Polycyclitols — Novel conduritol and carbasugar hybrids as new glycosidase inhibitors¹

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Abstract: A family of novel carbasugar analogues (bicyclitols) based on *cis*-hydrindane and *cis*-decalin frameworks has been conceptualized. These novel entities can be regarded as conduritol and carbasugar hybrids. Syntheses of these polyhydroxylated entities have been achieved in stereo- and regioselective manners, starting from the readily available Diels–Alder adducts of 5,5-dimethoxy-1,2,3,4-tetrachlorocyclopentadiene and appropriate dienophiles like cyclopentadiene or *p*-benzoquinone, that embody a masked 7-ketonorbornenone moiety. Thermally induced chelotropic elimination of CO from the appropriately functionalized 7-ketonorbornenone derivatives to deliver annulated bicyclic 1,3-cyclohexadiene derivatives was the key step in this synthetic endeavor. Further oxy-functionalization of the 1,3-cyclohexadiene moiety delivered the targeted polycyclitols. A preliminary investigation of the glycosidase inhibitory potency of these bicyclitols, identified compounds **18** and **54** as potent and selective inhibitors of α -glucosidase (yeast).

Key words: carbasugar, conduritol, glycomimics, glycosidase inhibitors.

Résumé : On a conceptualisé une nouvelle famille d'analogues de carbasucres (bicyclitols) reposant sur des squelettes de *cis*-hydrindane et de *cis*-décaline. Ces nouvelles entités peuvent être considérées comme des hybrides du conduritol et des carbasucres. Des synthèses de ces entités polyhydroxylées ont été réalisées de façons stéréo- et régiosélectives à partir d'adduits de Diels–Alder facilement accessibles du 5,5-diméthoxy-1,2,3,4-tétrachlorocyclopentadiène et de diéno-philes appropriés, tels le cyclopentadiène ou la *p*-benzoquinone, qui comportent une portion 7-cétonorbornénone marquée. L'étape clé de ces synthèses implique une élimination chélotropique thermiquement induite de CO à partir des dérivés fonctionnalisés d'une façon appropriée de la 7-cétonorbornénone qui permet d'obtenir les dérivés cyclohexa-1,3-diènes bicycliques annelés. Une oxy-fonctionnalisation subséquente de la portion cyclohexa-1,3-diène conduit aux polycyclitols recherchés. Une étude préliminaire du pouvoir inhibiteur de la glycosidase de ces bicyclitols a permis d'identifier les composés **18** et **54** comme actifs et comme inhibiteurs sélectifs de l' α -glucosidase (levure).

Mots clés : carbasucre, conduritol, glycomimique, inhibiteurs de la glycosidas.

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Introduction

Glycosidases and related enzymes are crucial in many biological processes that are vital for the normal functioning of cells. Hence, inhibitors of these carbohydrate-processing enzymes have received increased attention in the past few decades from a synthetic as well as a biological activity perspective. These pursuits have been largely fuelled by the realization of their potential as therapeutic agents for the treatment of a variety of carbohydrate-mediated diseases like diabetes, cancer, AIDS, and viral infections (1, 2). The basic premise for the design of glycosidase inhibitors has been to generate transition state mimics by replacing the endocyclic

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oxygen atom of a monosaccahride by a carbon (carbasugar, 1) or nitrogen atom (azasugar, 2). The lack of a glycosidic linkage in these entities, which otherwise resemble the monosaccharides in shape, size, and functionalization, renders them hydrolytically stable. In this context, a range of carbasugars, azasugars, and their analogues, with diverse functionalization and stereochemical features have been devised, and their glycosidase inhibitory activities evaluated (1, 2). Additionally, conduritols 3 (1,2,3,4-tetrahydroxy-5-cyclohexenes) and their various derivatives have also attracted attention because of their glycosidase inhibitory properties (3). Several bicyclic and tricyclic conduritol variants like 4 and 5 have also been synthesized and evaluated for their efficacy against various glycosidases (4).

Naturally occurring alkaloids like 1-deoxynojirimycin (6) can be regarded as simple azasugar analogues of D-glucose and have been found to be potent inhibitors of α - and β -



glucosidases. Besides nojirimycins, several other mono- and bicyclic alkaloids based on polyhydroxylated pyrrolidine, piperidine, pyrrolizidine, indolizidine, and quinolizidine frameworks have also been identified as carbohydrate mimics. Typical examples include bicyclic pyrrolizidine alkaloid alexine 7, indolizidine alkaloid castanospermine 8, and quinolizidine alkaloid 9 (5).



Spurred by the heightened interest in the design of potent inhibitors of glycosidases and inspired by the structures of bicyclic alkaloids 7-9, we have conceived of a new family of polyhydroxylated diquinanes 10, hydrindanes 11, and decalins 12, collectively termed as "polycyclitols", as potential glycomimics (6, 7). Among these bicyclitols, 11 and 12 can be considered as ring-annulated carbasugar entities and more interestingly, the decalin polyol 12 can be regarded as a hydrid of two carbasugars A and B (see bold portions in 13a and 13b). The presence of the additional hydroxyl groups on the annulus, it is presumed, may provide additional binding points for these analogues at the receptor site. The effect of annulation on the biological profile of biomolecules can be profound, as seen in the case of the natural glycosidase inhibitors, deoxynojirimycin 6 and castanospermine 8. The former, a nitrogen analogue of glucose, inhibits yeast α -glucosidase, but is inactive against almond β glucosidase. On the other hand 8, a ring-annulated analogue of 6, is a potent inhibitor of β -glucosidase, but exhibited no inhibition against yeast α -glucosidase (5). The bicyclitols 11 and 12 conceptualized here have a similar structural relationship to carbasugars as castanospermine 8 has to deoxynojirimycin 6. Indeed, these bicyclitols can be visualized as the carbon analogues of castanospermine 8 and other bicyclic alkaloids exhibiting glycosidase activity. Alternatively, these novel bicyclic entities can be regarded as annulated conduritol, with 12 being a hybrid of two conduritols with shared, common ring junction carbon atoms.



Herein, we report the stereo- and regioselective synthesis of the newly conceptualized polyhydroxylated carbocyclic entities, with as many as nine (in 11) and 10 (in 12) stereogenic centers, starting from readily available Diels–

Scheme 1. Retrosynthetic analysis of 11 and 12.



Alder adducts, following fairly general and flexible synthetic sequences. We also report on the glycosidase inhibitory activity of the newly synthesized polycyclitols, two of which have exhibited an impressive and selective inhibition of yeast α -glucosidase.

Results and discussions

Retrosynthetic analysis

A general retrosynthetic analysis for accessing hydrindane **11** and decalin **12** based bicyclitols is depicted in Scheme 1. It was visualized that the hydroxyl group pattern on the sixmembered ring could be generated through oxyfunctionalization of diene **14** (n = 1, 2), which in turn could be obtained by the thermal or photochemical decarbonylation of the 7-ketonorbornyl moiety in **15**. The precursor tricyclic ketones **15** (n = 1, 2) are readily obtainable via Diels–Alder cycloaddition between 5,5-dimethoxy-1,2,3,4-tetrachlorocyclopentadiene and an appropriate cyclic dienophile (Scheme 1).

Thus, tricyclic precursors 16 and 17 were obtained via a Diels-Alder reaction between 5,5-dimethoxy-1,2,3,4-tetrachlorocyclopentadiene and cyclopentadiene or *p*-benzoquinone, respectively. While the tricyclic endo-adduct 16 was to serve as the starting point for the hydrindane based polyols 11 (see bold portion), the benzoquinone derived Diels-Alder endo-adduct 17 was chosen as the starting material for the decalin based polyols 12 (bold portion) (Scheme 2). The main advantage in the choice of endoadducts 16 and 17 as our basic launching pad was that their tricyclic framework built on a norbornyl matrix confers on them a topological bias towards exo-selectivity. This built-in facial bias in 16 and 17 was, therefore, expected to ensure stereoselective operations, particularly oxyfunctionalization maneuvers. The foregoing retrosynthetic analysis provided Scheme 2.



the road map for the synthesis of the targeted new family of bicyclitols.

Synthesis of hydrindane based bicyclitols

The synthesis of the first galacto-configured *cis*-hydrindane-based polyol **18**, emanated from the tricyclic Diels–Alder adduct **16**, obtained by the [4 + 2] cycloaddition of 5,5,-dimethoxy-1,2,3,4-tetrachlorocyclopentadiene and cyclopentadiene (Scheme 3) (8). Adduct **16** was subjected to allylic oxidation, using in-situ-generated Mn(OAc)₃, to deliver the exo-acetate derivative **19** (Scheme 3) (9). The exo-acetate **19** was subjected to dihydroxylation following the Upjohn catalytic OsO₄–NMMO protocol (10) to generate diol **20**, which on reductive dehalogenation (**21**) followed by acetylation furnished the C_s -symmetric triacetate **22** (Scheme 3) (8*a*). All three acetate functionalities in **22** were cis disposed, owing to the exo-propensity of the topology of the endo-fused system **19**.

The tricyclic 7-ketal derivative 22 upon exposure to Amberlyst-15 in moist acetone, resulted in ketal deprotection and furnished the 7-ketonorbornenone derivative 23 (Scheme 3). As contemplated earlier, decarbonylation in 23 through thermal activation by heating in nitrobenzene (~160 °C) furnished the C_s -symmetric cyclohexa-1,3-diene derivative 24 in moderate yield (Scheme 3) (11). The cyclic 1,3-diene moiety in 24 was subjected to exhaustive dihydroxylation employing the catalytic OsO₄-NMMO protocol (10) to furnish a single tetrahydroxy-triacetate 25, which was devoid of any symmetry. Loss of symmetry during the double dihydroxylation reaction on the diene secured the stereochemistry of 25, which required the two hydroxylations on diene 24 to occur from the exo- and endo-faces, respectively, of the cis-hydrindane (Scheme 3). Mild base hydrolysis of triacetate 25 gave the targeted heptahydroxy bicyclitol 18, an annulated 5a-carbagalactose derivative.

