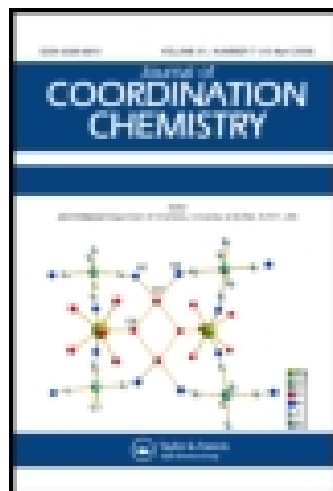


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Thermal stability of Pd(1,4-bis(2-hydroxyethyl)piperazine)Cl₂ and its role in the catalysis of base hydrolysis of α -amino acid esters

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Thermal stability of Pd(1,4-bis(2-hydroxyethyl)piperazine)Cl₂ and its role in the catalysis of base hydrolysis of α -amino acid esters

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Dedicated to Prof. Rudi van Eldik on the occasion of his 70th birthday.

Pd(BHEP)Cl₂ was synthesized and characterized (BHEP = 1,4-bis(2-hydroxyethyl)piperazine). The complex decomposes in two steps, leaving a residue of palladium metal. Amino acid ester (L) reacts with [Pd(BHEP)(H₂O)₂]²⁺ (BHEP = 1,4-bis(2-hydroxyethyl)piperazine), giving mixed ligand complexes, [Pd(BHEP)L]²⁺. The kinetics of hydrolysis of [Pd(BHEP)L]²⁺ have been studied by pH-stat technique and rate constants were obtained. Rate acceleration observed for glycine methyl ester is high. The effect with methionine methyl ester is much less marked, as the mixed-ligand complexes with these ligands do not involve alkoxy carbonyl donors. Possible mechanisms for these reactions are considered.

Keywords: 1,4-Bis(2-hydroxyethyl)piperazine; Thermal stability; Amino acid ester hydrolysis; Pd(II); pH-stat Technique

1. Introduction

Piperazines or cyclizines are generally considered as ethylenediamine derivatives or cyclic ethylenediamines (cyclizines); Piperazines are a broad class of compounds with many important pharmacological properties. This dinitrogen moiety has been a component of a plethora of drugs. Piperazines have chemical similarity with piperidine, a constituent of piperine in the black piper plant (*Piper nigrum*). Piperazine was introduced into medicine as a solvent for uric acid [1]. Metal ions in metalloenzymes such as carboxypeptidase A [2], carbonic anhydrase [3] and

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alkaline phosphatase [4] play key roles in many biochemical processes [5]. In these metal-containing metalloenzymes, metal ions at the active sites are considered to serve as a primary catalytic centre for bringing substrate and nucleophile together through formation of a coordination complex, to activate the substrate carbonyl group facilitating attack of the nucleophile as hydroxide. In order to probe the mechanism by which the metalloenzyme may operate and consequently provide a theoretical base for designing effective artificial metalloenzymes, previous reports [6-8] have developed biomimetic models for metalloenzyme, which catalyze the hydrolysis of carboxylic acid esters in biomimetic models.

Work in our laboratory [9-14] has focused on catalysis of the hydrolysis of various amino acid esters by metal complexes. $[\text{Pd}(\text{en})\text{L}]^{2+}$ undergoes hydrolysis by water and hydroxide ion [15]. It is therefore of considerable interest to extend this work to study the thermal stability of Pd(II) complexes with 1,4-bis(2-hydroxyethyl)piperazine and its role in the catalysis of amino acid esters. The results of this investigation are expected to support the biological activity of the piperazine derivative.

2. Experimental

2.1. Materials and reagent

All reagents were of Analar grade. K_2PdCl_4 and 1,4-bis(2-hydroxyethyl)piperazine are provided by Aldrich. The glycine- and methionine methyl esters were purchased from Fluka. Carbonate-free NaOH was prepared and standardized against potassium hydrogen phthalate solution. All solutions were prepared in deionized H_2O .

2.2. Preparation of the complex

$\text{Pd}(\text{BHEP})\text{Cl}_2$ was prepared by dissolving K_2PdCl_4 (0.92 g, 2.82 mmol) in 10 ml water with stirring. The clear solution of $[\text{PdCl}_4]^{2-}$ was filtered and 1,4-bis(2-hydroxyethyl)piperazine (0.491 g, 2.82 mmol) dissolved in 10 ml H_2O was added dropwise to the stirred solution. The pH was adjusted to 2-3 by addition of HCl and/or NaOH. A yellowish-brown precipitate of $\text{Pd}(\text{BHEP})\text{Cl}_2$ was formed and stirred for a further 30 minute at 50 °C. After filtering off the precipitate, it was thoroughly washed with H_2O , ethanol and diethyl ether. An orange crystalline precipitate was obtained after recrystallization from 2:1 methanol/water mixture. (Found: C, 27.3; H, 5.3; N, 7.8 %. Calc. for $\text{C}_8\text{H}_{18}\text{N}_2\text{O}_2\text{PdCl}_2$ (F.Wt = 351.55): C, 27.33; H, 5.17; N,

7.97 %). IR: ν_{O-H} (st) at 3429 cm^{-1} , ν_{C-H} (st) at 3018, 2983, 2949 and 2891 cm^{-1} , ν_{Pd-N} at 472 cm^{-1} , ν_{Pd-Cl} at 333 and 324 cm^{-1} .

