Reactions of α-Chloro Epoxides. 1. Hydrolysis of 1,3-Dichloropropene Oxide

Barry Gold* and Thomas Leuschen

Eppley Institute for Research in Cancer, University of Nebraska Medical Center, Omaha, Nebraska 68105

Received October 31, 1980

A reinvestigation of the hydrolysis of 1,3-dichloropropene oxide (1) is reported. In contrast to a previous report, the hydrolysis product is 2-hydroxy-3-chloropropanal (4), which dimerizes to 2,5-dihydroxy-3,6-bis(chloromethyl)-1,4-dioxane (5). The α -chloroacrylaldehyde (2) originally reported as the hydrolysis product is derived from the primary thermal isomerization product 2,3-dichloropropanal (3).

In a recent paper Kline, Solomon, and Van Duuren studied the hydrolysis and thermolysis of several α -chloro epoxides, including *cis*- and *trans*-1,3-dichloropropene oxide (1a and 1b, respectively).¹ They reported that both 1a and 1b afford α -chloroacrylaldehyde (2) upon hydrolysis in pH 7.4 buffer at 37 °C with pseudo- first-order rate constants ($k_{\rm H}$) determined to be 2.4 × 10⁻³ and 2.3 × 10⁻³ min⁻¹, respectively. They also indicated that thermolysis of 1b at 200 °C in xylene affords 2 and 2,3-dichloropropanal (3) with a measured rate constant (k_i) of 6.1 × 10⁻³ min⁻¹. No yields of 2 or 3 were given for either the hydrolysis or thermolysis experiments.

The results of Kline et al.¹ are not entirely consistent with those obtained in our laboratory. Thus, we report herein a more complete study of the reactions of 1a and 1b (50% each) in aqueous buffer. Reactions of 1a are the same as the cis and trans mixture. We conducted our experiment at 20 °C rather than 37 °C to discern between hydrolysis and thermolysis products. A 3.88×10^{-3} M solution of 1a and 1b in a 50% THF-pH 7.4 phosphate buffer was stirred for 168 h, at which time no epoxide starting material remained. The rate of disappearance of 1a and 1b under these conditions was measured by the method of Barbin et al.² and calculated to a half-life of 800 min. Upon workup the reaction afforded a 52% yield of 3 and 2 in trace amounts. The "true" hydrolysis product 2-hydroxy-3-chloropropanal (4) could not be isolated in pure form because of its propensity to dimerize to 2,5dihydroxy-3,6-bis(chloromethyl)-1,4-dioxane (5). The yield of 5, after silica gel chromatography, was 20%. Control reaction with aldehyde 3 showed that it is not a source of 5.



A CH₂Cl₂ solution of 4, initially contaminated with small amounts of 5, exhibits an IR carbonyl stretch at 1730 cm⁻¹. Upon standing overnight at 20 °C this band completely disappears and a white solid forms. A KBr disc of 5 exhibits intense bands at 3450 (OH) and 845 (CCl) cm⁻¹, but no carbonyl absorption at 1730. The ¹H NMR (Me₂SO-d₆) shows no aldehydic proton but does contain an exchangeable doublet at δ 6.73 (J_{ab} = 5.1 Hz) assigned to the hydroxyl proton (H_a), a doublet of doublets assigned to the C-2 proton H_b at δ 4.90 (J_{ab} = 5.1 Hz, J_{bc} = 1.3 Hz) which collapses to a doublet (J_{bc} = 1.3 Hz) upon addition



of D₂O, a triplet of doublets at δ 4.12 (J_{cd} = 6.2 Hz, J_{bc} = 1.3 Hz) which is unchanged by the addition of D₂O and is asigned to the C-3 proton (H_c), and a multiplet centered at δ 3.38 assigned to the nonequivalent methylene protons (H_d). The extremely intense band at 3450 cm⁻¹ suggests intramolecular hydrogen bonding which would require 5 to be in a twist-boat form. Unfortunately the poor solubility of 5 in nonpolar solvents prevents confirmation of structure by IR or NMR dilution studies. The fully decoupled ¹³C NMR (Me₂SO-d₆) of 5 shows resonances at δ 43.44 (CH₂Cl), 68.36 (C-3), and 88.46 (C-2).