The synthesis of diastereomeric *cis*-hydrindane-based carbasugar analogue **26** started from the readily available endotricyclic enone **27**. Enone **27** was obtained following the protocol reported by Schuda et al. (12) involving [4 + 2]dimerization of the ethylene ketal of 2,4-cyclopentadienone and chemoselective deketalization. Enone **27** was converted to endo-allylic alcohol **28** via DIBAL-H reduction from the exo-face and the norbornene double bond was protected Scheme 3. Reagents and conditions: (*a*) toluene, reflux, 24 h, 70%; (*b*) Mn(OAc)₃, AcOH, 70 °C, 1 h, 64%; (*c*) OsO₄ (1 mol%), NMMO, Me₂CO–H₂O–*t*-BuOH (5:5:2), rt, 12 h, 80%; (*d*) Na, liq. NH₃, THF, EtOH, -33 °C, 30 min, 72%; (*e*) Ac₂O, pyridine, DMAP, CH₂Cl₂, rt, 2 h, 88%; (*f*) Amberlyst-15, acetone, rt, 1 h, 89%; (*g*) C₆H₅NO₂, heat, 160 °C, 2 h, 50%; (*h*) OsO₄ (1 mol%), NMMO, Me₂CO–H₂O–*t*-BuOH (5:5:2), rt, 48 h, 88%; (*i*) aq. NaOH, rt, 1 h, 60%.



through intramolecular bromoetherification to give 29 (Scheme 4). The cyclopentene double bond in 29 was stereoselectively dihydroxylated and the resulting *cis*-diol 30 was protected as acetonide 31. The norbornene double bond

Scheme 4. Reagents and conditions: (*a*) DIBAL-H (1 mol/L in hexane), CH_2Cl_2 , -78 °C, 15 min, 90%; (*b*) NBS, CH_2Cl_2 , rt, 3 h, 80%; (*c*) OsO₄ (1 mol%), NMMO, Me₂CO-H₂O-*t*-BuOH, (5:5:2), rt, 12 h, 82%; (*d*) Amberlyst-15, acetone, 4 Å molecular sieves, rt, 3 h, 95%; (*e*) Zn dust, AcOH, MeOH, rt, 4 h, 85%; (*f*) Amberlyst-15, acetone, reflux, 3 h, 81%; (*g*) C₆H₅NO₂, 160 °C, 4 h, 67%; (*h*) OsO₄ (1 mol%), NMMO, Me₂CO-H₂O-*t*-BuOH (5:5:2), rt, 48 h, 83%; (*i*) 30% CF₃COOH, rt, 30 min, 89%.



was now unmasked with Zn metal to render **32** (Scheme 4) (8*a*). The ketal moiety in **32** was carefully deprotected in the presence of Amberlyst-15 to furnish the crystalline 7-norbornenone derivative **33** (Scheme 4). Tricyclic ketone **33**, on heating in nitrobenzene (160 °C), quite readily converted to the 1,3-cyclohexadiene derivative **34** through extrusion of CO (11). At this point, it was recognized that diene **34** was well suited for generating stereochemically diverse oxygenation patterns through appropriate sequencing of the oxyfunctionalization protocol.

Towards this end, the *cis*-hydrindane diene **34** was subjected to exhaustive dihydroxylation under catalytic OsO_4 conditions to furnish the pentahydroxy compound **35** (Scheme 4). The stereochemical assignment for **35** followed from an incisive analysis of the spectral data in analogy with that of compound **25**. In addition, it is expected that the directing influence of the free α -hydroxyl group at the peri position in diene **34** would lead to the preferred stereostructure of **35**. Mild acid hydrolysis of the acetonide protecting group in **35** furnished the targeted heptahydroxylated hydrindane **26**, which was an epimer of **18** at C1. Polyol **26**, like **18**, embodies a 5a-carbagalactose moiety, with the hydroxylated carbocyclic annulation spanning the C_{5a} - C_6 atoms of the carbasugar (Scheme 4).

In another stereochemical variation in the hydroxylation pattern on the hydrindane framework, [4 + 2] cycloaddition of singlet oxygen $({}^{1}O_{2})$ was effected on 34 to deliver endoperoxide 36 as the sole product in good yield with the addition of ${}^{1}O_{2}$ to 34 from the preferred convex face of the molecule (Scheme 5) (13). The peroxide linkage in 36 was cleanly cleaved with lithium aluminum hydride to yield the 1,4-diol derivative 37 and further dihydroxylated under the standard catalytic OsO₄ conditions to give a 3:1 mixture of tetrols 38 and 39, respectively. All attempts to separate the mixture of tetrols at this stage were unsuccessful. At this point, we reasoned that one of the stereoisomers from the dihydroxylation should have the hydroxyl groups in an all cis orientation while the other diastereomer would have a trans-syn-trans arrangement of the hydroxyl groups on the six-membered ring. Hence, it was reasoned that the protection of the hydroxyl groups in the mixture of 38 and 39 as the corresponding acetonides, would result in the formation of a tris-acetonide derivative in the case of 38 and a bisacetonide in the case of 39 and would thus be rendered amenable to separation. Consequently, the mixture of 38 and 39 was treated with Amberlyst-15 in acetone medium to furnish, as anticipated, tris-acetonide 40 and bis-acetonide 41, respectively, which were then readily separable by column chromatography (Scheme 5). The major isomer 40 originates from dihydroxylation of 37 from the syn-face of allylic alcohols, an outcome that is in conflict with Kishi's rule (14) for dihydroxylation of allylic alcohols. The stereochemical outcome in this case can be attributed to the overwhelming topological preference of the cis-hydrindane moiety in 37 to react from the less-hindered, convex face to yield syndihydroxylated 38 as the major product. The acetonide deprotection in 40 and 41 in the presence of 30% trifluoroacetic acid cleanly furnished the diastereomeric heptahydroxylated hydrindanes 42 and 43, respectively (Scheme 5). The bicyclic carbasugar analogues 42 and 43 correspond to the

Scheme 5. Reagents and conditions: (*a*) O₂, hv, methylene blue, CHCl₃, 20 °C, 8 h, 90%; (*b*) LiAlH₄, THF, rt, 1 h, 54%; (*c*) OsO₄ (1 mol%), NMMO, Me₂CO–H₂O–*t*-BuOH (5:5:2), rt, 1 h; (*d*) Amberlyst-15, acetone, 4 Å molecular sieves, followed by silica gel separation (62% for 40 and 21% for 41); (*e*) 30% CF₃COOH, 92% for 42, and 88% for 43.



 α -allopyranose and α -mannopyranose configurations, respectively.

Synthesis of *cis*-decalin-based bicyclitols

The synthetic venture towards *cis*-decalin-based carbasugar analogues of type **12** emanated from the readily available Diels–Alder adduct **17** of 5,5-dimethoxy-1,2,3,4tetrachlorocyclopentadiene and *p*-benzoquinone (Scheme 6) (15). Tricyclic quinone **17** was elaborated to the symmetrical tricyclic diene **44** essentially following a literature procedure (16). Adduct **17** was subjected to Luche reduction (NaBH₄-CeCl₃·7H₂O) (17) to give endo,endo-diol **45** and was further Scheme 6. Reagents and conditions: (*a*) toluene, reflux, 24 h, 83%; (*b*) NaBH₄, CeCl₃·7H₂O, MeOH, 0 °C, 10 min, 69%; (*c*) MsCl, pyridine, DMAP, CH₂Cl₂, rt, 24 h, 72%; (*d*) NaI, C₂H₅COCH₃, reflux, 8 h, 80%; (*e*) OsO₄ (1 mol%), NMMO, Me₂CO-H₂O-*t*-BuOH (5:5:2), rt, 48 h, 66%; (*f*) Amberlyst-15, acetone, 4 Å molecular sieves, rt, 3 h, 75%; (*g*) Na, liq. NH₃, THF, EtOH, -33 °C, 30 min, 49%; (*h*) Amberlyst-15, acetone, rt, 2 h, 98%; (*i*) C₆H₅NO₂, 160 °C, 4 h, 63%; (*j*) OsO₄ (1 mol%), NMMO, Me₂CO-H₂O-*t*-BuOH (5:5:2), rt, 48 h, 85%; (*k*) 30% CF₃COOH, rt, 30 min, 95%.



converted to dimesylate 46, and NaI-mediated 1,4elimination (via 47) in 46 led to the desired C_s -symmetric tricyclic diene 44 (Scheme 6). Exhaustive catalytic-OsO4mediated dihydroxylation on tricyclic diene 44 furnished tetrol **48** (Scheme 6). The symmetrical nature of the ¹H and ¹³C NMR spectra of **48** indicated an all cis relationship of the hydroxyl groups, which implied that the double dihydroxylation occurred from the exo-face of the tricyclic diene. Acetonide protection of the vicinal hydroxyl groups in tetrol 48 resulted in selective formation of the symmetrical monoacetonide 49 (Scheme 6). Acetonide 49 was subjected to reductive dechlorination in Na - liquid ammonia milieu to give the symmetrical olefin 50 in a modest yield (Scheme 6). The ketal deprotection in 50 proceeded quite uneventfully to furnish the crystalline 7-norbornenone derivative 51 in quantitative yield (Scheme 6). As contemplated earlier, thermally induced decarbonylation in tricyclic ketone 51 led to the C_s -symmetric bicyclic cyclohexadiene derivative 52(11).

Bicyclic diene 52 was subjected to exhaustive dihydroxylation employing catalytic OsO_4 to stereoselectively furnish the hexahydroxylated decalin 53 as a single diastereomer (Scheme 6). The stereochemical assignments for 53 were done on the basis of loss of symmetry (NMR) during dihydroxylation and in analogy with earlier observations for compound 25 (see Scheme 3). The acetonide protection in 53 was relieved by exposure to 30% trifluoroacetic acid to deliver the targeted octahydroxylated decalin based carbasugar analogue 54. Bicyclitol 54 can be regarded as embodying an α -carbagalactopyranose configuration on a *cis*-decalin framework (Scheme 6).

To generate stereochemical variations of hydroxyl groups on the decalin framework, tricyclic endo,endo-diol 45 was dihydroxylated stereoselectively to furnish tetrol 55 (Scheme 7). Dihydroxylation in 55 occurred exclusively from the convex face because of the exo-propensity of the tricyclic system built on a norbornyl scaffold, and the structure of 55 was secured on the basis of symmetry (C_s) revealed by its ¹H and ¹³C NMR spectra. The cis-diol moiety in 55 was protected using Amberlyst-15 in acetone medium to furnish acetonide 56. Reductive dechlorination in 56 under Birch reduction conditions delivered the C_s -symmetric olefin 57 (Scheme 7). The ketal moiety in 57 was next deprotected to unmask the carbonyl group and furnish norbornenone derivative 58 (Scheme 7). On thermal activation, tricyclic ketone 58 underwent smooth decarbonylation to furnish the C_s -symmetric bicyclic cyclohexadiene derivative 59 (Scheme 7) (11). Bicyclic diene 59 was diastereomeric with 52 (Scheme 6), and hence the protocol adopted earlier for the elaboration of 52 could be extended to diene 59 for the realization of the targeted octahydroxy polyol.