Aqueous solutions of the diaqua form of $\text{Pd}(\text{BHEP})\text{Cl}_2$ were prepared *in situ* by addition of slightly less than two mole equivalents of AgNO_3 to a solution of a known amount of the dichloro complex and stirred overnight. The white precipitate of AgCl that formed was filtered off using a 0.1 μm pore membrane filter. Great care was taken to ensure that the resulting solution was free of Ag^+ ion and that the dichloro complex had been converted completely into the diaqua species.

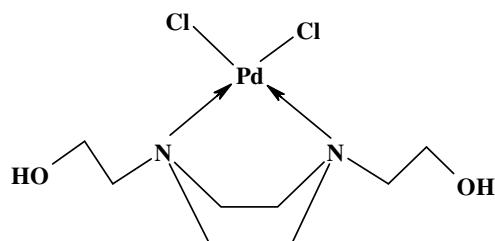
2.3. Instrumentation and measurements

IR spectra were recorded on a 8001-PC FT-IR Shimadzu spectrophotometer using KBr pellets. Thermogravimetric measurements (TGA, DTA, 20–1000 $^\circ\text{C}$) were recorded on a DTG-50 Shimadzu thermogravimetric analyzer at a heating rate of 10 $^\circ\text{C}/\text{min}$ and nitrogen flow rate of 20 ml/min.

The kinetics of hydrolysis was monitored using a Metrohm 751 Titrino operated with the SET mode (titration to a preset end point). The titroprocessor and electrode were calibrated with standard buffer solutions according to NIST [16]. Hydrolysis kinetics of glycine- and methionine methyl esters in the presence of $[\text{Pd}(\text{BHEP})(\text{H}_2\text{O})_2]^{2+}$ are investigated by pH-stat technique [17, 18]. After equilibrating a solution mixture (40 cm^3) containing $[\text{Pd}(\text{BHEP})(\text{H}_2\text{O})_2]^{2+}$ (5.0×10^{-3} M) (the concentration of Pd complex was taken twice that of ester to ensure that the ester is completely coordinated), ester (2.5×10^{-3} M) and NaNO_3 (0.1 M) at the required temperature under nitrogen flow and the pH was brought to the desired value by addition of 0.05 M NaOH solution. The hydrolysis was then followed by addition of 0.05 M NaOH solution to maintain the given pH constant. The data fitting was performed with the OLIS KINFIT set of programs [19] as described previously [20]. The precision of the kinetic data was estimated from the plot as obtained from the OLIS program output. The accepted residual values are less than 10^{-2} . Values of the hydroxide ion concentration were estimated from the pH using $\text{pK}_w = 13.997$ and an activity coefficient of 0.772 was determined from the Davies equation [21]. At the variable temperature studies, the following values of pK_w and γ were employed [22] at 15 $^\circ\text{C}$ ($\text{pK}_w = 14.35$, $\gamma = 0.776$), at 20 $^\circ\text{C}$ ($\text{pK}_w = 14.16$, $\gamma = 0.774$), at 25 $^\circ\text{C}$ ($\text{pK}_w = 14.00$, $\gamma = 0.772$), at 30 $^\circ\text{C}$ ($\text{pK}_w = 13.83$, $\gamma = 0.770$) and at 35 $^\circ\text{C}$ ($\text{pK}_w = 13.68$, $\gamma = 0.768$).

3. Results and discussion

The analytical data indicated that the complex is of 1:1 stoichiometry and of formula Pd(BHEB)Cl₂, which is given in scheme 1. IR spectra of BHEP and Pd(BHEP)Cl₂ are compared (figure 1). The stretching vibration of OH group of BHEP shows a broad band at 3151 cm⁻¹. This may be explained on the premise that the OH groups are involved in hydrogen bonding. This band is sharp in the complex and occurs at 3429 cm⁻¹, which indicates that the OH groups are not involved in coordination. The stretching vibration band corresponding to $\nu_{\text{Pd-N}}$ was assigned at 472 cm⁻¹. The stretching vibration band corresponding to $\nu_{\text{Pd-Cl}}$ was found to be a doublet at 333 and 324 cm⁻¹, which confirm *cis* configuration of the two Pd-Cl bonds [23, 24].



