.8 Hz), 8.21 (m, 1 H), 8.40 (br s, 1 H), 8.60 (m, 1 H), 8.91 (d, 1 H, J = 2.0 Hz), 9.02 (br s, 1 H), 10.08 (br s, 2 H). Anal. (C<sub>14</sub>H<sub>10</sub>N<sub>2</sub>O<sub>2</sub>·0.2H<sub>2</sub>O).

**Benzyl(6,7-dimethoxyquinolin-3-yl)amine, 19,** was obtained by reaction of 8a with benzaldehyde and NaCNBH<sub>3</sub> in methanol: mp (HCl salt) 207-209.5 °C (methanol/Et<sub>2</sub>O); NMR (CDCl<sub>3</sub>, HCl salt)  $\delta$  3.97 (s, 3 H), 4.04 (s, 3 H), 4.56 (s, 2 H), 6.86 (s, 1 H), 7.35 (br m, 6 H), 7.72 (s, 1 H), 9.19 (br s, 1 H). Anal. (C<sub>18</sub>H<sub>18</sub>N<sub>2</sub>O<sub>2</sub>·HCl·0.5H<sub>2</sub>O).

**3-(Benzyloxy)-6,7-dimethoxyquinoline, 20,** was obtained by reaction of **9a** with benzyl bromide and sodium hydride in THF: mp 163.5-165 °C (hexane/EtOAc); NMR (CDCl<sub>3</sub>)  $\delta$  4.03 (s, 3 H), 4.05 (s, 3 H), 5.14 (s, 2 H), 7.10 (m, 3 H), 7.41 (m, 6 H), 7.62 (d, 2 H, J = 8.7 Hz), 8.10 (d, 1 H, J = 2.2 Hz), 8.95 (d, 1 H, J = 2.2 Hz). Anal. (C<sub>24</sub>H<sub>21</sub>NO<sub>3</sub>).

4-(6,7-Dimethoxyquinolin-3-yl)benzoic acid, 22, was obtained by reaction of 15n with NaOH in ethanol: mp (HCl salt) 273-275 °C dec (methanol/CH<sub>2</sub>Cl<sub>2</sub>); NMR (DMSO- $d_6$ , HCl salt)  $\delta$  4.00 (3 H, s), 4.03 (3 H, s), 7.68 (s, 1 H), 7.73 (s, 1 H), 8.03 (d, 2 H, J = 8.3 Hz), 8.10 (d, 2 H, J = 8.3 Hz), 9.15 (br s, 1 H), 9.34 (br s, 1 H). Anal. (C<sub>18</sub>H<sub>15</sub>NO<sub>4</sub>·HCl·0.9H<sub>2</sub>O).

5-(6,7-Dimethoxyquinolin-3-yl)-1*H*-pyridin-2-one, 23, was obtained by reaction of 150 with pyridine hydrochloride at 160 °C for 5 min: mp 259–261 °C (methanol/EtOAc); NMR (DMSO-  $d_{\rm s}$ )  $\delta$  3.91 (s, 3 H), 3.93 (s, 3 H), 6.50 (br d, 1 H, J = 9.5 Hz), 7.31 (br s, 1 H), 7.36 (br s, 1 H), 7.90 (br s, 1 H), 7.97 (br d, 1 H, J = 9.5 Hz), 8.32 (br s, 1 H), 8.92 (br s, 1 H), 11.97 (br s, 1 H). Anal.  $(C_{16}H_{14}N_2O_3).$ 

4-(6,7-Dimethoxyquinolin-3-yl)phenol, 24, was obtained by reaction of 17g with  $H_2$  and Pd/C in methanol and DMF: mp 250–251 °C (methanol); NMR (CDCl<sub>3</sub> + CD<sub>3</sub>OD)  $\delta$  4.05 (s, 3 H), 4.07 (s, 3 H), 6.98 (d, 2 H, J = 8.7 Hz), 7.17 (s, 1 H), 7.38 (s, 1 H), 7.56 (d, 2 H, J = 8.7 Hz), 8.19 (d, 1 H, J = 2.1 Hz), 8.86 (d, 1 H, J = 2.1 Hz). Anal. (C<sub>17</sub>H<sub>15</sub>NO<sub>3</sub>).

