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Synthesis, antiproliferative activity and apoptosis-promoting effects of arene ruthenium(II) complexes with N, O chelating ligands

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7 A B S T R A C T

New half sandwich arene ruthenium(II) complexes of the type [Ru(arene)Cl(L)] 8 (where arene= benzene and p-cymene, L = thiophene benzhydrazone ligands) have been 9 synthesized from the reactions of the neutral precursor [Ru(arene) (μ -Cl) Cl]₂ and the 10 corresponding benzhydrazone ligand. All the complexes were completely characterized by 11 elemental analysis and additionally by IR, UV-Vis, ¹H NMR and ESI-MS spectroscopic 12 methods. The solid state structures of the complexes 6 and 7 were determined by single-13 crystal X-ray diffraction analysis, which exhibit typical pseudo-octahedral geometry around 14 15 the metal center. The antiproliferative activity of the complexes was evaluated on cancerous (HeLa, MDA-MB-231, and Hep G2) and noncancerous (NIH3T3) cell lines. In general, 16 17 complexes containing electron releasing OCH₃ substituent have potential anticancer activity than those incorporating H, Cl and Br substituents. Moreover, the p-cymene complexes show 18 more cytotoxicity than benzene derivatives, suggesting that the substituent at arene plays a 19 vital role in the biological activity of the compounds. Further, an apoptotic mechanism of 20 cell death in MDA-MB-231 was confirmed by AO-EB, Hoechst 33258 staining and annexin-21 V/PI double-staining techniques. In addition, the extent of DNA fragmentation in cancer cells 22 was studied by comet assay. 23

24

25 Keywords:

26 Benzhydrazone; η^6 -arene ruthenium(II) complex; Crystal structure; Cytotoxicity; Apoptosis

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33 **1. Introduction**

Although platinum-based drugs cisplatin, carboplatin and oxaliplatin have been 34 widely used for anticancer agents for the past few decades, the problems of high toxicity, 35 platinum resistance and undesirable side-effects are appealing the search for different 36 transition metal anticancer drugs. It is to note that ruthenium-complexes have attracted 37 significant attention among the other various metal complexes for their potential anticancer 38 activity. In this regard ruthenium complexes exhibit evidence of low toxicity compared to 39 traditional cisplatin agents. The ruthenium(III) complexes particularly [imiH₂] [trans-Ru(N-40 imiH)(S-dmso)Cl4] NAMI-A and [indH2][trans-Ru(N-indH)2Cl4] KP1019 [1,2] and its 41 sodium analogue Na[trans-Ru(N-indH)2Cl4] (NKP-1339 or IT-139) are the most promising 42 ruthenium complexes reaching clinical trials [3]. Notably, the activation method depends on 43 the redox potential of the Ru(III)/Ru(II) oxidation states, which in turn strongly depends on 44 the ligands coordinated to the metal centre. The activation by reduction results in a reactive 45 ruthenium(II) complex, which can react with numerous biomolecules [4-7]. 46

Particular attention has been paid to half sandwich arene ruthenium complexes 47 because of the π -ligated arene which confers great stability to Ru in the +2 oxidation state 48 and influences the hydrophobicity and interaction with biomolecules [8-10]. Substitutions at 49 50 arene moiety and variations in the chelating ligands will be able to fine tune their biological 51 properties [11]. Tocher et al. have reported that cytotoxicity was enhanced by coordinating the antibacterial agent metronidazole [1-β-(hydroxyethyl)-2-methyl-5-nitro-imidazole] to a 52 benzene ruthenium dichloro fragment [12]. At first, the prototype arene ruthenium(II) 53 $[(p-MeC_6H_4Pr^{i})RuCl_2(P-pta)]$ 54 complexes (pta = 1,3,5-triaza-7-phospha-tricyclo-[3.3.1.1]decane), termed RAPTA-C [13] which displays pH dependent DNA damage due to 55 the hypoxic (low pH) nature of cancer cells, and $[(C_6H_5Ph)RuCl(N,N-en)][PF6]$ (en = 1,2-56 ethylenediamine) exhibits selective binding to guanine bases on DNA, forming 57 monofunctional adducts [14], though many various categories have since been reported [15]. 58

Hydrazones are versatile ligands with fascinating ligation properties with many transition metals. Moreover, these ligands represent an important class of compounds for new drug development because hydrazone moiety was selected for its high stability at physiological pH and lability under strongly acidic and basic conditions as incontestable by drug delivery agents in tumor targeting. Thus, all the hydrazones possess the azomethine (-CONHN=CH-) group have been revealed to exhibit antiproliferative activities and act as cytotoxic agents

with the ability to stop cell progression in cancerous cells through different mechanisms [16].
Aroylhydrazones are magnificent multidentate ligands for transition metals. They have been
exhibit to reveal a variety of biological e.g. antiamoebic activity [17] and DNA synthesis
inhibition or antiproliferative behaviour [18-20]. Herein, we present a systematic
investigation of half-sandwich Ru(II) complexes bearing benzhydrazone ligands (Fig. 1)
with respect to their antiproliferative activity on human cancer cells.



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Fig. 1 Design of arene ruthenium(II) benzhydrazone complexes.