The 1,3-cyclohexadiene moiety in **59** was double dihydroxylated (OsO₄–NMMO) to furnish the hexahydroxylated *cis*-decalin derivative **60**, which was devoid of any symmetry as indicated by its NMR (¹H and ¹³C) spectral analysis. Hence, as observed in the case of **25** and **53** (vide supra), the two sequential dihydroxylations on the diene moiety had occurred from opposite faces, thereby generating a cis–trans–cis arrangement of the hydroxyl groups in **60** (Scheme 7). The acetonide deprotection in **60** delivered octahydroxylated decalin **61**, a diastereometic sibling of **54**.

Scheme 7. Reagents and conditions: (a) OsO₄ (1 mol%), NMMO, Me₂CO-H₂O-t-BuOH (5:5:2), rt, 1.5 h, 75%;
(b) Amberlyst-15, acetone, 4 Å molecular sieves, rt, 1 h, 82%;
(c) Na, liq. NH₃, THF, EtOH, -33 °C, 40 min, 88%;
(d) Amberlyst-15, acetone, rt, 2 h, 95%; (e) C₆H₅NO₂, 160 °C, 3 h, 43%; (f) OsO₄ (1 mol%), NMMO, Me₂CO-H₂O-t-BuOH (5:5:2), rt, 48 h, 73%; (g) 30% CF₃COOH, rt, 30 min, 90%.



With the availability of the designer polycyclitols 18, 26, 42, 43, 54, and 61, our attention was turned towards the evaluation of their efficacy as glycosidase inhibitors.

Glycosidase inhibitory potencies of bicyclitols

Evaluation of the glycosidase inhibitory activity of the newly synthesized bicyclitols **18**, **26**, **42**, **43**, **54**, and **61** was carried out against various commercially available glycosidases, α -glucosidase (yeast), β -glucosidase (sweet almond), α -galactosidase (green coffee beans), β -galactosidase (*Escherichia coli*), α -mannosidase (jack bean) and β -mannosidase (snail acetone powder), using their corresponding *ortho-* or *para*-nitrophenylglycosides at the optimum pH and temperature for each enzyme (see Experimental) (18). All assays

were done up to 1 mmol/L concentration of the polyols. Gratifyingly, the decalin-based carbasugar analogue 54 exhibited an impressive inhibition of yeast α -glucosidase, with an inhibition constant $K_i = 12 \,\mu \text{mol/L}$ (cf. $K_i = 25 \,\mu \text{mol/L}$ for deoxynojirimycin). However, the same compound 54 did not show any significant inhibitory activity against almond β-glucosidase up to 1 mmol/L concentration, thus highlighting the selectivity of 54 towards α -glucosidase. To our further delight, the hydrindane-based polyol 18 exhibited a moderate and selective inhibition of yeast α -glucosidase $(K_i = 84 \,\mu \text{mol/L})$ but, like 54, was totally inert to the almond β -glucosidase. Both these compounds failed to elicit any inhibition of the galactosidases and mannosidases. The other polyols 26, 42, 43, and 61 failed to exhibit any significant (<1 mmol/L) inhibitory activity against the glycosidases examined. The lack of inhibitory activity of polyols 26, 42, 43, and 61 when compared with their diastereomeric siblings 18 and 54, which inhibited α -glucosidase, brings forth the effect of subtle variations in the stereochemical dispositions of the hydroxyl groups on their inhibitory potency against the carbohydrate processing enzymes. At present, the precise reasons for the selective affinity of 18 and 54 against α glucosidase and the effect of subtle stereochemical variations on their inhibitory activity is not clear and remains a matter that still tickles our curiosity. Further investigations will hopefully illuminate this puzzling observation.

Conclusion

We have delineated a relatively short and fairly general synthetic access to novel bicyclic carbasugar analogues based on *cis*-decalin and *cis*-hydridane frameworks with as many as eight to seven hydroxyl groups, respectively. The synthetic approaches leading to secured stereochemistry at all the stereogenic centers (nine in **11** and 10 in **12**) in the bicyclitols are notable for their simplicity and stereoselectivity. The decalin-based polyol **54** and hydrindane bicyclitol **18** exhibited significant and selective inhibition of yeast α -glucosidase.

Experimental section

General

Melting points were recorded on a Büchi B-540 apparatus and are uncorrected. IR spectra were recorded on JASCO FT-IR 410 spectrometer. ¹H NMR spectra were recorded on a JEOL JNM-LA 300 or a Bruker AMX 400 or a Bruker DRX 500 instrument in CDCl₃ solutions, unless otherwise stated. Chemical shifts are reported with respect to tetramethylsilane (Me₄Si) as the internal standard (for ¹H NMR) and the central line (77.0 ppm) of CDCl₃ (for ^{13}C NMR). The chemical shifts are expressed in parts per million (δ) downfield from Me₄Si. The standard abbreviations s, d, t, q, and m refer to singlet, doublet, triplet, quartet, and multiplet, respectively. Coupling constants (J), whenever discernible, have been reported in Hz. Low resolution mass spectra (LR-MS) were recorded either on a Shimadzu GC-MS-QP 5050A spectrometer (EI or CI mode) or on a Q-TOF Micromass mass spectrometer. High resolution mass spectra (HR-MS) were recorded on a Q-TOF Micromass mass spectrometer. Elemental analyses were obtained on a Carlo Erba 1106 CHN analyzer. Silica gel from Acme (100–200 mesh particle size) was used for column chromatography. All moisture- and air-sensitive reactions were performed under an argon atmosphere with dry, freshly distilled solvents under anhydrous conditions using standard syringe–septum techniques. Yields reported are of isolated materials judged homogeneous by thin layer chromatography (TLC) and NMR. All solvent extracts were washed with water/brine as appropriate and dried over anhydr. Na₂SO₄ before removal of the solvent on a rotary evaporator under reduced pressure. The glycosidase enzymes and their corresponding substrates (*ortho-* or *para*-nitrophenolates) were purchased from the Sigma Chemical Company, USA.

1,7,8,9-Tetrachloro-10,10-dimethoxytricyclo[5.2.1.0^{2,6}]deca-4,8-dien-3-ylacetate (19)

Cyclopentadiene (12 g, 0.18 mol) was added to a solution of 5,5-dimethoxy-1,2,3,4-tetrachlorocyclopentadiene (50 g, 0.18 mol) in dry toluene (80 mL) and the resulting solution was refluxed for 24 h. Excess solvent was distilled off from the reaction mixture and the residue obtained was distilled under reduced pressure (130-140 °C/5 torr, 1 torr =133.3224 Pa) to yield adduct **16** (41.6 g, 70%). To a solution of the Mn(OAc)₂·H₂O (10.8 g, 44 mmol) in AcOH (40 mL) was added Ac₂O (14 mL) and the mixture was refluxed for 20 min. Solid KMnO₄ (1.74 g, 10.1 mmol) was carefully added and the resulting mixture was refluxed for an additional 30 min. After cooling to 70 °C, adduct 16 (12 g, 36.4 mmol) and KBr (720 mg, 6 mmol) were added and the reaction mixture was stirred at the same temperature until complete consumption of the starting material. The reaction mixture was cooled to room temperature and filtered through a Celite pad. The filtrate was extracted with hexane $(3 \times$ 100 mL) and the residue obtained after the usual work-up, was chromatographically purified on a silica gel column (5% ethyl acetate - hexane) to afford the exo-acetate 19 (9 g, 64%), mp 111 to 112 °C (lit. value (8a) mp 110 °C). IR (Nujol, cm^{-1}): 1740.

10,10-Dimethoxy-3,5-di(acetyloxy)tricyclo[5.2.1.0^{2,6}]dec-8-en-4-yl acetate (22)

To the solution of exo-acetate 19 (4 g, 10.3 mmol) in acetone-water-t-BuOH (5:5:1, 25 mL), OsO4 (26 mg, 1 mol%), and 50% aq. NMMO solution (2.65 mL, 11.3 mmol) were added sequentially. The pale yellow reaction mixture was stirred at room temperature for 12 h prior to quenching of the reaction by addition of solid NaHSO₃ (1 g). The resulting mixture was filtered through a Celite pad and the filtrate was concentrated under reduced pressure to give a residue, which on chromatographic purification on a silica gel column furnished diol 20 (3.5 g, 80%). The solution of diol 20 (2.2 g, 5.2 mmol) in dry THF (25 mL) and absolute EtOH (10 mL) was added to freshly distilled liquid ammonia (250 mL) and to this solution, freshly cut pieces of sodium (1 g) were carefully added, until the blue color of the solution persisted. The reaction mixture was stirred for 15 min, prior to quenching the reaction with solid NH₄Cl (1 g). Ammonia was allowed to evaporate overnight, the residue obtained was diluted with ice-cooled water (100 mL) and extracted with ethyl acetate $(3 \times 100 \text{ mL})$. The usual workup and evaporation of volatiles under reduced pressure furnished a residue (21) that was dissolved in dry pyridine (10 mL), then Ac₂O (1 mL) and catalytic DMAP (100 mg) were added. The reaction mixture was stirred at room temperature for 2 h before diluting with water. The reaction mixture was extracted with CH_2Cl_2 (3 × 60 mL) and after the usual work-up was purified by silica gel column chromatography (20% ethyl acetate – hexane) to furnish triacetate 22 (1.2 g, 64%), mp 140–142 °C. IR (Nujol, cm⁻¹): 1725.

3,4,5-Tri(acetyloxy)tricyclo[5.2.1.0^{2,6}]dec-8-en-10-one (23)

A solution of ketal **22** (200 mg, 0.543 mmol) in moist acetone (8 mL) was stirred with Amberlyst-15 (~30 mg) at room temperature for 1 h. The resin was filtered through a cotton plug and the filtrate was concentrated to give a residue, which was subjected to chromatography on a silica gel column (15% ethyl acetate – hexane) to furnish ketone **23** (155 mg, 89%). IR (Nujol, cm⁻¹): 3036, 1778, 1743. ¹H NMR (300 MHz, CDCl₃) δ : 6.69 (t, *J* = 2.4 Hz, 2H), 5.37 (t, *J* = 3.4 Hz, 1H), 4.55–4.49 (m, 2H), 3.16–3.13 (m, 2H), 2.98–2.96 (m, 2H), 2.14 (s, 3H), 2.05 (s, 6H). LR-MS (CI) *m/z*: 323 [M⁺ + 1]. Anal. calcd. for C₁₆H₁₈O₇: C 59.62, H 5.63; found: C 59.66, H 5.69.