6,7-Dimethoxy-3-[2-(4-methoxyphenyl)ethyl]quinoline, 25, was obtained by reaction of 17h with H<sub>2</sub> and Pd/C in methanol and acetic acid: mp 140–141 °C (ethanol); NMR (CDCl<sub>3</sub>)  $\delta$  2.98 (m, 4 H), 3.78 (s, 3 H), 3.99 (s, 3 H), 4.02 (s, 3 H), 6.82 (d, 2 H, J = 8.6 Hz), 6.97 (s, 1 H), 7.08 (d, 2 H, J = 8.6 Hz), 7.39 (s, 1 H), 7.71 (d, 1 H, J = 1.8 Hz), 8.53 (d, 1 H, J = 1.8 Hz). Anal. (C<sub>20</sub>H<sub>21</sub>-NO<sub>3</sub>).

3-(2-Cyclohexylethyl)-6,7-dimethoxyquinoline, 26, was obtained by reaction of 17p with  $H_2$  and  $\mbox{Pd}/\mbox{C}$  in methanol and acetic acid: oil (silica chromatography eluting with 4:1 hexane/ EtOAc); NMR (CDCl<sub>3</sub>) δ 0.96 (m, 2 H), 1.24 (m, 4 H), 1.55–1.81 (m, 7 H), 2.75 (t, 2 H, J = 8.0 Hz), 4.01 (s, 3 H), 4.03 (s, 3 H), 7.00(s, 1 H), 7.40 (s, 1 H), 7.77 (s, 1 H), 8.57 (d, 1 H, d, J = 1.9 Hz).Anal. (C19H25NO2.0.25H2O).

3-Cyclopentyl-6,7-dimethoxyquinoline, 27, was obtained by reaction of 17n with H<sub>2</sub> and Pd/C in acetic acid: mp (HCl salt) 213.5–215 °C (methanol/Et<sub>2</sub>O); NMR (CDCl<sub>3</sub>, free base)  $\delta$  1.80 (m, 6 H), 2.16 (m, 2 H), 3.14 (m, 1 H), 4.01 (s, 3 H), 4.03 (s, 3 H), 7.01 (s, 1 H), 7.39 (s, 1 H), 7.80 (d, 1 H, J = 2.0 Hz), 8.64 (d, 1 H, J = 2.0 Hz). Anal. (C<sub>16</sub>H<sub>19</sub>NO<sub>2</sub>·HCl·0.1H<sub>2</sub>O).

Biology. Materials. Recombinant human PDGF-BB was obtained from R & D Systems (Minneapolis, MN),  $\gamma$ -<sup>32</sup>P-ATP (6000 Ci/mmol) from NEN (Wilmington, DE), and cell culture reagents from GIBCO. Rabbit polyclonal anti-PDGF receptor antibodies were raised against a synthetic peptide from the COOH-terminal region (amino acids 1094-1106) of the human PDGF- $\beta$  receptor.

Cell Culture and Lysis. NIH 3T3 cells (ATCC) were grown in fibronectin-coated 15-cm plates in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% calf serum, 100 units/ mL penicillin-streptomycin and 2 mM glutamine. At  $\sim 90\%$ confluency, the cells were rendered quiescent by switching to DMEM containing 0.5% calf serum for 24 h. The cells were lysed in a detergent buffer (50 mM HEPES, 1.5 mM MgCl<sub>2</sub>, 150 mM NaCl, 1 mM EGTA, 10% glycerol, 1% Triton X-100, pH 7.5) containing 1 mM PMSF, 20 µg/mL aprotinin, and 2 µg/mL leupeptin (all reagents from Sigma), clarified by centrifugation at 16000g for 5 min at 4 °C and the lysates stored at -70 °C.

