



Stereoselective entry into the D-GalNAc series starting from the D-Gal one: a new access to N-acetyl-D-galactosamine and derivatives thereof[☆]

Lorenzo Guazzelli, Giorgio Catelani^{*}, Felicia D'Andrea, Alessia Giannarelli

Dipartimento di Chimica Bioorganica e Biofarmacia, Università di Pisa, Via Bonanno, 33-I-56126 Pisa, Italy

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ABSTRACT

A new stereoselective preparation of N-acetyl-D-galactosamine (**1b**) starting from the known *p*-methoxyphenyl 3,4-*O*-isopropylidene-6-*O*-(1-methoxy-1-methylethyl)-β-D-galactopyranoside (**10**) is described using a simple strategy based on (a) epimerization at C-2 of **10** via oxidation–reduction to give the *talo* derivative **11**, (b) amination with configurational inversion at C-2 of **11** via a S_N2-type reaction on its 2-imidazolate, (c) anomeric deprotection of the *p*-methoxyphenyl β-D-galactosamine glycoside **14**, (d) complete deprotection. Applying the same protocol to 2,3:5,6:3',4'-tri-*O*-isopropylidene-6'-*O*-(1-methoxy-1-methylethyl)-lactose dimethyl acetal (**4**), directly obtained through acetonation of lactose, the disaccharide β-D-GalNAcp-(1→4)-D-Glcp (**1a**) was obtained with complete stereoselectivity in good (40%) overall yield from lactose.

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1. Introduction

D-Galactosamine (D-GalNH₂) is, after D-glucosamine, the second most abundant natural aminosugar, isolated first in 1914 by Levene and La Forge² from chondroitin, a mucopolysaccharide in which D-GalNH₂ is present, as in the structurally related dermatan sulfate,³ as its acetamido derivative (2-acetamido-2-deoxy-D-galactopyranose, D-GalNAc, **1b**). D-GalNAc is also a constituent of several complex glycoproteins, as for examples: (i) the monosaccharide 'core' of mucines,⁴ (ii) oligosaccharide determinants of human blood group antigens,⁵ (iii) anti-freeze glycoproteins of anthartic fishes.⁵ Furthermore, D-GalNAc has been recently recognised as one of the strongest agonist of the NKR-P1 rat Natural Killer cells receptor.⁶ Owing to the biological relevance of D-GalNAc, several approaches to its synthesis in the free form and/or to the synthesis of its derivatives have been proposed, using different carbohydrate starting materials. The two most exploited synthetic channels to D-GalNAc and its derivatives are those based on (a) C-4 epimerization of the largely and cheaply available D-GlcNAc,⁷ mimicking thus the biosynthetic pathway; and (b) the amination at C-2 of D-galactose with formal retention of configuration.^{8–11} The first D-Gal to D-GalNAc transformation was described in 1976⁸ by Paulsen and co-workers using as key reaction the sodium azide opening of the epoxide ring of the intermediate 1,6:2,3-dian-

hydro-β-D-talopyranoside. A few years later, Lemieux and Ratcliffe⁹ used the azidonitration of tri-*O*-acetyl-D-galactal for the synthesis of some 2-azido-2-deoxy-D-galactopyranose derivatives and free D-GalNAc (**1b**). After this key paper, several addition reactions to D-galactal derivatives have been proposed to obtain D-GalNAc and its glycosides, as the Danishefsky's sulfamidoglycosylation,¹⁰ the Gin's acetamidoglycosylation.¹¹ Furthermore, specific syntheses of D-GalNH₂ starting from L-lyxose¹² and, very recently, D-tagatose¹³ have been proposed.

In the frame of a general project on the synthesis of β-D-hexosaminy-(1→4)-D-Glcp disaccharides,¹⁴ we considered the synthesis of β-D-GalNAcp-(1→4)-D-Glcp (**1a**), a natural disaccharide present in minute amount in the bovine colostrum.¹⁵ The sole reported synthesis of **1a** is based on an enzymatic glycosylation involving a β-(1→4)-N-acetylgalactosaminyltransferase.¹⁶ However, the above synthesis does not appear suitable for preparative purposes. Presented herein is a new efficient chemical method for the preparation of the disaccharide **1a** starting from lactose, involving a two-step amination with overall retention of configuration at C-2' (Chart 1), through the formation of a β-D-talopyranoside intermediate of type **2**. The potentiality of the method has also been demonstrated in the case of a monosaccharide β-D-galactopyranoside, that has been transformed into a known precursor of free D-GalNAc (**1b**) and of some 2-azido-2-deoxy-D-galactopyranoside glycosyl donors.

2. Results and discussion

A straightforward route to β-D-GalNAcp-(1→4)-D-Glcp (**1a**) was envisaged (Scheme 1) taking advantage of the easy availability of

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^{*} Corresponding author. Tel.: +39 0502219700; fax: +39 0502219660.

E-mail address: giocate@farm.unipi.it (G. Catelani).