1,3-Di(acetyloxy)-2,3,3a,4,5,7a-hexahydro-1*H*-2-indenyl acetate (24)

Ketone 23 (85 mg, 0.264 mmol) was heated to 160 °C in nitrobenzene solvent (2 mL) for 2 h. The reaction mixture was cooled to 80 °C and the nitrobenzene was distilled under reduced pressure. The residue obtained was chromatographed on a silica gel column (10% ethyl acetate – hexane) to deliver the crystalline cyclohexadiene triacetate 24 (39 mg, 50%), mp 198.6–199.9 °C. IR (Nujol, cm⁻¹): 1740. ¹H NMR (300 MHz, CDCl₃) δ: 5.88–5.84 (m, 2H, C=CH), 5.70–5.66 (m, 2H, C=CH), 5.25 (t, J = 4.5 Hz, 1H, CHOAc), 5.09-5.05 (m, 2H, CHOAc), 3.10 (br s, 2H), 2.09 (s, 3H, COCH₃), 2.07 (s, 6H, COCH₃). ¹³C NMR (75 MHz, CDCl₃) δ: 170.1 (2C, COCH₃), 170.0 (COCH₃), 124.7 (2C, C=CH), 122.4 (2C, C=CH), 78.0 (2C, CHOAc), 69.5 (CHOAc), 38.9 (2C, CH_{bridgehead}), 20.7 (2C, COCH₃), 20.6 (COCH₃). LR-MS (EI, 70° eV) m/z: 252 [M⁺ – Ac + 1]. Anal. calcd. for C₁₅H₁₈O₆: C 61.22, H 6.16; found: C 61.70, H 6.42.

1,3-Di(acetyloxy)-4,5,6,7-tetrahydroxyperhydro-2-indenyl acetate (25)

To a solution of diene 24 (35 mg, 0.119 mmol) in acetone-water-t-BuOH (5:5:2, 2 mL), catalytic OsO4 (0.3 mg, 1 mol%) and 50% aq. NMMO (83 µL, 0.357 mmol) were sequentially added at 0 °C. The resulting pale yellow reaction mixture was stirred at room temperature for 48 h. The reaction was cooled to 0 °C, prior to quenching by addition of solid NaHSO₃ (10 mg). The resulting mixture was diluted with ethyl acetate (15 mL), filtered through Celite, and the filtrate was concentrated under reduced pressure. The residue obtained was subjected to silica gel column chromatography (5% methanol - ethyl acetate) to afford tetrol 25 (38 mg, 88%). IR (Nujol, cm⁻¹): 3290, 1743. ¹H NMR $(300 \text{ MHz}, D_2\text{O}) \delta$: 5.38 (t, J = 6.3 Hz, 1H), 5.27 (dd, J =6.3, 1.8 Hz, 1H), 5.08 (t, J = 6.9 Hz, 1H), 3.97 (br s, 1H), 3.90 (t, J = 3.9 Hz, 1H), 3.78 (dd, J = 7.8, 3.6 Hz, 1H), 3.66(dd, J = 8.1, 3 Hz, 1H), 2.52 (m, 1H), 2.40 (br s, 1H), 1.99(s, 6H, COCH₃), 1.95 (s, 3H, COCH₃). ¹³C NMR (75 MHz, D₂O) δ : 174.2 (COCH₃), 174.1 (COCH₃), 173.2 (COCH₃), 75.0, 74.7, 71.7, 71.1, 70.0, 69.0, 68.6, 45.6 (CH_{bridgehead}), 43.5 (CH_{bridgehead}), 20.9 (COCH₃), 20.8 (COCH₃), 20.5 (COCH₃). Anal. calcd. for C₁₅H₂₂O₁₀: C 49.72, H 6.12; found: C 49.53, H 6.52.

Bicyclitol 18

Tetrol **25** (22 mg, 0.061 mmol) was stirred with aq. NaOH (0.5 mol/L, 200 µL) at room temperature for 1 h and the volatiles were removed under reduced pressure. The residue obtained was dissolved in double distilled water and the solution was filtered with Dowex cation-exchange resin (50 W × 8, H⁺ form). The filtrate was concentrated to furnish **18** (8 mg, 60%). ¹H NMR (300 MHz, D₂O) δ : 4.13 (t, J = 4.3 Hz, 1H), 3.98–3.85 (m, 4H), 3.73 (dd, J = 7.3, 4.0 Hz, 1H), 3.61 (dd, J = 7.6, 2.8 Hz, 1H), 2.21–2.08 (m, 2H). ¹³C NMR (100 MHz, D₂O) δ : 76.4 (CHOH), 75.0 (CHOH), 74.6 (CHOH), 74.4 (CHOH), 73.1 (CHOH), 71.9 (CHOH), 71.8 (CHOH), 48.3 (CH_{bridgehead}), 47.2 (CH_{bridgehead}). LR-MS (CI) m/z: 237 [M⁺ + 1]. Anal. calcd. for C₉H₁₆O₇: C 45.76, H 6.83; found: C 45.84, H 6.79.

(2*R**,3*R**,7*S**,8*R**,9*S**)-5,5-Dimethylspiro[dihydro[1,3]dioxolane]-2,13-4,6-dioxatetracyclo[8.2.1.0^{2,9}.0^{3,7}]tridecen-8-ol (32)

To a magnetically stirred solution of alcohol 28 (8) (2 g, 9.7 mmol) in CH₂Cl₂ (20 mL), NBS (2.1 g, 11.6 mmol) was added in one portion at room temperature and the resulting suspension was stirred for 3 h. Upon complete consumption of the starting material (TLC), the reaction mixture was diluted with CH₂Cl₂ (80 mL) and water (50 mL). Following the usual work-up, subsequent evaporation of the solvent gave a residue, which on chromatographic purification on a silica gel column furnished 29 (2.2 g, 80%). Osmium tetroxide (16 mg, 1 mol%) and 50% aq. NMMO (2.28 mL, 9.75 mmol) was added to a solution of bromoether 29 (1.85 g, 6.5 mmol) in an acetone-water-t-BuOH (5:5:2, 25 mL) solvent system, and the resulting pale yellow reaction mixture was stirred at room temperature for 12 h. The reaction mixture was cooled to ice-bath temperature, diluted with ethyl acetate (70 mL), and the osmate ester was hydrolyzed by addition of solid NaHSO₃ (500 mg). The solution was filtered through a Celite pad and the filtrate was concentrated under reduced pressure to furnish the crude diol 30 (1.76 g). The crude diol 30 was dissolved in dry acetone (20 mL) and Amberlyst-15 (300 mg) and powdered molecular sieves (1 g) were added. The reaction mixture was stirred for 3 h at room temperature and then was filtered through a Celite pad. The filtrate obtained on complete removal of the solvent was chromatographed on a silica gel column to furnish acetonide 31 (1.76 g, 75%). To a solution of bromoether 31 (1 g, 2.78 mmol) in MeOH (15 mL), acetic acid (0.1 mL) and Zn dust (550 mg, 8.34 m atom) were added sequentially at room temperature and the resulting suspension was stirred for 4 h. The reaction mixture was filtered through a Celite pad and the filtrate was concentrated under reduced pressure. The residue obtained was diluted with CH₂Cl₂ (60 mL) and evaporation of the solvent furnished a residue, which on purification on a silica gel column gave alcohol **32** (660 mg, 85%), mp 96 to 97 °C (lit. value (8*a*) mp 98 °C). IR (Nujol, cm⁻¹): 3400.

(2*R**,3*R**,7*S**,8*R**,9*S**)-8-Hydroxy-5,5-dimethyl-4,6dioxatetracyclo[8.2.1.0^{2,9}.0^{3,7}]tridec-11-en-13-one (33)

Ketal 32 (350 mg, 1.24 mmol) was stirred with Amberlyst-15 (~50 mg) in moist acetone (10 mL), under reflux for 3 h. Upon complete consumption of the starting material, the reaction mixture was cooled, diluted with acetone (25 mL), and the resin was filtered through a cotton plug. The filtrate was concentrated under reduced pressure and the residue obtained was chromatographed on a silica gel column (30% ethyl acetate – hexane) to give ketone 33 (237 mg, 81%), mp 179 to 180 °C. IR (Nujol, cm⁻¹): 1771. ¹H NMR (300 MHz, CDCl₃) δ : 6.64 (m, 1H), 6.48 (m, 1H), 4.36 (dd, J = 9.3, 5.4 Hz, 1H), 4.10 (d, J = 5.4 Hz, 1H), 4.03(t, J = 5.4 Hz, 1H), 3.20 (dt, J = 8.7, 3.9 Hz, 1H), 3.14-2.95(m, 3H), 1.44 (s, 3H), 1.23 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ: 200.1 (C=O), 135.2 (C=CH), 130.3 (C=CH), 110.8 (C_q), 89.4 (CH_{acetonide}), 82.0 (CH_{acetonide}), 81.3 (CHOH), 49.2, 49.0 (CH), 47.1 (CH), 44.4 (CH), 27.9 (CH_{3acetonide}), 25.3 (CH_{3acetonide}). LR-MS (EI, 70 eV) m/z: 237 [M⁺ + 1]. Anal. calcd. for $C_{13}H_{16}O_4$: C 66.09, H 6.83; found: C 66.46, H 7.22.