Cell-Free PDGF Receptor Autophosphorylation Assay. Lysates were diluted 1:2 with lysis buffer devoid of Triton X-100, stimulated with 10 ng/mL PDGF for 30 min at 4 °C and immunoprecipitated with anti-PDGF receptor antibodies adsorbed to Protein A Sepharose beads (Sigma). The immunoprecipitates were washed first with HNTG (20 mM HEPES, 150 mM NaCl, 0.1% Triton X-100, 10% glycerol, pH 7.5) followed by 10 mM Tris, pH 7.5, and suspended in ice-cold reaction buffer (50 mM Tris, 10 mM MnCl<sub>2</sub>, pH 7.5). Test compounds were freshly prepared for each experiment in 100% DMSO as a 40 mM stock solution. The compounds were diluted in reaction buffer and added to the immunoprecipitates for 10 min at 4 °C. Autophosphorylation reactions were initiated by adding an ATP mixture (10  $\mu$ Ci  $\gamma$ -<sup>32</sup>-P-ATP plus 2 mM unlabeled ATP) to the samples, followed by a 5-min incubation at 4 °C. The reactions were terminated by addition of 2X SDS sample buffer (0.14 M Tris, pH 6.8, 22.4% glycerol, 6% SDS, 0.02% bromophenol blue, 10% 2-mercaptoethanol), and the samples were separated by SDS-PAGE on 10% gels. The amount of incorporated ATP was quantitated by densitometry of the resulting autoradiographs. Each compound was tested in a minimum of two dose-response assays. The autophosphorylation reaction occurs essentially on a solid matrix support, and each compound is evaluated repeatedly in several experiments. Since interexperiment variability is significantly higher than the variability within a given experiment (e.g. duplicates), our overall experience with an inhibitory compound is best described by the range of  $IC_{50}$  values observed.

Acknowledgment. We thank Dr. Michael Myers, Ms. Natalie Setzer, and Ms. Allison Zulli for technical support and Dr. Dan McGarry for helpful chemistry discussions. We also thank Dr. Susan Johnson for helping to organize the biological data and for testing 15y.