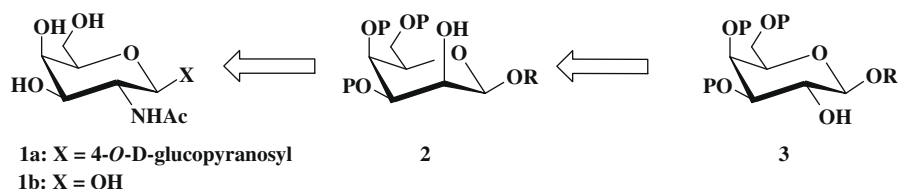
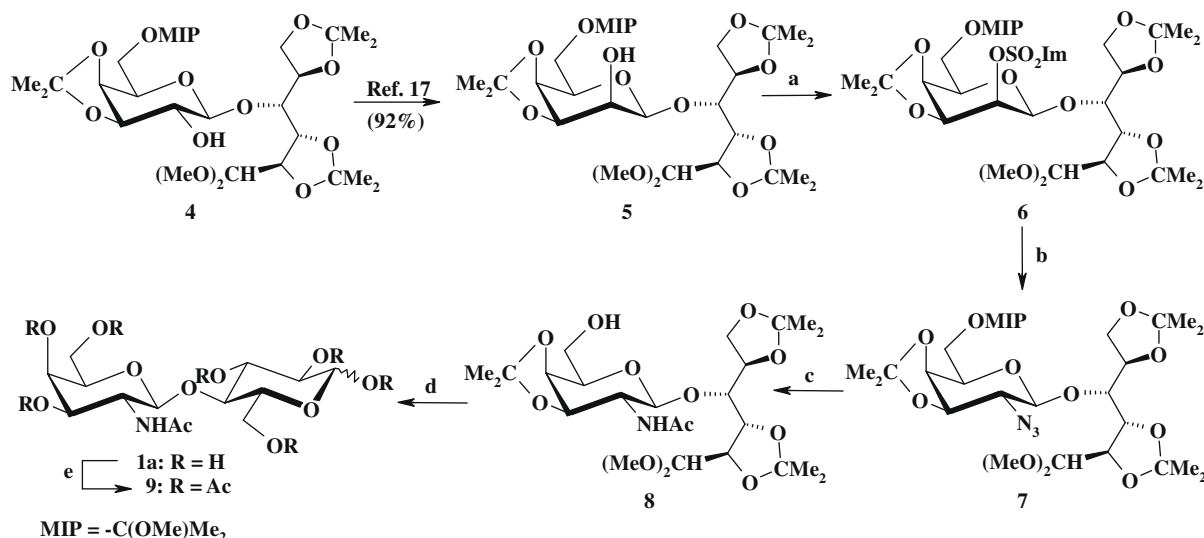


Chart 1. Retrosynthetic approach for transforming a β -D-galactopyranoside unit into a D-GalNAc one.



Scheme 1. Synthesis of β -D-GalNAcp-(1 \rightarrow 4)-D-Glcp disaccharide from lactose. Reagents and conditions: (a) Im₂SO₂, NaH, DMF, -30 °C, 4 h (86%); (b) NaN₃, DMF, 100 °C, 1.2 h (94%); (c) (1) NiCl₂·6H₂O, NaBH₄, MeOH, 0 °C, rt, 2 h; (2) Ac₂O, MeOH, rt, 2 h (86%); (d) 80% aq AcOH, 80 °C, 4 h (90%); (e) Ac₂O, Py, rt, 18 h (95%).

the tetraacetone **5**,¹⁷ prepared from its C-2' lactose epimer (**4**) through a completely stereoselective, high yielding (92%) oxidation–reduction sequence.¹⁷ The activation of **5** was made by treatment with imidazyl sulfate (Im₂SO₂) and NaH in DMF at -30 °C, resulting in the corresponding imidazylate **6**, isolated pure by flash chromatography in 86% yield. Compound **6** was then subjected to a S_N2 substitution reaction, with NaN₃ in DMF at 100 °C, which gave, after flash chromatography, the azido derivative **7** in 94% yield. This result confirms the usefulness of the imidazylate leaving group for performing efficient substitution in position 2 of a pyranoside, where other aryl and alkyl sulfonates are known to give unsatisfactory results.¹⁸

The reduction of the azido group of **7**, employing nickel chloride hexahydrate and sodium borohydride, afforded a single product which was directly submitted to N-acetylation (Ac₂O in MeOH). In the slightly acidic reaction conditions, the labile 6'-mixed acetal group was removed and compound **8** was obtained in 86% yield, after chromatographic purification. The target compound **1a** was instead prepared by complete deprotection of **8** using 80% aq AcOH at 80 °C: the reaction involves the O-deisopropylidenation and the C-1 aldehyde group exposition with con-

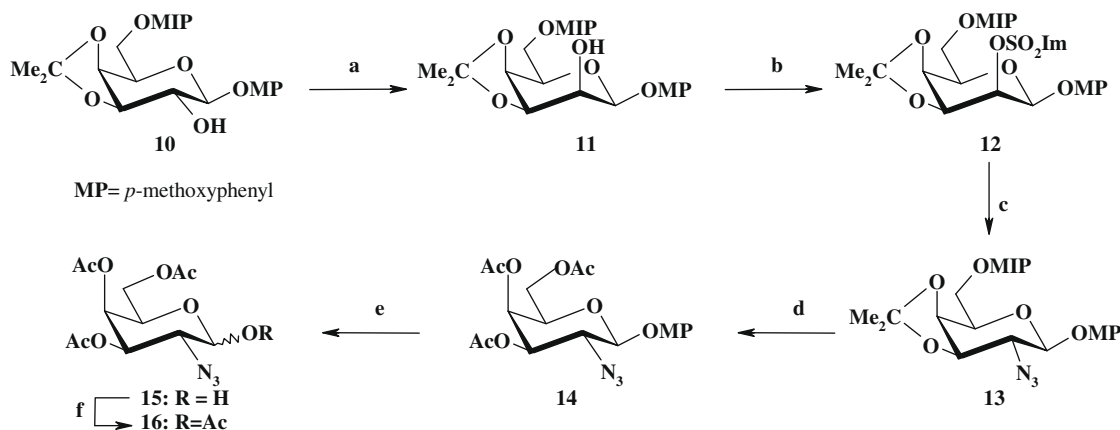
comitant six-membered ring closure. The structure of **1a** as well as its anomeric composition (α/β ratio about 2:3) was established on the basis of its NMR spectra. In particular, the ¹H NMR spectrum was identical to the reported one,¹⁵ while the ¹³C NMR signals, which have not yet been reported, were assigned by comparison (see Table 1) with those of α - and β -lactose,¹⁹ for the *gluco* portion, and with those of methyl 2-acetamido-2-deoxy- β -D-galactopyranoside²⁰ (not shown). NMR analysis of the acetylated disaccharide **9**, obtained by treatment of **1a** with Ac₂O in pyridine for 18 h, further confirmed the parent compound structure. It is worthy of note that the transformation of lactose into the 2'-aminated analogue is obtained with a seven-step process involving common reagents and simple manipulations in a very good overall yield (40%), certainly not easily achieved through chemical means starting from the two deprotected monosaccharide components. On the basis of this positive result, the same protocol of C-2 amination with retention of configuration was applied (Scheme 2) to *p*-methoxyphenyl 3,4-*O*-isopropylidene-6-*O*-(1-methoxy-1-methylethyl)- β -D-galactopyranoside (**10**), easily obtainable from commercial penta-*O*-acetyl- β -D-galactopyranose through a simple sequence.²¹ Also in this case, the introduction of the azido group