Scheme 1. Structure of [Pd(BHEP)Cl₂].

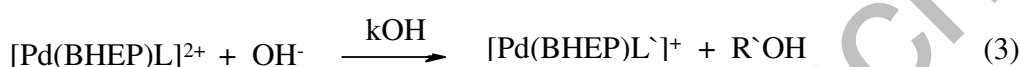
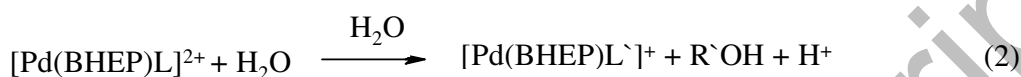
3.1. Thermal measurements

Thermogravimetric studies (TG) for the complex were carried out from room temperature up to 1000 °C with a heating rate of 10 degree/min. TG data are displayed in figure 2. The determined temperature ranges and percent mass losses of the solid complex on heating are given table 1. Analysis of the data revealed the following findings:

The TG curve of Pd(BHEP)Cl₂ exhibits a first estimated mass loss of 49.1% (calcd. 49.56%) at 150–360 °C, which may be attributed to the liberation of C₈H₁₈N₂O₂ as gases. The second estimated mass loss of 20.3% (calcd. 20.17 %), at 360–560 °C, may be attributed to the liberation of 2Cl atoms as gases. The residue corresponds to palladium metal 30.6% (calcd. 30.26%).

3.3. The catalysis of base hydrolysis of α -amino acid esters

α -Amino acid esters react with $[\text{Pd}(\text{BHEP})(\text{H}_2\text{O})_2]^{2+}$ according to equilibrium (1). The equilibrium constant is expected to be $\gg 1$. This is due to the high affinity of Pd^{II} to N-ligands [10]. The resulting mixed-ligand complexes $[\text{Pd}(\text{BHEP})\text{L}]^{2+}$ [$\text{L} = \text{NH}_2\text{CH}(\text{R})\text{CO}_2\text{R}'$] undergo hydrolysis by water and hydroxide ion according to equations (2) and (3).



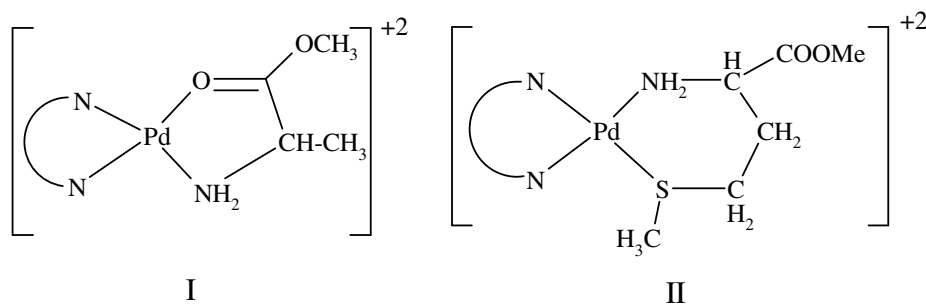
where $\text{L} = \text{NH}_2\text{CH}(\text{R})\text{CO}_2\text{R}'$ and $\text{L}' = \text{NH}_2\text{CH}(\text{R})\text{CO}_2^-$.

The kinetic data, *viz.* the volume of base added to keep the pH constant versus time, could be fitted by a single exponential function as shown in figure 3. Various other kinetic models were tested without satisfactory fits of the data. A plot of k_{obs} versus hydroxide ion concentration is linear as shown in figures 4 and 5 (for data see table 2) and follows the rate expression given in equation (4):

$$k_{\text{obs}} = k_o + k_{\text{OH}}[\text{OH}^-] \quad (4)$$

The k_o term is suggested to arise from spontaneous hydrolysis with water. The linear dependence of the rate on the OH^- concentration is consistent with direct attack of OH^- on the ester group (figures 4 and 5).

For the coordinated glycine methyl ester, the catalytic ratio is 1.45×10^5 . A catalytic ratio of this magnitude is consistent with the formation of the mixed ligand complex of structure I, where there is a direct interaction between Pd^{II} and the carbonyl group of the ester.



The catalytic ratio for methionine methyl ester complexes is 13.2 (table 3). The relatively low catalytic ratio suggests that the carbonyl group is not bound to Pd(II). The methionine methyl ester complex is expected to have structure II, where the amino and the sulfur are donors. Earlier studies [25-27] have shown that the formation of such complexes with a pendant ester group leads only to relatively small catalytic effects.