(3a*R**,3b*S**,7a*R**,8*R**,8a*S**)-2,2-Dimethyl-3b,7a,8,8atetrahydro-3a*H*-indeno[1,2-*d*][1,3]dioxol-8-ol (34)

A solution of ketone 33 (220 mg, 0.932 mmol) in nitrobenzene (6 mL) solvent was heated, with vigorous stirring, to 160 °C and reacted for 4 h. Upon complete consumption of the starting material (TLC), the reaction mixture was cooled to 80 °C and the solvent was distilled off under reduced pressure. The resulting dark brown residue was charged on the silica gel column and chromatographed (25%) ethyl acetate - hexane) to give diene 34 (130 mg, 67%) as a white solid, mp 190.6–191.2 °C. IR (Nujol, cm⁻¹): 3470. ¹H NMR (300 MHz, CDCl₃) δ : 6.17 (dd, J = 9.9, 5.1 Hz, 1H, C = CH, 5.80 (dd, J = 9.9, 4.8 Hz, 2H, C = CH), 5.72–5.70 (m, 1H), 4.74 (d, J = 6.0 Hz, 1H, H_{acetonide}), 4.45 (d, J =6 Hz, 1H, $H_{acetonide}$), 4.03 (br s, 1H, CHOH), 3.08–3.05 (m, 2H, $H_{bridgehead}$), 2.26 (d, J = 2.4 Hz, 1H, $H_{bridgehead}$), 1.42 (s, 3H), 1.31 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ: 129.2 (C=CH), 127.6 (C=CH), 123.3 (C=CH), 122.9 (C=CH), 110.4 (C_q), 87.3 (CH_{acetonide}), 83.9 (CH_{acetonide}), 79.6 (CHOH), 45.3 (CH), 39.6 (CH), 26.8 (CH_{3bridgehead}), 24.3 (CH_{3bridgehead}). LR-MS (EI, 70 eV) m/z: 208 [M⁺]. Anal. calcd. for C₁₂H₁₆O₃: C 69.21, H 7.74; found: C 68.88, H 7.52.

(3a*R**,3b*R**,4*S**,5*S**,6*S**,7*S**,7a*S**,8*R**,8a*S**)-2,2-Dimethylperhydroindeno[1,2-*d*][1,3]dioxole-4,5,6,7,8pentaol (35)

To a solution of diene **34** (45 mg, 0.216 mmol) in an acetone–water–*t*-BuOH (5:5:2, 4 mL) solvent system, catalytic OsO₄ (1 mol%, 0.5 mg) and 50% aq. NMMO (151 μ L, 0.648 mmol) were added at 0 °C. The resulting pale yellow reaction mixture was stirred at room temperature for 48 h, before diluting with ethyl acetate (10 mL) and quenching with solid NaHSO₃ (10 mg). The resulting mixture was filtered through a Celite pad and the filtrate was concentrated under reduced pressure. The residue was subjected to silica gel column chromatography (5% methanol – ethyl acetate) to afford pentaol **35** (49 mg, 83%). ¹H NMR (300 MHz, D₂O) δ : 4.70–4.65 (m, 1H), 4.35 (dd, *J* = 6.0, 1.8 Hz, 1H),

4.06 (m, 1H), 3.98–3.93 (m, 2H), 3.69–3.58 (m, 2H), 2.59–2.51 (m, 1H, $H_{bridgehead}$), 2.41–2.37 (m, 1H, $H_{bridgehead}$), 1.32 (s, 3H), 1.20 (s, 3H). ¹³C NMR (75 MHz, D₂O) δ : 111.0 (C_q), 87.9 (CH_{acetonide}), 83.0 (CH_{acetonide}), 77.1 (CHOH), 70.5, 70.4, 70.3, 69.2, 45.8, 45.6, 25.8 (C_{bridgehead}), 23.4 (C_{bridgehead}). LR-MS (EI, 70 eV) *m*/*z*: 258 [M⁺ – H₂O].

Bicyclitol 26

Acetonide **35** (25 mg, 0.09 mmol) was stirred in 30% trifluoroacetic acid (1 mL) at room temperature for 30 min until all the starting material had been consumed. The volatiles were removed under reduced pressure to furnish polyol **26** (19 mg, 89%). ¹H NMR (300 MHz, D₂O) δ : 4.19–4.15 (m, 1H), 4.01–3.81 (m, 4H), 3.68–3.64 (m, 1H), 3.61–3.56 (m, 1H), 2.40–2.33 (m, 1H), 2.13–2.07 (m, 1H). ¹³C NMR (75 MHz, D₂O) δ : 77.9, 77.2, 73.6, 72.9, 72.3, 68.4, 67.4, 45.9, 41.8. LR-MS (CI) *m*/*z*: 237 [M⁺ + 1]. Anal. calcd. for C₉H₁₆O₇: C 45.76, H 6.83; found: C 45.88, H 7.32.

(2*S**,3*R**,7*S**,8*R**,9*R**)-5,5-Dimethyl-4,6,11,12tetraoxatetracyclo[8.2.2.0^{2,9}.0^{3,7}]tetradec-13-en-8-ol (36)

A slow stream of oxygen gas was bubbled through a solution of diene 34 (60 mg, 0.288 mmol) in chloroform (6 mL), contained in a specially designed reaction flask with an outer water jacket, in the presene of methylene blue (~5 mg) as the sensitizer and under irradiation from a 500 W tungsten lamp. The temperature of the reaction mixture was maintained at 20 °C by circulating cold water through the outer jacket of the reaction flask. Upon complete consumption of the starting material (TLC, 8 h), the solvent was evaporated under reduced pressure. The residue obtained was chromatographed over a silica gel column (20% ethyl acetate - hexane) to furnish the endo-peroxide 36 (62 mg, 90%). IR (KBr, cm⁻¹): 3445. ¹H NMR (300 MHz, CDCl₃) δ: 6.78 (m, 1H, C=CH), 6.59 (m, 1H, C=CH), 4.85-4.80 (m, 2H, H_{peroxy}), 4.36 (dd, J = 9.0, 5.2 Hz, 1H, $H_{acetonide}$), 4.17 (t, J = 5.2 Hz, 1H, H_{acetonide}), 4.11 (d, J = 6.0 Hz, 1H), 3.33 (dt, J = 8.7, 4.0 Hz, 1H, H_{bridgehead}), 3.08 (dd, J = 8.7, 5.2 Hz, 1H), 1.47 (s, 3H, CH_3), 1.26 (s, 3H, CH_3). ¹³C NMR (75 MHz, CDCl₃) δ: 135.3 (C=CH), 130.2 (C=CH), 111.7 (C_q), 88.0, 80.9, 79.9, 70.9 (CH_{peroxy}), 70.8 (CH_{peroxy}), 44.5, 44.0, 27.8, 25.2. LR-MS (CI) m/z: 241 [M⁺ + 1]. Anal. calcd. for C₁₂H₁₆O₅: C 59.99, H 6.71; found: C 60.21, H 6.83.

(3a*R**,3b*R**,4*R**,7*S**,7a*S**,8*R**,8a*S**)-2,2-Dimethyl-3b,4,7,7a,8,8a-hexahydro-3a*H*-indeno[1,2-*d*][1,3]dioxole-4,7,8-triol (37)

LiAlH₄ (12 mg, 0.312 mmol) was added to a solution of the endo-peroxide **36** (50 mg, 0.208 mmol) in dry THF (6 mL) at 0 °C under an atmosphere of N₂. The resulting suspension was stirred at room temperature for 1 h, before quenching the reaction by careful addition of ethyl acetate (15 mL) followed by addition of a cold, satd. Na₂SO₄ solution (2 mL). After dilution with ethyl acetate, the aluminium salts were filtered through a Celite pad. Evaporation of the solvent under reduced pressure, after the usual work-up, gave a residue that was chromatographed over a silica gel column (60% ethyl acetate – hexane) to furnish the 1,4-diol **37** (27 mg, 54%), mp 210 to 211 °C. IR (Nujol, cm⁻¹): 3444. ¹H NMR (300 MHz, CDCl₃) δ : 5.69 (br s, 2H, C=CH), 4.71 (d, J = 5.7 Hz, 1H), 4.42 (d, J = 5.7 Hz, 1H), 4.13 (br s, 1H), 4.07 (d, J = 4.8 Hz, 1H), 3.81 (d, J = 7.8 Hz, 1H), 2.36 (m, 1H), 1.90 (t, J = 8.4 Hz, 1H), 1.33 (s, 3H), 1.22 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ : 133.6 (C=CH), 129.9 (C=CH), 111.5 (C_q), 86.0, 83.2, 77.7, 66.5, 62.5, 46.3, 46.1, 25.7, 23.5. LR-MS (CI) *m/z*: 243 [M⁺ + 1]. Anal. calcd. for C₁₂H₁₈O₅: C 59.49, H 7.49; found: C 59.63, H 7.52.

(3a*R**,3b*R**,4*S**,5*S**,6*R**,7*R**,7a*S**,8*R**,8a*S**)-2,2-Dimethylperhydroindeno[1,2-d][1,3]dioxole-4,5,6,7,8-pentaol (38) and (3a*R**,3b*R**,4*S**,5*R**,6*S**,7*R**,7a*S**,8*R**,8a*S**)-2,2dimethylperhydroindeno[1,2-d][1,3]dioxole-4,5,6,7,8pentaol (39)

To a solution of diol **37** (55 mg, 0.227 mmol) in an acetone–water–*t*-BuOH (5:5:2, 5mL) solvent system, catalytic OsO₄ (0.6 mg, 1 mol%), and 50% aq. NMMO (64 μ L, 0.272 mmol) were added sequentially at 0 °C and the resulting mixture was stirred at room temperature for 1 h. The reaction was quenched by addition of solid NaHSO₃ (10 mg), diluted with ethyl acetate (15 mL), and filtered through a Celite pad. The filtrate was concentrated under reduced pressure to yield a residue, whose ¹H NMR spectrum was found to be a mixture of diastereomers **38** and **39** (60 mg) in a 3:1 ratio. However, their chromatographic separation was unsuccessful and the mixture was used as such for the next step.