Table 1
¹³C NMR chemical shifts of α - and β -**1a** and related compounds^a

Compound	C-1'	C-2'	C-3'	C-4'	C-5'	C-6'	C-1	C-2	C-3	C-4	C-5	C-6
α -Lactose	103.6	72.0	73.5	69.5	76.2	62.0	92.7	72.2	72.4	79.3	71.0	61.0
β -Lactose	103.6	72.0	73.5	69.5	76.2	62.0	96.6	74.8	75.3	79.2	75.6	61.1
Me β -D-GalNAcp	103.1	53.0	71.9	68.6	75.8	61.7						
α - 1a	104.5	55.4	73.5	70.5	78.2	63.8	94.6	74.3	73.8	82.1	72.6	62.9 ^b
β - 1a	104.5	55.4	73.5	70.5	78.2	63.8	98.5	76.5	77.4 ^b	81.9	77.3 ^b	63.0 ^b

^a Spectra taken in D₂O. Internal reference: 1,4-dioxane for lactose (Ref. 19) and Me β -D-GalNAc (Ref. 20), TMSP for **1a**.

^b Assignments may be reversed.



Scheme 2. Formal synthesis of D-GalNAc and of 2-deoxy-2-azido-D-galactopyranosyl glycosyl donors. Reagents and conditions: (a) (1) TPAP, NMO, CH₂Cl₂, rt, 2 h; (2) NaBH₄, MeOH, 0 °C, 2 h (81% overall); (b) Im₂SO₂, NaH, DMF, −30 °C, 20 min (92%); (c) NaN₃, DMF, 100 °C, 2 h (96%); (d) (1) 80% aq AcOH, 80 °C, 2 h; (2) Ac₂O, Py, rt, 2 h (97% overall); (e) CAN, 3:1 Me₂CO–H₂O, 0 °C, rt, 30 min (81%); (f) Ac₂O, Py, rt, 15 h (95%).

in position 2 was achieved by a high yielding double stereoselective inversion sequence, providing first the preparation of the *talo* derivative **11** through an oxidation at C-2 (TPAP–NMO) followed by reduction of the crude uloside with NaBH₄ in MeOH. The stereoselectivity of the reduction was again complete leading to the exclusive formation of **11** (81% isolated yield) due to the β face shielding of the acetonide bridge, which completely inhibits the hydride attack on the same face. The axial hydroxyl function was then activated as sulfonyl imidazole (Im₂SO₂, and NaH in DMF at −30 °C) and **12** was obtained in 92% yield after chromatographic purification. The second inversion–amination was performed, as described for the disaccharide analogue, by treatment with NaN₃ in DMF at 100 °C, resulting in the desired azido derivative **13** in 96% yield. The two consecutive inversions of configuration were easily checked by ¹H NMR spectroscopy, on the basis of *J*_{1,2} and *J*_{2,3} values (2.1 and 4.5 Hz, respectively, for the *talo* derivative **11**, and 8.6 and 7.5 for the *galacto* one **13**).

Although some reported methods²² allow the direct transformation of the anomeric *p*-methoxyphenyl into other more reactive leaving groups, our next target was the known azido derivative **15**²³ and its peracetylated derivative **16**.²⁴

This goal was achieved by removing first the acetal functions with 80% aq AcOH and acetylation of the crude residue (Ac₂O–Py), resulting in **14** in almost quantitative yield (97%) after chromatographic purification. Removal of the anomeric *p*-methoxyphenyl group was performed with CAN, resulting in **15** (81% yield), which was quantitatively transformed into the peracetate **16** by routine acetylation.

Transformation of α-**16** into D-GalNAc hydrochloride has been described by Lemieux,⁹ while **15** constitutes the key precursor of a series of 2-azido-2-deoxy-D-galactopyranosyl donors.²⁵

In conclusion, we have performed an easy D-galactose to D-galactosamine transformation with complete stereoselectivity avoiding thus difficult diastereoisomeric separations. Using this new methodology, the first chemical synthesis of the β-D-GalNAc-(1→4)-D-Glc disaccharide has been achieved with excellent yield starting from lactose, while in the monosaccharide series, a new synthesis of D-GalNAc and of some 2-azido-2-deoxy-D-galactopyranosyl donors has been accomplished, complementing thus the existing approaches. This strategy of amination with overall retention of configuration (double inversion) gave good results in both the mono- and disaccharide series and seems to have general applicability and good potentiality in the construction of complex β-D-GalNAc containing oligosaccharides through a first β-galac-

tosylation followed by its C-2 amination with retention of configuration.

3. Experimental

3.1. General methods

General methods are those reported in Ref. 1. ¹H and ¹³C NMR spectra were recorded with an Avance II 250 spectrometer operating at 250.13 MHz (¹H) and 62.9 MHz (¹³C) in the reported solvent (internal standard Me₄Si) and the assignments were made, when possible, with DEPT, HETCOR and COSY experiments. Compounds **5**¹⁷ and **10**²¹ were prepared according to the reported procedures.