Comparative values of k_{OH} at 25 °C for the base hydrolysis of the glycine methyl ester incorporated in $[Pd(en)L]^{2+}$ is $4.88 \times 10^4 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ [18], where en = ethylenediamine. The k_{OH} value for $[Pd(BHEP)L]^{2+}$ (1.85×10^5) is higher than that of $[Pd(en)L]^{2+}$. The enhanced hydrolysis is probably due to stabilization of the mixed-ligand complex through involvement of the OH groups of BHEP in hydrogen bonding with the coordinated amino acid ester.

The activation parameters for the hydrolysis of coordinated glycine methyl ester were determined using Eyring plots of $\ln(k/T)$ versus $1/T$, figure 6 (for data see table 4). The activation parameters for k_{OH} were calculated to be $\Delta H^\ddagger = 28.0(\pm 0.2) \text{ kJ mol}^{-1}$ and $\Delta S^\ddagger = -49.5(\pm 0.3) \text{ JK}^{-1} \text{ mol}^{-1}$. The activation parameters for k_o may be inaccurate since the rate constants were determined by an extrapolation procedure. For base hydrolysis of the free glycine methyl ester, the activation parameters were reported [18, 28] to be $\Delta H^\ddagger = 39.7 \text{ kJ mol}^{-1}$ and $\Delta S^\ddagger = -117 \text{ JK}^{-1} \text{ mol}^{-1}$. The enhanced rate for base hydrolysis of the ester coordinated to the complex is therefore due to a decrease in ΔH^\ddagger and an increase in ΔS^\ddagger .

4. Conclusion

Thermogravimetric studies (TG) for $Pd(BHEP)Cl_2$ confirmed the structure and the absence of water of crystallization. The kinetic parameters (Supplementary Material) of thermal degradation process calculated using Coats–Redfern and Horowitz–Metzger methods show high values of the energy of activation of the complex reveal the high stability of such chelates and all the

decomposition steps are nonspontaneous processes. Also, the negative values of ΔS^* for the degradation process indicates more ordered activated complex than the reactants or the reaction is slow.

The hydrolysis of glycine methyl esters is catalyzed by $[\text{Pd}(\text{BHEP})(\text{H}_2\text{O})_2]^{2+}$ with catalysis ratio 1.45×10^5 . The catalytic effect is due to a direct interaction between Pd(II) and the alkoxy carbonyl group of the ester species. The activation parameters for the hydrolysis of coordinated glycine methyl ester were determined and compared with those of free glycine methyl ester. The enhanced rate for base hydrolysis of the ester coordinated to the complex is due to a decrease in ΔH^\ddagger and an increase in ΔS^\ddagger . The hydrolysis of methionine methyl ester is not significantly catalyzed. The relatively small catalysis ratio value suggests that the alkoxy carbonyl group is not bonded to the metal ion.

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Table 1. Scheme for TGA mass loss of Pd(BHEP)Cl₂ in the temperature range ~33 to 600 °C with heating rate of 10 degree/min.

Assignment loss	TGA °C	DrTGA °C	%Wt. Loss	
			Found	(Calcd.)
L(C ₈ H ₁₈ N ₂ O ₂)	150-360	255	49.1	(49.56)
2Cl	360-560	457	20.3	(20.17)
Pd (residue)	> 600		30.6	(30.26)

Table 2. Kinetics of hydrolysis of $[\text{Pd}(\text{BHEP})(\text{ester})]^{2+}$ at 25 °C and 0.1 M ionic strength.

Ester	pH	$[\text{OH}^-]^a$	k_{obs} (s^{-1})
Glycine methyl ester	4.8	7.94E-10	6.64E-04
	5.0	1.26E-09	8.19E-04
	5.2	2.00E-09	9.28E-04
	5.4	3.16E-09	1.17E-03
	5.6	5.01E-09	1.47E-03
Methionine methyl ester	8.8	7.94E-06	1.66E-04
	9.0	1.26E-05	2.25E-04
	9.2	2.00E-05	2.89E-04
	9.4	3.16E-05	4.06E-04
	9.6	5.01E-05	5.97E-04

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Table 3. Hydrolysis data ($\text{dm}^3\text{mol}^{-1}\text{s}^{-1}$) of $[\text{Pd}(\text{BHEP})(\text{ester})]^{2+}$ at 25 °C and 0.1 M ionic strength.

Ester	k_{OH}	$k_{\text{H}_2\text{O}}$	$k_{\text{OH}}^{(\text{ester})\text{a}}$	$k_{\text{OH}}/k_{\text{OH}}^{(\text{ester})}$
Glycine methyl ester	1.86E+05	1.00E-05	1.28	1.45E+05
Methionine methyl ester	10.1	1.71E-06	0.767	13.2

^aData from ref. (9).

Table 4. Hydrolysis data ($\text{dm}^3\text{mol}^{-1}\text{s}^{-1}$) of $[\text{Pd}(\text{BHEP})(\text{GlyOMe})]^{2+}$ at different temperatures in aqueous solution at pH=5.4.