(3a*R**,3b*S**,6a*S**,6b*S**,6*R**,9a*S**,10*R**,10a*R**,10b*R**)-2,2,5,5,8,8-Hexamethylperhydrodi[1,3]dioxolo-[4',5':1, 2:4',5':6,7]indeno[4,5-*d*][1,3]dioxole-10-ol (40) and (3a*R**,3b*R**,4*S**,4a*R**,7a*S**,8*R**,8a*R**,9*R**,9a*S**)-2,2,6,6tetramethylperhydro[1,3]dioxolo[4',5':1,2]indeno[5,6*d*][1,3]dioxole-4,8,9-triol (41)

The solution of mixture of **38** and **39** (60 mg) in dry acetone (10 mL) was treated sequentially with Amberlyst-15 resin (~5 mg) and 4 Å molecular sieves (~30 mg) and the resulting mixture was stirred at room temperature for 3 h. The reaction mixture was filtered through a Celite pad and the filtrate was concentrated under reduced pressure. The residue obtained was charged on a silica gel column and chromatographed with a 10% ethyl acetate - hexane solvent system to furnish trisacetonide 40 (48 mg, 62%). Further elution of the column with 30% ethyl acetate -hexane gave bisacetonide 41 (14 mg, 21%). Compound 40: mp 210 to 211 °C. IR (Nujol, cm⁻¹): 3361. ¹H NMR (300 MHz, CDCl₃) δ : 4.75 (d, J = 6.0 Hz, 1H), 4.42 (dd, J = 9.6, 5.1 Hz, 2H), 4.36 (dd, J = 13.5, 5.1 Hz, 1H), 4.28 (dd, J =6.9, 4.5 Hz, 1H), 4.08 (d, J = 5.1 Hz, 1H), 3.93 (t, J =6.9 Hz, 1H), 2.64-2.55 (m, 2H), 1.53 (s, 3H), 1.51 (s, 3H), 1.41 (s, 3H), 1.37 (s, 3H), 1.35 (s, 3H), 1.26 (s, 3H). LR-MS (EI, 70 eV) m/z: 356 [M⁺]. Compound 41: mp 190 to 191 °C. IR (Nujol, cm⁻¹): 3372. ¹H NMR (300 MHz, $CDCl_3$) δ : 4.74 (br d, J = 5.1 Hz, 1H), 4.49 (d, J = 5.1 Hz, 1H), 4.41 (br d, J = 5.4 Hz, 1H), 4.15–4.10 (m, 2H), 4.02– 3.96 (m, 1H), 3.73 (dd, J = 12.3, 6.9 Hz, 1H), 2.53–2.50 (m, 2H), 1.51 (s, 3H), 1.43 (s, 3H), 1.34 (s, 3H), and 1.30 (s, 3H). LR-MS (EI, 70 eV) m/z: 317 [M⁺ + 1].

Bicyclitol 42

Trisacetonide **40** (35 mg, 0.098 mmol) was stirred with 30% trifluoroacetic acid (500 μ L) at room temperature for

30 min. Upon complete consumption of the starting material, the volatiles were removed under reduced pressure to afford **42** (21 mg, 92%). ¹H NMR (300 MHz, D₂O) δ : 4.03 (dd, *J* = 6.9, 4.5 Hz, 1H), 3.89–3.85 (m, 2H), 3.74–3.70 (m, 3H), 3.62 (dd, *J* = 6.6, 3 Hz, 1H), 2.46 (dd, *J* = 14.4, 6.9 Hz, 1H), 2.11 (dd, *J* = 12.3, 6.9 Hz, 1H). ¹³C NMR (75 MHz, D₂O) δ : 78.9, 77.6, 72.7, 72.2, 71.5, 70.8, 70.1, 47.5, 41.0. LR-MS (CI) *m/z*: 237 [M⁺ + 1]. Anal. calcd. for C₉H₁₆O₇: C 45.76, H 6.83; found: C 45.53, H 6.79.

Bicyclitol 43

Bisacetonide **41** (10 mg, 0.032 mmol) was treated with 30% trifluoroacetic acid (150 µL) and the resulting biphasic mixture was stirred vigorously at room temperature for 30 min. The volatiles were removed under reduced pressure to furnish **43** (7 mg, 88%). ¹H NMR (300 MHz, D₂O) δ : 4.10 (t, 1H, *J* = 5.7 Hz), 4.00 (t, 1H, *J* = 5.4 Hz), 3.86 (t, *J* = 5.4 Hz, 1H), 3.83 (dd, *J* = 14, 7.2 Hz, 1H), 3.75–3.70 (m, 1H), 3.66–3.61 (m, 2H), 2.36 (dd, *J* = 15, 6.9 Hz, 1H), 1.93 (dd, *J* = 14.4, 6.9 Hz, 1H). ¹³C NMR (75 MHz, D₂O) δ : 77.8, 77.1, 74.0, 73.2, 72.3, 72.0, 68.8, 48.6, 44.2. LR-MS (CI) *m/z*: 237 [M⁺ + 1]. Anal. calcd. for C₉H₁₆O₇: C 45.76, H 6.83; found: C 45.77, H 6.98.

(2*R**,3*S**,6*R**,7*S**)-1,8,9,10-Tetrachloro-11,11-dimethoxytricyclo[6.2.1.0^{2,7}]undeca-4,9-diene-3,6-diol (45)

To a solution of 1,2,3,4-tetrachloro-5,5-dimethoxycyclopentadiene (63 mL, 0.36 mol) in dry toluene (300 mL), pbenzoquinone (40 g, 0.37 mol) was added and the resulting solution was refluxed for 24 h. Excess solvent was distilled off under reduced pressure and the residue obtained was recrystallized from hexane to furnish adduct 17 (110 g, 83%). Adduct 17 (10 g, 26.8 mmol) was dissolved in dry MeOH (150 mL) and to this solution at 0 °C under an inert atmosphere, CeCl₃·7H₂O (1 g, 10 mol%) and NaBH₄ (2 g, 53.6 mmol) were added in sequential portions. The resulting solution was stirred at the same temperature for 10 min until the starting material was completely consumed. The reaction mixture was diluted with ethyl acetate (150 mL) and quenched by the addition of water (80 mL). The organic layer was separated and after the usual work-up furnished residue 45 (6.9 g, 69%), mp 162 to 163 °C (lit. value (16) mp 167 to 168 °C). IR (Nujol, cm⁻¹): 3450.

(2*R**,7*S**)-1,8,9,10-Tetrachloro-11,11-

dimethoxytricyclo[6.2.1.0^{2,7}]undeca-3,5,9-triene (44)

To a magnetically stirred solution of diol 45 (5 g, 13.3 mmol) in dry CH_2Cl_2 (25 mL), triethylamine (5.6 mL, 39.9 mmol), DMAP (162 mg, 10 mol%), and methanesulphonyl chloride (2.5 mL, 29.2 mmol) were added sequentially at 0 °C under an inert atmosphere and the resulting solution was stirred at room temperature for a further 24 h. The reaction mixture was diluted with CH_2Cl_2 (50 mL) prior to quenching the reaction by adding water (30 mL). The organic layer was separated and the aqueous layer was extracted with CH_2Cl_2 (3 × 60 mL). The residue obtained by the usual work-up was subjected to chromatographic purification on a silica gel column (35% ethyl acetate – hexane) to furnish dimesylate **46** (5.1 g, 72%). Dimesylate **46** (8.4 g, 15.7 mmol) was dissolved in 2-butanone (60 mL) and so-

dium iodide (8.2 g, 54.9 mmol) was added to this solution. The dark brown solution was then refluxed for 8 h. The reaction mixture was cooled to ice bath temperature and diluted with diethylether (100 mL) and water (80 mL). The usual work-up and evaporation of the solvent under reduced pressure gave a residue, which on purification on a silica gel column (hexane) furnished the tricyclic diene **44** (4.3 g, 80%), mp 162 to 163 °C (lit. value (16) mp 167 to 168 °C).

(2*R**,3*S**,4*S**,5*R**,6*R**,7*S**)-1,8,9,10-Tetrachloro-11,11dimethoxytricyclo[6.2.1.0^{2,7}]undec-9-ene-3,4,5,6-tetraol (48)

A solution of diene 44 (1.2 g, 3.51 mmol) in an acetonewater-t-BuOH solvent system (5:5:2, 50 mL) was treated sequentially with catalytic OsO₄ (8 mg, 1 mol%) and 50% aq. NMMO (2.5 mL, 10.53 mmol). The resulting pale yellow solution was stirred vigorously at room temperature for 48 h, during which all the starting material was consumed (TLC). The reaction was quenched by addition of solid NaHSO₃ (500 mg), diluted with ethyl acetate (100 mL), and filtered through a Celite pad. The filtrate was concentrated under reduced pressure to give a residue, which was chromatographed on a silica gel column (70% ethyl acetate – hexane) to furnish tetrol **48** (950 mg, 66%). IR (Nujol, cm⁻¹): 3430. ¹H NMR (300 MHz, CDCl₃) δ: 3.76 (br s, 2H), 3.66 (br s, 2H), 3.63 (s, 3H), 3.56 (s, 3H), 3.14 (m, 2H). ¹³C NMR (75 MHz, CDCl₃) δ: 129.7 (2C, C=CCl), 111.7 (C₀), 70.7 (2C, CHOH), 68.0 (2C, CHOH), 52.8 (2C, C-Cl), 52.5 (2C, OMe), 51.8 (2C, CH). LR-MS (EI, 70 eV) *m/z*: 410 [M⁺ + 1].

(2*R**,3*S**,4*S**,8*R**,9*R**,10*S**)-1,11,12,13-Tetrachloro-14,14-dimethoxy-6,6-dimethyl-5,7-dioxatetracyclo[9.2.1.0^{2,10}.0^{4,8}]tetradec-12-ene-3,9-diol (49)

Tetrol **48** (950 mg, 2.32 mmol) was stirred with Amberlyst-15 (~100 mg) and 4 Å molecular sieves (500 mg) in dry acetone (30 mL) at room temperature for 3 h. Upon complete consumption of the starting material, the reaction mixture was filtered through a Celite pad. The filtrate was concentrated under reduced pressure to give a residue, which was chromatographed over a short silica gel column (25% ethyl acetate – hexane) to afford acetonide **49** (783 mg, 75%), mp 180.1–181.4 °C. IR (Nujol, cm⁻¹): 3346. ¹H NMR (300 MHz, CDCl₃) δ : 4.44 (s, 2H), 3.63 (s, 3H), 3.59 (s, 3H), 3.11–3.01 (m, 2H), 2.29 (d, *J* = 6.6 Hz, 2H), 1.57 (s, 3H), 1.37 (s, 3H). LR-MS (EI, 70 eV) *m/z*: 451 [M⁺ + 1]. Anal. calcd. for C₁₆H₂₀O₆Cl₄: C 42.69, H 4.48; found: C 42.84, H 4.80.