3.2. 4-O-[2-O-(1-Imidazolylsulfonyl)-3,4-O-isopropylidene-6-O-(1-methoxy-1-methylethyl)-β-D-talopyranosyl]-2,3:5,6-di-O-isopropylidene-aldehyde-D-glucose dimethyl acetal (**6**)

To a suspension of NaH in mineral oil (60%, 632 mg, 15.8 mmol) pre-washed with hexane under argon atmosphere and cooled to 0 °C, a soln of **5**¹⁷ (1.84 g, 3.17 mmol) in dry DMF (55 mL) was slowly added. The mixture was stirred at 0 °C for 30 min, cooled to −30 °C, treated with Im₂SO₂ (940 mg, 4.74 mmol) and further stirred until TLC analysis (EtOAc) revealed the complete disappearance of the starting material (4 h). The reaction mixture was then cooled to −40 °C, excess of NaH was destroyed by addition of MeOH (0.5 mL) followed by 10 min stirring, and partitioned between Et₂O (50 mL) and crushed iced-water. The organic phase was separated, and the aq layer extracted with Et₂O (4 × 50 mL). The collected organic phases were dried (MgSO₄), filtered and concentrated under diminished pressure. The flash chromatographic purification over silica gel of the reaction product (1:3 hexane–EtOAc + 0.1% Et₃N) gave pure **6** (1.93 g, 86%) as a syrup; [α]_D +2.7 (c, 1.06, CHCl₃); R_f 0.33 (1:3 hexane–EtOAc); ¹H NMR (CD₃CN): δ 7.98 (dd, 1H, *J*_{2,4} 1.3 Hz, *J*_{2,5} 0.9 Hz, Im-H-2), 7.44 (dd, 1H, *J*_{4,5} 1.7 Hz, Im-H-4), 7.05 (dd, 1H, Im-H-5), 4.92 (dd, 1H, *J*_{1',2'} 1.0 Hz, *J*_{2',3'} 5.7 Hz, H-2'), 4.82 (d, 1H, H-1'), 4.40 (dd, 1H, *J*_{3',4'} 5.8 Hz, H-3'), 4.36 (d, 1H, *J*_{1,2} 6.0 Hz, H-1), 4.27 (dd, 1H, *J*_{2,3} 7.7 Hz, H-2), 4.15 (dt, 1H, *J*_{4,5} 5.9 Hz, *J*_{5,6a} = *J*_{5,6b} 6.1 Hz, H-5), 4.12 (dd, 1H, *J*_{4',5'} 2.7 Hz, H-4'), 4.04 (dd, 1H, *J*_{3,4} 1.9 Hz, H-3), 3.94 (dd, 1H, *J*_{6a,6b} 8.6 Hz, H-6b), 3.85 (dd, 1H, H-6a), 3.84 (ddd, 1H, *J*_{5',6'a} 6.5 Hz, *J*_{5',6'b} 6.3 Hz, H-5'), 3.83 (dd, 1H, H-4), 3.63 (dd, 1H, *J*_{6'a,6'b} 9.6 Hz, H-6'b), 3.55 (dd, 1H, H-6'a), 3.41, 3.40 (2s, each 3H, 2 × OMe-1), 3.14 [s, 3H, C(OMe)Me₂], 1.37, 1.36, 1.32, 1.30, 1.29, 1.28 (6s, each 3H, 3 × CMe₂), 1.22, 1.20 [2s, each 3H, C(OMe)Me₂]; ¹³C NMR

(CD₃CN): δ 138.1 (Im-C-2), 130.9 (Im-C-5), 120.1 (Im-C-4), 110.7, 110.3, 109.2 (3 \times CMe₂), 106.5 (C-1), 100.9 [C(OMe)Me₂], 99.1 (C-1'), 79.9 (C-2'), 78.5 (C-4), 77.9 (C-3), 77.1 (C-5), 76.3 (C-2), 73.5 (C-5'), 71.9 (C-3'), 71.1 (C-4'), 66.5 (C-6), 60.3 (C-6'), 56.6, 54.7 (2 \times OMe-1), 48.9 [C(OMe)Me₂], 27.3, 27.1, 27.0, 25.8, 25.4, 25.3 (3 \times CMe₂), 24.7, 24.6 [C(OMe)Me₂]. Anal. Calcd for C₃₀H₅₀N₂O₁₅S: C, 50.69; H, 7.09; N, 3.94. Found: C, 50.89; H, 7.43; N, 5.29.

3.3. 4-O-[2-Azido-2-deoxy-3,4-O-isopropylidene-6-O-(1-methoxy-1-methylethyl)- β -D-galactopyranosyl]-2,3,5,6-di-O-isopropylidene-aldehydo-D-glucose dimethyl acetal (7)

A soln of **6** (3.85 g, 5.41 mmol) and NaN₃ (707 mg, 10.9 mmol) in dry DMF (100 mL) was stirred under argon atmosphere at 100 °C. After 1 h and 20 min, TLC analysis (EtOAc) revealed the complete disappearance of the starting material, the mixture was cooled to rt and partitioned between satd aq NaHCO₃ (50 mL) and Et₂O (50 mL). The organic phase was separated and the aq layer extracted with Et₂O (3 \times 50 mL). The organic extracts were dried (MgSO₄), filtered and concentrated under diminished pressure. Purification of the residue by flash chromatography over silica gel (7:3 hexane–EtOAc + 0.1% of Et₃N) gave pure **7** (3.08 g, 94%) as a clear syrup; $[\alpha]_D^{25}$ –30.0 (c, 1.0, CHCl₃); R_f 0.64 (EtOAc); ¹H NMR (CD₃CN): δ 4.63 (d, 1H, J_{1,2'} 8.5 Hz, H-1'), 4.34 (d, 1H, J_{1,2} 6.0 Hz, H-1), 4.32 (t, 1H, J_{2,3} 6.0 Hz, H-2), 4.25 (dt, 1H, J_{4,5} 4.7 Hz, J_{5,6a} = J_{5,6b} 6.4 Hz, H-5), 4.15 (dd, 1H, J_{3,4'} 5.4 Hz, J_{4,5'} 2.1 Hz, H-4'), 4.09 (dd, 1H, J_{6a,6b} 8.5 Hz, H-6b), 4.06 (dd, 1H, J_{3,4} 1.3 Hz, H-3), 4.01 (dd, 1H, H-6a), 3.94 (dd, 1H, H-4), 3.90 (dd, 1H, J_{2,3'} 8.3 Hz, H-3'), 3.84 (ddd, 1H, J_{5,6'a} 6.2 Hz, J_{5,6'b} 6.7 Hz, H-5'), 3.61 (dd, 1H, J_{6'a,6'b} 9.4 Hz, H-6'b), 3.53 (dd, 1H, H-6'a), 3.39, 3.38 (2s, each 3H, 2 \times OMe), 3.30 (dd, 1H, H-2'), 3.15 [s, 3H, C(OMe)Me₂], 1.49, 1.40, 1.33, 1.32, 1.30, 1.29 (6s, each 3H, 3 \times CMe₂), 1.29, 1.30 [2s, each 3H, C(OMe)Me₂]; ¹³C NMR (CD₃CN): δ 110.9, 110.7, 108.9 (3 \times CMe₂), 106.4 (C-1), 102.2 (C-1'), 100.9 [C(OMe)Me₂], 78.5 (C-3), 77.9 (C-5), 77.8 (C-3'), 76.7 (C-2), 76.2 (C-4), 73.8 (C-4'), 73.0 (C-5'), 67.3 (C-2'), 66.0 (C-6), 60.4 (C-6'), 56.3, 54.4 (2 \times OMe), 48.9 [C(OMe)Me₂]; 28.4, 27.6, 27.1, 26.7, 26.3, 25.2 (3 \times CMe₂), 24.7, 24.5 [C(OMe)Me₂]. Anal. Calcd for C₂₇H₄₇N₃O₁₂: C, 53.54; H, 7.82; N, 6.94. Found: C, 53.47; H, 7.62; N, 7.12.