T °C	k_{OH}	$k_{\text{H}_2\text{O}}$
15	1.24E+05	9.664E-06
20	1.51E+05	9.832E-06
25	1.86E+05	1.000E-05
30	2.41E+05	1.017E-05
35	2.76E+05	1.034E-05

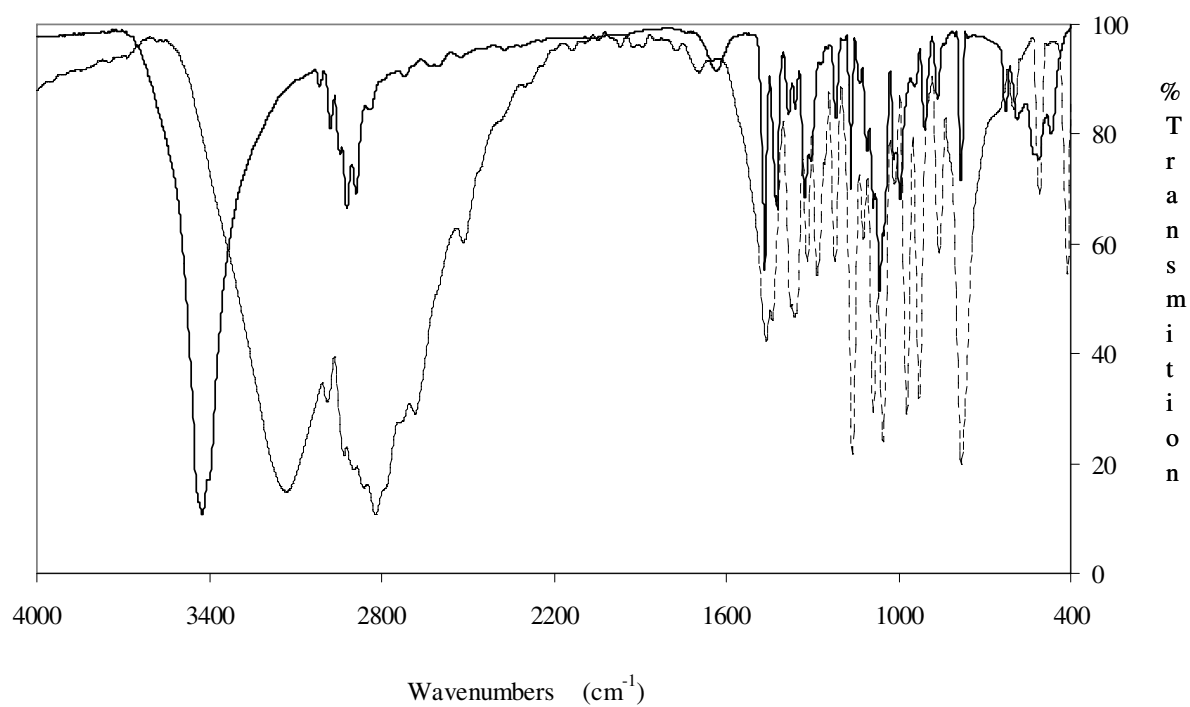


Figure 1a. Infrared spectrum of $[\text{Pd}(\text{BHEP})\text{Cl}_2]$ (-) and ligand BHEP (. . .).

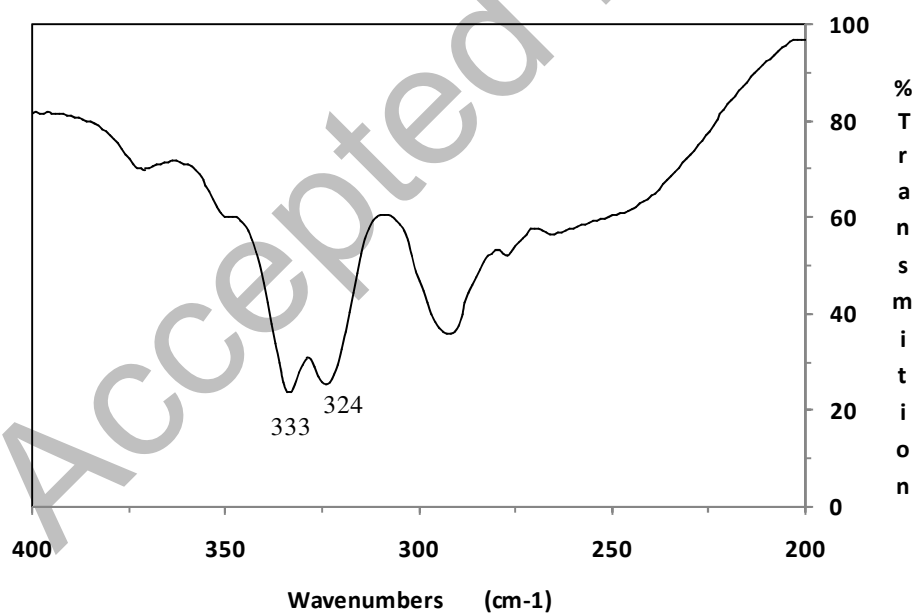


Figure 1b. Far-infrared spectrum of $[\text{Pd}(\text{BHEP})\text{Cl}_2]$.