(2*R**,3*R**,4*R**,8*S**,9*S**,10*S**)-14,14-Dimethoxy-6,6dimethyl-5,7-dioxatetracyclo[9.2.1.0^{2,10}.0^{4,8}]tetradec-12ene-3,9-diol (50)

Liquid ammonia (300 mL) was distilled into a 250 mL three-necked round-bottomed flask equipped with a condenser, KOH guard tube, and septum. Acetonide **49** (10 g, 22.22 mmol) in dry THF (40 mL) and absolute EtOH (40 mL) were added to the flask. Small pieces of freshly cut sodium were introduced into the reaction mixture with stirring at -33 °C until the blue color persisted. The mixture was stirred for 30 min and solid NH₄Cl (1 g) was added. The excess ammonia was allowed to evaporate, and a cold

aq. satd. NH₄Cl solution (50 mL) and water (100 mL) were added to the resulting residue, which was subsequently extracted with ethyl acetate (3 × 200 mL). Removal of the volatiles, followed by purification of the residue by silica gel column chromatography (50% ethyl acetate – hexane) gave the dechlorinated product **50** (3.4 g, 49%), mp 173–175 °C. IR (Nujol, cm⁻¹): 3460, 3380. ¹H NMR (300 MHz, CDCl₃) δ : 6.93 (s, 2H, C=CH), 4.38 (s, 2H), 3.24 (s, 3H), 3.17 (s, 5H), 3.05 (s, 2H), 2.60–2.50 (m, 2H), 1.91 (s, 1H), 1.88 (s, 1H), 1.53 (s, 3H), 1.35 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ : 133.8 (2C, C=CH), 118.9 (C_q), 110.0 (C_q), 76.6 (2C, C_{acetonide}), 68.8 (2C, CHOH), 52.0 (OCH₃), 50.0 (OCH₃), 46.8 (2C, CH), 40.4 (2C, CH), 25.9 (CH), 24.0 (CH). LR-MS (EI, 70 eV) *m/z*: 312 [M⁺]. Anal. calcd. for C₁₆H₂₄O₆: C 61.52, H 7.74; found: C 61.83, H 7.90.

(2*R**,3*R**,4*R**,8*S**,9*S**,10*S**)-3,9-Dihydroxy-6,6-dimethyl-5,7-dioxatetracyclo[9.2.1.0^{2,10}.0^{4,8}]tetradec-12-en-14-one (51)

A solution of ketal **50** (1 g, 3.20 mmol) in moist acetone (40 mL) was stirred with Amberlyst-15 (~150 mg) at room temperature for 2 h. Upon complete consumption of the starting material, the reaction mixture was filtered through a cotton plug. The filtrate on concentration gave a solid residue, which on purification by chromatography over a short silica gel column (50% ethyl acetate – hexane) afforded ketone **51** (834 mg, 98%), mp 213–215 °C. IR (Nujol, cm⁻¹): 3270, 1777.

(3a*R**,4*R**,4a*S**,8a*R**,9*S**,9a*S**)-2,2-Dimethyl-

3a,4,4a,8a,9,9a-hexahydronaphtho[2,3-*d*][1,3]dioxole-4,9diol (52)

A solution of ketone 51 (300 mg, 1.13 mmol) in nitrobenzene (15 mL) was heated, with vigorous stirring, at 160 °C for 4 h. Upon complete consumption of the starting material (TLC), the reaction mixture was cooled to 80 °C and the solvent was distilled off under reduced pressure. The residue left behind was chromatographed over a silica gel column (30% ethyl acetate - hexane) to furnish diene 52 (169 mg, 63%) as a colorless solid, mp 189 to 190 °C. IR (KBr, cm⁻¹): 3356. ¹H NMR (300 MHz, CDCl₃) δ: 5.87– 5.83 (m, 2H, C=CH), 5.65–5.61 (m, 2H, C=CH), 4.42–4.40 (m, 2H, $H_{acetonide}$), 3.74 (br s, 2H, CHOH), 3.00–2.98 (m, 2H, OH), 2.70–2.67 (m, 2H), 1.55 (s, 3H), 1.40 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ: 125.8 (2C, C=CH), 122.6 (2C, C=CH), 109.3 (C_q), 74.8 (2C, C_{acetonide}), 69.0 (2C, CHOH), 35.4 (2C, CH), 26.0 (CH), 24.4 (CH). LR-MS (EI, 70 eV) m/z: 239 [M⁺ + 1]. Anal. calcd. for C₁₃H₁₈O₄: C 65.53, H 7.61; found: C 65.90, H 7.72.

(3a*R**,4*R**,4a*R**,5*S**,6*S**,7*S**,8*S**,8a*S**,9*S**,9a*S**)-2,2-Dimethylperhydronaphtho[2,3-*d*][1,3]dioxole-4,5,6,7,8,9hexol (53)

Diene **52** (25 mg, 0.105 mmol) was treated sequentially with catalytic amounts of OsO_4 (0.2 mg, 1 mol%) and 50% aq. NMMO (54 µL, 0.231 mmol) in an acetone–water–*t*-BuOH solvent system (5:5:2, 5 mL) at 0 °C and the resulting pale yellow reaction mixture was stirred at room temperature for 48 h. The reaction mixture was cooled to 0 °C and diluted with ethyl acetate (20 mL) prior to quenching with

solid NaHSO₃ (10 mg). The mixture was filtered through a Celite pad and the filtrate was concentrated. The residue thus obtained was chromatographed over a silica gel column (8% methanol – ethyl acetate) to furnish hexol **53** (27 mg, 85%). IR (Nujol, cm⁻¹): 3350. ¹H NMR (300 MHz, D₂O) δ : 4.46–4.43 (m, 1H), 4.35–4.31 (m, 1H), 4.13–4.11 (br t, J = 3.3 Hz, 1H), 3.97 (br dd, J = 6.6, 2.7 Hz, 1H), 3.92 (br s, 1H), 3.85 (br dd, J = 12.0, 3.0 Hz, 1H), 3.71 (br dd, J = 8.7, 2.1 Hz, 1H), 3.64–3.61 (m, 1H), 2.21 (m, 1H), 2.06 (m, 1H), 1.39 (s, 3H), 1.26 (s, 3H). ¹³C NMR (75 MHz, D₂O) δ : 111.1 (C_q), 77.9, 76.6, 72.8, 71.3, 70.9, 70.7, 68.4, 66.2, 40.0 (CH), 38.9 (CH), 26.2, 24.9. LR-MS (EI, 70 eV) *m/z*: 289 [M⁺ – H₂O + 1]. Anal. calcd. for C₁₃H₂₂O₈: C 50.97, H 7.24; found: C 50.63, H 7.52.

Bicyclitol 54

Acetonide **53** (20 mg, 0.065 mmol) was stirred vigorously with 30% trifluoroacetic acid (800 µL) at room temperature for 30 min. The volatiles were removed under reduced pressure to furnish **54** (16 mg, 95%). ¹H NMR (300 MHz, D₂O) δ : 4.19 (br s, 1H), 4.00 (br s, 1H), 3.96–3.81 (series of m, 4H), 3.75–3.60 (m, 2H), 2.22–2.18 (m, 2H). ¹³C NMR (75 MHz, D₂O) δ : 77.0, 76.7, 76.0, 74.2, 73.2, 71.2 (3C), 43.1 (CH), 40.5 (CH). LR-MS (EI, 70 eV) *m*/*z*: 264 [M⁺ – 2]. Anal. calcd. for C₁₀H₁₈O₈: C 45.11, H 6.81; found: C 45.32, H 7.01.

(2*R**,3*S**,4*S**,5*R**,6*S**,7*S**)-1,8,9,10-Tetrachloro-11,11dimethoxytricyclo[6.2.1.0^{2,7}]undec-9-ene-3,4,5,6-tetrol (55)

A solution of diol 45 (1 g, 2.66 mmol) in acetone-water*t*-BuOH (5:5:2, 15 mL) was treated sequentially with OsO_4 (7 mg, 1 mol%) and 50% aq. NMMO (747 µL, 3.19 mmol) at 0 °C. The resulting pale yellow reaction mixture was stirred at room temperature for 1.5 h, at which point all the starting material had been consumed (TLC). The reaction mixture was cooled to 0 °C and quenched by the addition of solid NaHSO₃ (500 mg). The mixture was filtered through a Celite pad and the filtrate was concentrated under reduced pressure. The residue obtained was chromatographed over a silica gel column (30% ethyl acetate - hexane) to furnish 55 (817 mg, 75%). IR (Nujol, cm⁻¹): 3420, 3315, 3245. ¹H NMR (300 MHz, DMSO) δ: 3.83 (br s, 4H), 3.50 (s, 3H), 3.43 (s, 3H), 3.00 (s, 2H). ¹³C NMR (75 MHz, DMSO) δ : 128.7 (2C, C=CCl), 114.6 (C_q), 77.1 (2C, C-Cl), 68.4 (2C, CHOH), 67.3 (2C, CHOH), 52.4 (OCH₃), 51.5 (OCH₃), 46.2 (2C, CH). LR-MS (EI, 70 eV) *m/z*: 374 [M⁺ – Cl + 1]. Anal. calcd. for C₁₃H₁₆Cl₄O₄: C 38.08, H 3.93; found: C 37.88, H 3.88.

(2*R**,3*R**,4*S**,8*R**,9*R**,10*S**)-1,11,12,13-Tetrachloro-14,14-dimethoxy-6,6-dimethyl-5,7-dioxatetracyclo[9.2.1.0^{2,10}.0^{4,8}]tetradec-12-ene-3,9-diol (56)

To a solution of tetrol **55** (750 mg, 1.829 mmol) in dry acetone (25 mL), Amberlyst-15 (~100 mg) and 4 Å molecular sieve powder (250 mg) were added sequentially and the resulting mixture was stirred at room temperature for 1 h. Upon complete consumption of the starting material (TLC), the reaction mixture was filtered through a Celite pad and the filtrate was concentrated under reduced pressure to give a solid residue. The residue was chromatographed over a silica gel column (6% ethyl acetate – hexane) to afford acetonide **56** (675 mg, 82%), mp 210–212 °C. IR (KBr, cm⁻¹): 3260. ¹H NMR (300 MHz, CDCl₃) δ : 4.43 (s, 2H, H_{acetonide}), 4.29 (br s, 2H, CHOH), 3.79 (s, 3H, OCH₃), 3.57 (s, 3H, OCH₃), 3.04 (s, 2H, H_{bridgehead}), 1.45 (s, 3H), 1.34 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ : 129.4 (2C, C=CCl), 114.7 (C_q), 108.4 (C_q), 76.7 (2C, C_{acetonide}), 75.6 (2C, *C*-Cl), 65.1 (2C, *C*HOH), 53.1 (OCH₃), 51.7 (OCH₃), 46.3 (2C, *C*H), 26.1 (*C*H_{3acetonide}), 23.3 (*C*H_{3acetonide}). LR-MS (EI, 70 eV) *m/z*: 451 [M⁺ + 1]. Anal. calcd. for C₁₆H₂₀O₄Cl₆: C 42.69, H 4.48; found: C 43.04, H 4.62.