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74 **2. Results and discussion**

The benzhydrazones were obtained by condensation of equimolar amounts of 75 thiophene-2-carboxyaldehyde and substituted benzhydrazide [21]. The arene complexes of 76 77 the type [Ru(arene)Cl(L)] (arene= benzene and *p*-cymene and L = thiophene benzhydrazone ligands) (Scheme 1) have been synthesised from the reactions of the ligands and ruthenium 78 79 arene dimers [Ru(arene) (μ -Cl)Cl]₂ in a 2 : 1 molar ratio in benzene for 5h at reflux temperature in the presence of triethylamine as a base. The isolated complexes were yellow, 80 brown in colour, air stable solids, partially soluble in water and completely soluble in polar 81 82 organic solvents like methanol, ethanol, acetone, chloroform, dichloromethane, acetonitrile, dimethylformamide and dimethylsulfoxide. The elemental analysis of all the ruthenium(II) 83 complexes are in good agreement with the molecular formula of the proposed structure. 84



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Scheme 1. Synthesis of arene ruthenium(II) benzhydrazone complexes.

FT-IR spectra of the ligands and the complexes (1-8) furnished significant 89 information about coordination of the ligand to metal. A medium to strong band in the range 90 3191-3280 cm⁻¹ was assigned to the N-H functional group of the ligand. The ligands also 91 exhibit absorptions due to $v_{C=N}$ and $v_{C=O}$ within the range 1632-1649 cm⁻¹. Upon 92 complexation the bands associated with v_{N-H} and $v_{C=0}$ stretching vibrations are disappeared 93 and indicating that the ligands undergo tautomerization and consequent coordination of the 94 imidolate oxygen. The appearance of new bands in the range 1259-1272 and 1594-1620 cm⁻¹ 95 attributed to the C-O and C=N-N=C fragments which give further support for the 96 97 coordination of the ligand. Hence, the coordination through imine nitrogen and the imidolate oxygen of the ligand to ruthenium was confirmed by IR spectra of all the complexes [22]. All 98 the complexes show three bands in the region 234-366 nm in acetonitrile at room 99 temperature. Bands due to ligand-centered (LC) transitions are appeared around 234-304 nm 100 and have been designated as $\pi - \pi^*$ and $n - \pi^*$ transitions. The lowest energy bands that 101 appeared in the region 360-366 nm were attributed to the charge transfer due to metal to 102 ligand transitions [23]. The pattern of the electronic spectra of all the complexes is very 103 similar to other previously reported octahedral complexes. Fig. S1-S8 (ESI⁺). 104

105 The binding of the benzhydrazone ligand to the ruthenium(II) ion is further verified 106 by NMR spectra of the complexes. All the complexes show multiplets in the region δ 6.7- 8.1 107 ppm and have been assigned to the aromatic protons of benzhydrazone ligands. A sharp 108 singlet in the region δ 8.8-8.9 ppm is assigned to azomethine proton which shifted to 109 downfield on comparison with those of the free ligands, indicating deshielding of the 110 azomethine proton upon coordination to ruthenium. In addition, the absence NH proton of the

free ligands in all the complexes confirmed the coordination to Ru(II) ion via imidolate 111 oxygen. An upfield shift of η^6 -C₆H₆ protons of 1-4 has been observed in the region of δ 5.5 112 ppm. Two sets of doublets have been observed in the region δ 1.0-1.3 ppm for the methyl 113 protons of isopropyl group in *p*-cymene moiety. The methine proton of the isopropyl group 114 appears as a septet in the range of δ 2.5-2.6 ppm. Further, a singlet at δ 2.2 ppm is attributed 115 to the methyl protons of the *p*-cymene moiety. Moreover, four sets of doublets in the range 116 of δ 4.6-5.3 ppm were assigned to the aromatic protons of the *p*-cymene ligand. In addition, 117 for complexes 4 and 8 the methoxy signals of the benzhydrazone ring were observed as a 118 singlet at δ 3.8 and δ 3.7 ppm. Thus the ¹H NMR spectra of all the complexes confirm the 119 coordination mode of the benzhydrazone ligand to the ruthenium(II) ion through the 120 azomethine nitrogen and the imidolate oxygen Fig. S9-S16 (ESI⁺). 121

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123 **2.1 Crystal structures**

Single crystal X-ray diffraction analysis of the complexes 6 and 7 were grown from 124 CH₂Cl₂/ Pet .ether by slow evaporation method. The ORTEP diagrams for the two structures 125 are shown in Fig. 2, crystallographic data and selected bond parameters are listed in Table 1 126 and 2. Both complexes 6 and 7 crystallize in the monoclinic space group $P2_1/c$. In the 127 complex 6, the (η^6 -*p*-cymene) ligand occupying three coordination sites in η^6 -fasion and the 128 remaining coordination sites are occupied by N, O donor atoms from chelating ligand and 129 one chloride. Thus the crystallographic structure of complex confirms pseudo octahedral 130 geometry around the ruthenium metal [24]. The Ru-N, Ru-O and Ru-Cl bond lengths are 131 2.107(4), 2.056(3) and 2.398(13) Å, respectively. The Ru-C (p-cymene) bond lengths ranging 132 from 2.157-2.221 Å and p-cymene ring C-C bond lengths ranging from 1.398-1.432 Å. Bond 133 angles of 86.18(11)⁰, 85.73(11)⁰ and 76.23(13)⁰ are observed for Cl-Ru-O, Cl-Ru-N, and N-134 Ru-O respectively. A similar structural feature has been found in complex 7 with marginal 135 changes in bond lengths and bond angles. 136

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ΈD CRIPT



Fig. 2 Molecular structures of complexes 6 and 7; thermal ellipsoids are drawn at the 30% probability level. All 140 hydrogen atoms were omitted for clarity.