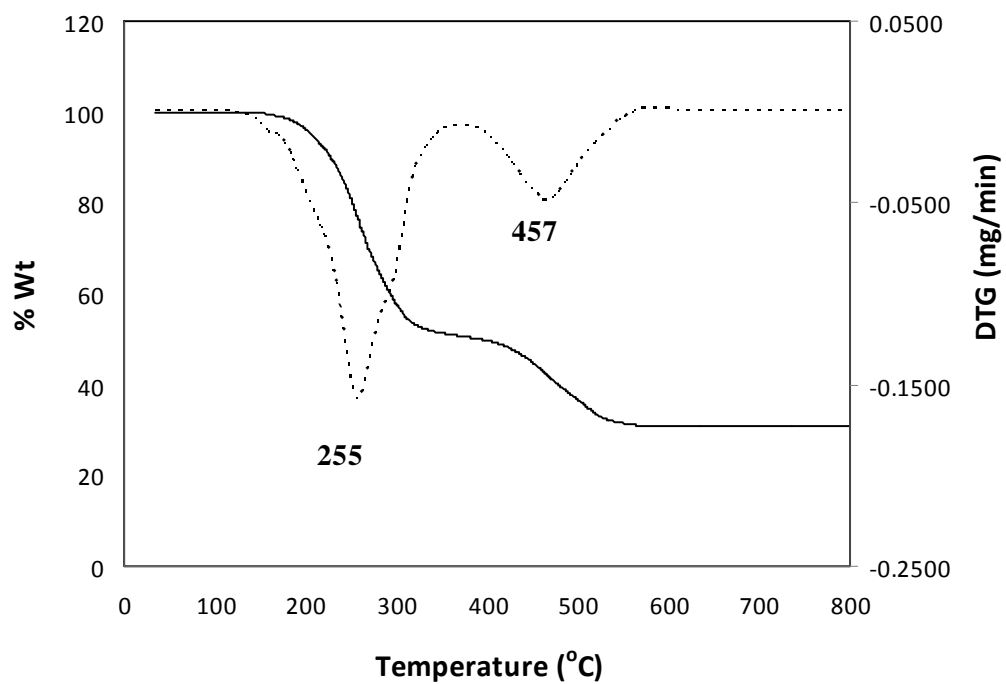


Figure 2. TGA (solid) and DrTGA (dotted) curves of Pd(BHEP)Cl₂.

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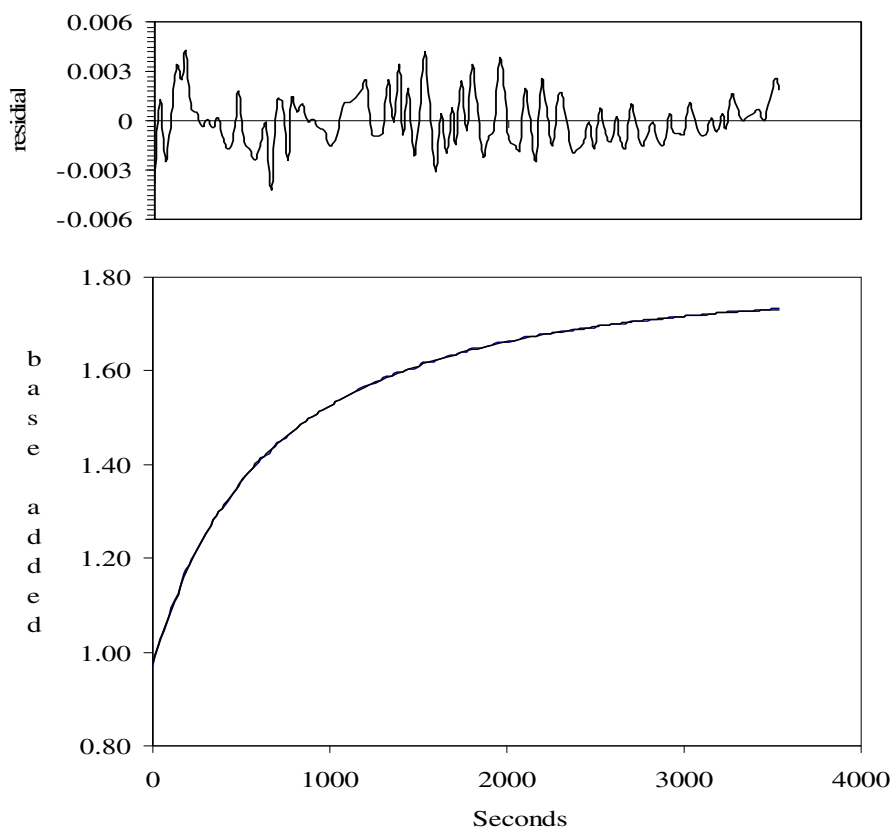


Figure 3. Typical plot of volume of base added vs. time in seconds fitted with a single exponential function for the hydrolysis of $[\text{Pd}(\text{BHEP})(\text{GlyOMe})]^{2+}$ at 25 °C and 0.1 M ionic strength.