#### References

- (1) Claesson-Welsh, L. PDGF Receptors: Structure and Mechanism of Action. Cytokines 1993, 5, 31-43. (2) Raines, E. W.; Ross, R. Platelet-Derived Growth Factor in vivo.
- Cytokines 1993, 5, 74-114.
- (3) Jawien, A.; Bowen-Pope, D. F.; Lindner, V.; Schwartz, S. M.; Clowes, A. W. Platelet-derived Growth Factor Promotes Smooth Muscle Migration and Intimal Thickening in a Rat Model of Balloon Angioplasty. J. Clin. Invest. 1992, 89, 507-511.
- (a) Schwartz, R. S.; Holmes, D. R., Jr.; Topol, E. J. The Restenosis Paradigm Revisited: An Alternative Proposal for Cellular Mechanisms. J. Am. Coll. Cardiol. 1992, 20, 1284-93. (b) DeFeudis, F. V. PDGF Antibody and Restenosis. Drug News Perspect. 1992, 5, 49-51. (c) Ferns, G. A.; Raines, E. W.; Sprugel, K. H.; Motani, A.S.; Reidy, M.A.; Ross, R. Inhibition of Neointimal Smooth Muscle Accumulation After Angioplasty by an Antibody to PDGF. Science 1991, 253, 1129-32.
- (5) Spada, A. P.; Persons, P. E.; Levitzki, A.; Gilon, C.; Gazit, A. Styrylsubstituted Monocyclic and Bicyclic Heteroaryl Compounds which Inhibit EGF Receptor Tyrosine Kinase. International Patent Application WO 91/16051, October 31, 1991 (Priority date: April L6, 1990)
- (6) Osborne, C. K.; Reddy, K. B.; Mangold, G.; Ciocca, D. R.; Zilberstein, A. A Tyrosine Kinase Inhibitor Antagonizes In-vitro and In-vivo Growth of Human Breast Cancer Cells. J. Cell Biochem. 1992, Suppl. 16D, 108.
- (7) EGF-RTK in vitro assay protocol described in: Lyall, R. M.; Zilberstein, A.; Gazit, A.; Gilon, C.; Levitzki, A.; Schlessinger, J. Tyrphostins Inhibit Epidermal Growth Factor (EGF)-Receptor Tyrosine Kinase Activity in Living Cells and EGF-stimulated Cell Proliferation. J. Biol. Chem. 1989, 264, 14503-09.
- (8) A similar approach was reported subsequent to our undertaking of this project: (a) Burke, T. R.; Lim, B.; Escobedo, J.; Williams, L. T.; Marquez, V. E. Carboxynaphthyl, -Quinolyl and Isoquinolyl Compounds as Bicyclic Ring-Constrained Analogues of Styryl-Based Protein Tyrosine Kinase Inhibitors Fourth Chem. Cong. North America 1991, Abstr. MEDI 23. (b) Burke, T. R.; Ford, H.; Osherov, N.; Levitzki, A.; Stefanova, I.; Horak, I. D.; Marquez, V. E. Aryl Amides of Hydroxylated Isoquinolines as Protein-Tyrosine Kinase Inhibitors. Bioorg. Med. Chem. Lett. 1992, 2, 1771-4.
- (9) Shamshurin, A. A. Veratral-6-arsonic acid. Chem. Abstr. 1941, 35, 5869.
- (10) Borsche, W.; Ried, W. Über, 2-Amino-veratrumaldehyde und 7.8-Dioxychinaldin. (2-Aminoveratraldehyde and 7,8-dihydroxyquinaldine.) Chem. Ber. 1943, 76, 1011-16. This compound is more easily prepared by the method described in ref 9.
- (11) Tröger, J.; Cohaus, C. Über mittels 6-Amino-3-methoxybenzaldehyd ausgeführte Chinolinsynthesen und über ein aus disem Aldehyde entstehendes Kondensationsprodukt. (Quinoline syntheses carried out with 6-amino-3-methoxybenzaldehyde and a condensation product resulting from this aldehyde.) J. Prakt. Chem. 1927, 117, 97 - 116
- (12) Hey, D. H.; Williams, J. M. New Therapeutic Agents of the Quinoline Series. Part VII. 2-, 3-, and 4-Pyridylquinolines, 4-pyridylquin-aldines, and 2-Pyridyllepidines. J. Chem. Soc. 1950, 1678-83.
- (13) Ihara, M.; Noguchi, K.; Fukumoto, K.; Kametani, T. Conversion of indoles into quinolines through the N-1-C-2 fission by singletoxygen as a model experiment of biomimetic synthesis of quinine alkaloids. *Tetrahedron* 1985, 41, 2109–14.
- (14) Prepared by the method described for 4c in ref 13.
  (15) Kalinin, V. N. Carbon-Carbon Bond Formation in Heterocycles Kamini, V. H. Carbon-Carbon Done of Communication in Proceedings Using Ni- and Pd-Catalyzed Reactions. Synthesis 1992, 413-432.
   (16) (a) Stille, J. K. The Palladium-Catalyzed Cross-Coupling Reactions
- of Organotin Reagents with Organic Electrophiles. Angew. Chem., Int. Ed. Engl. 1986, 25, 508–524. (b) Echavarren, A. M.; Stille, J. K. Palladium-Catalyzed Coupling of Aryl Triflates with Organo-stannanes. J. Am. Chem. Soc. 1987, 109, 5478-86.
- (17) Arcadi, A.; Burini, A.; Cacchi, S.; Delmastro, M.; Marinelli, F.; Pietroni, B. The Palladium-Catalyzed Cross Coupling of Vinyl and Aryl Triflates with 2-Furylzinc Chloride: An Efficient Route to 2-Vinyl- and 2-Arylfurans. Synlett 1990, 47-48. (18) (a) Popp, F. D.; Schuyler, P. Polyphosphoric Acid-catalyzed
- Reactions. Part II. The Synthesis of 3-Nitroquinolines. J. Chem. Soc. 1964, 522-523. (b) Bickerton, R. K. Hypotensive Quinolines. U.S. Pat. 3 178 348, 1965.
- (19) Clemo, G. R.; Swan, G. A. Some Derivatives of 3-Aminoquinoline. J. Chem. Soc. 1945, 867-870.
- Comins, D. L.; Killpack, M. O. Lithiation of Methoxypyridines (20)Directed by  $\alpha$ -Amino Alkoxides. J. Org. Chem. 1990, 55, 69-73.