3.4. 4-O-(2-Acetamido-2-deoxy-3,4-O-isopropylidene- β -D-galactopyranosyl)-2,3,5,6-di-O-isopropylidene-aldehydo-D-glucose dimethylacetal (8)

To a soln of **7** (1.40 g, 2.31 mmol) in MeOH (24 mL) cooled to 0 °C, NiCl₂·6H₂O (2.75 g, 11.5 mmol) and NaBH₄ (699 mg, 18.4 mmol) were added. The soln was warmed to rt and stirred for 2 h. To the mixture were then added brine (50 mL) and, after 10 min, water (50 mL) and CHCl₃ (50 mL). The organic phase was separated and the aq layer extracted with CHCl₃ (4 \times 50 mL). The collected organic extracts were dried (MgSO₄), filtered and concentrated at diminished pressure. The residue was dissolved in MeOH (30 mL), Ac₂O (7.5 mL) was added and the mixture was stirred at rt for 2 h when TLC analysis (EtOAc) showed the formation of a new product. The reaction mixture was repeatedly co-evaporated with toluene (4 \times 30 mL) under diminished pressure and purified by flash chromatography over silica gel (49:1 CHCl₃–MeOH) affording pure **8** (1.09 g, 86%) as a clear syrup; $[\alpha]_D^{25}$ +8.15 (c, 1.46, MeOH); R_f 0.10 (EtOAc); ¹H NMR (CD₃CN): δ 6.47 (d, 1H, J_{2',NH} 8.9 Hz, NH), 4.58 (d, 1H, J_{1,2'} 8.6 Hz, H-1'), 4.50 (dd, 1H, J_{1,2} 6.9 Hz, J_{2,3} 7.0 Hz, H-2), 4.34 (d, 1H, H-1), 4.36–4.05 (m, 4H, H-5, H-3', H-6'a, H-6'b), 3.97–3.70 (m, 4H, H-5', H-3, H-6a, H-6b), 3.51 (m, 2H, H-2, H-4), 3.43, 3.42 (2s, each 3H, 2 \times OMe), 1.90 (s, 3H, MeCON), 1.45, 1.40, 1.32, 1.31, 1.28, 1.26 (6s, each 3H, 3 \times CMe₂); ¹³C NMR (CD₃CN): δ 170.6 (MeCO), 110.2, 110.0, 108.5 (3 \times CMe₂), 107.6 (C-1), 101.6 (C-1'), 78.6 (C-3), 77.4, 77.3 (C-3', C-5), 75.7, 75.6 (C-

4, C-2), 74.6 (C-4'), 73.5 (C-5'), 65.6 (C-6), 62.5 (C-6'), 57.5, 54.6 (2 \times OMe), 55.2 (C-2'), 28.2, 27.4, 26.8, 26.4, 26.3, 24.8 (3 \times CMe₂), 23.3 (MeCON). Anal. Calcd for C₂₅H₄₃NO₁₂: C, 54.63; H, 7.89; N, 2.55. Found: C, 54.72; H, 7.91; N, 2.58.

3.5. 4-O-(2-Acetamido-2-deoxy- β -D-galactopyranosyl)- α , β -D-glucopyranose (1a)

A soln of **8** (450 mg, 0.82 mmol) in 80% aq AcOH (15 mL) was stirred at 80 °C for 4 h and then concentrated under diminished pressure by co-evaporation with toluene (4 \times 35 mL). The residue was triturated with EtOAc to give an amorphous white solid (283 mg, 90%) composed (¹³C NMR, D₂O) by a 2:3 α / β anomeric mixture of **1a**, as established on the basis of the integration of the H-1 signals; $[\alpha]_D^{25}$ +55.8 (c, 0.92, water); selected ¹H NMR (D₂O) data of α -**1a**: δ 5.21 (d, 1H, J_{1,2} 3.8 Hz, H-1), 4.52 (d, 1H, J_{1',2'} 8.4 Hz, H-1'); β -**1a**: δ 4.65 (d, 1H, J_{1,2} 8.4 Hz, H-1), 4.51 (d, 1H, J_{1',2'} 8.3 Hz, H-1'), 3.27 (dd, 1H, J_{2,3} 8.8 Hz, H-2); ¹³C NMR (D₂O) data of α -**1a** and β -**1a** see Table 1 and δ : 177.6 (MeCO), 25.0 (MeCO). Anal. Calcd for C₁₄H₂₅NO₁₁: C, 43.86; H, 6.57; N, 3.65. Found: C, 43.95; H, 6.59; N, 3.66.