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Table 1 Selected Bond Lengths (Å) and Angles (deg) for the Complexes 6 and 7 145

	6		7	
	Bond lengths (Å)			
	N(1)-N(2)	1.413(5)	N(1)-N(2)	1.410(6)
	N(1)- $Ru(1)$	2.107(4)	N(2)-Ru(1)	2.107(4)
	O(1)-Ru(1)	2.056(3)	O(1)-Ru(1)	2.053(3)
	Cl(1)- $Ru(1)$	2.398(13)	Cl(1)-Ru(1)	2.400(15)
	C(7)-O(1)	1.305(5)	C(7)-O(1)	1.300(6)
	C(7)-N(2)	1.299(6)	C(7)-N(1)	1.300(6)
	C(8)-N(1)	1.288(6)	C(8)-N(2)	1.296(6)
	Bond angles (⁰)			
	N(2)-N(1)-Ru(1)	113.5(3)	N(1)-N(2)-Ru(1)	113.7(3)
	C(7)-N(2)-N(1)	110.9(4)	C(7)-N(1)-N(2)	110.9(4)
	C(7)-O(1)-Ru(1)	112.6(3)	C(7)-O(1)-Ru(1)	112.9(3)
	O(1)-Ru(1)-N(1)	76.23(13)	O(1)-Ru(1)-N(2)	76.09(15)
	O(1)-Ru(1)-Cl(2)	86.18(11)	O(1)-Ru(1)-Cl(1)	85.76(12)
	N(1)-Ru(1)-Cl(2)	85.73(11)	N(2)-Ru(1)-Cl(1)	85.80(12)
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Table 2 Crystal data and structure refinement for complexes **6** and **7**

Compound	6	7
Empirical formula Formula weight	C ₂₂ H ₂₂ Cl ₂ N ₂ O ₂ Ru S 550.45	C ₂₂ H ₂₂ Br Cl N ₂ O ₂ Ru S 594.91
Temperature Wavelength Crystal system	296(2) K 0.71073Å Monoclinic	296(2) K 0.71073 Å Monoclinic
Space group Unit cell dimensions	P21/c a = 13.9604(5)Å $alpha = 90 deg.$	P21/c a = $13.9337(6)$ Å alpha = 90 deg.
	b = 17.0717(6) Å beta = 100.359(2) deg.	b = $17.2743(8)$ Å beta = $101.267(2)$ deg.
Volume	c = 10.2349(4)A gamma = 90 deg.	c = 10.3593(4) A gamma = 90 deg.
Z, Calculated density Absorption coefficient	4, 1.521Mg/m ³ 0.981 mm ⁻¹	4, 1.616 Mg/m ³ 2.490 mm ⁻¹
F(000) Crystal size	1112 0.30 x 0.30 x 0.25 mm	1184 0.35 x 0.30 x 0.30mm
Theta range for data collection Limiting indices	1.48 to 28.35 deg. $-18 \le h \le 12$, $-22 \le k \le 22$, $13 \le l \le 13$	1.49 to 28.32 deg. -17 $\leq h \leq 18$, -19 $\leq k \leq 22$, 13 $\leq 1 \leq 10$
Reflections collected / unique	19987 / 5955 [R(int) = 0.0291]	19409 / 5975 [R(int) = 0.0324]
Completeness to theta = 28.44 Absorption correction	99.2 % Semi-empirical from equivalents	98.3 % Semi-empirical from equivalents
Max. and min. transmission	0.7915 and 0.7573	0.5221 and 0.4761
Refinement method	Full-matrix least-squares on F ²	Full-matrix least-squares on F ²
Data / restraints / parameters	5955 / 0 / 271	5975 / 0 / 271
Goodness-of-fit on F ² Final R indices [I>2sigma(I)]	1.124 R1 = 0.0505, wR2 = 0.1655	1.088 R1 = 0.0505, wR2 = 0.1631
R indices (all data)	R1 = 0.0679, wR2 = 0.1888	R1 = 0.0765, wR2 = 0.1816
Largest diff. peak and hole	2.583 and -0.677 e.Å ⁻³	2.342 and -0.753 e.Å ⁻³

2.2 Stability of the complexes (time-dependent spectra)

Stability of compounds in solution is an essential requirement for drug candidates. 160 The stability of complexes (1-8) in a solution of buffer-DMSO was explored using UV-Vis 161 162 spectroscopy Fig.S9-S16 (ESI⁺). The spectra did not exhibit any noticeable changes during a period of 24 hour indicate the stability of the complexes. Further, ESI-MS spectral studies of 163 the complexes confirm the composition. All the complexes showed the characteristic peaks at 164 m/z 410.00 (1, M-Cl⁺), 444.96 (2, M-Cl⁺), 486.89 (3, M - Cl⁺), 439.00 (4, M-Cl⁺), 465.05 165 (5, M-Cl⁺), 499.06 (6, M-Cl⁺), 544.96 (7, M-Cl⁺), and 495.06 (8, M-Cl⁺) Fig. S25-S32 166 (ESI[†]). The results strongly indicate that the chlorine atom in these complexes is highly labile 167 168 and the resulting species easily interacts with biomolecules [25].