#### PDGF Receptor Tyrosine Kinase Inhibitors

- (22) Bandodakar, B. S.; Nagendrappa, G. A Simple Preparation of Cyclic Vinylic Bromides (1-Bromocycloalkenes and 1-Bromo-1,5-cyclooctadiene) from 1,2-Dibromocycloalkanes. Synthesis 1990, 843– 844.
- (23) Bilder, G. E.; Kasiewski, C. J.; Walczac, E. M.; Perrone, M. H. PDGF-Receptor Protein Tyrosine Kinase Activity in Carotid Artery Is Enhanced by Injury and Inhibited In Vivo by Tyrphostin RG 13291. Drug Develop. Res. 1993, 29, 158-66.
  (24) The naphthalene analogue of 2e was synthesized by Dr. Paul Persons
- (24) The naphthalene analogue of 2e was synthesized by Dr. Paul Persons and Mr. Cuong Ly of these laboratories and will be further detailed in subsequent publications.
- in subsequent publications.
  (25) Pratesi, P.; Castorina, G. Su alcune N-(β-piperidinoetil)-1.2.3.4-tetraidrochinoline. (1-(2-Piperidinoethyl)-1,2,3,4-tetrahydroquino-line compounds.) Ed. Sci. 1954, 9, 205-211.
- (26) (a) Thakkar, K.; Geahlen, R. L.; Cushman, M. Synthesis and Protein-Tyrosine Kinase Inhibitory Activity of Polyhydroxylated Stilbene Analogues of Piceatannol. J. Med. Chem. 1993, 36, 2950–55. (b) Smyth, M. S.; Stefanova, I.; Hartmann, F.; Horak, I. D.; Osherov,

N.; Levitzki, A.; Burke, T. R. Non-Amine Based Analogues of Lavendustin A as Protein-Tyrosine Kinase Inhibitors. J. Med. Chem. 1993, 36, 3010-14. (c) Hau, C. J.; Jacoski, M. V.; Maguire, M. P.; Spada, A. P.; Zilberstein, A. Inhibition Kinetics of the Tyrosine Kinase Inhibitor Erbstatin and a Pyridone-Based Analog. Biochem. Pharmacol. 1992, 43, 2471. (c) Yaish, P.; Gazit, A.; Gilon, C.; Levitzki, A. Science 1988, 242, 933-935.

- (27) Protein kinase A in vitro assay protocol described in the following: Hagiwara, M.; Inoue, S.; Tanaka, T.; Nunoki, K.; Ito, M.; Hidaka, H. Differential Effects of Flavonoids as Inhibitors of Tyrosine Protein Kinases and Serine/Threonine Protein Kinases. Biochem. Pharm. 1988, 37, 2987-92.
- (28) For an initial disclosure of these compounds see: Spada, A. P.; Maguire, M. P.; Persons, P. E.; Myers, M. R. Bis Mono- and Bicyclic Aryl and Heteroaryl Compounds Which Inhibit EGF and/or PDGF Receptor Tyrosine Kinase. International Patent Application WO 92/20642, November 26, 1992. (Priority date: May 10, 1991).
- (29) Neumann, H.; Seebach, D. Stereospecific preparation of terminal vinyllithium derivatives by Br/Li-exchange with t-butyllithium. Tetrahedron Lett. 1976, 4839-42.