3.6. 4-O-(2-Acetamido-2-deoxy-3,4,6-tri-O-acetyl- β -D-galactopyranosyl)- α , β -1,2,3,6-tetra-O-acetyl-D-glucopyranose (9)

Compound **1a** (50 mg, 0.13 mmol) was dissolved in 2:1 pyridine–Ac₂O (3 mL) and the resulting soln was stirred at rt for 18 h and then co-evaporated with toluene (3 \times 5 mL) under diminished pressure. Flash chromatographic purification, eluting with EtOAc, afforded pure **9** (84 mg, 95%) as an 1:1 α / β anomeric mixture, as established on the basis of the integration of the H-1 signals; syrup, $[\alpha]_D^{25}$ +16.0 (c, 1.04, CHCl₃); R_f 0.23 (EtOAc); ¹H NMR (CD₃CN) of α -**9**: δ 6.38 (d, 1H, J_{2',NH} 9.3 Hz, NH), 6.12 (d, 1H, J_{1,2} 3.8 Hz, H-1), 5.32 (dd, 1H, J_{2,3} 10.4 Hz, J_{3,4} 8.7 Hz, H-3), 5.02 (dd, 1H, J_{2,3'} 11.3 Hz, J_{3',4'} 3.5 Hz, H-3'), 4.90 (dd, 1H, H-2), 4.58 (d, 1H, J_{1',2'} 8.4 Hz, H-1'), 1.82 (s, 3H, MeCON); β -**9**: δ 6.36 (d, 1H, J_{2',NH} 9.3 Hz, NH), 5.74 (d, 1H, J_{1,2} 8.3 Hz, H-1), 5.24 (m, 1H, H-3), 5.03 (dd, 1H, J_{2,3'} 11.2 Hz, J_{3',4'} 3.5 Hz, H-3'), 4.92 (dd, 1H, J_{2,3} 9.7 Hz, H-2), 4.59 (d, 1H, J_{1',2'} 8.4 Hz, H-1'), 1.83 (s, 3H, MeCON); cluster of signals for both anomers: δ 4.40–3.95 (m, 14H, H-4, H-5, H-6a, H-6b, H-5', H-6'a, H-6'b), 3.86 (m 2H, H-2') 2.13, 2.08, 2.07, 2.06, 2.05, 2.04, 2.03, 2.02, 2.01, 2.00, 1.98, 1.97, 1.96, 1.90 (14s, each 3H, 14 \times MeCOO); ¹³C NMR (CD₃CN) α -**9**: δ 102.1 (C-1'), 89.6 (C-1), 76.1 (C-4), 71.3 (C-5, C-5'), 71.2 (C-3'), 70.6 (C-3), 70.2 (C-2), 67.5 (C-4'), 62.4 (C-6), 62.1 (C-6'), 51.4 (C-2'); β -**9**: δ 102.1 (C-1'), 92.2 (C-1), 75.8, 74.3, 73.3 (C-3, C-4, C-5), 71.3 (C-5'), 71.2 (C-3'), 70.3 (C-2), 68.5 (C-4'), 62.8 (C-6), 62.1 (C-6'), 51.4 (C-2'); cluster of signals for both anomers: δ 171.4–169.9 (MeCO), 23.2 (MeCON), 21.2–20.8 (MeCOO). Anal. Calcd for C₂₈H₃₉NO₁₈: C, 49.63; H, 5.80; N, 2.07. Found: C, 49.65; H, 5.83; N, 2.08.

3.7. 4-Methoxyphenyl 3,4-O-isopropylidene-6-O-(1-methoxy-1-methylethyl)- β -D-talopyranoside (11)

A mixture of **10**²¹ (320 mg, 0.803 mmol), pre-dried 4-methylmorpholine N-oxide (NMO, 165 mg, 1.41 mmol) and 4 Å powdered molecular sieves (500 mg) in anhyd CH₂Cl₂ (15 mL) was stirred for 30 min at rt under argon atmosphere. Tetrapropylammonium per-ruthenate (TPAP, 14.1 mg, 0.04 mmol) was then added and the resulting mixture was stirred for 2 h at rt until TLC (9:1 CH₂Cl₂–Me₂CO) revealed complete oxidation of **10**. The mixture was filtered through a Celite-silica gel–Celite triple alternate pad, the filter was washed with CH₂Cl₂ and then with EtOAc, and the organic phase was concentrated under diminished pressure. The residue was dissolved in MeOH (15 mL), NaBH₄ (121.4 mg, 3.21 mmol) was added and the mixture was stirred at 0 °C. After 2 h, TLC analysis

(9:1 CH₂Cl₂–Me₂CO) showed the complete disappearance of the 2-ulose. Water (8 mL) was added and the resulting soln was stirred for additional 30 min and then extracted with CH₂Cl₂ (3 × 30 mL). The organic extracts were collected, dried (MgSO₄), filtered and concentrated under diminished pressure. Purification of the residue by flash chromatography over silica gel (9:1 CH₂Cl₂–Me₂CO + 0.1% Et₃N) afforded pure **11** (259.2 mg, 81%) as a clear syrup; $[\alpha]_D^{25}$ –49.5 (c, 0.99, CHCl₃); *R*_f 0.43 (9:1 CH₂Cl₂–Me₂CO); ¹H NMR (CD₃CN): δ 7.02, 6.95 (AA'XX' system, 4H, Ar–H), 5.08 (d, 1H, *J*_{1,2} 2.1 Hz, H-1), 4.37 (dd, 1H, *J*_{3,4} 7.0 Hz, *J*_{2,3} 4.5 Hz, H-3), 4.30 (dd, 1H, *J*_{4,5} 2.1 Hz, H-4), 3.84 (ddd, 1H, *J*_{5,6a} 6.9 Hz, *J*_{5,6b} 5.2 Hz, H-5), 3.82 (dd, 1H, H-2), 3.73 (s, 3H, OMe), 3.59 (dd, 1H, *J*_{6a,6b} 10.0 Hz, H-6a), 3.55 (dd, 1H, H-6b), 3.12 [s, 3H, C(OMe)Me₂], 1.52, 1.32 (2s, each 3H, CMe₂), 1.28 (s, 6H, C(OMe)Me₂); ¹³C NMR (CD₃CN): δ 155.9, 152.4 (Ar–C), 118.9, 115.3 (Ar–CH), 110.5 (CMe₂), 100.8 [C(OMe)Me₂], 99.5 (C-1), 73.9 (C-3), 72.8 (C-4), 72.1 (C-5), 67.0 (C-2), 61.4 (C-6), 56.1 (OMe), 48.8 [C(OMe)Me₂], 25.8, 25.4 (CMe₂), 24.7, 24.6 [C(OMe)Me₂]. Anal. Calcd for C₂₀H₃₀O₈: C, 60.29; H, 7.59. Found: C, 60.47; H, 7.61.