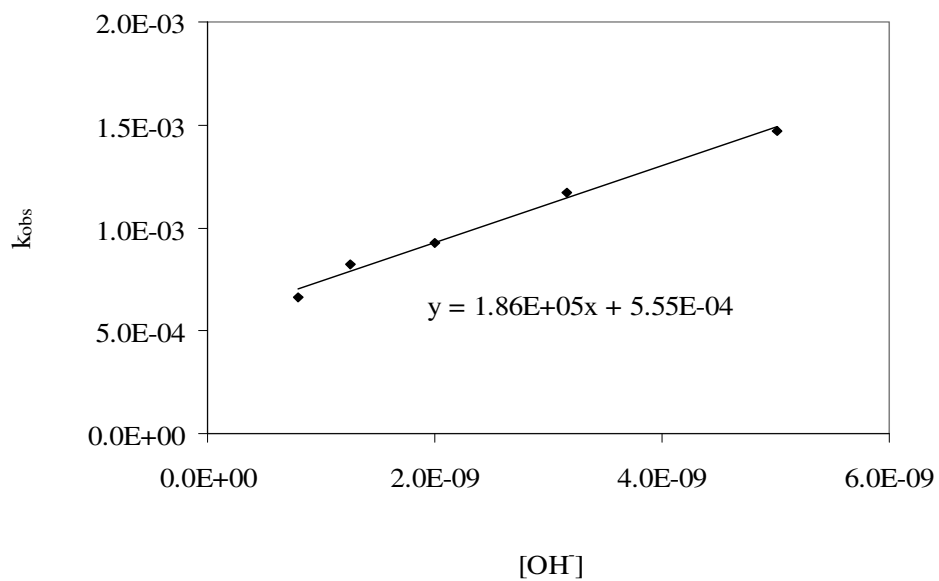


Figure 4. Kinetic data for the hydrolysis of Pd(BHEP)-glycine methyl ester at 25 °C and 0.1 M ionic strength.

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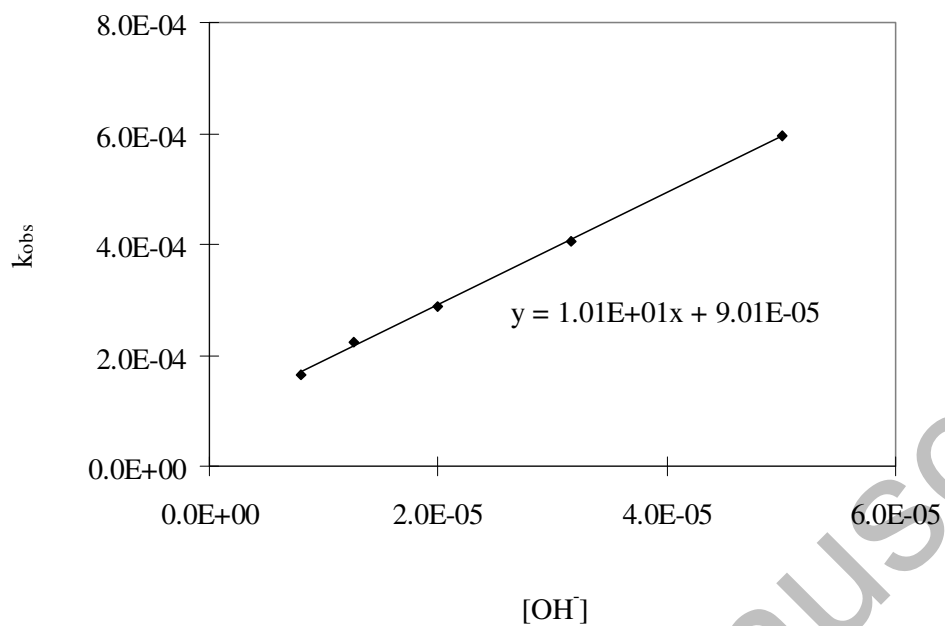


Figure 5. Kinetic data for the hydrolysis of Pd(BHEP)-methionine methyl ester at 25 °C and 0.1 M ionic strength.