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170 2.3 Partition Coefficient Determination

171 Hydrophobicity is the basic physiochemical parameters in the design of drugs and 172 their biological processes [26] and is determined by the n-octanol/water partition coefficient 173 (P) method [27]. Moreover, Log P, were measured to explain the permeability of complexes 174 (1-8) through a biological system [28] based on solubility of a given compound in a two-175 phase system [29]. The log P results are presented in Table S1(ESI[†]). The partition 176 coefficient values (log P) of the complexes suggested that hydrophobicity can be arranged in 177 the order 8 > 4 > 6 > 7 > 5 > 2 > 3 > 1.

178

179 **2.4 Cytotoxicity studies**

The cytotoxicities of the metallic precursors, ligands and complexes were 180 determined by spectrofluorimetric MTT assay. The plot of percentage of cell death versus 181 concentration is illustrated in Fig. S33&34 (ESI[†]). The cytotoxicity of the complexes was 182 expressed by IC_{50} values and are reported in Table 3. It is to be noted that the precursor and 183 the ligand did not show any inhibition even up to 100 µM and the observed cytotoxicity of 184 the complexes is mainly due to chelation of the ligand to ruthenium. The *in vitro* anticancer 185 activity of the Ru-arene complexes 1-8 towards several human cancer cell lines (HeLa, 186 MDA-MB-231, and Hep G2) and a normal human cell line (NIH3T3) were determined after 187 24 h inhibition and cisplatin was used as a positive control. Based on IC₅₀ values obtained, in 188 *vitro* anticancer activity of the complexes follows the order: 8>4>6>7>5 cisplatin = 1>2>3. 189 These results are also consistent with hydrophobicity of the complexes [30]. Complexes 1–8 190 191 show markedly increased cytotoxic potencies compared with the respective hydrazone

ligands. A comparison of the IC₅₀ values of these complexes against MDA-MB-231cells 192 indicates that complexes 4 and 8 exhibits comparatively better than the other complexes 193 under same experimental conditions. The complexes containing methoxy substituent exhibit 194 higher hydrophobicity and enables permeation of complexes across cell membranes [31]. 195 196 Further, the arene group plays significant role in the antiproliferative activity of these complexes. In general *p*-cymene complexes show higher cell killing activities which may be 197 198 due to the higher hydrophobic interactions between p-cymene complexes and the biomolecules. Thus, the in vitro anticancer activity of the complex towards NIH-3T3 (non-199 cancerous cells) was determined to be above 221 µM, confirms that these complexes are 200 specific for cancer cells. 201

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203	Table 3 The cytotoxic activit	v of arene ruthenium(II) benzhv	vdrazone complexes after 24 h expo	sure
200		, or arone radiumann(ii) conzi	farabone comprehes arter 2 in enpo	bare

Complexes	^a IC ₅₀ values (μ M)			
Complexes	HeLa	MDA-MB-231	Hep G2	NIH3T3
LI	<mark>>100</mark>	<mark>>100</mark>	<mark>>100</mark>	<mark>>100</mark>
<mark>L2</mark>	>100	>100	<mark>>100</mark>	<mark>>100</mark>
L3	<mark>>100</mark>	>100	<mark>>100</mark>	<mark>>100</mark>
L4	>100	>100	<mark>>100</mark>	<mark>>100</mark>
[(benzene)RuCl ₂] ₂	>100	>100	>100	<mark>>100</mark>
[(p-Cymene)RuCl ₂] ₂	<mark>>100</mark>	>100	>100	>100
1	32.5 ± 0.3	19.6 ± 0.3	26.9 ± 0.1	223.7 ± 0.8
2	28.6 ± 0.4	18.7 ± 0.2	21.3 ± 0.5	232.4 ± 0.3
3	31.2 ± 0.2	19.1 ± 0.3	22.0 ± 0.2	236.1 ± 0.3
4	10.2 ± 0.5	9.8 ± 0.2	12.0 ± 0.3	272.9 ± 0.4
5	22.9 ± 0.5	16.8 ± 0.2	21.9 ± 0.6	261.3 ± 0.9
6	15.4 ± 0.3	10.5 ± 0.1	18.4 ± 0.3	242.6 ± 0.2
7	17.2 ± 0.5	11.8 ± 0.2	18.5 ± 0.2	243.6 ± 0.3
8	9.4 ± 0.2	8.3 ± 0.4	10.9 ± 0.2	288.0 ± 0.5
Cisplatin	22.6 ± 0.8	14.9 ± 0.5	21.3 ± 0.9	221.3 ± 0.6

^{204 &}lt;sup>a</sup>IC₅₀ = concentration of the drug required to inhibit growth of 50% of the cancer cells (μ M).