3.8. 4-Methoxyphenyl 2-O-(1-imidazolylsulfonyl)-3,4-O-isopropylidene-6-O-(1-methoxy-1-methylethyl)-β-D-talopyranoside (**12**)

To a suspension of NaH in mineral oil (60%, 94 mg, 2.34 mmol) pre-washed with hexane under argon atmosphere and cooled to 0 °C, a soln of **11** (186.8 mg, 0.469 mmol) in dry DMF (8 mL) was added. The mixture was stirred at 0 °C for 30 min, cooled to –30 °C, treated with Im₂SO₂ (139 mg, 0.703 mmol) and further stirred at –30 °C. After 20 min, TLC analysis (3:7 hexane–EtOAc) revealed the complete disappearance of the starting material. The reaction mixture was then cooled to –40 °C, excess of NaH was destroyed by addition of MeOH (0.5 mL) followed by 10 min stirring, then partitioned between Et₂O (16 mL) and crushed iced-water. The organic phase was separated, and the aq layer extracted with Et₂O (2 × 16 mL). The collected organic phases were dried (MgSO₄), filtered and concentrated under diminished pressure. Flash chromatographic purification over silica gel of the reaction product (2:3 hexane–EtOAc + 0.1% of Et₃N) gave pure **12** (226.6 mg, 92%) as a clear syrup; $[\alpha]_D^{25}$ +4.2 (c, 1.18, CHCl₃); *R*_f 0.22 (2:3 hexane–EtOAc); ¹H NMR (CD₃CN): δ 8.07 (dd, 1H, *J*_{2,4} 1.3 Hz, *J*_{2,5} 0.8 Hz, Im–H-2), 7.50 (dd, 1H, *J*_{4,5} 1.7 Hz, Im–H-4), 7.10 (dd, 1H, Im–H-5), 6.83, 6.72 (AA'XX' system, 4H, Ar–H), 5.03 (dd, 1H, *J*_{1,2} 1.0 Hz, *J*_{2,3} 5.5 Hz, H-2), 4.93 (d, 1H, H-1), 4.51 (dd, 1H, *J*_{3,4} 5.8 Hz, H-3), 4.23 (dd, 1H, *J*_{4,5} 2.7 Hz, H-4), 4.01 (ddd, 1H, *J*_{5,6a} 7.4 Hz, *J*_{5,6b} 4.7 Hz, H-5), 3.71 (s, 3H, OMe), 3.65 (dd, 1H, *J*_{6a,6b} 10.3 Hz, H-6a), 3.57 (dd, 1H, H-6b), 3.09 [s, 3H, C(OMe)Me₂], 1.49, 1.33 (2s, each 3H, CMe₂), 1.29, 1.28 [2s, each 3H, C(OMe)Me₂]; ¹³C NMR (CD₃CN): δ 156.3, 151.3 (Ar–C), 138.3 (Im–C-2), 131.1 (Im–C-5), 120.0 (Im–C-4), 118.6, 115.2 (Ar–CH), 111.4 (CMe₂), 100.8 [C(OMe)Me₂], 97.3 (C-1), 79.3 (C-2), 73.8 (C-5), 71.9 (C-3), 71.4 (C-4), 60.7 (C-6), 56.1 (OMe), 48.8 [C(OMe)Me₂], 25.9, 25.4 (CMe₂), 24.7, 24.6 [C(OMe)Me₂]. Anal. Calcd for C₂₃H₃₂N₂O₁₀S: C, 52.26; H, 6.10; N, 5.30; S, 6.07. Found: C, 52.46; H, 6.18; N, 5.33; S, 6.11.

3.9. 4-Methoxyphenyl 2-azido-2-deoxy-3,4-O-isopropylidene-6-O-(1-methoxy-1-methylethyl)-β-D-galactopyranoside (**13**)

A soln of **12** (91.2 mg, 0.173 mmol) and NaN₃ (22.5 mg, 0.345 mmol) in dry DMF (4 mL) was stirred at 100 °C under argon atmosphere. After 2 h, the mixture was cooled to rt and partitioned between satd aq NaHCO₃ (8 mL) and Et₂O (15 mL). The organic phase was separated and the aq layer extracted with Et₂O (3 × 15 mL). The organic extracts were collected, dried (MgSO₄), filtered and concentrated under diminished pressure. Purification of the residue by flash chromatography over silica gel (7:3 hexane–EtOAc + 0.1% of Et₃N) gave pure **13** (71 mg, 96%) as a clear syrup; $[\alpha]_D^{25}$ +35.5 (c, 0.67, CHCl₃);

*R*_f 0.32 (7:3 hexane–EtOAc); ¹H NMR (CD₃CN): δ 7.03, 6.92 (AA'XX' system, 4H, Ar–H), 4.80 (d, 1H, *J*_{1,2} 8.6 Hz, H-1), 4.19 (dd, 1H, *J*_{3,4} 5.3 Hz, *J*_{4,5} 2.0 Hz, H-4), 4.02 (ddd, 1H, *J*_{5,6a} 7.0 Hz, *J*_{5,6b} 5.2 Hz, H-5), 3.98 (dd, 1H, *J*_{2,3} 7.5 Hz, H-3), 3.75 (s, 3H, OMe), 3.64 (dd, 1H, *J*_{6a,6b} 10.0 Hz, H-6a), 3.63 (dd, 1H, H-2), 3.59 (dd, 1H, H-6b), 3.12 [s, 3H, C(OMe)Me₂], 1.52, 1.32 (2s, each 3H, CMe₂), 1.31, 1.30 [2s, each 3H, C(OMe)Me₂]; ¹³C NMR (CD₃CN): δ 156.5, 151.8 (Ar–C), 119.1, 115.5 (Ar–CH), 111.1 (CMe₂), 101.4 (C-1), 100.8 [C(OMe)Me₂], 77.9 (C-3), 73.9 (C-4), 73.3 (C-5), 66.0 (C-2), 60.9 (C-6), 56.1 (OMe), 48.8 [C(OMe)Me₂], 28.4, 26.4 (CMe₂), 24.7, 24.6 [C(OMe)Me₂]. Anal. Calcd for C₂₀H₃₁N₃O₇: C, 56.46; H, 7.34; N, 9.88. Found: C, 56.48; H, 7.38; N, 9.91.