The electron-ionization mass spectrum of 5 did not show a molecular ion, although peaks were observed at m/e137.0006 (59%, C4HeClO3), 109.0029 (43%, C3HeClO2), and 78.9917 (100%, C_2H_4ClO) attributed to the ions resulting from the possible ring fragmentations. A small peak at m/e 107.9967 (2%, $C_3H_5ClO_2$) was observed and is consistent with structure 4. The chemical-ionization mass spectrum exhibits a base peak at m/e 109, resulting from protonated monomer 4, as well as an intense peak (65%) at m/e 91 corresponding to $4H^+ - H_2O$. 4 was characterized by treating the reaction solution with pentafluorophenylhydrazine (6) in ethanol. As anticipated, because of dehydrochlorination, the product from this derivatization was identical with that obtained from authentic pyruvaldehyde. The yield of pyruvaldehyde bis-(pentafluorophenylhydrazone) (7) was 33% after correcting for recovery based on reactions with authentic pyruvaldehyde. Derivatization of 2 with 6 yielded only 4% of the bishydrazone 7.

Aldehyde 2, reported to be the hydrolysis product of 1a and 1b, is actually formed by a thermal rearrangement pathway (see Scheme I).³ We assume that the rate of

⁽¹⁾ S. A. Kline, J. J. Solomon and B. L. Van Duuren, J. Org. Chem., 43, 3596 (1978).

⁽²⁾ A. Barbin, H. Bresil, A. Croisy, P. Jacquignon, C. Malaveille, R. Montesano, and H. Bartsch, *Biochem. Biophys. Res. Commun.*, 67, 596 (1975).

⁽³⁾ R. N. McDonald and T. E. Tabor, J. Am. Chem. Soc., 89, 6573 (1967).

formation of rearrangement product 3 is greatly accelerated in the polar aqueous solvent, as compared to the rate measured in xylene. Recently McDonald and Cousins have reported that the rate of thermolysis for a series of α chlorostyrene oxides is accelerated by a factor of 180 in nitrobenzene relative to CCl₄.⁴ Since the kinetic measurements reported by Kline et al.¹ were based on the assumption that the disappearance of 1a and 1b was directly proportional to the formation of hydrolysis product, the reported $k_{\rm H}$ are incorrect, in that they actually represent the rate of isomerization (k_i) in aqueous solution. Differentation between k_i and k_H is extremely important since $k_{\rm H}$ can be used to relate the electrophilicity of the chloro epoxides to their observed carcinogenic and mutagenic activity.⁵ In contrast to that reported by Kline et al., the initial hydrolysis products of vinyl chloride oxide, cis- and trans-1-chloropropene oxide, trichloroethylene oxide, and tetrachloroethylene oxides are glycolaldehyde, α -hydroxypropionaldehyde, 1,1,2-trichloro-1,2-propanediol, and oxalyl chloride, respectively.6-8 We conclude that the "hydrolysis" rate constants reported by Kline et al. include a large isomerization component.

Experimental Section

Melting points are uncorrected. Spectra were obtained on Beckmann IR-9 (IR), Varian HA-100 and CFT-20 (¹H and ¹³C NMR), and AEI MS-9 (mass spectrum) spectrometers. GLC analyses were carried out on a Varian 3700, using 6 ft \times 2 mm glass columns and a flow rate of 30 mL of N₂/min.

cis- and trans-1,3-dichloropropene oxide (1a and 1b) were synthesized by the method of Kline et al.¹ Approximately 25% of 1b was present in 1a. In addition 10% of 1,3-dichlorobenzene, identified by GLC, ¹H NMR, ¹³C NMR, and mass spectrometry, was formed during the synthesis and removed by a second distillation.

Hydrolysis of 1a and 1b. A solution of 1.97 g (155 mmol) of 1a, or a 50–50 mixture of 1a and 1b, in 250 mL of 50% THF–0.1 M pH 7.4 phosphate buffer was stirred for 168 h, at which time no epoxide was detected (GLC). A 200-mL aliquot of the aqueous solution was extracted with CH₂Cl₂ (2 × 100 mL). The CH₂Cl₂ layer was concentrated to dryness in vacuo and the residue