²⁰⁵ The sign (>) indicates that IC_{50} value was not obtained up to given concentration.

²⁰⁶

²⁰⁷

209 **2.5 Morphological changes in AO and EB dual staining**

An Acridine Orange–Ethidium Bromide (AO–EB) dual fluorescent staining method was used to investigate apoptosis in a MDA-MB-231cell line treated with complex 4 and 8. After treatment of cells with the complexes 4 and 8 for 24 h and irradiated with visible light showed significant reddish-orange emission with condensed chromatin and membrane blebbing. In the control, the cells of MDA-MB-231 were stained bright green in spots. Henceforth, the morphological changes clearly indicate that the complexes induce cell death through apoptosis.



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Fig. 3 Morphological assessment of AO and EB dual staining of MDA-MB-231cells treated with complex 4 & 8
(IC₅₀ concentration) for 24 h. The scale bar 20 mm.

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221 2.6 Morphological changes in Hoechst 33258 staining

To investigate the nuclear morphologic characteristics, MDA-MB-231 cells were stained with Hoechst 33258 and treated with complexes 4 and 8 using fluorescence microscopy. After 24 h, complexes treated cells showed fragmented nuclei and chromatin condensation which are features of apoptosis different from control cells (Fig.4).

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Fig. 4 Morphology of the nuclei of MDA-MB-231 cells observed by fluorescence microscopy (Hoechst 33258
 staining, 24 h incubation at IC₅₀ concentrations) after treatment with control complexes 4 and 8.

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233 2.7 Evaluation of apoptosis – Flow cytometry

As shown in Fig. 5 and 6, MDA-MB-231cells were treated with complex 4 and 8 at two different concentrations for 24 h. The increase of annexin V+/PI+ (Q2) population from 3.7% to 6.7% for 4 and 5.0% to 8.2% for 8 at 50 and 100 μ M concentrations of the complexes respectively represent cells undergoing apoptosis. Taken together, these results indicate that cell death induced by complexes is mainly caused by induction of apoptosis.

239

240



Fig. 5 Annexin V/propidium iodide assay of MDA-MB-231cells treated by complex 4 (50 and 100 μM
 concentration) measured by flow cytometry.

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249 **2.8 Comet assay**

The comet assay was used to detect the DNA strand breaks with high sensitivity at the single-cell level [32]. As shown in Fig. 7, MDA-MB-231cells treated with IC₅₀ concentration

of the complexes 4 and 8 for 24h show the increase in the length of the comet tail and illustrate that the complexes induce a remarkable DNA damage in a time-dependent manner, the percentage of DNA damage presented in Fig.S35 (ESI[†]). Further, the results of comet assay demonstrate that the complexes are capable of eliciting DNA damaging effects, as evidenced by the comet assays on MDA-MB-231 cells

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Fig. 7 Comet assay of EB-stained control, complex 4 and 8 treated breast cancer cells at 24h incubation.

260

261 **3.** Conclusions

In summary, we have described the synthesis of a series of arene ruthenium(II) 262 benzhydrazone complexes. All the complexes have been completely characterized by 263 analytical techniques and spectroscopic methods. Crystallographic studies of the complexes 6 264 265 and 7 have shown that the benzhydrazone ligands are coordinated to Ru(II) in a bidentate fashion via azomethine nitrogen and imidolate oxygen atoms. Besides, all the complexes 266 were tested for anticancer activity against HeLa, MDA-MB-231, and Hep G2 cancer cell 267 lines, and they were found to show excellent cytotoxicity to cancer cells without affecting the 268 normal NIH 3T3 cells. Remarkably, complexes 4 and 8 display high cytotoxicity against 269 cancer cell lines tested with very low IC₅₀ values. Moreover, fluorescence staining 270 techniques, flow cytometry and comet assays demonstrated that complexes induce apoptosis 271 in MDA-MB-231 cells. Hence, confirming that these arene ruthenium(II) benzhydrazone 272 complexes have promising biological properties and are worth investigating further. 273

274

275 **4. Experimental**

276 **4.1 Reagents and materials**

277 RuCl₃.3H₂O was purchased from Loba Chemie Pvt. Ltd. and used as received.
278 Aldehydes and benzhydrazide derivatives were obtained from Aldrich. All other chemicals

were purchased from commercial sources and used without further purification. The Solvents were distilled following the standard procedures [33] and degassed prior to use. [Ru(arene) (μ -Cl)Cl]₂ (arene= benzene and *p*-cymene) was prepared by reported procedure [34].