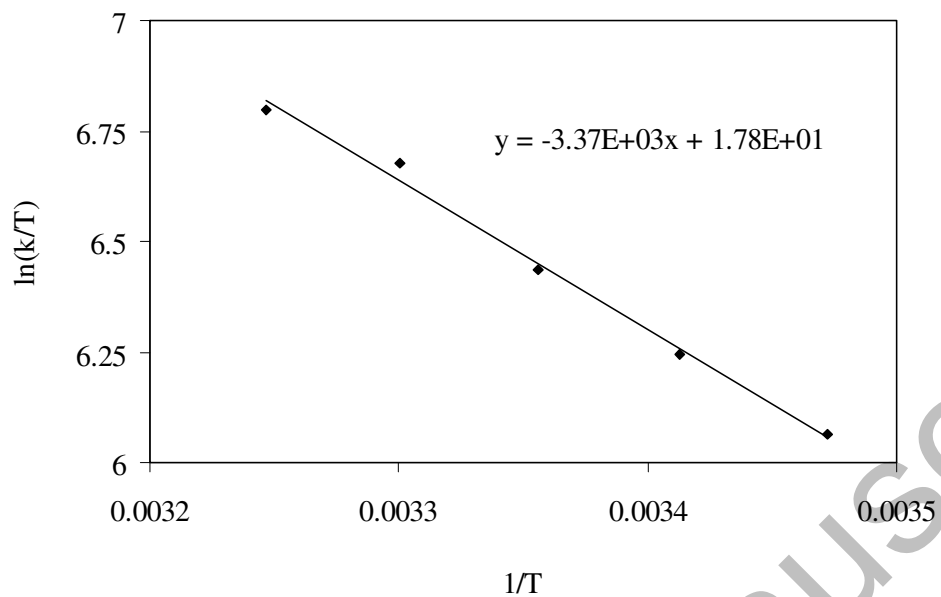


Figure 6. Effect of temperature on the hydrolysis of the coordinated glycine methyl ester.

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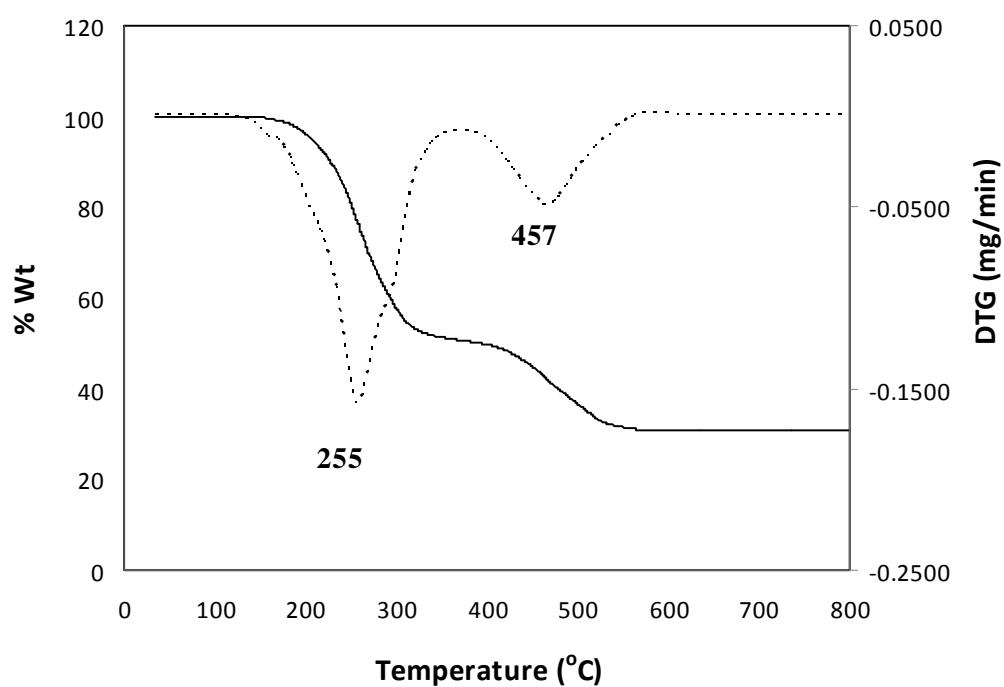
Graphical Abstract

THERMAL STABILITY OF Pd(1,4-BIS(2-HYDROXYETHYL)PIPERAZINE)Cl₂ AND ITS ROLE IN THE CATALYSIS OF BASE HYDROLYSIS OF α -AMINO ACID ESTERS

MOHAMED R. SHEHATA[†], MOHAMED M. SHOUKRY*[†], SHOKRY A. SHOKRY[‡] and M.A. MABROUK[‡]

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The thermal stability of Pd(BHEP)Cl₂ complex was investigated. The role of the complex in the catalysis of amino acid esters was studied.