chromatographed on silicic acid (Bio-Rad Laboratories) with methanol-CH₂Cl₂. The oil obtained afforded a white solid upon standing at room temperature. Additional dimer could be isolated from the aqueous layer by concentration in vacuo and trituration of the residue with CH₂Cl₂, followed by concentration of the CH₂Cl₂. The total yield varied from 13% to 20%: mp 135 °C dec; IR(KBr disk) 3450 (OH) and 845 cm⁻¹ (CCl); ¹H NMR (Me₂SO-d₆) δ 3.38 (m, 4 H, Cl-CH₂), 4.12 (t of d, 2 H, CHCH₂Cl, J = 6.2 and 1.3 Hz), 4.90 (d of d, 2 H, CH–O, J = 5.1 and 1.3 Hz, collapses to d upon addition of D_2O), 6.73 (d, 2 H, OH, J = 5.1Hz, exchanges with D_2O); ¹³C NMR (Me₂SO-d₆) δ 43.44 (CH₂Cl), 68.36 (CCH₂Cl), 88.46 (OCO); mass spectrum (chemical ionization), $m/e 109 [(C_3H_5O_2Cl + H)^+, 100\%], 91 [(C_3H_3OCl + H)^+, 100\%]$ 65%]; mass spectrum (electron ionization), m/e 137.0006 (C4H6ClO3, 59%), 109.0029 (C3H6ClO2, 43%), 78.9917 (C2H4ClO, 100%).

Anal. Calcd for $C_{e}H_{10}O_{4}Cl_{2}$: C, 33.18; H, 4.65; Cl, 32.68. Found: C, 33.35; H, 4.60; Cl, 32.55.

A 10-mL aliquot of the original aqueous reaction solution was extracted with CH_2Cl_2 and analyzed for 3 by GLC (10% SP-2100 on 100/120 Supelcoport, 50 °C/5 min-5 °C/min to 100 °C). Based on recovery experiments with authentic 3 (K+K Labs), the yield was 52%. The trace amount of 2 present (<1%) was detected by ¹H NMR. 3 has chromatographic and spectroscopic properties identical with those of authentic 2,3-dichloropropanal. Neither 4 or 5 could be detected by GLC.

Derivatization with Pentafluorophenylhydrazine. A 10-mL aliquot of the aqueous reaction solution and 50 mL of 0.05 M pentafluorophenylhydrazine (Aldrich) in ethanol was refluxed for 15 h. The solution was then gently concentrated in vacuo and the residue triturated with 5 mL of THF. The yield of pyruvaldehyde bis(pentafluorophenylhydrazone) (7), determined by GLC (3% SE-30 on 80/100 Chromosorb WHP, 220 °C), was 33%. Authentic 3 was treated under identical conditions to yield 4% 7. Isolated 7 was identical with that obtained from derivatization of authentic pyruvaldehyde: mp 122-123 °C (hexane); ¹H NMR (Me₂SO-d₆) 2.08 (s, 3 H, CH₃), 7.73 (s, 1 H, N=CH), 9.00 and 10.63 (br s, NH's); mass spectrum, m/e 432.0436 (M⁺, C_{1b}H₆N₄F₁₀, 43%), 249.0317 (C₉H₄N₃F₅, 63%).

Acknowledgment. We thank Dr. Michael Gross of the Midwest Center for mass spectroscopy at the University of Nebraska—Lincoln, which is supported under the National Science Foundation Regional Instrumentation Facilities Program, and Mr. James Nielsen for mass spectra (Eppley) and Ms. Mardelle Susman for editorial assistance. This work was supported by NIH Grant 1 RO1 CA24554 and USPHS Contract NO1 CP33278.

Registry No. 1a, 66826-72-0; **1b**, 66826-73-1; **2**, 683-51-2; **3**, 10140-89-3; **4**, 69519-13-7; **5**, 76429-86-2; **7**, 76429-87-3.

 ⁽⁴⁾ R. N. McDonald and R. C. Cousins, J. Org. Chem., 45, 2976 (1980).
(5) S. A. Kline, B. L. Van Duuren, E. C. McCoy, H. S. Rosenkranz, and

J. A. DiPaolo, Proc. Am. Assoc. Cancer Res., 21, 78 (1980).
(6) F. P. Guengerich, W. M. Crawford, Jr., and P. G. Watanabe, Bio-

 ⁽⁷⁾ D. Henschler, W. R. Ross, H. Fetz, E. Dallmeier, and M. Metzler,
Biochem Pharmacol. 28, 543 (1979).

Biochem. Pharmacol., 28, 543 (1979). (8) D. M. Frankel, C. E. Johnson, and H. M. Pitt, J. Org. Chem., 22, 1119 (1957).