283

4.2 Physical measurements

285 FT-IR spectra in KBr pellets were recorded on a JASCO 400 plus spectrometer. Microanalysis of carbon, hydrogen, nitrogen and sulphur were carried out by Vario EL III 286 CHNS elemental analyzer. UV- visible spectra was recorded on a CARY 300 Bio UV- Vis 287 spectrometer. The ¹H NMR spectra were carried out with Bruker 400 MHz instruments. 288 Melting points were determined on a Boetius micro-heating table and are corrected. ESI-MS 289 spectra were obtained by micro mass Quattro II triple quadrupole mass spectrometer. The 290 annexin V-FITC kit (APOAF-20TST) from Sigma-Aldrich was used based on manufacturer 291 instructions. 292

293

4.3 Preparation of thiophene benzhydrazone ligands

A solution of thiophene-2-carboxyaldehyde (5 mmol) in ethanol (10mL) was added drop wise to the ethanol solution (10 mL) of 4-substituted benzhydrazide (5 mmol) and the reaction mixture was refluxed for about 3 h. The solution was concentrated to 5 ml and cooled to room temperature. The cream or pale brown solid formed was filtered, washed with cold methanol (5mL) and dried in air. Yield 83-88%.

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301 **4.4 Synthesis of arene ruthenium(II) benzhydrazone complexes**

A mixture of $[Ru (\eta^6-C_6H_6) (\mu-Cl)Cl]_2$ or $[Ru (\eta^6-p-cymene) (\mu-Cl)Cl]_2 (0.04 mmol)$ and benzhydrazone ligand (0.08 mmol) was refluxed in benzene in the presence of triethylamine (0.5 mL) for 5 h. After removing the triethylammonium chloride by filtration, the solution was concentrated and light petroleum ether (bp 60-80 °C) was added whereby the solid separated out. The resulted solids were recrystallized from CH₂Cl₂/petroleum ether and dried under vacuum.

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4.4.1 [**Ru**(η^6 -**C**₆**H**₆)(**Cl**)(**L1**)] (1). Colour: Brown; Yield: 80%; M.p.: 165 ^oC; Anal. Calc. For C₁₈ H₁₅ Cl N₂ O Ru S: C, 48.70; H, 3.40; N, 6.31; S, 7.22%. Found: C, 48.52; H, 3.45; N, 6.30; S, 7.25%. IR (KBr, cm⁻¹):1598 v_(C=N-N=C), 1265 v_(C-O). UV–Vis (CH₃CN, λ max/nm;

312 $ε/dm^3 mol^{-1} cm^{-1}$): 354(3415), 274(4254), 236(6652). ¹H NMR (400 MHz, CDCl₃) (δ ppm):3138.9 (s, 1H, N=CH), 7.1–8.1 (m, 8H, aromatic), 5.5(s, 6H). ESI-MS (CH₃CN): calcd for C₁₈314H₁₅ Cl N₂ O Ru S m/z 443.96; found [M - Cl]⁺:410.00.

315

4.4.2 [**Ru**(η⁶-C₆H₆)(**Cl**)(**L2**)] (**2**). Colour: Brown; Yield: 77%; M.p.: 163 ^oC; Anal. Calc. For C₁₈ H₁₄ Cl₂ N₂ O Ru S: C, 45.19; H, 2.94; N, 5.85; S, 6.70%. Found: C, 45.35; H, 2.90; N, 5.88; S, 6.72%. IR (KBr, cm⁻¹):1620 $\nu_{(C=N-N=C)}$, 1260 $_{\nu(C-O)}$. UV–Vis (CH₃CN, λ max/nm; ε/dm³ mol⁻¹ cm⁻¹): 366(1855), 281(2394), 238(4473). ¹H NMR (400 MHz, CDCl₃) (δ ppm): 8.9 (s, 1H, N=CH), 7.1–8.1 (m, 7H, aromatic), 5.5 (s, 6H). ESI-MS (CH₃CN): calcd for C₁₈ H₁₄ Cl₂ N₂ O Ru S m/z 477.92; found [M - Cl]⁺:444.96.

322

4.4.3 [**Ru**(η^{6} -C₆H₆)(**Cl**)(**L3**)] (**3**). Colour: Brown; Yield: 74%; M.p.: 161 ⁰C; Anal. Calc. For C₁₈ H₁₄ Br Cl N₂ O Ru S: C, 41.35; H, 2.69; N, 5.35; S, 6.13%. Found: C, 41.58; H, 2.67; N, 5.36; S, 6.19%. IR (KBr, cm⁻¹):1612 v_(C=N-N=C), 1256 v_(C-O). UV–Vis (CH₃CN, λ max/nm; ε/dm³ mol⁻¹ cm⁻¹): 361(7045), 273(8836), 244(12972). ¹H NMR (400 MHz, CDCl₃) (δ ppm): 8.9 (s, 1H, N=CH), 7.1–8.1 (m, 7H, aromatic), 5.5(s, 6H). ESI-MS (CH₃CN): calcd for C₁₈ H₁₄ Br Cl N₂ O Ru S m/z 521.87; found [M - Cl]⁺:486.89.

329

4.4.4 [**Ru**(η^{6} -C₆H₆)(**Cl**)(**L4**)] (**4**). Colour: Brown; Yield: 72%; M.p.: 157 ^oC; Anal. Calc. For C₁₉ H₁₇ Cl N₂ O₂ Ru S: C, 48.15; H, 3.61; N, 5.91; S, 6.76%. Found: C, 48.25; H, 3.67; N, 5.95; S, 6.71%. IR (KBr, cm⁻¹):1594 v_(C=N-N=C), 1272 v_(C-O). UV–Vis (CH₃CN, λ max/nm; ϵ/dm^{3} mol⁻¹ cm⁻¹): 360(5095), 279(5742), 253(7314). ¹H NMR (400 MHz, CDCl₃) (δ ppm): 8.9 (s, 1H, N=CH), 6.8–8.1 (m, 7H, aromatic), 5.5(s, 6H), 3.8 (s, 3H, OCH₃). ESI-MS (CH₃CN): calcd for C₁₉ H₁₇ Cl N₂ O₂ Ru S m/z 473.97; found [M + H]⁺ :474.98, [M - Cl]⁺ :439.00.