(2*R**,3*R**,4*R**,8*S**,9*R**,10*S**)-14,14-Dimethoxy-6,6dimethyl-5,7-dioxatetracyclo[9.2.1.0^{2,10}.0^{4,8}]tetradec-12ene-3,9-diol (57)

Liquid ammonia (100 mL) was distilled into a threenecked round-bottomed flask equipped with a Dewar condenser, KOH guard tube, and septum. The solution of acetonide 56 (1 g, 2.22 mmol) in dry THF (10 mL) and absolute EtOH (5 mL) was added to the reaction flask. Small pieces of freshly cut sodium were introduced, with stirring, into the resulting mixture until the blue color of the solution persisted. The mixture was stirred for 40 min and quenched by addition of solid NH_4Cl (~1 g). The excess ammonia was allowed to evaporate and the residue obtained was diluted with cold aq. satd. NH₄Cl solution (20 mL) and water (50 mL). The residue, resulting from the usual work-up, was purified by silica gel column chromatography (70% ethyl acetate – hexane) to furnish the olefin diol 57 (610 mg, 88%), mp 200–202 °C. IR (Nujol, cm⁻¹): 3420, 1260. ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta$: 6.17 (t, 2H, J = 3.0 Hz, C=CH), 3.94 (br s, 2H, $H_{acetonide}$), 3.32 (s, 3H, OCH₃), 3.13 (br s, 3H, OCH₃), 3.04 (br s, 2H, CHOH), 2.92 (s, 2H), 1.65 (s, 2H), 1.44 (s, 3H), 1.30 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ : 132.0 (2C, C=CH), 119.4 (C_q), 108.4 (C_q), 78.5 (2C, Cacetonide), 71.4 (2C, CHOH), 52.0 (2C, OCH₃), 49.8 (2C), 46.7 (2C), 26.7 (CH_{3acetonide}), 23.0. LR-MS (EI, 70 eV) *m/z*: 312 [M⁺]. Anal. calcd. for C₁₆H₂₄O₆: C 61.52, H 7.74; found: C 61.90, H 7.52.

(2*R**,3*R**,4*R**,8*S**,9*R**,10*S**)-3,9-Dihydroxy-6,6dimethyl-5,7-dioxatetracyclo[9.2.1.0^{2,10}.0^{4,8}]tetradec-12en-14-one (58)

Ketal **57** (220 mg, 0.705 mmol) was dissolved in moist acetone (20 mL) and stirred with Amberlyst-15 resin (~50 mg) for 2 h, when all the starting material had been consumed (TLC). The reaction mixture was filtered through a Celite pad and the filtrate was concentrated under reduced pressure to give a solid residue, which was chromatographed over a silica gel column (50% ethyl acetate – hexane) to furnish the 7-ketonorbornene **58** (178 mg, 95%), mp 220–221.5 °C. IR (KBr, cm⁻¹): 3443, 1778.

(3aR*,4S*,4aS*,8aR*,9R*,9aS*)-2,2-Dimethyl-

3a,4,4a,8a,9,9a-hexahydronaphtho[2,3-*d*][1,3]dioxole-4,9diol (59)

A solution of ketone **58** (90 mg, 0.338 mmol) in nitrobenzene (10 mL) was heated to 160 $^{\circ}$ C, with vigorous stirring, for 3 h. Upon complete consumption of the starting material (TLC), the reaction mixture was cooled to 80 $^{\circ}$ C and the solvent was distilled off under reduced pressure. The resulting residue was subjected to silica gel column chromatography (30% ethyl acetate – hexane) to afford diene **59** (35 mg, 43%), mp 188.5–190 °C. IR (KBr, cm⁻¹): 3321, 3034. ¹H NMR (300 MHz, CDCl₃) δ : 5.97–5.94 (m, 2H, C=CH), 5.54–5.50 (m, 2H, C=CH), 4.50–4.49 (m, 2H, H_{acetonide}), 3.86 (br s, 2H), 3.53 (d, *J* = 6.9 Hz, 2H), 3.20 (br s, 2H, H_{bridgehead}), 1.46 (s, 3H), 1.37 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ : 125.8 (2C, C=CH), 123.8 (2C, C=CH), 108.6 (C_q), 74.9 (2C, C_{acetonide}), 69.7 (2C, CHOH), 32.4 (2C), 26.6 (CH_{3acetonide}), 24.0 (CH_{3acetonide}). LR-MS (EI, 70 eV) *m/z*: 239 [M⁺ + 1]. Anal. calcd. for C₁₃H₁₈O₄: C 65.53, H 7.61; found: C 65.90, H 7.52.

(3a*R**,4*S**,4a*R**,5*S**,6*S**,7*S**,8*S**,8a*S**,9*R**,9a*S**)-2,2-Dimethylperhydronaphtho[2,3-*d*][1,3]dioxole-4,5,6,7,8,9hexol (60)

To a solution of diene 59 (25 mg, 0.105 mmol) in acetone-water-t-BuOH (5:5:2, 6 mL), catalytic amounts of OsO_4 (0.5 mg, 1 mol%) and 50% aq. NMMO (75 µL, 0.315 mmol) were sequentially added at 0 °C. The resulting pale yellow reaction mixture was stirred at room temperature for 48 h. The reaction mixture was cooled to 0 °C prior to quenching with solid NaHSO₃ (15 mg). The resulting mixture was diluted with ethyl acetate (20 mL), filtered through Celite, and the filtrate was concentrated under reduced pressure. The residue was subjected to silica gel column chromatography (10% methanol – ethyl acetate) to afford hexol 60 (23 mg, 73%). IR (Nujol, cm⁻¹): 3460. ¹H NMR (300 MHz, D_2O) δ : 4.27 (d, J = 3 Hz, 1H), 4.20 (dd, J = 8.4, 5.4 Hz, 1H), 4.14 (dd, J = 5.1, 1.2 Hz, 1H), 4.03 (t, J = 3 Hz, 1H), 3.97-3.92 (m, 1H), 3.86 (dd, J = 10.2, 3.6 Hz, 1H), 3.79 (dd, J = 8.4, 5.4 Hz, 1H), 3.52 (dd, J = 10.2, 3.3 Hz, 1H), 2.54– 2.20 (m, 2H), 1.39 (s, 3H), 1.26 (s, 3H). ¹³C NMR (75 MHz, D_2O) δ : 110.5 (C_q), 80.1 (CH_{acetonide}), 78.3 (CH_{acetonide}), 72.7, 72.3, 72.0, 71.5, 67.4, 66.4, 41.9 (CH_{bridgehead}), 39.3 (CH_{bridgehead}), 28.6, 26.6. LR-MS (EI, 70 eV) m/z: 306 [M⁺]. Anal. calcd. for C₁₃H₂₂O₈: C 50.97, H 7.24; found: C 50.90, H 7.51.

Bicyclitol 61

Acetonide hexol **60** (14 mg, 0.046 mmol) was stirred vigorously with 30% TFA (500 µL) at room temperature for 30 min. The volatiles were removed under reduced pressure to furnish **61** (11 mg, 90%). ¹H NMR (300 MHz, D₂O) δ : 4.06 (m, 1H), 3.93 (dd, J = 10.5, 3.0 Hz, 1H), 3.87 (br s, 1H), 3.81 (m, 1H), 3.75 (t, J = 3.9 Hz, 1H), 3.73 (t, J = 3.0 Hz, 1H), 3.68 (t, J = 3.3 Hz, 1H), 3.52 (dd, J = 10.5, 3.0 Hz, 1H), 2.35 (dt, J = 6.6, 2.7 Hz, 1H), 2.28 (m, 1H). ¹³C NMR (75 MHz, D₂O) δ : 73.6, 72.8, 71.3, 70.7, 70.4, 69.3, 69.1, 67.4, 40.2, 38.4. LR-MS (CI) m/z: 266 [M⁺]. Anal. calcd. for C₁₀H₁₈O₈: C 45.11, H 6.81; found: C 45.45, H 7.12.

General procedure for enzymatic assays

The enzymes assayed were α -glucosidase (EC 3.2.1.20) from yeast, β -glucosidase (EC 3.2.1.21) from sweet almonds, α -galactosidase (EC 3.2.1.22) from green coffee beans, β galactosidase (EC 3.2.1.23) from *E. coli*, α -mannosidase (EC 3.2.1.24) from jack bean, and β -mannosidase (EC 3.2.1.25) from snail acetone powder. The substrates for the glycosidase were the corresponding ortho- or para-nitrophenylglycosides. The α -glucosidase (0.135 units/mL)³ was assayed using *p*-nitophenyl- α -D-glucopyranoside (2.53 mmol/L) as the substrate at pH 6.8 (100 mmol/L phosphate buffer) at 37 °C, while β -glucosidase (0.133 units/mL) was assayed using p-nitrophenyl- β -D-glucopyranoside (2.57 mmol/L) as the substrate at pH 5.0 (10 mmol/L acetate buffer) at 37 °C. The potency of α -galactosidase (0.1 units/mL) and β-galactosidase (0.91 units/mL) was studied using p-nitrophenyl-\alpha-D-galactopyranoside (2.12 mmol/L) and onitrophenyl-B-D-galactopyranoside (2.34 mmol/L) as substrate at pH 6.5 (100 mmol/L phosphate buffer) and pH 7.3 (100 mmol/L phosphate buffer) at 25 and 37 °C, respectively. The assay involving α -mannosidase (0.116 units/mL) was carried out using p-nitrophenyl-a-d-mannopyranoside (2.01 µmol/L) as substrate at pH 4.5 (10 mmol/L acetate buffer) at 25 °C, while β -mannosidase (0.125 units/mL) was assayed using p-nitrophenyl-a-D-mannopyranoside (2.15 mmol/L) as substrate at pH 4 (10 mmol/L acetate buffer) at 25 °C.

Typical assays were carried out in 1.5 mL disposable Eppendorf tubes, with the reaction volume being maintained at 200 μ L. The mixture of enzyme and potential inhibitor in the optimum buffer medium was preincubated for 5 min and the reaction was initiated by adding the corresponding substrate solution (prepared in double-distilled water) to the reaction mixture. After incubation for 30 min at the optimum temperature for the enzyme, the reaction was quenched by addition of 800 μ L of Na₂CO₃ buffer (400 mmol/L) solution. The enzyme activity was determined by measuring the absorption of the nitrophenolate ion released at 400 nm (in the case of *p*-nitrophenol) or 410 nm (in the case of *o*-nitrophenol). In the control experiments, the inhibitor was replaced by a corresponding buffer solution. The *K*_i was calculated using the Lineweaver–Burk plot.

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³One unit of enzyme is defined as the amount that liberated 1μ mol of corresponding nitrophenol per min.

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