3.10. 4-Methoxyphenyl 2-azido-2-deoxy-3,4,6-tri-O-acetyl-β-D-galactopyranoside (**14**)

A soln of **13** (68 mg, 0.16 mmol) in 80% aq AcOH (3 mL) was stirred at 80 °C for 2 h, then concentrated under diminished pressure and co-evaporated with toluene (3 × 8 mL). The residue was dissolved in 2:1 pyridine–Ac₂O (3 mL) and the resulting soln was stirred at rt for 2 h and then co-evaporated with toluene (3 × 8 mL) under diminished pressure. Flash chromatographic purification over silica gel (7:3 hexane–EtOAc) afforded pure **14** (67.8 mg, 97%) as a clear syrup; $[\alpha]_D^{25}$ +6.02 (c, 0.83, CHCl₃); *R*_f 0.56 (2:3 hexane–EtOAc); ¹H NMR (CD₃CN): δ 7.03, 6.90 (AA'XX' system, 4H, Ar–H), 5.33 (dd, 1H, *J*_{3,4} 3.4 Hz, *J*_{4,5} 1.0 Hz, H-4), 5.00 (d, 1H, *J*_{1,2} 8.1 Hz, H-1), 4.92 (dd, 1H, *J*_{2,3} 10.8 Hz, H-3), 4.24–4.04 (m, 3H, H-6a, H-6b, H-5), 3.98 (dd, 1H, H-2), 3.75 (s, 3H, OMe), 2.21, 2.00, 1.99 (3s, each 3H, 3 × MeCO); ¹³C NMR (CD₃CN): δ 171.2, 171.1, 170.7 (3 × MeCO), 156.6, 151.6 (Ar–C), 119.0, 115.6 (Ar–CH), 101.6 (C-1), 71.9 (C-5), 71.8 (C-3), 67.5 (C-4), 62.4 (C-6), 61.6 (C-2), 56.1 (OMe), 20.8 (MeCO). Anal. Calcd for C₁₉H₂₃N₃O₉: C, 52.17; H, 5.30; N, 9.61. Found: C, 52.20; H, 5.33; N, 9.63.

3.11. 2-Azido-2-deoxy-3,4,6-tri-O-acetyl-α,β-D-galactopyranose (**15**)

To a soln of **14** (67 mg, 0.153 mmol) in 3:1 Me₂CO–water (6 mL), CAN (587 mg, 1.07 mmol) was added at 0 °C, the soln was warmed to rt and stirred for 30 min. It was then concentrated to 3 mL, diluted with CH₂Cl₂ (25 mL), washed with sat aq NaHCO₃ (2 × 15 mL), dried (MgSO₄), filtered and concentrated under diminished pressure. Purification of the residue by flash chromatography over silica gel (7:3 hexane–EtOAc) afforded known **15**²³ (41 mg, 81%) as a clear syrup composed (NMR, CDCl₃) by a mixture of α- and β-anomers in a 3:2 ratio calculated on the basis of the relative intensities of C-1 signals (δ 92.3 and 96.3, respectively); $[\alpha]_D^{25}$ +61.7, $[\alpha]_D^{25}$ +55.5 (c, 0.91, MeOH); *R*_f 0.20 (7:3 hexane–EtOAc); ¹H NMR (CDCl₃) of α-**15**: δ 5.43 (d, 1H, *J*_{1,2} 3.3 Hz, H-1), 5.37 (dd, 1H, *J*_{2,3} 10.0 Hz, *J*_{3,4} 3.2 Hz, H-3), 4.47 (m, 1H, H-5), 3.74 (dd, 1H, H-2), 3.91 (bs, 1H, OH-1); β-**15**: δ 4.83 (dd, 1H, *J*_{2,3} 10.9 Hz, *J*_{3,4} 3.3 Hz, H-3), 4.72 (dd, 1H, *J*_{1,2} 7.8 Hz, *J*_{1,OH} 5.3 Hz, H-1), 3.91 (m, 1H, H-5), 3.67 (dd, 1H, H-2), 4.58 (d, 1H, OH-1); cluster of signals for both anomers: δ 5.45 (m, 1H, H-4), 4.08–4.22 (m, 2H, H-6a, H-6b); 2.20–1.97 (m, 9H, 3 × MeCO); ¹³C NMR (CDCl₃) α-**15**: δ 92.3 (C-1), 67.6 (C-4), 66.4, 66.3 (C-3, C-5), 57.9 (C-2); β-**15**: δ 96.3 (C-1), 71.1 (C-3), 70.8 (C-5), 68.3 (C-4), 61.9 (C-2); cluster of signals for both anomers: δ 170.7–170.0 (MeCO), 61.7, 61.3 (C-6), 20.7–20.5 (MeCO). Anal. Calcd for C₁₂H₁₉N₃O₈: C, 43.24; H, 5.75; N, 12.61. Found: C, 43.26; H, 5.76; N, 12.62.

3.12. 2-Azido-2-deoxy-1,3,4,6-tetra-O-acetyl-α,β-D-galactopyranose (**16**)

A soln of **15** (32 mg, 0.10 mmol) in 2:1 pyridine–Ac₂O (2 mL) was stirred at rt for 15 h and then co-evaporated with toluene

(3 × 8 mL) under diminished pressure. Flash chromatographic purification over silica gel (1:1 hexane–EtOAc) afforded pure **16** (34 mg, 95%) as a clear syrup, composed (NMR, CDCl₃) by a mixture of α- and β-anomers in 1:1 ratio calculated on the basis of the relative integral of H-1 signals (δ 6.28 and 5.50, respectively); *R*_f 0.65 (1:1 hexane–EtOAc); [α]_D +49.7 (c, 0.92, CHCl₃); lit.²⁴ [α]_D +36.7 (c, 1.6, CHCl₃) for a 2:3 mixture of α- and β-anomers; NMR data were in full accordance with those reported.²⁴

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