337

4.4.5 [**Ru**(η⁶-*p*-cymene)(Cl)(L1)] (5). Colour: Yellow; Yield: 85%; M.p.: 188 ⁰C; Anal. Calc. For C₂₂ H₂₃ Cl N₂ O Ru S: C, 52.84; H, 4.63; N, 5.60; S, 6.41%. Found: C, 52.67; H, 4.60; N, 5.64; S, 6.44%. IR (KBr, cm⁻¹):1596 $v_{(C=N-N=C)}$, 1259 $v_{(C-O)}$. UV–Vis (CH₃CN, λ max/nm; ε/dm³ mol⁻¹ cm⁻¹): 364(5516), 280(6007), 234(6622). ¹H NMR (400 MHz, CDCl₃) (δ ppm): 8.8 (s, 1H, N=CH), 7.1–8.0 (m, 8H, aromatic), 5.3 (d, *J* = 5.6 Hz, 1H, cymene Ar-H), 5.3 (d, *J* = 6 Hz, 1H, cymene Ar-H), 5.0 (d, *J* = 5.6 Hz, 1H, cymene Ar-H), 4.6 (d, *J* = 5.6 Hz, 1H, cymene Ar-H), 2.5 (m, 1H, CH of *p*-cymene), 2.2 (s, 3H, CH₃ of *p*-cymene), 1.0-

345 1.3 (dd, J = 94.8 Hz, J = 7.2 Hz, 6H, 2CH₃ of *p*-cymene). ESI-MS (CH₃CN): calcd for C₂₂ 346 H₂₃ Cl N₂ O Ru S *m*/*z* 500.02; found [M + H]⁺:501.03, [M - Cl]⁺:465.05.

347

4.4.6 [**Ru**(η^6 -*p*-cymene)(**Cl**)(**L2**)] (6). Colour: Yellow; Yield: 82%; M.p.: 180 °C; Anal. 348 Calc. For C₂₂ H₂₂ Cl₂ N₂ O Ru S: C, 49.43; H, 4.14; N, 5.24; S, 5.99%. Found: C, 49.28; H, 349 4.15; N, 5.22; S, 5.98%. IR (KBr, cm⁻¹):1599 $v_{(C=N-N=C)}$, 1262 $v_{(C-O)}$. UV–Vis (CH₃CN, λ 350 max/nm; ɛ/dm³ mol⁻¹ cm⁻¹): 364(3579), 282(3771), 245(5264).¹H NMR (400 MHz, CDCl₃) 351 (δ ppm): 8.8 (s, 1H, N=CH), 7.1–8.0 (m, 7H, aromatic), 5.3 (d, J = 6.4 Hz, 1H, cymene Ar-352 H), 5.3 (d, J = 6 Hz, 1H, cymene Ar-H), 5.0 (d, J = 5.6 Hz, 1H, cymene Ar-H), 4.6 353 5.6 Hz, 1H, cymene Ar-H), 2.5 (m, 1H, CH of *p*-cymene), 2.2 (s, 3H, CH₃ of *p*-cymene), 1.0-354 1.3 (dd, J = 100.8 Hz, J = 14.4 Hz, 6H, 2CH₃ of *p*-cymene). ESI-MS (CH₃CN): calcd for C₂₂ 355 $H_{22} Cl_2 N_2 O Ru S m/z 533.98$; found = $[M - Cl]^+$:499.02. 356

357

4.4.7 [**Ru**(η⁶-*p*-cymene)(**Cl**)(**L3**)] (7). Colour: Yellow; Yield: 78%; M.p.: 178 ⁰C; Anal. 358 Calc. For C₂₂ H₂₂ Br Cl N₂ O Ru S: C, 45.64; H, 3.83; N, 4.83; S, 5.53%. Found: C, 45.43; H, 359 3.85; N, 4.81; S, 5.54%. IR (KBr, cm⁻¹):1607 $v_{(C=N-N=C)}$, 1260 $v_{(C=O)}$. UV–Vis (CH₃CN, λ 360 max/nm; $\epsilon/dm^3 mol^{-1} cm^{-1}$): 360(6761), 304(7395), 246(10158). ¹H NMR (400 MHz, CDCl₃) 361 362 (δ ppm): 8.8 (s, 1H, N=CH), 7.1–8.0 (m, 7H, aromatic), 5.3 (d, J = 6 Hz, 1H, cymene Ar-H), 5.3 (d, J = 6 Hz, 1H, cymene Ar-H), 5.0 (d, J = 5.6 Hz, 1H, cymene Ar-H), 4.6 (d, J = 5.6363 Hz, 1H, cymene Ar-H), 2.5 (m, 1H, CH of *p*-cymene), 2.2 (s, 3H, CH₃ of *p*-cymene), 1.0-1.3 364 (dd, J = 104 Hz, J = 7.2 Hz, 6H, 2CH₃ of *p*-cymene). ESI-MS (CH₃CN): calcd for C₂₂ H₂₂ Br 365 Cl N₂ O Ru S *m*/*z* 577.93; found [M - Cl]⁺:544.96. 366

367

4.4.8 [**Ru**(η^6 -*p*-cymene)(**Cl**)(**L4**)] (8). Colour: Yellow; Yield: 76%; M.p.: 168 ^oC; Anal. 368 Calc. For C₂₃ H₂₅ Cl N₂ O₂ Ru S: C, 52.11; H, 4.75; N, 5.28; S, 6.04%. Found: C, 52.35; H, 369 4.70; N, 5.25; S, 6.08%. IR (KBr, cm⁻¹):1592 $v_{(C=N-N=C)}$, 1259 $v_{(C-O)}$. UV–Vis (CH₃CN, λ 370 max/nm; ɛ/dm³ mol⁻¹ cm⁻¹): 361(4930), 291(5508), 246(7594). ¹H NMR (400 MHz, CDCl₃) 371 (δ ppm): 8.8 (s, 1H, N=CH), 6.7–7.9 (m, 9H, aromatic), 5.3 (d, J = 6 Hz, 1H, cymene Ar-H), 372 5.3 (d, J = 6 Hz, 1H, cymene Ar-H), 5.0 (d, J = 5.6 Hz, 1H, cymene Ar-H), 4.6 (d, J = 5.6373 Hz, 1H, cymene Ar-H), 3.7 (s, 3H, OCH₃), 2.5 (m, 1H, CH of *p*-cymene), 2.2 (s, 3H, CH₃ of 374 *p*-cymene), 1.0-1.3 (dd, J = 101.6 Hz, J = 7.2 Hz, 6H, 2CH₃ of *p*-cymene). ESI-MS 375 (CH₃CN): calcd for C₂₃ H₂₅ Cl N₂ O₂ Ru S m/z 530.04; found $[M + H]^+$:531.04, $[M - Cl]^+$ 376 377 :495.06.

378	4.5 X-ray crystallography
379	A Single crystal of [Ru(η^6 - <i>p</i> -cymene)Cl(L2)] (6) and [Ru(η^6 - <i>p</i> -cymene)Cl(L3)] (7)
380	were obtained Dichloromethane-Petroleum ether solution at room temperature by slow
381	evaporation technique. X-Ray data were collected with a Bruker AXS Kappa APEX II single
382	crystal X-ray diffractometer using monochromated Mo-K α radiation (λ =0.71073). The
383	structure solution was obtained by direct methods (SIR-97) [35] and refined using (SHELXL-
384	97) full matrix least-squares calculations on F^2 [36]. All non-hydrogen atoms were refined
385	anisotropically, hydrogen atoms were fixed geometrically and refined by riding model. The
386	Bruker SAINT-Plus (Version 7.06a) software were used to analyse the Frame integration and
387	data reduction. The multiscan absorption corrections were applied using SADABS software.
388	CCDC reference number is 1449681-1449682.
389	
390	4.6 Stability Studies
391	The stability of the complexes were carried out as described previously [37].
392	
393	4.7 Partition Coefficient Determination
394	Partition coefficients (P) between n-octanol and water phases were carried out as
395	described previously [27,38].

396

397 **4.8 Cell culture**

HeLa human cervical cancer cell line, MDA-MB-231 Triple negative breast 398 399 carcinoma, Hep G2 human liver carcinoma cell line and NIH 3T3 noncancerous cell, mouse embryonic fibroblast were supplied by the National Centre for Cell Science 400 (NCCS), Pune. The cell lines were cultured as a monolayer in RPMI-1640 medium 401 (Biochrom AG, Berlin, Germany), supplemented with 10% fetal bovine serum (Sigma-402 Aldrich, St. Louis, MO, USA) and with 100 U mL⁻¹ penicillin and 100 µg mL⁻¹ streptomycin 403 as antibiotics (Himedia, Mumbai, India), at 37 °C in a humidified atmosphere of 5% CO₂ in a 404 CO₂ incubator (Heraeus, Hanau, Germany). 405

- 406 MTT assay, AO-EB staining, Hoechst 33258 staining, Flow cytometry and 407 comet assay were evaluated as described previously [39-42].
- 408

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414

415 Appendix A. Supplementary material

¹H spectra of the complexes (1-8), The ESI-MS of the complexes (1-8) and log P
Values for Complexes 1-8. CCDC 1449681-1449682 contains the supplementary
crystallographic data for this paper. These data can be obtained free of charge from The
Cambridge Crystallographic Data Centre via <u>www.ccdc.cam.ac.uk/data_request/cif</u>.

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- Synthesis and characterization of arene ruthenium(II) benzhydrazone complexes.
- > The single-crystal X-ray structure analysis of two complexes is depicted.
- > The complexes have been screened for their *in vitro* antiproliferative activities.
- \blacktriangleright The mechanism of action of the most potent complexes was evaluated.